

# 1                    **Encoding of locomotion kinematics in the mouse cerebellum**

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## 8                    **Abstract**

9                    The cerebellum has a well-established role in locomotion control, but how the cerebellar network  
10                    regulates locomotion behaviour is still not well understood. We therefore characterized the activity  
11                    of cerebellar neurons in awake mice engaged in a locomotion task, using high-density silicon electrode  
12                    arrays. We characterized the activity of over 300 neurons in response to locomotion, finding tuning  
13                    to speed of locomotion, turning, and phase of the step cycle. We found that the cerebellar neurons  
14                    we recorded mainly encoded information about future locomotor activity. We were able to decode the  
15                    speed of locomotion with a simple linear algorithm, needing relatively few well-chosen cells to provide  
16                    an accurate estimate of locomotion speed. Our observation that cerebellar neuronal activity predicts  
17                    locomotion in the near future, and encodes the required kinematic variables, points to this activity  
18                    underlying the efference copy signal for vertebrate locomotion.

## 19                    **Introduction**

20                    An animal's survival relies heavily upon its ability to locomote through space. The ethological im-  
21                    portance of locomotion is reflected in the large proportion of the central nervous system involved in  
22                    locomotor activity. One such area is the cerebellum, whose function was long ago established through  
23                    clinical and lesion studies to be essential for learning and controlling movements (Flourens, 1824;  
24                    Luciani, 1891; Holmes, 1939). Being located, in circuit terms, between higher cortical centres and the  
25                    periphery, the cerebellum acts as a strategic relay point for sensorimotor integration.

26                    Electrophysiological studies combined with the analysis of behaviour provided direct evidence for  
27                    the role of the cerebellum in locomotor control and learning. The spinocerebellum, the central part of  
28                    the cerebellum, receives projections from the spinal cerebellar tract neurons which convey peripheral  
29                    sensory signals and information from the spinal pattern generator (Arshavsky et al., 1983; Fedirchuk

30 et al., 2013). In particular, the medial zone of the spinocerebellum, the vermis, has been identified as  
31 the area involved in controlling posture, tone, flexion and extension of limbs (Chambers and Sprague,  
32 1955).

33 The spinocerebellar tracts, which are part of the locomotion circuitry (Goulding, 2009), were found  
34 to be essentially preserved across animal species, including mice (Oscarsson, 1965; Berretta et al., 1991;  
35 Sengul et al., 2015). The mouse is a model organism of particular interest due to its suitability for the  
36 use of transgenic technology to dissect out the contributions of individual circuit elements. In recent  
37 years, the application of transgenic techniques to mouse experiments provided new insights into the  
38 neural circuits involved in locomotion (Akay et al., 2014; Bellardita and Kiehn, 2015; Kiehn, 2016),  
39 and the role of the cerebellum in motor and cognitive functions (Reeber et al., 2013; Zhou et al., 2014;  
40 Galliano and De Zeeuw, 2014; Hoogland et al., 2015).

41 Observation of neural activity in the cerebellum has revealed that many cerebellar neuron types  
42 carry locomotion-related information. Purkinje cells, the only output of the cerebellum, are essential  
43 to interlimb coordination, adaptation to external perturbation, and they tend to fire rhythmically  
44 with the stepping cycle (Yanagihara and Kondo, 1996; Ichise et al., 2000; Orlovsky, 1972; Armstrong  
45 and Edgley, 1984). Although Purkinje cells in the cat were not observed to have substantial firing  
46 rate modulation by walking speed on a treadmill (Armstrong and Edgley, 1988), it was recently  
47 observed that the firing rate of Purkinje cells, averaged within single steps, can be modulated both  
48 negatively and positively with locomotion speed in freely running rats (Sauerbrei et al., 2015). Golgi  
49 cells were also shown to discharge rhythmically during locomotion, however no modulation by the  
50 speed of locomotion was observed in this case (Edgley and Lidieth, 1987). In contrast, granule cells  
51 and molecular layer interneurons of mice on a spherical treadmill increased their firing rate during  
52 locomotion relative to stationary periods (Ozden et al., 2012; Powell et al., 2015), leaving open the  
53 question of whether and how cerebellar neurons are tuned to the speed of locomotion.

54 Electrophysiological recordings of single units in the cerebellum validated the relationship between  
55 behaviour and neural activity, but have thus far failed to account for population dynamics, providing a  
56 confined view of the cerebellar neural code. In fact, cerebellar circuitry is characterised by a high degree  
57 of feedback, feed-forward and collateral connections (Ito, 2006; Coddington et al., 2013; Mathews et  
58 al., 2012; Rieubland et al., 2014; Astorga et al., 2015), and a distinctive divergence-convergence  
59 configuration of inputs and outputs (Napper and Harvey, 1988; Person and Raman, 2012). There are  
60 therefore important unanswered questions as to how locomotion-related information is conveyed by  
61 ensembles of cerebellar neurons, and what type of neural population code is employed.

62 To address these questions, we recorded from movement-sensitive populations of neurons in lobules

63 V and VI of the cerebellar vermis of mice navigating in a virtual reality (VR) environment. We  
64 characterised neurons whose activity is modulated by kinematics parameters such as locomotion speed  
65 and yaw rotation. The combined activity of these neurons linearly decodes locomotor speed with  
66 an accuracy proportional to the population size, providing new insight on the neural code of the  
67 cerebellum.

## 68 **Results**

69 Mice (n=14, 16-24 weeks old) were head-fixed on an air-supported spherical treadmill inside a demi-  
70 spherical screen (Figure 1A; Holscher et al., 2005; Harvey et al., 2009). Sphere pitch and yaw move-  
71 ments generated by the mouse were read by two optical computer mice, and integrated in order to  
72 determine the translation and heading of the animal in the virtual space. Visual stimuli were con-  
73 trolled in closed-loop using custom-developed LabView software (see Methods). The mice navigated  
74 through a virtual corridor along which they received water at defined reward points (green cylinders,  
75 Figure 1B). Following behavioural training (Figure 1C and Figure 1 - figure supplement 1), multi-  
76 electrode array extracellular recordings were made from lobules V and VI of the cerebellar vermis  
77 (Figure 1D). Four animals did not receive behavioural training, but instead were allowed to run in  
78 the dark, with the virtual reality stimulus switched off, as a control group to discern the influence  
79 of visual feedback on locomotor speed encoding. Animals spent on average  $57 \pm 3\%$  (mean  $\pm$  s.e.m.,  
80 n=39) of each recording period running (defined as speed exceeding 1 cm/s). For each animal, the  
81 recording electrode was positioned at a number of different depths (39 recordings in total, 311 units;  
82 see Figure 1E). Action potentials were detected and clustered to perform spike sorting (Rossant et al.,  
83 2016). For each identified unit, cell type was determined according to recently published classification  
84 criteria (Figure 1F and Figure 1 - figure supplement 2, Van Dijck et al., 2013; Hensbroek et al., 2014).

### 85 **Cerebellar neurons respond to speed of locomotion**

86 The activity of many units correlated with the behavioural status of the mouse, i.e. the firing rate  
87 changed with speed (Figure 2A-C). To determine whether neural activity correlated with speed of  
88 locomotion, we computed speed tuning curves for the firing rate of each unit (Figure 2D-F). To assess  
89 the significance of speed modulation, we shuffled the data 100 times and compared the variance of  
90 the original curve with that computed from the shuffled data. If this variance was greater than the  
91 values of at least 99 shuffled curves, the unit would be considered to be significantly speed modulated  
92 (Saleem et al., 2013; Kropff et al., 2015). We also checked whether the changes in firing rate were  
93 due only to changes in excitability between stationary and moving periods (i.e. if cells were driven

94 by locomotion, but their activity was not modulated by speed) by repeating the above procedure  
95 but only considering speeds  $>1$  cm/s. 159 units were found to respond to locomotion movements:  
96 20 showed a binary response to movement and the remaining 139 were modulated by speed. For  
97 these units, three classes of modulation profile (tuning class) were observed: units whose firing rate  
98 monotonically increased ( $n=50$ ) or decreased ( $n=51$ ) with speed, and units whose firing rate reached  
99 its maximum at a preferred speed that is  $\leq 70\%$  the maximal speed achieved by the mouse ( $n=38$ ).  
100 Similar profile responses were observed in naïve (untrained) mice and no differences were found between  
101 the responses of units recorded from these and trained animals. These units were therefore analysed  
102 conjointly (Figure 2 - supplement figure 1A).

103 Positively modulated units, on average, had lower spontaneous (animal at rest) firing rates com-  
104 pared to those cells whose firing rate decreased with speed (Figure 2G,H). Firing rate changed from  
105  $24.7 \pm 4.6$  Hz during resting periods to  $52.2 \pm 7$  Hz (mean  $\pm$  s.e.m.,  $n=50$ ) at maximal locomotion speed.  
106 For negatively modulated units, firing rate decreased from  $50 \pm 7.6$  Hz under the resting condition to  
107  $30 \pm 6.1$  Hz (mean  $\pm$  s.e.m.,  $n=51$ ) during locomotion at maximal speed. Units showing a preferred speed  
108 had on average less marked changes going from  $19.3 \pm 4.2$  Hz at rest up to  $40 \pm 5.7$  Hz (mean  $\pm$  s.e.m.,  
109  $n=38$ ) at the maximum speed observed (Figure 2I).

110 We examined whether the modulation of single cells differed within and across response tuning  
111 classes (Figure 2 - supplement figure 2A). The modulation index was defined as the ratio between the  
112 difference and the sum of the maximum and minimum firing rates measured on each tuning curve.  
113 Modulation indexes for all response types varied heterogeneously across the whole range. We found  
114 that, for all response tuning classes, the modulation of firing rate was negatively correlated with  
115 spontaneous firing rate (Figure 2 - supplement figure 1B).

116 Units responsive to movement were observed in all animals, with no discernible dependence on  
117 depth of recording site (Figure 2 - supplement figure 3). Units belonging to the same response tuning  
118 class were not observed to cluster spatially: in only 8 out of 39 recordings did we find units belonging  
119 to the same response class in close proximity (i.e. in the same electrode shank). In all remaining  
120 recordings ( $n=27$ ) in which we found multiple units responsive to movement in close proximity, their  
121 response type was heterogeneous.

122 Taken together, these results demonstrate that cerebellar neurons respond to locomotion speed by  
123 either increasing or decreasing their firing rate or responding maximally to a particular speed. Further-  
124 more, we did not find spatial clustering of units with the same tuning profile class. This observation  
125 suggests that nearby cerebellar neurons, possibly belonging to the same micro-zone (Oscarsson, 1979),  
126 encode locomotion-related movements by combining different types of sensorimotor information.

## 127 **A subset of cerebellar neurons display yaw direction tuned responses**

128 While animals ran in the virtual corridor, they corrected their trajectories repeatedly to reach the  
129 target location marking the end of each trial. As the animals exerted more strength on either one or  
130 the other side of the body when turning the sphere, we examined whether this asymmetric use of limb  
131 muscles was reflected in cerebellar neuronal activity. By extracting the yaw movement information  
132 from the motion sensors, we examined how the firing rate changed with respect to sphere rotations  
133 around the clockwise (CW, negative yaw) and counter-clockwise (CCW, positive yaw) direction (ex-  
134 ample trace, top Figure 3A).

