

1 **Spores and soil from six sides: interdisciplinarity and the environmental biology of anthrax**
2 **(*Bacillus anthracis*)**

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35 **Abstract**

36 Environmentally transmitted diseases are comparatively poorly understood and managed, and
37 their ecology is particularly understudied. Here we identify challenges of studying
38 environmental transmission and persistence with a six-sided interdisciplinary review of the
39 biology of anthrax (*Bacillus anthracis*). Anthrax is a zoonosis that is capable of maintaining
40 infectious spore banks in soil for decades (or even potentially centuries), and the mechanisms of
41 its environmental persistence have been the topic of significant research and controversy. In sites
42 where anthrax is endemic, it plays an important ecological role, shaping the dynamics of entire
43 herbivore communities. The complex eco-epidemiology of anthrax, and the mysterious biology
44 of *Bacillus anthracis* during its persistence in the environment, have necessitated an
45 interdisciplinary approach to pathogen research. Here, we illustrate different disciplinary
46 perspectives through key advances made by researchers working in Etosha National Park, a long-
47 term ecological research site in Namibia that has exemplified the complexities of anthrax's
48 enzootic process over decades of surveillance. Through the lens of microbiologists, geneticists,
49 immunologists, ecologists, epidemiologists, and clinicians, we discuss how anthrax dynamics are
50 shaped at the smallest scale by population genetics and interactions within the bacterial
51 communities up to the broadest scales of ecosystem structure. We illustrate the benefits and
52 challenges of this interdisciplinary approach to disease ecology, and suggest ways anthrax might
53 offer insights into the biology of other important pathogens.

54

55 **Key Words:** Anthrax, *Bacillus anthracis*, Etosha National Park, environmental transmission,
56 interdisciplinarity, disease ecology, eco-epidemiology.

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72 **I. Introduction**

73

74 The East Asian parable of the six blind sages, immortalized in John Godfrey Saxe's "The Blind
75 Men and the Elephant," is an apt metaphor for the process of interdisciplinary research and
76 multidisciplinary synthesis. In the story, the sages each attempt to describe to each other an
77 elephant, a new and terrifying beast they have never encountered, based solely on what they can
78 feel. The first touches the elephant's side and describes it like a wall; the second feels the tusks
79 and concludes an elephant is like a spear, and so on. Saxe's penultimate verse thus concludes,

80 *“And so these men of Indostan / disputed loud and long, / each in his own opinion / exceeding*
81 *stiff and strong, / though each was partly in the right, / and all were in the wrong!”*

82 Every new epidemic and (re-)emerging pathogen represents a challenge for medical
83 communities, and like the blind sages, each of these diseases draws researchers together to assess
84 the nature of the beast. The sages brought to the table have of course changed over time: since
85 the Modern Synthesis in evolutionary biology, ecologists have come into play as important
86 counterparts of the “traditional” disease research fields, joining the ranks of microbiologists,
87 immunologists, epidemiologists, evolutionary biologists, and clinicians (among many others).
88 Paradigms for collaborative research like EcoHealth and One Health bring these disparate groups
89 together to achieve interdisciplinary synthesis¹⁻⁴—an important step towards outbreak
90 preparedness, given that the majority of emerging pathogens have some sort of environmental
91 origin. On a global scale, the majority of recently emerging human diseases, including those with
92 environmental reservoirs, originate in animal populations (zoonoses) and, of those, an estimated
93 70% originate in wildlife.⁵ Particularly challenging to study are pathogens that blur the
94 boundaries between direct transmission and indirect modes, including vector-borne transmission
95 and transmission from biotic (wildlife) or abiotic (water or soil) reservoirs. In response to the
96 significant role ecology plays in these modes, multidisciplinarity and interdisciplinarity aim to
97 integrate ecology into disease prevention and break down the barriers that prevent meaningful
98 communication between the “blind sages.” A number of recent high profile works have recently
99 called for better integration of ecosystem research into disease-management efforts^{6,7}, and the
100 need to increase interdisciplinary interaction has been recognized for more than a decade^{3,8}, as
101 evidenced by a number of publications over two decades that call for removing disciplinary
102 barriers in disease research.^{1,3,4,8-13}

103 In the most pronounced success stories, the synthesis of interdisciplinary research has
104 enabled two key advancements: statistical forecasting methods that enable anticipation of
105 outbreaks based on environmental and social data, and ecologically inspired tools for
106 intervention to mitigate and sometimes prevent outbreaks (**Figure 1**). For example, cholera
107 (*Vibrio cholerae*) was once thought to be only directly transmitted human-to-human via
108 contaminated water (a mechanical vector), until 1983 when a team of ecologists and
109 microbiologists discovered that copepods are aquatic hosts of cholera bacteria. Through
110 continued collaboration among ecologists, microbiologists and clinicians, this discovery
111 eventually enabled outbreak forecasting based on climatic data and led to the implementation of
112 simple filtration methods that reduce case burden by as much as 48%. A combination of oral
113 vaccines, water filtration techniques, improved sanitation, and predictive modeling has made the
114 ongoing seventh global pandemic of cholera more manageable than ever before.¹⁴⁻¹⁶

115 Wilcox & Colwell proposed a “cholera paradigm” for interdisciplinary research based on
116 these advances, arguing that even for the most complex and challenging-to-predict systems,
117 synthesis work focused on elucidating multi-component life cycles can help develop both
118 predictive tools and prevention or control measures.¹⁶ But among zoonotic diseases, cholera is
119 characterized by a simple ecology relative to its human health burden, and the key insight of its
120 copepod host was ultimately enough to revolutionize interventions and predictions. Compared to
121 cholera, many pathogens are still poorly understood. Newer or neglected diseases tend to show
122 less-integrated clinical and academic knowledge; and diseases with an uncertain ecology are
123 particularly difficult to control. Generalist pathogens with a complicated or uncertain natural
124 history pose a particular problem for predictive work, and more often than not, the most limiting
125 factor is a dearth of research on their ecology. In a similar undertaking to Wilcox & Colwell’s

126 study, we demonstrate use a multidisciplinary framework to illustrate how “six blind scientists”
127 from different disciplines would characterize recent developments in research on anthrax (*B.*
128 *anthracis*), a generalist pathogen with an extremely complex life cycle compared to cholera. We
129 endeavor to illustrate how and why the ecology of diseases like anthrax is comparatively
130 understudied and undersynthesized, and how interdisciplinary synthesis that includes ecology is
131 especially important for pathogens like anthrax, precisely because there are so many unknown
132 elements of their complex, nonlinear dynamics.

133

134 **II. Anthrax: A Case Study in Slow Integration**

135

136 Anthrax is a zoonosis caused by the gram-positive bacterium, *Bacillus anthracis*, that primarily
137 infects ungulates; other mammals, including humans, tend to be incidental hosts. Transmission
138 takes place through several pathways, the primary one for ungulates being ingestion of *B.*
139 *anthracis* spores during feeding at carcass sites. Other potential pathways include ingesting
140 emesis and feces deposited by necrophagous flies on vegetation after these flies have fed on
141 hosts that have died of anthrax; inhaling anthrax spores that have become airborne, (in nature
142 occurring from dust bathing hosts, though recent evidence cast doubt on this¹⁷); waterborne
143 transmission from waterholes and temporary ponds; cutaneous routes, which account for the
144 majority of human clinical cases globally; and gastrointestinal infections from eating infected
145 meat and blood directly. In some regions, anthrax outbreaks are a natural part of ecosystems and
146 occur predictably on a seasonal cycle; in other settings, epizootics are infrequent events, and can
147 be responsible for mass die-offs among wildlife and livestock.

148 While pandemic threats like Ebola and Zika rightly attract substantial attention in both
149 disease ecology and the public health realms, anthrax maintains a lower profile in terms of
150 cumulative impact on global health, normally only making headlines through frequent bioterror
151 scares. Despite its global presence, and its wide range of suitable hosts, little is known about the
152 dynamics of anthrax in most of the ecosystems and hosts it occupies. The pathogen is best
153 studied in a handful of regions, in particular the Midwestern United States, the former Soviet
154 Union, and sub-Saharan Africa. The insights we review here come from two decades of work
155 based especially in Namibia's Etosha National Park, a savannah ecosystem that is host to high
156 ungulate diversity and endemic anthrax. Data collection regarding anthrax outbreaks in bovids,
157 elephants, zebra, and other mammals in Etosha began in the 1960s, and has provided one of the
158 most continuous sources of documented anthrax dynamics in any natural system.
159 Interdisciplinary work has emerged from treating Etosha as a window into the complex and often
160 hidden dynamics of anthrax spores in the environment, and to date represents one of the most
161 successful ventures to better understand the ecology of the disease. Here, we review six
162 disciplinary perspectives (**Figure 2**) on anthrax dynamics in sub-Saharan Africa, especially in
163 Etosha¹⁸; and highlight how each has contributed to scientific understanding about anthrax's life
164 cycle.

165

166 **(1) Microbiology**

167

168 Louis Pasteur first proposed that carcass sites could function as the main route of anthrax
169 transmission. Long considered an obligate pathogen, *B. anthracis* was thought to replicate only
170 within a vertebrate host, where conditions were conducive for proliferation of vegetative cells.

171 When the host succumbs to the anthrax infection, vegetative cells of *B. anthracis* are released
172 (along with blood and other body fluids) into the environment, and produce infectious spores
173 capable of long-term survival. The environmental maintenance of anthrax is the least understood
174 part of its life cycle; despite the central role of the environment in its transmission, surprisingly
175 little is known about the survival and activity of *B. anthracis* outside of a host. Three
176 fundamental questions have been at the heart of most research on this topic: where and for how
177 long can *B. anthracis* persist in the environment, is it capable of germinating in the soil under
178 any normal conditions, and how does it interact with other microorganisms and plants?

