

A theory that predicts behaviors of disordered cytoskeletal networks

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SUMMARY

Morphogenesis in animal tissues is largely driven by tensions of actomyosin networks, generated by an active contractile process that can be reconstituted *in vitro*. Although the network components and their properties are known, the requirements for contractility are still poorly understood. Here, we describe a theory that predicts whether an isotropic network will contract, expand, or conserve its dimensions. This analytical theory correctly predicts the behavior of simulated networks consisting of filaments with varying combinations of connectors, and reveals conditions under which networks of rigid filaments are either contractile or expansile. Our results suggest that pulsatility is an intrinsic behavior of contractile networks if the filaments are not stable but turn over. The theory offers a unifying framework to think about mechanisms of contractions or

23 expansion. It provides a foundation for the study of a broad range of processes
24 involving cytoskeletal networks, and a basis for designing synthetic networks.

25

26 INTRODUCTION

27 Networks of cytoskeletal filaments display a variety of behaviors. A decisive
28 feature for the physiological role of networks is whether they contract or expand.
29 For instance, actomyosin cortices can contract, and the tensions thus created
30 determine the morphology of animal cells (Sanchez et al., 2011). Conversely, the
31 mitotic spindle at anaphase is a network of microtubules that extends to segregate
32 the chromosomes. Such behaviors are essential, but we still lack an intuitive
33 understanding of how they come about, as it is difficult to extrapolate between the
34 microscopic level, where filaments are moved by molecular motors and restrained
35 by crosslinking elements, and the level of the entire system. Cytoskeletal filaments
36 and many of their associated factors are well characterized biochemically. With
37 sufficient knowledge of the relevant properties of the components of a particular
38 network, it should be possible to predict the network behavior. Traditional
39 approaches were particularly successful in predicting passive systems composed of
40 reticulated polymers (Wolff and Kroy, 2012), and more recent developments in
41 active gel theories address networks containing molecular motors (Prost et al.,
42 2015). These latter theories however cannot explain the contractile or expansile
43 nature of the network, as it arises from microscopic interactions that are not

44 represented in the theory. To understand why contractility occurs, one must
45 describe the system at higher resolution, and consider motors and filaments
46 explicitly (Carlsson, 2006; Kruse and Jülicher, 2000; Lenz and Gardel, 2012;
47 Liverpool et al., 2009; Liverpool and Marchetti, 2003; Ziebert et al., 2009). Small
48 networks can also be studied with computer simulations (Bidone et al., 2017;
49 Ennomani et al., 2016; Hiraiwa and Salbreux, 2016; Mendes Pinto et al., 2012; Oelz
50 et al., 2015), but we miss a simpler approach that can make rapid predictions purely
51 based on analytical deduction. Such a theoretical framework would be particularly
52 valuable to classify the different behaviors that are seen experimentally. In search
53 for such a general theory, we chose initially to concentrate on the major factor
54 determining contraction of networks, that is the force created by molecular motors,
55 although we recognized that filament shortening could also lead to contractility
56 (Backouche et al., 2006; Mendes Pinto et al., 2012; Oelz et al., 2015). *In vitro*
57 experiments have shown that contractility can arise with stabilized filaments. In such
58 experiments, the filaments are initially distributed randomly, and molecular motors
59 or crosslinkers added to the mixture make random connections between
60 neighboring filaments. The active motions of molecular motors then drive network
61 evolution. With microtubules and kinesin oligomers, static patterns such as asters
62 (Köhler et al., 2011; Nedelec et al., 1997) or dynamic beating patterns (Katoh et al.,
63 1998; Sanchez et al., 2011; Takiguchi, 1991; Thoresen et al., 2011) can arise. While
64 radial (Backouche et al., 2006) and other patterns (Köhler et al., 2011) were also

65 observed with actin, F-actin networks activated with myosin are predominantly
66 contractile, as demonstrated in various geometries: bundles (Kato et al., 1998;
67 Takiguchi, 1991; Thoresen et al., 2011), rings (Reymann et al., 2012), planar
68 networks (Murrell and Gardel, 2012), spherical cortices (Carvalho et al., 2013; Shah
69 et al., 2014; Vogel et al., 2013) or 3D networks (Bendix et al., 2008; Koenderink et
70 al., 2009). Microtubule networks with NCD or dynein motors are also contractile
71 (Foster et al., 2015; Surrey et al., 2001). Several interesting mechanisms of
72 contraction have been proposed and reviewed recently (Murrell et al., 2015;
73 Salbreux et al., 2012), but each of these applies only to a particular system for which
74 it explains the behavior. We propose here a theory that describes general principles
75 of contractility and that can be applied to both microtubule and actin systems. We
76 also illustrate that contractile systems become pulsatile if filament turnover is
77 introduced in the model.

78

79 **RESULTS**

80 **A Simple Theory Predicts the Behavior of Random Networks**

81 Let us consider a disorganized set of filaments connected by active and
82 passive “connectors” made of two functional subunits (Fig. 1A, B). Possible passive
83 connectors are crosslinkers such as Ase1, Plastin, alpha-Actinin or Filamin, whereas
84 active connectors are oligomeric motors such as Myosin mini-filaments, Dynein
85 complexes, bivalent motors such as Kinesin-5 or Myosin VI. By walking along

86 filaments, bridging motors move the filaments relative to each other and change
87 the network. It is however not obvious *a priori* how the sum of their local effects will
88 influence the overall shape and size of the network. A computer can be used to
89 simulate the dynamics of a network, but because all biochemical parameters must
90 be specified in a simulation, only a finite set of conditions can be tested. We present
91 here an analytical theory that overcomes this limitation. Active networks have been
92 previously analyzed (Gowrishankar et al., 2012; Lenz, 2014; Liverpool and
93 Marchetti, 2003; Nedelec et al., 1997; Ziebert et al., 2007) by considering pairs of
94 filaments with one active connector (Fig. 1C). This approach is valid for sparsely
95 connected networks in which only a few motors are active, but physiological
96 networks must be well connected to exert force. In other words, the network should
97 be elastically percolated, and there must exist continuous paths through which
98 tension can be transmitted between any pair of distant points (Dasanayake et al.,
99 2011). Specifically, we assumed that filaments are connected to at least two other
100 filaments of the network. Focusing on one of these filaments (Fig. 1D), we see that
101 the section of the filament between two connectors acts as a mechanical bridge
102 between two points of the network. If the connectors are immobile, or if they both
103 move in the same direction at the same speed, their distance remains constant, the
104 section of the filament between them does not change in length, and the bridge is
105 neutral (Fig. 1E). By contrast, if the two connectors move towards each other, the

106 bridge exerts a contractile stress, whereas if they move apart, this produces an
107 expansile stress (Fig. 1E).

108 To predict if the whole network will contract or expand, we sum up the effects
109 of all mechanical bridges in the network. To do this, we first list all the possible
110 configurations for two connector subunits bound to a filament (shown below for
111 two specific examples). For each configuration, we then calculate da/dt , where a is
112 the distance between the two connectors measured along the filament. This
113 quantity ($v_i = \frac{da}{dt}$ for configuration i) follows from the relative movements of the
114 bound subunits of the connectors. When we sum all the contributions, we take into
115 account the probability p_i of each configuration to occur ($\sum_i p_i v_i$), which can be
116 calculated from the concentrations of components in the system, the binding and
117 unbinding rate of the subunits, and other characteristics of the network (see
118 Supplemental Information). We also distinguish the case where the filaments are
119 rigid and can support expansile stress from the case where the filaments are flexible
120 such that they buckle under compression. In the latter case, filament buckling spoils
121 part or all of the expansile forces (Fig. 1E), and we thus discard the contribution of
122 these expansile configurations. The summation of the weighted contributions of all
123 configurations can then be carried out algebraically, and the sign of the result
124 indicates the predicted network behavior.

125

126 **Actomyosin Networks with Motors and Crosslinkers**

127 To develop the theory, we first applied it to a much studied model of
128 cytoskeletal activity, that of actomyosin contraction, which has also been
129 reconstituted *in vitro* (Carvalho et al., 2013; Kato et al., 1998; Koenderink et al.,
130 2009; Mizuno et al., 2007; Murrell and Gardel, 2012; Reymann et al., 2012; Shah et
131 al., 2014; Takiguchi, 1991; Thoresen et al., 2011; Vogel et al., 2013). Actomyosin
132 networks consist of stabilized F-actin filaments and two types of connectors:
133 bivalent motors moving at speed v and passive crosslinkers (Fig. 2A). The
134 crosslinker is composed of two identical subunits that may bind anywhere on the
135 filaments, and that remain immobile until they detach. There are four possible ways
136 to arrange the two types of connectors on a filament (Fig. 2A). Their likelihood
137 depends on P_M and P_C , the probability of one or more motors and the probability
138 of one or more crosslinkers being bound at an intersection of filaments,
139 respectively. Two of the configurations are neutral in outcome: those with two
140 motors and those with two crosslinkers. The other configurations are active and
141 involve a motor and a crosslinker (Fig. 2A). There are two symmetric configurations
142 with opposite outcomes. In one, the motor and the crosslinker approach each other
143 at speed $-v$, and in the other they move apart at speed v . They have an equal
144 likelihood that is proportional to $P_M P_C (1 - P_C)$, reflecting that one of the crossings
145 should have at least one motor and no crosslinkers, with a likelihood $P_M (1 - P_C)$,
146 while the second crossing should have at least a crosslinker, with or without motor,
147 carrying a likelihood P_C . The net sum over the effects of all configurations in this

148 example is null, and this predicts that a system made of rigid filaments that remain
149 straight should neither contract nor expand. Contractile and expansile
150 configurations cancel each other out, as found previously in the case where only
151 motors were considered (Kruse and Jülicher, 2000). If the filament buckles,
152 however, the expansile configuration will not be able to drive network expansion
153 (Fig. 1E, last panel). Whether a filament buckles depends on the rigidity of the
154 filament, the amount of force generated by the motors, and the distance a between
155 the connectors. Under conditions in which the filaments always buckle, there are no
156 expansile configurations, and the net sum is $-P_M P_C (1 - P_C) v$. In this simple case,
157 the sign reveals that the system is contractile, and this is correctly recapitulated in
158 simulations, for example, when filaments are set to be very flexible (rigidity = 0.01
159 $\text{pN } \mu\text{m}^2$) (Fig. 2BD and Movies 1 to 4).

