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**Ethoscopes: an open platform for high-throughput *ethomics***

Quentin Geissmann<sup>1</sup>, Luis Garcia Rodriguez<sup>2</sup>, Esteban J. Beckwith<sup>1</sup>, Alice S. French<sup>1</sup>, Arian R. Jamasb,<sup>1</sup> and Giorgio F. Gilestro<sup>1\*</sup>

<sup>1</sup> Department of Life Sciences, Imperial College London, London, UK

<sup>2</sup> Polygonal Tree ltd. London, UK

\*To whom correspondence should be addressed. E-mail: [giorgio@gilest.ro](mailto:giorgio@gilest.ro); [g.gilestro@imperial.ac.uk](mailto:g.gilestro@imperial.ac.uk)

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## 29 **Abstract**

30 We present ethoscopes, machines for high-throughput ethomics in *Drosophila* and other small animals.  
31 Ethoscopes provide four unique features: they provide a software and hardware solution that is  
32 reproducible, scalable and open-source; they perform not just real-time tracking, but faithful real-time  
33 profiling of behaviour using a supervised machine learning algorithm; they can stimulate flies in a  
34 feedback-loop mode; they are highly customisable and open source.

35

36 Understanding how behaviour is encoded in the brain is one of the ultimate goals of neuroscience. In  
37 particular, much of modern neurobiology focuses on finding the genes and the neuronal circuits  
38 underlying simple and complex behaviours alike, aiming to describe and ultimately understand how the  
39 brain processes sensory inputs into motor outputs. For many years, starting from Seymour Benzer's  
40 seminal work<sup>1</sup>, the fruit fly *Drosophila melanogaster* has been considered the model organism of  
41 choice to dissect the genetics of behaviour. In the past decade, *Drosophila* has also emerged as an  
42 excellent model for studying not only the genes, but the neuronal circuitry of behaviour too: the  
43 combination of a rapidly delineating connectome together with an unrivalled repertoire of genetics  
44 tools has established fruit flies as the most promising animal model to study neuronal circuits.  
45 Optogenetics, thermogenetics, a genome wide collection of RNAi lines, and a plethora of crafted and  
46 carefully described GAL4 lines constitute an unprecedented arsenal for neurobiologists interested in  
47 studying the neuronal circuitry underpinning behaviour. The limiting factor for *ethomics* – the high-  
48 throughput approach to behavioural studies - is therefore not the availability of genetic tools, but the  
49 access to an objective, reproducible and scalable system to detect and categorise behaviour. In the past  
50 few years, several computational approaches were introduced to address this limitation: some  
51 specifically dedicated to a subset of behaviours, like sleep<sup>2-4</sup>, and others designed to be more versatile<sup>5-</sup>  
52 <sup>8</sup>. However, while computer-assisted analysis of behaviour has the potential of revolutionising the field,  
53 adoption and throughput of currently available techniques are limited by several factors: among others,  
54 the scope and versatility of the software itself and the requirement for a non standardised hardware-  
55 setup, with relevant problems of cost, footprint, and scalability. Here we present a tool aimed at solving  
56 these issues, providing an affordable and versatile complete suite to study *ethomics*: ethoscopes.

57 An ethoscope is a self-contained machine able to either record or detect in real-time the activity  
58 of fruit flies (and potentially other animals) using computerised video-tracking. It relies on an  
59 independent small single-board computer (Raspberry Pi<sup>9</sup>) and a high-definition camera (Raspberry Pi

60 camera) to capture and process infrared-illuminated video up to a resolution of 1920x1080, at 30  
61 frames per second (**Fig. 1a**). Ethoscopes are assembled in a 3D-printed case measuring approximately  
62 10x13x19 cm (**Fig. 1b**). Although we recommend 3D printed assembly for research-grade use, we also  
63 provide detailed instruction to build a fully functional ethoscope out of LEGO bricks (**Fig. 1c**, -  
64 LEGOScope **Supplementary Material S1**) or out of folded cardboard (**Fig. 1d**, - PAPERscope  
65 **Supplementary Material S2**). These latter two options are mainly aimed at education and outreach but  
66 they may also be adopted in a real laboratory environment. The technical drawings required to 3D-print  
67 and assemble an ethoscope, along with its software (Python code on a Linux instance) are released  
68 under the open source GPL3 license and freely available on the ethoscope website  
69 (<https://lab.gilest.ro/ethoscope>). The combination of consumer-grade electronics, 3D printing and free  
70 open source software results into a total cost of about £80 for each machine. Limited cost, along with  
71 the fact that each ethoscope relies on its own computing power, allows for easy scaling of the entire  
72 platform.

