COGNITIVE DOMAINS FUNCTION COMPLEMENTATION BY *NTNG* GENE PARALOGS

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ABSTRACT

Gene duplication was proposed by S.Ohno (Ohno, 1970) as a key mechanism of a gene function evolution. A pair of gene paralogs, *NTNG1* and *NTNG2*, sharing identical gene and protein structures and encoding similar proteins, forms a functional complement subfunctionalising (SF) within cognitive domains and forming cognitive endophenotypes, as detected by Intellectual Quotient (IQ) tests (Prosselkov et al., in press). *NTNG* paralogs are associated with autism spectrum disorder (ASD), bipolar disorder (BD) and schizophrenia (SCZ), with unique non-overlapping segregation among the other 15 cognitive disorders (CD), emphasizing an evolutionary gain-dependent link between advanced cognitive functions and concomitant cognitive pathologies. Complementary expression and human brain transcriptome composition of the paralogs explains the observed phenomena of their functional complementarity. The lowest identity among *NTNGs* is found in a middle of encoded by them proteins designated as uknown (Ukd) domain. *NTNG1* contains anthropoid-specific constrained regions and both genes contain non-coding conserved sequences underwent accelerated evolution in human. *NTNG* paralogs SF perturbates "structure drives function" concept at protein and gene levels. Their function diversification results in a so-called "Cognitive Complement (CC)" formation, a product of gene duplication and subsequent cognitive subfunction bifurcation among the *NTNG* gene duplicates.

INTRODUCTION

Gene duplication was proposed by S.Ohno (**Ohno, 1970**) as a key mechanism of a gene function evolution. Complex behaviors arise from a combination of simpler genetic modules that either have evolved separately or co-evolved. Many genes and the proteins they encode have been found to be involved in the cognitive information processing with a single variant or a single gene generally accounting for only a partial phenotypic variation in a complex trait. Cognitive processing as a quintessence of the brain functioning can be viewed as a product of intricately interlinked networks generated by deeply embedded into it players with specific or partially overlapping functions. The robustness of the cognitive processing towards its single elements genetic eliminations (to study their function) and its simultaneous fragility expressed in the multiple forms of neurological disorders manifest the existence of cognitive domains interlocked but SF within a unit of cognition formed upon these domains interaction. Previously, we have described a function of a pair of gene paralogs (*NTNG1* and *NTNG2*) involved in human IQ tests performance and underwent hominin-specific evolutionary changes (**Prosselkov et al., in press**). Hereby, we continue looking at these genes paralogs features focusing on underlying mechanisms of their function segregation and complementation within the cognitive domains.

RESULTS

The observed phenomena of functional complementation among the *NTNG* paralogs within cognitive domains (**Prosselkov et al., in press**) is also manifested in *NTNG*-associated human pathologies diagnosed in most cases (if only not in all) by a cognitive decline (**Figure 1A-1** and **A-2**). Both genes are associated with BD and SCZ – devastating disorders sharing similar etiology (**Lee et al., 2013**) with genetic correlation by multivariate analysis of 0.590 (**Maier et al., 2015**), linked to human creativity (**Power et al., 2015**), and characterized by impulsiveness as a common diagnostic feature (**Reddy et al., 2014**). Recently found associations of both paralogs with ASD (**Sanders et al., 2015**) supports the reported genetic correlation of 0.194 ASD/SCZ pair (**Maier et al., 2015**) and shared module eigengenes detected by PC1 among these two disorders (**Parikshak et al., 2015**). 12 *NTNG1*-linked CDs, ranging from AD to TS, span a broad spectrum of clinical features frequently involving reduced processing speed (PS) and verbal comprehension (VC, **Figure 1A-1**). As for *NTNG2*, working memory (WM) deficit and inability "to bind" events (perceptual organization, PO) are the most prominent diagnostic traits for the SLE and TLE patients (**Figure 1A-2**), with PN characterised by indolent behavior in 90% of the cases (**Cavard et al., 2009**). Thus, both *NTNG* paralogs are associated with a variety of CDs and mostly in a non-overlapping manner, except for ASD, BD and SCZ characterized by shared and wide spectrum of cognitive abnormalities. The clinical etiology of the aforementioned diseases supports the IQ-deduced functional complementation among the *NTNG* paralogs (**Prosselkov et al., in press**) with (VC/PS) and (WM/PO) deficits being also uniquely segregated among the associated cognitive pathologies.

Since both genes are expected to have identical gene exon/intron compositions but different in their intron lengths (**Yin et al., 2002**) we have reconstructed the paralogs transcriptomes by re-processing the publicly available RNA-seq dataset (**Wu et al., 2012**)

from healthy and SCZ human subjects superior temporal gyrus (STG) post-mortem brain tissue (Supplementary Table 1a=**ST1a**). A difference is noted instantly at the total expression levels (genes, exons, individual RNA transcripts) when two gene paralogs are compared (**Figure 1B-1** and **B-2**). *NTNG2* amount (as a whole gene) is 5 times larger comparing to *NTNG1*; exons (2-5) are 3 times, exons (8-9) are 18 times and exon 10 is 4 times higher expressed for *NTNG2* than for *NTNG1*. The only two exons outlaying the prevailing amount rule for the *NTNG2* mRNAs are exons 6 and 7, expressed nearly at the same absolute level as for the *NTNG1* exon paralogs, making them highly underrepresented within the whole *NTNG2* transcriptome. Next, distinct non-alternating splicing modules are formed by exons (2-5) for *NTNG1* (**Figure 1B-1**), while exons (4-5) and exons (8-9) for *NTNG2* (**Figure 1B-2**). Two structurally identical RNA transcript paralogs (*NTNG1a* = G1a and *NTNG2a* = G2a) have been found to exist in both *NTNG* transcriptomes with G2a being expressed at 8-9 times higher level than G1a. *NTNG1* is uniformly presented across the all analysed 16 human samples by 2 more protein coding RNAs (G1c and G1d, detected previously in mice brain, **Nakashiba et al., 2000**) and by 2 non-coding intron (9-10) derived transcripts (**Figure 1B-1**). At the same time, *NTNG2* transcriptome is comprised of one extra potentially coding RNA (G2a-like with exon 2 spliced out but in-frame coding preserved) and 2 assumed to be non-coding RNAs with exons 6 and 7 retained along with preceding and following them introns. Quite interesting that these two latter transcripts are the only RNA species with *NTNG2* exon 6 and 7 retained (**Figure 1B-2**). Two more coding (G1f and G1n) and 4 more non-coding for *NTNG1* and 9 extra non-coding for *NTNG2* RNA species have been also assembled from the available reads but due to inconsistency in their appearance across all 16 STG samples they are not presented on the figure but summarized in the table (**Figure 1C**, for details refer to **ST1d**). Summarising above said, it can be concluded that quantitative and qualitative complementary differences is a prominent feature characterising the brain RNA

105 transcriptome of human *NTNG* paralogs. However, no significant changes at the
106 transcription level of neither whole genes, nor individual exons, nor reconstructed RNA
107 transcripts have been found for SCZ and healthy subjects.

