Melanism patches up the defective cuticular morphological traits through promoting the up-regulation of cuticular protein-coding genes in *Bombyx mori*

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Abstract

Melanin and cuticular proteins are important cuticle components in insect. The cuticle defects caused by the loss function of cuticular protein-encoding genes could hinder melanin deposition. However, the effects of melanin variation on cuticular protein-encoding genes and the corresponding morphological traits associated with these genes remain largely unknown. Using *Bombyx mori* as a model, we showed that the level of melanism during larval cuticle pigmentation correlated positively with the expression of cuticular protein-encoding genes. This correlation stemmed from the simultaneous induction of these genes by the melanin precursors. More importantly, the effect of the melanism background on the cuticle induced the up-regulation of other functionally redundant cuticular protein-encoding genes to rescue the morphological and adaptive defects caused by the dysfunction of some mutated cuticular proteins, and the restorative ability increased with increasing melanism levels, which gives a novel evidence that melanism enhances insect adaptability. These findings deepen our understanding of the interactions among cuticle components, as well as their importance in the stabilization of the normal morphology and function of the cuticle.

Introduction

The prerequisite for the benefits of melanism to insect is not only the integrity of the melanin biosynthesis and regulatory pathway (Wilson *et al.* 2001; Liu *et al.* 2015; Mallet and Hoekstra 2016), but also the normal presence of the platform it relied on

(Wittkopp *et al.* 2003; Wittkopp and Beldade 2009; Andersen 2010; Moussian 2010; Van Belleghem *et al.* 2017). For the insects, the most important fundamental platform for the color pattern drawing is the cuticle (Hopkins and Kramer 1992; Andersen 2010; Moussian 2010). During shaping the cuticle, the maintenance and stability of the cuticle features depends on normal functional cuticular proteins and the interactions with other components (Hopkins and Kramer 1992; Guan *et al.* 2006; Suderman *et al.* 2006; Andersen 2010; Moussian 2010; Chaudhari *et al.* 2011; Noh *et al.* 2016). Due to the crucial roles of cuticular proteins on cuticle development, when their coding genes are loss of function, the abnormal or defective cuticle will likely affect the deposition and attachment of melanin, which is not conducive to the performance of color pattern (Kanekatsu *et al.* 1988; Oota and Kanekatsu 1993; Arakane *et al.* 2012; Jasrapuria *et al.* 2012; Wang 2013; Noh *et al.* 2015). Yet little is known about the corresponding response mode of cuticular protein-encoding genes via the alteration of the melanin biosynthesis or regulatory pathway.

Recently, several high throughput expression surveys showed that the abundance of cuticular protein-encoding genes in different colored integuments varied in some insect species, especially with evidently up-regulated in the melanic regions (Futahashi *et al.* 2012; He *et al.* 2016; Wu *et al.* 2016; Tajiri 2017), and some of those shared very similar expression patterns and functions (Nakato *et al.* 1994; Nakato *et al.* 1997; Shofuda *et al.* 1999; Okamoto *et al.* 2008; Liang *et al.* 2010; Tang *et al.* 2010; Qiao *et al.* 2014).These studies implied that there are probably some relationships between the promotion of melanism and the expression of cuticular

89 protein-encoding genes. Prior to this study, further exploring and understanding of the
90 potential relationships were unclear. Additionally, when melanism-promoting
91 instructions and defective cuticle proteins occur simultaneously, what are the effects
92 of their relationships on the presence of the corresponding morphological traits ?

93 In the Lepidoptera model, *Bombyx mori*, an intriguing phenomenon has been
94 reported that a larval melanic mutant, *Striped* (p^S, 2-0.0) is able to rehabilitate the
95 malformed body shape, as well as the adaptability defects of the *stony* mutant (*st*,
96 8-0.0) (Xiang 1995; Banno *et al.* 2005). A recent study clarified that a transcription
97 factor, *Apontic-like*, which boosts the expression of melanin synthesis genes in
98 epidermal cells, is responsible for the p^S mutant (Yoda *et al.* 2014). Besides this, there
99 are also multiple alleles with different melanism degrees at the *p* locus, including p^B,
100 p^M, *etc* (Xiang 1995; Banno *et al.* 2005; Yoda *et al.* 2014). And the *stony* mutant (*st*,
101 8-0.0) which precisely caused by the deletion of a RR1-type larval cuticular
102 protein-encoding gene, *BmLcp17* (or *BmorCPR2*) showed hard and tight touch feeling,
103 uncoordinated ratios of the length of the internodes and the intersegment folds (I/IF),
104 bulges at intersegment folds, and severely defective locomotion and behavioral
105 activities in the larval stage (Qiao *et al.* 2014). In addition, the similarity of the gene
106 expression patterns and functional characteristics among some members of the
107 RR1-type larval cuticular protein-encoding gene family, such as *BmLcp18*, *BmLcp22*,
108 *BmLcp30* also suggest that they may play very similar roles as *BmLcp17* in shaping
109 the larvae cuticle (Nakato *et al.* 1994; Nakato *et al.* 1997; Shofuda *et al.* 1999;
110 Okamoto *et al.* 2008; Liang *et al.* 2010; Tang *et al.* 2010; Qiao *et al.* 2014). These

111 dispersed findings are linked through the epistasis of p^S to *stony*, then provide the
112 breakthrough and the exceptional genetic resources for exploring the interactions
113 between melanin and cuticular protein-encoding genes.

114 Here we illustrated that the transcripts levels of four cuticular protein-encoding
115 genes were positively correlated with the melanism degree of larval cuticle, which
116 were due to the simultaneous induction these genes by the intracellular melanin
117 precursors. Moreover, by importing melanism-promoting instruction into the *stony*
118 mutant, the cuticle deficiency rescued through functionally redundant compensation
119 by some other up-regulated cuticular protein-encoding genes, which a new evidence
120 that melanism as a beneficial trait. These findings deepen our understanding of the
121 interactions among genetic factors which shape morphological features in
122 lepidopteran, and emphasize the ecological and evolutionary significance of these
123 mutual interactions.

124

125 **Materials and Methods**

126 ***Silkworm strains***

127 Wild-type strains Dazao ($+^p$) and melanic mutant strains p^M, p^S and p^B (Xiang 1995;
128 Banno *et al.* 2005; Yoda *et al.* 2014) were analyzed in this study. The darkness of
129 pigment was measured as mean OD value using Image J (https://imagej.nih.gov/ij/).
130 In terms of melanism degree, the body color of an individual that is homozygous at
131 one of the aforementioned melanic loci is darker than that of a heterozygous
132 individual (Xiang 1995; Banno *et al.* 2005). The albinism mutant *albino* (*al*) (Banno

et al. 2005; Fujii *et al.* 2013), non-diapause wild-type strain N4 (used for melanin inhibition treatment) and *BmLcp17* deletion strain Dazao-*stony* (near isogeneic line of Dazao, which have been backcrossed with Dazao over 26 generations) were supplied by the Silkworm Gene Bank in Southwest University. The *al* mutant was fed with artificial die at 28℃, and the others were fed fresh mulberry leaves under a 12□h/12□h light/dark photoperiod at 24□°C.