73 In a typical usage scenario, several ethoscopes are placed in a climatic controlled chamber, each  
74 powered through a USB cable; ethoscopes connect via WIFI to a computer acting as data collecting  
75 station (“the hub” in **Supplementary Fig. 1a**) and are controlled remotely via a graphical web interface  
76 (**Supplementary Fig. 1** and **Supplementary Video S1**). If the hub is connected to the internet, the  
77 entire platform will receive automatic software updates from a central GIT repository. Flies to be  
78 tracked are loaded into a behavioural arena that slides and locks inside the lower part of the ethoscope  
79 (**Fig. 1a**). Alike the rest of the machine, arenas are also 3D-printed and the design of the arena to be  
80 employed depends on the nature of the experiment: some examples of arenas developed in our  
81 laboratory are provided in **Fig. 1e-k** and span arenas adopted for long term sleep experiments that may  
82 be lasting weeks (**Fig. 1e-g,j**) or short term assays such as decision making (**Fig. 1h**) and courtship  
83 (**Fig. 1i,k**). When starting an experiment, the experimenter can decide whether the activity of the  
84 animals should be tracked in real-time or whether the ethoscope should record a video to be analysed at  
85 a later time, with the ethoscope software or with another software of choice. In real-time tracking  
86 mode, ethoscopes will detect and record the position and orientation of each animal with a variable  
87 frame rate of 1-4 frames per second (**Fig. 2a-b**).

88 To validate the accuracy of tracking, we recorded 2736 hours of video and asked three  
89 independent experienced fly researchers to manually annotate the position of flies in 1413 frames. We  
90 then compared the manually annotated positions to the positions detected by ethoscopes, and found that

91 the median distance between the two was 0.3 mm, corresponding to a tenth of a fly body length. In no  
92 cases, did the error exceed one body length (2.5mm). To enrich the capabilities of ethoscopes, we also  
93 implemented a real-time behavioural annotator. We created a ground-truth of 1297 videos, each lasting  
94 10 seconds and each manually annotated by at least three experienced fly researchers (**Fig. 2c**,  
95 annotation labels were: “walking”, “micro-movement” or “immobile”). Random forest variable  
96 importance<sup>10</sup> was used to screen for possible predictors of mobility in a supervised manner and the two  
97 highest-ranking features - maximal velocity and cumulative walked distance – were selected for further  
98 analysis (**Fig. 2d,e**). Conveniently, ROC analysis showed that maximal velocity alone appeared to  
99 serve as faithful predictor of behaviour (**Fig. 2d,e**). Therefore, not only can ethoscopes annotate the  
100 position of flies, but also detect in real-time when an animal is immobile, performing a micro-  
101 movement (such as grooming or eating), or walking, with an accuracy of 94.3% for micro-movement  
102 detection and 99.0% for walking detection (  $\text{accuracy} = (\text{N\_true\_positive} + \text{N\_true\_negative}) / \text{N}$  ).