108 Upon calling the presence of IQ-affecting SNPs (**Prosselkov et al., in press**) across
109 all STG samples (**ST1c**) it has been revealed that 15 out of 16 subjects were positive for the
110 T-allele of rs2149171 (exon 4-nested), shown above to attenuate the WM score in SCZ
111 patients, making a comparison among the allele carrier *vs* non-carrier impossible. Four
112 healthy and three SCZ samples carry a T-allele of rs3824574 (exon 3-nested, non-affecting
113 IQ), and 1 healthy and 1 SCZ sample each contains a C-allele of rs4915045 (exon10, non-
114 coding part-nested, and non-affecting IQ). Thus, among the eleven cognitive endophenotype-
115 associated SNPs only 3 were possible to call out of the available *NTNG* transcriptome.

116 Distinctly complementary nature of the *NTNG* paralogs segregation within
117 neurological disorders and RNA transcriptome usage in STG (**Figure 1**) has prompted us to
118 analyse both genes expression across the entire human brain. We have reconstructed both
119 genes expression profiles in the human brain areas over the life span from conception (pcw =
120 post-conception week) to mature age (30-40 yrs old) using the RNA-seq data from BrainSpan
121 (www.brainspan.org). Similarities and differences are easily noted when the age-dependent
122 phases of *NTNG1* and *NTNG2* expression profiles are matched (**Figure 2**). Based on the
123 visual inputs three distinct classifiers have been elaborated: 1. predominantly synchronous
124 (**Figure 2A(1-4))**, characteristic mostly for the cortical areas; 2. predominantly mixed and
125 asynchronous (**Figure 2B**), characteristic for the cerebellar cortex and subcortical formations;
126 and 3. anti-phasic (complementary, **Figure 2C**), characteristic for the MD of thalamus and
127 hippocampus. All analysed brain areas demonstrated an elevated level of *NTNG2* expression
128 in comparison to *NTNG1* except for thalamus (**Figure 2C**) with the largest difference
129 observed is at the time of birth (35-37 pcw) or soon after (4 mo) for the synchronous

130 classifiers (**Figure 2A**), oscillating increment values across the life span for the mixed
131 (**Figure 2B**) and anti-phasic (**Figure 2C**) classifiers. It is quite intriguing to note that
132 essentially all brain areas show a trend towards the expression difference being negated
133 between the paralogs by reaching the mature age of 30-40 yrs old (nearly or above the mean
134 age used for the IQ testing), except MD where the expression discrepancy is increased. Thus,
135 the observed functional complementation among the *NTNG* paralogs is supported by the
136 anatomical distribution of the genes in human brain and their expression pattern modality
137 over the human subjects lifetime.

138 A direct comparison of the *NTNG* paralogs shows not only identical intron-exon gene
139 structure (**Figure 1B-1, 2B-2**) but also closely matched exon sizes (**Figure 3A**). There are
140 three exons of identical sizes (exons 4, 8 and 9), another three exons differed by one encoded
141 aa (exons 3, 5 and 6) and there are exons of different sizes (exons 2, 7 and 10). In terms of
142 size the largest difference among the genes is visually presented by the introns: intron (9-10)
143 of *NTNG1* is 52.7 times larger its *NTNG2* paralogous intron with intron (6-7) of *NTNG1*
144 being only 1.43-times larger pointing towards non-equilibria process of non-coding elements
145 elaborations as the process of gene paralogs SF proceeded. Nevertheless, it can be
146 generalised that in average all *NTNG1* introns are several times larger their *NTNG2* analogs
147 (**Figure 3A**). We have shown previously that exons 6 and 7 are differentially used within the
148 brain *NTNG* transcriptome (**Figure 1B-1** and **B-2**) and to explore their potential contribution
149 into the paralogs SF we have built identity matrices with these exons excluded and included
150 (but still producing in-frame existing transcripts, **Figure 3B-1** left and right panels,
151 respectively). Exclusion of both exons from the full-lengths transcripts (thus converting
152 *NTNG1m* to *NTNG1a* and *NTNG2b* to *NTNG2a*, respectively) increases the identity of DNA
153 on 2% (a relatively large effect since both exons together represent only 7.22 and 9.69% of
154 the total coding part of the full-length RNA transcripts, *NTNG1m* and *NTNG2b*, respectively).

155 This effect becomes even stronger when the encoded by these transcripts proteins are also
156 compared (**Figure 3B-2**). The spliced out Ukd protein domains (encoded by the exons 6 and
157 7) increases the proteins identity on 3.8% thus making the middle of both genes (and encoded
158 proteins) substantially more different among the both gene paralogs. To corroborate this
159 observation and to explore the importance of other protein parts we have directly compared
160 the sequences encoded by the full-length transcripts and producing Netrin-G1m and Netrin-
161 G2b (**Figure 3C**). Similarly to what has been shown on **Figure 3B-1** and **3B-2**, the lowest
162 identity (17.5%) is represented by the Ukd domain (encoded by the exons 6 and 7) and by the
163 preceding it exon 5 (a 3'-part of the LE1 domain). Two other areas also show a substantially
164 low identity, namely the N-terminus (it includes the protein secretory signal indicated by an
165 arrow) and the outmost C-terminus responsible for the unique feature of Netrin-Gs – the GPI
166 attachment. Thus, based on the percent identity comparisons among the Netrin-G paralogs it
167 can be predicted that there are several potential protein parts contributing to the paralogs SF.
168 As it has been reported by **Seiradake et al. (2011)**, identical gene and protein domain
169 compositions result in the identical structural motif with differences only in the spatial
170 arrangement of the loops facing the post-synaptic Netrin-G's interacting partners, NGL-1 and
171 NGL-2, respectively (**Figure 3D**). Loop I binding surfaces alignment (**Figure 3C**, blue color)
172 shows a high level of conservation (with at least 5 amino acids 100% conserved) among the
173 Netrin-G paralogs, indicating that it is unlikely to be responsible for the cognate ligand
174 binding specificity. Neither Loop II (**Figure 5C**, yellow color) nor Loop III (**Figure 5C**,
175 orange color) display a single conserved amino acid shared among the paralogous binding
176 interfaces (as it originally has been described in **Seiradake et al., 2011**). Thus the
177 complementary pattern of the pre-postsynaptic interactions mediated via specific Netrin-
178 G/NGL pairs is reflected in the reciprocally different sizes of the loops binding interfaces
179 representing another element of the *NTNG*-encoded protein paralogs SF.
180