179 How long do anthrax spores persist in the environment? Evidence from a cross-scale
180 study by Turner *et al.*¹⁹ suggests that cultivable *B. anthracis* spore concentrations exponentially
181 decline over time in the soil. In the first two years after a carcass site is formed and spores are
182 deposited, any grazing at carcass sites is likely to infect hosts, even just from eating above-
183 ground plant parts of grasses; anthrax spores tend to decay to negligible concentrations on
184 grasses after more than two years. However, after 2-4 years, anthrax spores still persist in the soil
185 at high enough concentrations that herbivores ingesting soil directly, or indirectly along with
186 grass roots, are probably still at significant risk of infection. Overall, transmission through
187 grazing appears to be most likely in the 1-2 year window when grass growing at former carcass
188 sites is more abundant and nutritious.¹⁹ Nevertheless, data from Etosha shows that spores can be
189 detected for more than seven years after decomposition of the carcass²⁰, and it is likely that
190 longer-term persistence drives anthrax dynamics in other more episodic systems (like western
191 Canada's Wood Bison National Park), where exponential rates of spore decay in more heavily
192 vegetation-covered, less intensively radiated soils may be substantially slower than in Etosha. In
193 some natural conditions, spores are known to persist for decades; spores of the Vollum strain of

194 *B. anthracis* were detected more than forty years after soils were experimentally inoculated at
195 Gruinard Island.²¹ Similarly, re-emergence of anthrax in reindeer in the Yamal region of Siberia
196 in 2016 after more than seventy years after the last known case, together with sporadic cases
197 originating from unknown environmental sources in Sweden, strongly suggests that persistence
198 times can exceed one or more centuries under certain conditions. The rate at which a spore bank
199 decays likely depends heavily on local environmental conditions: for example, larger
200 concentrations of spores are found in soils having slightly alkaline pH, higher organic matter and
201 higher calcium content.²² Features of the exosporium also have been shown to affect the ability
202 of *B. anthracis* spores to bind to different soil types.²³

203 Originally, data from spore-contaminated soil samples in some areas indicated that *B.*
204 *anthracis* has a tendency to lose the pXO2 plasmid (encoding the poly- γ -D-glutamic acid
205 capsule) over time (5-8 years), suggesting that at least a minimum amount of replication, and
206 therefore genetic evolution, takes place in the soil environment; subsequent work has confirmed
207 that *B. anthracis* can in fact replicate in the soil. However, the conditions under which this can
208 happen are highly controversial. Van Ness proposed that under conditions of alkaline pH, high
209 soil moisture, and the presence of organic matter, *B. anthracis* can maintain a high population
210 density by replicating in the environment²⁴. However, this “incubator hypothesis” remains
211 controversial because Van Ness did not provide empirical support. Moreover, laboratory studies
212 suggest that although vegetative cells may potentially flourish outside of a host, their survival in
213 the environment may be significantly influenced by antagonistic interactions with other
214 microbes. In early experimental studies, Minett and Dhanda²⁵ and Minett²⁶ found that *B.*
215 *anthracis* spores germinate and multiply in moist sterile soil but not in soil with a microbially-
216 diverse population, and suggested that bacterial antagonists may restrict its activity. In further

217 work, several species of soil bacteria were found to impede growth of vegetative *B. anthracis*
218 cells²⁷, and a separate study found that some bacteria typically present in soil inhibited
219 multiplication of vegetative *B. anthracis* cells in unsterilized soil.²⁸ Conversely, the effect of *B.*
220 *anthracis* on other soil bacteria has also proved interesting, if complex. In a groundbreaking
221 study, Saile and Koehler demonstrated that *B. anthracis* germinates in the rhizosphere of plants
222 (but did not find evidence for multiplication), suggesting replicative cycles in the rhizosphere of
223 grass plants to play a potential role in the evolution of *B. anthracis* (as it does other members of
224 the *B. cereus* group).²⁹ It is interesting in this respect to note that *B. subtilis* recently has been
225 shown to protect plants against bacterial pathogens in the plant rhizosphere, and that the
226 protective effect requires biofilm formation.^{30,31} Although knowledge is lacking about a potential
227 role for *B. anthracis* biofilm formation, in the rhizosphere or during infection, the bacterium is
228 capable of biofilm formation *in vitro*.³²

229 Recent ecological studies have shown that *B. anthracis* also interacts more broadly with
230 some other members of the grassland-soil community, including plants²⁹, earthworms³³⁻³⁵, and
231 soil amoeba.³⁶ Pasteur was the first to propose that earthworms vector *B. anthracis* from buried
232 livestock carcasses, and he isolated *B. anthracis* from earthworms collected in surface soils at a
233 burial site.^{33,34} Following up on these observations, Schuch and Fischetti³⁵ found that
234 bacteriophages can generate phenotypic changes in *B. anthracis* that enable it to persist as an
235 endosymbiont in earthworms and to act as a saprophyte in soil and water. Under simulated
236 environmental conditions, Dey *et al.*³⁶ showed that a fully virulent Ames strain (pXO1+,
237 pXO2+) of *B. anthracis* germinates and multiplies intracellularly within a free-living soil
238 amoeba living in moist soils and stagnant water, and that the pXO1 plasmid was essential for
239 growth. This may indicate that amoebae and possibly other soil-borne protozoa contribute to *B.*

240 *anthracis* amplification and spore persistence by providing an intracellular environment that
241 allows the completion of a life cycle from spore germination through vegetative growth and back
242 to spore formation. Several other mammalian pathogens (e.g. *Francisella* spp. and *Legionella*
243 *pneumophila*) are known to be capable of parasitic or symbiotic relationships with amoeba^{37,38},
244 and there may be a direct link between a pathogen's ability to survive within amoeba and its
245 ability to survive encounters with primary phagocytic cells (macrophages and neutrophils). Some
246 researchers have even gone as far as to suggest that co-evolution between these bacteria and
247 amoeba seems to have allowed the parasitism of mammals.³⁸

248

249 **(2) Genetics, Genomics, & Evolution**

250

251 *Bacillus anthracis* shares a common chromosomal framework with all six main species of the
252 *Bacillus cereus* (*sensu lato*) super-species group, including the non-pathogenic soil bacterium *B.*
253 *cereus*, and the entomopathogenic *B. thuringiensis*, thereby blurring species boundaries.³⁹ The
254 chromosomal elements principally separating *B. anthracis* (*sensu stricto*) from the other closely
255 related species are: 1) the presence of four distinctive chromosomal prophage elements; 2) a
256 specific, inactivating nonsense mutation in the transcription factor PlcR, a positive regulator
257 mainly of extracellular insect virulence factors; and 3) being part of the genetically
258 monomorphic *B. anthracis* cluster by phylogenetic analysis.⁴⁰ In addition, *B. anthracis* requires
259 two large plasmids for full virulence: pXO1, which encodes the anthrax toxins, and pXO2, which
260 encodes the protective poly- γ -D-glutamate capsule. Large-scale, whole-genome sequencing
261 studies suggest there has been no recent large-scale gene loss in *B. anthracis* or unusual
262 accumulation of non-synonymous DNA substitutions in the chromosome.⁴¹ The fact that *B.*

263 *anthracis* spends large parts of its evolutionary time as a dormant spore (on average the
264 bacterium carries out 0.28–1 generation per year⁴²), presumably contributes to its highly
265 monomorphic nature. During infection of a host, however, mutations accumulate, and it is
266 thought that genetic evolution of *B. anthracis* is mainly limited to the roughly week-long periods
267 between exposure and host death, estimated to cover 20–40 bacterial generations.⁴³ Selection in
268 *B. anthracis* can subsequently act upon phenotype with regards to the spore, based on mutations
269 acquired during its last infective cycle.²²

270 Furthermore, based on genetic data, the *B. anthracis* species can be divided into three
271 distinct subpopulations, the A-, B-, and C-branch, respectively, of which C-branch isolates seem
272 to be strikingly rare, and A-branch isolates have been hugely successful with respect to lineage
273 multiplication and geographical spread worldwide. In addition, a number of different genotypes
274 of *B. anthracis* may be present in endemic regions⁴⁴, which also may potentially give rise to co-
275 infection with more than one genotype.⁴⁵ In the context of the relationship between *B. anthracis*
276 genetics and transmission, however, little is known regarding the relationship between different
277 strains and virulence levels, and hence estimates of LD₅₀ by and large fail to account for variation
278 between different host species, or between different immunological and physiological states of
279 individuals.

280 Although *B. anthracis* is thought to have gone through a genetic bottleneck fixating it as
281 a genetically monomorphic pathogen, it is interesting to note that consolidation of clinical and
282 eco-evolutionary (DNA sequencing) data indicates that what presents as “anthrax” also may
283 include specific isolates of *B. cereus* (*B. cereus* biovar *anthracis*), causing opportunistic anthrax-
284 like infections in humans and great apes.^{46,47} During the past decade, several *B. cereus* group
285 variants have been isolated from cases of human or animal infections involving anthrax-like

286 symptoms and/or disease. These cases are genetically different from classical *B. anthracis* in that
287 they carry the pXO1 and pXO2 (or other capsule-producing) plasmids in a *B. cereus*
288 chromosomal background.^{46,48,49} The recurring presence of such strains in natural settings shows
289 that active transfer of pXO1 and pXO2 variants was not a one-off evolutionary event for a
290 lineage leading up to *B. anthracis*. On the contrary, these observations allow for the possibility of
291 pXO1 and pXO2 transfer into *B. cereus* having taken place as multiple independent evolutionary
292 events, leading to multiple lineages of strains with the capacity to evolve into contemporary
293 strains of variable genetic composition, but producing similar anthrax-like symptoms during
294 human or animal infections. However, it is interesting to note that the *B. cereus* *vs anthracis*
295 isolates from animals in Cote d'Ivoire, Cameroon, Central African Republic and Democratic
296 Republic of Congo appear to belong to the same phylogenetic clade, and that the pXO1 and
297 pXO2 plasmids hosted by the strains followed the same phylogenies with the same branching
298 order.¹⁸ These data indicate that both the chromosomes and the plasmids in the isolated strains
299 are closely related. In addition, strains isolated from Cote d'Ivoire and Cameroon were tested in
300 toxicity and vaccination experiments and found to be just as toxic as fully virulent *B. anthracis*,
301 and protective vaccination in mice and guinea pigs were just as efficient as for *B. anthracis*.⁵⁰

302

303 **(3) Immunology**

304

305 Anthrax infections end either in recovery, or death of the host. When *B. anthracis* spores are
306 ingested, the spores germinate into fast-multiplying vegetative forms that produce three soluble
307 factors that assemble to form toxic complexes: edema factor (EF), an adenylate cyclase that
308 impairs immune cell function⁵¹⁻⁵⁴; lethal factor (LF), a zinc metalloprotease that cleaves MAP-

309 kinase-kinases thereby suppressing production of several types of cytokines and immune cell
310 functions⁵⁵⁻⁶⁰; and protective antigen (PA), which complexes with the other two factors and
311 allows them to enter host cells through oligomeric PA pores.⁶¹ The PA and LF bind to form the
312 anthrax lethal toxin (LeTx), the key virulence factor of *B. anthracis* that kills macrophages and
313 dendritic cells through a caspase-1-dependent cell death program known as pyroptosis.⁶² The
314 collective actions of these toxins may ultimately result in the peracute-to-acute death of
315 susceptible hosts from edema, vascular collapse, and inflammation, combined with an
316 overwhelming septicemia of up to 10^9 bacterial cells per milliliter of blood.⁶³⁻⁷⁰

