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Using connectivity to identify climatic drivers of local adaptation

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Abstract

Despite being able to conclusively demonstrate local adaptation, we are still often unable to objectively determine the climatic drivers of local adaptation. Given the rapid rate of global change, understanding the climatic drivers of local adaptation is vital. Not only will this tell us which climate axes matter most to population fitness, but such knowledge is critical to inform management strategies such as translocation and targeted gene flow. Targeted gene flow, for example, requires knowledge of where in the landscape we can find populations with pre-adapted climate-relevant traits; we cannot find these populations if we do not know the dominant climatic drivers of local adaptation. While simple assessments of geographic trait variation are useful, geographic variation — and its associations with environment — may represent plastic, rather than evolved differences. Additionally, the vast number of environment–trait combinations makes it difficult to determine which aspects of the environment populations adapt to. Here we argue that by incorporating a measure of landscape connectivity as a proxy for gene flow, we can differentiate between environment–trait relationships that are under selection versus those that reflect phenotypic plasticity. By doing so, we can rapidly shorten the list of environment–trait combinations that may be of adaptive significance. We demonstrate this method using data on geographic trait variation in a lizard species from Australia’s Wet Tropics rainforest.

54 **Significance Statement**

55 Despite 150 years of evolutionary research, we still do not have a good method for
 56 determining the climatic drivers of local adaptation. Having such a method is critical
 57 if we are to understand and mitigate the impact of climate change. Here we exploit the
 58 fact that local adaptation is eroded by gene flow to develop a new method for
 59 identifying the environmental drivers of local adaptation. Our method will allow
 60 workers around the world, for the first time (and often using existing datasets), to
 61 determine the climatic drivers of local adaptation in their system. As such, the method
 62 has powerful implications not only for pure evolutionary research, but also for
 63 management.

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Introduction

It is only recently that we have begun to appreciate the speed with which evolution can happen; not only over relatively short timespans (e.g., 1, 2-4), but also at small spatial scales (5). Rapid local adaptation has been recorded in response to a wide suite of environmental drivers, including invasive species, and pollution (6). We expect climate to also be a major driver of local adaptation (e.g., 7, 8), and understanding the way in which species respond to climate is of increasing importance because anthropogenic climate change is proceeding at such a rate that there are concerns that many species will be unable to evolve rapidly enough to avoid extinction (9, 10).

Evolution typically optimizes phenotypes, but the optimum will vary through both time and space (11, 12), in turn leading to populations ('demes') that have, on average, higher fitness in their home environment than an immigrant would: local adaptation. While adaptive optima for traits almost always vary geographically, it does not follow that all geographic trait variation is due to local adaptation. Geographic trait variation can arise due to other factors, such as phenotypic plasticity (including developmental plasticity and maternal effects) and environmental factors (such as geographic variation in fitness-reducing parasites). These factors, and particularly phenotypic plasticity, can give the appearance of local adaptation, despite having no underlying heritable basis (10, 11), complicating our identification of climate-relevant adaptive variation.

To circumvent these issues, evolutionary biologists use experimental approaches to demonstrate local adaptation (12, 13). Experiments designed to detect local adaptation typically utilise one of two techniques: 1) reciprocal transplants, which are done *in*

90 *situ*, and are considered the gold standard for demonstrating local adaptation; or 2)
 91 common garden experiments, which are usually done in the lab where it is easier to
 92 control each environmental variable (12). Both of these techniques can be difficult,
 93 for reasons of time, expense, logistics, or ethics. This difficulty increases as the
 94 number of separate demes and environmental variables to be tested increases and as
 95 the generation time of the organism increases (12). Additionally, although reciprocal
 96 transplants will detect signs of local adaptation, they are not necessarily suited to
 97 identifying the environmental drivers of that local adaptation (14). This is because *in*
 98 *situ* reciprocal transplants necessarily encompass all the environmental variables that
 99 differ between the transplant locations. Lab-based common garden approaches may,
 100 in principle, be more suited to identifying environmental drivers (because the
 101 environment may be under a degree of control), but in practice it often remains
 102 impossible to identify the environmental drivers of trait variation seen in the wild.
 103 Thus, the best experimental tools we have for studying local adaptation are
 104 demanding in terms of time and cost, and are unsuitable for assigning environmental
 105 drivers (such as climate variables) to adaptive variation. If we are looking for climate-
 106 driven local adaptation, this is a problem: we want to know which climate variable or
 107 variables are the main drivers of adaptation, and we urgently need this information for
 108 many species.