103 The ability to recognize simple behaviour in real-time opens a new perspective: animal  
104 -dependent feedback-loop stimuli delivered upon behavioural trigger. Interfering with behaviour  
105 through external stimuli is an important tool for neuroscientists as it pushes beyond description and  
106 allows for more manipulative analysis. In principle, feedback loops can be used for multiple purposes:  
107 to reinforce learning, to sleep deprive animals, to stimulate or silence circuits using optogenetics, to  
108 study operant conditioning, *etc.* Systems operating feedback-loop stimuli on fruit flies were proposed  
109 before and already proved to be useful, but they are not easily compatible with a high-throughput  
110 approach and are focused on very specific usage<sup>11,12</sup>. We therefore designed ethoscopes so that they  
111 could be extended with modules that seamlessly connect with the machine and react to real-time  
112 analysis to trigger an action whenever a condition is satisfied. Figure 3 shows three examples of such  
113 modules: an air/gas/odour (AGO) delivery module (**Fig. 3a,b**), a rotational module capable of variable  
114 intensity(**Fig. 3d,e**), and an opto-motor module combining optogenetic stimulation and motor  
115 disturbance (**Fig. 3g,h**). Modules plug into the bottom part of the machine and are configured through  
116 the graphical web-interface, where the experimenter can set the trigger conditions that will activate the  
117 stimulus. A trigger can be a combinatorial ensemble of position, time, and behaviour (e.g. “*micro-*  
118 *movement for at least 20 seconds within 5mm from the food*” or “*immobile for at least 5 minutes*  
119 *anywhere*”). As proof of principle, we provide representative evidence of how single flies react to three  
120 different stimuli: a 5 seconds delivery of CO<sub>2</sub>, triggered by crossing of the midline tube (**Fig. 3c**); a 2  
121 seconds fast rotation of the tube, triggered by 20 seconds of immobility (**Fig. 3f**); a 5 seconds opto-

122 stimulation on “moon-walker”<sup>13</sup> receptive flies, manually triggered (**Supplementary Video S2**). In  
123 general, flies responses to stimuli will obviously depend on the experimental paradigm of choice. On  
124 the ethoscope website, we provide detailed instruction on how to build all three modules and a  
125 description of the API needed to interface any new custom module to the ethoscope platform.  
126 Ultimately, we expect and encourage users to build modules based on their own scientific needs. One  
127 unprecedented strength of the feedback-loop module system is the ability to interact with single flies  
128 rather than with the entire population, which provides, among other things, the option of performing  
129 sham treatments.

130 Ethoscopes emerge from the maker culture to combine three of the most revolutionary  
131 innovations of the last decades - 3D printing, small single-board computers and machine learning - into  
132 a novel paradigm for behavioural researchers. They were designed to be easy to build, inexpensive,  
133 compatible with high-throughput research, and able to generate reproducible results. Moreover, in their  
134 LEGO and paper versions, they can serve as excellent tool for education and citizens science.  
135 Ethoscopes rely heavily on Raspberry Pis, the third best-selling computer of all time, currently running  
136 at their 3<sup>rd</sup> hardware version. We expect Raspberry Pis to continue in their evolution, and we therefore  
137 expect ethoscopes’ computing power to grow accordingly. A standardised, plug-and-play, inexpensive  
138 tool for behavioural analysis – like ethoscopes are – can be instrumental for future development of the  
139 behavioural field, similarly to how activity monitors have been instrumental for the success of  
140 circadian biology.

141

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152

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## 154 **Figure Legends**

155

156 **Figure 1** | The ethoscope. **(a)** Exploded drawing of a prototypical ethoscope. The machine is composed  
157 of two main parts: an upper case containing the Raspberry Pi and its camera, and a lower case  
158 providing infrared light (IR) and support for the experimental arena. The two cases are separated by  
159 spacers maintaining a fix focal distance (140mm). **(b)** A rendered drawing of the assembled model  
160 showing actual size. The arena slides in place through sliding guides and locks into position. A WebGL  
161 interactive 3D model is available as **Supplementary Material 1**. **(c)** The LEGOScope, a version of the  
162 ethoscope entirely built using LEGO bricks. A detailed instruction manual is provided in  
163 **Supplementary Material 2**. **(d)** A paper and cardboard version of the ethoscope, assembled using  
164 220gsm paper and 1mm grayboard. Blueprints are provided in **Supplementary Material 3**. In all  
165 cases, ethoscopes must be powered with a 5Vdc input using a common USB micro cable either  
166 connected to the main or to a portable power-pack. Cables not shown for sake of simplicity. **(e-k)**  
167 Versatility of use with custom behavioural arenas. Examples of 7 different behavioural arenas  
168 developed in our laboratory. **(e)** Sleep arena. Most commonly used arena for sleep studies, lodging 20  
169 individual tubes. **(f)** Long tubes arena. Used for odour delivery studies or, more generally, for  
170 behaviours where longer walking is required. **(g)** Food bullet arena. Animals are placed directly on the  
171 arena and food can be replaced by pushing in a new bullet. **(h)** Decision making arena. Can be used to  
172 study simple decision making behaviours. **(i)** Square wells arena. Can be used for courtship assay or to  
173 record activity in a bi-dimensional environment. **(j,k)** Conceptually analogs to E and I, but designed to  
174 work in high-resolution (full-HD) settings. Note that all arenas are marked with three visible reference  
175 points (indicated by a red circle in **e.**) that are used by the ethoscope to automatically align images for  
176 tracking, providing a degree of physical flexibility.