160

161 **Networks of Semi-Flexible Filaments Contract at Predicted Rates**

162 If the filaments are semi-flexible, which is the case for F-actin, with a rigidity
163 of $0.075 \text{ pN } \mu\text{m}^2$, the contribution of expansile configurations may not always be
164 negligible, since a filament may or may not buckle depending on the length over
165 which it is compressed. Therefore, to be able to predict the behavior of a network,
166 it is necessary to know the conditions under which filaments buckle.

167 For an empirical assessment of this effect, we thus simulated networks in
168 which the length and density of the filaments, and the number of crosslinkers and

169 molecular motors were systematically varied. For this, we used Cytosim, an Open
170 Source simulation engine that is based on Brownian Dynamics (Nedelec and
171 Foethke, 2007). In brief, each filament is represented by a set of equidistant points,
172 subject to bending elasticity (Fig. 3A). Crosslinkers and motors are represented by
173 diffusing point-like particles, which bind stochastically to neighboring filaments
174 (Fig. 3CD). Connectors with a stiffness k are formed when motors or crosslinkers are
175 bound with their two subunits (Fig. 3E). The movement of motors follows a linear
176 force-speed relationship (Fig. 3F). For simplicity, the unbinding rate is constant for
177 this study, and a motor reaching the end of a filament immediately unbinds (Fig.
178 3G). Given a random network as initial condition, Cytosim simulates the movement
179 of all the filaments in the system, and a contraction rate is extracted automatically
180 (Supplemental Information C).

181 Guided by the results of many simulations, we concluded that network
182 contraction depends on the ratio between the mesh size L_1 and the threshold
183 distance d_0 above which buckling occurs, which in turn can be calculated from the
184 filament rigidity and the maximum force exerted by the motor (Supplemental
185 Information D). If $L_1 < d_0$, then any filament segment of length d_0 will be intersected
186 by $\beta_0 = d_0/L_1$ filaments. If any of these intersections is bridged by a crosslinker, this
187 fixes the filament laterally and prevents it from buckling under the force of the
188 motor(s) and crosslinker positioned at its ends. From these considerations, we can
189 calculate the probability for a filament segment to buckle as $P_M P_C (1 - P_C)^{\beta_0}$ (Fig.

190 2C), where $(1 - P_C)^{\beta_0}$ is the probability that none of the intersections between the
191 motor and the crosslinkers are bridged by a crosslinker (Supplemental Information
192 D). The theory predicts a dependence of the contraction rate on the number of
193 connectors for a variety of conditions (Fig. S12–S14). For cases where the density
194 of filaments is low, the contraction rate also depends on the contribution from a
195 mechanism that has been anticipated long ago (Weisenberg and Cianci, 1984). In
196 this configuration (Fig. 7H), a motor acts on an intersection that is already
197 connected by a crosslinker, producing a contractile force by ‘zippering’ two
198 filaments. For a network containing bivalent motors and bivalent crosslinkers, the
199 dependence of contractility on the number of these connectors is thus predictable
200 from first principles.

201

202 **Networks of Rigid Filaments Can Contract or Expand**

203 We next explored systems composed of rigid filaments such as microtubules.
204 Because some molecular motors are associated with microtubule ends in nature,
205 we investigated the behavior induced by connectors comprised of motors and end-
206 binding subunits (Fig. 4). As predicted by the theory (Fig. 4A), the simulations
207 showed that the system is expansile if plus-end directed motors are combined with
208 minus-end binding subunits (Fig. 4B, Movie 5), and contractile if plus-end directed
209 motors are associated with plus-end binding subunits (Fig. 4C, Movie 6). A system
210 composed of these two types of connectors can be either contractile or expansile

211 depending on the relative concentrations of the connectors (Fig. 4DE, Movie 7).
212 This reveals exciting avenues for the development of synthetic materials (Henkin et
213 al., 2014) that could be tuned to be expansile or contractile. Using light-switchable
214 molecular motors (Nakamura et al., 2014) would be particularly exciting in this
215 respect.

216

217 **The Effects of Many Combinations of Connectors is Predicted**

218 To probe the general applicability of the theory, we simulated networks with
219 mixtures of connectors containing 5 different types of subunits (Fig. 1B). A subunit
220 can bind, and then either remain bound at the initial position, or move. Non-moving
221 binders may be of a type that can bind anywhere on the filament, or they may be
222 restricted in their binding to a region near the plus or the minus end. Moving
223 elements (motors) can bind anywhere, but can be of two types, those moving to the
224 plus and those moving to the minus end. By combining any two of these subunits,
225 one can make 15 types of connectors. Simulated networks containing any one type
226 of connector all behaved as predicted by the theory (see examples on Fig. 5A). We
227 also simulated systems containing two different types of connectors (in equal
228 quantities), both for flexible and rigid filaments. There are 210 possible
229 combinations, and for every one of them, the simulations closely matched the
230 prediction of the theory (Fig. 5BC, see Supplemental Information for details of the
231 calculation). Many types of molecular elements that are found in nature, such as

232 end-binding proteins, have not been used in reconstituted networks, but we can
233 now predict what their effects on a network should be.

234

235 **Heterogeneous systems composed of different types of filaments**

236 So far, we have considered systems made of one type of filament, but some
237 networks *in vivo* contain different types of filaments. In particular myosin II motors
238 are organized into thick antiparallel 'mini-filaments' with an average length of 300
239 nm (Verkhovsky and Borisy, 1993). Networks such as the actomyosin meshwork of
240 the cell cortex and the contractile actin cables in cells are thus heterogeneous
241 systems in which F-actin filaments are mixed with mini-filaments, which are also the
242 motors driving the system out of equilibrium. To probe if the theory could hold for
243 such heterogeneous systems, we listed all the possible combinations of two
244 connectors for the two types of filaments (Fig. 6A). Similar to the homogeneous
245 case (Fig. 2) this analysis predicts that the system should be contractile if
246 crosslinkers are also present, and neutral otherwise. We then simulated such a
247 system of actin-like filaments and mini-filaments composed of a rigid backbone of
248 length $0.5\mu\text{m}$ with a motor subunit at each end. The results confirmed the predicted
249 behaviors (Fig. 6B and 6C), suggesting that the theory can be applied to
250 heterogeneous networks. It is tempting to think that the approach can also be
251 extended to anisotropic networks if the probabilities of the configurations are
252 calculated locally.

253

254 **Contractile Systems Pulse if Filaments Turn Over**

255 So far, we have considered systems made of filaments of fixed length that
256 persist infinitely. Under these conditions, network contraction and expansion are
257 non-reversible events, that only occur once. This is indeed what happens with most
258 *in vitro* reconstituted cytoskeletal networks, obtained with stabilized filaments
259 (Carvalho et al., 2013; Foster et al., 2015; Katoh et al., 1998; Murrell and Gardel,
260 2012; Shah et al., 2014; Surrey et al., 2001; Takiguchi, 1991; Thoresen et al., 2011;
261 Vogel et al., 2013). But how does this relate to networks *in vivo*, which manage to
262 avoid such a collapse? The simulations described above do not perfectly mimic the
263 situation *in vivo*, since cytoskeletal filaments elongate by self-assembly and if they
264 remain dynamic will eventually vanish, such that both the length of the filaments,
265 and their abundance are fluctuating quantities that can be regulated. In some cells,
266 actomyosin networks persist for several minutes, and exhibit pulsatile behavior that
267 is reminiscent of contractility (He et al., 2010; Martin et al., 2009; Munro et al., 2004;
268 Solon et al., 2009). To test the relationship between this dynamic behavior and
269 contractility, we extended our simulations to include filament turnover. Specifically,
270 to implement an average lifetime T for the filament, we randomly selected and
271 deleted one of the N filaments at a rate N/T , and replaced it with a new one placed
272 at a random position (Fig. 6D). We typically observed that within $3s < T < 200s$, a
273 configuration that was contractile without turnover now displayed pulsed

274 contractions (Fig. 6E, Movie 8). Note that these new simulations used periodic
275 boundary conditions (PBC), because if the system is allowed to contract freely,
276 pulses cannot be observed. Use of PBC imposes a constant surface on the system
277 and thereby forces the network to build up tension. It corresponds best to a network
278 that is attached at the cell boundaries, without requiring additional assumptions on
279 the nature of the attachment. These results confirm earlier models that considered
280 filament dynamics (Bidone et al., 2017) or turnover (Hiraiwa and Salbreux, 2016;
281 Kumar et al., 2014), illustrating that with filament turnover, a system that was
282 contractile otherwise can be pulsatile. As suggested by a coarse-grained model
283 (Kumar et al., 2014), we wondered if pulsatility was a general consequence of
284 turnover. We thus simulated networks with all the combinations of connectors that
285 were contractile on Fig. 5B and varied systematically the filament turnover rate. All
286 displayed pulsatile behavior, confirming the universal nature of the phenomenon
287 (data not shown).

