

1 **Title:**

2 Genetics of cortico-cerebellar expansion in anthropoid primates: a comparative approach

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35 **Abstract**

36 What adaptive changes in brain structure and function underpin the evolution of increased
37 cognitive performance in humans and our close relatives? Identifying the genetic basis of
38 brain evolution has become a major tool in answering this question. Numerous cases of
39 positive selection, altered gene expression or gene duplication have been identified that may
40 contribute to the evolution of the neocortex, which is widely assumed to play a predominant
41 role in cognitive evolution. However, the neocortex co-evolves with other, functionally inter-
42 dependent, regions of the brain, most notably the cerebellum. The cerebellum is linked to a
43 range of cognitive tasks and expanded rapidly during hominoid evolution, independently of
44 neocortex size. Here we demonstrate that, across primates, genes with known roles in
45 cerebellum development are just as likely to be targeted by selection as genes linked to
46 cortical development. In fact, cerebellum genes are more likely to have evolved adaptively
47 during hominoid evolution, consistent with phenotypic data suggesting an accelerated rate of
48 cerebellar expansion in apes. Finally, we present evidence that selection targeted genes with
49 specific effects on either the neocortex or cerebellum, not both. This suggests cortico-
50 cerebellar co-evolution is maintained by selection acting on independent developmental
51 programs.

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69 **Introduction**

70 The proximate basis of primate brain expansion and increased cognitive performance lies in
71 changes in gene function and regulation. Identifying the genetic basis of phenotypic change
72 can provide insights into how developmental mechanisms evolve, how they are constrained,
73 and how changes at a cellular level contribute to broad scale anatomical evolution (1). This
74 potential for dissecting the biological basis of brain and behavioural evolution motivates
75 many genomic comparisons across primates (2). These have identified numerous genes
76 associated with brain development with high rates of evolution (3–9), divergent expression
77 profiles (10–13) or duplicated sequence (14–17), either across primates or during recent
78 human evolution. In several of these cases the genetic changes have been demonstrated to
79 have functional effects on neuronal proliferation or maturation (7,8,15,18,19). These results
80 highlight potential cellular adaptations driving changes in brain size, and provide a powerful
81 means of investigate human-specific adaptations.

82 The majority of these examples investigate genes linked to neocortical evolution,
83 reflecting the widely held assumption that the neocortex has a predominant role in ‘higher’
84 cognition (20). However, the neocortex co-evolves with other brain components with which it
85 is functionally connected, suggesting a complete understanding of primate brain expansion
86 will not be found by focusing solely on neocortex development (21–23). Of particular
87 importance is the relationship between the neocortex and cerebellum. Across primates
88 cortico-cerebellar co-evolution pervades biological levels, occurring at a coarse volumetric
89 scale, at the level of individual nuclei, and at the cellular level (21,22,24).

90 In humans, the cerebellum is increasingly recognised to be important for both motor
91 and ‘higher cognitive’ function, including the capacity to plan and execute complex
92 behavioural sequences (20,23,25). Similarly, across primates, cerebellar expansion is linked
93 to extractive foraging, independently of neocortex volume (23). The importance of the
94 cerebellum in primate brain evolution is further bolstered by comparative analyses that
95 demonstrate a rapid, non-allometric expansion of the cerebellum relative to neocortex size in
96 hominoids that is indicative an adaptive change in the cortico-cerebellar functional
97 relationship (26–29). Hominin evolution is also characterised by reciprocal expansion of the
98 neocortex and cerebellum, with recent modern humans being distinguished from our early
99 modern humans by an increase in cerebellum volume relative to the neocortex (30).

100 The importance of coordinated cortico-cerebellum expansion suggests that we must
101 look beyond neocortical evolution to obtain a full picture of the genetic architecture of
102 primate brain evolution. The likely action of positive selection on genes involved in

103 cerebellum development is further suggested by the accelerated rate of evolution of *AHII*, a
104 gene associated with developmental disorders of the cerebellum, during human evolution
105 (31). In addition, genetic approaches may be useful in addressing long-standing debates about
106 the constraints governing co-evolving brain networks. For example, do the neocortex and
107 cerebellum co-evolve through genetically independent developmental changes maintained by
108 selection (*sensu* Barton and Harvey (21))? Or are they the result of pleiotropic genetic effects
109 shared across anatomical boundaries (*sensu* Finlay and Darlington (32))? If the latter is true,
110 do these pleiotropic effects reflect an evolutionary constraint, or have they evolved to
111 maintain the functional relationship between components during brain expansion? These
112 questions will be central to providing a full understanding of brain evolution, and also
113 address questions of long standing interest in evolutionary biology on the trade-offs between
114 adaptation and constraint in the genetic basis of composite traits.

115 Here we ask three questions that provide an initial assessment of the role of genes
116 controlling the development of the neocortex and cerebellum in anthropoid brain evolution.
117 First, we ask whether genes with known roles in the development of the cerebral cortex,
118 which is predominantly composed of the neocortex, are more likely to be targets of positive
119 selection than those affecting cerebellum development. Second, whether patterns of
120 molecular evolution mirror non-allometric changes in component size. Finally, whether these
121 genes evolve in a manner suggesting a specific evolutionary association with either brain
122 component to explore whether cortico-cerebellar co-evolution is maintained by selection
123 acting on a common or independent set of genes.