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110 By definition, local adaptation has a genetic basis and is consequently weakened by
 111 gene flow (11, 15, 16). Demes with high levels of inward gene flow are therefore
 112 likely to be less optimally adapted, causing an observable mismatch between optimal
 113 and actual phenotypes. Some examples of this are birds dispersing and producing
 114 clutch sizes that are not optimised for the habitat quality in which they are now

nesting (17), larval salamander colouration not matching streambed colouration due to high levels of gene flow from nearby but predator-free streams (18), and stick insects in smaller habitat patches having non-cryptic colouration when the surrounding patches are larger and environmentally dissimilar (19). These observations of "migrant load" suggest an alternative technique for identifying and assessing local adaptation. First, we look across populations for correlations between the environment (e.g., mean annual temperature) and traits (e.g., morphology, physiology). By themselves, these correlations are not sufficient evidence of local adaptation — they could also be caused by phenotypic plasticity. Second, knowing that local adaptation is hindered by gene flow, we can look at whether gene flow diminishes the environmental effect. In cases where data on gene flow are absent (which is often the case), habitat connectivity can be used as a substitute for gene flow. Environment–trait correlations that are strong, but which are also weakened by connectivity, are indicative of environment–trait correlations that have arisen through local adaptation.

If we now collect data on a large number of environment–trait correlations, and their interaction with connectivity, we can imagine several possible patterns emerging. These possibilities are depicted in Figure 1. Each panel represents a possible relationship between environment–trait correlations (environmental coefficients on the x-axis) and the interaction between environment and connectivity (y-axis). Panel A shows a set of environment–trait relationships that are strong, but that are not influenced by connectivity (i.e., no environment–connectivity interaction). This pattern is indicative of a system in which environment–trait correlations are predominantly driven by plastic responses of traits to their environment (i.e., traits always match the local environment, regardless of the level of inward gene flow).

Plastic responses are still of interest, and many species are likely to show such plasticity as the climate changes (20). Panel B shows a system in which environment–trait relationships are eroded by connectivity: increased connectivity diminishes the correlation between the environment and the trait. In this situation, the interaction between the environmental variable and connectivity is negative when the environmental coefficient is positive (i.e., greater connectivity causes the environmental coefficient to decrease towards zero; bottom-right quadrant), and positive when the environmental coefficient is negative (i.e., greater connectivity causes the environmental coefficient to increase towards zero; top-left quadrant). This is the pattern we would expect if there is a genetic basis to the environment–trait correlation, i.e., local adaptation. Panel C shows the situation where the effect of the environment tends to be enhanced by connectivity. This pattern might arise in organisms that are highly mobile and can actively move to their ideal environment, thus avoiding the selective pressures that would lead to local adaptation.

Understanding how species respond to specific aspects of their environment is vital if we are to have any hope of halting the current rapid loss of biodiversity. Climate change is undoubtedly one of the biggest threats to global biodiversity (21, 22), and conservation biologists are looking to a variety of techniques to assess and help mitigate the impacts of climate change on vulnerable species (23–25). One technique that is likely to see increasing use is targeted, or assisted gene flow [TGF; for review, see (23, 26)]. This technique involves the spatial redistribution of long-standing adaptations, and acts to increase genetic diversity in recipient populations, thereby bolstering capacity for evolutionary adaptation (10, 23, 25, 26). When applying TGF to help species adapt to climate change, we need to find an existing location that

matches the future climate at our recipient site, and then translocate animals from that source location. It is a simple idea, but climate is multidimensional and species will not be adapting equally to each climate axis: is a difference of 0.5°C in mean temperature more important than a difference of 100mm in annual rainfall? The answer depends upon which axes have the strongest influence on fitness.

Here we explore the idea of using connectivity to infer local adaptation. To do this we develop a case study of a lizard species from northern Australia. We use this system to examine the relationship, across sites, between traits and climatic variables. We assess how habitat connectivity affects these relationships and use the interaction between the environmental variable and connectivity to rank environment–trait combinations. In doing so, we reveal a set of environment–trait relationships dominated by local adaptation.

Methods

Study species and site selection

The Rainforest Sunskink (*Lampropholis coggeri*) is a small (snout–vent length up to 45 mm), diurnal scincid lizard restricted to the rainforests of the Wet Tropics region of northeastern Australia (27). The rainforests of this region cover a wide range of environmental conditions, spanning significant elevation (0–1600 m ASL), precipitation (annual mean precipitation of 1432–8934 mm, not including input from cloud stripping), and temperature (annual mean temperature of 16.3–25.8°C) gradients. This heliothermic skink is active year-round, often seen basking in patches

of sunlight on the rainforest floor. Lizards were captured by hand from sites that were selected to maximize the environmental heterogeneity sampled (Fig. 2).

Morphological measurements were obtained from 532 skinks from 32 sites. Physiological measurements were obtained from a smaller subset of these lizards: 259 skinks from 12 sites. Between 8–20 skinks were caught at each site on each collecting trip. Following capture, skinks were transported to James Cook University (JCU) in Townsville for trait measurement. All procedures involving lizards were approved by the JCU animal ethics committee (projects A1976 and A1726).

