Genetic networks of the oxytocin system in the human brain: A gene expression and large-scale fMRI meta-analysis study

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Abstract

Oxytocin is a neuropeptide involved in animal and human reproductive and social behaviour, with potential implications for a range of psychiatric disorders. However, the therapeutic potential of oxytocin in mental health care suggested by animal research has not been successfully translated into clinical practice, partly due to a poor understanding of the expression and distribution of the oxytocin signaling pathway in the human brain, and its complex interactions with other biological systems. Among the genes involved in the oxytocin signaling pathway, three genes have been frequently implicated in human social behavior: OXT (structural gene for oxytocin), OXTR (oxytocin receptor), and CD38 (central oxytocin secretion). We characterized the distribution of the OXT, OXTR, and CD38 mRNA across the brain, identified putative gene pathway interactions by comparing gene expression patterns across 29131 genes, and assessed associations between gene expression patterns and cognitive states via large-scale fMRI meta-analysis. In line with the animal literature, oxytocin pathway gene expression was enriched in central, temporal, and olfactory regions. Across the brain, there was high co-expression of the oxytocin pathway genes with both dopaminergic (DRD2) and muscarinic acetylcholine (CHRM4) genes, reflecting an anatomical basis for critical gene pathway interactions. Finally, fMRI meta-analysis revealed that oxytocin pathway maps correspond with motivation and emotion processing, demonstrating the value of probing gene expression maps to identify brain functional targets for future pharmacological trials.

Keywords: Oxytocin, oxytocin receptor, mRNA, brain
Oxytocin is an evolutionarily conserved neuropeptide implicated in an array of social and reproductive behaviors, and its role in the pathophysiology of complex psychological traits and as a putative therapeutic target in mental health conditions has attracted considerable attention (1). Oxytocin is mostly synthesized in neurons in the supraoptic nucleus and the paraventricular nucleus and released both systemically and centrally. Human research has shown beneficial effects of intranasal oxytocin on performance on tests assessing social cognition (2), gaze to the eye region (3), and the retrieval of social cues (4). Moreover, single nucleotide polymorphisms in oxytocin pathway genes have been associated with social behavior and psychiatric disorders (5). Emerging evidence also points to the oxytocin system’s role in energy metabolism (6), which may contribute to the increased risk of metabolic dysfunction in mental illness (7).

Despite some encouraging clinical trial results (e.g., 8, 9), unsuccessful trials and failures to replicate early human research have curbed the initial expectations for intranasal oxytocin as a therapeutic target (10). These translation failures have been partly attributed to a poor understanding of human central OXTR expression (11). As oxytocin can influence brain circuitry by binding to central oxytocin receptors, the anatomical distribution and density of oxytocin receptors in the brain can denote which neural regions and circuits would be most responsive to oxytocin. Given that OXTR mRNA distribution corresponds with OXTR locations (12, 13), the distribution of OXTR mRNA across the brain can therefore provide a map of central oxytocin binding.

Early animal work using histochemistry and immunohistochemistry revealed high concentrations of OXTR mRNA in the hypothalamus, amygdala, olfactory bulb, ventral pallidum, and the dorsal vagal nucleus (14, 15). Experimentally increasing
or decreasing (17) OXTR expression in the prairie vole nucleus accumbens also modulated partner preference behavior. While central OXTR mRNA localization in the rodent brain has been well-described (18), the anatomical distribution across the human brain is poorly characterized in general, as investigations have tended to sample very few brain regions (19, 20).

In addition to the anatomical distribution of OXTR mRNA, characterizing its interactions with other elements of the oxytocin signaling pathway and biological systems beyond this pathway is critical for determining oxytocin’s behavioral and functional relevance. Along with OXTR, CD38 and oxytocin-neurophysin I (OXT) genes in the oxytocin signaling pathway have been implicated in human social behavior (5). Specifically, CD38 is involved in central oxytocin secretion (21), and OXT encodes the oxytocin prepropeptide containing the nonapeptide oxytocin and the carrier protein neurophysin-I (22). Recent research (23) has reported whole-brain mRNA patterns of two out of three gene constituents of the oxytocin pathway system: OXTR and OXT. This analysis revealed increased OXT gene expression in the paraventricular nucleus of the hypothalamus, the lateral hypothalamic area, and the supraoptic nucleus, along with co-localization of OXT and OXTR mRNA with μ-opioid and κ-opioid receptor mRNA (23).