135 To do so, we computed tuning curves for neural activity in response to CW and CCW yaw direc-  
136 tions, and calculated the modulation index for each cell in the two directions. For a few units, the  
137 neural response was clearly one-sided (Figure 3A). This was reflected in their tuning curves, and in  
138 the absolute difference between the modulation indexes of the tuning curves computed for the neg-  
139 ative (CW) and positive (CCW) yaw directions (delta yaw modulation index - Figure 3B,C). Most  
140 cells, however, responded equally to either yaw direction, showing a decrease (Figure 3D) or increase  
141 (Figure 3E) in firing rate to either CW or CCW yaw movement, in an almost symmetric fashion.  
142 This is reflected in the distribution of the delta yaw modulation indexes: 63% (195 out of 311) had  
143 a difference in modulation indexes smaller than 0.1, while only 19% (58 out of 311) was greater than  
144 0.2 (Figure 3F), indicating sensitivity to yaw direction. In fact many units had similar modulation  
145 indexes for both the CW and CCW yaw directions (Figure 3G). We also examined whether tuning  
146 to speed influenced the yaw-direction selectivity of the 58 cells that had delta yaw modulation index  
147 greater than 0.2. Interestingly, 32 out of the 58 yaw direction sensitive units did not modulate their  
148 activity with speed (Figure 3 - supplement figure 3A). There was also no evident relationship between  
149 the speed modulation index and delta yaw modulation index (Figure 3 - supplement figure 3B). These  
150 results suggest that there are units in the cerebellum that respond selectively to the use of one side of  
151 the body in preference to the other, and that these cells are not necessarily influenced by changes in  
152 speed.

## 153 **A subset of cerebellar neurons are tuned to phase of stepping cycle**

154 Previous studies of the role of the cerebellum in locomotion control found that Purkinje cells are  
155 rhythmically modulated by stepping cycle (Orlovsky, 1972; Armstrong and Edgley, 1984; Armstrong  
156 and Edgley, 1988; Edgley and Lidierth, 1987; Powell et al., 2015; Sauerbrei et al., 2015). We reasoned  
157 that one way to produce speed tuning would be for units to respond by spiking at a fixed phase per  
158 step cycle, thus producing higher firing rates the more step cycles occur per unit of time. We therefore

159 examined whether this was the reason behind the tuning of activity to locomotion speed. Since pitch  
160 velocity is measured by motion sensors parallel to the vertical axis, with velocity sampled at a high  
161 polling rate ( $f=200$  Hz), it was possible to detect the vertical oscillations caused by the mouse stepping  
162 on the sphere (Figure 4A). The high frequency periodicity of the signal was extracted from the original  
163 pitch velocity signal by transforming the pitch velocity with the Hilbert operator. At least 393 putative  
164 step cycles were found for each recording session ( $1724\pm 158$ , mean $\pm$ s.e.m.,  $n=39$ ). We then measured  
165 the correlation between the firing rate (bin width = 5ms, smoothed with a 20-ms Gaussian window)  
166 and the phase of the step cycle for each unit (Figure 4B) finding that only 57 units out of 311 showed  
167 a significant modulation with the stepping cycle ( $p\leq 0.001$ ,  $\chi^2$  test for uniformity). Of these, thirty-  
168 two units were also significantly modulated by speed. Mean preferred phases of the modulated units  
169 were distributed across the stepping cycle, with approximately two-thirds of the response covering on  
170 average  $2.2\pm 0.06$  (mean $\pm$ s.e.m,  $n=57$ ) radians, as measured by the standard circular deviation (Drew  
171 and Doucet, 1991) (Figure 4B).

172 To quantify the step phase modulation of the response, we used an approach commonly used  
173 in the study of orientation tuning, an analogous problem (where here phase within a step cycle  
174 replaces orientation within a circular stimulus space). We calculated the normalised phase orientation  
175 vector and computed the orientation selectivity index (Mazurek et al., 2014), renamed here the phase  
176 selectivity index, PSI. Units significantly modulated by step phase have higher phase selectivity indexes  
177 in comparison to non-modulated units ( $p=2e^{-16}$ , Mann-Whitney U-test, Figure 4D). These results  
178 suggest that cerebellar neurons' activity encodes kinematic information, i.e. locomotion speed is not  
179 a by-product of rhythmic modulation of the stepping cycle as shown in a previous study (Sauerbrei  
180 et al., 2015).

## 181 **Cerebellar units compute multiple kinematic parameters**

182 We have described units tuned for speed of locomotion, yaw (including some tuned for direction of yaw  
183 motion), and phase of stepping cycle. It is important to determine whether these constitute separate  
184 classes of neurons, or if instead, each of the neurons displays tuning to a lesser or greater extent across  
185 each dimension.

186 For each cell, we therefore compared the modulation index for speed with the mean modulation  
187 index for CW and CCW yaw (see Methods), and with the phase selectivity index. These are depicted  
188 in a tri-plot in Figure 5A. It is apparent that speed, yaw and phase selective units do not cluster  
189 in this space, but are instead distributed relatively uniformly. A similar picture arises when instead  
190 comparing the mutual information units convey about speed, yaw and step phase (Figure 5B). As

191 speed and yaw are not independent - indeed, speed is comprised of both a pitch and a yaw component  
192 - we broke speed tuning up into these components and assessed them separately. Figure 5C shows that  
193 units that conveyed substantial information about pitch also tended to convey substantial information  
194 about yaw - and that in fact, more of the speed information arose from yaw than from pitch signals.  
195 As bimodal distributions were not found in the information conveyed about speed, pitch, yaw nor  
196 step phase, we conclude that the cerebellar units examined were a relatively homogeneous population  
197 encoding all of these quantities to a greater or lesser extent, rather than a heterogeneous population  
198 comprising clusters of units encoding different kinematic variables.

### 199 **Cerebellar neurons mainly encode motor information**

200 In order to understand whether the recorded units encode information about descending motor com-  
201 mands, we analysed the timing of the information conveyed by neuronal responses about locomotion  
202 speed, reasoning that for motor units, the firing rate should provide predictive information about  
203 locomotion speed, whereas for sensory units, the information should be largely retrospective. We  
204 computed the mutual information (Schultz, Ince, et al., 2015) between the firing rate and locomotion  
205 speed time courses for each neuron, for a range of imposed time lags. Firing rates were shifted with  
206 respect to the speed signal by 10 ms from -500 to +500 ms (Figure 6A). While there were retrospective  
207 units (peak mutual information at negative time lag), the majority of speed modulated units showed  
208 a peak in the mutual information at a positive time lag of  $108.1 \pm 17.2$  ms (mean  $\pm$  s.e.m.,  $n=139$ ), sug-  
209 gesting that the neurons observed may primarily encode descending motor signals rather than sensory  
210 feedback (Figure 6B).

211 The values of maximal mutual information were significantly higher than those of non-modulated  
212 units ( $p=2.3e^{-9}$ , Mann-Whitney U-test):  $0.24 \pm 0.01$  bits (mean  $\pm$  s.e.m.,  $n=139$ ) for speed modulated  
213 units and  $0.13 \pm 0.01$  bits (mean  $\pm$  s.e.m.,  $n=172$ ) for unresponsive units. In fact, the lower information  
214 values - calculated at zero lag - for the latter class are consistent with the classification based on the  
215 tuning curve modulation approach (Figure 6 - figure supplement 1).

### 216 **Speed of locomotion can be linearly decoded from cerebellar neuronal ensembles**

217 It has been previously shown that neural activity of single Purkinje cells encode multiple kinematic  
218 parameters of multi-joint movements during arm reaching tasks in primates (Roitman, 2005; Pasalar  
219 et al., 2006; Hewitt et al., 2011). Similarly, locomotion requires the coordination of multiple joints and  
220 muscles. Since the majority of units we found was shown to encode multiple kinematics parameters,  
221 we set out to find out whether locomotion speed could be accurately reconstructed by populations of

222 cerebellar neurons.

223 To this end, we developed an optimal linear estimator (OLE) with the aim to reconstruct the  
224 locomotion speed time course from a weighted sum of the firing rates of each unit (Figure 7A). The  
225 weights were adjusted by linear regression, so as to minimise the mean squared error (MSE) between  
226 the decoded and the original recorded trace. The decoder was trained on 70% of the data of each  
227 recording and tested on the remaining 30%. To assess the scaling of decoder performance with ensemble  
228 size, we selected recordings comprising at least eight units ( $n=6$ ).

229 We investigated how the population size affected the accuracy of the decoder. To avoid any bias  
230 in the choice of units used in the reconstruction, these were selected randomly for any given size of  
231 neural population.  $2N$  possible population combinations were tested for ensembles of 1, 2,  $N - 1$  and  
232  $N$  units ( $N$  number of units recorded in the experiment), and  $N^2$  otherwise. As the number of units  
233 increased, so did the decoder accuracy whilst the decoding performance range of the random selected  
234 ensembles reduced (Figure 7B). Furthermore, as the population size increased, the median accuracy of  
235 locomotion speed reconstruction approached the one obtained using populations formed by the most  
236 correlated units only, as measured by the Pearson correlation coefficient.

237 These results are consistent across all experiments that contained more than seven units (Figure  
238 7C) suggesting that the cerebellum encodes kinematic information related to instantaneous locomotion  
239 speed by linearly summing the contributions of single neurons.

## 240 Discussion

241 In this study, we used a virtual reality behavioural task, together with multi-unit electrophysiological  
242 recording, to investigate the neuronal population activity underlying locomotion. Our approach al-  
243 lowed us to assess the activity of multiple neighbouring neurons during behaviour while maintaining  
244 a high degree of experimental control over behavioural parameters. Our recordings from lobules V  
245 and VI of the vermis indicate that most cells in this area encode kinematic parameters of locomotion.  
246 We found that while 6% of the cells showed significantly different firing rates during locomotion as  
247 opposed to rest, but no significant modulation by speed (similar to the classical results of Armstrong  
248 and Edgley (1984)), 45% of neurons were specifically tuned for speed of locomotion. This included  
249 cells that increased their firing rate with increasing running speed, cells that decreased their firing  
250 rate below the spontaneous (rest) level with increasing speed, and cells showing a preferred loco-  
251 motion speed. We also found that some cells were tuned to yaw (turning) and also to particular phases of  
252 the locomotion step cycle. The different responses of individual cells that we observed to locomotion  
253 speed, turning and stepping may reflect interdependent information about behavioural state, laterality

254 and type of muscles being exerted during locomotion.

255 While we have employed and extended recently developed cell classification algorithms (Van Dijk  
256 et al., 2013; Hensbroek et al., 2014), our approach is limited by the lack of certainty about cell identity.  
257 We were able to identify Purkinje cells, granule cells, and interneurons, however we were not able to  
258 classify interneurons into specific classes with any degree of confidence, and our approach did leave  
259 a significant number of unclassified neurons. In the present study, for technical reasons we did not  
260 attempt to analyse complex spike (CS) waveforms from Purkinje cells, restricting our attention to  
261 simple spike (SS) waveforms. CS will be the subject of a future manuscript. Further improvements  
262 to cell classification algorithms will probably require obtaining ground truth validation data through  
263 simultaneous MEA and sharp electrode or whole cell patch clamp recording in the awake animal,  
264 with histological validation, a technically challenging task. In our study, however, we did not find the  
265 encoding of locomotion kinematic parameters to be dependent upon cell class.