Physiological trials

Physiological trials commenced within seven days of skinks being collected from the field; skinks being used only for morphology were measured and released back at their point of capture within seven days. The following measures were taken from each skink ($n = 259$) during laboratory trials: critical thermal minimum (CT_{min}), critical thermal maximum (CT_{max}), thermal-performance breadth for sprinting (breadth₈₀), maximum sprint speed (R_{max}), temperature at which sprint speed is optimized (T_{opt}), active body temperature as measured in a thermal gradient (T_{active}), and desiccation rate (des) (see Table S1 for further details). Details of trait measurement procedures are detailed elsewhere (see 28, 29).

Morphological measurements

The following measurements were taken from each skink ($n = 532$) using digital calipers: head width (HeadW); head length (HeadL); interlimb length (ILimbL); hindlimb length (HindLL). Left and right measurements were averaged to obtain one measurement for that trait. We also recorded snout–vent length (SVL), total length, and mass (see Table S1 for further details). All measurements were taken by one person (SLM) to minimize observer bias. All morphological variables were log-transformed prior to regression analyses.

Climatic variables, and connectivity

As our study aimed to assess adaptation to local climate, various temperature and precipitation variables were extracted for each site (see Table S2 for details). We considered both means and extremes. It is important to consider climatic extremes, because temperature extremes may be increasing faster than mean temperatures (30), and many species are likely to be particularly vulnerable to these extremes (31). Many environmental variables are highly correlated (28), so only the less-derived variables were used in analyses, specifically: annual mean precipitation (AMP); seasonality of precipitation (Pcov); precipitation of the driest quarter (Pdry); annual mean temperature (AMT); coefficient of variation of temperature (Tcov); average minimum daily temperature (Tmin); average maximum daily temperature (Tmax); average variance of daily maximum temperature (TmaxVar); and average variance of daily Tmin (TminVar).

Our connectivity index was designed to capture the flux of individuals through a location and is detailed in the Supplementary Material. Briefly, it is a measure of habitat suitability for our focal skink species, averaged over space using a species-specific estimate of dispersal potential. As our species is an obligate rainforest-dweller, grid cells in the landscape that are rainforest and that are surrounded by rainforest have high connectivity indices, while grid cells of rainforest surrounded by non-rainforest matrix have low indices. See Table S2 for further details on all variables, and Figure S1 for correlations between all variables.

Analysis

Our analysis aimed to assess: 1) the relationship, across sites, between each trait and each environmental variable; and 2) how connectivity affected each of these relationships (i.e., the interaction between connectivity and environment). To allow comparison of coefficients across variables, and to make interaction effect-sizes meaningful, all trait and environmental variables were standardized so they had a mean of 0 and a standard deviation of 1. Linear models were fitted for each pair of environment–trait variables, with all models including the effect of lizard body size and sex, as well as the interaction between environment and connectivity:

$$\text{trait}_i = A + B_{svl} \times \text{SVL}_i + B_{sex} \times \text{Sex}_i + B_{env} \times \text{Env} + B_{conn} \times \text{Conn} + B_{int} \times \text{Env} \times \text{Conn} + \text{error}_i$$

Where:

trait_i = standardized trait value of interest for lizard *i*

A = intercept

B_{svl} = coefficient of SVL

SVL = lizard snout–vent length, to control for effect of body size

B_{sex} = coefficient of Sex

259 Sex = lizard sex (this species is sexually dimorphic in some morphological
 260 traits)
 261 B_{env} = coefficient of environmental variable
 262 Env = environmental variable (e.g., annual mean temperature) at the lizard's
 263 site
 264 B_{conn} = coefficient of connectivity
 265 Conn = connectivity index at the lizard's site
 266 B_{int} = coefficient of interaction between Env and Conn
 267 $error_i$ = deviation between expected value and trait value of lizard i
 268

A score for ranking the strength of local adaptation was then calculated as:

$$L = -B_{env} B_{int}$$

If the signs of the two coefficients (B_{env} and B_{int}) are opposite (which indicates an environmental effect being eroded by high connectivity, i.e., evidence for local adaptation), L will be positive. If the signs are the same (which indicates an environmental effect being enhanced by high connectivity, a situation not consistent with local adaptation), L will be negative. Thus, higher numbers on this scale equate to stronger evidence for local adaptation in that environment–trait pair. This score can, in theory, range from $-\infty$ to $+\infty$. Once many environment–trait combinations have been assessed, the coefficients for all pairs can be plotted (see Fig. 1). As described in the Introduction, in a system dominated by local adaptation, we expect to see a negative relationship between B_{env} and B_{int} (Fig. 1B). All analyses were conducted in R v3.2 (32).

Results

There was substantial variation in the effect of environment (B_{env}) and its interaction with connectivity (B_{int}) across climate and trait variables, with B_{env} ranging from -1.64 to 1.37, and B_{int} ranging from -0.89 to 0.74 (Fig. 3). Despite this variation, a clear pattern is evident, with most points in Figure 3 appearing in the top-left or bottom-right quadrants: the quadrants in which the two coefficients have opposing signs, and where we would expect points to fall if environment–trait relationships are due to local adaptation. Across these environment–trait combinations there is a distinct negative linear trend (slope= -0.36, $p < 0.001$). It is especially noteworthy that the environment–trait pairs with the largest coefficients (i.e., those points furthest away from the origin) are in the two quadrants indicative of local adaptation.