317 Lethal dose is a host-population concept, and within populations, hosts will vary in their
318 susceptibility to anthrax because of inherited genetic factors, as well as current immunological
319 status, coinfection, and physiological condition. Most studies regarding host immune responses
320 to anthrax have been conducted in laboratory settings. These studies have demonstrated that
321 humoral immunity, particularly against the PA toxin, plays a very important role in a host's fight
322 against anthrax; the presence of anti-PA antibodies appears to be essential for adaptive
323 protection, and several studies have demonstrated that the magnitude of a host's anti-PA IgG
324 antibody titer is correlated with level of protection against the disease.⁶³⁻⁷⁰ Furthermore, anthrax
325 vaccine studies have indicated that T cells may also play a role in immunity to anthrax.⁷¹ While
326 anthrax spores require phagocytosis by macrophages for germination, macrophages have also
327 been found to play a primary role in limiting and clearing anthrax infection.^{72,73} Following
328 infection, macrophages engulf and destroy invading pathogens, recognizing *B. anthracis* through
329 toll-like receptor 2 (*TLR2*).⁷⁴ Genetic studies in mouse models and cell lines have identified
330 several host genes that modulate susceptibility to *B. anthracis* infection and support a multigenic
331 contribution to the host response.⁷⁵ The myeloid differentiation factor (*myD88*), a downstream

332 mediator of the TLR pathway, has been shown to confer susceptibility to anthrax⁷⁶, and
333 polymorphisms in the *Nlrp1b* (*Nalp1b*) gene have also been shown to influence susceptibility to
334 the anthrax toxin in mouse macrophages⁷⁷ and human fibroblasts.⁷⁸ However, other studies have
335 demonstrated that the LT-sensitive *Nlrp1b* allele induces early inflammation that protects against
336 anthrax.^{79,80} *TEM8* and *CMG2* genes encode host transmembrane proteins that function as
337 anthrax LeTx receptors^{81,82}, binding with PA and mediating delivery of LF into host cells. *CMG2*
338 has a considerably higher affinity for LeTx than does *TEM8*, and *CMG2*-null mice are highly
339 resistant to *B. anthracis* infection.⁸³ In fact, *CMG2* variation significantly alters toxin uptake and
340 sensitivity in humans, with lethality differing up to 30,000-fold among cells from people of
341 different ethnic backgrounds.⁸⁴

342 While these studies provide a wealth of mechanistic knowledge about the host's
343 immunological response to anthrax, the scaled application of that information to anthrax
344 dynamics is far more complex—especially as laboratory studies on mice can only reveal so much
345 about the dynamics of infection in large herbivores. To remedy this, field studies are needed to
346 address gaps in our understanding of anthrax infections, and of how immunology scales up to
347 produce broader eco-epidemiological patterns. One particularly important problem is the effect
348 that sub-lethal doses may have in promoting adaptive immune responses to anthrax.^{85–89} Recent
349 evidence suggests that sublethal anthrax infections in species known for high apparent
350 mortality—including the herbivores like zebra and springbok that are most abundant and most
351 important in anthrax outbreaks in Etosha—are more common than previously thought.⁹⁰ In fact,
352 frequent anthrax contact can act as an immunity booster in both carnivores and herbivores,
353 strengthening their anti-anthrax protection over time and possibly lessening the overall morbidity
354 and mortality within the population (endemic stability).^{89,90} A similar key problem is that anthrax

355 infections exist in an ecosystem of pathogens, and the within-host role that co-infection plays in
356 anthrax immunology is complicated at best. Field studies in Etosha have shown apparent
357 tradeoffs in zebra between Th-1 and Th-2 type immune responses, where Th-2 responses seem to
358 peak in the wet season in response to gastrointestinal helminths.⁹¹ These immune responses
359 appear to make zebra and springbok more tolerant of helminth infections when they peak during
360 the wet season, decreasing host resistance to anthrax infection and thereby potentially
361 contributing to the overall seasonality of anthrax outbreaks.⁹²

362

363 **(4) Ecology**

364

365 Landscape ecology research on anthrax has had the greatest successes by studying the locations
366 and processes of herbivorous hosts that have died of anthrax (**Figure 3**). These carcass sites act
367 as “locally infectious zones” (LIZs), and come to have a demography of their own as these zones
368 appear and fade over time. Rather than passively acting as a fomite, evidence suggests that
369 anthrax carcass sites have a complex set of biotic interactions that determine their persistence
370 and infectiousness throughout a landscape. Nutrient deposition from carcass decomposition
371 appears to be the primary correlate of overall plant growth in green-ups; zebra carcasses in
372 Etosha substantially increase soil phosphorus and nitrogen that persists over at least three years.¹⁹
373 However, experimental evidence suggests *B. anthracis* spores facilitate the germination of grass
374 seeds.¹⁹ The mechanism through which that occurs is still uncertain, but Saile and Koehler
375 demonstrated that anthrax germinates in the plant rhizosphere²⁹; as *B. anthracis* is a member of
376 the *B. cereus* group, it is possible that *B. anthracis* retains some of its ancestral capabilities to
377 engage in beneficial plant-microbe interactions. There has been no evidence, however, that

378 plants facilitate persistence of *B. anthracis* in the soil, potentially suggesting that its association
379 with vegetation may serve to attract herbivores that ultimately become infected and spread the
380 pathogen across the landscape.⁹³

381 Herbivores in Etosha face a tradeoff between the benefits of foraging at green-ups and the
382 obvious costs associated with lethality, with possible selective pressure acting on foraging
383 strategy.^{19,94} Evidence based on camera trap data from Etosha suggests that most herbivores
384 avoid carcass sites early in the first year of establishment because they are denuded of vegetation
385 by scavengers, but that they in fact favor green-ups following the first rains after the nutrition
386 influx from the carcass. This likely contributes to the importance of the 1-3 year window after
387 establishment in transmission.¹⁹ Anthrax dynamics are also seasonal within years, peaking in
388 March-April, which more generally aligns with the later part of the warm wet season in Etosha.
389 While some research previously suggested that nutritional stress might drive the seasonality of
390 anthrax, evidence directly contradicts the idea that nutritional stress is worse during the anthrax
391 season.^{92,95} Instead, it appears that soil ingestion increases during the wet season for a handful of
392 species including zebra, directly increasing anthrax exposure; in contrast, elephant deaths (while
393 rare) in fact peak in October-November, suggesting there are interspecific heterogeneities in
394 exposure pathways that still require investigation. Similarly, interspecific variation in movement
395 still requires investigation, given the wealth of movement data collected in Etosha over the past
396 two decades; for example, elephants largely migrate away from the known anthrax areas of
397 Etosha during the anthrax season, and return in the dry season. Intraspecific variation also
398 requires further investigation study; evidence suggests that there may be a link between partial
399 migration of zebra herds in Etosha and avoidance of the anthrax season. It has been suggested

400 that this phenomenon could be linked to dominance structure, as dominant groups migrate, while
401 resident (submissive) herds are encouraged to stay by decreased competition.⁹⁶

402 How do non-herbivorous mammals affect anthrax dynamics? Some work had suggested
403 that scavengers might play an important role in the dispersal and creation of LIZs, but work in
404 Etosha has often proved counter to those ideas. Previous theory had suggested vultures might
405 disperse bacteria from carcass sites to their nesting sites and thereby help spread disease.
406 However, research in Etosha failed to find higher *B. anthracis* concentrations at anthrax nests,
407 perhaps because vultures' acidic droppings produce soil unsuitable for anthrax spores.⁹⁷
408 Furthermore, prior work indicates that during the first 72 hours after carcass deposition, if a
409 carcass remains unopened, vegetative cells fail to sporulate, ending the life cycle. Consequently,
410 scavengers were likely to play a significant role in anthrax dynamics, by tearing open carcasses
411 and promoting blood flow into the soil. In contrast, an alternative hypothesis suggested that
412 scavengers—especially vultures and other birds, which are less prone to anthrax-related deaths
413 due to acquired immunity⁸⁶—could “cleanse” carcass sites, reducing LIZ formation and
414 establishment. However, experimental exclusion of scavengers from zebra carcasses in Etosha
415 revealed that scavengers had no effect on soil spore density, failing to find evidence for either
416 hypothesis, and further challenging the role of scavengers in anthrax ecology.⁹⁸

417

418 **(5) *Epidemiology***

419

420 The epidemiology of directly-transmitted pathogens draws upon ecology, particularly behavioral
421 ecology, to better understand how susceptible and infected individuals come into contact with
422 one another; by comparison, the epidemiology of environmentally transmitted pathogens, such as

423 anthrax, requires a much wider understanding of the relevant host and pathogen ecologies,
424 particularly the interactions of hosts and pathogens within their environments. Thus, in the case
425 of indirect transmission, it is more difficult to separate the ecological and epidemiological
426 components. Appropriately complex epidemiological models are needed that can unpack
427 different aspects of transmission, including the dose of pathogen that hosts are exposed to, the
428 immunological variation between individual hosts and between species, and the interplay
429 between the two. Modeling the dose-exposure process requires an understanding of individual
430 pathogen shedding into the environment, the movement of susceptible individuals through space,
431 the internal milieu of the host, and in some cases the behavior of susceptibles once the source has
432 been encountered. However, studies that explicitly consider the movement of individuals across
433 landscapes, the ability of pathogens to persist in the environment, the immunological status of
434 susceptible individuals, and issues of dose or prior low-dose exposure are rare.

435 Studies in Etosha have provided the tools to begin to develop models that cross these
436 different scales, and thereby unravel the false “lethal dose paradox,” in which the experimentally
437 determined lethal dose required to kill herbivores appears to be far higher than would be
438 encountered in nature. Work that combines field experiments and modeling shows that,
439 especially in the first two years after deposition, carcasses should provide ample infection risk
440 for grazing herbivores, even when soil ingestion is minimal.²⁰ Even though spore concentrations
441 begin to decline rapidly after two years, they may still be sufficient to produce sporadic
442 outbreaks, especially in drought years that intensify herbivore soil contact during grazing. These
443 small outbreaks may set off (and often precede) epidemic years, illustrating how the long tail of
444 LIZ persistence could ultimately play an important role in long-term anthrax dynamics.²⁰ Studies
445 like this allow future work that considers the epidemiology of anthrax at large scales more

446 explicitly, such as by using agent-based models to simulate the effects of host heterogeneity and
447 landscape structure on outbreak dynamics, and measuring the relative utility of different host
448 movement metrics as predictors of anthrax risk.

449 While the epidemiology of anthrax in Etosha has recently been successfully modeled (at
450 least, in a piecemeal fashion²⁰), Etosha is only a single landscape, and anthrax outbreaks behave
451 very differently around the world. The reasons for differences in the frequency, timing, and
452 intensity of anthrax outbreaks globally are poorly understood, but stem from some combination
453 of microbiological, immunological, and ecological factors discussed above. Not all possible
454 transmission modes are important in Etosha; for example, vector enhancement by necrophagous
455 flies has been implicated as an important mode of spores being spread from carcasses onto
456 above-ground vegetation, but appears to play a minimal role in Etosha. However, some universal
457 patterns can be noted. For instance, soil type and alkalinity is known to affect spore persistence²²,
458 and other climatic conditions may play roles in determining when and how often outbreaks
459 occur. Anthrax outbreaks in the middle latitudes appear to be seasonal across host systems.^{1,14,15}
460 For example, deer outbreaks in Texas appear in summer months¹⁶, with the severity of outbreaks
461 increasing in response to early and intense spring green-up.¹⁷ Similarly, anthrax outbreaks tend
462 to be observed in Etosha (and elsewhere) with dry conditions following periods of intense
463 rainfall, for a number of potential reasons, including changing animal movement patterns
464 (without water-restrictions, animals range more widely at lower densities, whereas in drier
465 periods, they aggregate at waterholes), changing vegetation growth or processes changing spore
466 density on vegetation (such as splashing of spore-laden soil onto grasses)⁹⁵, and increasing
467 exposure to other potentially interacting microparasites and macroparasites (altering host
468 susceptibility⁹²).