177

178 **Figure 2** | Tracking and validation of behavioural classification. **(a)** In real-time tracking mode,  
179 ethoscopes record Cartesian coordinates ( $x,y$ ) of each animal relative to their ROI (region of interest),  
180 along with the numbers describing an ellipsis circumscribing the animal ( $w,h,\phi$ ). **(b)** A screenshot of  
181 the data table recorded by ethoscope, showing four data points for a single fly. **(c)** To build a statistical  
182 model of activity, we used ethoscopes to record offline 2736 hours of video (144 hours x 19 flies) at  
183 resolution of 1280x960pixels and frame rate of 25FPS. Video fragments of the duration of 10 seconds  
184 were sampled every hour for all 19 animals and then scored by at least three experienced fly

185 researchers in a randomized order. Consensual annotations – with a margin greater than 0.5 – were  
186 kept, resulting in a ground truth of 1297 video fragments (116 ambiguous annotations were excluded  
187 using this latter criteria). Scorers manually annotated both the position of the animal in the tube and the  
188 perceived behavioural state (i.e. immobile, micro-moving or walking). Ethoscope video-tracking were  
189 run independently on the whole video resampled at 1-5FPS, all realistic frame rates for real-time  
190 analysis. **(d)** Performance of maximal velocity and cumulative walked distance as features for 1297  
191 annotated videos of behaviours. **(e)** Marginal distribution of maximal velocity for each behaviour,  
192 showing the thresholds used to detect movement (dotted line) and walking (dashed line). **(f)** ROC  
193 curves for movement detection. A conservative threshold of 0.36mm/s, indicated by a red dot on the  
194 curve, yields a sensitivity (TPR) of 91.26% and specificity (1-FPR) of 99.65%. **(g)** ROC curve for  
195 walking detection. A threshold of 0.9mm/s (red dot on the curve) results in sensitivity and specificity of  
196 99.34 and 99.39, respectively.

197

198 **Figure 3** | Versatility of use with behavioural feedback-loop modules. **(a)** Diagram and **(b)** detail of the  
199 air/gas/odour (AGO)-delivery module. Two independent air flows (blue and purple in the drawing) are  
200 fed into the module using external sources. The module features 10 LEGO valves, each independently  
201 controlled through a servo motor. The motor switches the air source on the valve, selecting which  
202 source will be relayed to the tube containing the fly. Available positions are: blue source, purple source,  
203 no source. **(c)** Representative response of three flies subjected to CO<sub>2</sub> administration using the AGO  
204 module. CO<sub>2</sub> release lasts 5 seconds (gray bar) and it is triggered by midline crossing (red dot). The  
205 blue line indicates the fly position in the tube over the 150 second period. **(d)** Model and **(e)** detail of  
206 the rotational module. The module employs a servo motor to turn the tube hosting the fly. Direction,  
207 speed, duration and angle of the rotation can be modulated to change the quality of the stimulus. **(f)**  
208 Representative response of three flies upon stimulation using the rotational module shown in **(d,e)**.  
209 Rotation of the tube is triggered by 20 consecutive seconds of immobility (dashed line) and it is  
210 followed by 5 seconds of masking during which tracking is suspended to avoid motion artefacts (cyan  
211 area). The bottom panel shows traces of a dead fly. **(g)** Model of the optomotor module, able to  
212 simultaneously stimulate single flies with motion and light. **(h)** Detailed view of the optomotor  
213 principle. Light is directed into the tube using optical fiber. **Supplementary Videos 2** shows the  
214 optomotor module in action.