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 295 Overall, physiological traits showed substantially stronger environmental effects (i.e.,
 296 larger values of B_{env}) than did morphological traits, with the largest environmental
 297 effects being exhibited by CTmin (AMP: -1.64; Tmax: 1.31; Pdry: 1.28) and CTmax
 298 (Pdry: 1.37). Physiological traits also showed stronger interactions between
 299 environmental effects and connectivity, again with CTmin and CTmax showing the
 300 largest interactions. These trends are apparent when we examine our index of local
 301 adaptation, L . Figure 4 shows a heatmap of all environment–trait pairs, ranked via
 302 reciprocal averaging according to the strength of their local adaptation index. The
 303 environment–trait pairs that show the strongest signature of local adaptation appear at
 304 the top-left in red. There is a rough divide, with most of the physiological traits on the
 305 left and most of the morphological traits on the right. The exceptions are the
 306 physiological traits T_{opt} and R_{max}, which appear at the far right of the figure.
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308 The two environmental variables that produced the strongest effects (topmost rows in
 309 Fig 4) were both precipitation related: annual mean precipitation (AMP) and
 310 precipitation of the driest quarter (Pdry). In our system, AMP and Pdry are both
 311 highly correlated with connectivity (see Fig. S1). This is expected, because our
 312 connectivity index is largely a measure of where rainforest is, and the distribution of
 313 rainforest in our study region is driven to a large degree by rainfall.

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316 Discussion

317 Understanding environment–trait correlations will help us plan management
 318 strategies, such as targeted gene flow (TGF), that can mitigate the impact of climate
 319 change on vulnerable species. Numerous studies have looked for (and found)

environment–trait correlations (e.g., 18, 19, 33, 34–36), but the interpretation of these associations is plagued with uncertainty: are they associations due to local adaptation, or plasticity? By acknowledging that gene flow undermines adaptation, we can incorporate connectivity (a proxy for the flux of genes) into our analysis, and in doing so, separate correlations due to local adaptation from those due to plasticity.

Local adaptation

In the environment–trait combinations we assessed, physiological traits typically showed a substantially stronger effect of environment than did morphological traits, with the largest environmental effects shown in CT_{max} and CT_{min} (Figs. 3 & 4). Physiological traits also generally showed stronger environment–connectivity interactions, again with CT_{min} and CT_{max} showing the largest interactions. Overall, physiological traits generally showed stronger evidence of local adaptation than did morphological traits. This result is intuitive: we would expect an ectotherm’s physiological traits to be under strong selection from climate (37–39), but the fitness link between morphology and climate is much less clear. Had we also included some environmental variables that had a clearer bearing on morphology, we might have detected stronger environment–trait relationships for morphology. For example, skinks that occur in rockier habitats show various morphological adaptations to that environment (40). Including a measure of rockiness in our set of environmental variables, for example, might have allowed us to detect environmental correlations with limb length.

Of the environmental variables used, our analysis suggests that precipitation is a very strong driver of local adaptation, even in thermal traits that might not seem obviously

related to precipitation (e.g., CTmin, CTmax). Although this may seem a surprising result, precipitation has been shown to directly affect growth rate, body temperature, activity patterns, and thermoregulatory opportunities in lizards (38, 41-45). Wetter areas also have higher thermal inertia (and so lower cyclical thermal fluctuations (46)), and changed environmental variance in temperature potentially has a strong influence on thermal limits (47). Additionally, Bonebrake and Mastrandrea (48) found that changes in precipitation can significantly affect modeled fitness and performance curves. Finally, comparative analyses also suggest that precipitation can influence thermal traits in many species (38). Thus, although the mechanisms linking precipitation to thermal limits are diffuse and poorly resolved, they do exist, and our analyses suggest that precipitation is a strong driver of local adaptation at thermal physiological traits.

Our analysis also suggests that temperature is an important driver of local adaptation in this system, but that extremes of temperature (encapsulated in minimum and maximum temperatures) are at least as strongly associated with local adaptation as is mean temperature. Again, this result is intuitive (natural selection from climate is likely stronger during extreme events than during normal daily temperatures) and agrees with results of empirical studies (38). Finally, the environmental variables with the weakest signals of local adaptation are Tcov (temperature seasonality), TminVar, and TmaxVar (variance of minimum and maximum daily temperatures, respectively). These variables represent predictable environmental variation occurring within an individual's lifespan and so are variables to which we might expect individuals to develop plastic responses, rather than adaptive responses (49-51).