469

470 *(6) Clinical & Public Health*

471

472 Like all environmentally transmitted diseases, anthrax poses a unique challenge for surveillance
473 efforts, as the majority of anthrax dynamics are unobserved either due to the difficulties of
474 studying anthrax in the soil, or the limited resources available for epizootic surveillance. In
475 Etosha, the acuteness of anthrax infections makes the window of detectability very narrow for
476 infected animals, and even once carcasses are deposited, many are not found for days or are
477 never found at all, depending on scavenger presence and the location of death. Even still, the
478 Etosha site has been nearly unique in the depth and detail of coverage, with over 50 years of data
479 collection. Anthrax is globally cosmopolitan, and outbreak data for human, wildlife and
480 agricultural cases are most commonly collected from passive surveillance following both
481 wildlife and livestock mortality. While these data are limited, they can be used in combination
482 with our understanding of anthrax dynamics at local scales to create public health-relevant
483 predictive infrastructure. In particular, while the subtleties of community ecology and herbivore
484 movement may influence anthrax landscape ecology at the scale of Etosha, the microbiological
485 factors that determine spore production, dispersal, persistence and amplification will ultimately
486 determine the location and persistence of LIZs at much more flexible spatial scales, and these
487 abiotic constraints ultimately determine broader-scale patterns of presence or absence.

488 Through the use of ecological niche models (ENMs), anthrax occurrence data can be used
489 to study and predict the environmental covariates driving persistence at continental, national, and
490 sub-national scales. Studies using ENMs to map anthrax have been done in at least 11 countries,
491 including Australia⁹⁹; Cameroon, Chad, and Nigeria¹⁰⁰; China¹⁰¹; Italy and Kazakhstan¹⁰²;

492 Kyrgyzstan¹⁰³; Mexico and the United States¹⁰⁴; and Zimbabwe.¹⁰⁵ However, it is worth noting
493 that while anthrax is effectively cosmopolitan on a global scale, no global map of its distribution
494 has ever been constructed. Instead, most studies have mapped its distribution using ENMs
495 constructed at the regional or national scale, often in close partnership with public health efforts.
496 ENM-based methodology is also incredibly flexible, and can be used in combination with other
497 tools such as resource selection functions to improve predictions of how herbivore movement
498 drives cases, or hotspot analyses to study clusters of human and livestock case data. By
499 combining predictive understanding of environmental persistence (with model selection and
500 variable selection heavily informed by studies and understanding at the microbiology and
501 landscape ecology scale) with studies at the human-wildlife-livestock interface, regional
502 surveillance tools can be developed that appropriately map anthrax risk. These studies can even
503 be used to project future scenarios, including the role climate change will play in altering anthrax
504 transmission.¹⁰⁶

505 Anthrax eradication is, for any given landscape, an essentially impossible task given the
506 soil spore banks and the often-cryptic enzootic process. However, animal vaccination programs,
507 carcass removal and avoidance of high-risk locations have been shown to greatly improve
508 regional outcomes. Where combined with surveillance, public health and veterinary
509 infrastructure to deal with outbreaks when they occur, regional patterns of emergence have been
510 kept intermittent and low-impact. An anthrax vaccine is currently available for both animals and
511 humans⁶⁷, but its memory response—as well as the memory response to natural, sub-lethal
512 anthrax infection—tends to remain elevated for only a few months^{70,90}, the reasons for which are
513 unknown. In addition to vaccination, control efforts focus on sanitary carcass disposal. However,

514 in areas where dead animals will not be discovered before body fluids have leaked into the
515 ground, the success is limited, as soil sterilization is costly and inefficient.

516 Around the world, control efforts are highly variable, and necessarily correspond to the
517 local ecology. In the Etosha, control efforts centered on sanitizing of carcasses, but the
518 remoteness of the area, lack of local firewood sources and fragility of vegetation to heavy
519 machinery caused control efforts to be discontinued in the 1980's. Since then, anthrax has been
520 endemic and seen as a natural part of the ecosystem with annual outbreaks in wildlife in the area.
521 All host species seem capable of keeping stable populations however, though elephants may be
522 vulnerable due to fluctuations in their smaller population when combined with poaching
523 pressure. In Russia, anthrax is known as the “Siberian plague” (сибирская язва) due to its
524 historical high prevalence in Siberia. It has been largely controlled in the last half century due to
525 large-scale vaccination of domestic reindeer herds, combined with efforts at tracking and
526 avoiding burials of infected animals. In the Yamalo-Nenets region, reindeer vaccination efforts
527 started in 1928 were discontinued in 2007 because no new cases had been observed since 1941.
528 Following unusual permafrost thawing in the summer of 2016, three simultaneous anthrax
529 outbreaks killed 2500 reindeer and caused the culling of several hundred thousand more during
530 control efforts. A hundred people were hospitalized and one boy died. Further developments as
531 more permafrost thaws are under investigation, but the spore banks of Siberia seem unlikely to
532 be eradicated in even if every new carcass was sanitized new infections will occur for the
533 foreseeable future. In the industrialized countries of western Europe, large-scale anthrax
534 outbreaks have been absent in modern times due to sanitation and vaccination, but even in
535 Sweden the summer of 2016 saw outbreaks in domestic cattle from old environmental sources of
536 unknown locations. The outbreaks were rapidly controlled through the normal efforts of carcass

537 disposal, culling, and vaccinations, but it remains clear that environmental spore banks will
538 continue to exist indefinitely even in modern agricultural areas, ready to emerge as epidemics
539 should veterinary infrastructure falter.

540

541 **III. Conclusions**

542 In the face of global change, hidden rules that have produced extant landscapes of disease (on
543 which theories are based) are liable to change, producing patterns that current interdisciplinary
544 syntheses will sometimes fail to anticipate. In the face of these accelerating threats, we worry
545 that the pace at which knowledge is collected and synthesized for pathogens like anthrax is not
546 sufficient to keep pace, even with emerging interdisciplinary frameworks like One Health. The
547 perspective on anthrax dynamics we present here has been loosely modeled off previous work on
548 cholera, which has been widely noted as a model for One Health, interdisciplinarity, and the
549 value of ecologists' involvement in global health. Could anthrax be a similar template for the
550 value of One Health-oriented investigative research? And if so, how far does that model extend?
551 Is the relevance of the “anthrax paradigm” only significant for other environmentally maintained
552 pathogens like plague and brucellosis, or are there broader lessons for pandemic prediction and
553 prevention in anthrax ecology? To borrow a commonly formatted question from the literature,
554 can anthrax tell us anything about Ebola or Zika, and similar prominent threats to global health?

555 To an even greater extent than is true for better-studied pathogens like malaria or cholera,
556 data on anthrax is still incomplete, and established knowledge is subject to change in many
557 disciplinary perspectives (**Figure 4**)—and the overall integration of knowledge is comparatively
558 limited. Work from Etosha highlights the strength of interdisciplinary research on the
559 environmental biology of neglected diseases like anthrax; the ecological and epidemiological

560 insights we discuss above have the potential to develop tentative early warning systems, or
561 encourage the development of ecologically-minded interventions. Even though eco-
562 epidemiological insights are usually somewhat place-specific, lessons from Etosha may still
563 provide insights outside of savannah ecosystems. For example, a recent heat wave passing
564 through Siberia released at least three separate outbreaks of *B. anthracis* preserved in now-
565 melting permafrost. While little is currently known about the dynamics of anthrax in changing
566 tundra ecosystems, work in Etosha still suggests basic rules for system dynamics, addresses basic
567 information about pathogen biology and genetics, and offers a template for experimental and
568 modeling work needed to understand the potentially different and unexpected eco-
569 epidemiological characteristics of anthrax in a novel ecosystem. Similarly, the frameworks that
570 have been used to study *B. anthracis* in Etosha could be invaluable to develop a research
571 program assessing how widespread *B. cereus* biovar *anthracis* is in West and Central Africa, the
572 basic aspects of its transmission, and the magnitude of a threat it poses to conservation,
573 agriculture, and human health.

574

575 *(1) Lessons from Anthrax for Studying Environmental Transmission*

576 On a broader scale, the seminal novelty of our multi-decade work on anthrax is a deeper
577 understanding of how environmental maintenance and transmission affect the biology of a
578 complex, multi-host pathogen. This has substantial relevance to research on other pathogens,
579 including some of the most serious threats to human health; while some diseases like anthrax are
580 primarily characterized by environmental modes of transmission, a far greater diversity of
581 diseases are occasionally maintained in fomites and reservoirs. The questions we have
582 highlighted here apply to any of these systems, and highlight key uncertainties in the role

583 environmental transmission plays in pathogen life cycles. Many other bacteria appear to be
584 capable of dormancy for extended periods of time, with no reproductive activity, within soil or
585 aquatic environments; but our work here shows that environmental persistence can often be
586 longer and more complex than thought. For example, recent work has confirmed the
587 *Mycoplasma bovis*, a bacterial pathogen associated with mastitis in cattle and bison, can persist
588 for long periods and possibly replicate, most likely through the formation of biofilms associated
589 with gram negative bacteria, in sandy soil used as cattle bedding under certain moisture
590 conditions.¹⁰⁷ Even plague (*Yersinia pestis*), conventionally studied as a vector-borne disease,
591 has been recently shown to persist in the soil for weeks.¹⁰⁸ In cases like these, the role that soil
592 microbiota play in the dynamics and duration of persistence is predominantly unexplored and
593 could represent a key future research direction. Similarly, specific environmental conditions,
594 such as soil alkalinity, moisture, or specific mineral content, are required to allow environmental
595 maintenance (possibly not through direct toxicity to the spores but through shaping the microbial
596 community they interact with)—but for some systems, such as plague, those factors are still
597 understudied or entirely untested. Ecological niche modeling tools that have been used to map
598 anthrax persistence could easily be applied to other soil-borne bacteria, to elucidate the role of
599 different drivers in persistence landscapes, and extrapolate transmission risk from
600 microbiological knowledge.