124

125 **Methods**

126 *Data collection*

127 We obtained a list of human genes with known roles in ‘cerebral cortex development’ or
128 ‘cerebellum development’ from two ontology databases; Amigo (33) (GO:0021987 and
129 GO:0021549) and EBI’s QuickGO (34) (GO:0021987 and GO:0021549). These GO terms
130 have the best-matched definitions among those relevant for each component. In anthropoids,
131 the neocortex comprises the vast majority of the cerebral cortex (~90%; Stephan et al. 1981).
132 This GO search resulted in 198 Amigo and 300 QuickGO genes associated with cerebral
133 cortex development, and 144 Amigo and 222 QuickGO genes associated with cerebellum
134 development, which are not mutually exclusive, that were then combined to form the starting
135 gene set. This starting human gene set was then used to obtain 1:1 orthologs from 11
136 anthropoid genomes (Figure 1A) using a reciprocal best hit BLASTn (36) approach between

137 each species and the human coding sequence with an e-value cut-off of $1e^{-10}$ and a minimum
138 percentage identity of 30. 11-way 1:1 ortholog sets were aligned using codon aware PRANK
139 v 140615 (37), converted to phylip format in preparation for PAML analyses (38) and filtered
140 using a conservative alignment filtering program SWAMP v 1.0 (39) to remove regions of
141 poorly aligned or error-rich sequences on a branch-specific basis. SWAMP was run twice,
142 first using a threshold of 5 and a window size of 15, and then a second run with a threshold of
143 2 with a window size of 3, with a minimum sequence length of 300 bases and *interscan*
144 masking for both runs. The final, strict and conservatively filtered 11-species dataset
145 consisted of three non-overlapping groups: 53 genes with known roles cerebral cortex
146 development, 47 genes with known roles in cerebellum development, and 10 genes with
147 known roles in both (Table S1).

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149 *Selection analyses*

150 Estimation of dN/dS ratios (ω), a common measure of the strength of selection acting on a
151 protein coding gene, was carried out using a codon-based maximum likelihood method
152 (PAML v.4) (38). Nested models were compared using the likelihood ratio test statistic
153 ($-2[\log\text{likelihood1} - \log\text{likelihood2}]$) to critical values of the χ^2 distribution and degrees of
154 freedom as the difference in the number of parameters estimated in each model. We
155 compared the frequency of positive selection acting on genes associated with cerebral cortex
156 and cerebellum development in two ways. First, we used the site model tests of positive
157 selection ('Model 8/8a') to identify genes evolving under positive selection across
158 anthropoids. The site models allow ω to vary across sites, but not across branches. Second,
159 we used the branch-site models ('new model A') to identify genes under increased positive
160 selection in hominoids (Figure 1A). The branch-site models allow ω to vary across both sites
161 and branch categories defined a priori. We repeated this test for accelerated rates of evolution
162 in hominoids using a branch model test, where ω is fixed across sites but varies between
163 branch categories. For each analysis, the percentage of each category of gene that
164 experienced selection were compared using a Z-test and more conservative Fisher's Exact
165 Test.

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167 *Gene-phenotype co-evolution*

168 We sought to test the link between the molecular evolution of our gene-set and the evolution
169 of neocortex and cerebellum size using a phylogenetic, comparative approach (40). Branch
170 models in PAML were used to calculate the root-to-tip dN/dS for each species. These were

171 then regressed against the phenotypic trait of interest using a Phylogenetic Generalised Least
172 Squares (PGLS) regression implemented in BayesTraits (41), that corrects for the non-
173 independence of inter-specific data. Each gene was regressed against neocortex volume and
174 cerebellum volume as predictor variables simultaneously. This permits the identification of
175 genes with either a specific co-evolutionary association with either neocortex or cerebellum
176 size, or genes that co-vary with both traits independently. We also repeated the analyses
177 including rest-of-brain volume as an additional predictor variable, which lead to similar
178 results. In all cases root-to-tip dN/dS and brain component volumes were \log_{10} -transformed.

179

180 **Results**

181 *What proportion of genes with known roles in cerebral cortex or cerebellum development are*
182 *targeted by positive selection?*

183 Across anthropoids, the average rate of evolution of genes with known roles in the
184 development of the cerebellum does not differ from those that function in the development of
185 the cerebral cortex, which is predominantly comprised of neocortex (Mann-Whitney $U_{(98)} =$
186 1167.5 , $Z = 0.539$, $p = 0.590$) (Table S2). Genes linked to the development of both brain
187 components do not differ from those specifically linked to cerebral cortex ($U_{(61)} = 296.5$, $Z =$
188 0.592 , $p = 0.554$) or cerebellum development ($U_{(55)} = 241.5$, $Z = 0.136$, $p = 0.892$). Site-
189 model tests for positive selection acting at a subset of codons also identify a similar
190 proportion of genes associated with cerebral cortex or cerebellum development with evidence
191 of positive selection across anthropoids. 3/47 cerebellum genes (6.4%) are significant at a
192 nominal α of 0.05, compared to 5/53 cerebral cortex genes (9.4%). These proportions are not
193 significantly different (Fisher's Exact Test, $p = 0.716$, Z-test: $z = 0.5613$, $p = 0.575$) (Table,
194 1; Figure 1B; Table S3). The same conclusion is reached after correcting for multiple testing
195 using the false discovery rate (FDR) (42), after which the site-model test is significant for two
196 genes linked to cerebellum development (*RPGRIP1L*, *PCNT*) and one gene linked to cerebral
197 cortex development (*TACC2*). None of the 10 genes with annotated function in the
198 development of both brain components show evidence of positive selection. These results
199 suggest genes affecting cerebellum development are just as likely to be targeted by positive
200 selection as those affecting cerebral cortex development.