Evidence suggests that the oxytocin system also interacts with other neurotransmitter circuits, such as the dopaminergic (24) and muscarinic cholinergic (19) circuits, with possible implication for mental health. For instance, the dopamine D2-receptor subtype (DRD2), has been implicated with dysfunctional motivational processing in psychiatric illness (25) and pair bonding in animal models (26, 27). Moreover, the cholinergic muscarinic M4 receptor (CHRM4) has been associated with schizophrenia (28) and implicated in cognitive flexibility (29), social impairment
(30), and dopamine release (31). Finally, oxytocin can also bind to AVPR1A receptors (32), which has also been linked to social functioning (33, 34). However, the mRNA co-expression of these systems, and potentially others, with the oxytocin system are not well characterized.

While research has identified brain regions associated with high oxytocin pathway gene expression (19, 20, 23), inferring specific mental states to a single brain region can be problematic due to poor specificity. For instance, commonly observed increases in medial and lateral frontal region activity during emotion and pain processing seem to be better explained by more general sustained attention processes (35). The Neurosynth framework provides a tool (35) which can be used to decode which mental states are associated with the whole-brain oxytocin pathway network. Via the reverse inference of data from over 11,000 fMRI studies, this approach can reveal the probability of specific mental state patterns for a given whole-brain oxytocin pathway map with high specificity. Establishing the specific functional role of central oxytocin pathway network will provide a deeper understanding of the central human oxytocin system and aid translational research by more accurately identifying mental state targets for oxytocin treatment.

By leveraging the Allen Human Brain Atlas, which offers a uniquely comprehensive gene expression survey from six neurotypical adult human brains, we first characterize the anatomical distribution of the central oxytocin network by assessing mRNA expression of OXT, OXTR and CD38. Second, we explore putative gene pathway interactions by identifying genes with overlapping anatomical distributions with our target genes. Third, we decode the functional relevance of the oxytocin gene expression network using quantitative reverse inference via large-scale fMRI meta-analysis.
Materials and Methods

Post-mortem brain samples. mRNA distribution data was collected from the Allen Human Brain Atlas (http://human.brain-map.org/). Three donors were Caucasian males, one donor was a Hispanic female and two donors were African-American males. Mean donor age was 42.5 (S.D.=11.2) years. Data was collected on average 22.3 (S.D.=4.5) hours after demise (See Table S1 for details). Each brain was sampled in 363-946 distinct locations, either in the left hemisphere only (4 donor brains), or over both hemispheres (2 donor brains) using a custom Agilent 8 × 60K cDNA array chip. More details on procedures are available at http://help.brain-map.org/display/humanbrain/Documentation.

mRNA expression maps. The Allen Human Brain Atlas provides open-access expression profiles for oxytocin pathway genes, reporting log₂ transformed measures of the amount of hybridized RNA found through microarray analysis. While this is a comprehensive dataset, expression data is missing from some regions or not available from all donor samples. To assess accurate cognitive correlates of gene expression using meta-analysis via the Neurosynth tool, voxel-by-voxel mRNA maps are necessary. To create these voxel-by-voxel volumetric expression maps, we first marked all the sample locations and expression values in native image space. To interpolate missing voxels, we labeled brain borders with the sample expression value that had the closest distance to a given border point (Fig. S1A). Next, we linearly interpolated the space between scattered points by dividing that space into simplices based on Delaunay triangulation (Fig S1B), then linearly interpolated each simplex with values to yield a completed map (Fig. S1C). All maps were computed in
Matlab 2015a (The Mathworks Inc., Natick, MA, USA). Next, we registered all the individual brains to the MNI152 (Montreal Neurological Institute) template using ANTs (Advanced Normalization Tools; 36). To extract region specific statistics, we used regions extracted based on the Automated Anatomical Label (AAL) atlas.