215



216 **Supplementary Figure S1** | The ethoscope platform. **(a)** A diagram of the typical setup. Ethoscopes,  
217 powered through a USB adapter, are connected in an intranet mesh through an Access Point (AP) or a  
218 WI-FI router. A server computer in the network acts as hub, downloading data from ethoscopes and  
219 serving a web-based user interface (UI). Ethoscopes can be controlled through the web-UI, either  
220 locally or remotely. **(b)** Screenshot of the homepage of the web-UI, showing a list of running machines  
221 and some associated experimental metadata (e.g. username and location). **(c)** Screenshot of an  
222 ethoscope control page on the web-UI, providing metadata about the experiment and a real-time  
223 updated snapshot from the ethoscope point of view.

224

225 **Supplementary material 1** | Interactive 3D rendering of the assembled ethoscope – requires WebGL  
226 capable browser (e.g. Google Chrome)

227

228 **Supplementary material 2** | Instruction booklet for building a LEGOScope

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230 **Supplementary material 3** | Instruction booklet for building a PAPERscope

231

232 **Supplementary Videos S1** | An overview of how the ethoscope platform works

233

234 **Supplementary Videos S2** | The optogenetics component of the optomotor module in action.  
235 Moonwalking flies ( VT200107-Gal4 :: UAS-CsChrimson ) are illuminated for 5-7 seconds using a red  
236 LED (630nm) through an optical fibre.