201

202 *Do rates of molecular evolution reflect rates of brain component expansion?*

203 A significantly greater proportion of cerebellum genes (14/47, 29.8%) than cerebral cortex
204 genes (6/53, 11.3%) experienced an accelerated rate of evolution in hominoids, taking a

205 nominal α of 0.05 (Fisher's Exact Test, $p = 0.026$, Z-test: $z = 2.304$, $p = 0.021$) (Table 1;
206 Figure 1C; Table S4). This result is also reflected in the branch-site test where the proportion
207 of cerebellum genes (6/47, 12.8%) with evidence of episodic positive selection in hominoids
208 again exceeds the proportion of cerebral cortex genes (2/53, 3.8%). In this case the trend does
209 not reach significance (Fisher's Exact Test, $p = 0.143$, Z-test: $z = 1.654$, $p = 0.099$) (Table
210 S5). However, one assumption of the branch-site test is the absence of positive selection in
211 the background (non-hominoid) branches (43). After excluding genes with evidence of
212 positive selection across anthropoids under the site-model test (Table S3), the trend observed
213 in the branch-site test becomes significant at $p < 0.05$ (Fisher's Exact Test, $p = 0.049$, Z-test:
214 $z = 2.136$, $p = 0.032$) (Table 1; Figure 1D). These contrasting proportions suggest the strength
215 of positive selection acting on genes controlling cerebellum development increased during
216 hominoid evolution, a clade in which the rate of cerebellar expansion significantly
217 accelerated (29), without a corresponding acceleration in cerebral cortex expansion (21,44).

218

219 *Is selection associated with interspecific variation in the size of specific brain components?*

220 We identify 11/47 cerebellum genes (23.4%) that co-evolve with cerebellum volume, after
221 controlling for neocortex volume, of which 4 are significant at $p < 0.001$ and 2 (*RPGRIP1L*,
222 *ATRN*) remain significant after FDR-correction (Table 1; Figure 2; Table S6). We re-
223 analysed the top four genes associated with cerebellum volume, separating dN and dS whilst
224 controlling variation in neocortex volume, to test if the association is driven by variation in
225 dN . For 3/4 genes we find a significant partial regression with dN (*ATRN* $t_5 = 3.789$, $p =$
226 0.006 ; *EZH2* $t_5 = 3.990$, $p = 0.005$; *KNDC1* $t_5 = 3.586$, $p = 0.008$), the remaining locus
227 showed a non-significant trend (*RGRIP1L* $t_5 = 1.582$, $p = 0.087$). Only one cerebellum gene
228 (2.1%) shows an association with neocortex volume, which is a significantly lower
229 proportion (Fisher's Exact Test, $p = 0.004$, Z-test: $z = 3.091$, $p = 0.002$). 1/53 cerebral cortex
230 genes (*DICER1*) shows evidence of co-evolution with neocortex volume (1.9%), whilst 6
231 (11.3%) show evidence of an association with cerebellum size (Table S6). This is not a
232 significant difference in proportion (Fisher's Exact Test, $p = 0.113$, Z-test: $z = 1.955$, $p =$
233 0.050), only one of these associations is significant at $p < 0.001$ and none survive FDR-
234 correction. One of the ten genes annotated as functioning in both neocortex and cerebellum
235 development (*GART*) shows an association with cerebellum, but not neocortex, volume
236 (Table S6). Similar results were obtained when rest-of-brain was included in the regression
237 model. No gene shows an association with variation in both neocortex and cerebellum

238 volume, regardless of the gene category or regression model. These results suggest selection
239 acts on genes associated with phenotypic variation specific to each brain component.

240

241 **Discussion**

242 Primate brain expansion reflects increases in the volume and neuron number of co-evolving
243 structures (21). This pattern of distributed adaptation must be reflected in the molecular
244 evolution of genes controlling brain development. Our results provide two contributions to
245 understanding the genetic basis of brain evolution. First, we provide evidence that genes
246 associated with cerebral cortex development are no more likely to have evolved adaptively
247 across anthropoids than cerebellum genes. Indeed, significantly more cerebellum genes
248 experienced an increased rate of evolution in hominoids, consistent with the non-allometric
249 expansion of the cerebellum, but not the neocortex, in apes (29). Second, we provide
250 evidence that the selection regimes shaping these genes are linked to the evolution of specific
251 brain components.

252 Our results raise the possibility that a significant proportion of the genetic changes
253 that underpin adaptive evolution of primate brain size, structure and cognition will affect
254 aspects of non-cortical development. This conclusion is consistent with evidence of adaptive
255 expansion in cerebellum volume during hominoid evolution (29), and is further supported by
256 a phylogenetic analysis of lineage-specific shifts in gene expression across mammals that
257 found an over-abundance of hominoid-specific expression shifts in genes expressed in the
258 cerebellum (10). These results suggest future comparative studies of primate gene expression
259 should not be limited to samples derived from cortical tissue, and functional tests of
260 candidate genes targeted by positive selection should consider the phenotypic relevance of
261 changes in gene function in non-cortical structures.