288

289 **DISCUSSION**

290 The theory we present here predicts the initial evolution of a network from
291 the properties of its connectors. We have confirmed these predictions with
292 simulations for all tested conditions. The model implemented in the simulation is
293 intentionally minimalistic, as subunit binding, unbinding and filament turnover
294 occurred at constant rates and independently of other events. All simulations where

295 done in 2D, and did not consider steric interactions between the filaments, which
296 in 2D would induce artifacts. We expect our theoretical arguments to hold also for
297 other types of networks such as filament bundles or 3D networks. However, the
298 calculation presented in the supplemental information depends on the geometry
299 of the network, and would need to be revised to apply to these different conditions.
300 Our analytical prediction of network behavior was based on the characteristics of
301 the connectors but did not include physical parameters such as the viscosity of the
302 medium. Rather than an absolute contraction rate, this theory predicts the relative
303 contraction rates of two networks when the parameters of the connectors (numbers,
304 types, binding rates, unbinding rates) are different between them. Such a
305 prediction is immediately valuable, as it can be readily tested experimentally by
306 systematically varying the concentration of both motors and crosslinkers in
307 reconstituted *in vitro* networks.

308 For a system containing crosslinkers and bivalent motors (Fig. 2), our analysis
309 indicates that the 'active' contractile configurations involve both a crosslinker and a
310 motor. We thus expect these two types of activities always to be found in a
311 contractile system. However, we note that in an *in vitro* experimental system, the
312 assumedly pure preparation of motors that is added may in fact contain damaged
313 'dead' motor proteins that act as passive connectors. Nevertheless, addition of
314 crosslinkers indeed dramatically enhances the effect of myosin, a phenomenon
315 observed more than 50 years ago (Ebashi and Ebashi, 1964). For 2D networks (Fig.

316 2CD and Supplemental Information), we could explain why the maximum
317 contractility is obtained *in vitro* with approximately equal amounts of motors and
318 crosslinkers (Bendix et al., 2008; Ennomani et al., 2016; Köhler and Bausch, 2012).
319 Our theory also explains that under the action of myosin VI, a branched network
320 made with Arp2/3, which represents an example of a connector combining end-
321 binding and side-binding activities together, is more contractile than a network
322 connected by crosslinkers binding anywhere along the filaments (Ennomani et al.,
323 2016). This is because, as myosin VI is directed to the pointed end, the configuration
324 involving a crosslinker bound to the pointed end (Arp2/3) is always contractile.
325 Thus, at equal levels of connectivity, a network made with Arp2/3 is more
326 contractile than a network made with a non-specific crosslinker, and less contractile
327 than a network made with only end-to-end crosslinkers.

328 Because we focused on the initial behavior of a network, it seemed justified
329 to ignore mechanisms that regulate the characteristics of the cell cortex, for instance
330 its thickness. We used however simulations to explore the effects of filament
331 turnover (Fig. 6DE). The fact that introducing filament turnover was sufficient to
332 induce pulsing in all the contractile scenarios leads to the surprising conclusion that
333 pulsatility is an intrinsic behavior of contractile networks of non-stable filaments,
334 and that no other external elements are necessary. This of course does not mean
335 that in the natural biological situation there may not be regulatory elements
336 superimposed on the underlying mechanism that suppress or enhance pulsing

337 (Nishikawa et al., 2017). Pulsing is seen only over a certain range of filament
338 lifetimes (Fig. 6D), indicating that one such regulatory input could be via the
339 stability of filaments: for example, stabilizing filaments should, according to our
340 simulations, arrest pulsing, and the ability of myosin to destroy filaments (Matsui et
341 al., 2011) could on the contrary enhance pulsing. The characteristics of the pulses
342 can be further tuned by modulating the other characteristics of the network
343 components, as has also been seen *in vivo* for the regulation of myosin (Munjal et
344 al., 2015). Pulses may be a desired feature or an inevitable consequence of having
345 filament turnover when the connectors lead to contractility. This important feature
346 of *in vivo* cytoskeletal networks deserves more investigation in the future.

347 Importantly, our theory unifies ideas previously proposed for various
348 biological systems, and we will discuss now how various mechanisms giving rise to
349 contraction or extension fit within the new theory (Fig. 7).

350 In the sarcomeres found in striated muscles, myosin II mini-filaments pull on
351 filaments arranged in an anti-parallel manner (Fig. 7A). This system can be seen as
352 containing two types of connectors: a passive one linking the barbed ends of the
353 filament, and a motor directed towards the barbed end. Three possible
354 combinations can be made with these two connectors (Fig. 7, right column). In the
355 absence of any expansile configuration, the system is bound to always be
356 contractile. A sarcomeric system is highly ordered, but less organized systems
357 made of the same subunits, for example bipolar filaments that are discussed for

358 smooth muscles (Fig. 7B) or a fully disorganized system (Fig. 5A, fourth column) are
359 also contractile.

360 For a system in which the crosslinkers are not restricted to binding to the
361 filament ends, but can bind anywhere along the length (Fig. 7C) both contractile
362 and expansile configurations arise. Following the discussion on how buckling
363 promotes contraction of a disorganized actin network (Liverpool et al., 2009;
364 Mizuno et al., 2007) we argued that buckling can spoil some of the expansile
365 configurations, tipping the balance in favor of contraction.

366 One popular mechanism to explain the contraction of microtubule networks
367 (Fig. 7D) does not require filament bending, but involves a motor that has an affinity
368 for the end of the filaments (Hyman and Karsenti, 1996; Nedelec et al., 1997).
369 Because the motor walks towards the end, where it may be transiently trapped,
370 configurations are contractile or neutral, but never expansile, and the entire
371 network itself is therefore contractile (Foster et al., 2015). Looking at the set of
372 configurations (Fig. 7, right column), the similarity of this mechanism with
373 sarcomeric contractility (Fig. 7AB) is noticeable. In the case of the end-dwelling
374 motor, however, the same molecular type is involved in generating the active and
375 neutral end-binding connections.

376 Although we did not consider filament disassembly in this study, we would
377 suggest that the theory can be applied also to this situation. For example, a
378 molecule that tracks and remains bound to the depolymerizing end of a filament

379 (Fig. 7E) will reduce the distance between itself and a connector located elsewhere
380 on the filament, thereby creating a pulling force. By calculating the likelihood of
381 such a configuration, one may be able to predict the overall contractility of the
382 network. We also did not consider filament elongation, which is a prominent
383 mechanism by which actin networks expand, but identify conditions where, even
384 without this mechanism, a network can increase its surface area.

385 A system with expansile configurations will extend if the filaments are
386 sufficiently rigid to resist buckling, which is more likely to be the case for
387 microtubules than for actin. A mitotic spindle is a complicated structure, which can
388 be found in multiple states. In most cells metaphase corresponds to a steady state
389 in which contractile contributions must on average compensate expansile ones. In
390 *Xenopus laevis*, contraction is driven by the motor minus-end directed Dynein
391 (Foster et al., 2015) while expansion is driven by the plus-end motor Kinesin5
392 (Needleman and Brugués, 2014). When anaphase is induced, this balance is broken
393 leading to spindle elongation, and chromosome segregation. For the sake of the
394 argument, we consider here that the spindle at anaphase is purely driven by
395 kinesin5, and thus assume only two types of connectors (Fig. 7F): passive
396 complexes containing the protein NuMA, which connect the minus-ends of
397 microtubules at the spindle poles, and plus-end directed motors connecting
398 adjacent, antiparallel microtubules. Since Kinesin5 walks away from the minus-
399 ends, the configurations are symmetric to the sarcomeric systems (Fig. 7A), and

400 expand the anaphase spindle. A disorganized network made of the same
401 connectors is also expansile (Fig. 4B).

402 Additional expansile microtubule systems can be found in blood platelets
403 and their progenitor cells, the megakaryocytes. During pro-platelet generation, the
404 microtubules assemble into bundles that elongate, and this elongation is
405 dependent on the activity of the molecular motors Dynein (Patel, 2005). In the
406 mature platelets, microtubules are organized into a closed circular bundle, and this
407 bundle must be able to resist contractile forces as it pushes outward on the plasma
408 membrane. It was recently reported that the microtubule ring elongates after
409 platelet activation, in a manner that is dependent on microtubules motors, but we
410 lacked until now a microscopic picture of the elongation mechanism (Diagouraga
411 et al., 2014). Our systematic exploration of random networks (Fig. 5) suggests
412 different scenarios that could explain why the system is expansile. Beyond the
413 relevance to these *in vivo* systems, it will be exciting to follow these principles to
414 create expansile networks of microtubules *in vitro*, since end-binders are available
415 to synthetic biologists.

416 Finally, in a system where the symmetry provides an equal number of
417 contractile and expansile configurations, any imbalance in the probabilities of these
418 configurations may lead to overall contraction or expansion (Gao et al., 2015).
419 Following this principle, we can suggest here an explanation for the expansile
420 nature of *in vitro* networks (Sanchez et al., 2012). Particularly, if the motors are

421 sufficiently processive, they may run a distance that is comparable to the length of
422 the filament, and in this case their distribution along the length of the filament will
423 be non-uniform (Fig. 7G). This effect has been called the antenna effect (Varga et
424 al., 2006), and arises as a consequence of the motility, in a situation where binding
425 has the same probability at every position of the filament. A plus-end directed
426 motor would accumulate towards the plus ends of microtubules (Fig. 7G). In the
427 presence of crosslinkers, such an effect will increase the likelihood of the expansile
428 configurations, and lower the likelihood of the contractile configurations, thus
429 promoting expansion. It is interesting to note that even if the motors were directed
430 towards the minus-end of microtubules, the antenna effect would still lead to a bias
431 in favor of expansion. The net imbalance will depend on the biophysical properties
432 of the motors (speed, unbinding rate), and the length of the microtubules, and
433 could provide tunable expansibility for networks made *in vivo* with microtubules
434 (Sanchez et al., 2012).