Chemicals and cell lines

L-dopa (D9628), dopamine (H8502), tetrahydrofolic acid (BH_4) (T3125) and 2,4-Diamino-6-hydroxypyrimidine (DAHP) (D19206) were purchased from Sigma-Aldrich (St. Louis, MO, USA). *BmNs* cell line was kept in our laboratory.

Mating combinations and progeny phenotypes identification

The p^S and p^M strains were crossed with the Dazao-*stony* strain to generate the F_1 generation, respectively. The F_2 generation were produced by an F_1 self-cross, and individuals of day 5 of the fifth-instar were collected to further use. The p^B strain was crossed with the Dazao-*stony* strain to generate F_1 progeny, which mated with the Dazao-*stony* strain to generate the BC_1 generation, and fed them until at day 5 of the fifth-instar. Firstly, individuals of the F_2 or BC_1 generations were separated by their cuticle pigmentation. Subsequently, their phenotypes were further classified by morphological characteristics, touch feeling, and the ratios between the number of internodes and intersegmental folds in the second, third, and fourth abdominal segments based on a earlier method (Qiao *et al.* 2014).

Genotyping

Because the p^S, p^M and p^B mutations are multiple alleles at the *p* locus, they should be located in proximity each other on the chromosome 2 (Xiang 1995; Banno *et al.* 2005; Yoda *et al*. 2014). Based on the reported sequence of the gene corresponding to the p^S allele, a polymerase chain reaction (PCR)-based molecular marker within of the genomic region of the *Apontic-like* was designed. PCR screening were performed in the p^X (*X*=*M*, *S* and *B*) and Dazao-*stony* to obtain the polymorphism molecular marker for *p* locus. Similarly, molecular marker was also designed within genomic region of the *BmLcp17* for polymorphism screening of the *stony* locus. The primers used in this study are listed in Table S1.

Association analysis of gene expression, phenotype and genotype

Day 5 fifth-instar larvae of the Dazao or Dazao-*stony* strains were selected for cuticle dissection. The cuticles of the semi-lunar marking region and the non-melanic portion between the paired semi-lunar marking were finely dissected. Then, gene expression levels of *BmLcp17*, *BmLcp18*, *BmLcp22* and *BmLcp30* in these regions were compared. Gene expression patterns in the dorsal epidermis of abdominal segments (from semi-lunar marking to star marking) from day 5 of fifth instar larvae (Dazao, p^S/+, p^M/+, p^B/+). Similarly, day 1 of second instar larvae of the *al* and Dazao strains were also investigated. In addition, the same regions of dorsal epidermis were collected from F_2 generation individuals with the p^S/p^S, *st/*st, and p^S/+, *st/st* genotypes, as well as the p^M/p^M, *st/st* and p^M/+, *st/st* genotypes for gene expression analyses. For all genotyped individuals, the ratios of the length of the intersegment and the intersegment fold were also analysed using image J.

177 ***Melanin precursors-promoting and inhibition treatments***

178 The preparation and concentration of L-dopa and dopamine solutions were slightly
179 modified according to the description by Futahashi (Futahashi and Fujiwara 2005),
180 and filtered with a 0.22 μm filter prior to use. Cells were washed three times with
181 Grace medium without melanin precursors to remove metabolites and other
182 contaminants on the cell surface. Then, 0.8mL medium containing L-dopa or
183 dopamine was added separately into each well, and the medium without melanin
184 precursors was used as the control. The plate was sealed with tape, wrapped with foil
185 and incubated at 28℃ for 24 h for a gene expression analysis. In BH_4 feeding assays,
186 the 30mM working solution was prepared by dissolving tetrahydrofolic acid into
187 ddH_2O, and then smeared on the artificial diet to feed the *al* mutant, and ddH_2O was
188 used as the control. Phenotype were observed and recorded from the second instar and
189 the expression of cuticular protein-encoding genes was also analysed. For the
190 melanism-inhibition experiment, the wild-type strain N4 was selected. Newly-hatched
191 larvae were divided into treatment and control groups. Individuals in the treatment
192 group fed with DAHP dissolved in 0.1M NaOH (15g/L), and individuals in the
193 control group fed 0.1M with NaOH. Phenotypes observation and gene expressions
194 detection were performed on day 1 of the second-instar larvae.

195 ***Quantitative RT–PCR***

196 Total RNA extraction, reverse transcription and qRT-PCR conducted as described
197 previously (Qiao *et al.* 2014). Three biological replicates were prepared for each
198 condition, and *BmRPL3* was used as the internal control. Primers are listed in Table

S1.

Results

Entirely distinct expression patterns of cuticular protein-encoding genes and the cuticle appearance between melanic and non-melanic regions

The expression patterns of *BmLcp17*, *BmLcp18*, *BmLcp22* and *BmLcp30* in melanic or non-melanic epidermal regions are shown in Figure 1. These results, together with earlier studies (Futahashi *et al.* 2012; He *et al.* 2016; Wu *et al.* 2016), revealed that the gene expressions were significantly higher in melanic parts of the cuticle than in non-melanic parts. Moreover, micro protrusions were more intensive in the melanic regions than in the non-melanic regions, and accompanied by a higher chitin content (another important cuticle component (Hopkins and Kramer 1992; Moussian *et al.* 2006; Andersen 2010; Moussian 2010; Chaudhari *et al.* 2011; Qiao *et al.* 2014), and the reduction chitin content was reported to impede cuticle melanism (Moussian *et al.* 2005)) (Figure S1A). These results showed that, regardless of the different genetic basis of the melanic mutants or the melanic markings in the non-melanic strains, excessive accumulation of melanin in the cuticle (accompany by the changes in the surface structure and the chitin content of the melanic cuticle) was closely related to the high expression levels of the cuticular protein-encoding genes.

Expression level of larval cuticular protein-encoding genes positively correlated with the degree of cuticle melanism

To obtain further insights into the relationships between the accumulation of

melanin and the expression of cuticular protein-encoding genes, we investigated gene expression patterns using four different mutations at the *p* locus (Dazao ($+^{p}$), p^{M}, p^{S} and p^{B}), which showed gradually increasing melanin accumulation (Figure 2A). Our results revealed that the gene expression levels were gradually and significantly up-regulated with the increase in the degree of melanism of the cuticle (Figure 2B). They further showed that the expression levels correlated positively with the degree of melanism. Thus, the quantities of cuticular proteins with similar or redundant functions could be increased greatly by the increasing the degree of melanism.