262 Of course, our results are wholly dependent on the quality of gene ontology
263 annotation. Our incomplete knowledge of gene function means that it is inevitable that the
264 genes included in this study reflect a minority of those that influence cerebral cortex and
265 cerebellum development. However, gene ontology datasets are routinely used in post-hoc
266 tests for functional enrichment where the same caveats apply, here we have simply used the
267 available data to facilitate tests of specific hypotheses. We know of no reason why either
268 gene set may be biased in a way that would produce contrasting patterns of results, but cannot
269 formally rule out this possibility. Finally, in this study we adopted a conservative approach,
270 analysing only genes with strict 1:1 orthologues in published anthropoid genomes

271 and removing short and/or poorly aligned sequence. This further reduces the gene set, but
272 produces more reliable estimates of selection regimes (39).

273 Our analyses highlight several genes with patterns of molecular evolution that link
274 them to inter-specific differences in cerebellum volume. These include two genes, *RGRIPIL*
275 and *ATRN*, with a particularly strong signal of co-evolution between the strength of selection
276 acting on their coding sequence and cerebellum volume, independently of variation in
277 neocortex or rest-of-brain volume, in our gene-phenotype association tests (Figure 2).
278 *RGRIPIL* is one of a small number of loci linked to Joubert Syndrome, a rare genetic
279 disorder associated with severe hypoplasia of the mid-hindbrain and cerebellum (45,46). The
280 cellular role of *RGRIPIL* appears to be in the correct function of the cilia and basal bodies
281 (47,48) which are necessary for expansion of the cerebellar neural progenitor pool (49).
282 Disruption of *ATRN*, an E3 ubiquitin ligase, in *Mus* causes vacuolization and degeneration of
283 the cerebellum (50,51). Another ubiquitin ligase, *UBE3A*, has been implicated in brain and
284 cognitive development (52) and interacts with *ASPM*, a key regulator of brain size (53). A
285 third gene, *EZH2*, also shows evidence of a phenotypic association with cerebellum volume
286 but is narrowly non-significant after FDR correction. *EZH2*, functions to regulate neuronal
287 migration of precerebellar neurons during the development of the cortico-cerebellar
288 connectivity (54).

289 We also find multiple genes with significantly accelerated rates of evolution in
290 hominoids, coincident with an accelerated rate of cerebellar expansion (29). Several of these
291 genes also show an association with cerebellum volume in our gene-phenotype association
292 tests at a nominal α threshold of 0.05. These include *AGTPBP1*, disruption of which causes
293 cerebellar Purkinje cell degeneration (55), *PCNT*, which causes primordial dwarfism with
294 microcephaly (56), *TH*, a gene linked to two disorders which affect motor control, Segawa
295 Syndrome and Parkinson's (57,58), and *MYO16* and *GART* which have putatively been
296 linked to Autistic Spectrum Disorders and Down Syndrome respectively (59,60).

297 Finally, we identify patterns of molecular evolution that may implicate a separate
298 group of genes in the evolution of neocortex size. Only one locus shows a phenotypic
299 association with neocortex volume; *DICER1*, which regulates neurogenesis in the developing
300 cerebral cortex in a time dependent manner (61,62). Notably, the two genes with the strongest
301 evidence for episodic positive selection in hominoids belong to the same gene family. *TACCI*
302 and *TACC2* are both associated with regulation of nuclear migration and are required for
303 normal patterns of self-renewal in neural progenitors. TACCs interact with a centrosomal
304 protein, CEP120, to regulate nuclear migration and the self-renewal of cortical neural

305 progenitor cells by controlling microtubule growth (63). A similar function is thought to
306 mediate the influence of microcephaly genes on brain development (18,64).

307 Beyond functional affects of individual genes, our approach has the potential to tackle
308 fundamental questions about how composite or modular tissues, such as the brain, evolve.
309 For example, two models of brain evolution dominate debates surrounding the adaptive
310 significance of variation in brain structure. One model proposes a conserved developmental
311 program drives a ‘concerted’ pattern of brain evolution, with selection shaping the overall
312 size of the system but not individual components(32,65). An alternative model instead argues
313 that different brain regions evolve independently of overall brain size to meet species-specific
314 behavioural needs, resulting in a ‘mosaic’ pattern of brain evolution, but may co-evolve due
315 to functional interdependence (21,66). These two models implicitly make contrasting
316 predictions about the genetic architecture of brain structure. The concerted model predicts
317 variation in the size of different brain regions will be determined by genetic correlations
318 between those structures, i.e. variation at a common set of genes. The mosaic model instead
319 predicts that the development of different brain regions must be at least partially distinct in
320 order to facilitate independent evolution.

321 In recent years these predictions have been tested using quantitative genetics within a
322 range of vertebrates, either using wild pedigrees (67–69), inbred strains (70) or divergent
323 populations/domestic breeds (71,72). In support of the mosaic brain hypothesis, these have
324 found little evidence for widespread genetic co-variation between major brain components.
325 Similarly, large genome-wide association studies within humans that have identified
326 independent genetic bases associated with brain regions (73,74). Quantitative genetics
327 assesses the phenotypic associations of standing genetic variation within populations and
328 their relevance to macroevolution depends on the relative frequency at which selection acts
329 on *de novo* mutations that may cause different patterns of genetic correlation. Our results
330 therefore complement these intra-specific studies, providing the first inter-specific test
331 designed to identify genes associated with neocortex and/or cerebellum evolution. These two
332 structures show a consistent pattern of co-evolution across primates (21,22,24), reflecting
333 their functional inter-dependence (23,25). Our analyses did not identify any gene that co-
334 varies with both structures, but does identify multiple genes with a specific association with
335 either the cerebellum or neocortex. This suggests the coevolution of these structures is
336 unlikely to be solely due to genetic integration or pleiotropy. If more broadly true, this
337 conclusion bolsters the interpretation of cortico-cerebellar coevolution as indicating adaptive
338 co-evolution, maintained by selection acting on distinct developmental pathways.