601 From an eco-epidemiological angle, the role of environmental transmission in pathogen
602 dynamics remains understudied and rarely modeled, especially in the case of diseases for which
603 environmental transmission is not the primary mode of transmission.¹⁰⁹ Key epidemiological
604 concepts like R_0 rarely have the ability to accommodate environmental transmission, especially
605 for generalist bacterial pathogens like anthrax. Environmental maintenance has a substantial

606 effect on epidemiological dynamics even when a small part of a pathogen life cycle; for instance,
607 Lowell *et al.* examined the spatial genetic diversity of plague in the western U.S. and
608 demonstrated that widespread plague epizootics are driven by local persistence in the soil for up
609 to weeks at a time, a finding that can inform anticipatory surveillance of local factors (e.g.
610 climate) known to increase plague outbreak risk.¹⁰⁸ Unusual consequences of environmental
611 transmission are also especially important; for example, horizontal gene transfer from
612 environmental reservoir strains can cause abrupt changes in human outbreaks. Just as pathogenic
613 *B. cereus* outbreaks can be driven by long-term genetic cross-talk with *B. anthracis*, in the case
614 of cholera, virulence genes transferred between benign and pathogenic strains in aquatic
615 reservoirs are hypothesized to be the cause of some epidemics¹¹⁰; genotype surveillance of this
616 evolutionary process, focused on virulence-mediating genes, offers a promising predictive tool
617 for severe outbreaks.¹¹¹

618 While bacterial and macroparasitic diseases more commonly evolve environmental
619 modes of transmission, environmental persistence also plays an important role in the spatial
620 epidemiology of some viruses, such as Hendra virus, for which viral shedding locations become
621 spillover sites between bats and horses.¹¹² Influenza can persist in waterways, and just like
622 regulatory cross-talk has been important for anthrax and cholera outbreaks, strains of influenza
623 that circulate in aquatic reservoirs and wild birds are likely to contribute new pathogenic strains
624 to domestic poultry and potentially humans.¹¹³ Similarly, strains of polio can be transmitted in
625 the environment from the live oral polio vaccine (OPV), leading to recurrent environmentally
626 transmitted outbreaks that reinitiate polio transmission.¹¹⁴ Even many prionic diseases have an
627 environmental transmission mode, such as scrapie¹¹⁵ or chronic wasting disease¹¹⁶, which persist
628 in the soil for unknown durations, historically making pasture unusable for decades. As prionic

629 diseases continue to spread through wild ungulates, and as grazing land infringes on natural
630 areas, ecologists will be tasked with identifying and tracking agriculturally unsuitable, prion-
631 contaminated land. If prions bond differently to different soil or vegetation types (as they appear
632 to do^{117,118}), or host heterogeneity and movement determine the distribution of environmental
633 reservoirs, the same One Health approaches that have succeeded in tracking anthrax
634 emergence¹¹⁹ will need to be applied to the challenging problem of prion surveillance.

635

636 *(2) Lessons from Anthrax for Integrative Thinking*

637 On a broader scale, our case study highlights the need for targeted interdisciplinarity in
638 disease ecology. Interdisciplinary work, especially at long-term ecological research sites, has the
639 potential to revise key ideas about pathogen biology and illuminate the hidden dynamics of
640 pathogens in the environment. Some pathogens, like plague and cholera, are now well enough
641 understood that forecasting can be done successfully across scales, ranging from local early
642 warning systems to global projections under climate change. Anthrax poses a comparatively
643 more serious challenge especially at broader scales, as locally developed scientific understanding
644 becomes less transferrable; thus, expanding research across ecosystems with different dynamics
645 and local drivers to extract generalities of environmental transmission dynamics represents a key
646 next direction for synthesis work. But the majority of threats to public health are nowhere near
647 that stage of synthesis. Ebola virus's reservoirs are still uncertain, and the drivers of Ebola
648 outbreaks have been recently studied but remain controversial at best.¹²⁰⁻¹²² The enzootic cycle
649 of Zika is even more poorly studied; the role primate reservoirs play in the enzootic process has
650 been the subject of some speculation, but the ecology of the disease in its native range (Africa
651 and potentially south Asia) remains essentially undocumented. The scope of complexity inherent

652 to these pathogens' life cycles cannot be fully understood until the enzootic process is better
653 studied; and the ongoing value of interdisciplinarity as a tool for organizing that research is clear.
654 We caution against a focus on research that pushes the cutting edges of disparate fields in
655 isolation, which risks overlooking important insights gained from tying disparate fields together,
656 and could leave the task of synthesis to policy makers with little or no scientific training. In the
657 face of global change, interdisciplinary research is the only option for more rapid advances that
658 keep pace with the accelerating threats that public health must face.

659

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667 **V. References**

668

- 669 1. Manlove, K. R. *et al.* ‘One Health’ or Three? Publication Silos Among the One Health
670 Disciplines. *PLoS Biol* **14**, e1002448 (2016).
- 671 2. Coker, R. *et al.* Towards a conceptual framework to support one-health research for policy
672 on emerging zoonoses. *Lancet Infect. Dis.* **11**, 326–331 (2011).
- 673 3. Parkes, M. W. *et al.* All hands on deck: transdisciplinary approaches to emerging infectious
674 disease. *EcoHealth* **2**, 258–272 (2005).
- 675 4. Borer, E. T. *et al.* Bridging taxonomic and disciplinary divides in infectious disease.
676 *EcoHealth* **8**, 261–267 (2011).
- 677 5. Jones, K. E. *et al.* Global trends in emerging infectious diseases. *Nature* **451**, 990–993
678 (2008).
- 679 6. Johnson, P. T., De Roode, J. C. & Fenton, A. Why infectious disease research needs
680 community ecology. *Science* **349**, 1259504 (2015).
- 681 7. Ezenwa, V. O. *et al.* Interdisciplinarity and infectious diseases: An ebola case study. *PLoS*
682 *Pathog* **11**, e1004992 (2015).
- 683 8. Wilcox, B. A. & Colwell, R. R. Emerging and reemerging infectious diseases:
684 biocomplexity as an interdisciplinary paradigm. *EcoHealth* **2**, 244 (2005).
- 685 9. Wilcox, B. A. & Gubler, D. J. Disease ecology and the global emergence of zoonotic
686 pathogens. *Environ. Health Prev. Med.* **10**, 263–272 (2005).
- 687 10. Daszak, P. *et al.* Collaborative research approaches to the role of wildlife in zoonotic
688 disease emergence. in *Wildlife and emerging zoonotic diseases: the biology, circumstances*
689 *and consequences of cross-species transmission* 463–475 (Springer, 2007).

- 690 11. Levin, B. R., Lipsitch, M. & Bonhoeffer, S. Population biology, evolution, and infectious
691 disease: convergence and synthesis. *Science* **283**, 806–809 (1999).
- 692 12. Moore, C. G. Interdisciplinary research in the ecology of vector-borne diseases:
693 Opportunities and needs. *J. Vector Ecol.* **33**, 218–224 (2008).
- 694 13. Plowright, R. K., Sokolow, S. H., Gorman, M. E., Daszak, P. & Foley, J. E. Causal
695 inference in disease ecology: investigating ecological drivers of disease emergence. *Front.*
696 *Ecol. Environ.* **6**, 420–429 (2008).
- 697 14. Huq, A. *et al.* Ecological relationships between *Vibrio cholerae* and planktonic crustacean
698 copepods. *Appl. Environ. Microbiol.* **45**, 275–283 (1983).
- 699 15. Colwell, R. R. *et al.* Reduction of cholera in Bangladeshi villages by simple filtration. *Proc.*
700 *Natl. Acad. Sci.* **100**, 1051–1055 (2003).
- 701 16. Colwell, R. R. Global climate and infectious disease: the cholera paradigm. *Science* **274**,
702 2025 (1996).
- 703 17. Barandongo, Z. R. Dust bathing behaviours of Elephants, Zebras and Wildebeest and the
704 potential risk of inhalational anthrax in Etosha National Park. (2015).
- 705 18. Antonation, K. S. *et al.* *Bacillus cereus* biovar anthracis causing anthrax in sub-Saharan
706 Africa—chromosomal monophyly and broad geographic distribution. *PLoS Negl. Trop.*
707 *Dis.* **10**, e0004923 (2016).
- 708 19. Turner, W. C. *et al.* Fatal attraction: vegetation responses to nutrient inputs attract
709 herbivores to infectious anthrax carcass sites. *Proc. R. Soc. Lond. B Biol. Sci.* **281**,
710 20141785 (2014).
- 711 20. Turner, W. C. *et al.* Lethal exposure: An integrated approach to pathogen transmission via
712 environmental reservoirs. *Sci. Rep.* **6**, (2016).

- 713 21. Manchee, R., Broster, M., Melling, J., Henstridge, R. & Stagg, A. *Bacillus anthracis* on
714 Gruinard island. (1981).
- 715 22. Hugh-Jones, M. & Blackburn, J. The ecology of *Bacillus anthracis*. *Mol. Aspects Med.* **30**,
716 356–367 (2009).
- 717 23. Williams, G., Linley, E., Nicholas, R. & Baillie, L. The role of the exosporium in the
718 environmental distribution of anthrax. *J. Appl. Microbiol.* **114**, 396–403 (2013).
- 719 24. Van Ness, G. B. Ecology of anthrax. *Science* **172**, 1303–1307 (1971).
- 720 25. Minett, F. & Dhanda, M. Multiplication of *B. anthracis* and *Cl. chauvei* in soil and water.
721 *Ind J Vet Sci Anim Husb* **11**, 308–328 (1941).
- 722 26. Minett, F. Sporulation and viability of *B. anthracis* in relation to environmental temperature
723 and humidity. *J. Comp. Pathol. Ther.* **60**, 161–176 (1950).
- 724 27. Vasil'eva, V. Soil bacteria as antagonists of anthrax bacilli. *Vet. Bull.* **9**, 149–153 (1958).
- 725 28. Zarubkinskii, V. Self purification of soil and water from anthrax bacilli. *Vet. Bull.* **9**, 51–58
726 (1958).
- 727 29. Saile, E. & Koehler, T. M. *Bacillus anthracis* multiplication, persistence, and genetic
728 exchange in the rhizosphere of grass plants. *Appl. Environ. Microbiol.* **72**, 3168–3174
729 (2006).
- 730 30. Beauregard, P. B., Chai, Y., Vlamakis, H., Losick, R. & Kolter, R. *Bacillus subtilis* biofilm
731 induction by plant polysaccharides. *Proc. Natl. Acad. Sci.* **110**, E1621–E1630 (2013).
- 732 31. Chen, Y. *et al.* Biocontrol of tomato wilt disease by *Bacillus subtilis* isolates from natural
733 environments depends on conserved genes mediating biofilm formation. *Environ.*
734 *Microbiol.* **15**, 848–864 (2013).