Typical stony phenotyped individuals masked after the introduction of melanic loci into the BmLcp17 defection strain

We assessed the effects of modulating the melanic background on the phenotypic defects caused by the deletion of *BmLcp17*. After mating the p^{B} and *stony* parental, the percentage of BC_1 individuals with melanism and the normal body shape in the backcrossed population of the p^{B}×*stony* cross was 52 % (290/553; and theoretically, it was 25 %), yet individuals with the melanism cuticle and *stony*-type body shape were not found (theoretically should be almost equivalent to the number of individuals with melanism cuticle and normal body shape) (Figure 3). In the cross of p^{S}×*stony*, ~10.8% (36/331) of F_2 progeny had an lighter melanism body color and smaller body size (Figure 3). These individuals exhibited a little hard and tight touched body, but the body was much softer than that of the *stony* mutant. Their intersegment folds bulged slightly, and the length were significantly shorter than that of the internodes; accordingly, their phenotypes slightly resembled the morphological features of the

stony mutant (Figure 3 and 4A). Even so, We did not find individuals with the typical *stony*-type body shape and also defective adaptability under the melanism background (theoretical ratio is 3/16). (Figure 3). Similarly, in the p^M×*stony* corss, only approximately ~11.7% (51/437) of the individuals of the F_2 population were very lightly melanic, but they exhibited obviously unusual morphological features (Figure 3, Figure 4A). When compared with the ~10.8% F_2 individuals (aforementioned) from the p^S×*stony* cross, the touch feelings of individuals from the p^M×*stony* cross were tighter and firmer, and the intersegment folds bulged more obviously and had a higher proportion among the segments, meaning that their body features were closer to the phenotype of the *stony* mutant (Figure 3 and 4A) (Qiao *et al.* 2014). Nevertheless, melanic individuals showing the typical *stony*-type body features and defective adaptability still did not appear (theoretical ratio is 3/16) in the progeny from the p^M × *stony* cross. Therefore, induction of the melanic mutation into individuals with a defective cuticular protein-encoding gene could mask the adverse phenotypes, and the masking effect was more remarkable with the increasing degree of melanism.

Other larval cuticular protein-encoding genes up-regulated evidently in the melanic and non-stony phenotypic, but with mutated BmLcp17 genotypic offspring

Using the molecular markers (closely linked to the *p* and *st* loci, respectively), we further genotyped the progenies with melanic color pattern and non-*stony* (including ambiguous *stony*-like) body shape. The results revealed that approximately 49% of the individuals showing a melanic color and normal body shape in the BC_1 population from the p^B×*stony* cross was the $p^B/+^{pB}$, *st/st* genotype (Figure 4A). The ratios of the

length of the internodes and the intersegment folds (I/IF) were ~4, which is very similar to that in $p^{B}/+^{pB}$, $+^{st}/st$ individuals, and also no significant differences as that in the wild-type individuals (Figure 4B) (Qiao *et al*. 2014). In the F_2 generation from the p^{S}×*stony* cross, approximately 9.3% of the individuals with the p^{S}/p^{S}, *st*/*st* genotype and very few individuals (~1%) with the genotype $p^{S}/+^{pS}$, *st/st* exhibited a melanic color and the normal body shape (Figure 4A). For p^{S}/p^{S}, *st/st* genotyped individuals, the I/IF value was approximately 3.3 (Figure 4B). Although the I/IF value was lower than that in $p^{S}/\_$, $+^{st}/\_$ individuals (approximately 4, Figure 4B), it was much higher than that in the *stony* mutant (approximately 1.6 (Qiao *et al*. 2014)). Despite the slightly smaller body size of the p^{S}/p^{S}, *st/st* individuals, their body shape was essentially normal (Figure 4A). The genotypes of those lightly melanic individuals, whose body shape was slightly like that of the *stony* phenotype (mentioned in Result 3), were all the $p^{S}/+^{pS}$, *st*/*st*, and the I/IF value of these individuals was approximately 2.7 (Figures 4A and 4B). In progeny of the p^{M}×*stony* cross, ~10.5% of the individuals with the p^{M}/p^{M}, *st/st* genotype and ~0.7 % of the individuals with the $p^{M}/+^{pM}$, *st/st* genotype showed a melanic color and subtle *stony* features (just very slight bulges) (Figure 4A). The body size of the p^{M}/p^{M}, *st/st* individuals were smaller than those of the $p^{M}/\_$, $+^{st}/\_$ individuals. They exhibited a certain sense of hardness, and the I/IF ratio was approximately 2.8, which is in good agreement with their phenotypes (Figure 4B). Additionally, individuals showing very slight melanism and morphological features that were more similar to that of the *stony* mutant (mentioned in Result 3) were all the $p^{M}/+^{pM}$, *st/st* genotype, and their I/IF values were

287 approximately 1.8, which is closer to that of the *stony* mutant (Figures 4A and 4B). In
288 addition, the expression of cuticular protein-encoding genes in p^{S}/p^{S}, *st/st* individuals
289 were significantly higher than that in $p^{S}/+^{pS}$, *st/st* individuals (Figure. 4C); a similar
290 result was also obtained from the p^{M}/p^{M}, *st/st* and $p^{M}/+^{pM}$, *st/st* individuals (Figure 4C).
291 Taken together, these results revealed that more cuticular proteins with similar
292 functions were accumulated in the cuticle of melanic homologous individuals at the *p*
293 locus. Based on the comprehensive and association analysis, we infer that melanic
294 background effectively drove the expressions of cuticular protein-encoding genes
295 with similar expression patterns and redundant functions, which compensated for the
296 morphological and adaptability defects caused by the dysfunctional *BmLcp17* gene;
297 and the law of compensatory abilities was $p^{B}/+^{pB}$, *st/st* > p^{S}/p^{S}, *st/st* > p^{M}/p^{M}, *st/st* ≮
298 $p^{S}/+^{pS}$, *st/st* > $p^{M}/+^{pM}$, *st/st*, which corresponds well with the gradual weakening of the
299 degree of melanism (Figures 4 and Figure S3).

300 ***Content variations of melanin precursors affect the transcript amount of the***
301 ***cuticular protein-encoding genes***

302 Due to the crucial material basis for the cuticle melanism (no matter what kind of
303 genetic basis caused by) is the extensive accumulation of melanin precursors in the
304 epidermal cells; thus, the essence that melanism tend to increase the expression of
305 some cuticular protein-encoding genes should be driven by the relations between the
306 accumulation of melanin precursors and the transcripts amount of the cuticular
307 protein-encoding genes. The basal expressions of four cuticular protein-encoding
308 genes were detected in *BmNs* cells (Figure S5), indicating that there are regulatory

309 pathways controlling the expression of cuticular protein-coding genes in this cell line;
310 therefore, this cell line can be used to examine the effect of melanin precursors on
311 gene expressions. After incubating *BmNs* cells with melanin precursors, the
312 expression of cuticular protein-encoding genes was significantly higher in cells
313 treated either by L-dopa or dopamine, compare with that in the control group (Figure
314 5 top (left)). In addition, when treated with BH_4, the second-instar *al* mutant (which is
315 characterized by albinism and a porous cuticle due to a mutation of sepiapterin
316 reductase, which leads to the insufficient synthesis of the co-factor BH_4 during the
317 synthesis of melanin precursors (Banno *et al.* 2005; Fujii *et al.* 2013) was rescued to a
318 melanic body color (Lab unpublished contribution) (Fujii *et al.* 2013), and the gene
319 expressions were obviously higher than that in the control group (Figure 5 top (right)).
320 These results suggest that the expression of cuticular protein-encoding genes can be
321 induced by increasing amounts of melanin precursors. Furthermore, adding DAHP (an
322 inhibitor of guanylate cyclase hydrolase (GTPCHI), which is an important
323 rate-limiting enzyme in the synthesis of BH_4 (Hamadate *et al.* 2008), inhibited the
324 synthesis of BH_4 in wild-type second-instar larvae by blocking the synthesis of
325 melanin precursors in epidermal cells, which caused individuals to lose their original
326 melanic body color (Lab unpublished contribution). The gene expressions were also
327 significantly reduced when compared with that in the control group (Figure 5 bottom).
328 Thus, the content and variation of the intracellular melanin precursors are important
329 factors regulating the expression of cuticular protein-encoding genes. We concluded
330 that the inducing effect of the melanin precursors on the expression of cuticular

331 protein-encoding genes is the basis for melanism promoting genes‘ transcription.