339 Comparative functional analysis of the genes highlighted by our analyses will be
340 necessary to confirm and extend these conclusions. These will also be needed to address key
341 questions beyond functional effects. For example, when disrupted, several of the genes
342 highlighted by our analyses affect the development of multiple organs. For example, both
343 *AGTPBP1* and *PAFAH1B* have known roles in spermatogenesis (75,76), whilst disruption of
344 *PCNT* can affect global somatic growth (56). If these genes do have a specific evolutionary
345 role in cerebellar development, how are these pleiotropic effects avoided? Similar questions
346 have been raised over previous candidate genes (3,18), further emphasising the importance of
347 coupling comparative and functional data.

348 In summary, we have presented an analyses aimed at providing an initial assessment
349 of the strength of selection targeting genes with development roles in distinct brain regions.
350 Although our understanding of gene ontology is incomplete, we illustrate how this
351 information can be used to test hypotheses in a phylogenetic comparative setting, in addition
352 to post-hoc enrichment analyses. We highlight that there is currently no evidence that
353 selection is limited or biased towards genes affecting cerebral cortex development, and
354 encourage evolutionary geneticists to adopt a cohesive view of brain evolution that
355 encompasses the recognised importance or non-allometric expansion of non-cortical regions,
356 and to tackle the central question of co-evolution and relative genetic independence of brain
357 components. Finally, we further illustrate the potential of hypothesis driven comparative
358 genetics in dissecting the genetic basis of phenotypic evolution. The ever-increasing numbers
359 of sequenced genomes will permit increasingly powerful analyses of the targets of selection
360 and their phenotypic relevance.

361

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366

367 **References**

- 368 1. Rausher MD, Delph LF. When does understanding phenotypic evolution require
369 identification of the underlying genes? *Evolution*. 2015;69(7):1655–64.
- 370 2. Enard W. Comparative genomics of brain size evolution. *Front Hum Neurosci*.
371 2014;8:345.
- 372 3. Montgomery SH, Capellini I, Venditti C, Barton RA, Mundy NI. Adaptive evolution
373 of four microcephaly genes and the evolution of brain size in anthropoid primates. *Mol*

- 374 Biol Evol. 2011;28(1):625–38.
- 375 4. Montgomery SH, Mundy NI. Evolution of *ASPM* is associated with both increases and
376 decreases in brain size in primates. *Evolution*. 2012;66(3):927–32.
- 377 5. Montgomery SH, Mundy NI. Positive selection on *NIN*, a gene involved in
378 neurogenesis, and primate brain evolution. *Genes, Brain Behav*. 2012;11(8):903–10.
- 379 6. Pollard KS, Salama SR, King B, Kern AD, Dreszer T, Katzman S, et al. Forces
380 shaping the fastest evolving regions in the human genome. *PLoS Genet*.
381 2006;2(10):1599–611.
- 382 7. Kamm GB, López-Leal R, Lorenzo JR, Franchini LF. A fast-evolving human *NPAS3*
383 enhancer gained reporter expression in the developing forebrain of transgenic mice.
384 *Philos Trans R Soc Lond B Biol Sci*. 2013;368:20130019.
- 385 8. Boyd JL, Skove SL, Rouanet JP, Pilaz L-J, Bepler T, Gordân R, et al. Human-
386 chimpanzee differences in a *FZD8* enhancer alter cell-cycle dynamics in the
387 developing neocortex. *Curr Biol*. 2015;772–9. A
- 388 9. Enard W, Przeworski M, Fisher SE, Lai CSL, Wiebe V, Kitano T, et al. Molecular
389 evolution of *FOXP2*, a gene involved in speech and language. *Nature*. 2002;418:869–
390 72.
- 391 10. Brawand D, Soumillon M, Necsulea A, Julien P, Csárdi G, Harrigan P, et al. The
392 evolution of gene expression levels in mammalian organs. *Nature*. 2011;478:343–8.
- 393 11. Enard W, Khaitovich P, Klose J, Zöllner S, Heissig F, Giavalisco P, et al. Intra- and
394 interspecific variation in primate gene expression patterns. *Science*. 2002;296:340–3.
- 395 12. Bauernfeind AL, Soderblom EJ, Turner ME, Moseley MA, Ely JJ, Hof PR, et al.
396 Evolutionary divergence of gene and protein expression in the brains of humans and
397 chimpanzees. *Genome Biol Evol*. 2015;7(8):2276–88.
- 398 13. Khaitovich P, Muetzel B, She X, Lachmann M, Hellmann I, Dietzsch J, et al. Regional
399 patterns of gene expression in human and chimpanzee brains. *Genome Res*.
400 2004;14:1462–73.
- 401 14. Burki F, Kaessmann H. Birth and adaptive evolution of a hominoid gene that supports
402 high neurotransmitter flux. *Nat Genet*. 2004;36(10):1061–3.
- 403 15. Florio M, Albert M, Taverna E, Namba T, Brandl H, Lewitus E, et al. Human-specific
404 gene *ARHGAP11B* promotes basal progenitor amplification and neocortex expansion.
405 *Science*. 2015;347(6229):1465–70.
- 406 16. Zimmer F, Montgomery SH. Phylogenetic analysis supports a link between DUF1220
407 domain number and primate brain expansion. *Genome Biol Evol*. 2015;7(8):2083–8.
- 408 17. Keeney J, Dumas L, Sikela J. The case for DUF1220 domain dosage as a primary
409 contributor to anthropoid brain expansion. *Front Hum Neurosci*. 2014;8:1–11.
- 410 18. Pulvers JN, Bryk J, Fish JL, Wilsch-Bräuninger M, Arai Y, Schreier D, et al.
411 Mutations in mouse *Aspm* (abnormal spindle-like microcephaly associated) cause not
412 only microcephaly but also major defects in the germline. *Proc Natl Acad Sci USA*.
413 2010;107(38):16595–600.
- 414 19. Enard W, Gehre S, Hammerschmidt K, Hölter SM, Blass T, Somel M, et al. A
415 humanized version of *Foxp2* affects cortico-basal ganglia circuits in mice. *Cell*.
416 2009;137:961–71.
- 417 20. Parvizi J. Corticocentric myopia: old bias in new cognitive sciences. *Trends Cogn Sci*.