435 In conclusion, our theory offers a framework for elementary mechanisms of
436 expansion or contraction. It is a starting point for further exploration, since in its
437 current state, the theory does not explain all the phenomena observed in
438 simulations. In particular, for very flexible filaments a configuration involving two
439 connectors starting from the same point of contact between two filaments seems to
440 contribute to contraction (Fig. 7H). In this case, the connectors can pull the network
441 together, even though the distance between them is growing. This interesting

442 effect, which is analogous to a zipper, can only be understood by considering two
443 filaments and two connectors, whereas our theory considered one filament and two
444 connectors. Because this effect only contributes little in most cases (see
445 Supplemental Information D5), the theory is still able to make accurate predictions,
446 even for complex systems (Fig. 5BC). Mathematically, the theory could be
447 understood as providing a first-order approximation of the exact contractile rate.

448 From the theoretical framework presented here, with its clear predictions,
449 perhaps a classification of the different types of active networks found in nature will
450 emerge. Our approach may also inspire novel avenues for synthetic filament
451 networks with enhanced functionalities.

452

453 **AUTHOR CONTRIBUTIONS**

454 This work arose from regular discussions between the authors, which all have
455 contributed significantly to the findings. All authors together wrote the article.

456

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465

466 REFERENCES

- 467 Backouche, F., Haviv, L., Groswasser, D., and Bernheim-Groswasser, A. (2006).
468 Active gels: dynamics of patterning and self-organization. *Phys Biol* 3, 264-273.
- 469 Bendix, P.M., Koenderink, G.H., Cuvelier, D., Dogic, Z., Koeleman, B.N., Briehar,
470 W.M., Field, C.M., Mahadevan, L., and Weitz, D.A. (2008). A quantitative analysis of
471 contractility in active cytoskeletal protein networks. *Biophys J* 94, 3126-3136.
- 472 Bidone, T.C., Jung, W., Maruri, D., Borau, C., Kamm, R.D., and Kim, T. (2017).
473 Morphological Transformation and Force Generation of Active Cytoskeletal
474 Networks. *PLoS Comput. Biol.* 13, e1005277.
- 475 Carlsson, A.E. (2006). Contractile stress generation by actomyosin gels. *Phys Rev E*
476 *Stat Nonlin Soft Matter Phys* 74, 051912.
- 477 Carvalho, K., Tsai, F.-C., Tsai, F.C., Lees, E., Voituriez, R., Koenderink, G.H., and
478 Sykes, C. (2013). Cell-sized liposomes reveal how actomyosin cortical tension
479 drives shape change. *Proc Natl Acad Sci USA* 110, 16456-16461.
- 480 Dasanayake, N.L., Michalski, P.J., and Carlsson, A.E. (2011). General mechanism of
481 actomyosin contractility. *Phys Rev Lett* 107, 118101.
- 482 Diagouraga, B., Grichine, A., Fertin, A., Wang, J., Khochbin, S., and Sadoul, K.
483 (2014). Motor-driven marginal band coiling promotes cell shape change during
484 platelet activation. *J Cell Biol* 204, 177-185.
- 485 Ebashi, S., and Ebashi, F. (1964). A New Protein Factor Promoting Contraction of
486 Actomyosin. *Nature* 203, 645-646.
- 487 Ennomani, H., Letort, G., Guérin, C., Martiel, J.-L., Cao, W., Nedelec, F., La Cruz,
488 De, E.M., Théry, M., and Blanchoin, L. (2016). Architecture and Connectivity Govern
489 Actin Network Contractility. *Curr Biol* 26, 616-626.
- 490 Foster, P.J., Fürthauer, S., Shelley, M.J., and Needleman, D.J. (2015). Active
491 contraction of microtubule networks. *Elife* 4.
- 492 Gao, T., Blackwell, R., Glaser, M.A., Betterton, M.D., and Shelley, M.J. (2015).
493 Multiscale modeling and simulation of microtubule-motor-protein assemblies.

- 494 Phys Rev E Stat Nonlin Soft Matter Phys 92, 062709.
- 495 Gowrishankar, K., Ghosh, S., Saha, S., C, R., Mayor, S., and Rao, M. (2012). Active
496 remodeling of cortical actin regulates spatiotemporal organization of cell surface
497 molecules. *Cell* 149, 1353-1367.
- 498 He, L., Wang, X., Tang, H.L., and Montell, D.J. (2010). Tissue elongation requires
499 oscillating contractions of a basal actomyosin network. *Nat Cell Biol* 12, 1133-
500 1142.
- 501 Henkin, G., DeCamp, S.J., Chen, D.T.N., Sanchez, T., and Dogic, Z. (2014). Tunable
502 dynamics of microtubule-based active isotropic gels. *Philos Trans a Math Phys Eng*
503 *Sci* 372.
- 504 Hiraiwa, T., and Salbreux, G. (2016). Role of Turnover in Active Stress Generation in
505 a Filament Network. *Phys Rev Lett* 116, 188101.
- 506 Hyman, A.A., and Karsenti, E. (1996). Morphogenetic properties of microtubules
507 and mitotic spindle assembly. *Cell* 84, 401-410.
- 508 Katoh, K., Kano, Y., Masuda, M., Onishi, H., and Fujiwara, K. (1998). Isolation and
509 contraction of the stress fiber. *Mol Biol Cell* 9, 1919-1938.
- 510 Koenderink, G.H., Dogic, Z., Nakamura, F., Bendix, P.M., MacKintosh, F.C., Hartwig,
511 J.H., Stossel, T.P., and Weitz, D.A. (2009). An active biopolymer network controlled
512 by molecular motors. *Proc Natl Acad Sci USA* 106, 15192-15197.
- 513 Köhler, S., and Bausch, A.R. (2012). Contraction mechanisms in composite active
514 actin networks. *PLoS ONE* 7, e39869.
- 515 Köhler, S., Schaller, V., and Bausch, A.R. (2011). Structure formation in active
516 networks. *Nat Mater* 10, 462-468.
- 517 Kruse, K., and Jülicher, F. (2000). Actively contracting bundles of polar filaments.
518 *Phys Rev Lett* 85, 1778-1781.
- 519 Kumar, K.V., Bois, J.S., Jülicher, F., and Grill, S.W. (2014). *Phys. Rev. Lett.* 112,
520 208101 (2014) - Pulsatory Patterns in Active Fluids. *Physical Review Letters*.
- 521 Lenz, M., and Gardel, M. (2012). Requirements for contractility in disordered
522 cytoskeletal bundles - Abstract - *New Journal of Physics* - IOPscience. *New Journal*
523 *of Physics*.
- 524 Lenz, M. (2014). Geometrical Origins of Contractility in Disordered Actomyosin
525 Networks. *Phys. Rev. X* 4, 041002.

- 526 Liverpool, T.B., Marchetti, M.C., Joanny, J.F., and Prost, J. (2009). Mechanical
527 response of active gels. *Epl* 85, 18007.
- 528 Liverpool, T.B., and Marchetti, M.C. (2003). Instabilities of isotropic solutions of
529 active polar filaments. *Phys Rev Lett* 90, 138102.
- 530 Martin, A.C., Kaschube, M., and Wieschaus, E.F. (2009). Pulsed contractions of an
531 actin-myosin network drive apical constriction. *Nature* 457, 495-499.
- 532 Matsui, T.S., Kaunas, R., Kanzaki, M., Sato, M., and Deguchi, S. (2011). Non-muscle
533 myosin II induces disassembly of actin stress fibres independently of myosin light
534 chain dephosphorylation. *Interface Focus* 1, 754-766.
- 535 Mendes Pinto, I., Rubinstein, B., Kuchavy, A., Unruh, J.R., and Li, R. (2012). Actin
536 depolymerization drives actomyosin ring contraction during budding yeast
537 cytokinesis. *Dev Cell* 22, 1247-1260.
- 538 Mizuno, D., Tardin, C., Schmidt, C.F., and Mackintosh, F.C. (2007). Nonequilibrium
539 mechanics of active cytoskeletal networks. *Science* 315, 370-373.
- 540 Munjal, A., Philippe, J.-M., Munro, E., and Lecuit, T. (2015). A self-organized
541 biomechanical network drives shape changes during tissue morphogenesis.
542 *Nature* 524, 351-355.
- 543 Munro, E., Nance, J., and Priess, J.R. (2004). Cortical flows powered by
544 asymmetrical contraction transport PAR proteins to establish and maintain
545 anterior-posterior polarity in the early *C. elegans* embryo. *Dev Cell* 7, 413-424.
- 546 Murrell, M.P., and Gardel, M.L. (2012). F-actin buckling coordinates contractility
547 and severing in a biomimetic actomyosin cortex. *Proc Natl Acad Sci USA* 109,
548 20820-20825.
- 549 Murrell, M., Oakes, P.W., Lenz, M., and Gardel, M.L. (2015). Forcing cells into
550 shape: the mechanics of actomyosin contractility. *Nat Rev Mol Cell Biol*.
- 551 Nakamura, M., Chen, L., Howes, S.C., Schindler, T.D., Nogales, E., and Bryant, Z.
552 (2014). Remote control of myosin and kinesin motors using light-activated
553 gearshifting. *Nature Nanotechnology* 9, 693-697.
- 554 Nedelec, F.J., Surrey, T., Maggs, A.C., and Leibler, S. (1997). Self-organization of
555 microtubules and motors. *Nature* 389, 305-308.
- 556 Nedelec, F., and Foethke, D. (2007). Collective Langevin dynamics of flexible
557 cytoskeletal fibers. *New Journal of Physics* 9, 499-510.
- 558 Needleman, D., and Brugués, J. (2014). Determining physical principles of