System-wide signal of local adaptation

The clear negative linear trend displayed in Fig. 3 is precisely what we would expect in a set of environment–trait combinations dominated by local adaptation. Migrant load (the negative effect of the immigration of less-locally adapted individuals) scales positively with immigration as well as with the strength of selection [see equation 5 in Polechová, Barton and Marion (52)]. The reason for this is that, when the strength of selection is moderately high, the environment will have a large effect on relevant traits, and therefore any immigrants coming from differing environments will be particularly maladapted and will therefore have a large and negative impact on the local phenotype. Thus, we expect environment–trait combinations with strong local adaptation to show strong effects of connectivity on the environment–trait correlation (52).

We used long-term climatic averages and found strong evidence that local adaptation dominates over plasticity in our system. If we had included different environmental variables, such as the conditions the lizards had recently encountered, signals of plasticity may have been more apparent. Clearly environmental variables that are similar across generations should lead to local adaptation, while environmental variables that fluctuate within generations should have a strong influence on phenotypic plasticity

Phenotypic plasticity

The importance of accounting for phenotypic plasticity is exemplified in our dataset by the relatively strong effect of precipitation of the driest quarter (Pd_{dry}) on the temperature at which maximum sprint speed is achieved (T_{opt}) and on maximum

sprint speed (R_{\max}) itself. On their own, these strong correlations might traditionally be considered evidence for local adaptation. Our analysis, however, suggests that the environmental effect is largely unaffected by connectivity, implying that variation in these traits is due to plasticity. Other work (28) has shown little temporal variation in T_{opt} (within generations) despite clear geographic variation and this, together with our results, suggests that this trait undergoes developmental plasticity, but is fixed in adult lizards. In principle, this non-effect of connectivity could also arise due to selection that is so strong that it maintains local adaptation despite high levels of gene flow [i.e., immigrants are selected against so strongly that they do not contribute to the effective population (11)]. The environment–trait relationships for T_{opt} and R_{\max} are, however, weaker than those for some other traits (e.g., CT_{\max} and CT_{\min}) that show clear effects of connectivity, so extremely strong selection seems an unlikely explanation for the pattern we see here.

Assumptions and limitations

Our analysis requires an index of population connectivity across the landscape, something that can be calculated relatively easily for many species by using broad scale habitat mapping datasets [e.g., vegetation mapping from DERM (53)]. Where possible, these measures of connectivity should be calculated at a scale relevant to the scale of dispersal of the species in question [as was ours, using dispersal rate data for *Lampropholis coggeri* from Singhal and Moritz (54)]. For highly mobile species, such as plants or insects that have wind-assisted dispersal and for which calculating connectivity between populations may be difficult, it is possible our approach will not work. But for many species of animals with relatively low vagility (i.e., the species for which TGF is most needed), our approach should be generally applicable.

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421 Our approach implicitly assumes that traits have a linear response to the environment
 422 (at least at the environmental scale across which we are looking). In most instances,
 423 this will be a reasonable assumption: it seems unlikely, for example, that a trait such
 424 as desiccation resistance would be high in dry environments, low in moderately wet
 425 environments, and then high again in very wet environments. The reason our
 426 assumption of linear environment–trait relationships bears mentioning is that a
 427 limitation of our technique arises when the connectivity index is strongly correlated
 428 with one or more of the other environmental variables being used. In our system, for
 429 example, AMP and Pdry are correlated with connectivity (Fig. S1). Where the
 430 environment–connectivity correlation is very strong, the interaction term in our model
 431 ($\text{Conn} \times \text{Env}$) could be interpreted as a quadratic term for environment (i.e., Env^2). In
 432 these cases, it is possible that a strong connectivity interaction is, in fact, pointing to a
 433 non-linear environment–trait relationship. Thus, for environmental variables that
 434 correlate with connectivity (and there will always be some), careful consideration
 435 needs to be given to the possibility of a quadratic fitness function between
 436 environment and trait. In our case, it remains possible, for example, that the strong
 437 influence of precipitation on local adaptation in our system is spurious, and instead
 438 reflects non-linear relationships between optimal trait values and precipitation. We
 439 can, however, think of no obvious reason why thermal limits should respond
 440 quadratically to precipitation, nor why desiccation rates and other physiological traits
 441 should also do so. Thus we are inclined to accept the importance of this
 442 environmental variable in driving local adaptation in our system.

443

Finally, our approach, by examining one environment–trait combination at a time, may potentially miss relationships that only appear in multivariate analyses. For example, if two environmental variables are negatively correlated but both have a positive effect on a trait, it is possible that these countergradients can obscure the univariate relationship. Similar problems are encountered when examining response to selection over time (55), and with our approach may lead us to underestimate the number of important environmental drivers of local adaptation. To minimize this effect, care should be taken in future work to sample environmental spaces in such a way as to minimize correlations between environmental variables. Such an aim can be achieved by, for example, strategically exploiting latitudinal and altitudinal gradients.

