

General methods for evolutionary quantitative genetic inference from generalised mixed models

Pierre de Villemereuil^{*}, Holger Schielzeth[†], Shinichi Nakagawa[‡], and Michael B. Morrissey[§]

^{*}Laboratoire d'Écologie Alpine, CNRS UMR 5553, Université Joseph Fourier, BP53
2233 rue de la piscine, 38041 Grenoble, Cedex 9, France
+33476635437
`bonamy@horus.ens.fr`

[†]Department of Evolutionary Biology, Bielefeld University
Morgenbreede 45, 33615 Bielefeld, Germany
+49-521-106 2820
`holger.schielzeth@uni-bielefeld.de`

[‡]Evolution and Ecology Research Centre
School of Biological, Earth and Environmental Sciences, University of New South Wales
Sydney, NSW 2052, Australia
+61 293 859 138
`s.nakagawa@unsw.edu.au`

[§]School of Biology, University of St Andrews
Dyers Brae House, St Andrews, UK, KY16 9TH
+44(0)1334463738
`michael.morrissey@st-andrews.ac.uk`

January 30, 2016

Keywords: quantitative genetics, generalised linear mixed model, statistics, theory, evolution, additive genetic variance, G matrix

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23

Abstract

Methods for inference and interpretation of evolutionary quantitative genetic parameters, and for prediction of the response to selection, are best developed for traits with normal distributions. Many traits of evolutionary interest, including many life history and behavioural traits, have inherently non-normal distributions. The generalised linear mixed model (GLMM) framework has become a widely used tool for estimating quantitative genetic parameters for non-normal traits. However, whereas GLMMs provide inference on a statistically-convenient latent scale, it will often be desirable to estimate quantitative genetic parameters on the scale upon which traits are expressed. The parameters of a fitted GLMM, despite being on a latent scale, fully determine all quantities of potential interest on the scale on which traits are expressed. We provide expressions for deriving each of such quantities, including population means, phenotypic (co)variances, variance components including additive genetic (co)variances, and parameters such as heritability. The expressions require integration of quantities determined by the link function, over distributions of latent values. In general cases, the required integrals must be solved numerically, but efficient methods are available and we provide an implementation in an R package, QGGLMM. We show that known formulae for quantities such as heritability of traits with Binomial and Poisson distributions are special cases of our expressions. Additionally, we show how a fitted GLMM can be incorporated into existing methods for predicting evolutionary trajectories. We demonstrate the accuracy of the resulting method for evolutionary prediction by simulation, and apply our approach to data from a pedigreed vertebrate population.

24 Introduction

25 Additive genetic variances and covariances of phenotypic traits determine the response to selec-
26 tion, and so are key determinants of the processes of adaptation in response to natural selection
27 and of genetic improvement in response to artificial selection (Fisher, 1918; Falconer, 1960;
28 Lynch and Walsh, 1998; Walsh and Lynch, forthcoming). While the concept of additive genetic
29 variance (Fisher, 1918; Falconer, 1960) is very general, being applicable to any type of character
30 with any arbitrary distribution, including, for example, fitness (Fisher, 1930), techniques for
31 estimating additive genetic variances and covariances are best developed for Gaussian traits
32 (i.e., traits with a normal distribution; Henderson 1950; Lynch and Walsh 1998). Furthermore,
33 quantitative genetic theory for predicting responses to selection are also best developed and
34 established for Gaussian characters (Walsh and Lynch, forthcoming), but see Morrissey (2015).
35 Consequently, although many characters of potential evolutionary interest are not Gaussian
36 (e.g. survival or number of offspring), they are not well-handled by existing theory and meth-
37 ods. Comprehensive systems for estimating genetic parameters and predicting evolutionary
38 trajectories of non-Gaussian traits will hence be very useful for quantitative genetic studies of
39 adaptation.

40 For Gaussian traits, a linear mixed model allows various analyses of factors that contribute to
41 the mean and variance of phenotype. In particular, a formulation of a linear mixed model called
42 the ‘animal model’ (Henderson, 1973; Kruuk, 2004; Wilson *et al.*, 2010) provides a very general
43 method for estimating additive genetic variances and covariances, given arbitrary pedigree
44 data, and potentially accounting for a range of different types of confounding variables, such as
45 environmental effects, measurement error or maternal effects. A general statement of an animal
46 model analysis decomposing variation in a trait, \mathbf{z} , into additive genetic and other components
47 would be

$$\mathbf{z} = \mu + \mathbf{X}\mathbf{b} + \mathbf{Z}_a\mathbf{a} + \mathbf{Z}_1\mathbf{u}_1 + \dots + \mathbf{Z}_k\mathbf{u}_k + \mathbf{e}, \quad (1)$$

48 where μ is the model intercept, and \mathbf{b} is a vector of fixed effects such as sex and age, relating
49 potentially to both continuous and categorical effects to observations via the fixed effects design
50 matrix \mathbf{X} , just as in an ordinary linear model. An arbitrary number of random effects can be
51 modelled, with design matrices \mathbf{Z} , where effects ($\mathbf{a}, \mathbf{u}_1 \dots \mathbf{u}_k$) are assumed to be drawn from

52 normal distributions with variances to be estimated. The key feature of the animal model is
53 that it includes individual additive genetic effects, or breeding values, conventionally denoted
54 \mathbf{a} . These additive genetic effects and, critically, their variance, are estimable given arbitrary
55 pedigree data, which defines the relatedness of all individuals in an analysis. The covariances
56 of breeding values among individuals can be modelled according to

$$\mathbf{a} \sim N(\mathbf{0}, \mathbf{A}V_A), \quad (2)$$

57 where \mathbf{A} is the additive genetic relatedness matrix derived from the pedigree and V_A is the
58 genetic additive variance.

59 Many non-Gaussian traits, however, cannot be strictly additive on the scale on which they are
60 expressed. Consider, for example, survival probability that is bounded at 0 and 1 so that effects
61 like the substitution effect of one allele for another necessarily must be smaller when expressed
62 in individuals that otherwise have expected values near zero or one. In such a scenario, it may
63 be reasonable to assume that there exists an underlying scale, related to survival probability,
64 upon which genetic and other effects are additive.

65 In addition to inherent non-additivity, analysis of many non-Gaussian traits will have com-
66 plex patterns of variation. Over and above sources of variation that can be modelled with fixed
67 and random effects, as in a LMM (e.g., using Eqs. 1 and 2), residual variation may include
68 both inherently stochastic components, and components that correspond to un-modelled sys-
69 tematic differences among observations. In a LMM, such differences are not distinguished, but
70 contribute to residual variance. However, for many non-Gaussian traits it may be desirable
71 to treat the former as arising from some known statistical distribution, such as the binomial
72 or Poisson distribution, and to deal with additional variation via a latent-scale residual (i.e.
73 an overdispersion term). Separation of these two kinds of variation in residuals may be very
74 generally useful in evolutionary quantitative genetic studies. For example, when observed data
75 represent observations (e.g., calling rate), but interest is in long-run average values (e.g., call-
76 ing effort over a season), it may be useful to exclude stochastic observation variance from
77 assessments of differences among individuals.

78 Generalised linear mixed model (GLMM) analysis can be used for inference of quantitative

79 genetic parameters, and provide pragmatic ways of dealing with inherent non-additivity and
80 with complex sources of variation. A latent scale is assumed (Fig. 1), on which effects on the
81 propensity for expression of some trait are assumed to be additive. A function, called a ‘link
82 function’, is applied that links expected values for a trait to the latent scale. For example,
83 a trait that is expressed in counts, say, number of behaviours expressed in a unit time, is a
84 strictly non-negative quantity. As depicted in Fig. 1, a strictly positive distribution of expected
85 values may be related to latent values ranging from $-\infty$ to $+\infty$ by a function such as the log
86 link. Finally, a distribution function is required to model the “noise” of observed values around
87 expected values (Fig. 1). Different distributions are suitable for different traits. For example,
88 with a count trait such as that depicted in Fig. 1, observed values may be modelled using the
89 Poisson distribution, with expectations related to the latent scale via the log link function.

90 The GLMM framework thus involves three scales on which we can think of variation in a
91 trait occurring. More formally, these three scales of the GLMM (see also Fig. 1) can be written:

92

$$\boldsymbol{\ell} = \boldsymbol{\mu} + \mathbf{X}\mathbf{b} + \mathbf{Z}_a\mathbf{a} + \mathbf{Z}_1\mathbf{u}_1 + \dots + \mathbf{Z}_k\mathbf{u}_k + \mathbf{o}, \quad (3a)$$

93

$$\boldsymbol{\eta} = g^{-1}(\boldsymbol{\ell}), \quad (3b)$$

94

$$\mathbf{z} \sim \mathcal{D}(\boldsymbol{\eta}, \boldsymbol{\theta}), \quad (3c)$$

95 where Eq. 3a is just as for a LMM (Eq. 1), except that it describes variation on the latent
96 scale $\boldsymbol{\ell}$, rather than the response directly. Note that we now refer to the “residual” (noted \mathbf{e}
97 in Eq. 1) as “overdispersion” (denoted \mathbf{o} , with a variance denoted V_O), as residuals (variation
98 around expected values) are defined by the distribution function, \mathcal{D} , in this model. Eq. 3b
99 formalises the idea of the link function. Any link function has an associated inverse link
100 function, g^{-1} , which is often useful for converting specific latent values to expected values. The
101 level of expected values is what we call the expected value scale. For example, where the log
102 link function translates expected values to the latent scale, its inverse, the exponential function,
103 translates latent values to expected values. Finally, Eq. 3c specifies the distribution by which
104 observations scatter around the expected values according to some distribution function, that
105 may involve parameters (denoted $\boldsymbol{\theta}$) other than the expectation. We call this the observed data

106 scale. Some quantities of interest, such as the mean, are the same on the expected data scale
107 and on the observed data scale. When parameters are equivalent on these two scales, we will
108 refer to them together as the data scale.