**Data extraction.** Data for the oxytocin pathway was extracted from the Allen Human Brain Atlas for 3 different mRNA probes that have been associated with social behavior: **OXTR, CD38, and OXT** (5). Three other mRNA probes, which were hypothesized to co-express with oxytocin pathway mRNA, were also extracted a priori: **DRD2 (24-27)**, **CRHM4 (28-31)**, and **AVPR1A (33, 34)**. Since more than one probe for each mRNA was available, we selected the probe with the highest signal-to-noise ratio (mean/standard deviation), which represented the probe with least amount of spatial variability among donors (see Table S2).

**Central gene expression.** Statistical analysis was performed with R statistical package (version 3.3.2). Across the brain, gene expression for the right hemisphere ($n = 2$) was highly correlated with the corresponding left hemisphere ($n = 6$) gene expression values for all oxytocin pathway genes (**OXTR, OXT, CD38**; all $p$ values $< .00001$, all $r$ values $≥ 0.9$; Fig S2). Thus, the left hemisphere was used for analysis given its higher sample size. One-sample t-tests (two-tailed) were conducted to assess which of the 54 left hemisphere regions from six donor samples expressed mRNA to a significantly greater or lesser degree compared to average expression across the brain. To correct for multiple tests (54 in total), reported $p$-values were adjusted using a false discovery rate (FDR) threshold. Cohen’s $d$ values for one-sample t-tests were calculated to yield a measure of effect size. Pearson correlation...
coefficients were also calculated to assess the relationship of mRNA for 54 brain regions for OXTR, OXT, CD38, DRD2, CHRM4, and AVPR1A. P-values for correlational analyses were adjusted using a FDR threshold to correct for multiple tests (15 in total). Finally, we performed exploratory correlational analysis computing relationship between brain sample set of each subject and averaging over six individuals of all available mRNA probes (n = 58691) against three oxytocin pathway mRNA probes to assess putative gene-gene interactions based on spatial similarities in gene expression.

**Cognitive term correlates.** To identify the cognitive correlates of mRNA whole-brain expression maps, we performed quantitative reverse inference via large-scale meta-analysis of functional neuroimaging data using the NeuroSynth framework (35). As NeuroSynth favors the input of whole-brain voxel maps, we created a composite brain map representing an average of 6 individuals on the left hemisphere and average of 2 individuals on right hemisphere. Using naïve Bayes classification, NeuroSynth estimated the likelihood that mRNA maps were associated with mental state terms based on text-mining a corpus of 11,406 neuroimaging studies. The top 5 strongest non-anatomical correlations (Pearson’s r) were identified for OXTR, CD38, and OXT mRNA.

**Data and analysis script availability.** mRNA expression data is available from Allen Human Brain Atlas (http://human.brain-map.org/). The Matlab script for producing the brain region-specific data, the resulting dataset, and the R script used for statistical analysis is available at https://osf.io/jp6zs/.
Results

Central gene expression. Compared to average expression across the brain, high expression of OXTR mRNA levels were observed in the olfactory bulbs ($p = .0007, d = 6.9$), caudate ($p = .0009, d = 5.7$), putamen ($p = .002, d = 3.8$), pallidum ($p = .006, d = 3$), amygdala ($p = .009, d = 2.6$), rectus ($p = .02, d = 2.2$), parahippocampal region ($p = .02, d = 2.1$), hippocampus ($p = .02, d = 2$), and anterior cingulate ($p = .03, d = 1.9$; Fig 1A; Fig 2A; Supplemental Dataset S1 contains all test statistics). There was lower OXTR expression in the cerebral crus I ($p = .009, d = 2.5$), cerebral crus II ($p = .001, d = 4.9$), Cerebellum 7b ($p = .002, d = 3.7$), and Cerebellum 8 ($p = .001, d = 4.5$). High expression of CD38 mRNA was observed in the caudate ($p = .003, d = 5.7$), pallidum ($p = .003, d = 3$) olfactory bulbs ($p = .01, d = 6.9$), putamen ($p = .01, d = 3.8$), thalamus ($p = .02, d = 1.5$), and cingulate anterior ($p = .02, d = 1.9$; Fig 1B; Fig 2A). There was lower CD38 expression in the middle temporal pole ($p = .02, d = .7$) and Cerebellum 7b ($p = .03, d = 3.7$).