Conclusion

There is increasing urgency to identify populations that will act as suitable sources for targeted gene flow efforts in the face of climate change. To identify these populations, we need to know which traits influence sensitivity to climate and are locally adapted. Traditional approaches to unearthing local adaptation (reciprocal transplants and common garden experiments) are time consuming, and often cannot attribute adaptation to any particular environmental driver. Local adaptation is, however, undermined by gene flow, and we should be able to use this fact to sort patterns of local adaptation from patterns with other causes. Here we have demonstrated this approach: using connectivity as a proxy for gene flow, and looking for its effect on environment–trait correlations. Our analysis, using a species of lizard from Australia’s Wet Tropics rainforest, suggests the approach has merit: the results we achieve are coherent and suggest local adaptation is the overwhelming signal in the set of environment-trait correlations tested. As well as confirming a strong role for local

adaptation, we have effectively ranked environmental drivers of local adaptation, finding evidence that precipitation and temperature are important environmental variables with regard to local adaptation. Our analysis also suggests that some traits exhibit strong plastic responses to the environment, particularly in response to precipitation of the driest quarter and the seasonality of temperature and precipitation. These specific results will likely apply to other species that are phylogenetically or ecologically similar to our focal species, but the method has the potential to apply much more broadly. Given the potential of this method to provide evidence of local adaptation, and rapid ranking of the climatic drivers of local adaptation, assessment of the method in a broader array of systems is warranted.

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Figure Captions

Figure 1. Graphs showing the concepts illustrated by plotting a set of environment–trait coefficients (x-axis) and the corresponding environment–connectivity interaction coefficients (y-axis). A) Phenotypic plasticity is suggested when environment–trait relationships are strong, but are not influenced by connectivity. B) Local adaptation is suggested when increasing connectivity diminishes the correlation between the environment and the trait. C) The effect of the environment is enhanced by connectivity. This latter pattern might arise in organisms that are highly mobile and can actively move to their ideal environment, thus avoiding the selective pressures that would lead to local adaptation.

Figure 2. Map of the southern Australian Wet Tropics bioregion, showing the distribution of rainforest and the sampling locations.

Figure 3. Scatterplot showing the results of 99 linear models run to assess the relationship between each pair of environment–trait variables and the environment–connectivity interaction. Environment–trait coefficients are on the x-axis, and environment–connectivity interaction coefficients are on the y-axis. Local adaptation is suggested in environment–trait pairs that show a strong environmental effect that is eroded by increasing connectivity.

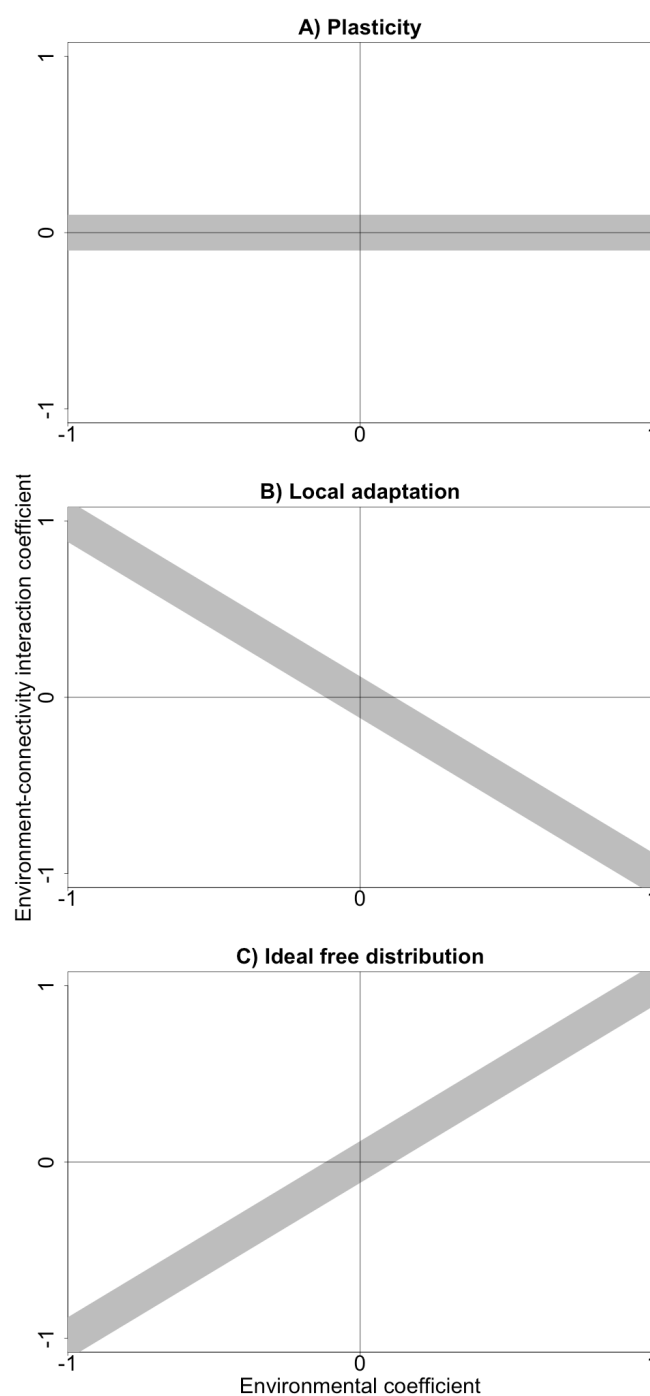
Figure 4. Heatmap showing the relative rankings of climate variables (rows) and morphological and physiological traits (columns). The matrix has been sorted (by reciprocal averaging) and coloured according to the strength of local adaptation, with higher values coloured red and being sorted to the top/left. See Tables S1 and S2 for explanations of the trait and environmental variables used.