109 As for the LMM (Eq. 1), all random effects in a GLMM are assumed to follow normal
110 distributions, but on the latent scale. Particularly, the variance of additive genetic effects \mathbf{a}
111 are assumed to follow Eq. 2 on the latent scale. The expected data scale can be thought of
112 as the “intrinsic” value of individuals (shaped by both genetic and environmental effects), but
113 this intrinsic value can only be studied through random realisations. What matters more is a
114 topic of the problem at hand. For example, individuals (given their juvenile growth and genetic
115 value) might have an intrinsic annual reproductive success of 3.4, but can only produce integer
116 values of offspring each year (e.g. 2, or 3, or 5).

117 Linear mixed model-based inferences of genetic parameters, using the ‘animal model’, have
118 become common practice, particularly in evolutionary studies on wild populations (Kruuk,
119 2004; Wilson *et al.*, 2010). The use of generalised linear mixed animal model analysis is also
120 growing (e.g. Milot *et al.*, 2011; Wilson *et al.*, 2011; Morrissey *et al.*, 2012; de Villemereuil
121 *et al.*, 2013; Ayers *et al.*, 2013). However, whereas Gaussian animal model analysis directly
122 estimates additive genetic parameters on the scale on which traits are expressed and selected,
123 and upon which we may most naturally consider their evolution, this is not so for generalised
124 analyses. Genetic variance components estimated in a generalised animal model are obtained
125 on the latent scale. Hence, the “conventional” formula to compute heritability,

$$h_{\text{lat}}^2 = \frac{V_{A,\ell}}{V_{A,\ell} + V_{\text{RE}} + V_{\text{O}}} \quad (4)$$

126 where V_{RE} is the summed variance of all random effects apart from the additive genetic variance,
127 and V_{O} is the overdispersion variance, h_{lat}^2 is the heritability on the latent scale, not on the
128 observed data scale. Here, and throughout this paper, $V_{A,\ell}$ stands for the additive genetic
129 variance on the latent scale. Although it might sometimes be sensible to measure the heritability
130 of a trait on the latent scale (for example, in animal breeding, where selection might be based
131 on latent breeding values), it is natural to seek inferences on the scale upon which the trait
132 is expressed, and on which we may think of selection as acting. Some expressions exist by

133 which various parameters can be obtained or approximated on the observed data scale. For
134 example, various expressions for the intra-class correlation coefficients on the data scale exist
135 (reviewed in Nakagawa and Schielzeth, 2010), but these do not provide inferences of additive
136 genetic variance on the data scale. Exact analytical expressions exist for the additive genetic
137 variance and heritability on the observed data scale for two specific and important families of
138 GLMMs (i.e. combinations of link functions and distribution functions): for a binomial model
139 with a probit link function (i.e., the “threshold model,” Dempster and Lerner, 1950) and for
140 a Poisson model with a logarithm link function (Foulley and Im, 1993). A general system for
141 calculating genetic parameters on the expected and observed data scales for arbitrary GLMMs
142 is currently lacking.

143 In addition to handling the relationship between observed data and the latent trait via
144 the link and distribution functions, any system for expected and observed scale quantitative
145 genetic inference with GLMMs will have to account for complex ways in which fixed effects
146 can influence quantitative genetic parameters. It is currently appreciated that fixed effects
147 in LMMs explain variance, and that variance associated with fixed effects can have a large
148 influence on summary statistics such as repeatability (Nakagawa and Schielzeth, 2010) and
149 heritability (Wilson, 2008). This principle holds for GLMMs as well, but fixed effects cause
150 additional, important complications for interpreting GLMMs. While random and fixed effects
151 are independent in a GLMM on the latent scale, the non-linearity of the link function renders
152 them inter-related on the expected and observed scales. Consider, for example, a GLMM with
153 a log link function. Because the exponential is a convex function, the influence of fixed and
154 random effects will create more variance on the expected and observed data scales for larger
155 values than for smaller values.

156 While it will undoubtedly be desirable to develop a comprehensive method for making data-
157 scale inferences of quantitative genetic parameters with GLMMs, such an endeavour will not
158 yield a system for predicting evolution in response to natural or artificial selection, even if a
159 particular empirical system is very well served by the assumptions of a GLMM. This is because
160 systems for evolutionary prediction, specifically the Breeder’s equation (Lush, 1937; Fisher,
161 1924) and the Lande equation (Lande, 1979; Lande and Arnold, 1983), assume that breeding
162 values (and in most applications, phenotypes) are multivariate normal (Walsh and Lynch,

163 forthcoming). Even if it is possible to estimate additive genetic variances of traits on the
164 expected and observed data scales, we will show that these quantities will not strictly be usable
165 for evolutionary prediction. However, the latent scale in a GLMM does, by definition, satisfy
166 the assumptions of the Breeder's and Lande equations. Thus, for the purpose of predicting
167 evolution, it may be useful to be able to express selection of non-Gaussian traits, not on the
168 observed scale, but rather on the latent scale. Such an approach could yield a system for
169 evolutionary prediction of characters that have been modelled with a GLMM, requiring no
170 more assumptions than those that are already made in applying the statistical model.

171 We propose a system for making inferences of quantitative genetic parameters on the ex-
172 pected and observed scales, for arbitrary GLMMs. We show how to estimate genotypic and
173 additive genetic variances and covariances on the expected and observed data scale, accounting
174 for fixed effects as necessary. We lay out the formal theory underlying the system, apply it to an
175 empirical dataset, and provide software for implementation. The relationships between existing
176 analytical formulae and our general framework are also highlighted. Next, we outline a system
177 of evolutionary prediction for non-Gaussian traits that capitalises on the fact that the latent
178 scale in a GLMM satisfies the assumptions of available equations for the prediction of evolution.
179 We show in a simulation study that (i) evolutionary predictions using additive genetic variances
180 on the observed data scale represent approximations, and can, in fact, give substantial errors,
181 and (ii) that making inferences via the latent scale provides unbiased predictions, insofar as a
182 GLMM may provide a pragmatic model of variation in non-Gaussian traits. We also provide
183 software for making evolutionary predictions using the latent scale. Although all examples and
184 most equations in this article are presented in a univariate form, all our results are applicable
185 to multivariate analysis, which is implemented in our software. Together, these approaches
186 provide a comprehensive treatment of the evolutionary quantitative genetics of traits that may
187 be modelled with GLMMs.

188 **Quantitative genetic parameters in GLMMs**

189 Throughout this section we will refer to the additive genetic variance as defined on the latent
190 scale as $V_{A,\ell}$, the summed variance of other random effects on the latent scale as V_{RE} and the

191 overdispersion variance (i.e. the variance of \mathbf{o}) as V_{O} .

192 Phenotypic mean and variances

193 **Expected population mean** The expected mean phenotype on the data scale (i.e., applying to
194 both the mean expected value and mean observed value) is given by

$$\bar{z} = \int g^{-1}(\ell) f(\ell) d\ell, \quad (5)$$

195 where $f(\ell)$ is the probability density of ℓ . Typically, and especially in the absence of fixed
196 effects, the distribution of ℓ will be normal with a mean μ and variance $V_{\text{A},\ell} + V_{\text{RE}} + V_{\text{O}}$.

197 In the presence of fixed effects, it is necessary to average over the components of the predictive
198 values marginalised over the random effects (i.e. $\mathbf{X}\hat{\mathbf{b}}$, where $\hat{\mathbf{b}}$ are the fixed effects estimates)
199 as well as integrating over the random parts of ℓ ,

$$\bar{z} = \frac{1}{N} \sum_{i=1}^N \int g^{-1}(\ell) f_{\mathcal{N}}(\ell, \mu + \hat{\ell}_i, V_{\text{A},\ell} + V_{\text{RE}} + V_{\text{O}}) d\ell, \quad (6)$$

200 where N is the number of predicted latent values in $\hat{\ell} = \mathbf{X}\hat{\mathbf{b}}$. Typically, \mathbf{X} will be the fixed
201 effects design matrix used when fitting the generalised animal model (equations 1, 2, and 3),
202 and N will be the number of data observations. However, \mathbf{X} could profitably be modified to
203 a general prediction matrix in some scenarios. For example, if a model included a fixed effect
204 for sex, and if the population in question had an equal sex ratio but the data did not, an \mathbf{X}
205 matrix might be used that represented both sexes equally. Throughout the rest of this section,
206 and for the sake of clarity, we will assume the simple case of no fixed effects, but all equations
207 can easily be transformed as for Eq. 6. We will only specify versions of a few fundamental
208 equations that account for fixed effects.