DISCUSSION

Complementary contribution of *NTNG* paralogs into human cognitive pathologies. Involvement of the pre-synaptically expressed axon-localised *NTNGs* in SCZ diagnosis supports the established view of SCZ as a result of distorted trans-synaptic signaling (**Lips et al., 2012**), with a recent study proving that axonal connectivity-associated genes form a functional network visualisable by fMRI (**Richiardi et al., 2015**), and that brain connectivity predicts the level of fluid intelligence (**Finn et al., 2015; Pamplona et al., 2015**). Both *NTNGs* have been found to participate in the brain functional connectivity by the parcellated connectome reconstruction ((**Hawrylycs et al., 2015**). Most of the reported disease associations link *NTNG1* to SCZ with a variety of other neurologic pathologies (15 in total, **Figure 1A-1**), while *NTNG2* pathologic associations (6 in total, **Figure 1A-2**) are quite limited to those affecting WM or PO. Among them is SLE frequently characterized by WM deficit (**Shucard et al., 2011**) and also known to represent schizoid-type abnormalities characteristic for autoimmune pathologies (**Guilloux et al., 2010**; **Eaton et al., 2006**). Immune activation is known to lead to altered pre-pulse inhibition (a key diagnostic trait for SCZ) reversed by antipsychotics (**Romero et al., 2007**). The three diseases associated with both paralogs (ASD, BD and SCZ) are also a primary focus of the recently initiated PsychENCODE project (**PsychENCODE et al., 2015**). It is also worth to mention the resemblance of the reported disease associations with the behavioral phenotypes of *Ntng1* and *Ntng2* gene knockout mice (**Qi et al., in press**).

It is a known fact that a gene content associated with IQ score often relates to numerous diseases, such as SCZ, ASD, depression, and others (see **Zhao et al., 2014** for ref.). Several genes associated with SCZ have undergone positive selection following the human brain evolution (**Xu et al., 2015**). Despite the global network properties of the brain transcriptome are highly conserved among the species there are robust human-specific

disease-associated modules (**Miller et al., 2010**) and human accelerated regions (HARs) - highly conserved parts of genome that underwent accelerated evolution in humans (**Pollard et al., 2006**). HARs can serve as genomic markers for human-specific traits underlying a recent acquisition of modern human cognitive abilities by brain (**Boyd et al., 2015**) but that also "might have led to an increase in structural instability… resulted in a higher risk for neurodegeneration in the aging brain" (**Zhou et al., 2015**), rendering our intellectual abilities genetically fragile (**Crabtree, 2013**) and resulting in a variety of CDs. The role genomic context, epistasis (**Hemani et al., 2014**), plays in the evolution and pathology is manifested by frequently found disease-causing alleles present in animals without obvious pathological symptoms for the host (**Jordan et al., 2015**). Any CD is characterized by general intellectual disability (GID) plus psychiatric symptoms. A genetic perturbation-exerted behavioral cognitive deficit (BCD) in an animal model organism is a poor match to a human CD *per se* due to very poor contextual resemblance between the human GID and animal BCD together with the absence of interpretable psychiatric symptoms. No wonder that the compounds that "cure" mice models consistently fail in human trials (discussed in **Hyman, 2014**).

***NTNG* paralogs brain transcriptome intrinsic complementarity and possible mechanism for the IQ-affecting mutation alleles effect.** There is no global change at the mRNA level between healthy subjects and SCZ patients (**Figure 1B**). This conclusion is supported by previously published works stating that globally altered mRNA expression of *NTNG1* or *NTNG2* is unlikely to confer disease susceptibility, at least in the temporal lobe (**Eastwood and Harrison, 2008**), and Brodmann's area (**Aoki-Suzuki et al., 2005**). However, the original paper-source of the STG samples RNA-seq along with many other genes (>1,000) found that *NTNG1* (but not *NTNG2*) falls under the group of genes with significant alternative promoter usage (**Wu et al., 2012**: **ST6**, p<9.05E-10 at FDR <0.5) and *NTNG2* (but not *NTNG1*) clusters with genes (>700) with significant alternative splicing change (**Wu**

et al., 2012: **ST7**, p<6.15E-12 at FDR<0.5) when SCZ and controls are compared. Such GWAS observation adds an extra layer of complementary regulation to both *NTNG* paralogs on a top of the described in the results section complementary usage rule for the exons, formed unspliced splicing modules, resulting transcripts and their comprising exons (**Figure 1B**). Based on the available RNA-seq dataset it was almost impossible to detect RNA with the matched position of *NTNG* SNPs used for the IQ testing (**ST2c** and **Prosselkov et al., in press**) except for two coding exons located (rs2149171 and rs3824574) and exon 10 non-coding area located but transcribed rs4915045 (in 2 out of 16 samples). This fact points towards indirect effect of the IQ-affecting mutation alleles which can be associated with the shorter (secretable) isoforms generation (**Prosselkov et al., unpublished**) lacking two of the most prominent *NTNG* features: GPI-link and the Ukd domain through an aberrant splicing factor binding. The GPI-link is a hallmark of Netrin-G family members (**Nakashiba et al., 2000, 2002**) and without it the aberrant Netrin-G isoforms are likely to mimic the action of their releasable ancestry molecules - netrins, still being able to bind to their cognate postsynaptic ligand – NGL but without forming an axonal-postsynaptic contact. The Ukd domain of Netrin-G1, despite its so-far unknown function, is involved in lateral binding to the pre-synaptically localised LAR modulating the binding strength between NGL-1 and Netrin-G1 (**Song et al., 2013**). Work is currently underway in search for a similar lateral interaction partner for the Netrin-G2 Ukd domain (**Kim E, personal communications, April 2014**). The inclusion of Ukd encoding exons 6 and 7 is regulated by the Nova splicing factor (**Ule et al., 2005**) affecting the cortex Netrin-G1 exon 7 but not exon 6, and, simultaneously, Netrin-G2 paralog exons exhibiting an opposite pattern. In general, it is tempting to speculate that deregulation of *NTNG* transcripts processing may have a role in the brain-controlled cognitive abilities and associated CDs. Supporting such notion, a decreased level of Netrin-G1c mRNA (exons 6-9 excluded, **Figure 1B-1**) has been reported for BD and SCZ

(**Eastwood and Harrison, 2008**) with Netrin-G1d (exons 6 and 7 included but 8-9 excluded, **Figure 1B-1**) and Netrin-G1f (a secretable short isoform consisted of domain VI only and lacking the Ukd and GPI-link) being increased in BD, but not in SCZ, in anterior cingulated cortex (**Eastwood and Harrison, 2010**). Higher Netrin-G1d mRNA expression in fetal brain but low for the Netrin-G1c isoform in the human adult (**Eastwood and Harrison, 2008**) indicates different functionality of these two splice variants joggling with the Ukd domain inclusion. And, according to our data in the forthcoming manuscript, if Netrin-G1 Ukd-containing isoforms are the dominant isoforms in adult mouse brain, Netrin-G2 Ukd-containing isoforms are present at the trace level (**Prosselkov et al., forthcoming**), resembling a similar transcriptome pattern for the human STG samples (**Figure 1B-1** and **B-2**). A similar "dynamic microexon regulation" associated with the protein interactome misregulation has been reported to be linked to ASD (**Irimia et al., 2014**).