High expression of CHRM4 mRNA was observed in the thalamus ($p = .0002, d = 9.6$), pallidum ($p = .02, d = 2.8$), and olfactory bulbs ($p = .02, d = 2.7;$ Fig S3C). There was lower CHRM4 expression in the cerebral crus I ($p = .04, d = 2.2$), cerebral crus II ($p = .001, d = 6.8$) and Cerebellum 7b ($p = .002, d = 4.8$) High expression of DRD2 mRNA was observed in the putamen ($p < .0001, d = 13.1$), caudate ($p < .0001, d = 10.3$), thalamus ($p = .0002, d = 7.2$), pallidum ($p = .004, d = 3.3$), olfactory bulbs ($p = .009, d = 2.4$), insula ($p = .01, d = 2.4$), amygdala ($p = .01, d = 2.1$), and hippocampus ($p = .01, d = 2.1; $ Fig S3B). There was lower DRD2 expression in several areas in the cerebellum (cerebellum 7b, cerebellum 8, cerebellum 9, cerebral crus I, cerebral crus II), frontal region (orbital superior frontal gyrus, precentral area, dorsolateral superior frontal gyrus, orbital superior frontal gyrus, triangular inferior
frontal gyrus, supplementary motor area, medial superior frontal gyrus, medial orbital superior frontal gyrus), Occipital lobe (calcarine, cuneus), and Parietal lobe (postcentral gyrus, superior parietal gyrus, precuneus, paracentral lobule; all FDR corrected p-values < .05; Fig S3B; Dataset S1). There were no brain areas with significantly increased OXT (Fig. 1C; Fig 3A) or AVPR1A (Fig. S3A) expression which survived FDR correction (Table S1), however there was a considerable increase of OXT mRNA expression in the olfactory region (p = .13, d = 2.5).

Analysis revealed statistically significant relationships in gene expression over 54 brain regions (FDR corrected p-values) between all six genes (Fig. 3). Gene expressions were positively correlated for all gene pairs, except for the relationship in gene expression between AVPR1A and CHRM4, which was negatively related. Exploratory correlation analyses with all available 58692 gene probes in the Allen Human Brain Atlas and each of the three oxytocin pathway gene probes are summarized in Table 1 (full details in Table S3), with the top 10 strongest positive and top 10 strongest negative correlations. Histograms of the correlations between the three oxytocin pathway mRNA probes and all other available mRNA probes are presented in Figure 4.

**Cognitive correlates.** Decoding mental states meta-analytically from mRNA maps via quantitative reverse inference revealed that OXTR and CD38 mRNA expression maps were most highly correlated with functional imaging maps with motivation/dopaminergic topics such as “reward”, “anticipation”, and “dopaminergic”, and emotion-processing topics (Figure 2B, Table S4). Examination of these patterns revealed that the CD38 and OXTR mRNA maps were most associated with motivation-related topics whereas the OXT mRNA map was most associated with
emotion processing-related topics. Other topic categories with strong associations included pain and learning.

Discussion
The anatomical distribution of gene expression in the brain is heterogeneous and highly coordinated (37). Whereas dynamic alterations in gene expression are abundant in response to environmental demands, and are critically involved in range of cognitive functions, learning and diseases (38, 39), the basic organization likely partly reflects the evolutionary conserved modular layout of the brain (40). Indeed, gene-gene co-expression patterns form specific genetic signatures in the brain, representing distinct tissues, biological systems and associations with various brain disorders (37), and likely reflect the potential of complex and differential gene-gene interactions with implications for brain disorders and mental health. Here, we leverage the unique human brain mRNA expression library from Allen Brain Atlas to show that mRNA reflecting specific genes in the oxytocin pathway are highly expressed in central and temporal brain structures, along with the olfactory region. We also show reduced expression of OXTR and CD38 in the cerebellum, consistent with prior animal research (41). Oxytocin pathway genes showed considerable co-expression of DRD2 and CHRM4 mRNA, providing evidence for putative interactions between dopaminergic and cholinergic systems with the central oxytocin pathways. Exploratory analysis across 58692 RNA probes revealed strong positive spatial correlations between oxytocin pathway mRNA expression and mRNA associated with metabolic functions (GLUD1, GLUD2, C12orf39, SIM1), and strong negative correlations with GABRD, CAMKK2, PAK7, KCNJ3, KCNK1, and RTN4R, which have been associated with autism (42-44) and schizophrenia (45-49). Lastly,
quantitative reverse inference via meta-analysis of a corpus of 11,406 neuroimaging studies revealed that the distribution of the central genetic oxytocin pathway is most strongly associated with terms related to motivation and emotion, supporting the gene-system interactions suggested by the gene expression patterns. Together, these findings are consistent with a role of oxytocin in the processing of social stimuli and corroborate emerging evidence for interactions between the oxytocin, cholinergic (19), and dopaminergic systems (24), and for oxytocin’s role in metabolic regulation (7).