209 **Expected-scale phenotypic variance** Phenotypic variance on the expected data scale can be
210 obtained analogously to the data scale population mean. Having obtained \bar{z} , the phenotypic
211 variance is

$$V_{\text{P,exp}} = \int (g^{-1}(\ell) - \bar{z})^2 f(\ell) d\ell. \quad (7)$$

212 **Observed-scale phenotypic variance** Phenotypic variance of observed values is the sum of the
213 variance in expected values and variance arising from the distribution function. Since these
214 variances are independent by construction in a GLMM, they can be added. This distribution
215 variance is influenced by the latent trait value, but might also depend on additional distribution
216 parameters included in $\boldsymbol{\theta}$ (see Eq. 3c). Given a distribution-specific variance function v :

$$V_{P,\text{obs}} = V_{P,\text{exp}} + \int v(\ell, \boldsymbol{\theta}) f(\ell) d\ell. \quad (8)$$

217 **Genotypic variance on the data scale, arising from additive genetic variance on the**
218 **latent scale**

219 Because the link function is non-linear, additive genetic variance on the latent scale is manifested
220 as a combination of additive and non-additive variance on the data scale. Following Falconer
221 (1960) the genotypic variance, as opposed to (additive) genetic, on the data scale is the variance
222 of genotypic values on that scale. Genotypic values are the expected data scale phenotypes,
223 given latent scale genetic values. The expected phenotype of an individual with a given latent
224 genetic value a , i.e., its genotypic value on the data scale $E[z|a]$, is given by

$$E[z|a] = \int g^{-1}(\ell) f(\ell|a) d\ell, \quad (9)$$

225 where $f(\ell|a)$ is the density of the latent trait for a given value of a . For example, in absence of
226 fixed effects, $f(\ell|a)$ would be $f_{\mathcal{N}}(\ell, \mu + a, V_{\text{RE}} + V_{\text{O}})$.

227 The genotypic variances on the expected and observed data scales are the same, since geno-
228 typic values are expectations that do not change between the expected and observed scales.
229 The genotypic variance on both the expected and observed data scales is then

$$V(E[z|a]) = \int (E[z|a] - \bar{z})^2 f_{\mathcal{N}}(a, 0, V_{A,\ell}) da. \quad (10)$$

230 This is the genotypic variance on the data scale, arising from strictly additive genetic variance
231 on the latent scale. If non-additive genetic effects are modelled on the latent scale, they would
232 be included in the expectations and integrals in Eqs. 9 and 10.

233 Additive genetic variance on the data scale

234 That part of the genotypic variance on the data scale (arising under a model that is additive on
235 the latent scale) that is additive is the variance of breeding values on the data scale. Following
236 Robertson (1950; see also Fisher 1918), breeding values on the data scale, i.e., a_{exp} and a_{obs} , are
237 the part of the phenotype z that depends linearly on the latent breeding values. The breeding
238 values on the data scale can then be defined as the predictions of a least-squares regression of
239 the observed data on the latent breeding values,

$$a_{\text{obs}} = \hat{z}|a = m + ba, \quad (11)$$

240 where \hat{z} is the value of z predicted by the regression. Thus, we have $V_{A,\text{obs}} = b^2 V_{A,\ell}$ and, from
241 standard regression theory:

$$b = \frac{\text{cov}(z, a)}{V_{A,\ell}}. \quad (12)$$

242 Because of the independence between the expected values of z (i.e. the expected data scale
243 $g^{-1}(\ell)$) and the distribution “noise” (see Eq. 8), we can obtain the result that $\text{cov}(z, a) =$
244 $\text{cov}(g^{-1}(\ell), a)$, hence:

$$b = \frac{\text{cov}(g^{-1}(\ell), a)}{V_{A,\ell}}. \quad (13)$$

245 Stein’s (1973) lemma states that if X and Y are bivariate normally distributed random variables,
246 then the covariance of Y and some function of X , $f(X)$, is equal to the expected value of $f'(X)$
247 times the covariance between X and Y , so,

$$\text{cov}(g^{-1}(\ell), a) = E \left[\frac{dg^{-1}(\ell)}{d\ell} \right] \text{cov}(\ell, a) = E \left[\frac{dg^{-1}(\ell)}{d\ell} \right] V_{A,\ell}, \quad (14)$$

248 noting that the covariance of latent breeding values and latent values is the variance of breeding
249 values. Finally, combining Eq. 13 with Eq. 14, we obtain:

$$b = E \left[\frac{dg^{-1}(\ell)}{d\ell} \right]. \quad (15)$$

250 To avoid confusion with various uses of b as other forms of regression coefficients, and for
251 consistency with Morrissey (2015), we denote the average derivative of expected value with

252 respect to latent value as Ψ . In the absence of fixed effects in the model, Ψ is

$$\Psi = \int \frac{dg^{-1}(\ell)}{d\ell} f_{\mathcal{N}}(\ell, \mu, V_{A,\ell} + V_{RE} + V_O) d\ell. \quad (16)$$

253 If fixed effects (other than the intercept μ) are included in the model, the equation above should
254 be modified accordingly:

$$\Psi = \frac{1}{N} \sum_{i=1}^N \int \frac{dg^{-1}(\ell)}{d\ell} f_{\mathcal{N}}(\ell, \mu + \hat{\ell}_i, V_{A,\ell} + V_{RE} + V_O) d\ell. \quad (17)$$

255 The additive genetic variance on the data scale is given by

$$V_{A,obs} = V_{A,exp} = \Psi^2 V_{A,\ell}. \quad (18)$$

256 An alternative derivation of equation 17, and its associated definition of Ψ , for the general
257 calculation of the additive genetic variances following a non-linear transformation, is given in
258 Morrissey (2015).

259 **Summary statistics and multivariate extensions**

260 Equations 5 through 18 give the values of different parameters that are useful for deriving other
261 evolutionary quantitative genetic parameters on the observed data scale. Hence, from them,
262 other parameters can be computed. The narrow-sense heritability on the observed data scale
263 can be written as

$$h_{obs}^2 = \frac{V_{A,obs}}{V_{P,obs}}. \quad (19)$$

264 Replacing $V_{P,obs}$ by $V_{P,exp}$ will lead to the heritability on the expected data scale h_{exp}^2 :

$$h_{exp}^2 = \frac{V_{A,exp}}{V_{P,exp}}. \quad (20)$$

265 Parameters such as additive genetic coefficient of variance and evolvability (Houle, 1992) can
266 be just as easily derived. The coefficient of variation on the expected and observed data scales

267 are identical and can be computed as

$$CV_{A,obs} = CV_{A,exp} = 100 \frac{\sqrt{V_{A,exp}}}{\bar{z}}, \quad (21)$$

268 and the evolvability on the expected and observed data scales will be

$$I_{A,obs} = I_{A,exp} = \frac{V_{A,exp}}{\bar{z}^2}. \quad (22)$$

269 The genetic basis of multivariate phenotype, especially as summarised by the \mathbf{G} matrix is
270 often of interest. For simplicity, all expressions considered to this point have been presented in
271 univariate form. However, every expression has a fairly simple multivariate extension. Multi-
272 variate phenotypes are typically analysed by multi-response GLMMs. For example, the vector
273 of mean phenotypes in a multivariate analysis on the expected data scale is obtained by defining
274 the link function to be a vector-valued function, returning a vector of expected values from a
275 vector of values on the latent scale. The phenotypic variance is then obtained by integrating
276 the vector-valued link function times the multivariate normal distribution total variance on
277 the latent scale, as in Eq. 5 and Eq. 8. Integration over fixed effects for calculation of the
278 multivariate mean is directly analogous to the extension of Eq. 5 given in Eq. 6. Calculation of
279 other parameters, such as multivariate genotypic values, additive-derived covariance matrices,
280 and phenotypic covariance matrices, have directly equivalent multivariate versions as well. The
281 additive genetic variance-covariance matrix (the \mathbf{G} matrix) on the observed scale is simply the
282 multivariate extension of equation 18, i.e., $\mathbf{G}_{obs} = \mathbf{\Psi} \mathbf{G}_\ell \mathbf{\Psi}^T$. Here, \mathbf{G}_ℓ is the latent \mathbf{G} matrix and
283 $\mathbf{\Psi}$ is the average gradient matrix of the vector-valued link function, which is a diagonal matrix
284 of Ψ values for each trait (simultaneously computed from a multivariate version of Eq. 16).

285 Relationships with existing analytical formulae

286 Binomial distribution and the threshold model

287 Heritabilities of binary traits have a long history of analysis with a threshold model (Wright,
288 1934; Dempster and Lerner, 1950), whereby an alternate trait category is expressed when a
289 trait on a latent “liability scale” exceeds a threshold. It can be shown (see Supplementary
290 Information, section A) that a GLMM with a binomial distribution and a probit link function

291 is exactly equivalent to such a model. Heritability can then be computed on this “liability”
292 scale (different from the expected data scale!) by adding a so-called “link variance” V_L to the
293 denominator (see, for example, Nakagawa and Schielzeth, 2010; de Villemereuil *et al.*, 2013):

$$h_{\text{thres}}^2 = \frac{V_{A,\ell}}{V_{A,\ell} + V_{\text{RE}} + V_{\text{O}} + V_L}. \quad (23)$$

294 Because the probit link function is the inverse of the cumulative standard normal distribution
295 function, the “link variance” V_L is equal to one in this case.