Synchronous and complementary expression of *NTNG* paralogs in the human brain supports the IQ-associated cognitive endophenotypes. Influential parieto-frontal integration theory (P-FIT, **Jung and Haier, 2007**) states that general intelligence ("g") depends on multiple brain cortical areas such as dlPFC, Broca's and Wernicke's areas, somatosensory and visual cortices (**Colom et al., 2009**). Despite "g" is widely accepted as the only correlate of the intelligence, its unitary nature was challenged by (**Hampshire et al., 2012)** claiming had indentified two independent brain networks (for memory and for reasoning) responsible for the task performance, the idea later criticised for the employed data processing approach (**Haier et al., 2014**). Higher IQ scores (a composite surrogate of "g") have been reportedly associated with the fronto-parietal network (FPN) connectivity (**Song et al., 2008**; **Glascher et al., 2009**). High level of *NTNG* paralogs expression within the cognition intensively loaded areas of the brain and the distinct patterns of expression profiles (synchronous, asynchronous/mixed, and complementary, **Figure 2A**) support

associations of *NTNG1* and *NTNG2* with the recorded cognitive endophenotypes (**Prosselkov et al., in press**). Based on the expression patterning over the human life-span, among the total 16 analysed brain areas we found two falling under the same "anti-phasic (complementary)" classifier (**Figure 2C**): HIP and MD. Adding more to that, MD is the only brain area (out of the 16 presented) where *NTNG1* expression level exceeds that of *NTNG2* making it a promising candidate for the phenomena of *NTNGs* SF explanation. Two other brain areas classified by a synchronous paralogs expression deserve a special attention, dlPFC and mPFC (**Figure 2A-4**). PFC circuitry has been known as a "hub of the brain's WM system" (**Kim et al., 2013; Markowitz et al., 2015**), which acts through direct HIP afferents (**Spellman et al., 2015**) and has many connections with other cortical and subcortical areas (**Riga et al., 2014**). mPFC may function as an intelligence-control switchboard and lPFC, part of the FPN global connectivity, predicts the WM performance and fluid intelligence (**Cole et al., 2012**). Interactions of the auditory recognition information fed by the vPFC stream with the sequence processing by the dorsal stream are crucial for the human language articulation (**Skeide and Friederici, 2015**). The fact that both *NTNG* paralogs are extensively expressed across PFC (**Figure 2A-2 and A-4**) pinpoints this area as a key for future molecular studies of the human-unique symbolic communications. And PFC is not only implicated in many psychiatric disorders, including SCZ (**Gulsuner and McClellan, 2014**; see also **Riga et al., 2014** for ref.), but is also the only brain structure unique to primates without known homologs in the animal kingdom (**Wise, 2008**).

Evolution of the protein paralogs encoded by the *NTNGs*. Forkhead box P2 (FOXP2) – a ubiquitously expressed transcription factor that has been reported to be linked to the evolution of human language through T303N, N325S substitutions when compared to a primate ortholog (**Enard et al., 2002**) and is 100% identical to Nea protein (**Krause et al., 2007**). FOXP2 regulates the expression of multiple genes and among them is *LRRC4C* (gene

encoding NGL-1 – a post-synaptic target of Netrin-G1) in human and chimpanzee (**Konopka et al., 2009**). Netrin-G1 similarly to FOXP2 is a 100% conserved protein among the hominins with only 1 mutation found in chimpanzee which is absent in marmoset (and other primates) and mice proteins (**Prosselkov et al., in press**). On the other hand, extinct hominins' Netrin-G2 relative to modern human contains T346A point mutation (as per current version of hg19), also found in primates and mouse and known as rs4962173 (dbSNP missense mutation) representing an ancient substitution from Nea genomes found in modern humans and reflecting a recent acquisition of the novel allele in Ice around 5,300 yrs BC. Nothing is known regarding the functional significance of this mutation but biochemically a substitution of alanine (A) on a polar threonine (T) could bring an extra point of regulation, e.g. a phosphorylation or glycosylation (NetPhos2.0 (**Blom et al., 1999**) assigns a low score for the T346 to be phosphorylated but NetOGlyc4.0 (**Steentoft et al., 2013**) robustly predicts it to be glycosylated, **SM**). Another mutation S371A/V reflects a selective sweep in Netrin-G2 protein from primates to hominins within a similar to T346A functional context when a hydrophobic alanine (in chimpanzee, A)/valine (in marmoset, V) is replaced by a polar serine (S) and a strong positive predictions for glycosylation but not phosphorylation (**SM**). This poses a question whether these two human-specific protein substitutions associate with advanced cognitive traits as they may represent a hidden layer of poorly studied so far protein glycosylation-associated regulatome known to affect the brain function and diseases (**Baenziger, 2012**; **Baenziger, 2013**). Adding more to this, T346 is nested on exon 5 just 20 nu away from the affecting WM score rs2274855 (**Prosselkov et al., in press**), and, together with S371A/V, they are both located within the lowest percent identity area (exons (5-7)) of Netrin-Gs (**Figure 3C**) and, proposedly, contributing to the *NTNG* duplicates SF. There are at least three more protein parts potentially contributing to the gene paralogs specialised function subdivision (based on the low identity scores, **Figure 3C**): the secretory peptide, the

GPI-link, and the outmost structurally elaborated unstructured loops (I-III) responsible for the reciprocal binding of Netrin-Gs to their post-synaptic cognate partners, NGL-1 or NGL-2, both containing a C-terminal PDZ-binding domain (**Kim et al., 2006**). An interesting finding was reported in (**Arbuckle et al., 2010**) found a presence of SH3(PSD95) domain binding site (required for the phosphatidylinositol-3-kinase recruitment) in mice Netrin-G2 (100% identical to human) but not in Netrin-G1. The detected SH3 binding site overlaps with the Netrin-G2-loop III responsible for the binding specificity to NGL-2 (**Seiradake et al., 2011; Soto et al., 2013; DeNardo et al., 2012**). A plausible working hypothesis would be that while internalised (and being GPI-link naïve/immature) the pre-synaptic Netrin-G2 is bound to SH3-PSD95 via loop III but as soon as being secreted extracellularly (and being attached to the membrane) it is bound to post-synaptic NGL-2. Corroborating this, in the absence of Netrin-G2 in the KO mice NGL-2 is unstable on the post-synaptic surface and gets quickly internalized (**Qi et al., in press**). We can only speculate regarding the potential importance of PSD-95(SH3)-Netrin-G2-NGL-2 scaffolding loop interaction/competition but the ability for Netrin-G1 to bind to SH3 has not been reported. Following this logic, Netrin-G1 should have a similar binding partner via loop II while internalised.