332

333 **Discussion**

334 Melanin is deposited in the cuticle, in which many of cuticular proteins also occur;
335 thus, overlapping distribution of cuticular proteins make it possible for them to
336 interact with melanin, thereby contributing to the phenotype of the cuticle (Hopkins
337 and Kramer 1992; Suderman *et al.* 2006; Andersen 2010; Moussian 2010; Hu *et al.*
338 2013). We clearly observed that extensive star-like protrusions were present on the
339 cuticle when a large amount of melanin accumulated (Figure S1). Similar correlations
340 between cuticle structure and body color have been reported multiple times (Futahashi
341 *et al.* 2012; Noh *et al.* 2016; Tan *et al.* 2016), suggest that there should be some
342 interactions between cuticular proteins and melanin. Although the exact interaction
343 pattern between these two cuticular components is unknown, microscopic observation
344 clearly shows that the deposition of an excessive amount of melanin affected the
345 cuticle characteristics.

346 The expression profile of cuticular protein-encoding genes in melanic silkworm
347 mutants and the black markings of *Papilio* larvae supported the view that
348 overexpressed cuticular proteins probably participate in the transport or maintenance
349 of the corresponding pigments in a specific painted cuticle (Figures 1, 2 and Figure S1)
350 (Futahashi *et al.* 2012; He *et al.* 2016; Wu *et al.* 2016). Yet over-expression of
351 cuticular protein-encoding genes *in vivo* cannot trigger (or induce) the formation of
352 the melanic cuticle (Tajiri *et al.* 2017). Therefore, we reasoned that once the

353 instructions for melanin accumulation are included in the developmental program
354 (melanism being the original factor), other cuticle features and structures should adapt
355 to the level of melanin accumulation, regardless of genetic background. The
356 up-regulation of cuticular protein-encoding genes should be a necessary adaptation for
357 the maintenance and stability of the structural characteristics and physical properties
358 of melanic cuticles. The interactions of melanin and cuticular proteins at the
359 ultrastructural level deserves special attention in follow-up studies.

360 Despite the elaborate regulatory mechanism by which melanin precursors affect the
361 expression of cuticular protein-encoding genes is unclear, this study really revealed
362 the existence of this regulatory phenomenon (Figure 5). Cuticle formation is regulated
363 stringently in temporal and spatial patterns, the accumulation and oxidization of
364 melanin precursors, and the interactions (such as cross-linking) among other
365 components should occur after the formation of the initial formation of the cuticle
366 (Moussian 2010; Sobala and Adler 2016; Tajiri 2017). Therefore, we propose that the
367 cuticular proteins induced by melanin precursors are used to set up a platform for
368 further accumulation and oxidization of melanin precursors. When melanin-associated
369 protein-encoding genes have similar expression patterns and functions (Nakato *et al.*
370 1994; Nakato *et al.* 1997; Shofuda *et al.* 1999; Okamoto *et al.* 2008; Liang *et al.* 2010;
371 Tang *et al.* 2010; Qiao *et al.* 2014), the production of a large amount of functionally
372 similar cuticular proteins would be driven by the melanic background to guarantee
373 construction and stability of the melanic cuticle. During this process, melanism
374 unlocks the complementary features of melanin-associated cuticular proteins (such as

375 *BmLcp18*, *BmLcp22*, *BmLcp30*), rescued cuticular malformation caused by the loss of
376 function of some cuticular proteins (such as defected *BmLcp17* in *stony* mutant).
377 Because it appears that a special cuticle-forming pattern is regulated by melanin
378 accumulation, melanism may enhance insect adaptability to avoid the impairment of
379 survival caused by the mutation and/or functional loss of some cuticular proteins, and
380 these results also add new evidence to explain how melanism can be a beneficial trait
381 (Wilson *et al.* 2001; True 2003; Wittkopp *et al.* 2003; Wittkopp and Beldade 2009;
382 Liu *et al.* 2015; Mallet and Hoekstra 2016).

383 As far as we known, as structural proteins, there is no evidence to suggest that the
384 four larval cuticular protein-encoding genes and their orthologous can enter the
385 nucleus and regulate gene expression by acting as transcription factors. In addition,
386 our findings and some other studies showed that changes in the expression of one
387 cuticular protein-encoding genes of R&R family did not affect the expression of other
388 members (Figure S6, S7) (Arakane *et al.* 2012; Noh *et al.* 2015). Moreover, several
389 studies revealed that organisms optimize resources use at gene or protein expression
390 level (Liebermeister *et al.* 2004; Dekel and Alon 2005); thus, the expression of
391 melanin associated cuticle protein-encoding genes might be appropriately and
392 simultaneously coordinated with the sufficient accumulation of intracellular melanin
393 precursors, as a relatively direct, economical, efficient and convenient strategy.
394 Furthermore, DAHP inhibits GTPCHI activity, but it does not directly impair the
395 extracellular accumulation of melanin and protein–protein interactions in the cuticle.
396 Finally, there is some evidence that melanin precursors regulate gene expressions via

the receptors (Konradi *et al.* 1996; Berke *et al.* 1998; Westin *et al.* 2001). In summary, we hypothesized that the up-regulation of the four *BmLcp* genes were induced simultaneously by excessive amounts of melanin precursors, but should not be the interaction among these four genes on regulation level. To test our hypothesis, further analyses will be performed to determine the expression of the remaining *BmLcp* genes in some *BmLcps* mutant (such as defective *BmLcp17*) cell line by increasing or decreasing the content of melanin precursors.

The markings in the *stony* mutant were lighter than those in the wild-type strain, and accompanied by the down-regulation of melanin synthase genes (Figure S4). If the dysfunction of some cuticular proteins cannot be effectively supplemented, abnormalities of the cuticle structure and interactions of various cuticular components may occur, which probably result in barriers for melanin deposition and metabolism. Perhaps this is the reason why the $p^{S}/+^{pS}$, *st/st* (or $p^{M}/+^{pM}$, *st/st*) genotypes are lighter than the $p^{S}/+^{pS}$, $+^{st}/\_$ (or $p^{M}/+^{pM}$, $+^{st}/\_$) genotypes, besides to their different body shape. Because the homozygous of p^{B} mutation is lethal (Xiang 1995; Banno *et al.* 2005), we were unable to obtain F_2 progeny with the p^{B}/p^{B}, *st/st* genotype. However, it is worth noting that the body shape and the melanism degree of $p^{B}/+^{pB}$, $+^{st}/st$ and $p^{B}/+^{pB}$, *st/st* genotype individuals were almost same (Figure 4A). Therefore, during cuticle formation, if the signal promoting melanism is sufficiently strong, plenty of melanin precursors should be generated. The contents of functionally redundant proteins induced by melanin precursors are enough to fill the gap generated by the dysfunction of some cuticular proteins in theoretically, which will be able to guarantees the normal

accumulation of melanin. Therefore, we believe that there should be a threshold of the melanism-promoting effect. When the accumulation of melanin precursors spans this threshold, the requirements for cuticular proteins with similar function are not weakened even when one of them loses the function, which ensuring the formation of a normal cuticle structure the later pigmentation.