296 When the heritability is computed using the threshold model, Dempster and Lerner (1950)
297 and Robertson (1950) derived an exact analytical formula relating this estimate to the observed
298 data scale:

$$h_{\text{obs}}^2 = \frac{t^2}{p(1-p)} h_{\text{thres}}^2, \quad (24)$$

299 where p is the probability of occurrence of the minor phenotype and t is the density of a
300 standard normal distribution at the p th quantile (see also Roff, 1997). It can be shown (see
301 SI, section A) that this formula is an exact analytical solution to Eqs. 5 to 20 in the case of
302 a GLMM with binomial distribution and a probit link. When fixed effects are included in the
303 model, it is still possible to use these formulae by integration over the marginalised predictions
304 (see SI, section A). Note that this expression applies only to analyses conducted with a probit
305 link; it does not apply to a binomial model with a logit link function.

306 **Poisson distribution with a logarithm link**

307 For a log link function and a Poisson distribution, both the derivative of the inverse link function,
308 and the variance of the distribution function, are equal to the expected value. Consequently,
309 analytical results are obtainable for a log/Poisson model for quantities such as broad- and
310 narrow-sense heritabilities. Foulley and Im (1993) derived an analytical formula to compute
311 narrow-sense heritability on the observed scale:

$$h_{\text{obs}}^2 = \frac{\lambda^2 V_{A,\ell}}{\lambda^2 [\exp(V_{A,\ell} + V_{\text{RE}} + V_{\text{O}}) - 1] + \lambda} = \frac{\lambda V_{A,\ell}}{\lambda [\exp(V_{A,\ell} + V_{\text{RE}} + V_{\text{O}}) - 1] + 1}, \quad (25)$$

312 where λ is the data scale phenotypic mean, computed analytically as:

$$\lambda = \exp\left(\mu + \frac{V_{A,\ell} + V_{RE} + V_O}{2}\right). \quad (26)$$

313 Again, it can be shown (see SI, section B) that these formulae are exact solutions to Eq. 5 to
314 20 when assuming a Poisson distribution with a log link. The inclusion of fixed effects in the
315 model make the expression slightly more complicated (see SI, section B). These results can also
316 be extended to the Negative-Binomial distribution with log link with slight modifications of
317 the analytical expressions (see SI, section B).

318 The component of the broad-sense heritability on the observed data scale that arises from
319 additive genetic effects on the latent scale can be computed as an intra-class correlation coeffi-
320 cient (i.e. repeatability) for this kind of model (Foulley and Im, 1993; Nakagawa and Schielzeth,
321 2010):

$$H_{\text{obs}}^2 = \frac{V(E[z|a])}{V_{P,\text{obs}}} = \frac{\lambda(\exp(V_{A,\ell}) - 1)}{\lambda[\exp(V_{A,\ell} + V_{RE} + V_O) - 1] + 1}. \quad (27)$$

322 If non-additive genetic component were fitted in the model (e.g. dominance variance), they
323 should be added to $V_{A,\ell}$ in Eq. 27 to constitute the total genotypic variance, and thus obtain
324 the actual broad-sense heritability. Note that Eqs. 27 and 25 converge for small values of $V_{A,\ell}$.

325 **Example analysis: quantitative genetic parameters of a non-normal** 326 **character**

327 We modelled the first year survival of Soay sheep (*Ovis aries*) lambs on St Kilda, Outer He-
328 brides, Scotland. The data are comprised of 3814 individuals born between 1985 and 2011,
329 and that are known to either have died in their first year, defined operationally as having died
330 before the first of April in the year following their birth, or were known to have survived be-
331 yond their first year. Months of mortality for sheep of all ages are generally known from direct
332 observation, and day of mortality is typically known. Furthermore, every lamb included in this
333 analysis had a known sex and twin status (whether or not it had a twin), and a mother of a
334 known age.

335 Pedigree information is available for the St Kilda Soay sheep study population. Maternal

336 links are known from direct observation, with occasional inconsistencies corrected with genetic
337 data. Paternal links are known from molecular data. Most paternity assignments are made
338 with very high confidence, using a panel of 384 SNP markers, each with high minor allele
339 frequencies, and spread evenly throughout the genome. Details of marker data and pedigree
340 reconstruction are given in Bérénos *et al.* (2014). The pedigree information was pruned to
341 include only phenotyped individuals and their ancestors. The pedigree used in our analyses
342 thus included 4687 individuals with 4165 maternal links and 4054 paternal links.

343 We fitted a generalised linear mixed model of survival, with a logit link function and a
344 binomial distribution function. We modelled fixed effects sex and twin status, and linear,
345 quadratic, and cubic effects of maternal age ($matAge_i$). Maternal age was mean-centred by
346 subtracting the overall mean. We also included an interaction of sex and twin status, and an
347 interaction of twin status with maternal age. We included random effects of breeding value (as
348 for equation 2), maternal identity, and birth year. Because the overdispersion variance V_O in
349 a binomial GLMM is unobservable for binary data, we set its variance to one. The model was
350 fitted in MCMCGLMM (Hadfield, 2010), with diffuse independent normal priors on all fixed
351 effects, and parameter-expanded priors for the variances of all estimated random effects.

352 We identified important effects on individual survival probability, i.e., several fixed effects
353 were substantial, and also, each of the additive genetic, maternal, and among-year random
354 effects explained appreciable variances (Table 1). The model intercept corresponds to the
355 expected value on the latent scale of a female singleton (i.e. not a twin) lamb with an average
356 age (4.8 years) mother. Males have lower survival than females, and twins have lower survival
357 than singletons. There were also substantial effects of maternal age, corresponding to a rapid
358 increase in lamb survival with maternal age among relatively young mothers, and a negative
359 curvature, such that the maximum survival probabilities occur among offspring of mothers aged
360 6 or 7 years. The trajectory of maternal age effects in the cubic model are similar to those
361 obtained when maternal age is fitted as a multi-level effect.

362 To illustrate the consequences of accounting for different fixed effects in expected and ob-
363 served data scale inferences, we calculated several parameters under a series of different treat-
364 ments of the latent scale parameters of the GLMM. We calculated the phenotypic mean, the
365 additive genetic variance, the total variance of expected values, the total variance of observed

366 values, and the heritability of survival on the expected and observed scales.

367 First, we calculated parameters using only the model intercept (μ in Eq. 1 and 3a). In
368 general, linear modelling software will essentially arbitrarily define a model's intercept. In the
369 current case, due to the details of how the data were coded, the intercept is the latent scale
370 prediction for female singletons with average aged (4.8 years) mothers. In an average year,
371 singleton females with average aged mothers have a probability of survival of about 80%. The
372 additive genetic variance $V_{A,obs}$, calculated with Eq. 18 is about 0.005, and corresponds to
373 heritabilities on the expected and observed scales of 0.115 and 0.042 (Table 2).

374 In contrast, if we wanted to calculate parameters using a different (but potentially equally
375 arbitrary) intercept, corresponding to twin males, we would obtain a mean survival rate of 0.32,
376 an additive genetic variance that is twice as large, but similar heritabilities (Table 1). Note
377 that we have not modelled any systematic differences in genetic parameters between females
378 and males, or between singletons and twins. These differences in parameter estimates arise
379 from the exact same estimated variance components on the latent scale, as a result of different
380 fixed effects.

381 This first comparison has illustrated a major way in which the fixed effects in a GLMM
382 influence inferences on the expected and observed value scales. For linear mixed models, it
383 has been noted that variance in the response is explained by the fixed predictors, and that
384 this may inappropriately reduce the phenotypic variance and inflate heritability estimates for
385 some purposes (Wilson, 2008). However, in the example so far, we have simply considered two
386 different intercepts (i.e. no difference in explained variance): female singletons vs male twins,
387 in both cases, assuming focal groups of individuals are all born to average aged mothers. Again
388 these differences in phenotypic variances and heritabilities arise from differences in intercepts,
389 not from any differences in variance explained by fixed effects. All parameters on the expected
390 and observed value scales, including the mean, the additive genetic variance and the total
391 variance, are dependent on the intercept. Heritability is modestly affected by the intercept,
392 because additive genetic and total variances are similarly, but not identically, influenced by the
393 model intercept.

394 Additive genetic effects are those arising from the average effect of alleles on phenotype,
395 integrated over all background genetic and environmental circumstances in which alternate

396 alleles might occur. Fixed effects are part of this background. Following, for example, Eq. 6 and
397 17, we can integrate our calculation of Ψ and ultimately $V_{A,obs}$ over all fixed effects. Considering
398 all fixed and random effects, quantitative genetic parameters on the expected and observed
399 scales are given in table 2, 3rd column. The calculation of $V_{A,obs}$ now includes an average slope
400 calculated over a wide range of the steep part of the inverse-link function (near 0 on the latent
401 scale, and near 0.5 on the expected value scale), and so is relatively high. The observed total
402 phenotypic variance $V_{P,obs}$ is also quite high. The increase in $V_{P,obs}$ has two causes. First the
403 survival mean is closer to 0.5, so the random effects variance is now manifested as greater total
404 variance on the expected and observed scales. Second, there is now variance arising from fixed
405 effects that is included in the total variance.