The overall identical structural scaffold among the Netrin-G paralogs (**Figure 3D**) is likely to represent an anciently preserved one of the primordial protein (encoded by a single gene in the primitive urochordate *C.intestinalis*) and its contribution to the process of SF among the *NTNG* paralogs goes against the "structure drives function" concept. It looks like that it is not the "structure" but rather the "evolution" itself that drives a selection for the best structural (or unstructural in our case) fit out of the available frameworks provided by the gene duplicates to fulfill the emerged functional demand in a new ecological niche. The intricate variability of phenotype is grounded by the conserved nature of genotype and constrained by the "structure-function" limitations of the coding DNA and is only possible

355 due to permissive evolutionary continuing elaborations of non-coding areas able to absorb the
356 most recently acquired elements (having a potential to become regulatory at some point, e.g.
357 like HAR5 (**Boyd et al., 2015**)) and carried over by neutral drift as proposed by Kimura but
358 for proteins (**Kimura, 1983**). At the same time, the multiple protein substitutions coinciding
359 with the SF labor segregation phenomena among the Netrin-G paralogs question their neutral
360 nature. Both of them undergo a purifying selection from mice to human through the reduction
361 in size of non-coding DNA (introns) and encoded proteins (the mice Netrin-G2 is 2 aa longer
362 its human ortholog) further contributing to the host-specific SF. Thus while the non-coding
363 sequences are used to explore the evolutionary space in time, the restrictive boundaries of the
364 paralogs SF are determined by the protein (unstructured) elements.
365 **Molecular evolution of the Cognitive Complement (CC).** Appearance of the neural crest
366 (**Abitua et al., 2012**), an event that "affected the chordate evolution in the unprecedented
367 manner" (**Green et al., 2015**), multipotent progenitor cells (**Stolfi et al., 2015**), and
368 neurogenic placodes (suggesting a chemosensory and neurosecretory activities, **Abitua et al.,**
369 **2015**) in first primitive urochordates/tunicates coincides with the presence of *Ntng* precursor
370 gene (ENSCING00000024925) later undergoing two rounds of duplication events in lamprey
371 and found to affect human cognitive abilities (**Prosselkov et al., in press**). *NTNG* paralogs
372 are expressed in the human neural crest-forming cells with *NTNG2* 10 times stronger than
373 *NTNG1* (**Rada-Iglesias et al., 2012**), both are differentially expressed in human comparing to
374 chimpanzee and rhesus monkey with *NTNG2* expression model showing stronger probability
375 than *NTNG1* (**Iskow et al., 2012**), and both are stronger expressed in human telencephalon
376 comparing to chimpanzee and macaque (**Konopka et al., 2012**). *NTNG1* has been classified
377 as a brain module hub gene "whose pattern fundamentally shifted between species"
378 (**Hawrylycs et al., 2015**). Belonging to distinct modules of brain expression regulation (**Liu**
379 **et al., 2012**, **Konopka et al., 2012**), *NTNGs* are classified as "genes with human-specific

380 expression profiles" (**Liu et al., 2012**). The nearby gene ~260 kbp upstream of *NTNG2* is
381 *MED27* (mediator of RNA polymerase II) has been proposed to be associated with the
382 evolution of human-specific traits (**McLean et al., 2011**). *NTNG1* has also been reported
383 among the "adaptive plasticity genes" (**Ghalambor et al., 2015**) potentiating rapid adaptive
384 evolution in guppies (*NTNG2* was not found among the input RNA for analysis).

385 Complementarity among the *NTNG* paralogs and encoded by them proteins has been
386 reported previously: brain expression complementary pattern (in almost self-exclusive
387 manner) defined by the 5'-UTR-localised *cis*-regulatory elements (**Yaguchi et al., 2014**);
388 complementary distribution within the hippocampal laminar structures (**Nishimura-Akiyoshi**
389 **et al., 2007**); axon-dendrite synaptic ending resulting in differential control over the neuronal
390 circuit plasticity (**Matsukawa et al., 2014**); mutually-exclusive binding pattern to post-
391 synaptic partners, NGL-1 and NGL-2, dictated by the protein unstructural elements
392 (**Seiradake et al., 2011**); alternative promoter usage *vs* alternative mRNA splicing in SCZ
393 patients (**Wu et al., 2012**); KO mice behavioral phenotypes and subcellular signaling partners
394 complementarity (**Qi et al., in press**); "differential stability" brain modules expression
395 (*NTNG1* is expressed in the dorsal thalamus (M11) as a hub gene (Pearson's 0.92) while
396 *NTNG2* is in neocortex and claustrum (M6, Pearson's 0.65)) (**Hawrylycs et al., 2015**);
397 module top-down *vs* bottom-up information flows gating in mice and differential
398 responsiveness to neuronal activity (**Prosselkov et al., forthcoming**); and human IQ-
399 compiling cognitive domains complementation (**Prosselkov et al., in press**). The current
400 study reports the *NTNGs* complementarity association with the CDs (**Figure 1A**); mRNA
401 splicing pattern complementary at the quantitative and qualitative levels via differential use
402 of the middle-located exons (**Figure 1B**); brain complementary oscillatory expression over
403 the human life span observed in the intensive cognitively loaded brain areas (**Figure 2**); AE
404 of the paralogs-segregated unique non-coding elements (**Figure 3A**); complementary pattern

405 of the protein orthologs (mice-to-human) protein sequence evolution. Such multi-level
406 complementation is likely to reflect a shared evolutionary origin from a single gene in a
407 primitive vertebrate organism 700 mln yrs ago and its subsequent functional segregation
408 among the evolution-generated gene duplicates in jawless fish, such as lamprey.

409 Occupying independent but intercalating functional niches, *NTNG1* and *NTNG2* do
410 not compensate but complement each other's function forming a "functional complement" of
411 genes. Half a billion yrs ago the doubled gene dosage led to the gradual SF and manifested in
412 a function complementation within the cognitive domains, at least in human. We would like
413 to coin such gene pair as a Cognitive Complement (CC).

414

415 CONCLUSION

416 The emerged functional redundancy, as an outcome of gene duplication, leads to function
417 subdivision and its bifurcation among the gene paralogs resulting in the paralogs SF. A
418 functional compensation is known to exist among the evolutionary unrelated genes but has
419 not been reported among the gene paralogs, more frequently characterized by the function
420 complementation. Gene paralogs structural identity (at both, gene and protein levels) does not
421 provide a substrate for functional compensation but rather for complementation perturbating
422 "structure drives function" rule. A gene duplication event of a tunicate *NTNG* primordial
423 gene and the subsequent process of its function specialisation (driven by the new ecological
424 niches appearance and evolution) among the gene duplicates made them to SF into distinct
425 cognitive domains in a complementary manner forming a CC. In our forthcoming work we
426 are to describe how *Ntng* mice genes function resembles that of human orthologs (**Prosselkov**
427 **et al., forthcoming**).