- 735 32. Lee, K. *et al.* Phenotypic and functional characterization of *Bacillus anthracis* biofilms.
736 *Microbiology* **153**, 1693–1701 (2007).
- 737 33. Debré, P. *Louis Pasteur*. (Johns Hopkins University Press, 2000).
- 738 34. Schwartz, M. The life and works of Louis Pasteur. *J. Appl. Microbiol.* **91**, 597–601 (2001).
- 739 35. Schuch, R. & Fischetti, V. A. The secret life of the anthrax agent *Bacillus anthracis*:
740 bacteriophage-mediated ecological adaptations. *PLoS One* **4**, e6532 (2009).
- 741 36. Dey, R., Hoffman, P. S. & Glomski, I. J. Germination and amplification of anthrax spores
742 by soil-dwelling amoebas. *Appl. Environ. Microbiol.* **78**, 8075–8081 (2012).
- 743 37. Barbaree, J. M., Fields, B. S., Feeley, J. C., Gorman, G. W. & Martin, W. T. Isolation of
744 protozoa from water associated with a legionellosis outbreak and demonstration of
745 intracellular multiplication of *Legionella pneumophila*. *Appl. Environ. Microbiol.* **51**, 422–
746 424 (1986).
- 747 38. El-Etr, S. H. *et al.* *Francisella tularensis* type A strains cause the rapid encystment of
748 *Acanthamoeba castellanii* and survive in amoebal cysts for three weeks postinfection. *Appl.*
749 *Environ. Microbiol.* **75**, 7488–7500 (2009).
- 750 39. Helgason, E. *et al.* *Bacillus anthracis*, *Bacillus cereus*, and *Bacillus thuringiensis*—one
751 species on the basis of genetic evidence. *Appl. Environ. Microbiol.* **66**, 2627–2630 (2000).
- 752 40. Kolstø, A.-B., Tourasse, N. J. & Økstad, O. A. What sets *Bacillus anthracis* apart from
753 other *Bacillus* species? *Annu. Rev. Microbiol.* **63**, 451–476 (2009).
- 754 41. Zwick, M. E. *et al.* Genomic characterization of the *Bacillus cereus* sensu lato species:
755 backdrop to the evolution of *Bacillus anthracis*. *Genome Res.* **22**, 1512–1524 (2012).
- 756 42. Pilo, P. & Frey, J. *Bacillus anthracis*: Molecular taxonomy, population genetics, phylogeny
757 and patho-evolution. *Infect. Genet. Evol.* **11**, 1218–1224 (2011).

- 758 43. Keim, P. *et al.* Anthrax molecular epidemiology and forensics: using the appropriate
759 marker for different evolutionary scales. *Infect. Genet. Evol.* **4**, 205–213 (2004).
- 760 44. Beyer, W. *et al.* Distribution and molecular evolution of *Bacillus anthracis* genotypes in
761 Namibia. *PLoS Negl. Trop. Dis.* **6**, e1534 (2012).
- 762 45. Beyer, W. & Turnbull, P. Co-infection of an animal with more than one genotype can occur
763 in anthrax. *Lett. Appl. Microbiol.* **57**, 380–384 (2013).
- 764 46. Hoffmaster, A. R. *et al.* Identification of anthrax toxin genes in a *Bacillus cereus* associated
765 with an illness resembling inhalation anthrax. *Proc. Natl. Acad. Sci. U. S. A.* **101**, 8449–
766 8454 (2004).
- 767 47. Leendertz, F. H. *et al.* Anthrax kills wild chimpanzees in a tropical rainforest. *Nature* **430**,
768 451–452 (2004).
- 769 48. Klee, S. R. *et al.* Characterization of *Bacillus anthracis*-like bacteria isolated from wild
770 great apes from Cote d’Ivoire and Cameroon. *J. Bacteriol.* **188**, 5333–5344 (2006).
- 771 49. Klee, S. R. *et al.* The genome of a *Bacillus* isolate causing anthrax in chimpanzees
772 combines chromosomal properties of *B. cereus* with *B. anthracis* virulence plasmids. *PLoS*
773 *One* **5**, e10986 (2010).
- 774 50. Brézillon, C. *et al.* Capsules, toxins and AtxA as virulence factors of emerging *Bacillus*
775 *cereus* biovar anthracis. *PLoS Negl. Trop. Dis.* **9**, e0003455 (2015).
- 776 51. Leppla, S. H. Anthrax Toxin Edema Factor: A Bacterial Adenylate Cyclase That Increases
777 Cyclic AMP Concentrations in Eukaryotic Cells. *Proc. Natl. Acad. Sci.* **79**, 3162–3166
778 (1982).
- 779 52. Collier, R. J. & Young, J. A. T. Anthrax Toxin. *Annu Rev Cell Dev Biol* **19**, 45–70 (2003).

- 780 53. Brien, J. O., Friedlander, A., Dreier, T., Ezzell, J. & Leppla, S. Effects of Anthrax Toxin
781 Components on Human Neutrophils. *Infect. Immun.* **47**, 306–310 (1985).
- 782 54. Comer, J. E., Chopra, A. K., Peterson, J. W. & Konig, R. Direct Inhibition of T-
783 Lymphocyte Activation by Anthrax Toxins In Vivo. *Infect. Immun.* **73**, 8275–8281 (2005).
- 784 55. Duesbery, N. S. Proteolytic Inactivation of MAP-Kinase-Kinase by Anthrax Lethal Factor.
785 *Science* **280**, 734–737 (1998).
- 786 56. Vitale, G. *et al.* Anthrax Lethal Factor Cleaves the N-Terminus of MAPKKs and Induces
787 Tyrosine/Threonine Phosphorylation of MAPKs in Cultured Macrophages. *Biochem.*
788 *Biophys. Res. Commun.* **248**, 706–711 (1998).
- 789 57. Erwin, J. L. *et al.* Macrophage-Derived Cell Lines Do Not Express Proinflammatory
790 Cytokines after Exposure to Bacillus anthracis Lethal Toxin. *Infect. Immun.* **69**, 1175–1177
791 (2001).
- 792 58. Pellizzari, R., Guidi-Rontani, C., Vitale, G., Mock, M. & Montecucco, C. Anthrax lethal
793 factor cleaves MKK3 in macrophages and inhibits the LPS/IFN γ -induced release of
794 NO and TNF α . *FEBS Lett.* **462**, 199–204 (1999).
- 795 59. Ribot, W. J. *et al.* Anthrax lethal toxin impairs innate immune functions of alveolar
796 macrophages and facilitates Bacillus anthracis survival. *Infect. Immun.* **74**, 5029–34 (2006).
- 797 60. Agrawal, A. *et al.* Impairment of dendritic cells and adaptive immunity by anthrax lethal
798 toxin. *Nature* **424**, 329–34 (2003).
- 799 61. Mogridge, J., Cunningham, K. & Collier, R. J. Stoichiometry of anthrax toxin complexes.
800 *Biochemistry (Mosc.)* **41**, 1079–1082 (2002).

- 801 62. Fink, S. L., Bergsbaken, T. & Cookson, B. T. Anthrax lethal toxin and Salmonella elicit the
802 common cell death pathway of caspase-1-dependent pyroptosis via distinct mechanisms.
803 *Proc. Natl. Acad. Sci.* **105**, 4312–4317 (2008).
- 804 63. Aloni-Grinstein, R. *et al.* Oral Spore Vaccine Based on Live Attenuated Nontoxinogenic
805 Bacillus anthracis Expressing Recombinant Mutant Protective Antigen. *Infect. Immun.* **73**,
806 4043–4053 (2005).
- 807 64. Cohen, S. N. *et al.* Attenuated Nontoxinogenic and Nonencapsulated Recombinant Bacillus
808 anthracis Spore Vaccines Protect against Anthrax. *Infect. Immun.* **68**, 4549–4558 (2000).
- 809 65. Little, S. F., Ivins, B. E., Fellows, P. F. & Fried. Passive protection by polyclonal antibodies
810 against Bacillus anthracis Infection in guinea pigs. *Infect. Immun.* **65**, 5171–5175 (1997).
- 811 66. Little, S. F. *et al.* Development of an in vitro-based potency assay for anthrax vaccine.
812 *Vaccine* **22**, 2843–2852 (2004).
- 813 67. Marcus, H. *et al.* Contribution of immunological memory to protective immunity conferred
814 by Bacillus anthracis protective antigen-based vaccine. *Infect. Immun.* **72**, 3471–3477
815 (2004).
- 816 68. Pitt, M. L. M. *et al.* In vitro correlate of immunity in a rabbit model of inhalational anthrax.
817 *Appl. Microbiol.* **19**, 4768–4773 (2001).
- 818 69. Reuveny, S. *et al.* Search for correlates of protective immunity conferred by anthrax
819 vaccine. *Infect. Immun.* **69**, 2888–2893 (2001).
- 820 70. Turnbull, P. C. B. Current status of immunization against anthrax: Old vaccines may be
821 here to stay for a while. *Curr. Opin. Infect. Dis.* **13**, 113–120 (2000).

- 822 71. Allen, J. S., Skowera, A., Rubin, G. J., Wessely, S. & Peakman, M. Long-lasting T cell
823 responses to biological warfare vaccines in human vaccinees. *Clin. Infect. Dis.* **43**, 1–7
824 (2006).
- 825 72. Cote, C. K., Rea, K. M., Norris, S. L., van Rooijen, N. & Welkos, S. L. The use of a model
826 of in vivo macrophage depletion to study the role of macrophages during infection with
827 *Bacillus anthracis* spores. *Microb. Pathog.* **37**, 169–175 (2004).
- 828 73. Cote, C. K., Van Rooijen, N. & Welkos, S. L. Roles of macrophages and neutrophils in the
829 early host response to *Bacillus anthracis* spores in a mouse model of infection. *Infect.*
830 *Immun.* **74**, 469–480 (2006).
- 831 74. Barton, G. M. & Medzhitov, R. Toll-like receptor signaling pathways. *Science* **300**, 1524–
832 1525 (2003).
- 833 75. Yadav, J. S. *et al.* Multigenic control and sex bias in host susceptibility to spore-induced
834 pulmonary anthrax in mice. *Infect. Immun.* **79**, 3204–3215 (2011).
- 835 76. Hughes, M. A. *et al.* MyD88-dependent signaling contributes to protection following
836 *Bacillus anthracis* spore challenge of mice: implications for Toll-like receptor signaling.
837 *Infect. Immun.* **73**, 7535–7540 (2005).
- 838 77. Boyden, E. D. & Dietrich, W. F. Nalp1b controls mouse macrophage susceptibility to
839 anthrax lethal toxin. *Nat. Genet.* **38**, 240–244 (2006).
- 840 78. Liao, K.-C. & Mogridge, J. Expression of Nlrp1b inflammasome components in human
841 fibroblasts confers susceptibility to anthrax lethal toxin. *Infect. Immun.* **77**, 4455–4462
842 (2009).
- 843 79. Terra, J. K. *et al.* Cutting edge: resistance to *Bacillus anthracis* infection mediated by a
844 lethal toxin sensitive allele of Nalp1b/Nlrp1b. *J. Immunol.* **184**, 17–20 (2010).