406 **Evolutionary prediction**

407 Systems for predicting adaptive evolution in response to phenotypic selection assume that the
408 distribution of breeding values is multivariate normal, and in most applications, that the joint
409 distribution of phenotypes and breeding values is multivariate normal (Lande, 1979; Lande
410 and Arnold, 1983; Morrissey, 2014; Walsh and Lynch, forthcoming). Breeding values on the
411 expected and observed scales will not be normal in GLMMs. Breeding values are normal by
412 construction on the latent scale, and the non-linear (inverse) link functions cause the corre-
413 sponding distributions on the expected and observed scales to be non-normal. Consequently,
414 even with quantities such as additive genetic variances, heritabilities, etc., calculated on the
415 expected and observed data scales, evolutionary predictions using statistical genetic machinery
416 developed assuming normality will not hold. The Breeder's and Lande equations may hold
417 approximately, and may perhaps be useful. However, having taken up the non-trivial task
418 of pursuing GLMM-based quantitative genetic analysis, the investigator has at their disposal
419 inferences on the latent scale. On this scale, the assumptions required to predict the evolution
420 of quantitative traits hold. In this section we first demonstrate by simulation how application
421 of the Breeder's equation on the expected and observed scales represents an approximation.
422 We then proceed to an exposition of some statistical machinery that can be used to generate
423 predictions of evolution on the latent scale (from which evolutionary predictions on the ex-

424 pected and observed scale can subsequently be derived, using Eq. 5), given inference of the
425 function relating traits to fitness. Insofar as as the assumptions of a GLMM may represent a
426 useful model of a given non-normal trait, this latter approach to evolutionary prediction can
427 outperform application of the Breeder's equation on the data scale.

428 **Direct application of the Breeder's and Lande equations on the data scale**

429 In order to explore the predictions of the Breeder's equation applied at the level of observed
430 phenotype, we conducted a simulation in which phenotypes were generated according to a
431 Poisson GLMM (Eqs. 3a to 3c, with a Poisson distribution function and a log link function), and
432 then selected the largest observed count values (positive selection) with a range of proportions
433 of selected individuals (from 5% to 95%, creating a range of selection differentials), a range
434 of latent-scale heritabilities (0.1, 0.3, 0.5 and 0.8, with a latent phenotypic variance fixed to
435 0.1), and a range of latent means μ (from 0 to 3). We simulated 10,000 replicates of each
436 scenario, each composed of a different array of 10,000 individuals. For each simulation, we
437 simulated 10,000 offspring. For each offspring, a breeding value was simulated according to
438 $a_{\ell,i} \sim \mathcal{N}((a_{\ell,d} + a_{\ell,s})/2, V_{A,\ell}/2)$, where $a_{\ell,i}$ is the focal offspring's breeding value, $a_{\ell,d}$ and $a_{\ell,s}$ are
439 the breeding values of simulated dams and sires and $V_{A,\ell}/2$ represents the segregational variance
440 assuming parents are not inbred. Dams and sires were chosen at random with replacement
441 from among the pool of simulated selected individuals. For each scenario, we calculated the
442 realised selection differential arising from the simulated truncation selection, S_{obs} , and the
443 average evolutionary response across simulations, R_{obs} . For each scenario, we calculated the
444 heritability on the observed scale using Eq. 19. If the Breeder's equation was strictly valid for
445 a Poisson GLMM on the observed scale, the realised heritability $R_{\text{obs}}/S_{\text{obs}}$ would be equal to
446 the observed-scale heritability h_{obs}^2 .

447 The correspondence between $R_{\text{obs}}/S_{\text{obs}}$ and h_{obs}^2 is approximate (Fig. 2), and strongly de-
448 pends on the selection differential (controlled here by the proportion of selected individuals).
449 Note that, although the results presented here depict a situation where the ratio $R_{\text{obs}}/S_{\text{obs}}$ is
450 very often larger than h_{obs}^2 , this is not a general result (e.g. this is not the case when using
451 negative instead of positive selection, data not shown). In particular, evolutionary predictions
452 are poorest in absolute terms for large μ and large (latent) heritabilities. However, because

453 we were analysing simulation data, we could track the selection differential of latent value (by
454 calculating the difference in its mean between simulated survivors and the mean simulated be-
455 fore selection). We can also calculate the mean latent breeding value after selection. Across all
456 simulation scenarios, the ratio of the change in breeding value after selection, to the change in
457 breeding value before selection was equal to the latent heritability (see Fig. 2), showing that
458 evolutionary changes could be accurately predicted on the latent scale.

459 **Evolutionary change on the latent scale, and associated change on the expected** 460 **and observed scales**

461 In an analysis of real data, latent breeding values are, of course, not measured. However,
462 given an estimate of the effect of traits on fitness, say via regression analysis, we can derive
463 the parameters necessary to predict evolution on the latent scale. The idea is thus to relate
464 measured fitness on the observed data scale to the latent scale, compute the evolutionary
465 response on the latent scale and finally compute the evolutionary response on the observed
466 data scale.

467 To relate the measured fitness on the observed scale to the latent scale, we need to compute
468 the expected fitness W_{exp} given latent trait value ℓ , which is

$$W_{\text{exp}}(\ell) = \sum_k W_P(k)P(Z = k|\ell), \quad (28)$$

469 where $W_P(k)$ is the measure of fitness for the k th data scale category (assuming the observed
470 data scale is discrete). Population mean fitness can then be calculated in an analogous way to
471 equation 5:

$$\bar{W} = \int W_{\text{exp}}(\ell)f_{\mathcal{N}}(\ell, \mu, V_{A,\ell} + V_{\text{RE}} + V_{\text{O}})d\ell. \quad (29)$$

472 These expressions comprise the basic functions necessary to predict evolution. Given a fitted
473 GLMM, and a given estimate of the fitness function $W_P(k)$, each of several approaches could
474 give equivalent results. For simplicity, we proceed via application of the breeder's equation at
475 the level of the latent scale.

476 The change in the mean genetic value of any character due to selection is equal to the
477 covariance of breeding value with relative fitness (Robertson, 1966, 1968). Using Stein's (1973)

478 lemma once more, this covariance can be obtained as the product of the additive genetic variance
479 of latent values and the average derivative of expected fitness with respect to latent value, i.e.,
480 $E \left[\frac{dW_{\text{exp}}}{d\ell} \right]$. Evolution on the latent scale can therefore be predicted by

$$\Delta\mu = V_A E \left[\frac{dW_{\text{exp}}}{d\ell} \right] \frac{1}{\bar{W}}. \quad (30)$$

481 In the case of a multivariate analysis, note that the derivative above should be a vector of
482 partial derivatives (first order partial derivative for each trait).

483 If fixed effects need to be considered, the approach can be modified in the same way as
484 integration over fixed effects is accomplished for calculating other quantities, i.e. the expression

$$\bar{W} = \frac{1}{N} \sum_{i=1}^N \int W_{\text{exp}}(\ell) f_{\mathcal{N}}(\ell, \mu + \hat{\ell}_i, V_{A,\ell} + V_{\text{RE}} + V_{\text{O}}) d\ell \quad (31)$$

485 would be used in calculations of mean fitness and the average derivative of expected fitness
486 with respect to latent value.

487 Phenotypic change caused by changes in allele frequencies in response to selection is calcu-
488 lated as

$$\Delta\bar{z} = \int g^{-1}(\ell) f_{\mathcal{N}}(\ell, \mu + \Delta\mu, V_{A,\ell} + V_{\text{RE}} + V_{\text{O}}) d\ell - \bar{z}. \quad (32)$$

489 Or, if fixed effects are included in the model:

$$\Delta\bar{z} = \frac{1}{N} \sum_{i=1}^N \int g^{-1}(\ell) f_{\mathcal{N}}(\ell, \mu + \hat{\ell}_i + \Delta\mu, V_{A,\ell} + V_{\text{RE}} + V_{\text{O}}) d\ell - \bar{z}. \quad (33)$$

490 Note that, in this second equation, \bar{z} must be computed as in Eq. 6 and that this equation
491 assumes that the distribution of fixed effects for the offspring generation is the same as for the
492 parental one.

493 Another derivation of the expected evolutionary response using the Price-Robertson identity
494 (Robertson, 1966; Price, 1970) is given in the Supplementary Information (section C).