266 Kinematic parameters of arm movements have been found to be related to single neuron activity  
267 in the cerebellum (Hewitt et al., 2011; Popa et al., 2012) and in the motor cortex (Ashe and Geor-  
268 gopoulos, 1994; Yu et al., 2007) of non-human primates. It has recently been reported that Purkinje  
269 cells discharge rhythmically during locomotion (Sauerbrei et al., 2015), and that granule cells fire in  
270 bursts at locomotion onset (Powell et al., 2015). Ozden et al. (2012) reported finding molecular layer  
271 interneuron activity to increase following transition from a resting to a moving (locomotion) state.  
272 Sauerbrei et al. (2015) did report modulation by speed of some of the cells in their dataset (recorded  
273 from the freely moving rat), but did not describe speed tuning further, instead focusing on step-phase  
274 dependent correlations with behaviour. Moreover, vestibular inputs were not taken into account in  
275 their modulation analysis; such inputs were controlled in our study, as we used head-fixed animals  
276 for which these inputs can be considered negligible. To the best of our knowledge, the current study  
277 is the first description of the tuning of cerebellar neurons to speed of locomotion. One question that  
278 may arise is why this was not observed previously, for instance in earlier studies of cats walking on  
279 a treadmill. While it is not at this stage possible to rule out that inter-species differences account  
280 for this, our view is that the discrepancy is more likely to arise from the fact that in these studies  
281 the cats passively stepped on a treadmill, which was rotating at a rate fixed by the experimenter.  
282 Instead, in our study, the animals actively locomoted at a speed of their own choice (starting and  
283 stopping as they wished), motivated by an increasing reward rate for more rapid progress down the  
284 corridor segments, which may engage cerebellar networks to a greater degree. In fact, a small amount  
285 of speed modulation is apparent in Fig. 2 of Armstrong and Edgley (1988) for most of the neurons  
286 they recorded, suggesting that the situation in the mouse and cat may not be completely dissimilar.

287 By allowing the spherical treadmill to rotate along any orientation (rather than constraining it  
288 to a single axis as with a treadmill), we aimed to create a navigation paradigm congruent with a  
289 real-world scenario. Animals had to intentionally engage many muscles relevant for locomotion, and  
290 were not constrained to run at fixed speeds. However, the locomotion task should be thought of as a  
291 sensorimotor control task, rather than as normal locomotion behaviour, because of the artificial nature  
292 of the head fixation and of the act of balancing on a frictionless ball, which is itself an acquired skill.  
293 Because the animals were head-fixed, we assumed that vestibular inputs were negligible during yaw  
294 (turning) movements. The correlation of neuronal activity with the direction of movement may be  
295 related to lateralised spinocerebellar inputs from muscles employed to steer clockwise or anti-clockwise.  
296 We did not find cells with a preference for a turning direction (i.e. a high Delta modulation index) to  
297 be highly modulated by speed, suggesting that speed and direction locomotion information are relayed  
298 separately. A similar result was observed in macaque monkeys performing a visually guided tracking  
299 task (Roitman, 2005): Purkinje cells were found in that study to respond to position and direction of  
300 arm movement but not to arm speed.

301 According to our information theoretic analysis, the majority of the units provided maximum  
302 information about the speed of locomotion a short time in the future ( $\sim 100$  ms). They can thus be  
303 thought of as providing predictive, rather than retrospective, information about locomotion, suggesting  
304 that they may be driven by internally generated rather than sensory signals. The cerebellum receives  
305 projections from the nucleus cuneiformis (Gioia and Bianchi, 1987) that, in turn, receives inputs from  
306 the mesencephalic locomotor region (MLR; Ryczko and Dubuc, 2013). Since the MLR is a region of  
307 the hind brain that is involved in initiating and modulating locomotion (Shik et al., 1966; Lee et al.,  
308 2014; Kiehn, 2016), the cerebellum might receive a copy of the motor signals sent to spinal locomotion  
309 centres (Orlovsky et al., 1999). Indeed, MLR neurons have been observed to show similar speed  
310 tuning profiles to those reported here (Lee et al., 2014). We see this in agreement with computational  
311 theories, based on forward internal models, according to which the cerebellum uses an efference copy  
312 to compensate for slow sensory feedback during fast movements (Wolpert et al., 1995; Pasalar et al.,  
313 2006). In addition, the cerebellum might use the efference copy to suppress sensory feedback in order  
314 to reduce motor noise during movements (Shergill et al., 2003; Kennedy et al., 2014; Laurens et al.,  
315 2013).

316 We were able to reconstruct locomotion speed to high accuracy by linearly summing (positive and  
317 negative) weighted firing rates. The performance of our decoder increased with population size, as it  
318 can be expected, suggesting that multiple motor commands and copies of the central pattern generator  
319 signals can be effectively combined to minimise noise in the output to the deep cerebellar nuclei (Eccles,

320 1973; Schultz, Kitamura, et al., 2009), while preserving the remarkable pattern recognition capacity  
321 of the individual Purkinje cell (Marr, 1969; Albus, 1971; Barlow, 2002). This strategy could provide a  
322 more accurate estimate of the real speed of the animal, and, in turn, optimise motor control, similarly  
323 to what has been described for population decoding of saccades duration in monkeys (Thier et al.,  
324 2000). It was striking, however, how few cells needed to be combined in order to obtain an accurate  
325 readout of locomotion speed, for well-chosen cells. This interpretation also agrees with the idea that a  
326 linear summation of different response contributions could underpin the cerebellar neural code (Walter  
327 and Khodakhah, 2009).

328 In this study we recorded from neurons in cerebellar vermis lobules V and VI whose activity con-  
329 veyed information about locomotion kinematics. Other areas (such as paravermal lobule V, Sauerbrei  
330 et al., 2015) have also been found to represent locomotion signals. Because of the peculiar fractured  
331 somatotopy and modular organization of the circuitry (see e.g. Apps and Hawkes, 2009), it is unclear  
332 how kinematic information processing is divided across zones and how their outputs are integrated  
333 by deep cerebellar nuclei to generate motor control. Perhaps such motor estimations are also relayed  
334 to, and exploited by, other nervous centres, to integrate behavioural information relevant for motor  
335 control and spatial navigation. In future work we hope that this may be elucidated.

## 336 **Materials and Methods**

### 337 **Virtual reality system**

338 Experiments were performed in a custom made virtual reality system for mice similar to the ones  
339 used in previous studies (Harvey et al., 2009; Schmidt-Hieber and Häusser, 2013). Mice ran on a  
340 polystyrene sphere of 20 cm in diameter free floating on a 3D-printed concave inset. The motion of  
341 the sphere was read by two USB laser mice (Razer Emperor, Razer Inc, USA) positioned ninety  
342 degrees apart on the equator of the sphere. The signals carrying the instantaneous velocities of the  
343 sphere were polled at 200Hz by the host computer (Windows 7 OS, Microsoft Corporation, USA) via  
344 a Labview custom software (National Instruments Corporation, USA). These were then integrated to  
345 update the position in the virtual environment. The virtual reality environment was rendered with  
346 the OpenGL API implemented in the C++ language and interfaced with Labview control software  
347 via Microsoft dynamic-link libraries (DLL). This was projected via a digital projector (PJD6553w  
348 ViewSonic Corporation, USA) onto a demispherical screen around the mouse (Talbot design Ltd,  
349 UK) with a refresh rate of 120Hz. The VR apparatus comprised an automated reward system for  
350 water delivery and air puff stimulation controlled via a data acquisition card (NI BNC-2090A and NI

351 PCIe-6321, National Instruments Corporation). Water rewards were delivered to the mouth of the  
352 mouse via a custom made water spout connected to a peristaltic pump (Model 80204-0.5, Campden  
353 Instruments). Low pressure air jets were puffed to the mouse trunk from two lateral copper tubes  
354 upon opening of two normally closed solenoid valves (model PU220AR-01, Shako Co. Ltd) connected  
355 to an air pressure regulator.

356 Some experiments were run in the dark by disabling the projection of the visual stimulus and only  
357 recording mouse movements on the sphere.

## 358 **Surgical procedures**

359 All experiments were performed in accordance with the regulations of the United Kingdom Home  
360 Office. Of the fourteen animals used, ten of these animals received behavioural training before the  
361 electrophysiological recording. For the remaining four, the head stage implant and recording prepara-  
362 tion was performed in a single surgery (as these animals did not receive any training they are referred  
363 to as 'naïve' in this paper). All mice were implanted with a metal plate for head fixation attached  
364 on the anterior cranial bone with histoacryl tissue adhesive (Williams Medical Supplies, product code  
365 D569).

366 Trained mice underwent a second surgery upon completion of the behavioural training. Water  
367 restriction was terminated at least 24 hours before the surgery to make sure the animals could undergo  
368 the surgery fully hydrated. A craniotomy and durotomy were made over the central part of the vermis.  
369 The craniotomy was covered with a 1.2% agarose in with phosphate-buffered saline (PBS) and then  
370 capped with Kwik-Cast sealant and nail varnish to preserve the brain tissue until the recording session.  
371 This happened at least 24 hours after the end of the surgery.

## 372 **Behavioural training**

373 Following a recovery period of at least 24 hours from the surgery, the mice were head-fixed on the  
374 spherical treadmill for up to 10 minutes to habituate to the system for two consecutive days. Water  
375 restriction began on day 2 post-surgery. From day 3, the virtual reality projection was switched on.  
376 This is the first session of behavioural training. To incentivize the mice participation to the task, a  
377 water reward was given through a lick port as the mouse walked underneath cylinders suspended over  
378 the corridor. The water reward was given only at the end of each trial, once the mouse reached the  
379 following two criteria: 1) when the mouse was observed to intentionally stop only to lick its water  
380 reward and 2) when the mouse could reach the end of the virtual corridor without running more than  
381 twice the length of the corridor throughout the trials. Corridor length was progressively increased up

382 to 500 cm depending on the mouse performance and motivation. Two lateral air-puffers were used to  
383 prevent the mouse from hitting the virtual walls of the corridor. They pointed to the rear part of the  
384 trunk of the mouse on either side.

385 One to two hours after the end of a training session we gave the mice water *ad libitum* until they  
386 stopped drinking. Mouse weight was monitored daily as to guarantee that the each animal did not  
387 lose more than 20% of its pre-training body weight.

388 Mice were trained every day for at least 2 weeks and until they were capable of running 20  
389 consecutive trials in under 30 minutes from the start of the session for two consecutive days.

## 390 **Electrophysiological recordings**

391 After 24 hr recovery from the second surgery for trained mice (or from the first surgery for naïve  
392 mice), the mice were head-fixed on the spherical treadmill, the craniotomy was exposed after gently  
393 removing all layers of nail varnish, Kwik-Cast and agarose. The electrode was inserted at a 45 degree  
394 angle along the coronal plane and allowed to stabilize in the cerebellum for approximately 10 minutes  
395 once good spike signals were detected. To record activity from lobules V and VI of the cerebellum, we  
396 used 4-shank, 32-channel multielectrode arrays (MEAs) (Neuronexus Technologies, USA, probe model  
397 Buzsaki32). Behaviour and electrophysiological activity were then recorded in parallel while the mice  
398 navigated in the virtual reality environment.

399 To maximise mechanical and electrical stability during the electrophysiological recording, the water  
400 spout was removed from the mouth of the mouse and airpuffers were diverted from the mouse trunk.  
401 We did not observe behavioural changes, as the mice were fully hydrated beforehand, and the airpuff  
402 noise was a stimulus strong enough to elicit trajectory corrections. Multiple recordings were acquired  
403 from each mouse (minimum duration 270 seconds, max duration 1833 seconds). The entire recording  
404 sessions lasted up to 50 minutes.

## 405 **Data analysis**

### 406 **Spike sorting and clustering**

407 Electrophysiological data from each shank were processed independently with the *SpikeDetekt / Klus-*  
408 *taKwik / Klustaviewa* software suite (Rossant et al., 2016). After the spike times were detected and  
409 sorted with the automated program, we ‘cured’ the outcome of the spike sorting with the built-in  
410 *KlustaViewa* program. At this stage, we selected the units with the highest clustering quality. We  
411 kept units that had a central portion of the auto-correlogram completely clean (Harris et al., 2000).  
412 We merged units that were separated due to shift of the signal in different channels with the help of

413 the clustering features viewing tool. These were also validated against each other by means of the  
414 cross-correlograms. No distinction between simple and complex spikes were considered as the time  
415 window within which each spike waveform was extracted was only 2 ms.