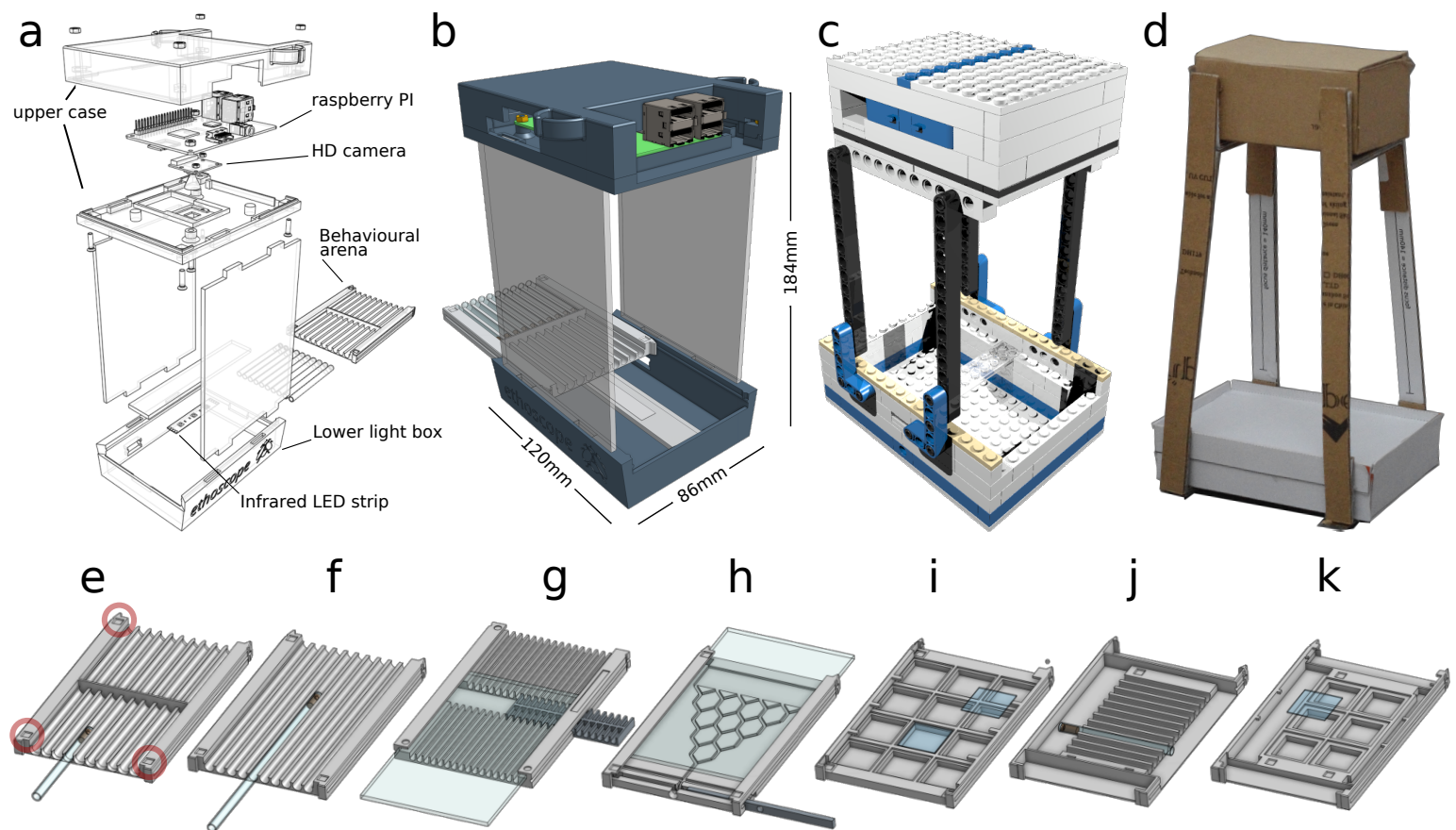


Figure 1  
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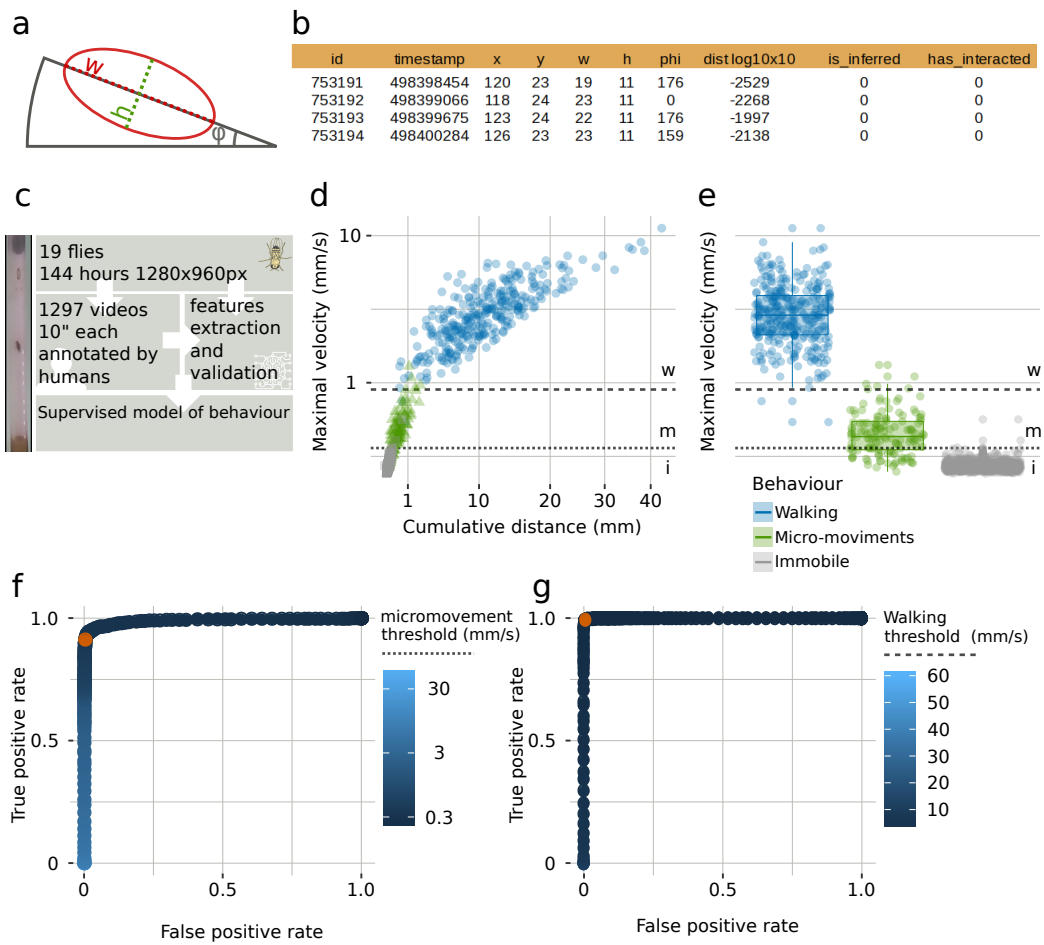


Figure 2  
Geissmann et al.