- 559 subcellular organization. *Dev Cell* 29, 135-138.
- 560 Nishikawa, M., Naganathan, S.R., Jülicher, F., and Grill, S.W. (2017). Controlling
561 contractile instabilities in the actomyosin cortex. *Elife* 6.
- 562 Oelz, D.B., Rubinstein, B.Y., and Mogilner, A. (2015). A Combination of Actin
563 Treadmilling and Cross-Linking Drives Contraction of Random Actomyosin Arrays.
564 *Biophys J* 109, 1818-1829.
- 565 Patel, S.R. (2005). Differential roles of microtubule assembly and sliding in
566 proplatelet formation by megakaryocytes. *Blood* 106, 4076-4085.
- 567 Prost, J., Jülicher, F., and Joanny, J.F. (2015). Active gel physics. *Nat Phys* 11, 111-
568 117.
- 569 Reymann, A.-C., Boujemaa-Paterski, R., Martiel, J.-L., Guérin, C., Cao, W., Chin,
570 H.F., La Cruz, De, E.M., Théry, M., and Blanchoin, L. (2012). Actin network
571 architecture can determine myosin motor activity. *Science* 336, 1310-1314.
- 572 Salbreux, G., Charras, G., and Paluch, E. (2012). Actin cortex mechanics and
573 cellular morphogenesis. *Trends in Cell Biology* 22, 536-545.
- 574 Sanchez, T., Welch, D., Nicastro, D., and Dogic, Z. (2011). Cilia-Like Beating of
575 Active Microtubule Bundles. *Science* 333, 456-459.
- 576 Sanchez, T., Chen, D.T.N., DeCamp, S.J., Heymann, M., and Dogic, Z. (2012).
577 Spontaneous motion in hierarchically assembled active matter. *Nature* 491, 431-
578 434.
- 579 Shah, E.A., Keren, K., and Balasubramanian, M. (2014). Symmetry breaking in
580 reconstituted actin cortices. *Elife* 3.
- 581 Solon, J., Kaya-Copur, A., Colombelli, J., and Brunner, D. (2009). Pulsed forces
582 timed by a ratchet-like mechanism drive directed tissue movement during dorsal
583 closure. *Cell* 137, 1331-1342.
- 584 Surrey, T., Nedelec, F., Leibler, S., and Karsenti, E. (2001). Physical properties
585 determining self-organization of motors and microtubules. *Science* 292, 1167-
586 1171.
- 587 Takiguchi, K. (1991). Heavy meromyosin induces sliding movements between
588 antiparallel actin filaments. *J Biochem* 109, 520-527.
- 589 Thoresen, T., Lenz, M., and Gardel, M.L. (2011). Reconstitution of Contractile
590 Actomyosin Bundles. *Biophysical Journal* 100, 2698-2705.

- 591 Varga, V., Helenius, J., Tanaka, K., Hyman, A.A., Tanaka, T.U., and Howard, J.
592 (2006). Yeast kinesin-8 depolymerizes microtubules in a length-dependent
593 manner. *Nat Cell Biol* 8, 957–962.
- 594 Verkhovsky, A.B., and Borisy, G.G. (1993). Non-sarcomeric mode of myosin II
595 organization in the fibroblast lamellum. *J Cell Biol* 123, 637–652.
- 596 Vogel, S.K., Petrasek, Z., Heinemann, F., and Schwille, P. (2013). Myosin motors
597 fragment and compact membrane-bound actin filaments. *Elife* 2, e00116.
- 598 Weisenberg, R.C., and Cianci, C. (1984). ATP-induced gelation--contraction of
599 microtubules assembled in vitro. *J Cell Biol* 99, 1527–1533.
- 600 Wolff, L., and Kroy, K. (2012). Minimal model for the inelastic mechanics of
601 biopolymer networks and cells. *Phys Rev E Stat Nonlin Soft Matter Phys* 86,
602 040901.
- 603 Ziebert, F., Vershinin, M., Gross, S.P., and Aranson, I.S. (2009). Collective alignment
604 of polar filaments by molecular motors. *Eur Phys J E Soft Matter* 28, 401–409.
- 605 Ziebert, F., Aranson, I.S., and Tsimring, L.S. (2007). Effects of cross-links on motor-
606 mediated filament organization. *New Journal of Physics* 9, 421–421.

607

608 **Figure 1: Elements of the theory**

609 **(A)** Networks are composed of polar filaments that may bend, and connectors
610 containing two subunits through which they can bridge two nearby filaments.

611 **(B)** Subunits may be minus-end or plus-end directed motors that can bind
612 anywhere to a filament, binders that can bind to any location along a filament, or
613 end-binders that attach only at the minus or the plus ends of filaments.

614 **(C, D)** To predict the behavior of a network, previous theories have considered a
615 pair of filaments with a single connector between them, while the theory
616 considered here is based on the effects that connectors bound to a single filament
617 have on the rest of the network.

618 (E) Pairs of connectors may generate local stress in the network depending on
619 how the subunits move relative to one another on the filament. If the initial
620 distance a_0 between the subunits is maintained, the network does not deform.
621 Local contraction is expected if the connectors move towards each other ($a < a_0$)
622 and expansion may occur if they move apart ($a > a_0$). If the filament is flexible,
623 however, the expansile stress can be reduced if the filament buckles.

624

625 **Figure 2: Predictions and simulations for actin-like networks of flexible**
626 **filaments.**

627 (A) A system composed of flexible filaments and two types of connectors:
628 crosslinkers and bivalent motors. The table lists the four possible configurations
629 for two connectors bound to a filament, the relative movement of the connectors
630 ($\frac{da}{dt}$), the likelihood, and the mechanical nature of each configuration. The
631 likelihoods are combinations of P_M and P_C , i.e. the probabilities of having a motor
632 or a crosslinker at an intersection of two filaments (see Supplemental Information
633 D3).

634 (B) The evolution of a simulated random network composed of 1500 flexible
635 filaments (bending rigidity = $0.01 \text{ pN } \mu\text{m}^2$) and 12000 connectors of each type,
636 distributed over a circular area of radius $15 \mu\text{m}$.

637 (C) The contraction rate of a simulated network as a function of the ratio of
638 crosslinkers to motors, with the total number of connectors kept constant. Each

639 symbol indicates the result of one simulation. The broken line indicates the
640 analytical prediction made by the theory (Supplemental Information D).

641 **(D)** Snapshots at $t=15s$ of networks similar to **(B)** containing varying numbers of
642 motors (vertical axis) and crosslinkers (horizontal axis). No contraction occurs
643 without crosslinkers or without motors, and the optimal contractile rate is
644 obtained here for 8000 motors and 10000 crosslinkers. The location of the
645 optimum can be predicted from the molecular properties of the connectors
646 (Supplemental Information D).

647 **Figure 3: Elements of the stochastic model of cytoskeletal dynamics.**

648 **(A)** Networks of flexible filaments are simulated using a Brownian dynamics
649 method. In brief, filaments are polar, thin and have a constant length. Each
650 filament is modeled with an oriented string of points, defining segments of equal
651 lengths. The movement of filament points follows Brownian dynamics, with elastic
652 forces such as the bending elasticity of the filament, and the elasticity of
653 connectors.

654 **(B)** In the simulations, connecting molecules are made of two independent
655 filament-binding subunits (a and b, which can be any one of those defined in Fig.
656 1B). When both subunits are unattached to filaments, the molecule diffuses within
657 the simulation space.

658 **(C)** Binding occurs at a constant rate k_{on} to any filament closer to than ϵ .
659 Attachment occurs on the closest point of the filament.

660 **(D)** End-binding follows the same rules as binding, but is restricted to a distance δ
661 from the targeted filament end.

662 **(E)** Connectors act mechanically as Hookean springs between two filaments, with
663 stiffness k and zero resting length.

664 **(F)** Motor subunits move towards either the plus- or minus-end of the filament with
665 a linear force-velocity relationship.

666 **(G)** All connector subunits detach with a force-independent rate k_{off} , and motors
667 detach immediately upon reaching a filament end.

668

669 **Figure 4: Predictions and simulations for microtubule-like networks of rigid**
670 **filaments.**

671 **(A)** A system composed of rigid filaments and two types of connectors. One
672 connector consists of a plus-end directed motor combined with a minus-end
673 binder, the other is a plus-end directed motor combined with a plus-end binder.
674 There are four possible configurations involving these two connectors.

675 **(B)** Three time points on the evolution of an expansile network of 1500 straight
676 filaments (their bending rigidity is set as "infinite" here) with 1500 motor-plus-end
677 binders and 48000 motor-minus-end binders initially distributed over a circular
678 area of radius $15\mu\text{m}$.

679 **(C)** Three time points on the evolution of a network similar as **(B)**, but with 48000
680 motor-plus-end binders and 1500 motor-minus-end binders.

681 **(D)** The contraction rate of a network as a function of the number of connectors,
682 which are inversely varied. Each symbol represents a simulated random network
683 of 4000 straight filaments initially distributed over a circular area of radius 25 μ m.
684 Details of methods as in Fig. 2C. The broken line indicates the analytical
685 prediction made by the theory (Supplemental Information F).

686

687 **(E)** Simulations of networks containing varying numbers of connectors. Networks
688 contain 1500 filaments initially distributed over a radius of 15 μ m. Depending on
689 the concentrations of the connectors, the network can be expansile (top left
690 corner) or contractile (bottom right corner). Snapshots at t=30s.

691

692 **Figure 5: Additional predictions of the theory**

693 The predictions of the theory for the various conditions shown here are
694 represented graphically as sets of green centripetal arrows for contraction, red
695 centrifugal arrows for expansion, and grey squares for neutral networks.

696 **(A)** Examples of simulations of networks with the indicated types of connectors.