428

MATERIALS AND METHODS

Human brain *NTNG* transcriptome reconstruction. Relates to **Figure 1B** and **1-C**. The original source of the dataset was produced by (**Wu et al., 2012**: E-MATB-1030) and the downloaded .bam files used for the re-processing are listed in **ST1a**. All reconstructed transcripts are presented in **ST1d** stand alone Excel file. Two samples were excluded from the analysis due to failed "per base sequence quality" measure, and zero expression level for *NTNG1a* and *NTNG1int(9-10)* otherwise consistently expressed throughout other samples (**ST1b**). SAMtools software was used for the SNPs calling from the available RNA-seq datasets (**ST1c**). For details refer to **SM**.

Human brain expression profiling for *NTNGs* across the life span. The original source of data was www.brainspan.org. All available samples were initially included into the analysis but two of them excluded at a later stage (MD for 12-13 pcw and mPFC for 16-19 pcw) due to high deviation (6-7 times) from the mean for other replicas. The mean expression values per each brain area as RPKM were plotted against the sampling age. Profiles classification was done visually considering the trend over the all plotted points as an average.

***NTNG1* (*NTNG1m*) and *NTNG2* (*NTNG2b*) full-length mRNA transcripts assembly.** Relates to **Figure 3B.** Human *NTNG1m* brain transcript has been reported previously (**Meerabux et al., 2005**) and we have also confirmed its ortholog presence in the mice brain via full-length cloning (**Prosselkov et al., unpublished**). Since NCBI contains only its partial CDS (AY764265), we used the RNA-seq-generated exons (**Figure 1B**) to reconstruct its full-length and to generate an ORF of the encoded Netrin-G1m. Similarly, human *NTNG2b* was reconstructed from the RNA-seq dataset and from Ensemble as follows. Exon 5 sequence was deduced from ENST00000372179, other exons were from ENST00000467453 (no longer available on the current version of Ensemble) except for exon 6 deduced by running three independent alignments against the human genomic DNA with the mice 3'-intron (5-6),

453 exon 6, and 5'-intron (6-7) concomitantly confirmed by the generated full-length ORF for
454 Netrin-G2b. The reconstructed protein was predicted to encode 587 amino acids, which is in
455 a close proximity to the mice netrin-G2b ortholog of 589 residues (**Prosselkov et al., forth.**).
456 **Full-lengths gene structures of *NTNG* paralogs reconstruction.** Relates to **Figure 3A.**
457 Both, the obtained above from the STG brain samples RNA-seq and the reconstructed full-
458 lengths transcripts carrying all stably expressed exons were used to confirm the intron-exon
459 junctions positioning for *NTNG1* and *NTNG2*. Due to observed variability in the intron (1-2)
460 and exon 10 sizes their boundaries were left unmarked.
461
462 SUPPLEMENTARY MATERIALS (**SM**)
463 Contain Supplementary Methods (RNA-seq of STG re-processing and SNPs detection) and
464 Supplementary Tables (**ST1a-d, ST2**) as a single compiled pdf file. Reconstructed RNA-seq
465 (.gtf) of the STG is presented as a standalone Excel file (**ST1d**). Also included: Netrin-G2b
466 predicted phosphorylation and *O*-glycosylation, Netrin-G1 *vs* Netrin-G2 Ukd alignment
467 (**McWilliam et al., 2013)**, predicted secretory peptide cleavage and GPI attachments.
468
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475
476 COMPETING INTERESTS
477 Authors would like to express a lack of any competing interests associated with the work.
478

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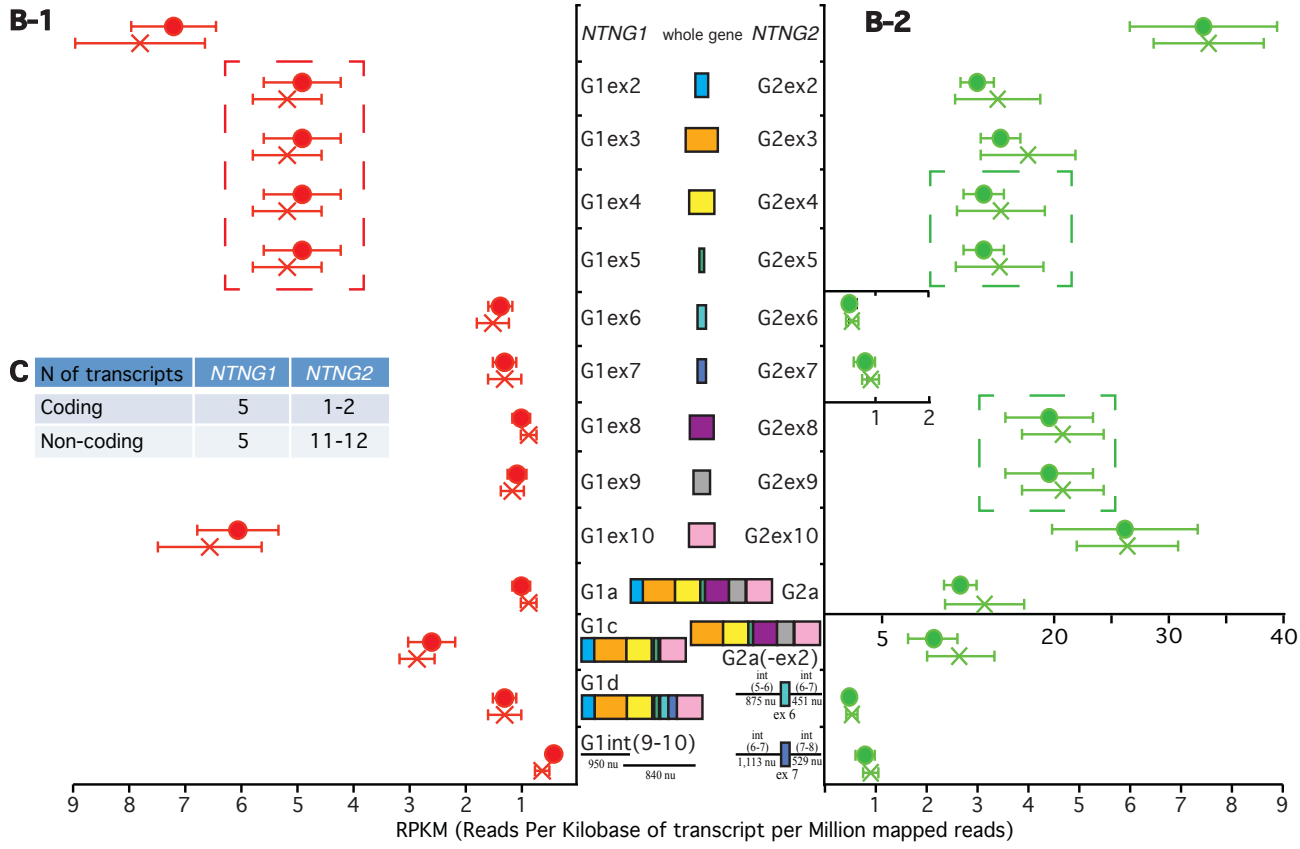
A-1 *NTNG1* reported associations	Cognitive disorder (CD)	**A-2** *NTNG2* reported associations
Voineagu et al., 2011; O'Roak et al., 2012a,b; King et al., 2013*; Iossifov et al., 2014; Sanders et al., 2015; D'Gamma et al., 2015	Autism sporadic disorder (ASD)	Sanders et al., 2015; D'Gamma et al., 2015
Eastwood and Harrison, 2008, 2010; Akula et al., 2014	Bipolar disorder (BD)	Eastwood and Harrison, 2008, 2010; Akula et al., 2014
Fukasawa et al., 2004; Aoki-Suzuki et al., 2005; JSSLG et al. 2005; Eastwood and Harrison, 2008; Ohtsuki et al., 2008; Zakharyan et al., 2011; Zhu et al., 2011; Ayalew et al., 2012; Wilcox and Quadri, 2014	Schizophrenia (SCZ)	Aoki-Suzuki et al., 2005; Eastwood and Harrison, 2008
Zhang et al., 2013	Alzheimer disease (AD)	
Lagier-Tourenne et al., 2012*; Ishigaki et al., 2012*; Rogelj et al., 2012*; Nakaya et al., 2013*	Amyotrophic lateral sclerosis (ALS)	
	Fronto-temporal lobe dementia (FTLD)	
Maurano et al., 2012; Wang et al., 2011; Boraska et al., 2014	Anorexia nervosa (AN)	
van Kuilenburg et al., 2009	Dihydropyrimidine dehydrogenase deficiency (DPD)	
Gilissen et al., 2014	Intellectual disability (ID)	
Stepanyan et al., 2013	Ischemic stroke (IS)	
Bisgaard et al., 2007	Mental retardation (MeR)	
Stewart et al., 2013*	Obsessive-compulsive disorder (OCD)	
Lesnick et al., 2007	Parkinson disease (PD)	
	Pseudopapillary neoplasm (PN)	Cavard et al., 2009
Borg et al., 2005; Archer et al., 2006; Nectoux et al., 2007	Rett syndrome (RS)	
	Systemic lupus erythematosus (SLE)	Maurano et al., 2012
	Temporal lobe epilepsy (TLE)	Pan et al., 2010
Scharf et al., 2013*	Tourette syndrome (TS)	