- 845 80. Moayeri, M. *et al.* Inflammasome sensor Nlrp1b-dependent resistance to anthrax is
846 mediated by caspase-1, IL-1 signaling and neutrophil recruitment. *PLoS Pathog* **6**,
847 e1001222 (2010).
- 848 81. Bradley, K. A., Mogridge, J., Mourez, M., Collier, R. J. & Young, J. A. Identification of the
849 cellular receptor for anthrax toxin. *Nature* **414**, 225–229 (2001).
- 850 82. Scobie, H. M., Rainey, G. J. A., Bradley, K. A. & Young, J. A. Human capillary
851 morphogenesis protein 2 functions as an anthrax toxin receptor. *Proc. Natl. Acad. Sci.* **100**,
852 5170–5174 (2003).
- 853 83. Liu, S. *et al.* Capillary morphogenesis protein-2 is the major receptor mediating lethality of
854 anthrax toxin in vivo. *Proc. Natl. Acad. Sci.* **106**, 12424–12429 (2009).
- 855 84. Martchenko, M., Candille, S. I., Tang, H. & Cohen, S. N. Human genetic variation altering
856 anthrax toxin sensitivity. *Proc. Natl. Acad. Sci.* **109**, 2972–2977 (2012).
- 857 85. Turnbull, P. C. B., Doganay, M., Lindeque, P. M., Aygen, B. & McLaughlin, J. Serology
858 and anthrax in humans, livestock and Etosha National Park wildlife. *Epidemiol. Infect.* **108**,
859 299–313 (1992).
- 860 86. Turnbull, P. C. B. *et al.* Naturally acquired antibodies to *Bacillus anthracis* protective
861 antigen in vultures in southern Africa. *Onderstepoort J. Vet. Res.* **75**, 95–102 (2008).
- 862 87. Lembo, T. *et al.* Serologic Surveillance of Anthrax in the Serengeti Ecosystem, Tanzania,
863 1996–2009. *Emerg. Infect. Dis.* **17**, 387–394 (2011).
- 864 88. Hampson, K. *et al.* Predictability of anthrax infection in the Serengeti, Tanzania. *J. Appl.*
865 *Ecol.* no-no (2011). doi:10.1111/j.1365-2664.2011.02030.x
- 866 89. Bellan, S. E. *et al.* Black-backed jackal exposure to rabies virus, canine distemper virus,
867 and bacillus anthracis in etosha national park, namibia. *J. Wildl. Dis.* **48**, 371–81 (2012).

- 868 90. Cizauskas, C. A., Bellan, S. E., Turner, W. C., Vance, R. E. & Getz, W. M. Frequent and
869 seasonally variable sublethal anthrax infections are accompanied by short-lived immunity
870 in an endemic system. *J. Anim. Ecol.* **83**, 1078–1090 (2014).
- 871 91. Cizauskas, C. A. *et al.* Gastrointestinal helminths may affect host susceptibility to anthrax
872 through seasonal immune trade-offs. *BMC Ecol.* **14**, 27 (2014).
- 873 92. Cizauskas, C. A., Turner, W. C., Pitts, N. & Getz, W. M. Seasonal patterns of hormones,
874 macroparasites, and microparasites in wild African ungulates: the interplay among stress,
875 reproduction, and disease. *PloS One* **10**, e0120800 (2015).
- 876 93. Ganz, H. H. *et al.* Interactions between *Bacillus anthracis* and plants may promote anthrax
877 transmission. *PLoS Negl Trop Dis* **8**, e2903 (2014).
- 878 94. Getz, W. M., Salter, R., Seidel, D. P. & Hooft, P. Sympatric speciation in structureless
879 environments. *BMC Evol. Biol.* **16**, 50 (2016).
- 880 95. Turner, W. C. *et al.* Soil ingestion, nutrition and the seasonality of anthrax in herbivores of
881 Etosha National Park. *Ecosphere*
- 882 96. Zidon, R., Garti, S., Getz, W. M. & Saltz, D. Zebra migration strategies and anthrax in
883 Etosha National Park, Namibia. *Ecosphere*
- 884 97. Ganz, H. H., Karaoz, U., Getz, W. M., Versfeld, W. & Brodie, E. L. Diversity and structure
885 of soil bacterial communities associated with vultures in an African savanna. *Ecosphere* **3**,
886 1–18 (2012).
- 887 98. Bellan, S. E., Turnbull, P. C., Beyer, W. & Getz, W. M. Effects of experimental exclusion
888 of scavengers from carcasses of anthrax-infected herbivores on *Bacillus anthracis*
889 sporulation, survival, and distribution. *Appl. Environ. Microbiol.* **79**, 3756–3761 (2013).

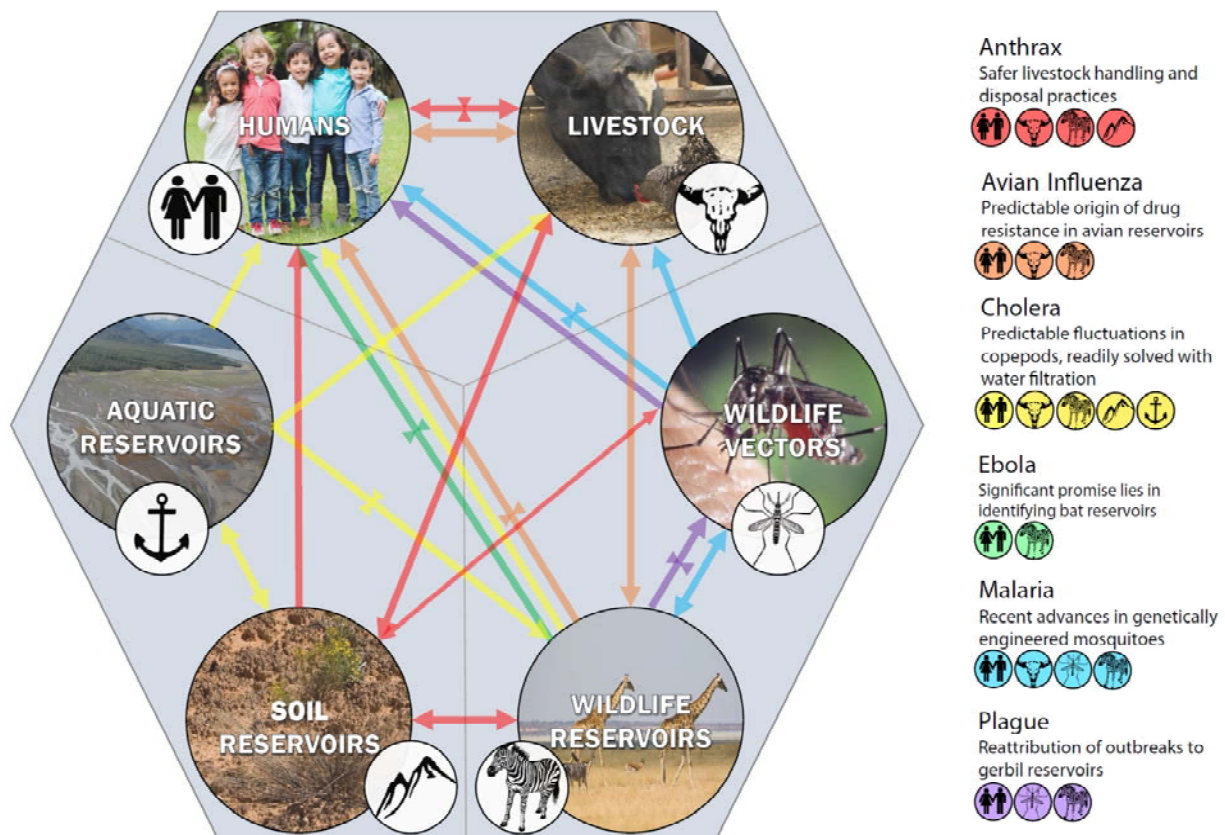
- 890 99. Barro, A. S. *et al.* Redefining the Australian anthrax belt: Modeling the ecological niche
891 and predicting the geographic distribution of *Bacillus anthracis*. *PLoS Negl Trop Dis* **10**,
892 e0004689 (2016).
- 893 100. Blackburn, J. K. *et al.* *Bacillus anthracis* diversity and geographic potential across Nigeria,
894 Cameroon and Chad: further support of a novel West African lineage. *PLoS Negl Trop Dis*
895 **9**, e0003931 (2015).
- 896 101. Chen, W.-J. *et al.* Mapping the Distribution of Anthrax in Mainland China, 2005–2013.
897 *PLoS Negl Trop Dis* **10**, e0004637 (2016).
- 898 102. Mullins, J. C. *et al.* Ecological niche modeling of *Bacillus anthracis* on three continents:
899 evidence for genetic-ecological divergence? *PloS One* **8**, e72451 (2013).
- 900 103. Blackburn, J. K. *et al.* Modeling the Ecological Niche of *Bacillus anthracis* to Map Anthrax
901 Risk in Kyrgyzstan. *Am. J. Trop. Med. Hyg.* **96**, 550–556 (2017).
- 902 104. Blackburn, J. K. Integrating geographic information systems and ecological niche modeling
903 into disease ecology: a case study of *Bacillus anthracis* in the United States and Mexico. in
904 *Emerging and Endemic Pathogens* 59–88 (Springer, 2010).
- 905 105. Chikerema, S., Murwira, A., Matope, G. & Pfukenyi, D. Spatial modelling of *Bacillus*
906 *anthracis* ecological niche in Zimbabwe. *Prev. Vet. Med.* **111**, 25–30 (2013).
- 907 106. Joyner, T. A. *et al.* Modeling the potential distribution of *Bacillus anthracis* under multiple
908 climate change scenarios for Kazakhstan. *PloS One* **5**, e9596 (2010).
- 909 107. Justice-Allen, A. *et al.* Survival and replication of *Mycoplasma* species in recycled bedding
910 sand and association with mastitis on dairy farms in Utah. *J. Dairy Sci.* **93**, 192–202 (2010).

- 911 108. Lowell, J. L. *et al.* Single-nucleotide polymorphisms reveal spatial diversity among clones
912 of *Yersinia pestis* during plague outbreaks in Colorado and the western United States.
913 *Vector-Borne Zoonotic Dis.* **15**, 291–302 (2015).
- 914 109. Saad-Roy, C., van den Driessche, P. & Yakubu, A.-A. A Mathematical Model of Anthrax
915 Transmission in Animal Populations. *Bull. Math. Biol.* **79**, 303–324 (2017).
- 916 110. Faruque, S. M. & Nair, G. B. Molecular ecology of toxigenic *Vibrio cholerae*. *Microbiol.*
917 *Immunol.* **46**, 59–66 (2002).
- 918 111. Rivera, I. N., Chun, J., Huq, A., Sack, R. B. & Colwell, R. R. Genotypes associated with
919 virulence in environmental isolates of *Vibrio cholerae*. *Appl. Environ. Microbiol.* **67**, 2421–
920 2429 (2001).
- 921 112. Plowright, R. K. *et al.* Ecological dynamics of emerging bat virus spillover. in *Proc. R. Soc.*
922 *B* **282**, 20142124 (The Royal Society, 2015).
- 923 113. Ip, H. S. *et al.* Novel Eurasian Highly Pathogenic Avian Influenza A H5 Viruses in Wild
924 Birds, Washington, USA, 2014. *Emerg. Infect. Dis. J.* **21**, 886 (2015).
- 925 114. Fine, P. E. & Carneiro, I. A. Transmissibility and persistence of oral polio vaccine viruses:
926 implications for the global poliomyelitis eradication initiative. *Am. J. Epidemiol.* **150**,
927 1001–1021 (1999).
- 928 115. Seidel, B. *et al.* Scrapie agent (strain 263K) can transmit disease via the oral route after
929 persistence in soil over years. *PLoS One* **2**, e435 (2007).
- 930 116. Almgren, E. S., Cross, P. C., Johnson, C. J., Heisey, D. M. & Richards, B. J. Modeling
931 Routes of Chronic Wasting Disease Transmission: Environmental Prion Persistence
932 Promotes Deer Population Decline and Extinction. *PLOS ONE* **6**, 1–11 (2011).