We proposed a possible regulatory model as follows: when large amounts of melanin precursors induced by endogenous- and/or exogenous melanism-promoting factors, the melanin pathway may directly or indirectly induce the up-regulation of some cuticular protein-encoding genes to guarantee the formation of normal structural features of the melanic cuticle. During this process, if some cuticular proteins lose their functions, other functionally redundant cuticular proteins induced by melanin precursors can compensate for the functional defects. The compensatory intensity is increased with increasing melanin accumulation. When melanin accumulation spans a certain threshold, this compensation can totally mask the defective phenotype caused by the malfunctioning genes, which adds to growing evidence that melanism may have pleiotropic effects that enhance fitness over and above the effect of melanin accumulation itself (Figure 6). Due to the coexistence of excess melanin and cuticular proteins is common in other insects, and homologues the four *BmLcp* gene are widely distributed in the Lepidoptera (Table S2), we presume that the above reciprocal action and its corresponding biological significance are conserved in other Lepidoptera insects to that in *Bombyx mori*.

In summary, we used crucial components of the cuticle to elucidate the mutual

441 effects among some key genetic factors, and the physiological significance of these
442 mutual effects during the development of the morphological features. Our findings not
443 only contribute a realistic basis to in-depth study of the interaction patterns of melanin
444 and cuticular proteins, but they will also inspire relevant studies of other Lepidopteran
445 insects or other insect species.

446

447

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Figure legends

Figure 1. Expression of four larval cuticular protein-encoding genes in melanic and non-melanic integuments. A and B represent the gene expressions between the semi-lunar marking (black box) and the non-melanic region (between the semi-lunar

598 marking, red box) in Dazao or Dazao-*stony*, respectively. Scale bar: 2 mm. C.
599 Comparative analysis of gene expressions in the dorsal side of abdominal segments
600 (from the third to the fourth segment, red box) between the p^S and Dazao strains.
601 Scale bar: 1 cm. D. Comparison of gene expressions between the second-instar *al*
602 mutant and the Dazao strain (melanic). The red hashtag symbol indicates the Fig. 1 D
603 we are showing is cited from the previous study of our lab group (Min *et al.* 2016)
604 with modification. Scale bar: 2 mm. *t*-test, n=3; * $p < 0.05$; ** $p < 0.01$; *** $p <$
605 0.001.
606
607 **Figure 2**. Expression of cuticular protein-encoding genes in integuments showing
608 different degree of melanism. A and B display comparisons of degree of melanism
609 and cuticular protein-encoding gene expression levels among four strains with mutant
610 alleles at the *p* locus. Scale bar: 1 cm. Ratios represent the ratios of gene expression
611 levels between two strains. Symbols (−, +, ++, and +++) represent the increment of
612 the degree of melanism. Star represents the melanin-associated cuticular
613 protein-encoding genes. *t*-test, n=3; * $p < 0.05$; ** $p < 0.01$.
614
615 Figure 3. Segregation patterns of the phenotypes of progenies from different crosses
616 of melanic mutant strains and the *stony* strain. In the segregated progenies, the first
617 item in the list (B,N), (B,st), (S,N), (S,st), (M,N), (M,st), (N,N) and (N,st) represents
618 p^B-, p^S- p^M- and Normal color patterns, respectively. The second item indicates body
619 shape features marked with non-*stony* type (N) and the *stony* type (st). It is

noteworthy that in (S, st-am+) and (M, st-am+), the second item represents the ambiguous *stony* body shape. The size of "+" symbol represents the corresponding degree of the ambiguous *stony* body shape. Superscript "T"s represent theoretical values. Superscript "A"s represent actual values. Backslashes indicates that a value was not obtained. Chi-squared test, * $p < 0.05$; ** $p < 0.01$.

Figure 4. Association analysis of the genotypes, phenotypes and gene expression levels in segregated progenies from different crosses. A. Correlation analysis between the genotypes and phenotypes in self-crossed or backcrossed progenies. Scale bar: 1 cm. White and red stars represent polymorphic bands at the $+^{p}$ and p^{X} loci (X = *B*, *S* or *M*), respectively. Red and white hash-tag represent polymorphic bands at the *st* and $+^{st}$ locus, respectively. Solid and dotted red arrows indicate the relative degree of bulging (solid > dotted), respectively. Slashes show the genotypes within one phenotypic category. The thickness of the slash represents the proportion of the corresponding genotypes. Dotted backslashes indicate that the number of individuals with the corresponding genotype is quite low. B. Ratios of the length of internodes and intersegmental folds in the second, third, and fourth abdominal segments of individuals with different genotypes (showing melanic body color) in the self-crossed or backcrossed progenies. n≥3, *t*-test, ** $p < 0.01$. C. Gene expression analysis of cuticular protein-encoding genes in homogeneous and heterogeneous individuals at the *p* locus from self-crossed progenies of p^{S} × *stony*, and p^{M} × *stony* under the condition that the cuticle was melanic and the genotype was homozygous recessive at

642 *st* locus. n = 3, *t* – test, * $p < 0.05$; ** $p \leq 0.01$.

643

644 Figure 5. Effect of melanin precursors (top left: in cells) and BH_4 (top right: in *vivo*)
645 treatments on the expression of cuticular protein-enconding genes (*t*-test; n = 3, * $p <$
646 0.05; ** $p < 0.01$), and variations of gene expression levels in larval cuticle treated
647 with the inhibitor DAHP (bottom). n = 3, *t*-test, ** $p < 0.01$.