Compelling animal evidence suggests that direct central oxytocin administration influences social behavior and cognition via action on central oxytocin receptors (50). In these animals, oxytocin receptors are located in regions that are crucial for social behavior and the processing of social cues (18). Our data reveals striking similarities with central oxytocin pathway mRNA distribution observed in rodents (14, 15) and non-human primates (51). Given these between-species similarities in oxytocin receptor distribution, we suggest that the lack of robust effects of oxytocin administration in human clinical studies are most likely due to issues surrounding intranasal oxytocin administration methods which may constrain the potential availability of central oxytocin for oxytocin receptor binding (52, 53).

Along with the oxytocin pathway expression, we also investigated co-expression with dopaminergic and cholinergic receptor networks in the brain. Our results indicate that brain regions with increased expression of oxytocin pathway mRNA also have increased expression of the dopamine D2 receptor. Moreover, reverse inference of our generated oxytocin pathway mRNA maps on cognition via large-scale fMRI meta-analysis demonstrate that these maps closely correspond with brain regions underpinning for motivational processes. Altogether, our data
provides mRNA evidence that oxytocin exerts its effects synergistically with the dopaminergic network (24). Indeed, evidence points to a bidirectional relationship between central oxytocin and dopaminergic functioning. For instance, the oxytocin and dopaminergic systems work in concert to promote rodent maternal behaviors (54) and central D2 receptors have been shown to modulate social functioning in prairie voles (55, 56). There is also evidence of direct dopaminergic fibers to the supraoptic nucleus and the paraventricular nucleus (57), consistent with demonstrations that dopamine may stimulate oxytocin release in the hypothalamus (58, 59). This is also consistent with reported deficits in dopaminergic signaling in schizophrenia (60) and autism (61). The muscarinic cholinergic system may also work with the oxytocin system to boost attention (62) and to assist with the release of oxytocin (63). Moreover, the muscarinic cholinergic system is also thought to play a role in the cognitive dysfunction observed in schizophrenia (64).

We performed an exploratory correlational analysis between oxytocin pathway mRNA and 58692 other mRNA probes. There are several relationships worth noting in the context of psychiatric illness. First, OXTR is highly co-expressed with Neurotensin receptor gene (NTSR2), which has been found to regulate ethanol consumption in mice (65), and Glutamate dehydrogenase 1 and 2 (GLUD1, GLUD2), which are involved in glutamate recycling and insulin secretion. Second, CD38 is highly co-expressed with NTSR2, GLUD1, and Spexin (C12orf39), with the latter associated with weight regulation (66). Third, OXT is highly co-expressed with AVP, and Single-minded homolog 1 (SIM1), which may regulate feeding behavior (67). Altogether, these results are consistent with emerging evidence that the oxytocin system may play a role in the metabolic dysfunction often observed in severe mental illnesses (7). Strikingly, the strongest negative correlation across 58692 mRNA
probes for both OXTR and OXT were found for the gamma-aminobutyric acid type A receptor delta subunit (GABRD) gene, which has been associated with autism (42, 43), mood disorders (68), and schizophrenia (69). Another strong negative correlation was CAMKK2, which has been identified as a schizophrenia susceptibility gene (45).

Our data revealed high expression of oxytocin pathway mRNA in the olfactory region, which is consistent with animal research (e.g., 70). Indeed, social olfactory deficits in mice without the oxytocin gene (71) are rescued with injection of oxytocin into rat olfactory bulbs (72). Increased mRNA in the human olfactory region may be vestigial, as olfaction is not as important for human conspecific identification compared to most other mammals due to species specialization, however, intranasal oxytocin has been shown to improve olfactory detection performance in schizophrenia (73), and intranasal oxytocin is thought to enter the brain via the olfactory bulbs (74).