C

N of transcripts	*NTNG1*	*NTNG2*
Coding	5	1-2
Non-coding	5	11-12

Figure 1. *NTNG* paralogs complementation within neurological disorders and brain transcriptome. (**A-1, A-2**) Reported cognitive disorders associations for *NTNG1* and *NTNG2*. *denotes rather an indirect association via a direct interaction with the research target. **(B-1, B-2)** RNA-seq of the STG of healthy (circle) and SCZ (cross) human subjects. The original dataset was produced by (**Wu et al., 2012**), accession number E-MTAB-1030 on ArrayExpress (**ST1a**) and reprocessed as described in **SM**. Five *NTNG1* and four *NTNG2* transcripts, consistently expressed across all 16 human samples are shown. Two samples (one healthy and one SCZ) have been omitted due to unsatisfactory quality of reads and expression profiling (**ST1b**). For the SNPs calling by SAMtools see **ST1c**. Data are presented as a mean RPKM±SEM. **(C)** Total number of the assembled transcripts across all samples for both paralogs (see **ST1d** for the completely reconstructed transcriptome). Dash-outlined are co-spliced exon clusters.

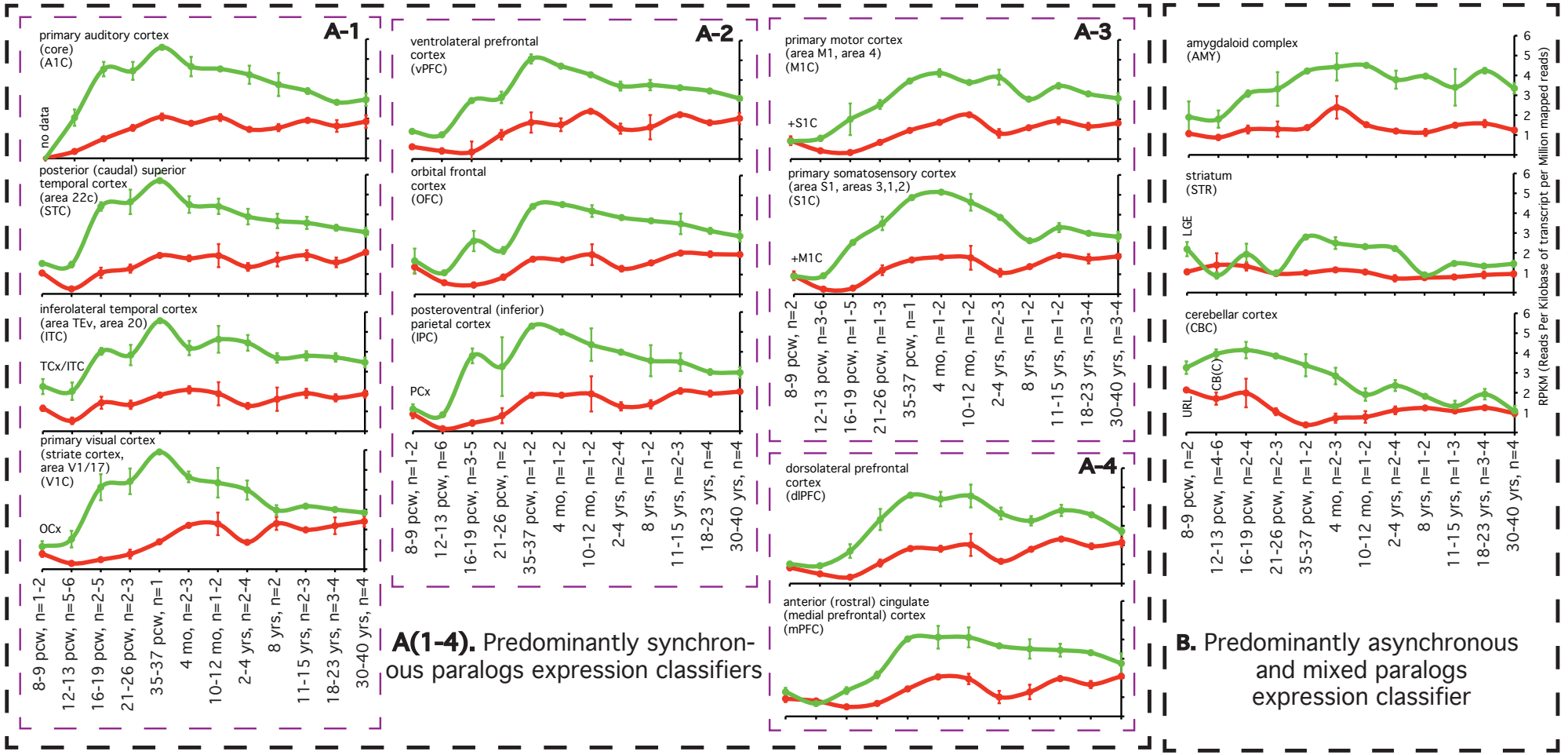


Figure 2. *NTNG* paralogs expression dynamics classification (A-C) in the human brain across the life span. A (1-4): further subdivision of the classifier. RNA-seq data are from the BrainSpan (www.brainspan.org) presented as a mean±SEM. TCx = temporary neocortex; OCx = occipital neocortex; PCx = parietal neocortex; LGE = lateral ganglionic eminence; CB(C) = cerebellum(cortex); DTH = dorsal thalamus; URL = upper (rostral) rombic limb; pcw = post-conception week. Two data points (MD, 12-13 pcw, and mPFC, 16-19 pcw) for *NTNG1* expression were omitted as they were 6-7 times different from the mean of other replicas. All processed brain samples are listed in **ST2**.

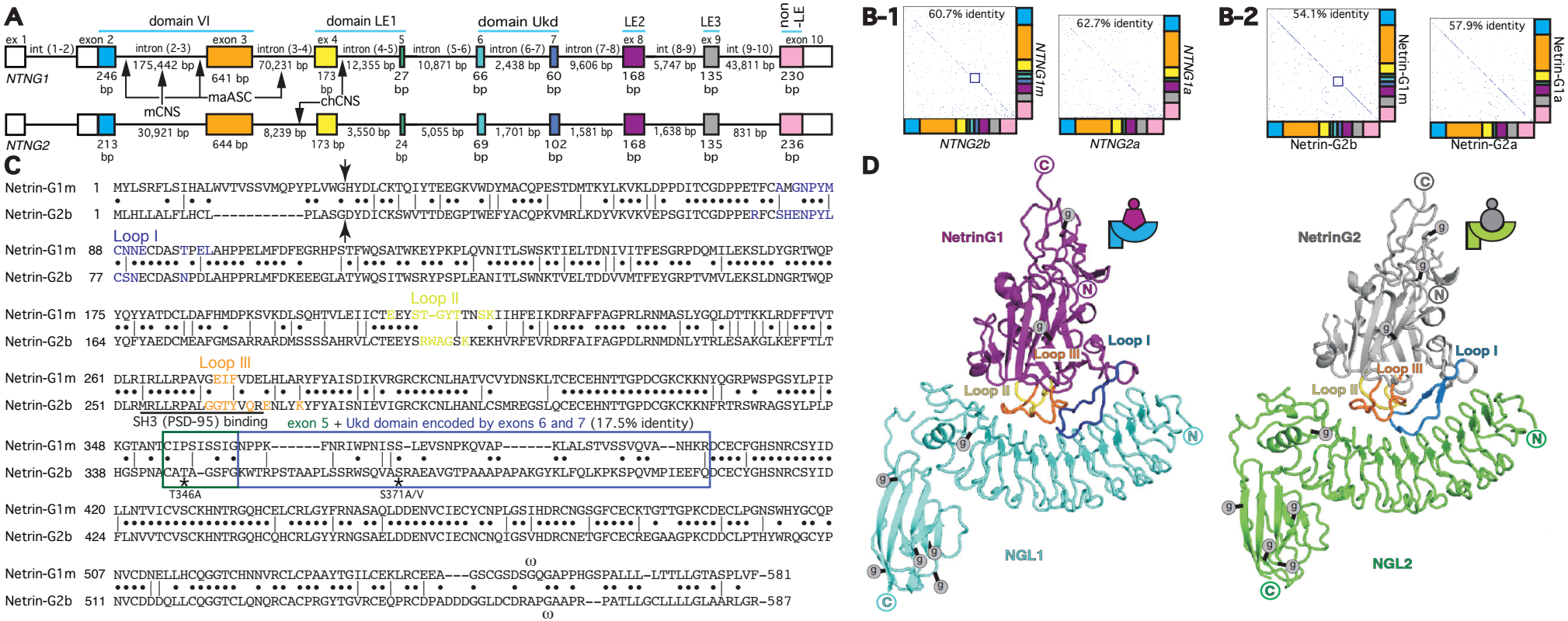