648

649 **Figure 6.** Schematic overview of the effect of melanin precursors on the expressions
650 of cuticular protein-encoding genes. Black solid circles represent the melanin.
651 Triangles represent the BmLcps with similar expression patterns and functions. Solid
652 and hollow triangles represent the normal and defective functions, respectively. Red
653 rhombi represent other components (such as chitin) in the cuticle. Solid and dotted
654 double-headed arrows indicate probable interactions and loss of interactions due to
655 the functional deficiency of some *BmLcps* (such as dysfunctional *BmLcp17*),
656 respectively. Purple arrows show the direction in which the melanin precursors or
657 cuticular proteins flow from the haemolymph to the epidermal cells as well as from
658 the epidermal cells to the cuticle. Red arrows indicate the increased contents of
659 melanin precursors or the up-regulation of cuticular protein-encoding genes. Purple
660 polyline arrows indicate melanism-promoting factors produced by other genetic
661 instructions. Double dovetail arrows indicate the effect of melanin precursors on
662 cuticular protein-encoding genes. The question mark indicates that the details of
663 induction (directly or indirectly) are unclear. Here the cuticle especially means the

664 exo-cuticles in which the melanin and cuticular proteins deposited in (Hopkins and

665 Kramer 1992; Suderman *et al.* 2006; Andersen 2010; Moussian 2010).

666

Figure 1

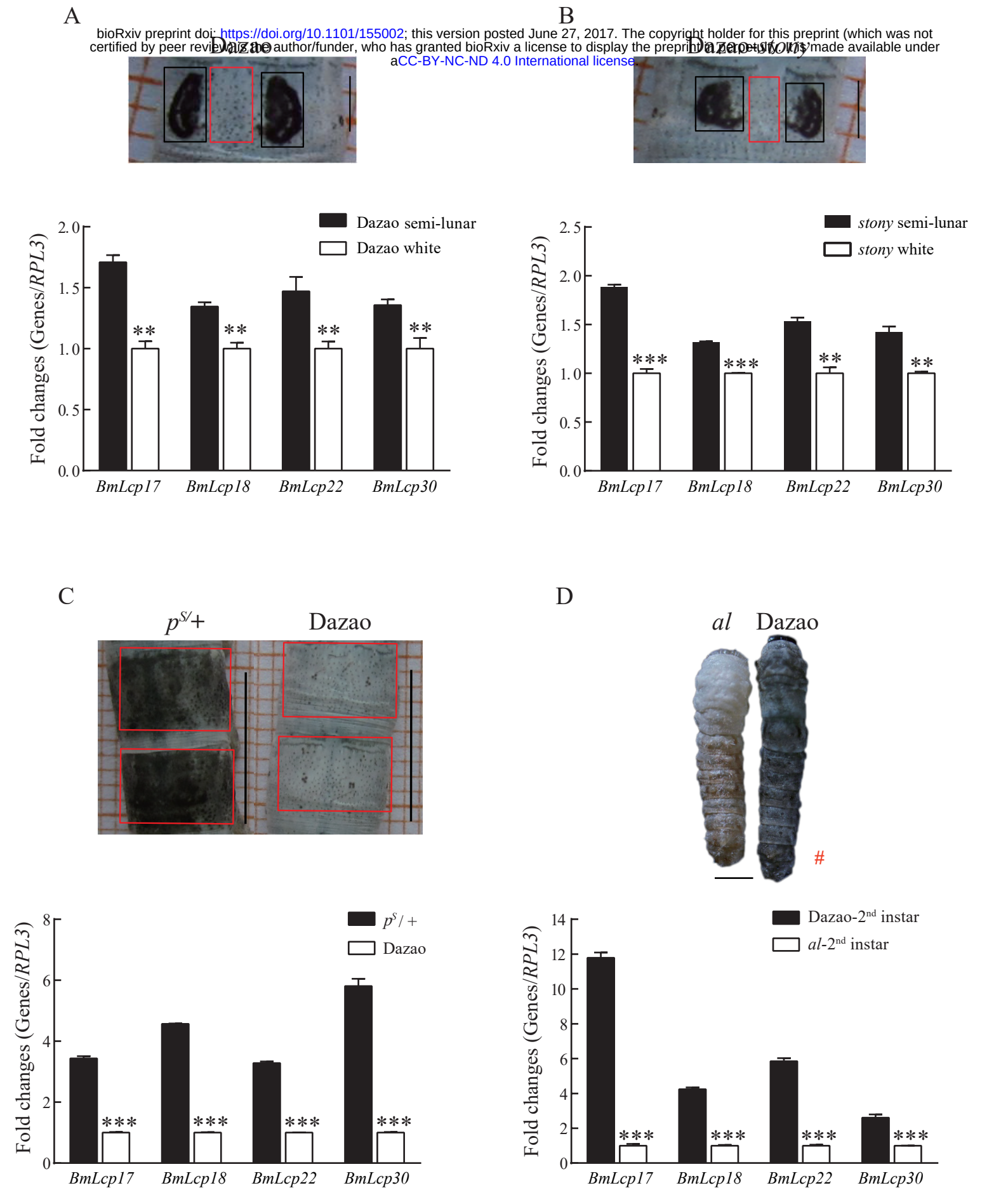
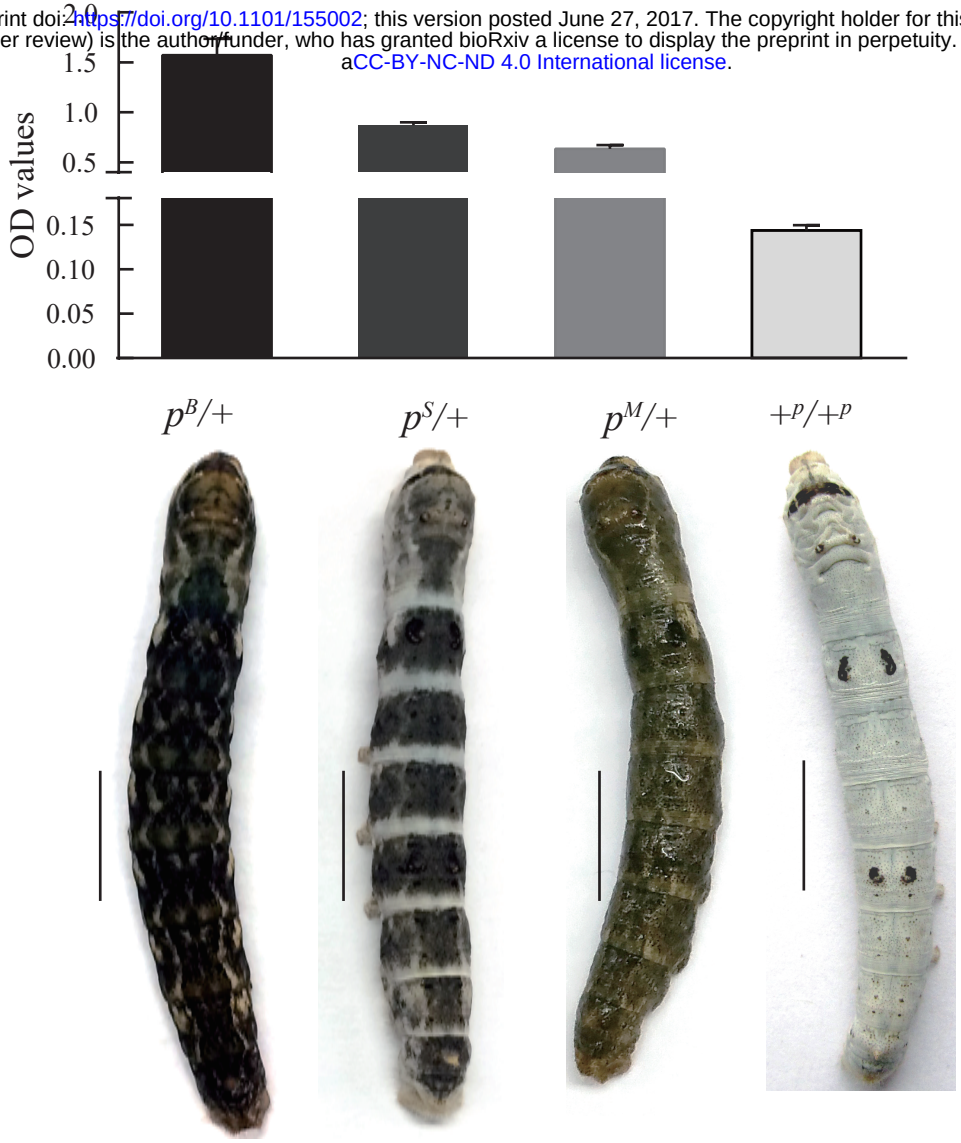