- 418 2009;13:354–9.
- 419 21. Barton RA, Harvey PH. Mosaic evolution of brain structure in mammals. *Nature*.
420 2000;405(6790):1055–8.
- 421 22. Whiting BA, Barton RA. The evolution of the cortico-cerebellar complex in primates:
422 Anatomical connections predict patterns of correlated evolution. *J Hum Evol*.
423 2003;44:3–10.
- 424 23. Barton RA. Embodied cognitive evolution and the cerebellum. *Phil Trans Roy Soc B*.
425 2012;2097–107.
- 426 24. Herculano-houzel S, Sherwood CC. Coordinated scaling of cortical and cerebellar
427 numbers of neurons. *Front Neuroanat*. 2010;4:1–8.
- 428 25. Ramnani N. The primate cortico-cerebellar system: anatomy and function. *Nat Rev*
429 *Neurosci*. 2006;7:511–22.
- 430 26. Balsters JH, Cussans E, Diedrichsen J, Phillips KA, Preuss TM, Rilling JK, et al.
431 Evolution of the cerebellar cortex: The selective expansion of prefrontal-projecting
432 cerebellar lobules. *Neuroimage*. 2010;49(3):2045–52.
- 433 27. Macleod CE, Zilles K, Schleicher A, Rilling JK, Gibson KR. Expansion of the
434 neocerebellum in Hominoidea. *J Hum Evol*. 2003;44:401–29.
- 435 28. Rilling J, Insel T. Evolution of the cerebellum in primates: Differences in relative
436 volume among monkeys, apes and humans. *Brain Behav Evol*. 1998(52):308–14.
- 437 29. Barton RA, Venditti C. Rapid evolution of the cerebellum in humans and other Great
438 Apes. *Curr Biol*. 2014;24(20):2440–4.
- 439 30. Weaver AH. Reciprocal evolution of the cerebellum and neocortex in fossil humans.
440 *Proc Natl Acad Sci USA*. 2005;102(10):3576–80.
- 441 31. Ferland RJ, Eyaid W, Collura R V, Tully LD, Hill RS, Al-nouri D, et al. Abnormal
442 cerebellar development and axonal decussation due to mutations in *AH11* in Joubert
443 syndrome. *Nat Genet*. 2004;36(9):1008–13.
- 444 32. Finlay B, Darlington R. Linked regularities in the development and evolution of
445 mammalian brains. *Science*. 1995;268(5217):1578–84.
- 446 33. Carbon S, Ireland A, Mungall CJ, Shu S, Marshall B, Lewis S, et al. AmiGO: online
447 access to ontology and annotation data. *Bioinformatics*. 2009;25(2):288–9.
- 448 34. Binns D, Dimmer E, Huntley R, Barrell D, O’Donovan C, Apweiler R. QuickGO: a
449 web-based tool for Gene Ontology searching. *Bioinformatics*. 2009;25(22):3045–6.
- 450 35. Stephan H, Frahm H, Baron G. New and revised data on volumes of brain structures in
451 Insectivores and Primates. *Folia Primatol*. 1981;35(1):1–29.
- 452 36. Altschup SF, Gish W, Miller W, Myers E, Lipman D. Basic Local Alignment Search
453 Tool. *J Mol Biol*. 1990;(215):403–10.
- 454 37. Löytynoja A, Goldman N. webPRANK: a phylogeny-aware multiple sequence aligner
455 with interactive alignment browser. *BMC Bioinform*. 2010;579.
- 456 38. Yang Z. PAML 4: Phylogenetic analysis by maximum likelihood. *Mol Biol Evol*.
457 2007;24(8):1586–91.
- 458 39. Harrison PW, Jordan GE, Montgomery SH. SWAMP : Sliding Window Alignment
459 Masker for PAML. *Evol Bioinform Online*. 2014;197–204.
- 460 40. Montgomery SH, Capellini I, Venditti C, Barton RA, Mundy NI. Adaptive evolution