697 The predicted outcomes of network contraction, expansion or neutrality (symbol
698 at the top left of each simulation) are confirmed by the behavior of the network in
699 simulations in each case. The networks are composed of 1500 flexible or rigid
700 filaments, and 24000 connectors. Snapshots at t=20s.

701 **(B)** Summary of the predictions for random networks with all possible
702 combinations of all possible types of connectors, either with flexible (bending
703 rigidity = $0.01 \text{ pN } \mu\text{m}^2$, below diagonal) or rigid filaments (straight filaments,
704 above diagonal). The networks contain 4000 filaments and 64000 connectors at a
705 1:1 ratio. The labels of the rows and columns of the table indicate the type of the
706 two connectors that are mixed.

707 **(C)** Comparison of the contraction rates predicted by the theory (horizontal axis)
708 with the rates obtained by simulation (vertical axis). Each data point indicates one
709 of the 210 systems considered in **(B)**. Networks are made of 4000 filaments and
710 64000 connectors initially distributed over a circular area of radius $25\mu\text{m}$. In this
711 case, all the binding parameters of the subunits and the concentration of
712 connectors are always equal, such that the prediction is simplified (Supplemental
713 Information E).

714

715 **Figure 6: Heterogeneous and pulsatile systems**

716 **(A)** Configurations present in a heterogeneous network containing rigid
717 minifilaments and flexible actin-like filaments. The motors are permanently
718 attached at the extremities of the minifilaments, such as to represent myosin
719 minifilaments. The system is predicted to be contractile in the presence of passive
720 crosslinkers connecting actin filaments directly, and neutral without crosslinkers.

721 **(B)** Detail of a simulation with minifilaments (green) and crosslinkers (blue).

722 (C, D) The simulated systems contract with crosslinkers, but not when they are
723 omitted.

724 (E) Time series of a simulation with filament turnover, 1400 filaments (rigidity
725 $0.075 \text{ pN } \mu\text{m}^2$), 22400 motors, 5600 crosslinkers within periodic boundary
726 conditions with size $16 \mu\text{m}$. Filament turnover was implemented by deleting a
727 randomly selected filament and placing a new filament at a random location,
728 stochastically with a rate $R=64\text{s}^{-1}$, corresponding to an average filament lifetime of
729 $\sim 21.8\text{s}$. The series shows the formation of a new contractile spot at the right side
730 (black arrowhead) and its downward movement and fusion with the contractile
731 spot at the lower right corner of the panels (green arrowheads).

732 (F) The local density of filaments in an arbitrarily chosen region covering $\sim 6\%$ of
733 the simulated space as a function of time. The data with filament lifetime 21.8s are
734 from the simulation shown in (A). The network continues to redistribute, showing
735 irregular variations of the local filament density, and does not contract into one
736 spot.

737

738 **Figure 7: Review of contractile and expansile mechanisms**

739 Previously described mechanisms can be considered by focusing on pairs of
740 connectors present on filaments. The sarcomeric (A) and the analogous
741 mechanism involving bipolar filaments (B) have a barbed-end directed multivalent
742 motor acting on filaments that are connected at their pointed ends by molecular

743 complexes that act as connectors. These systems are always contractile because
744 there are only two active configurations: one involving two motors, which is
745 neutral, and one with a motor and an end-binder, which is contractile since the
746 motor always moves towards the end binder.

747 The buckling-dependent mechanism (C) leads to contractility because the
748 flexibility of the filament spoils the expansile configuration. Thus, if the filaments
749 are sufficiently flexible, the net effect will be contractile (see Fig. 2).

750 In systems containing only end-dwelling multivalent motors (D), the motors
751 generate contraction without added passive connectors, because they eventually
752 come to a halt at the end of the filament and thereby act as end-binding
753 connectors. Configurations involving a motor halted at the end, and a motor
754 moving towards this end along the same filament result in contraction. Note that
755 since in this mechanism no configuration is expansile, and the net effect is always
756 contractile, irrespective of filament buckling.

757 A connector with a subunit that binds to a disassembling end of a filament (E)
758 generates only one active configuration which is always contractile, even in the
759 absence of motors. In this example, the end-tracker binds to the barbed end, and
760 moves towards the pointed end by tracking a depolymerizing end (or inducing its
761 disassembly).

762 (F) A mitotic spindle at anaphase may be considered as a network held together
763 by multivalent plus-end directed motors from the Kinesin5 family, and by factors

764 connecting the microtubule minus ends at the spindle pole. With these two types
765 of connectors, the configurations involving two connectors are neutral, passive, or
766 expansile.

767 **(G)** A system can be made expansile by the “antenna effect”, because motors
768 acquire an asymmetric distribution profile along the filaments. In the presence of
769 this effect, contractile configurations are less likely than expansile ones, and the
770 overall system can become expansile as a consequence.

771 **(H)** Some mechanisms of contraction involve two connectors acting on more than
772 one filaments. In the case depicted here, two crossing filaments will be “zippered
773 together”, by a pair of connectors moving apart. This configuration is able to
774 create a contractile force dipole in the direction perpendicular to the filaments.

775

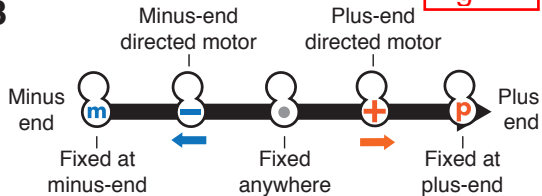


Polar filament
(e.g. actin, microtubule)

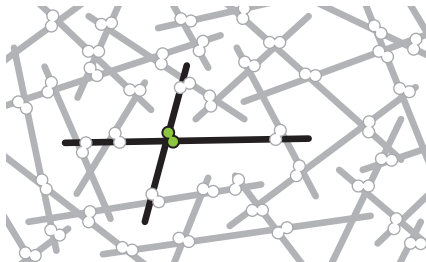


Connectors
(e.g. myosins, kinesins, dyneins
cross-linkers, Arp2/3)

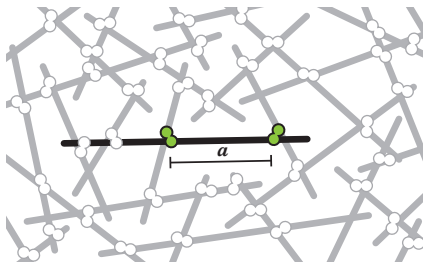
B



C



D



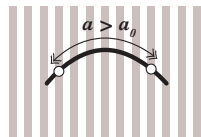
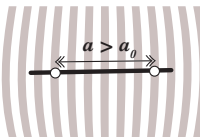
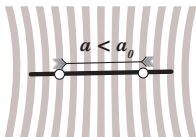
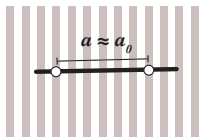
E

Neutral





Contraction

Expansion
rigid filament

Spoiled expansion
flexible filament



A Components:  +  + flexible filaments

Configuration	da/dt	Probability	Effect
	0	P_c^2	static
	0	$P_M^2 (1 - P_c)^2$	neutral
	-v	$P_M (1 - P_c) P_c$	pulling
	+v	$P_M (1 - P_c) P_c$	(buckling)

Net outcome: contraction

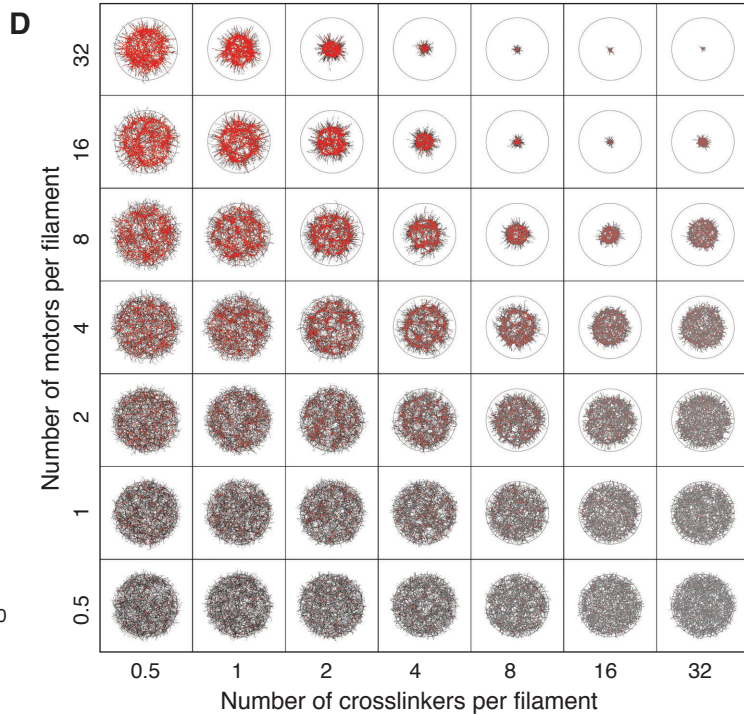
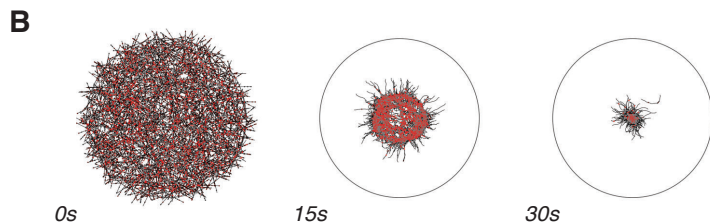
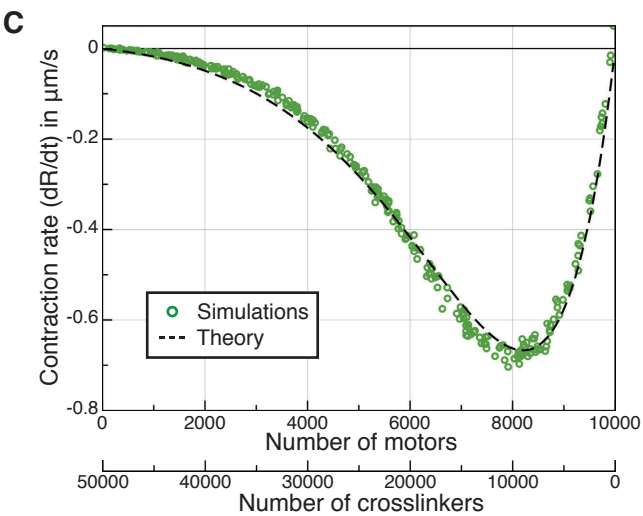
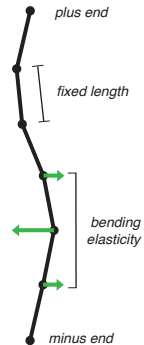