Figure 3. Human *NTNG* paralogs DNA and protein sequence comparisons and "structure-function" rule incongruency. (**A**) Identical gene structures with different sizes of introns. RNA-seq data from **Figure 1B** were used to precisely deduce the exon/intron junction boundaries. The sizes of exons 1, 10 and introns (1-2) are not indicated due to observed among the splice transcripts lengths variability (see **ST1a** for details). Arrows indicate location of CNS = conserved non-coding sequences underwent accelerated evolution in human compare to mice (mCNS) and chimpanzee (chCNS), as per **Prabhakar et al., 2006**; and ASC = anthropoid-specific constrained regions in human compare to marmoset (maASC), as per **del Rosario et al., 2014**. (**B**) Identical exonal composition of the longest *NTNG* encoded RNA paralog transcripts and corresponding proteins with relatively high percent of identity among them dependent on the included/excluded Ukd domain (**B-2**) encoded by the exons 6 and 7 (**B-1**). Notably, the protein sequence represents higher percent of the paralogs difference than encoded it DNA. The matrices were obtained by GeneJockey II (Biosoft). (**C**) Protein alignments for the longest human *NTNG* encoded proteins, Netrin-G1m and Netrin-G2b, with Loops I-III highlighting binding sites for their cognate post-synaptic binding partners NGL-1 (*Lrrc4c*) and NGL-2 (*Lrrc4*), respectively, as determined by **Seiradake et al. (2011)**. Arrow indicates a putative secretory cleavage site location, as calculated by SignalIP (**Petersen et al., 2011**), the blue rectangle delineates the area of the lowest identity (3'-domain LE1+Ukd domain); ω – denotes a point of putative GPI-attachment, as predicted by Big-PI (**Eisenhaber et al., 2000**). PSD-95 interaction site via the SH3-binding domain (**Arbuckle et al., 2010**, as determined for mice Netrin-G2) overlaps with the Loop III NGL-2 binding surface. Two stars indicate a modern human (T346A) and a hominin-specific (S371A/V) amino acid substitutions (**Prosselkov et al., in press**). **(D)** Identical structural motif of the Netrin-G1/NGL1 and Netrin-G2/NGL2 complexes as per **Seiradake et al. (2011)**. The figure's reproduction is covered by the Creative Commons license.