Figure 2

A



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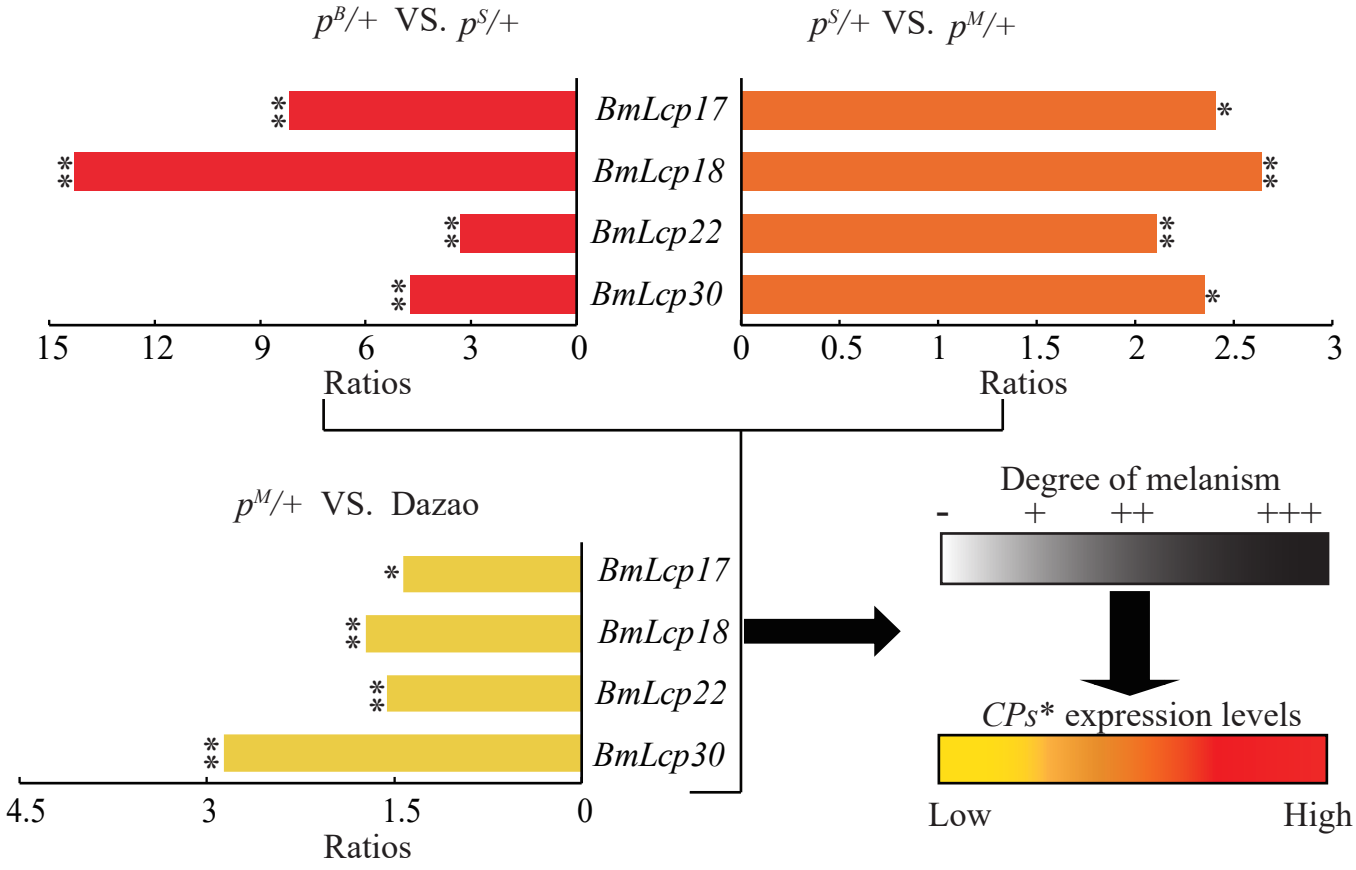
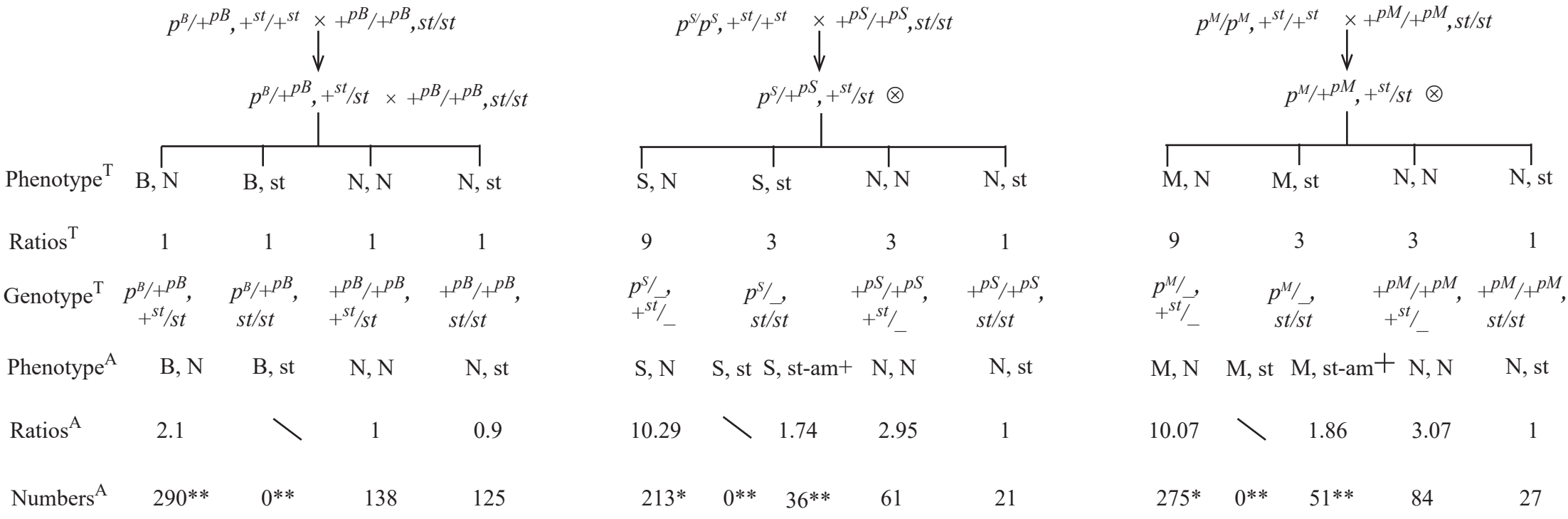