#### 416 **Cell classification**

417 We used a hybrid cell classifier based on recently published algorithms (Van Dijk et al., 2013; Hens-  
418 broek et al., 2014). The algorithm was applied only to those units from which it was possible to collect  
419 at least 60 inter-spike intervals (ISI) taken during periods of stillness (speed of mouse  $\leq 1$  cm/s). The  
420 first steps aimed to identify putative Purkinje and granule cells based on their mean spike frequency,  
421 entropy of ISI distribution and logarithmic coefficient of variation of ISIs (CVlog), (Figure 1 - figure  
422 supplement 3); the remaining units were considered to be putative interneurons.

#### 423 **Tuning curves**

424 Firing rate was calculated at the same frequency as the speed (5 ms bins) and then smoothed with a  
425 150 ms Gaussian filter. Data points for speeds greater than 1 cm/s of the tuning curves are comprised  
426 by 2000 bins. The data points for speed=0 cm/s are formed by all bins taken when the mouse is  
427 still. To evaluate the significance of a unit's firing rate modulation with speed of locomotion, we  
428 randomly shifted the spike times one hundred times by at least 20 seconds and up to the duration  
429 of the recording minus 20 seconds (Kropff et al., 2015). For each iteration, firing rate was calculated  
430 and a speed tuning curve computed, and its variance was measured. We then compared the variance  
431 of the original speed tuning curve with the ones from the shuffled data. If its value was greater than  
432 99% of the shuffled data values, then we considered the unit as significantly sensitive to movement  
433 (binary response). We repeated this calculation and applied the same criteria to speeds  $\geq 1$  cm/s to  
434 assess if each unit was significantly modulated by locomotion (Saleem et al., 2013).

435 A unit response type was defined according to the curve that best fits the original data points.  
436 Because of the different response profiles obtained from the original data, three different curves were  
437 fitted (linear, second-degree polynomial and double exponential). The inverse of the variance of each  
438 data point was used as weight for the fitting to compensate for the different number of data points in  
439 each bin at speed=0. The coefficients of the best fit curve were used to determine the response type.  
440 In addition, we classified a cell as:

- 441 • positively modulated if the maximum firing rate was greater than the firing rate during stationary  
442 periods, and this was recorded at a speed greater than 70% of the maximum speed of the mouse;
- 443 • negatively modulated if the minimum firing rate was smaller than the firing rate during stationary  
444 periods, and it was recorded at a speed greater than 70% of maximum speed of the mouse

- having a preferred speed if the maximum firing was greater than the firing rate during stationary periods and this was recorded at a speed smaller than 70% of the maximum speed of the mouse.

The tuning curves for yaw movement were calculated similarly for clockwise (CW) and counter-clockwise (CCW) turning of the sphere. We then fitted three different curves (linear, second-degree polynomial and double exponential), selected the best fitting, and calculated the modulation index for either yaw direction. Modulation indexes were calculated as:

$$\frac{yaw_{max} - yaw_{min}}{yaw_{max} + yaw_{min}}.$$

We also calculated the difference in Modulation Index (Delta Modulation Index: Modulation Index CW - Modulation Index CCW) between the CW and CCW direction to assess the 'asymmetry' of tuning curves. Cells with a Delta larger than 0.2 were apparently asymmetric on visual inspection.

#### Naïve vs. trained

In order to verify whether the virtual reality affected the responses to movement kinematics, we compared the population of units acquired from naïve (n of units = 58) or trained (n of units = 253) animals. We used the Mann-Whitney Test to compare the modulation index distributions in the two conditions for speed and yaw. We also compared the difference in yaw modulation index and the phase index.

#### Step cycle modulation

To look at the modulation with stepping cycle, the pitch velocity signal was high-pass filtered at 3 Hz to cancel the locomotor related changes of speed. The Hilbert transform was then computed and its phase was extracted as a function of time. To ensure that pitch velocity changes were due to stepping, only putative stepping cycles longer than 50 ms and occurring only during moving periods (speed  $\geq 1$  cm/s) were considered. Each cycle duration was normalised with respect to time and divided in 36 equal intervals. For each interval, the instantaneous firing rate was computed.

Because of the binning of each cycle, step phase modulation was tested for uniformity with the  $\chi^2$  test of uniformity (Fisher, 1995). The mean direction  $\theta$  (in radians) of the firing rate distribution of a cell around the step cycle was computed as:

$$\theta = \arctan \left( \frac{\sum_{i=1}^n \frac{\sin \alpha}{n}}{\sum_{i=1}^n \frac{\cos \alpha}{n}} \right) \quad (1)$$

where the numerator and denominator are the mean rectangular coordinates of the resulting phase angle,  $X$  and  $Y$  respectively,  $\alpha$  is the phase angle of the resultant vector  $R = \sqrt{X^2 + Y^2}$  for each cycle, and  $n$  is the number of cycles or steps.

473 We also calculated the circular standard deviation  $\sigma$ , which is a measure of the spread of the firing  
474 rate around the mean phase direction, and indicates where approximately 66% of the data lie (Drew  
475 and Doucet, 1991), as  $\sigma = \sqrt{-2 \ln R}$ . We calculated the Phase Selectivity Index (PSI). PSI is defined  
476 equivalently to the orientation selectivity index described by Mazurek et al. (2014),

$$PSI = 1 - \frac{\sum_{i=1}^n R(\theta_n)^{2i\theta_n}}{\sum_{i=1}^n R(\theta_n)}, \quad (2)$$

477 where  $R(\theta)$  is the magnitude of the firing rate for any given angle  $\theta = [0^\circ : 10^\circ : 360^\circ]$ , for each  
478 stepping cycle.

## 479 Mutual information

480 The Mutual Information between instantaneous firing rate and kinematic time-courses was computed  
481 using a continuous estimator based on the Kraskov, Stögbauer, and Grassberger (GSK) technique  
482 (Kraskov et al., 2004). We used the Matlab implementation of the GSK algorithm in the JIDT  
483 toolkit (Lizier, 2014). Firing rates and kinematics variables were computed as described above, and  
484 fed into the GSK algorithm, returning a mutual information value for each unit. For the step cycle  
485 calculation, the mutual information was computed between the firing rate and the phase angles of the  
486 Hilbert transform of the pitch velocity. Only periods during movement were considered, and mutual  
487 information was estimated for each cycle and then averaged. In this case, firing rate was computed  
488 every 5 ms and smoothed with a Gaussian filter of standard deviation 20 ms.

## 489 Decoding

490 For every chosen experiment, the recorded cells' spike trains were binned at 5 ms and then convolved  
491 with a Gaussian function ( $\sigma = 50$ , window width of  $3\sigma$ ) to obtain a time-course vector of instantaneous  
492 firing rates. The locomotion speed time course was convolved with the same Gaussian function. We  
493 considered all locomotion speeds  $\leq 1$  cm/s to be stationary; these were set to 0 cm/s. Both the firing  
494 rate and the locomotion speed time-courses were then normalised to obtain values between 0 and 1.

495 Only recording sessions with at least 8 units (resulting in inclusion of 6 sessions from 4 mice) were  
496 considered, in order to investigate the scaling of decoder performance with ensemble size. To decode,  
497 we used an optimal linear estimator (OLE) which weighted and linearly summed the instantaneous fir-  
498 ing rate of each neuron in its ensemble, then rectified the summed output. We tested the incorporation  
499 of an additional offset term prior to rectification, but found that it did not improve performance on  
500 our dataset. The decoder was trained on 70% of each locomotion session, and tested on the remaining

501 30%. The OLE reconstruction is given by

$$\hat{\mathbf{v}} = [\hat{\mathbf{R}}\mathbf{w}]^+ \quad (3)$$

502 where  $\hat{\mathbf{v}} = [v_1 \dots v_{T_{\text{test}}}]^T$  is the reconstructed locomotion speed time course,  $[\cdot]^+$  denotes the rectification  
503 operator,  $\hat{\mathbf{R}}$  is a matrix whose columns consist of the firing rates  $\mathbf{r}_i$  of each cell  $i$  for the  $T_{\text{test}}$  test time  
504 bins, and  $\mathbf{w}$  is the linear estimator given by

$$\mathbf{v} = \mathbf{R}\mathbf{w}, \quad (4)$$

505 with  $\mathbf{v} = [v_1 \dots v_{T_{\text{train}}}]^T$  being the measured speed time course vector for the  $T_{\text{train}}$  training data bins,  
506 and  $\mathbf{R}$  a matrix whose columns are the firing rates for the training data, with the addition of a column  
507 of ones for the  $y$  intercept. Training the decoder by linear least squares regression is equivalent to  
508 solving this equation to find the optimal value of the estimator:

$$\mathbf{w} = (\mathbf{R}^T\mathbf{R})^{-1}\mathbf{R}^T\mathbf{v}, \quad (5)$$

509 where  $\mathbf{v}$  is a column vector containing the locomotion speed values for the training data. The esti-  
510 mated speed is half wave rectified to reflect the fact that only positive speed values are possible. We  
511 assessed decoding performance by computing the Pearson correlation coefficient between the actual  
512 and reconstructed locomotion speed time-courses, for the test data.

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## 519 References

- 520 Akay, T., Tourtellotte, W. G., Arber, S., and Jessell, T. M. (2014), ‘Degradation of mouse locomotor  
521 pattern in the absence of proprioceptive sensory feedback.’ *Proceedings of the National Academy  
522 of Sciences of the United States of America*, **111**(47), 16877–82, DOI: 10.1073/pnas.1419045111.
- 523 Albus, J. S. (1971), ‘A theory of cerebellar function’, *Mathematical Biosciences*, **10**, 25–61.
- 524 Apps, R. and Hawkes, R. (2009), ‘Cerebellar cortical organization: a one-map hypothesis’, *Nature  
525 Reviews Neuroscience*, **10**(9), 670–681, DOI: 10.1038/nrn2698.
- 526 Armstrong, D. M. and Edgley, S. A. (1984), ‘Discharges of Purkinje cells in the paravermal part of  
527 the cerebellar anterior lobe during locomotion in the cat.’ *Journal of Physiology*, **352**(1), 403–424,  
528 DOI: 10.1113/jphysiol.1984.sp015300.
- 529 Armstrong, D. M. and Edgley, S. A. (1988), ‘Discharges of interpositus and Purkinje cells of the cat  
530 cerebellum during locomotion under different conditions’, *Journal of Physiology*, **400**, 425–445.
- 531 Arshavsky, Y. I., Gelfand, I., and Orlovsky, G. (1983), ‘The cerebellum and control of rhythmical  
532 movements’, *Trends in Neurosciences*, **6**, 417–422, DOI: 10.1016/0166-2236(83)90191-1.
- 533 Ashe, J. and Georgopoulos, A. P. (1994), ‘Movement parameters and neural activity in motor cortex  
534 and area 5’, *Cerebral Cortex*, **4**(6), 590–600, DOI: 10.1093/cercor/4.6.590.
- 535 Astorga, G., Bao, J., Marty, A., Augustine, G. J., Franconville, R., Jalil, A., Bradley, J., and Llano, I.  
536 (2015), ‘An excitatory GABA loop operating in vivo’, *Frontiers in Cellular Neuroscience*, **9**, 275,  
537 DOI: 10.3389/fncel.2015.00275.
- 538 Barlow, J. S. (2002), *The Cerebellum and adaptive control*, Cambridge University Press.
- 539 Bellardita, C. and Kiehn, O. (2015), ‘Phenotypic characterization of speed-associated gait changes in  
540 mice reveals modular organization of locomotor networks.’ *Current Biology*, **25**(11), 1426–36, DOI:  
541 10.1016/j.cub.2015.04.005.
- 542 Berretta, S., Perciavalle, V., and Poppele, R. E. (1991), ‘Origin of spinal projections to the anterior  
543 and posterior lobes of the rat cerebellum’, *The Journal of Comparative Neurology*, **305**(2), 273–  
544 281, DOI: 10.1002/cne.903050208.
- 545 Chambers, W. W. and Sprague, J. M. (1955), ‘Functional localization in the cerebellum. I. Orga-  
546 nization in longitudinal cortico-nuclear zones and their contribution to the control of posture,  
547 both extrapyramidal and pyramidal’, *Journal of Comparative Neurology*, **103**(1), 105–129, DOI:  
548 10.1002/cne.901030107.
- 549 Coddington, L. T., Rudolph, S., Vande Lune, P., Overstreet-Wadiche, L., and Wadiche, J. I. (2013),  
550 ‘Spillover-mediated feedforward inhibition functionally segregates interneuron activity.’ *Neuron*,  
551 **78**(6), 1050–62, DOI: 10.1016/j.neuron.2013.04.019.