495 **The simulation study revisited**

496 Using the same replicates as in the simulation study above (Fig. 2, top row), we used Eqs. 28
497 to 33 to predict phenotypic evolution. This procedure provides greatly improved predictions
498 of evolutionary change on the observed scale. However, somewhat less response to selection is
499 observed than is predicted. This behaviour occurs because, in addition to producing a perma-
500 nent evolutionary response in the mean value on the latent scale, directional selection creates
501 a transient reduction of additive genetic variance due to linkage disequilibrium. Because the
502 link function is non-linear, this transient change in the variance on the latent scale generates
503 a transient change in the mean on the expected and observed scales. Following several genera-
504 tions of random mating, the evolutionary change on the observed scale would converge on the
505 predicted values. We simulated such a generation at equilibrium by simply drawing breeding
506 values for the post-selection sample from a distribution with the same variance as in the parental
507 generation. This procedure necessarily generated a strong match between predicted and simu-
508 lated evolution (Fig. 2, bottom row). Additionally, the effects of transient reduction in genetic
509 variance on the latent scale could be directly modelled, for example, using Bulmer's (1971)
510 approximations for the transient dynamics of the genetic variance in response to selection.

511 **Discussion**

512 The expressions given here for quantitative genetic parameters on the expected and observed
513 data scales are exact, given the GLMM model assumptions, in two senses. First, they are not
514 approximations, such as might be obtained by linear approximations (Browne *et al.*, 2005).
515 Second, they are expressions for the parameters of direct interest, rather than convenient sub-
516 stitutes. For example, the common calculation of variance partition coefficients (intraclass
517 correlations) on an underlying scale with a logistic distribution, (as also suggested by Browne
518 *et al.* 2005) provides a value of the broad-sense heritability (e.g. using the genotypic variance
519 arising from additive genetic effects on the latent scale) when applied to genetic parameters
520 estimated in a logistic GLMM. The expressions given here can provide quantitative genetic
521 inferences of the additive genetic parameters, and on the scale on which the traits are observed.
522 The whole framework developed here (including univariate and multivariate parameters com-

523 putation and evolutionary predictions on the observed data scale) is implemented in the R
524 package QGGLMM available at <https://github.com/devillemereuil/qgglm>.

525 While the calculations we provide will often (i.e. when no analytical formulae exist) be
526 more computationally demanding than calculations on the latent scale, they will be direct
527 ascertainment of specific parameters of interest, since the scale of evolutionary interest is likely
528 to be the observed data scale, rather than the latent scale (unless some artificial selection is
529 applied to predicted latent breeding values as in modern animal breeding). Most applications
530 should not be onerous. Computations of means and (additive genetic) variances took less
531 than a second on a 1.7 GHz processor when using our R functions on the Soay sheep data
532 set. Summation over fixed effects, and integration over 1000 posterior samples of the fitted
533 model took several minutes. When analytical expressions are available (e.g. for Poisson/log,
534 Binomial/probit and Negative-Binomial/log; see the supplementary information and R package
535 documentation), these computations are considerably accelerated.

536 We have highlighted additional and important ways in which fixed effects influence quan-
537 titative genetic inferences with GLMMs, and developed an approach for handling these com-
538 plexities. In LMMs, the main consideration pertaining to fixed effects is that they explain
539 variance, and some or all of this variance might be inappropriate to exclude from an assessment
540 of V_P when calculating heritabilities (Wilson, 2008). This aspect of fixed effects is relevant
541 to GLMMs, but furthermore, all parameters on the expected and observed scales, not just
542 means, are influenced by fixed effects in GLMMs; this includes additive genetic and phenotypic
543 variances. This fact necessitates particular care in interpreting GLMMs.

544 In our example analysis in Soay sheep, $V_{A,\ell}$ and V_P changed substantially depending on
545 different treatments of the fixed effects (especially, arbitrary different definitions of the model's
546 latent intercept). We do not intend to suggest that any of these treatments of the fixed effects
547 is correct or wrong. Rather, any of the analyses we presented (and many other conceivable
548 variations) may be appropriate to any particular purpose. For this particular case, different
549 treatments of fixed effects changed $V_{A,\ell}$ and V_P in roughly, but not exactly, similar proportions.
550 Consequently, heritabilities on the data scales were not greatly different among treatments. This
551 need not necessarily be the case in all quantitative genetic analyses using GLMMs, although it
552 is likely to be common in binomial response models.

553 Consequently, users of GLMM-based quantitative genetic analyses should take great care in
554 defining intercepts. When a clear biological motivation for the definition of a model intercept is
555 not available, any data-scale GLMM-based inferences of quantitative genetic parameters should
556 be assessed for sensitivity to arbitrary choices about model intercepts. For many situations in
557 which the definition of the model intercept seems arbitrary, integrating over biologically-relevant
558 distributions of fixed effects (e.g., as in equations 6, 17, etc.) will probably be the best solution.
559 In some cases, there will be multiple meaningful values of parameters such as V_A on the data
560 scale, associated with a single value on the latent scale. For example, if the sexes have different
561 intercepts, but are modelled as having a common value of V_A on the latent scale, then there
562 are different sex-specific data scale values of V_A on the data scale, resulting from the different
563 intercepts.

564 Currently, with the increasing applicability of GLMMs, investigators seem eager to convert
565 to the observed data scale. It seems clear that conversions between scales are generally useful.
566 However, it is of note that the underlying assumption when using GLMMs for evolutionary
567 prediction is that predictions hold on the latent scale. Therefore, given an estimate of a fitness
568 function, no further assumptions are necessary to predict evolution via the latent scale (as
569 with equations 28, 30, and 32), over and above those that are made in the first place upon
570 deciding to pursue GLMM-based quantitative genetic analysis. The approach we suggest treats
571 the relationships between the levels of a GLMM as a simple developmental system, and the
572 approach described here is essentially the general theory laid out in Morrissey (2015), with
573 specific extensions to handle distribution functions and integration over fixed effects.

574 **Acknowledgements**

575 PdV was supported by a doctoral studentship from the French *Ministère de la Recherche et de*
576 *l'Enseignement Supérieur*. HS was supported by an Emmy Noether fellowship from the German
577 Research Foundation (DFG; SCHI 1188/1-1/2). SN is supported by a Future Fellowship, Aus-
578 tralia (FT130100268). MBM is supported by a University Research Fellowship from the Royal
579 Society (London). The Soay sheep data were provided by Josephine Pemberton and Loeske
580 Kruuk, and were collected primarily by Jill Pilkington and Andrew MacColl with the help of

581 many volunteers. The collection of the Soay sheep data is supported by the National Trust
582 for Scotland and QinetQ, with funding from NERC, the Royal Society, and the Leverhulme
583 Trust. We thank Kerry Johnson, Paul Johnson, Alastair Wilson, Loeske Kruuk and Josephine
584 Pemberton for valuable discussions and comments on this manuscript.

585 **References**

- 586 Ayers, D. R., R. J. Pereira, A. A. Boligon, F. F. Silva, F. S. Schenkel, V. M. Roso, and L. G.
587 Albuquerque, 2013 Linear and Poisson models for genetic evaluation of tick resistance in
588 cross-bred Hereford x Nellor cattle. *Journal of Animal Breeding and Genetics* 130: 417–424.
- 589 Béréños, C., P. A. Ellis, J. G. Pilkington, and J. M. Pemberton, 2014 Estimating quantitative
590 genetic parameters in wild populations: a comparison of pedigree and genomic approaches.
591 *Molecular Ecology* 23: 3434–3451.
- 592 Browne, W. J., S. V. Subramanian, K. Jones, and H. Goldstein, 2005 Variance partitioning
593 in multilevel logistic models that exhibit overdispersion. *Journal of the Royal Statistical*
594 *Society* 168: 599–613.
- 595 Bulmer, M. G., 1971 The effect of selection on genetic variability. *The American Natural-*
596 *ist* 105: 201–211.
- 597 de Villemereuil, P., O. Gimenez, and B. Doligez, 2013 Comparing parent–offspring regression
598 with frequentist and Bayesian animal models to estimate heritability in wild populations: a
599 simulation study for Gaussian and binary traits. *Methods in Ecology and Evolution* 4(3):
600 260–275.
- 601 Dempster, E. R. and I. M. Lerner, 1950 Heritability of Threshold Characters. *Genetics* 35(2):
602 212–236.
- 603 Falconer, D. S., 1960 *Introduction to Quantitative Genetics*. Oliver and Boyd.
- 604 Fisher, R. A., 1918 The correlation between relatives on the supposition of Mendelian inheri-
605 tance. *Transactions of the Royal Society of Edinburgh* 52: 399–433.
- 606 Fisher, R. A., 1924 The biometrical study of heredity. *Eugenics Review* 16: 189–210.