There are three important limitations worth noting. First, given the nature of human tissue collection for mRNA studies, the sample size was small. However, the relatively small amount of measurement error for the mRNA probes is consistent with high precision. Second, there was only 1 female in the sample, which prohibited an examination of sex differences of mRNA distribution. However, evidence suggests there is limited sex differences in OXTR expression in humans (75). Third, our reverse inference approach is better suited to identify broad domains, such as “emotion”, rather than more specific domains.

Comprehensive clinical trials need to demonstrate engagement of drug targets, such as OXTR occupancy reflected by regional brain activity changes (11). Without precise targets, it is unclear whether null effects are due to an inefficacious
drug or misidentified drug targets. By identifying accurate oxytocin pathway targets in the human brain, our study provides an oxytocin treatment target map that can facilitate efforts to better understand treatment efficacy and the most effective treatment dose. Using this map, research can move towards studying dose-dependent effects on neural targets rich in oxytocin receptors (e.g., 76).

Analysis of data from the Allen Brain Atlas database provides a unique opportunity to map the central oxytocin system and explore co-expression with other systems. Altogether, these results provide a proof-of-principle demonstration of corresponding cognitive and gene expression patterns of a neuropeptide pathway involved in complex human and animal behaviors, and identify brain regions and cognitive processing targets for future oxytocin trials (11).
Acknowledgements. This research was supported by an Excellence Grant from the Novo Nordisk Foundation (NNF16OC0019856 to D.S.Q), the Research Council of Norway (223273, 249795), KG Jebsen Stiftelsen and the South-Eastern Norway Regional Health Authority (2014097, 2015073).
References


10. Alvares GA, Quintana DS, & Whitehouse AJ (2016) Beyond the hype and hope: critical considerations for intranasal oxytocin research in autism spectrum disorder.


behavior in vasopressin V1a receptor knockout mice. *Neuropsychopharmacology* 29(3):483.


hypothalamus and extrahypothalamic regions of the human and primate brain. 


Figure 1. Central gene expression for OXTR (A), CD38 (B), and OXT (C). Each point represents mean expression with standard errors for a given brain region. The bolded dashed lines represent whole-brain mean expressions. One sample t-tests were performed to assess which regions expressed mRNA to a significantly greater degree compared to average expression across the brain. * p < 0.05 (FDR corrected for 54 tests).
Figure 2. Whole-brain gene expression maps for the oxytocin pathway and their cognitive correlates. Whole-brain gene expression maps are presented for OXT, OXTR and CD38 (A). A large-scale meta-analysis of functional neuroimaging using the NeuroSynth framework was performed to infer mental state correlates of oxytocin pathway gene expression, with plot units representing Pearson’s r (B). SD = Standard deviation.
Figure 3. Correlations between central gene expression of **OXTR**, **CD38**, **OXT**, **AVPR1A**, **DRD2**, and **CHRM4**. Each scatterplot point represents a location in the left hemisphere of the brain, with 54 regions in total. *P*-values are FDR corrected for multiple tests. See Figure 1 for a list of locations.
Figure 4. Distributions of correlations between oxytocin pathway genes and all other mRNA probes in the Allen Human Brain Atlas. Histograms are presented for OXTR (A), CD38 (B), and OXT (C). Each histogram visualises the correlation between the target oxytocin pathway mRNA probe and all other mRNA probes (n = 58691). For each oxytocin pathway gene, the position of the other two oxytocin pathway genes is shown.
### Table 1. Top correlations between oxytocin pathway probes and all remaining probes

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<th>OXT r</th>
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<tr>
<td>NTSR2</td>
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#### Top 10 positive

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<td>PAK7</td>
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<tr>
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<td>KCNJ3</td>
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<tr>
<td>CAMKK2</td>
<td>-0.651</td>
<td>-0.710</td>
<td>-0.333</td>
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<tr>
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#### Top 10 negative

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<td>GABRD</td>
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Note. Some genes had more than one probe feature in the top 10 positive or negative correlations (e.g., OXT). Full probe identification numbers available in Table S3. 

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