- 552 Drew, T. and Doucet, S. (1991), ‘Application of circular statistics to the study of neuronal discharge  
553 during locomotion’, *Journal of Neuroscience Methods*, **38**(2-3), 171–181, DOI: 10.1016/0165-  
554 0270(91)90167-X.
- 555 Eccles, J. C. (1973), ‘The cerebellum as a computer: patterns in space and time.’ *Journal of Physiology*,  
556 **229**(1), 1–32.
- 557 Edgley, S. A. and Lidieth, M. (1987), ‘The discharges of cerebellar Golgi cells during locomotion in  
558 the cat’, *The Journal of Physiology*, **392**, 315–332.
- 559 Fedirchuk, B., Stecina, K., Kristensen, K. K., Zhang, M., Meehan, C. F., Bennett, D. J., and Hultborn,  
560 H. (2013), ‘Rhythmic activity of feline dorsal and ventral spinocerebellar tract neurons during  
561 fictive motor actions’, *Journal of Neurophysiology*, **109**(2), 375–388.
- 562 Fisher, N. I. (1995), *Statistical analysis of circular data*, Cambridge Press.
- 563 Flourens, P. (1824), *Recherches experimentales sur les proprietes et les fonctions du systeme nerveux*  
564 *dans les animaux vertebres*, Paris : J.-B. Balliere.
- 565 Galliano, E. and De Zeeuw, C. I. (2014), ‘Questioning the cerebellar doctrine’, in: *Progress in brain*  
566 *research*, ed. by N. Ramnani, vol. 210, Elsevier, chap. 3, pp. 59–77.
- 567 Gioia, M. and Bianchi, R. (1987), ‘The cytoarchitecture of the nucleus cuneiformis. A Nissl and Golgi  
568 study.’ *Journal of Anatomy*, **155**, 165–76.
- 569 Goulding, M. (2009), ‘Circuits controlling vertebrate locomotion: moving in a new direction’, *Nature*  
570 *Reviews Neuroscience*, **10**(7), 507–518, DOI: 10.1038/nrn2608.
- 571 Harris, K. D., Henze, D. A., Csicsvari, J., Hirase, H., and Buzsaki, G. (2000), ‘Accuracy of tetrode spike  
572 separation as determined by simultaneous intracellular and extracellular measurements’, *Journal*  
573 *of Neurophysiology*, **84**(1), 401–414.
- 574 Harvey, C. D., Collman, F., Dombeck, D. A., and Tank, D. W. (2009), ‘Intracellular dynamics of  
575 hippocampal place cells during virtual navigation’, *Nature*, **461**(7266), 941–946, DOI: 10.1038/  
576 nature08499.
- 577 Hensbroek, R. A., Belton, T., Beugen, R. A. van, Maruta, J., Ruigrok, T. J., and Simpson, J. I.  
578 (2014), ‘Identifying Purkinje cells using only their spontaneous simple spike activity’, *Journal of*  
579 *Neuroscience Methods*, **232**, 173–180, DOI: 10.1016/j.jneumeth.2014.04.031.
- 580 Hewitt, A. L., Popa, L. S., Pasalar, S., Hendrix, C. M., and Ebner, T. J. (2011), ‘Representation of  
581 limb kinematics in Purkinje cell simple spike discharge is conserved across multiple tasks.’ *Journal*  
582 *of Neurophysiology*, **106**(5), 2232–2247, DOI: 10.1152/jn.00886.2010.
- 583 Holmes, G. (1939), ‘The cerebellum of man’, *Brain*, **62**, 1–30.

- 584 Holscher, C., Schnee, A., Dahmen, H., Setia, L., and Mallot, H. A. (2005), ‘Rats are able to navigate  
585 in virtual environments’, *Journal of Experimental Biology*, **208**(3), 561–569, DOI: 10.1242/jeb.  
586 01371.
- 587 Hoogland, T. M., De Griijl, J. R., Witter, L., Canto, C. B., and De Zeeuw, C. I. (2015), ‘Role  
588 of synchronous activation of cerebellar Purkinje cell ensembles in multi-joint movement control.’  
589 *Current Biology*, **25**(9), 1157–1165, DOI: 10.1016/j.cub.2015.03.009.
- 590 Ichise, T., Kano, M., Hashimoto, K., Yanagihara, D., Nakao, K., Shigemoto, R., Katsuki, M., and  
591 Aiba, A. (2000), ‘mGluR1 in cerebellar Purkinje cells essential for long-term depression, synapse  
592 elimination, and motor coordination’, *Science*, **288**(5472), 1832–1835, DOI: 10.1126/science.  
593 288.5472.1832.
- 594 Ito, M. (2006), ‘Cerebellar circuitry as a neuronal machine.’ *Progress in Neurobiology*, **78**(3-5), 272–  
595 303, DOI: 10.1016/j.pneurobio.2006.02.006.
- 596 Kennedy, A., Wayne, G., Kaifosh, P., Alviña, K., Abbott, L. F., and Sawtell, N. B. (2014), ‘A tem-  
597 poral basis for predicting the sensory consequences of motor commands in an electric fish.’ *Nature*  
598 *Neuroscience*, **17**(3), 416–422, DOI: 10.1038/nn.3650.
- 599 Kiehn, O. (2016), ‘Decoding the organization of spinal circuits that control locomotion’, *Nature Re-*  
600 *views Neuroscience*, **17**(4), 224–238, DOI: 10.1038/nrn.2016.9.
- 601 Kraskov, A., Stögbauer, H., and Grassberger, P. (2004), ‘Estimating mutual information.’ *Physical*  
602 *Review E*, **69**(6 Pt 2), 066138, DOI: 10.1103/PhysRevE.69.066138.
- 603 Kropff, E., Carmichael, J. E., Moser, M.-B., and Moser, E. I. (2015), ‘Speed cells in the medial  
604 entorhinal cortex’, *Nature*, **523**(7561), 419–424, DOI: 10.1038/nature14622.
- 605 Laurens, J., Meng, H., and Angelaki, D. E. (2013), ‘Computation of linear acceleration through an  
606 internal model in the macaque cerebellum.’ *Nature Neuroscience*, **16**(11), 1701–1708, DOI: 10.  
607 1038/nn.3530.
- 608 Lee, A. M., Hoy, J. L., Bonci, A., Wilbrecht, L., Stryker, M. P., and Niell, C. M. (2014), ‘Identification  
609 of a brainstem circuit regulating visual cortical state in parallel with locomotion.’ *Neuron*, **83**(2),  
610 455–66, DOI: 10.1016/j.neuron.2014.06.031.
- 611 Lizier, J. T. (2014), ‘JIDT: An Information-Theoretic Toolkit for Studying the Dynamics of Complex  
612 Systems’, *Frontiers in Robotics and AI*, **1**, DOI: 10.3389/frobt.2014.00011.
- 613 Luciani, L. (1891), *Il cervelletto: nuovi studi di fisiologia normale e patologica*, Le Monnier.
- 614 Marr, D. (1969), ‘A theory of cerebellar cortex’, *Journal of Physiology*, **202**(2), 437–470.

- 615 Mathews, P. J., Lee, K. H., Peng, Z., Houser, C. R., and Otis, T. S. (2012), ‘Effects of climbing fiber  
616 driven inhibition on Purkinje neuron spiking’, *Journal of Neuroscience*, **32**(50), 17988–17997, DOI:  
617 10.1523/JNEUROSCI.3916-12.2012.
- 618 Mazurek, M., Kager, M., and Van Hooser, S. D. (2014), ‘Robust quantification of orientation selectivity  
619 and direction selectivity.’ *Frontiers in Neural Circuits*, **8**, 92, DOI: 10.3389/fncir.2014.00092.
- 620 Napper, R. M. and Harvey, R. J. (1988), ‘Number of parallel fiber synapses on an individual Purkinje  
621 cell in the cerebellum of the rat.’ *Journal of Comparative Neurology*, **274**, 168–177.
- 622 Orlovsky, G. N. (1972), ‘Work of the Purkinje cells during locomotion’, *Biophysics*, **17**, 935–941.
- 623 Orlovsky, G., Deliagina, T., and Grillner, S. (1999), *Neural Control of Locomotion, from mollusc to*  
624 *man*, Oxford University Press.
- 625 Oscarsson, O. (1965), ‘Functional organization of the spino- and cuneocerebellar tracts.’ *Physiological*  
626 *Reviews*, **45**(3), 495–522.
- 627 Oscarsson, O. (1979), ‘Functional units of the cerebellum- sagittal zones and microzones’, *Trends in*  
628 *Neuroscience*, **2**, 144–145.
- 629 Ozden, I., Dombeck, D. A., Hoogland, T. M., Tank, D. W., and Wang, S. S.-H. (2012), ‘Widespread  
630 state-dependent shifts in cerebellar activity in locomoting mice’, *PLoS ONE*, **7**(8), DOI: 10.1371/  
631 journal.pone.0042650.
- 632 Pasalar, S., Roitman, A. V., Durfee, W. K., and Ebner, T. J. (2006), ‘Force field effects on cerebellar  
633 Purkinje cell discharge with implications for internal models.’ *Nature Neuroscience*, **9**(11), 1404–  
634 11, DOI: 10.1038/nn1783.
- 635 Person, A. L. and Raman, I. M. (2012), ‘Purkinje neuron synchrony elicits time-locked spiking in the  
636 cerebellar nuclei.’ *Nature*, **481**(7382), 502–505, DOI: 10.1038/nature10732.
- 637 Popa, L. S., Hewitt, A. L., and Ebner, T. J. (2012), ‘Predictive and feedback performance errors are  
638 signaled in the simple spike discharge of individual purkinje cells’, *Journal of Neuroscience*, **32**(44),  
639 15345–15358, DOI: 10.1523/JNEUROSCI.2151-12.2012.
- 640 Powell, K., Mathy, A., Duguid, I., and Häusser, M. (2015), ‘Synaptic representation of locomotion in  
641 single cerebellar granule cells.’ *eLife*, **4**, e07290, DOI: 10.7554/eLife.07290.
- 642 Reeber, S. L., Otis, T. S., and Sillitoe, R. V. (2013), ‘New roles for the cerebellum in health and  
643 disease.’ *Frontiers in systems neuroscience*, **7**, 83, DOI: 10.3389/fnsys.2013.00083.
- 644 Rieubland, S., Roth, A., and Häusser, M. (2014), ‘Structured connectivity in cerebellar inhibitory  
645 networks’, *Neuron*, **81**(4), 913–929, DOI: 10.1016/j.neuron.2013.12.029.