- 461 of four microcephaly genes and the evolution of brain size in anthropoid primates. *Mol*
462 *Biol Evol.* 2011;28(1):625–38.
- 463 41. Pagel M. Inferring the historical patterns of biological evolution. *Nature.*
464 1999;401(6756):877–84.
- 465 42. Benjamini Y, Hochberg Y. Controlling the false discovery rate: a practical and
466 powerful approach to multiple testing. *J Stat Soc B.* 1995;57(1):289–300.
- 467 43. Anisimova M, Yang Z. Multiple hypothesis testing to detect lineages under positive
468 selection that affects only a few sites. *Mol Biol Evol.* 2007;24(5):1219–28.
- 469 44. Barton RA, Venditti C. Human frontal lobes are not relatively large. *Proc Natl Acad*
470 *Sci USA.* 2013;110(22):9001–6.
- 471 45. Joubert M, Eisenring J, Robb JP, Andermann F. Familial agenesis of the cerebellar
472 vermis A syndrome of episodic hyperpnea, abnormal eye movements, ataxia, and
473 retardation. *Neurology.* 1969;19(9):813.
- 474 46. Doherty D. Joubert Syndrome: Insights into brain. *Semin Pediatr Neurol.*
475 2009;16(3):143–54.
- 476 47. Arts HH, Doherty D, Beersum SEC Van, Parisi MA, Letteboer SJF, Voesenek K, et al.
477 Mutations in the gene encoding the basal body protein *RPGRIP1L*, a nephrocystin-4
478 interactor, cause Joubert syndrome. *Nat Genet.* 2007;39(7):882–8.
- 479 48. Delous M, Baala L, Saloman R, Laclef C, Vierkotten J, Golzio C, et al. The ciliary
480 gene *RPGRIP1L* is mutated in cerebello-oculo-renal syndrome (Joubert syndrome type
481 B) and Meckel syndrome. *Nat Genet.* 2007;39(7):875–81.
- 482 49. Spassky N, Han Y, Aguilar A, Strehl L, Besse L, Laclef C, et al. Primary cilia are
483 required for cerebellar development and Shh-dependent expansion of progenitor pool.
484 *Dev Biol.* 2008;317:246–59.
- 485 50. Bronson RT, Donahue LR, Samples R, Kim JH, Naggert JK. Mice with mutations in
486 the mahogany gene *Atrn* have cerebral spongiform changes. *J Neuropathol Exp*
487 *Neurol.* 2001;60(7):724–30.
- 488 51. He L, Lu X, Jolly AF, Eldridge AG, Watson SJ, Jackson PK, et al. Spongiform
489 degeneration in mahoganoid mutant mice. *Science.* 2003;299:710–2.
- 490 52. Wilkinson LS, Davies W, Isles AR. Genomic imprinting effects on brain development
491 and function. *Nat Rev Neurosci.* 2007;8(11):832–43.
- 492 53. Singhmar P, Kumar A. Angelman syndrome protein *UBE3A* interacts with primary
493 microcephaly protein *ASPM*, localizes to centrosomes and regulates chromosome
494 segregation. *PLoS One.* 2011;6(5):e20397.
- 495 54. Meglio T Di, Kratochwil CF, Vilain N, Loche A, Vitobello A, Yonehara K, et al. *Ezh2*
496 orchestrates topographic migration and connectivity of mouse precerebellar neurons.
497 *Science.* 2013;339(6116):204–7.
- 498 55. Lalonde R, Strazielle C, Inserm U, Electronique SDM, Médecine F De. Spontaneous
499 and induced mouse mutations with cerebellar dysfunctions: Behavior and
500 neurochemistry. *Brain Res.* 2006;1140:51–74.
- 501 56. Rauch A, Thiel CT, Schindler D, Wick U, Crow YJ, Ekici AB, et al. Mutations in the
502 Pericentrin (*PCNT*) gene cause primordial dwarfism. *Science.* 2008;319:816–9.
- 503 57. Haavik J, Toska K. Tyrosine Hydroxylase and Parkinson's disease. *Mol*
504 *Neurobiol.* 1998;16(3):285–309.