- 607 Fisher, R. A., 1930 *The Genetical Theory of Natural Selection*. Oxford: Clarendon Press.
- 608 Foulley, J. L. and S. Im, 1993 A marginal quasi-likelihood approach to the analysis of Poisson
609 variables with generalized linear mixed models. *Genetics Selection Evolution* 25(1): 101.
- 610 Hadfield, J., 2010 MCMC Methods for Multi-Response Generalized Linear Mixed Models:
611 The MCMCglmm R Package. *Journal of Statistical Software* 33(2): 1–22.
- 612 Henderson, C. R., 1950 Estimation of genetic parameters. *Annals of Mathematical Statis-*
613 *tics* 21: 309–310.
- 614 Henderson, C. R., 1973 *Proceedings of the Animal Breeding and Genetics Symposium in*
615 *Honour of Dr. Jay L. Lush*, Chapter Sire evaluation and genetic trends. Published by the
616 American Society of Animal Science and the American Dairy Science Association.
- 617 Houle, D., 1992 Comparing Evolvability and Variability of Quantitative Traits. *Genet-*
618 *ics* 130(1): 195–204.
- 619 Kruuk, L. E. B., 2004 Estimating genetic parameters in natural populations using the ‘animal
620 model’. *Philosophical Transactions of the Royal Society of London B* 359: 873–890.
- 621 Lande, R., 1979 Quantitative genetic analysis of multivariate evolution, applied to brain:body
622 size allometry. *Evolution* 33: 402–416.
- 623 Lande, R. and S. J. Arnold, 1983 The measurement of selection on correlated characters.
624 *Evolution* 37: 1210–1226.
- 625 Lush, J. L., 1937 *Animal breeding plans*. Ames, Iowa: Iowa State College Press.
- 626 Lynch, M. and B. Walsh, 1998 *Genetics and analysis of quantitative traits*. Sunderland, MA:
627 Sinauer.
- 628 Milot, E., F. M. Mayer, D. H. Nussey, M. Boisvert, F. Pelletier, and D. Reale, 2011 Evidence for
629 evolution in response to natural selection in a contemporary human population. *Proceedings*
630 *of the National Academy of Sciences* 108(41): 17040–17045.
- 631 Morrissey, M. B., 2014 In search of the best methods for multivariate selection analysis.
632 *Methods in Ecology and Evolution* 5: 1095–1109.

- 633 Morrissey, M. B., 2015 Evolutionary quantitative genetics of non-linear developmental systems.
634 Evolution 69: 2050–2066.
- 635 Morrissey, M. B., C. A. Walling, A. J. Wilson, J. M. Pemberton, T. H. Clutton-Brock, and
636 L. E. B. Kruuk, 2012 Genetic Analysis of Life-History Constraint and Evolution in a Wild
637 Ungulate Population. The American Naturalist 179(4): E97–E114.
- 638 Nakagawa, S. and H. Schielzeth, 2010 Repeatability for Gaussian and non Gaussian data: a
639 practical guide for biologists. Biological Reviews 85(4): 935–956.
- 640 Price, G. R., 1970 Selection and covariance. Nature 227: 520–521.
- 641 Robertson, A., 1950 Heritability of Threshold Characters. Genetics 35(2): 212–236.
- 642 Robertson, A., 1966 A mathematical model of the culling process in dairy cattle. Animal
643 Production 8: 95–108.
- 644 Robertson, A., 1968 *Population Biology and Evolution*, Chapter The spectrum of genetic
645 variation, pp. 5–16. New York: Syracuse University Press.
- 646 Roff, D. A., 1997 *Evolutionary quantitative genetics*. New York, New York (US): Chapman
647 & Hall.
- 648 Stein, C. M., 1973 Estimation of the mean of a multivariate normal distribution. Proceedings
649 of the Prague Symposium on Asymptotic Statistics 1: 345–381.
- 650 Walsh, B. and M. Lynch, forthcoming *Evolution and selection of quantitative traits*. Sinauer.
- 651 Wilson, A. J., 2008 Why h^2 does not always equal V_A/V_P ? Journal of Evolutionary Biol-
652 ogy 21(3): 647–650.
- 653 Wilson, A. J., M. B. Morrissey, M. J. Adams, C. A. Walling, F. E. Guinness, J. M. Pemberton,
654 T. H. Clutton-Brock, and L. E. B. Kruuk, 2011 Indirect genetics effects and evolutionary
655 constraint: an analysis of social dominance in red deer, *Cervus elaphus*. Journal of Evolu-
656 tionary Biology 24(4): 772–783.

- 657 Wilson, A. J., D. Reale, M. N. Clements, M. B. Morrissey, C. A. W. E. Postma, L. E. B.
658 Kruuk, and D. H. Nussey, 2010 An ecologist's guide to the animal model. *Journal of Animal*
659 *Ecology* 79: 13–26.
- 660 Wright, S., 1934 An Analysis of Variability in Number of Digits in an Inbred Strain of Guinea
661 Pigs. *Genetics* 19(6): 506–536.

Table 1: Parameters from the GLMM-based quantitative genetic analysis of Soay sheep (*Ovis aries*) lamb first-year survival. All estimates are reported as posterior modes with 95% credible intervals. The intercept in this model is arbitrarily defined for female lambs without twins, born to average age (4.8 years) mothers.

Parameter	Posterior mode with 95% CI
(a) Fixed effects	
Intercept	2.555 (1.755 – 3.514)
Sex (male vs. female)	-1.141 (-1.441 – -0.943)
Twin (twin vs. singleton)	-2.434 (-3.377 – -1.755)
Maternal age, linear term	0.228 (0.089 – 0.390)
Maternal age, quadratic term	-0.169 (-0.194 – -0.148)
Maternal age, cubic term	0.015 (0.011 – 0.02)
Sex-twin interaction	0.652 (0.015 – 1.068)
Sex-maternal age interaction	-0.027 (-0.115 – 0.054)
(b) Random effects	
$V_{A,\ell}$	0.831 (0.275 – 1.664)
V_{mother}	0.408 (0.177 – 0.887)
V_{year}	3.025 (1.452 – 5.551)

Table 2: Estimates of expected and observed scale phenotypic mean and variances, and additive genetic variance, for three different treatments of the fixed effects, as modelled on the linear scale with a GLMM, and reported in table 1.

Quantity	Arbitrary intercept (singleton female)	Arbitrary intercept (twin male)	Averaging over all fixed effects
\bar{z}	0.788 (0.718 – 0.886)	0.371 (0.212 – 0.471)	0.430 (0.336 – 0.517)
$V_{A, \text{data}}$	0.006 (0.002 – 0.015)	0.011 (0.005 – 0.024)	0.014 (0.005 – 0.021)
$V_{P, \text{exp}}$	0.062 (0.033 – 0.096)	0.104 (0.069 – 0.123)	0.120 (0.106 – 0.138)
$V_{P, \text{obs}}$	0.167 (0.107 – 0.206)	0.241 (0.183 – 0.250)	0.250 (0.226 – 0.250)
h_{exp}^2	0.096 (0.036 – 0.202)	0.125 (0.045 – 0.227)	0.112 (0.036 – 0.170)
h_{obs}^2	0.051 (0.015 – 0.085)	0.048 (0.023 – 0.106)	0.047 (0.019 – 0.089)

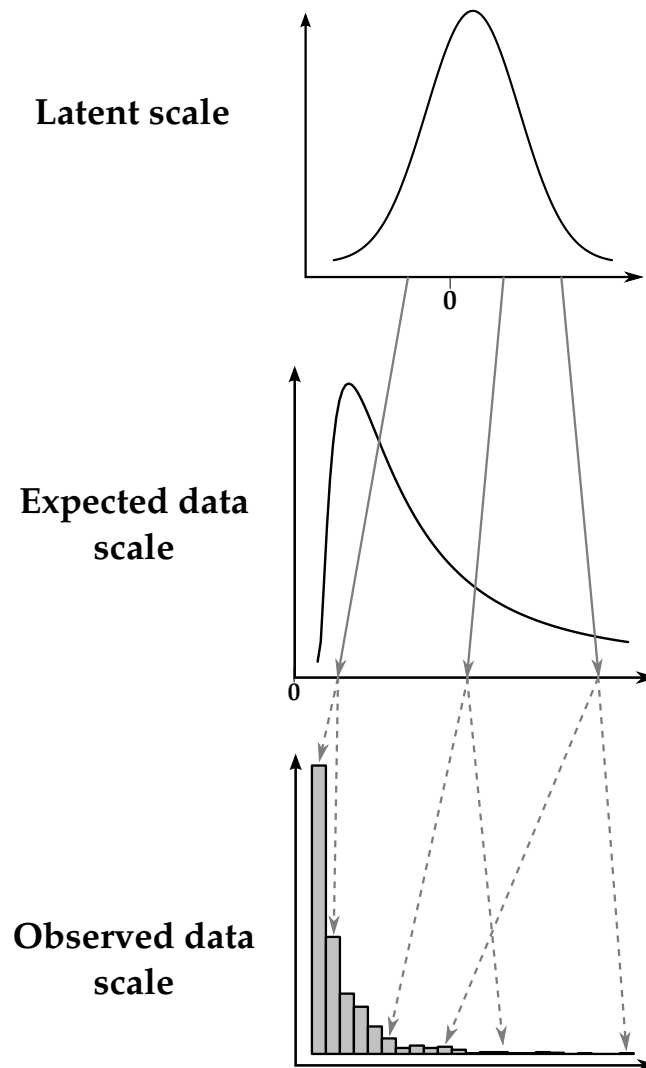


Figure 1: Example of the relationships between the three scales of the GLMM using a Poisson distribution and a logarithm link function. Deterministic relationships are denoted using grey plain arrows, whereas stochastic relationships are denoted using grey dashed arrows. Note that the latent scale is depicted as a simple Gaussian distribution for the sake of simplicity, whereas it is a mixture of Gaussian distributions in reality.