- 646 Roitman, A. V. (2005), ‘Position, direction of movement, and speed tuning of cerebellar Purkinje cells  
647 during circular manual tracking in monkey’, *Journal of Neuroscience*, **25**(40), 9244–9257, DOI:  
648 10.1523/JNEUROSCI.1886-05.2005.
- 649 Rossant, C., Kadir, S. N., Goodman, D. F. M., Schulman, J., Belluscio, M., Buzsaki, G., and Harris,  
650 K. D. (2016), ‘Spike sorting for large, dense electrode arrays’, *Nature Neuroscience*, **19**(4), 634–641,  
651 DOI: 10.1101/015198.
- 652 Ryczko, D. and Dubuc, R. (2013), ‘The Multifunctional Mesencephalic Locomotor Region’, *Current*  
653 *Pharmaceutical Design*, **19**(24), 4448–4470, DOI: 10.2174/1381612811319240011.
- 654 Saleem, A. B., Ayaz, A., Jeffery, K. J., Harris, K. D., and Carandini, M. (2013), ‘Integration of visual  
655 motion and locomotion in mouse visual cortex.’ *Nature Neuroscience*, **16**(12), 1864–1869, DOI:  
656 10.1038/nn.3567.
- 657 Sauerbrei, B. A., Lubenov, E. V., and Siapas, A. G. (2015), ‘Structured variability in Purkinje cell  
658 activity during locomotion’, *Neuron*, **87**(4), 840–852, DOI: 10.1016/j.neuron.2015.08.003.
- 659 Schmidt-Hieber, C. and Häusser, M. (2013), ‘Cellular mechanisms of spatial navigation in the medial  
660 entorhinal cortex.’ *Nature neuroscience*, **16**(3), 325–31, DOI: 10.1038/nn.3340.
- 661 Schultz, S. R., Ince, R. A., and Panzeri, S. (2015), ‘Applications of Information Theory to Analysis of  
662 Neural Data’, in: *Encyclopedia of Computational Neuroscience*, Springer, pp. 199–203.
- 663 Schultz, S. R., Kitamura, K., Post-Uiterweer, A., Krupic, J., and Häusser, M. (2009), ‘Spatial pattern  
664 coding of sensory information by climbing fiber-evoked calcium signals in networks of neighboring  
665 cerebellar Purkinje cells’, *The Journal of Neuroscience*, **29**(25), 8005–8015.
- 666 Sengul, G., Fu, Y., Yu, Y., and Paxinos, G. (2015), ‘Spinal cord projections to the cerebellum in the  
667 mouse.’ *Brain Structure & Function*, **220**(5), 2997–3009, DOI: 10.1007/s00429-014-0840-7.
- 668 Shergill, S. S., Bays, P. M., Frith, C. D., and Wolpert, D. M. (2003), ‘Two eyes for an eye: the  
669 neuroscience of force escalation.’ *Science*, **301**(5630), 187, DOI: 10.1126/science.1085327.
- 670 Shik, M. L., Severin, F. V., and Orlovski, G. N. (1966), ‘Control of walking and running by means of  
671 electric stimulation of the midbrain.’ *Biofizika*, **11**(4), 659–66.
- 672 Thier, P., Dicke, P. W., Haas, R., and Barash, S. (2000), ‘Encoding of movement time by populations  
673 of cerebellar Purkinje cells’, *Nature*, **405**(6782), 72–76, DOI: 10.1038/35011062.
- 674 Van Dijck, G., Van Hulle, M. M., Heiney, S. A., Blazquez, P. M., Meng, H., Angelaki, D. E., Arenz,  
675 A., Margrie, T. W., Mostofi, A., Edgley, S., Bengtsson, F., Ekerot, C. F., Jorntell, H., Dalley,  
676 J. W., and Holtzman, T. (2013), ‘Probabilistic identification of cerebellar cortical neurones across  
677 species’, *PLoS ONE*, **8**(3), e57669, DOI: 10.1371/journal.pone.0057669.

- 678 Walter, J. T. and Khodakhah, K. (2009), ‘The advantages of linear information processing for cerebel-  
679 lar computation.’ *Proceedings of the National Academy of Sciences of the United States of America*,  
680 **106**(11), DOI: 10.1073/pnas.0812348106.
- 681 Wolpert, D. M., Ghahramani, Z., and Jordan, M. I. (1995), ‘An internal model for sensorimotor  
682 integration’, *Science*, **269**(5232), 1880–1882, DOI: 10.1126/science.7569931.
- 683 Yanagihara, D. and Kondo, I. (1996), ‘Nitric oxide plays a key role in adaptive control of locomotion  
684 in cat.’ *Proceedings of the National Academy of Sciences of the United States of America*, **93**(23),  
685 13292–13297.
- 686 Yu, B. M., Kemere, C., Santhanam, G., Afshar, A., Ryu, S. I., Meng, T. H., Sahani, M., and Shenoy,  
687 K. V. (2007), ‘Mixture of trajectory models for neural decoding of goal-directed movements.’  
688 *Journal of Neurophysiology*, **97**(5), 3763–3780, DOI: 10.1152/jn.00482.2006.
- 689 Zhou, H., Lin, Z., Voges, K., Ju, C., Gao, Z., Bosman, L. W., Ruigrok, T. J., Hoebeek, F. E., De  
690 Zeeuw, C. I., and Schonewille, M. (2014), ‘Cerebellar modules operate at different frequencies.’  
691 *eLife*, **3**, e02536, DOI: 10.7554/eLife.02536.

692 **Supplementary Figures**

693 After main figures

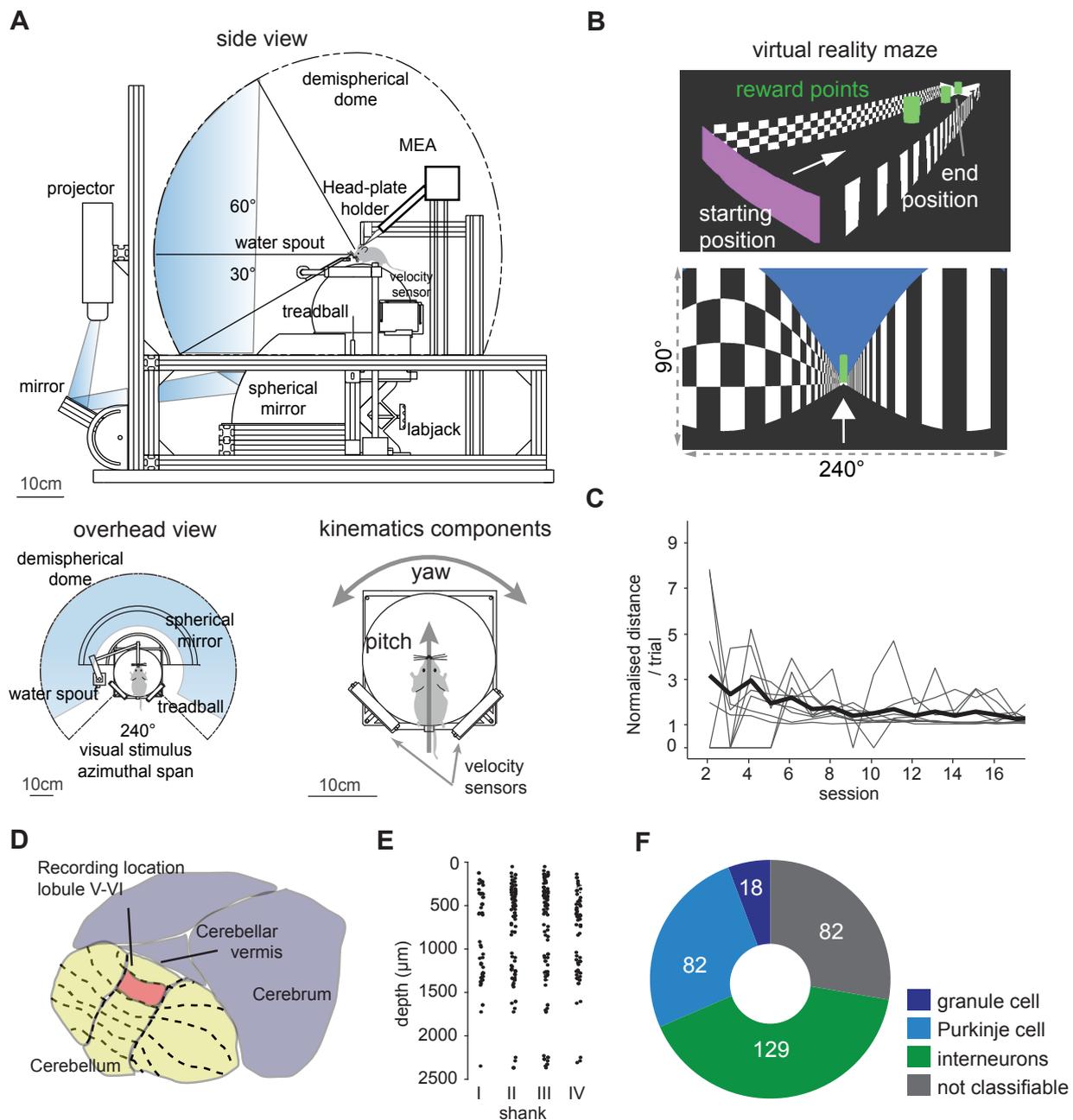


Figure 1: Virtual reality setup and electrophysiology recordings. (A) schematic view of the virtual reality system with approximate image path of the visual stimulus from the projector to the demispherical screen. Bottom left. Overhead view of the coverage of the mouse visual field. Bottom right. Mouse on the spherical treadmill, or treadball. The signals from the velocity sensors are integrated to determine translation (pitch) and heading (yaw) movements of the mouse in the virtual environment. Mouse drawings not to scale. (B) Virtual reality environment. Top, perspective view of the virtual corridor. Bottom, subject perspective of the VR maze with horizontal and vertical span of the virtual camera. (C) Mean normalised distance ran per trial in first 16 days. Gray lines are subjects, thick black line is average for all mice. (D) Recording location area. (E) Depth of single units ( $n=311$ ) from the cerebellar surface grouped by shank (I-IV) for all mice. Depth measurements are based on the position of the channel in which the greatest spike amplitude of the signal is recorded (F) Pie chart of cell classes as identified by a hybrid classification algorithm based on VanDijck et al., 2013 and Hensbroek et al. 2014