L = local adaptation index: $-B_{env} B_{int}$

B_{env} = coefficient of the environmental variable.

B_{int} = coefficient of the interaction between the environment and connectivity terms.

674 Figures



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676 **Figure 1**
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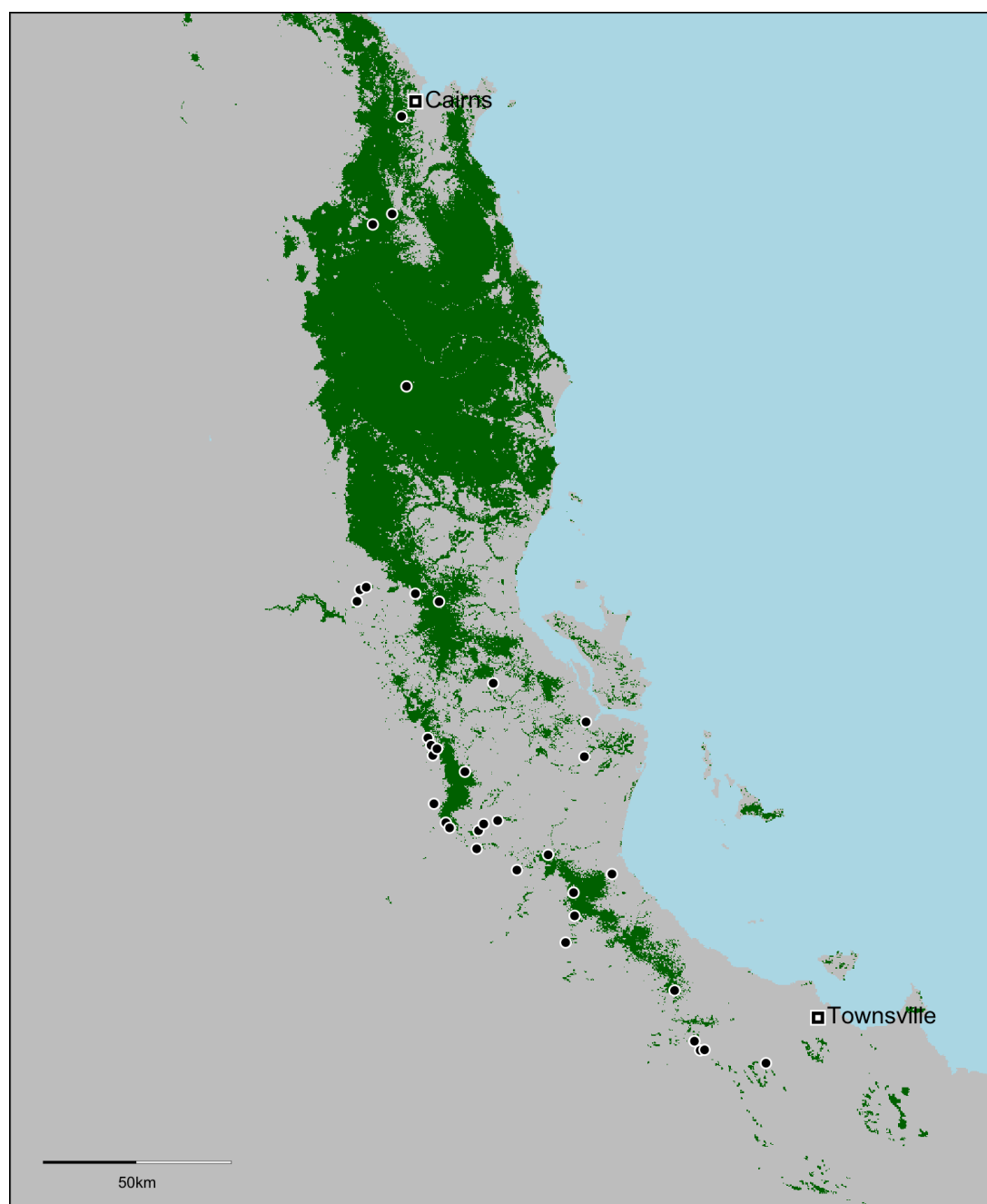
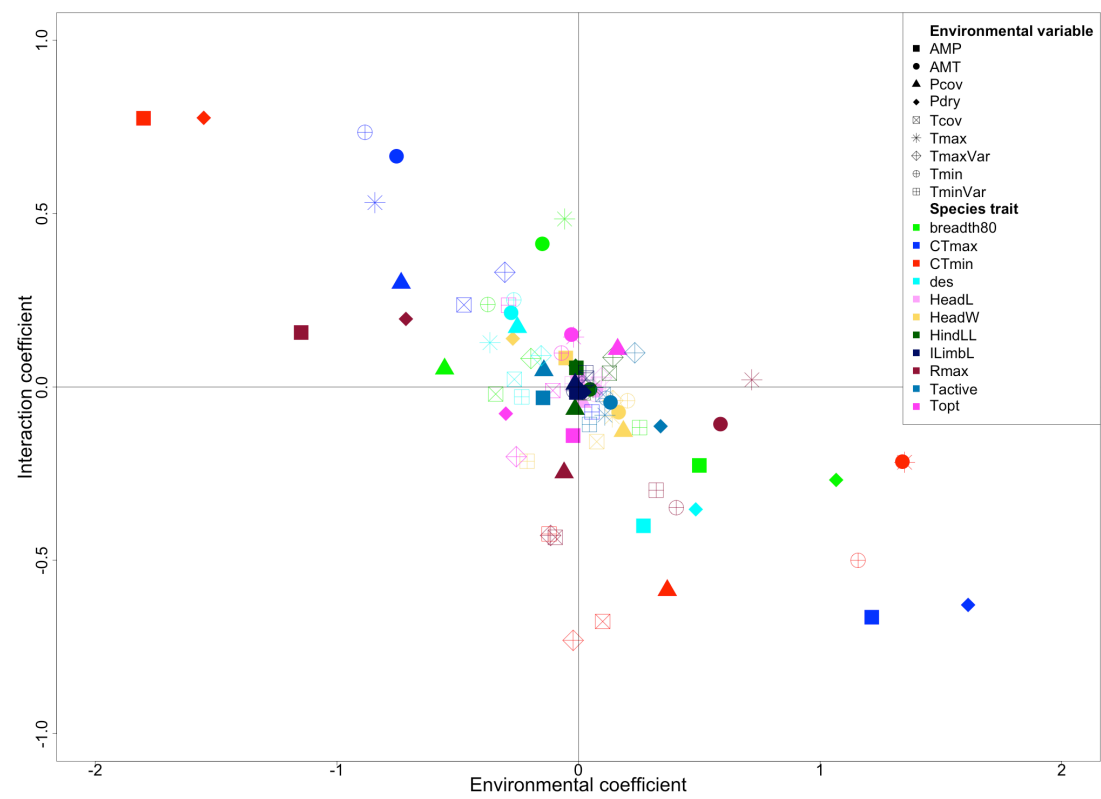