- 933 117. Johnson, C. J., Pedersen, J. A., Chappell, R. J., McKenzie, D. & Aiken, J. M. Oral
934 transmissibility of prion disease is enhanced by binding to soil particles. *PLoS Pathog.* **3**,
935 e93 (2007).
- 936 118. Pritzkow, S. *et al.* Grass plants bind, retain, uptake, and transport infectious prions. *Cell*
937 *Rep.* **11**, 1168–1175 (2015).
- 938 119. Blackburn, J. K., Kracalik, I. T. & Fair, J. M. Applying Science: Opportunities to Inform
939 Disease Management Policy with Cooperative Research within a One Health Framework.
940 *Front. Public Health* **3**, (2015).
- 941 120. Alexander, K. A. *et al.* What factors might have led to the emergence of Ebola in West
942 Africa? *PLoS Negl. Trop. Dis.* **9**, e0003652 (2015).
- 943 121. Wallace, R. G. *et al.* Did Ebola emerge in West Africa by a policy-driven phase change in
944 agroecology? in *Neoliberal Ebola* 1–12 (Springer, 2016).
- 945 122. Rulli, M. C., Santini, M., Hayman, D. T. & D’Odorico, P. The nexus between forest
946 fragmentation in Africa and Ebola virus disease outbreaks. *Sci. Rep.* **7**, (2017).
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949 **Figure 1.** A One Health approach applied to disease systems, showing the complexity of human
950 interactions with livestock, wildlife, and environmental reservoirs (including aquatic reservoirs,
951 and soil and plants). Arrows represent the directionality of transmission or spillover from one
952 compartment to another, highlighting that each disease has a unique complexity. The question
953 mark highlights that Ebola viral disease's biology is characterized by greater unknowns than the
954 other diseases. Major discoveries presented earlier that help contain disease are shown on the
955 transmission pathway (as bowties) they most significantly affect. Viewing diseases as organisms
956 in their own right, navigating this web, provides a more holistic and appropriate view than only
957 considering the human angle.

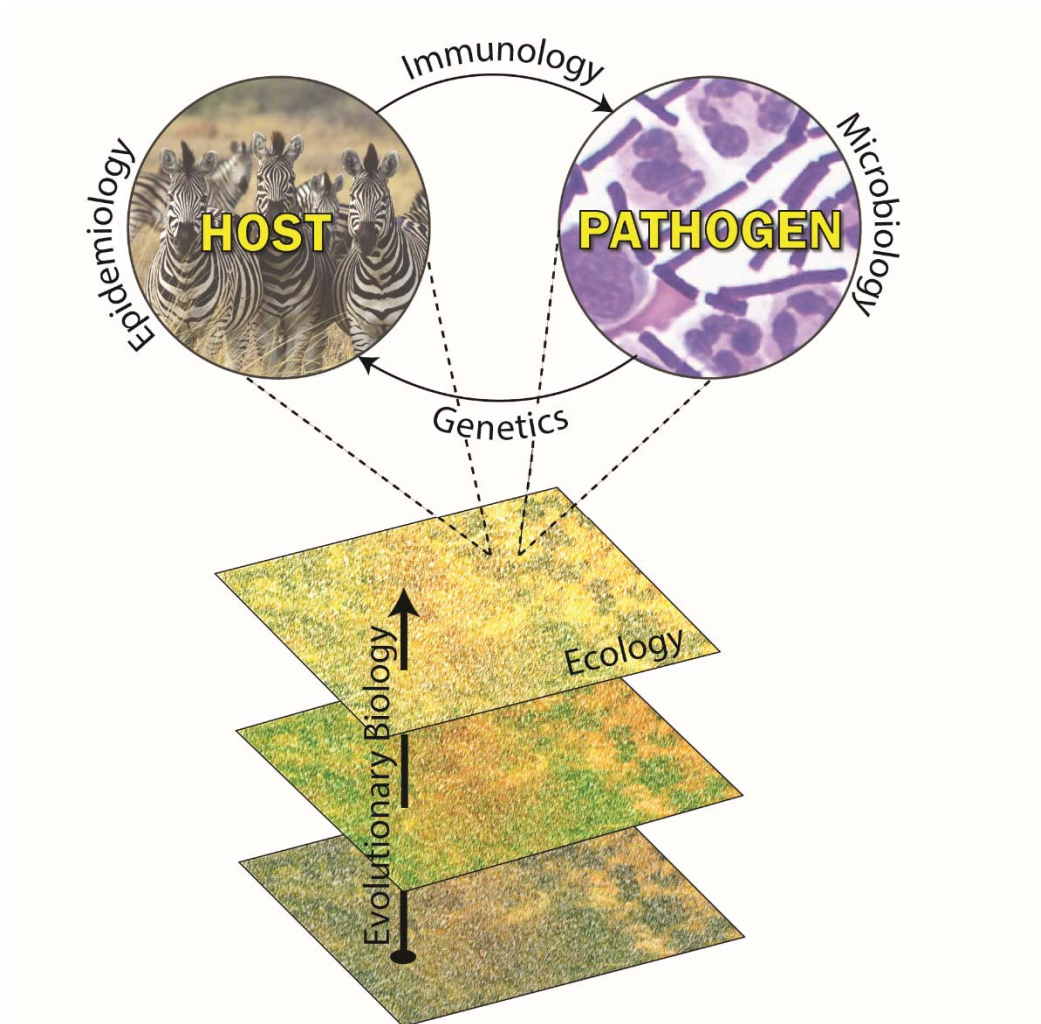
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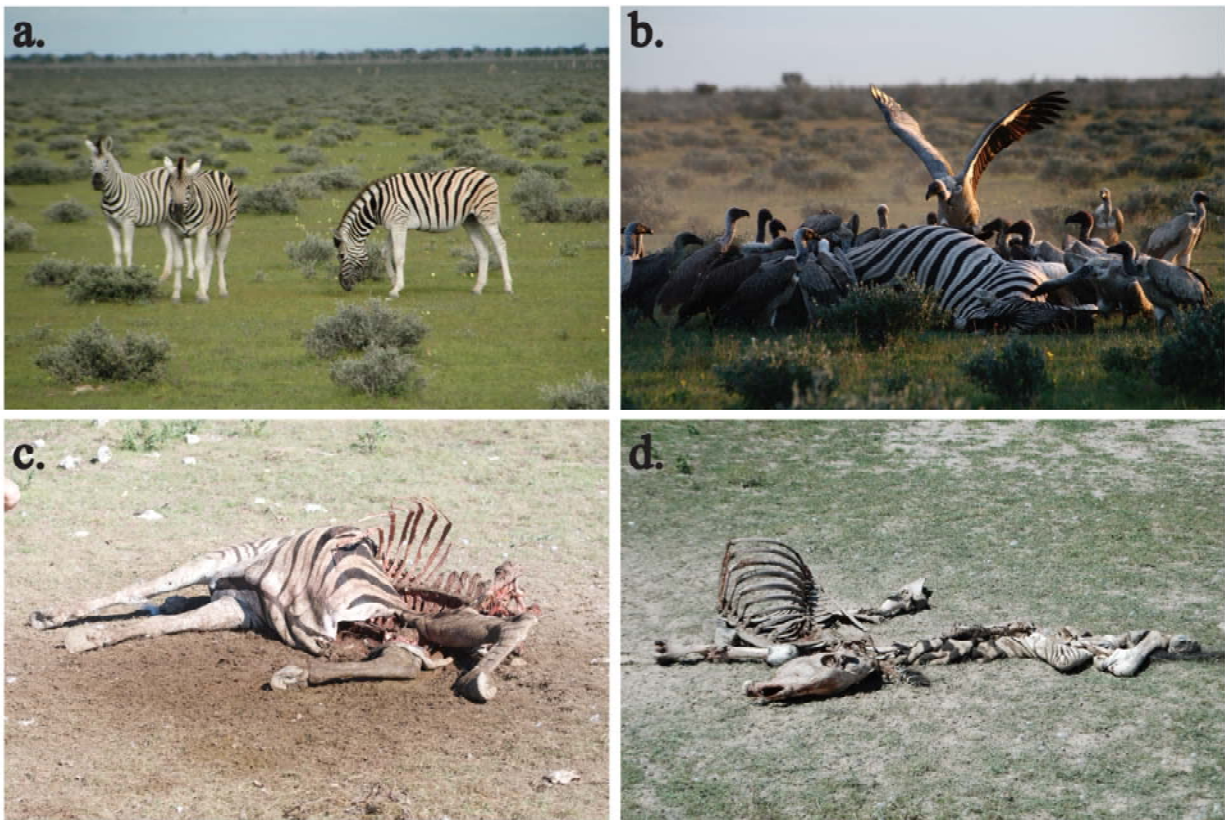
962 **Figure 2.** Different disciplinary perspectives fit together to provide a holistic perspective on
963 pathogen ecology. Some occur at multiple scales (e.g. while we aggregate genetics, genomics,
964 and evolution in the main text, their study may occur somewhat separately at different scales).
965 The sixth presented in our main text, clinical and public health, occurs in parallel with all the
966 processes depicted.
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970 **Figure 3.** The life cycle of anthrax in Etosha, viewed from the perspective of zebra (a), the most
971 common host. Zebra become infected while grazing, dying within approximately a week and
972 immediately attracting scavengers (b) that quickly open a carcass, depositing spores into the
973 ground. During the early stages of a carcass site, herbivores can fairly easily identify and avoid
974 partially decomposed carcasses (c), but as carcasses slowly blend into the environment over a
975 period of years and vegetation returns (d), herbivores return and once again become infected.
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978 **Figure 4.** The state of interdisciplinarity in disease research, as shown by Google Scholar results
 979 for papers published since 2000 and the nexus of seven disciplines (an approximate method for a
 980 top-down view of literature). For some diseases, like cholera, a strong interdisciplinary focus
 981 allows ecologists and clinicians to interact at the same intensity as researchers in more closely
 982 related fields. But for other neglected diseases, like anthrax, intra-host research (microbiology
 983 and immunology especially) dominate clinical collaboration. In the poorly-integrated literature
 984 on these diseases, ecological insights translate into human health solutions in a limited way.
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