- 505 58. Ludecke B, Dworniczak B, Bartolome K. A point mutation in the tyrosine hydroxylase
506 gene associated with Segawa's syndrome. *Hum Genet.* 1995;6:123–5.
- 507 59. Liu YF, Sowell SM, Luo Y, Chaubey A, Cameron RS, Kim H, et al. Autism and
508 intellectual disability-associated KIRREL3 interacts with neuronal proteins MAP1B
509 and MYO16 with potential roles in neurodevelopment. *PLoS One.*
510 2015;10(4):e0123106.
- 511 60. Brodsky G, Barnes T, Bleskan J, Becker L, Cox M, Patterson D. The human *GARS-*
512 *AIRS-GART* gene encodes two proteins which are differentially expressed during
513 human brain development and temporally overexpressed in cerebellum of individuals
514 with Down syndrome. *Hum Mol Genet.* 1997;6(12):2043–50.
- 515 61. Kawase-koga Y, Otaegi G, Sun T. Different timings of Dicer deletion affect
516 neurogenesis and gliogenesis in the developing mouse central nervous system. *Deve*
517 *Dyn.* 2009;238(11):2800–12.
- 518 62. Davis TH, Cuellar TL, Koch SM, Barker AJ, Harfe BD, McManus MT, et al.
519 Conditional loss of Dicer disrupts cellular and tissue morphogenesis in the cortex and
520 hippocampus. *J Neurosci.* 2008;17:4322–30.
- 521 63. Xie Z, Moy LY, Sanada K, Zhou Y, Buchman JJ, Tsai L. Cep120 and TACCs control
522 interkinetic nuclear migration and the neural progenitor pool. *Neuron.* 2007;56(1)79–
523 93.
- 524 64. Thornton GK, Woods CG. Primary microcephaly: do all roads lead to Rome? *Trends*
525 *Genet.* 2009;25(11):501–10.
- 526 65. Finlay BL, Darlington RB, Nicastro N. Developmental structure in brain evolution.
527 *Behav Brain Sci.* 2001;24:263–78; discussion 278–308.
- 528 66. de Winter W, Oxnard CE. Evolutionary radiations and convergences in the structural
529 organization of mammalian brains. *Nature.* 2001;409(6821):710–4.
- 530 67. Fears SC, Melega WP, Service SK, Lee C, Chen K, Tu Z, et al. Identifying heritable
531 brain phenotypes in an extended pedigree of vervet monkeys. *J Neurosci.*
532 2009;29(9):2867–75.
- 533 68. Rogers J, Kochunov P, Zilles K, Shelledy W, Lancaster J, Thompson P, et al. On the
534 genetic architecture of cortical folding and brain volume in primates. *Neuroimage.*
535 2010;53(3):1103–8.
- 536 69. Rogers J, Kochunov P, Lancaster J, Shelledy W, Glahn D, Blangero J, et al.
537 Heritability of brain volume, surface area and shape: An MRI study in an extended
538 pedigree of baboons. *Hum Brain Mapp.* 2007;28:576–83.
- 539 70. Hager R, Lu L, Rosen GD, Williams RW. Genetic architecture supports mosaic brain
540 evolution and independent brain–body size regulation. *Nat Commun.* 2012;3:1079.
- 541 71. Henriksen R, Andersson L, Jensen P, Wright D. From the jungle to the barn:
542 Independent genetic control for increased brain and body size and Mosaic brain
543 evolution in chickens during domestication. In review.
- 544 72. Noreikiene K, Herczeg G, Gonda A, Balazs G, Husby A, Merilä J. Quantitative
545 genetic analysis of brain size variation in sticklebacks: support for the mosaic model of
546 brain evolution. *Proc Roy Soc B.* 2015;282(1810):20151008.
- 547 73. Toro R, Poline J-B, Huguet G, Loth E, Frouin V, Banaschewski T, et al. Genomic
548 architecture of human neuroanatomical diversity. *Mol Psychiatry.* 2015;20(8):1011-16.

- 549 74. Hibar DP, Stein LP, Renteria ME, Arias-Vasquez A, Desrivieres S, Jahanshad N, et al.
550 Common genetic variants influence human subcortical brain structures. *Nature*.
551 2015;520(7546):224-229.
- 552 75. Kim N, Xiao R, Choi H, Jo H, Kim J, Uhm S, et al. Abnormal spermatid development in
553 *pcd 3J - / -* Mice: the Importance of *Agtpbp1* in spermatogenesis. *Mol Cell*.
554 2011;(2005):39-48.
- 555 76. Yan W, Assadi AH, Wynshaw-boris A, Eichele G, Matzuk MM, Clark GD. Previously
556 uncharacterized roles of platelet-activating factor acetylhydrolase 1b complex in
557 mouse spermatogenesis. *Proc Natl Acad Sci USA*. 2003;100(12):7189-94.
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585 **Figure legends:**

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587 **Figure 1 A)** Phylogeny of the 11 anthropoids included in the study: hominoid lineages are
588 shown in orange. **B-D)** Pie-charts showing the proportion of genes significant (darker colour)
589 under each test for cerebral cortex (red) and cerebellum genes (green). The p-value from a
590 Fisher's Exact test comparing the groups is shown below.

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592 **Figure 2 Phenotypic associations for two genes A) *RGRIP1L* and B) *ATRN*.** The \log_{10} -
593 transformed root-to-tip (rtt-) dN/dS is plotted against residual cerebellum volume, calculated
594 from a PGLS regression of cerebellum against neocortex volume. The black line shows the
595 result of a phylogenetically-controlled regression between rtt- dN/dS and residual cerebellum
596 volume. This figure is for illustrative purposes, in the main analyses the two brain
597 components were included as variables in a multiple regression.

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614 **Tables**

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616 **Table 1: summary of results of dN/dS analyses and tests of positive selection**

Test	Inference	Significant results (proportion)		Difference in proportion	
		Cerebral cortex (n = 47)	Cerebellum (n = 53)	Z-test	Fisher's Exact Test
Site model test	Pervasive positive selection	5 (9.4%)	3 (6.4%)	$z = 0.561, p = 0.575$	$p = 0.716$
Branch model test	Accelerated evolution in apes	6 (11.3%)	14 (29.8%)	$z = 2.304, p = 0.021$	$p = 0.026$
Branch-site model test (all)	Episodic positive selection in apes	2 (3.8%)	6 (12.8%)	$z = 1.654, p = 0.099$	$p = 0.143$
Branch-site model test (restricted ¹)	Episodic positive selection in apes	0 (0.0%)	4 (10.0%)	$z = 2.136, p = 0.049$	$p = 0.032$

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619 ¹ excludes genes with significant site-model results