Figure 2

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Figure 3

	CTmin	CTmax	breadth80	Rmax	des	HeadW	Tactive	HindLL	HeadL	ILimL	Topt
AMP	L = 0.7751	L = -0.6644	L = -0.2265	L = 0.1570	L = -0.4007	L = 0.0835	L = -0.0315	L = 0.0551	L = 0.0219	L = -0.0159	L = -0.1404
Pdry	L = 0.7763	L = -0.6289	L = -0.2683	L = 0.1961	L = -0.3532	L = 0.1395	L = -0.1134	L = 0.0613	L = 0.0279	L = -0.0150	L = -0.0771
Tmin	L = -0.5004	L = 0.7345	L = 0.2380	L = -0.3484	L = 0.2510	L = -0.0399	L = -0.0329	L = -0.0157	L = -0.0219	L = -0.0116	L = 0.0977
AMT	L = -0.2156	L = 0.6655	L = 0.4128	L = -0.1072	L = 0.2139	L = -0.0730	L = -0.0447	L = -0.0071	L = -0.0099	L = -0.0043	L = 0.1511
Tmax	L = -0.2173	L = 0.5316	L = 0.4846	L = 0.0206	L = 0.1275	L = -0.0864	L = -0.0813	L = -0.0020	L = -0.0041	L = 0.0002	L = 0.1438
Pcov	L = -0.5868	L = 0.2996	L = 0.0522	L = -0.2473	L = 0.1719	L = -0.1279	L = 0.0474	L = -0.0642	L = -0.0440	L = 0.0082	L = 0.1089
Tcov	L = -0.6766	L = 0.2366	L = -0.0206	L = -0.4338	L = 0.0219	L = -0.1579	L = -0.0219	L = 0.0397	L = 0.0083	L = 0.0253	L = -0.0104
TmaxVar	L = -0.7312	L = 0.3308	L = 0.0820	L = -0.4284	L = 0.0898	L = -0.0394	L = 0.0984	L = 0.0853	L = 0.0267	L = 0.0034	L = -0.2013
TminVar	L = -0.4241	L = -0.0721	L = -0.1174	L = -0.2979	L = -0.0289	L = -0.2144	L = -0.1083	L = -0.0172	L = 0.0095	L = 0.0409	L = 0.2357

Figure 4