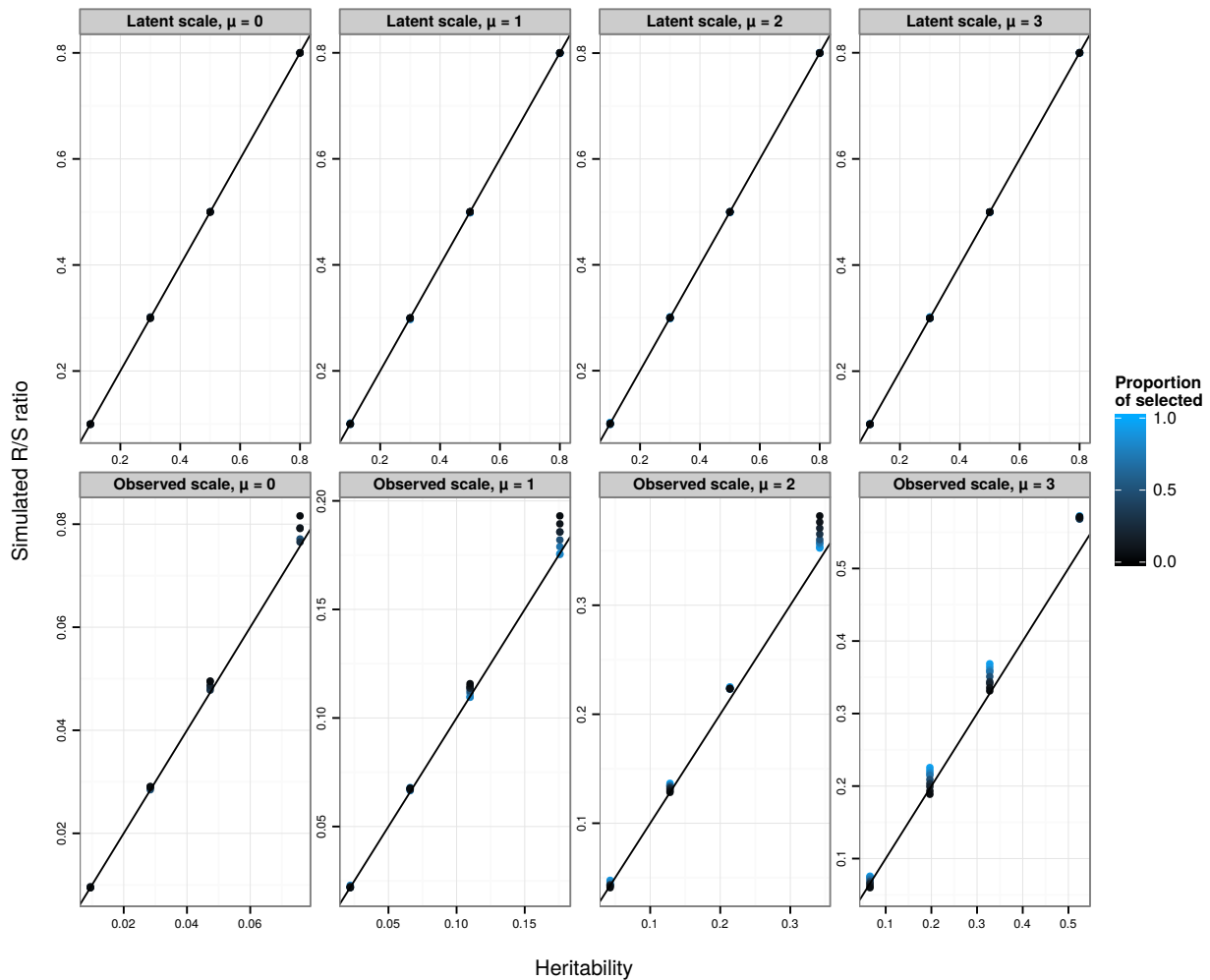


Figure 2: Simulated R/S (evolutionary response over selection differential, or the realised heritability) on the latent (upper panels) or observed date (lower panels) scales against the corresponding-scale heritabilities. Each data point is the average over 10,000 replicates of 10,000 individuals for various latent heritabilities h_{lat}^2 (0.1, 0.3, 0.5, 0.8), latent population mean (μ from 0 to 3, from left to right) and proportion of selected individuals (5%, 10%, 20%, 30%, 50%, 70%, 80%, 90%, 95%, varying from black to blue). The 1:1 line is plotted in black. The breeder's equation is predictive on the latent scale (upper panels), but approximate on the observed data scale (lower panels), because phenotypes and breeding values are not jointly multivariate normal on that scale.

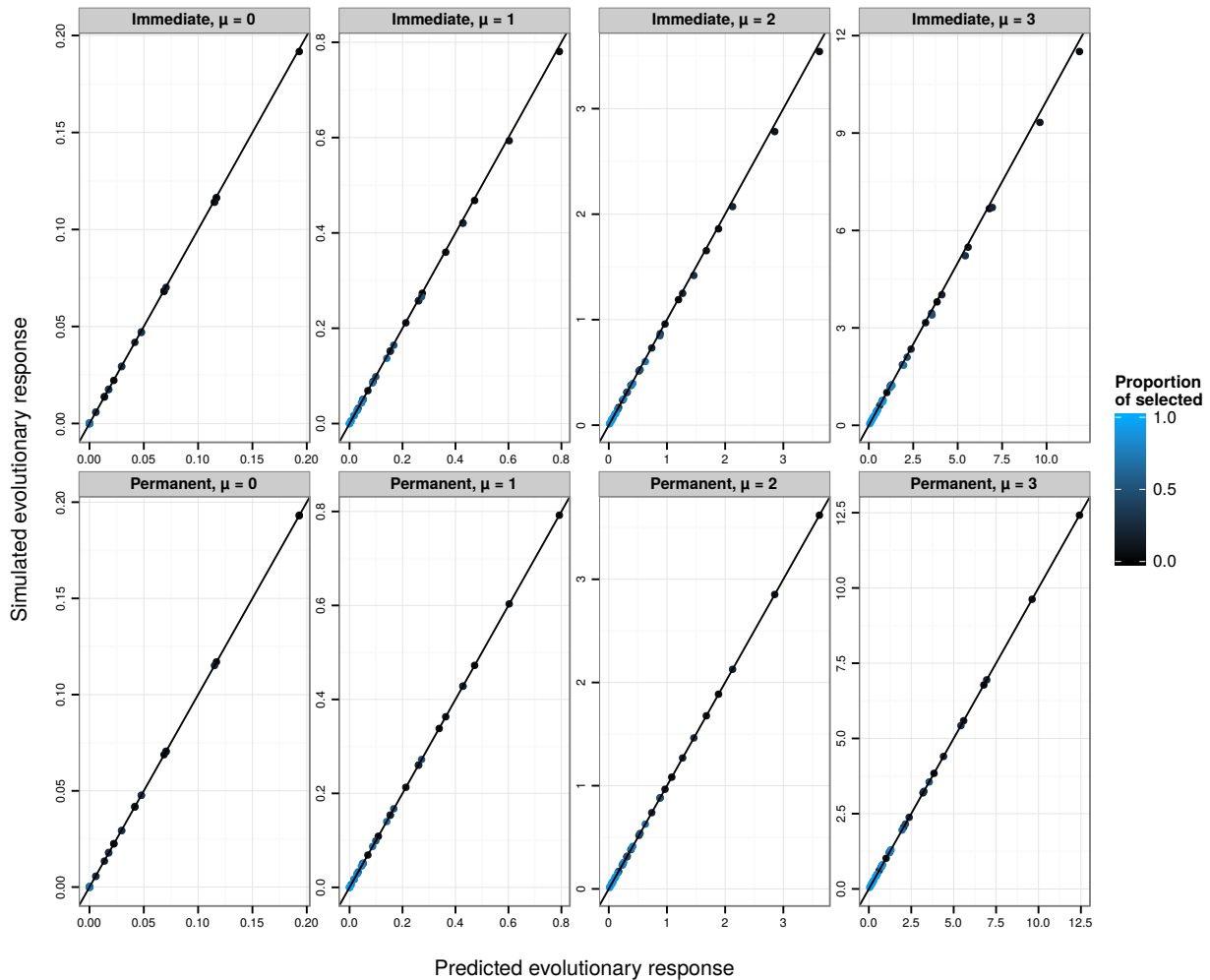


Figure 3: Predicted R_{obs} (phenotypic evolutionary response on the observed scale, see Eq. 33) against the simulated R_{obs} , via evolutionary predictions applied on the latent scale. Each data point is the average over 10,000 replicates of 10,000 individuals for various latent heritabilities h_{lat}^2 (0.1, 0.3, 0.5, 0.8), latent population mean (μ from 0 to 3) and proportion of selected individuals (5%, 10%, 20%, 30%, 50%, 70%, 80%, 90%, 95%, varying from black to blue). The 1:1 line is plotted in black. The upper panels (“Immediate”) show simulations for the response after a single generation, which include both a permanent and transient response to selection arising from linkage disequilibrium. The bottom panels (“permanent”) show simulation results modified to reflect only the permanent response to selection.