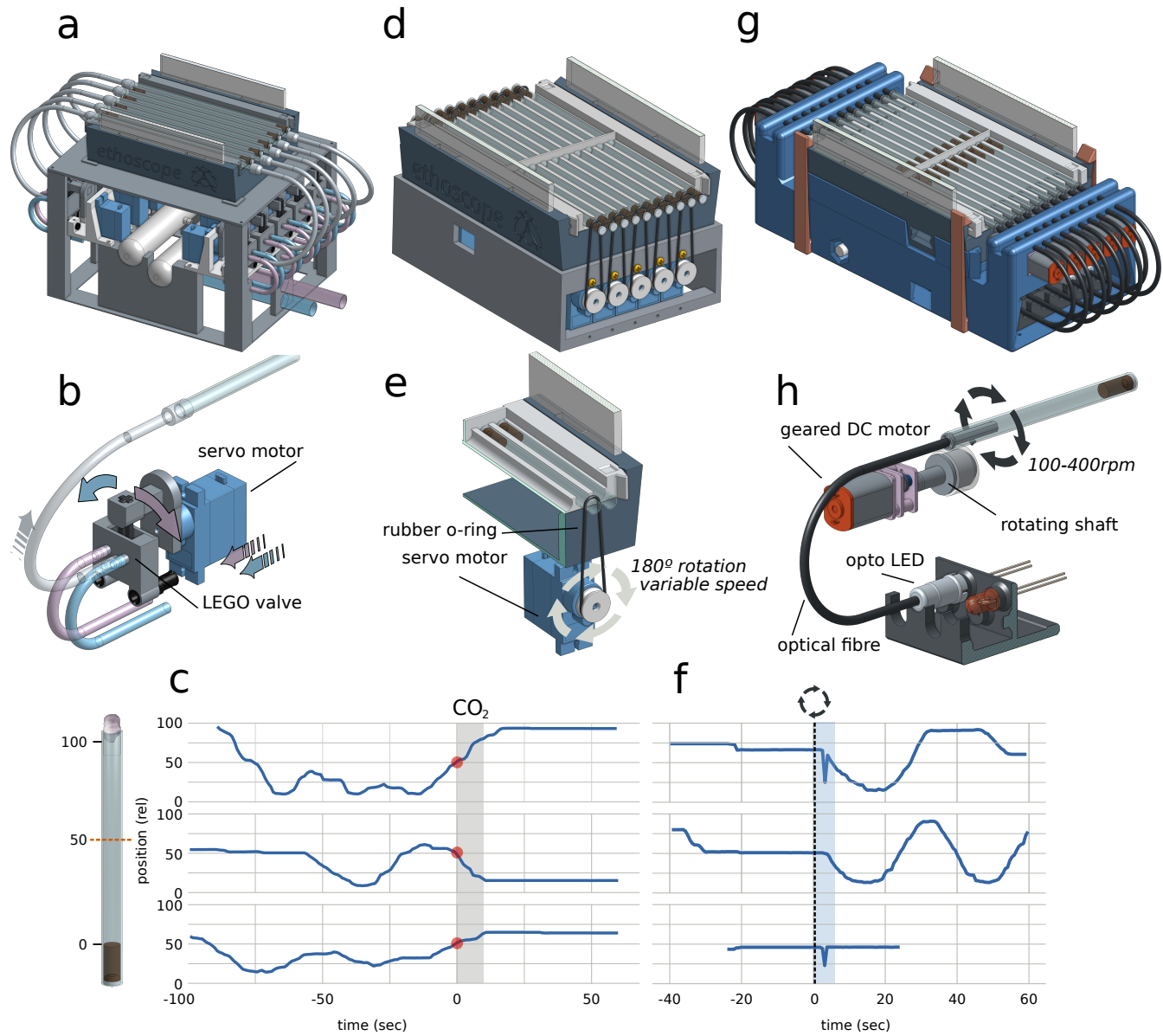


Figure 3  
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