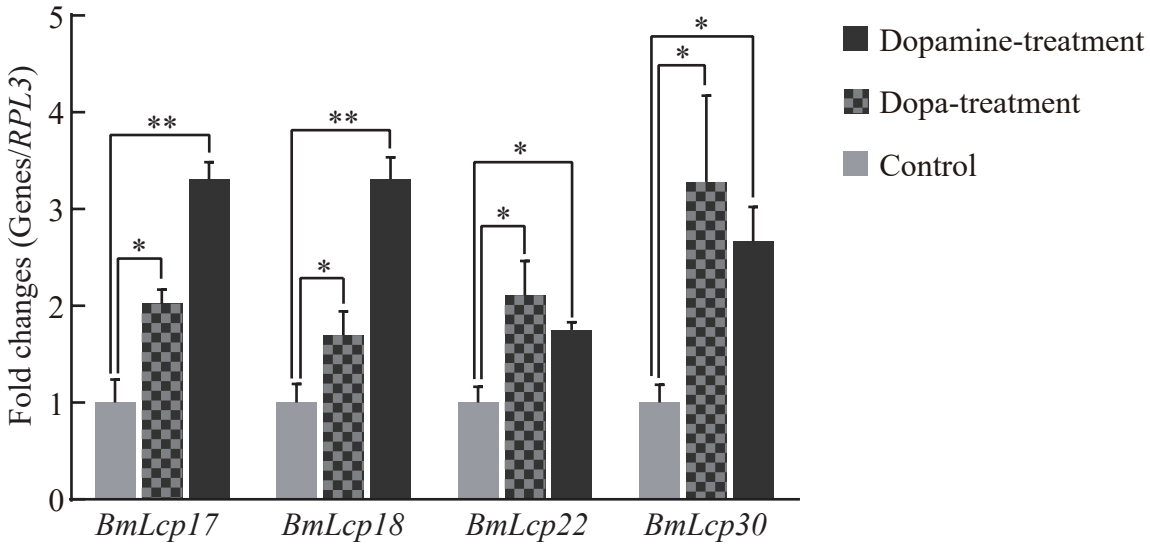
Figure 3

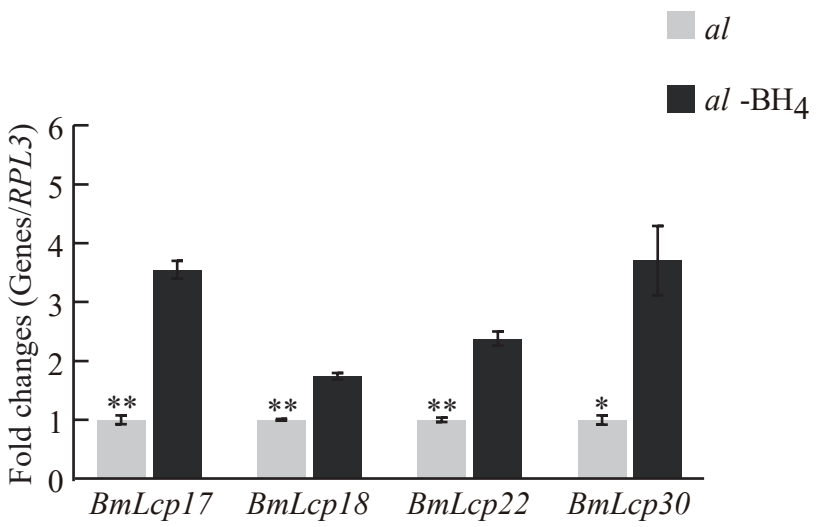


Phenotype^T	B, N	B, st	N, N	N, st	S, N	S, st	N, N	N, st	M, N	M, st	N, N	N, st
Ratios^T	1	1	1	1	9	3	3	1	9	3	3	1
Genotype^T	$p^B/+^{pB}$, $+^{st}/st$	$p^B/+^{pB}$, st/st	$+^{pB}/+^{pB}$, $+^{st}/st$	$+^{pB}/+^{pB}$, st/st	$p^S/\_$, $+^{st}/\_$	$p^S/\_$, st/st	$+^{pS}/+^{pS}$, $+^{st}/\_$	$+^{pS}/+^{pS}$, st/st	$p^M/\_$, $+^{st}/\_$	$p^M/\_$, st/st	$+^{pM}/+^{pM}$, $+^{st}/\_$	$+^{pM}/+^{pM}$, st/st
Phenotype^A	B, N	B, st	N, N	N, st	S, N	S, st	S, st-am+	N, N	N, st	M, N	M, st	M, st-am+	N, N	N, st
Ratios^A	2.1	\	1	0.9	10.29	\	1.74	2.95	1	10.07	\	1.86	3.07	1
Numbers^A	290**	0**	138	125	213*	0**	36**	61	21	275*	0**	51**	84	27

Figure 4

Figure 5





Contents of melanin precursors

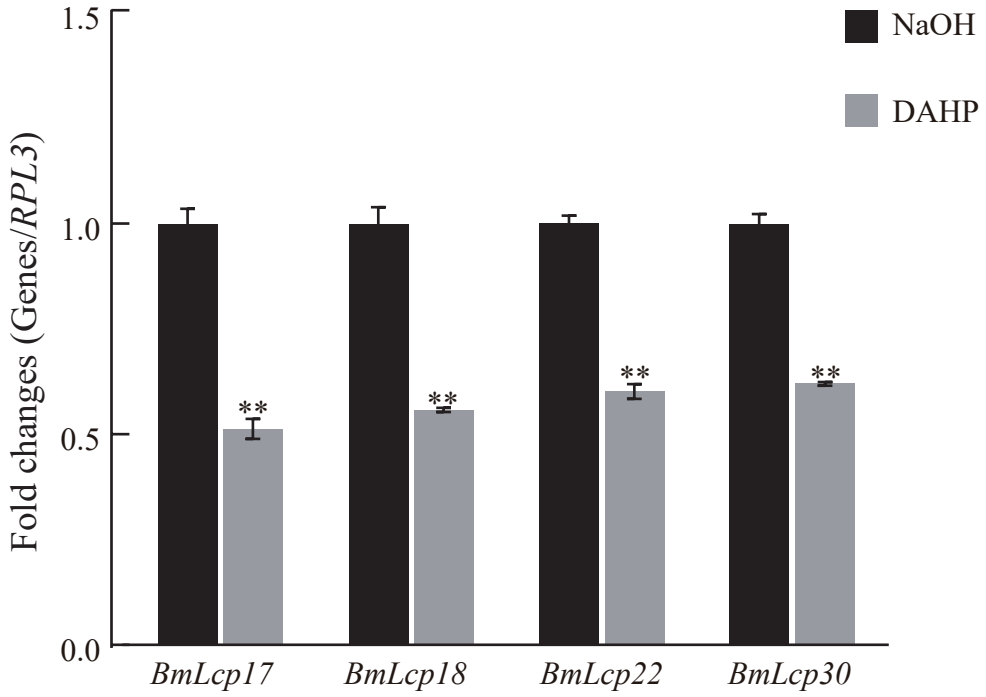


Figure 6