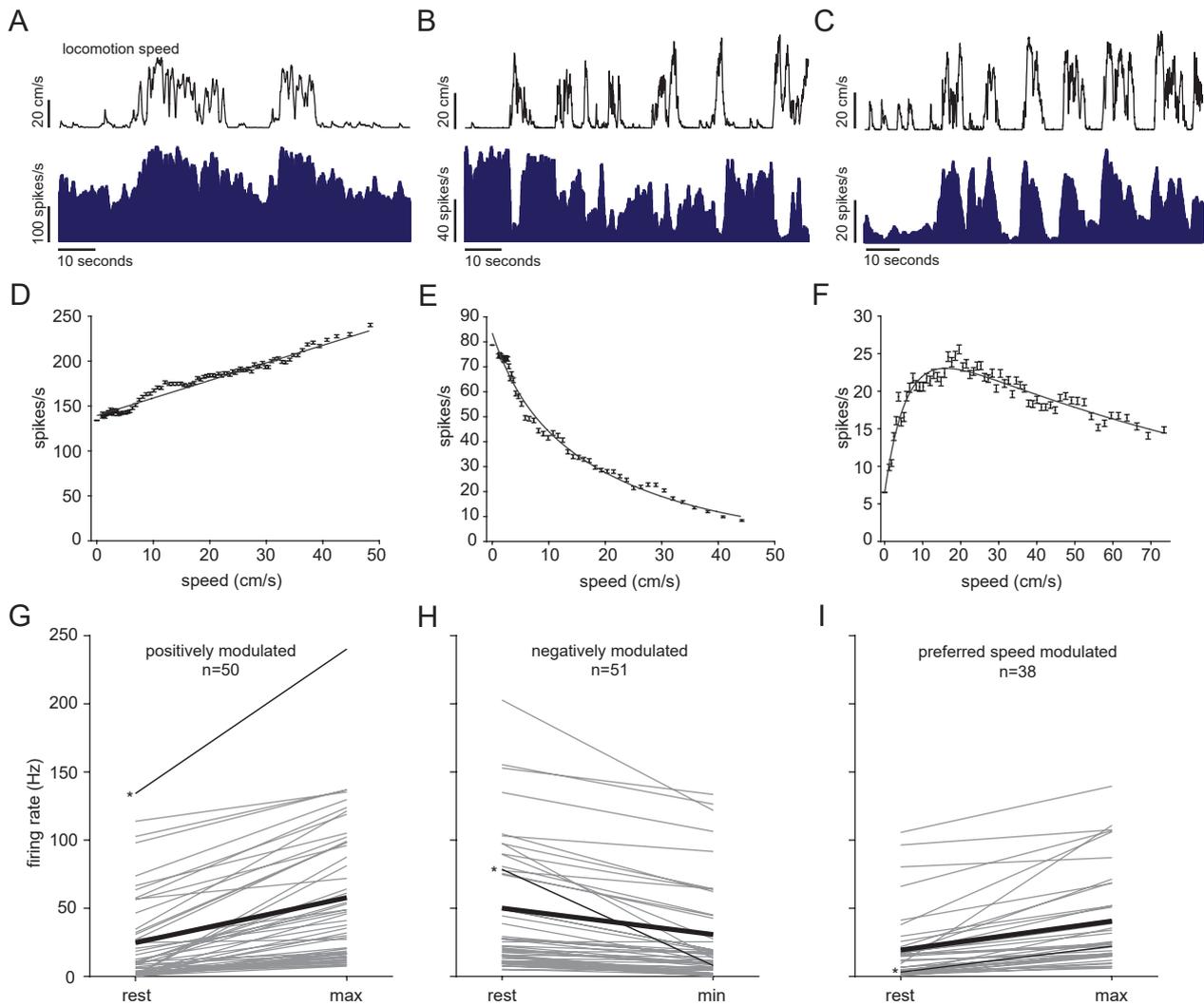


Figure 2: Cerebellar neurons response to locomotion speed. (A-C) Speed and instantaneous firing rate of three example units for the three response profiles observed (bins = 0.5 seconds). (D-F) Speed tuning curves of the examples in A-C; error bars are standard errors. (G-I) Mean firing rate during stationary periods and speeds at which maximal firing rate is recorded. Gray lines are single units; thin black lines with asterisks on the left indicate units shown in A-C and D-F; thick black lines are average firing rates of all units within each response class.

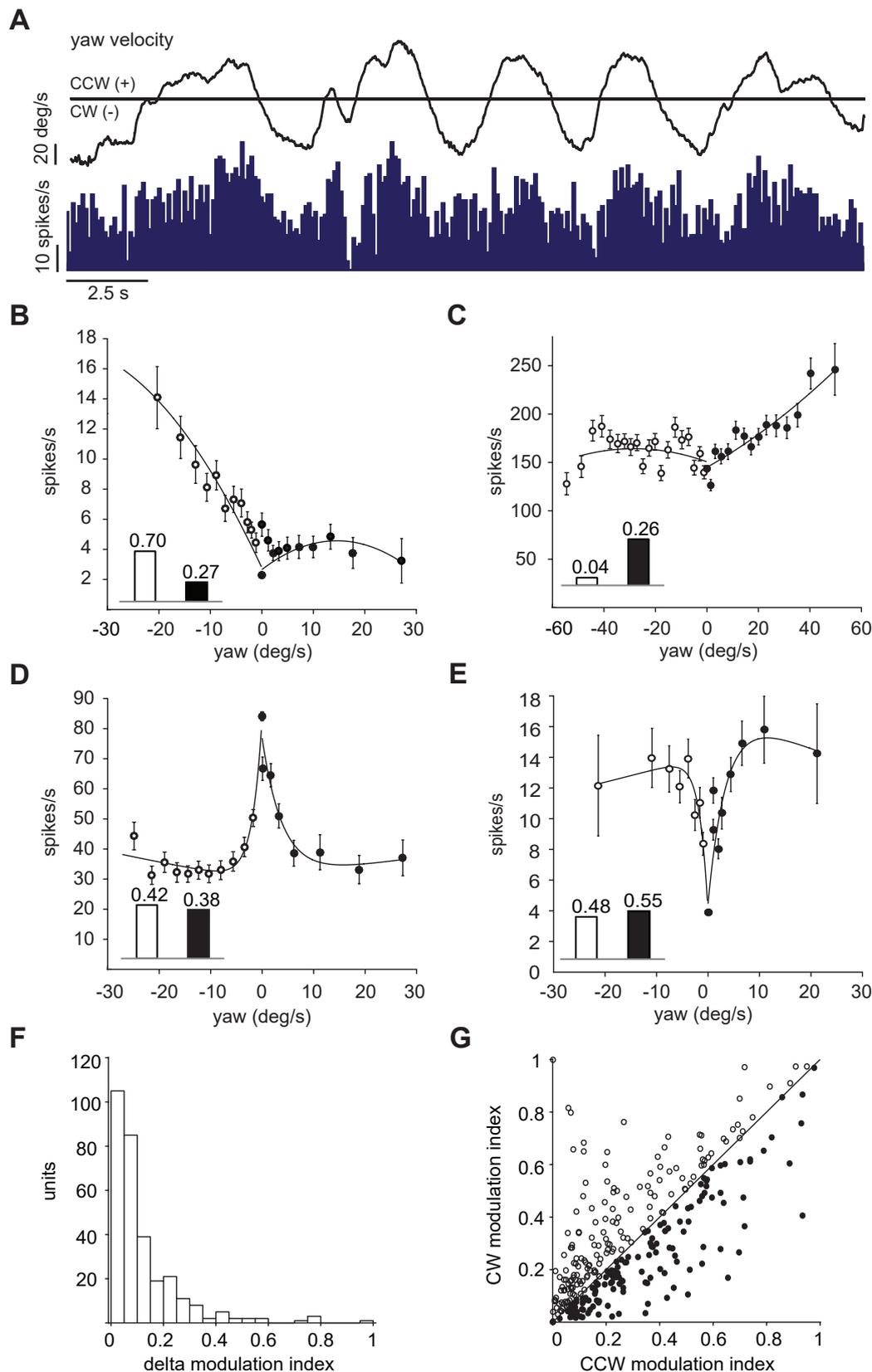


Figure 3: Cerebellar neurons response to yaw direction. (A) Example unit tuned to CCW yaw direction (positive): top, yaw trace in deg/s; bottom, mean instantaneous firing rate, bin width 100 ms. (B-C) Two example units preferentially responding to yaw-turning in the CW (B) or CCW (C) ; each data point is formed by more than 900 bins (100ms width); error bars are s.e.m.; inset bars indicate the modulation index values derived from the two curves. (D) Example unit decreasing its activity in both yaw directions. (E) Example unit that increases its activity in either yaw direction. (F) Distribution of the difference in modulation index between the CW and CCW yaw direction. (G) Population plot of modulation index values of each unit for the CW and CCW direction (n=311).

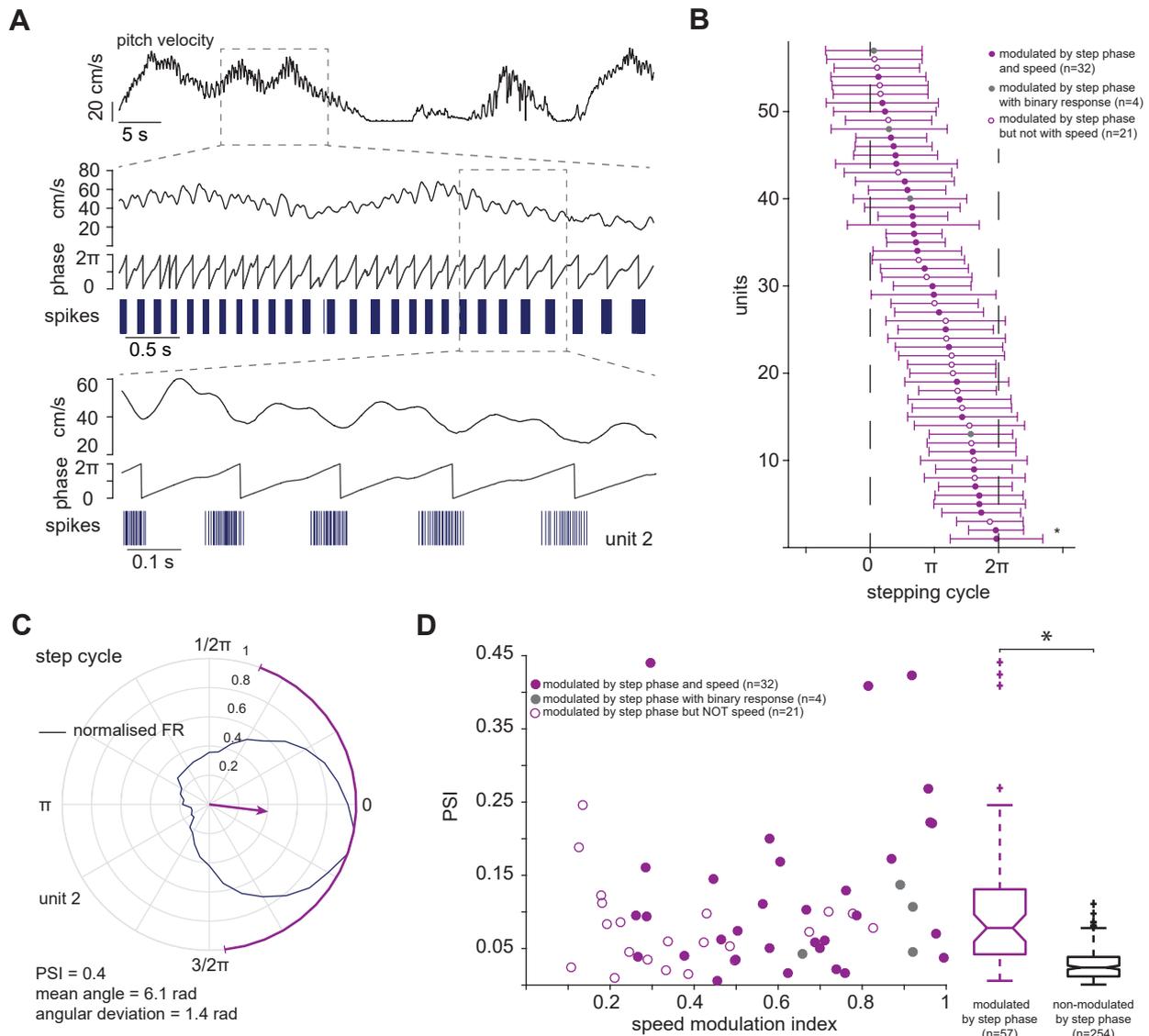


Figure 4: Cerebellar neurons response to stepping cycle. (A) Top: pitch velocity signal; centre: boxed pitch velocity signal aligned with its Hilbert transformed signal (middle) and spike events of an example unit; bottom: close-up of approximately one second. (B) Preferred phases of units significantly modulated by stepping cycle (n=57); horizontal bars indicate standard circular deviations. Unit 2, shown in A and C is marked by the asterisk. (C) Polar plot showing the preferred locomotion phase of normalised firing for the same unit shown in A; arrow indicates mean angle direction and phase selectivity index, PSI (magnitude of mean response); purple solid line around the circle is the circular standard deviation. (D) PSI of units modulated by step phase as a function of speed modulation index. Right, distribution of PSI of these units is significantly different from the distribution of the other units.