Fig. 3

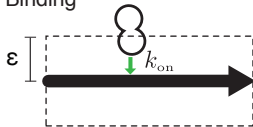
A Filaments



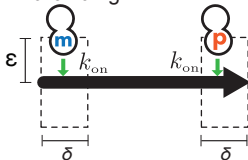
B Diffusion



C Binding



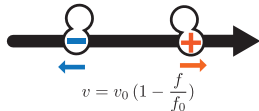
D End-binding



E Linking

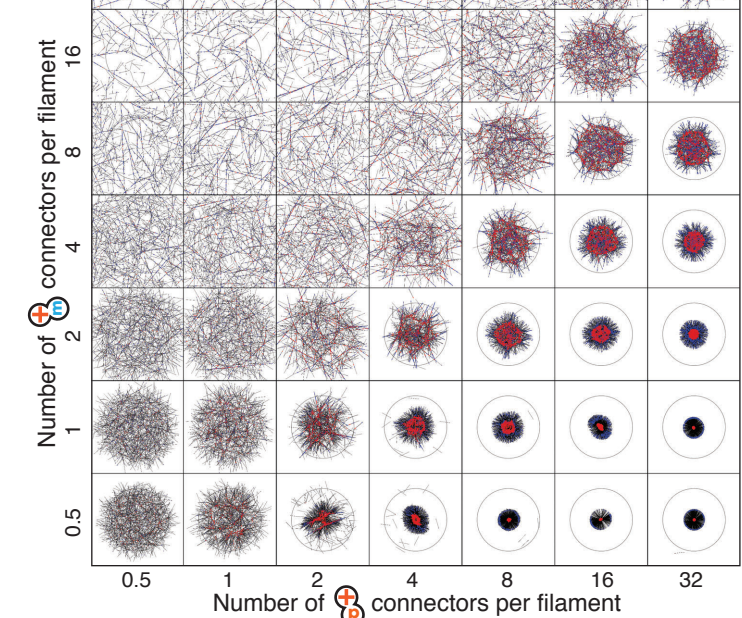
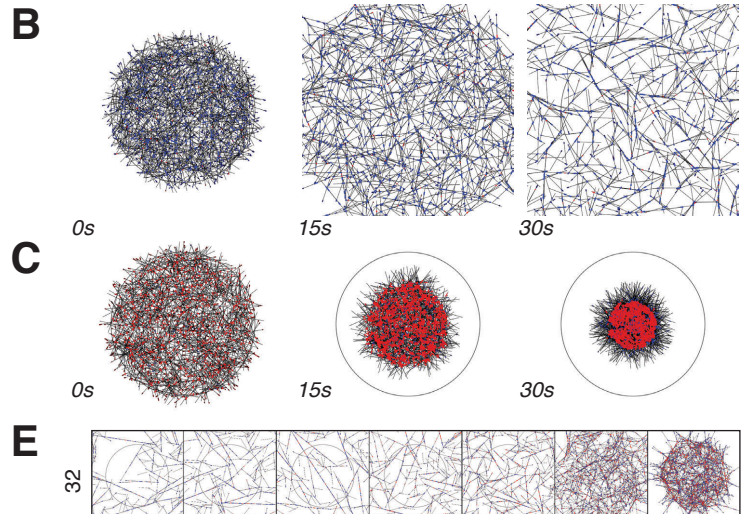
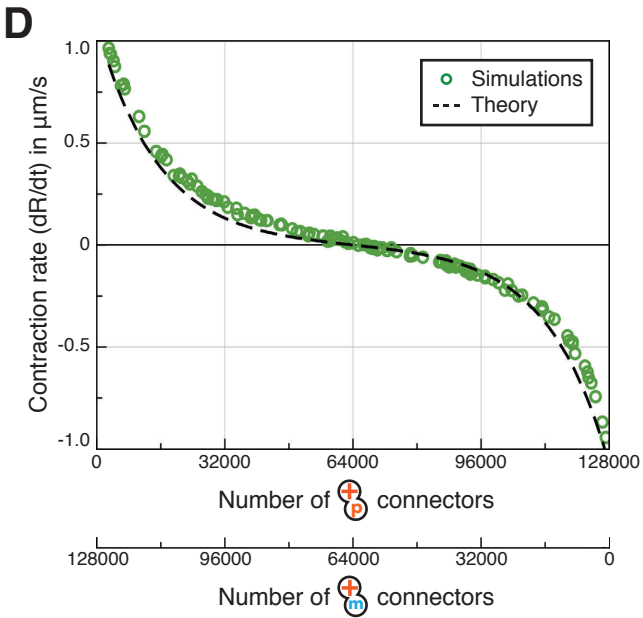
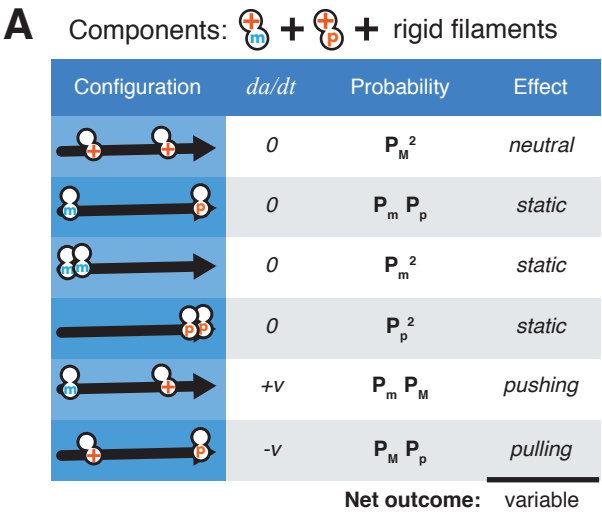


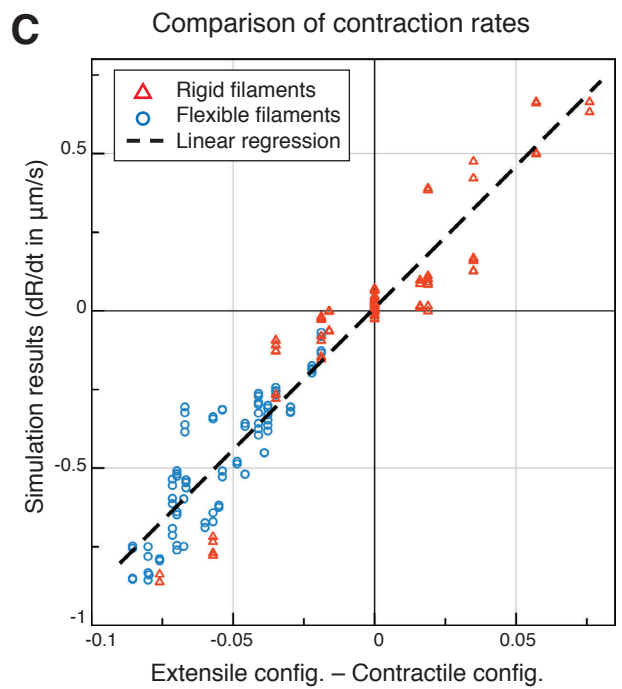
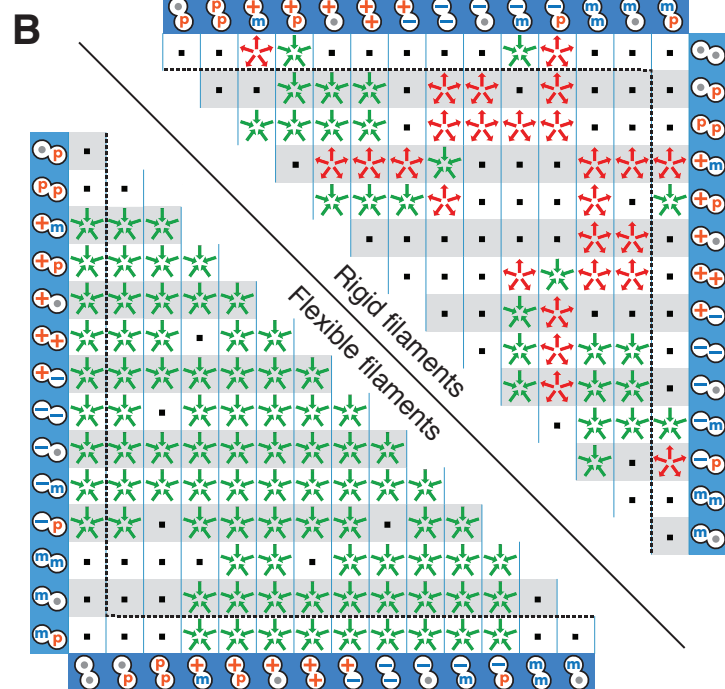
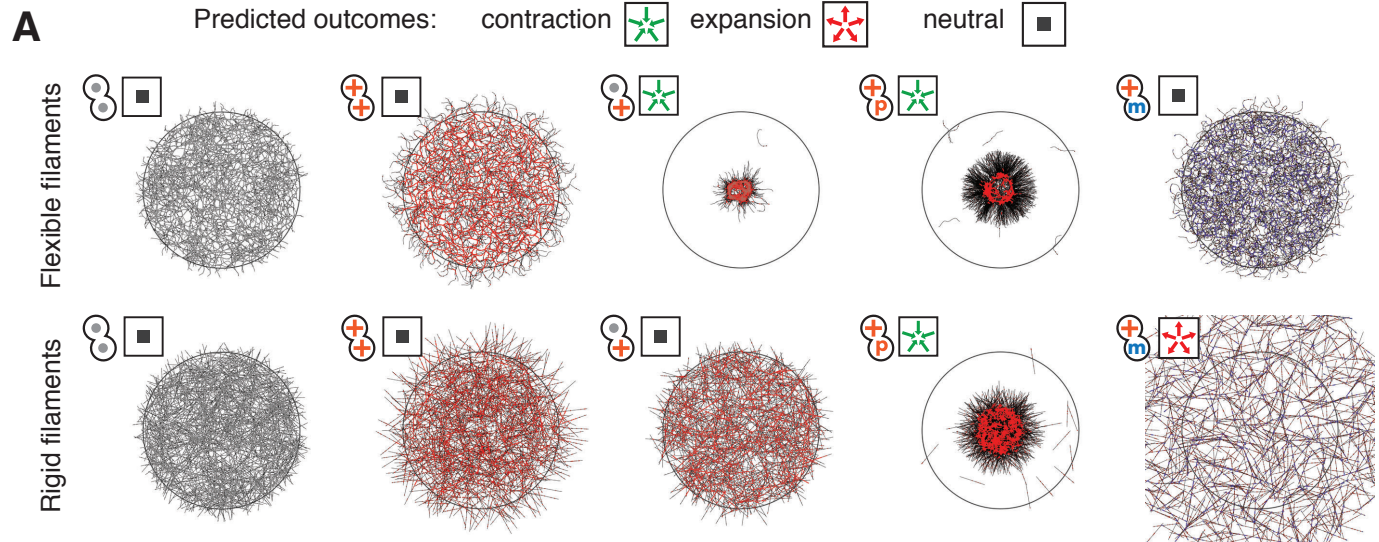
F Movement








G Unbinding

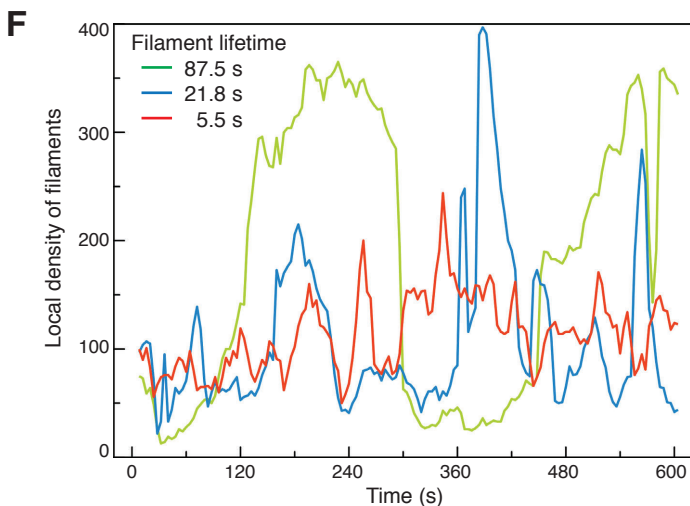
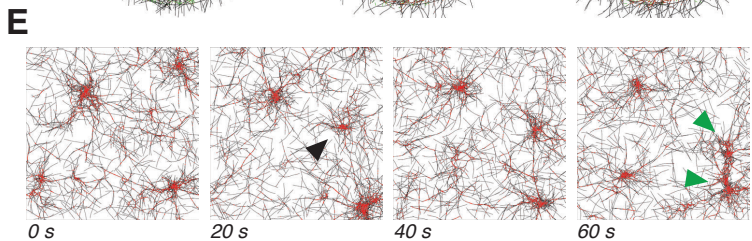
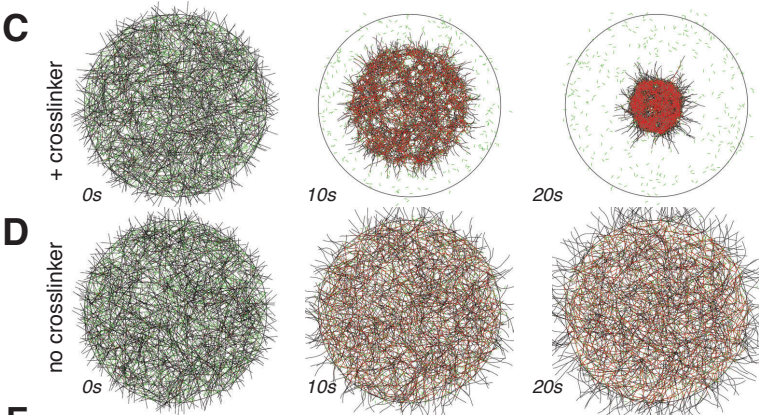
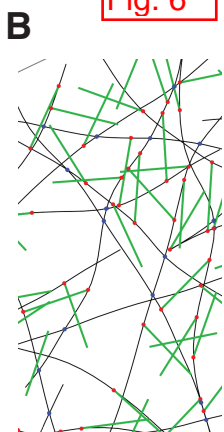






A Components:  + 
 + flexible filaments

Configuration	da/dt	Probability	Effect
	0	1	static
	0	$P_M^2 (1 - P_C)^2$	neutral
	0	P_C^2	static
	-v	$P_M (1 - P_C) P_C$	pulling
	+v	$P_M (1 - P_C) P_C$	(buckling)
Net outcome:			contractile

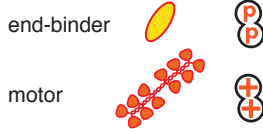
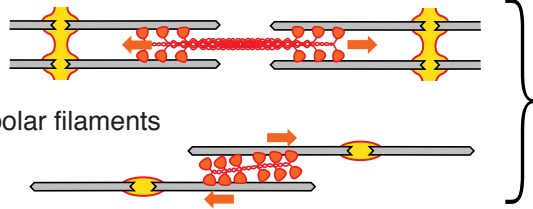


mechanisms

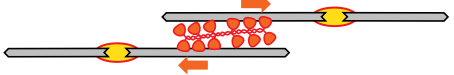
connectors

configurations

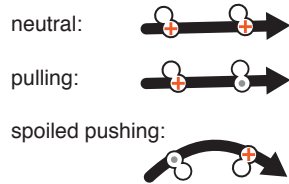
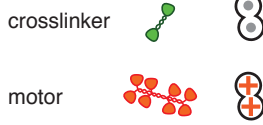
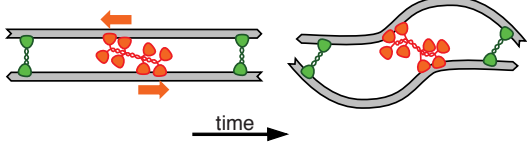
A Sarcomeric



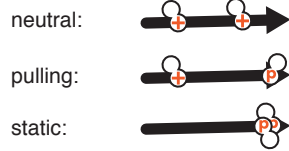
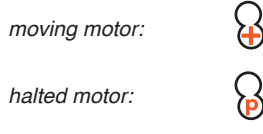
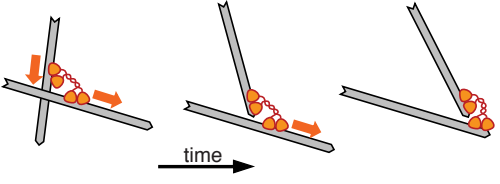
B Bipolar filaments



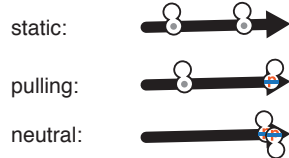
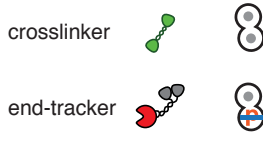
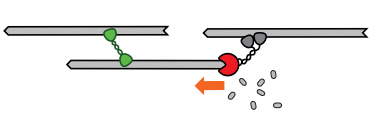
C Dependent on buckling



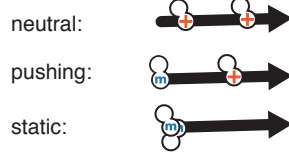
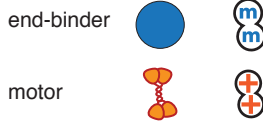
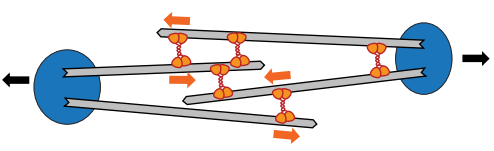
D End-dwelling multivalent motors



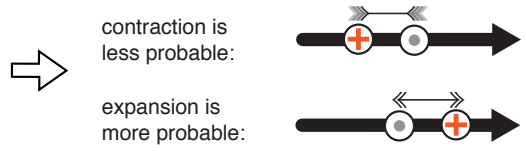
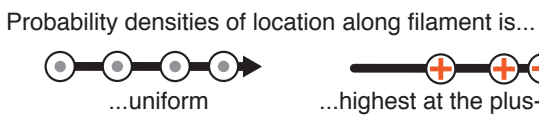
E Filament disassembly



F Mitotic spindle elongation



G Possible extension mechanism for disorganized networks



H Zippering pair of connectors