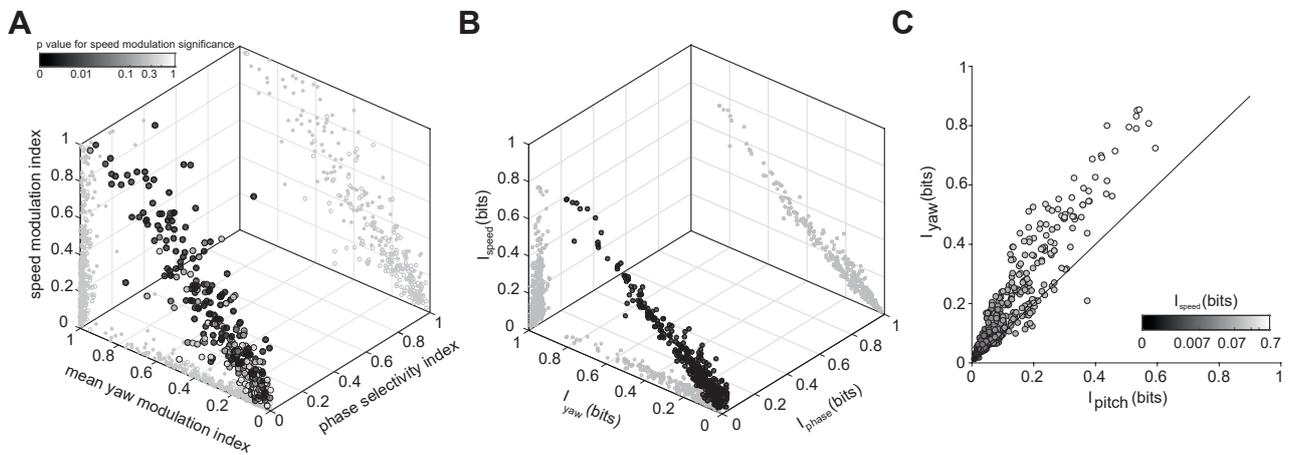


Figure 5: Modulation indexes and mutual information values of cerebellar units. **(A)** Modulation indexes for speed, yaw and phase selectivity index. Colour coding indicates p value of test for significance of speed modulation. Mean yaw modulation index is the mean between the modulation indexes of the yaw tuning curves for clockwise and counter-clockwise directions. **(B)** Mutual information values for speed, yaw and step cycle. **(C)** Mutual information values of the two vectorial components of speed, pitch and yaw. Colour coding indicates the value of mutual information of firing rate and speed.

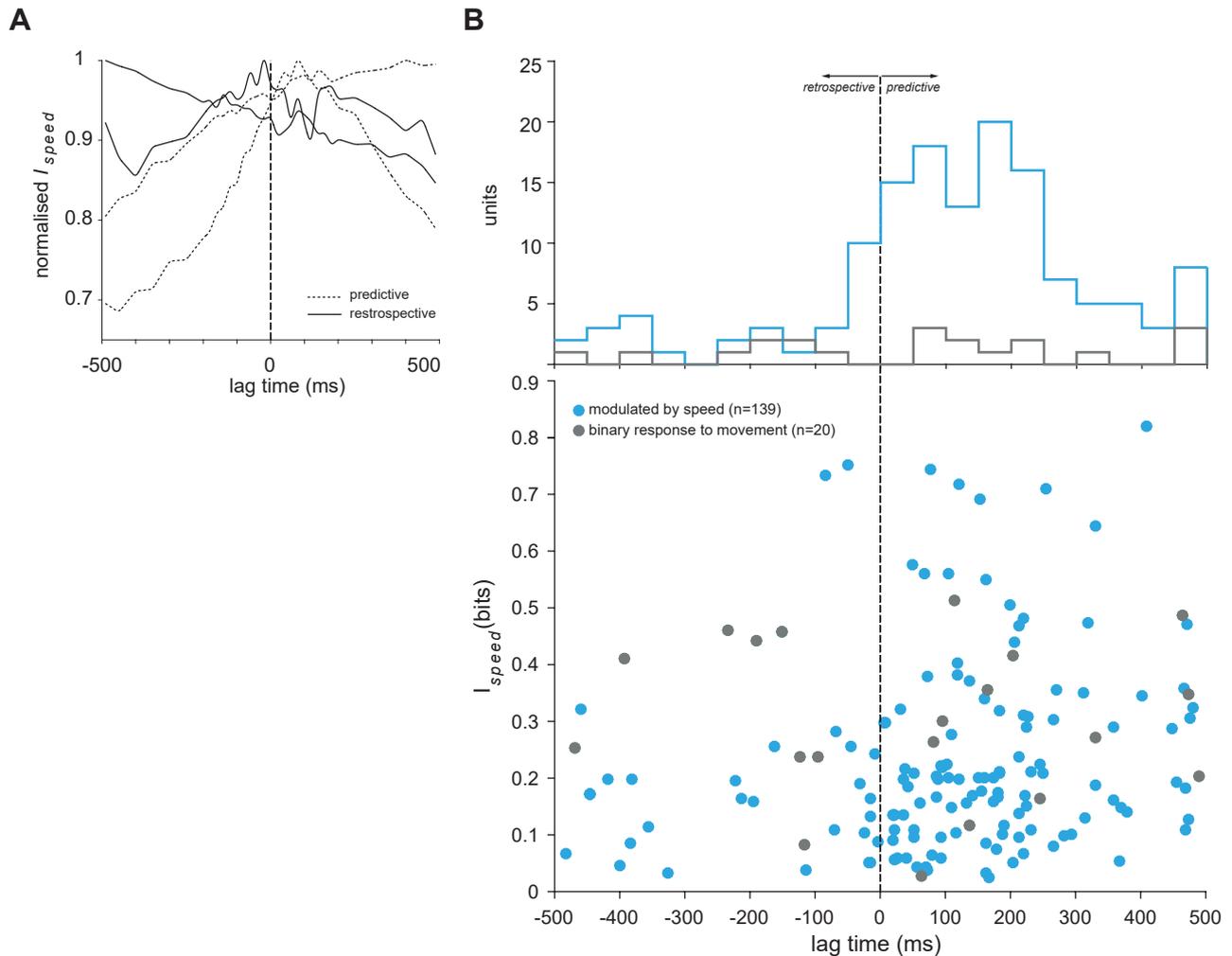


Figure 6: Mutual information of locomotion speed and firing rate. **(A)** Normalised mutual information of four example units for different time lags; predictive units are shown in dashed lines and retrospective units in solid lines; for each type, two examples are shown: one that either increases or decreases with time and one with a peak either before or after zero lag. **(B)** Top: Histogram of the distribution of the peaks at which maximal mutual information is found for units responsive to movement and modulated by speed (bin size = 50 ms). Bottom: maximal mutual information values of the units responsive to movement and modulated by speed with respect to the time lag.

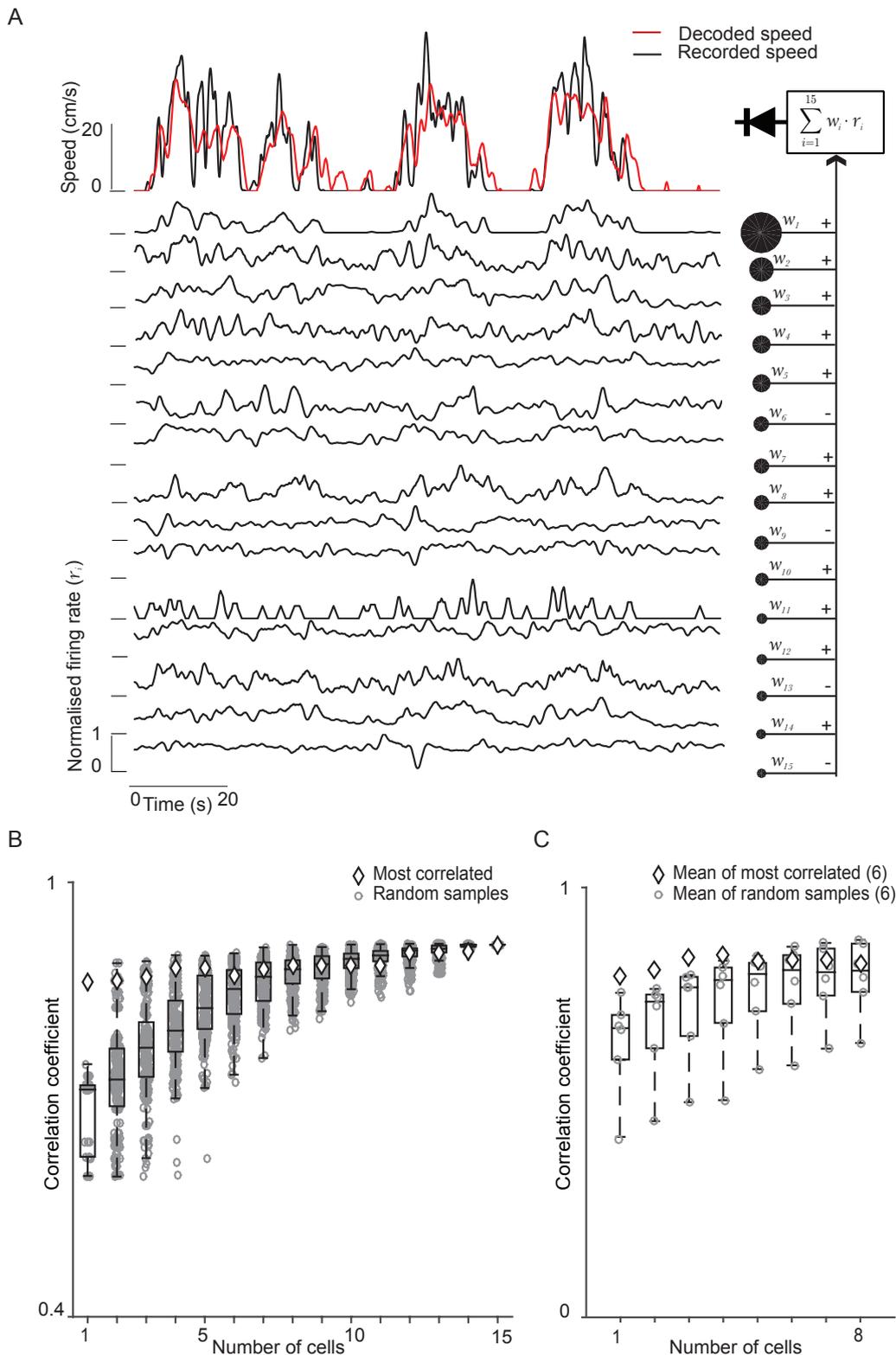
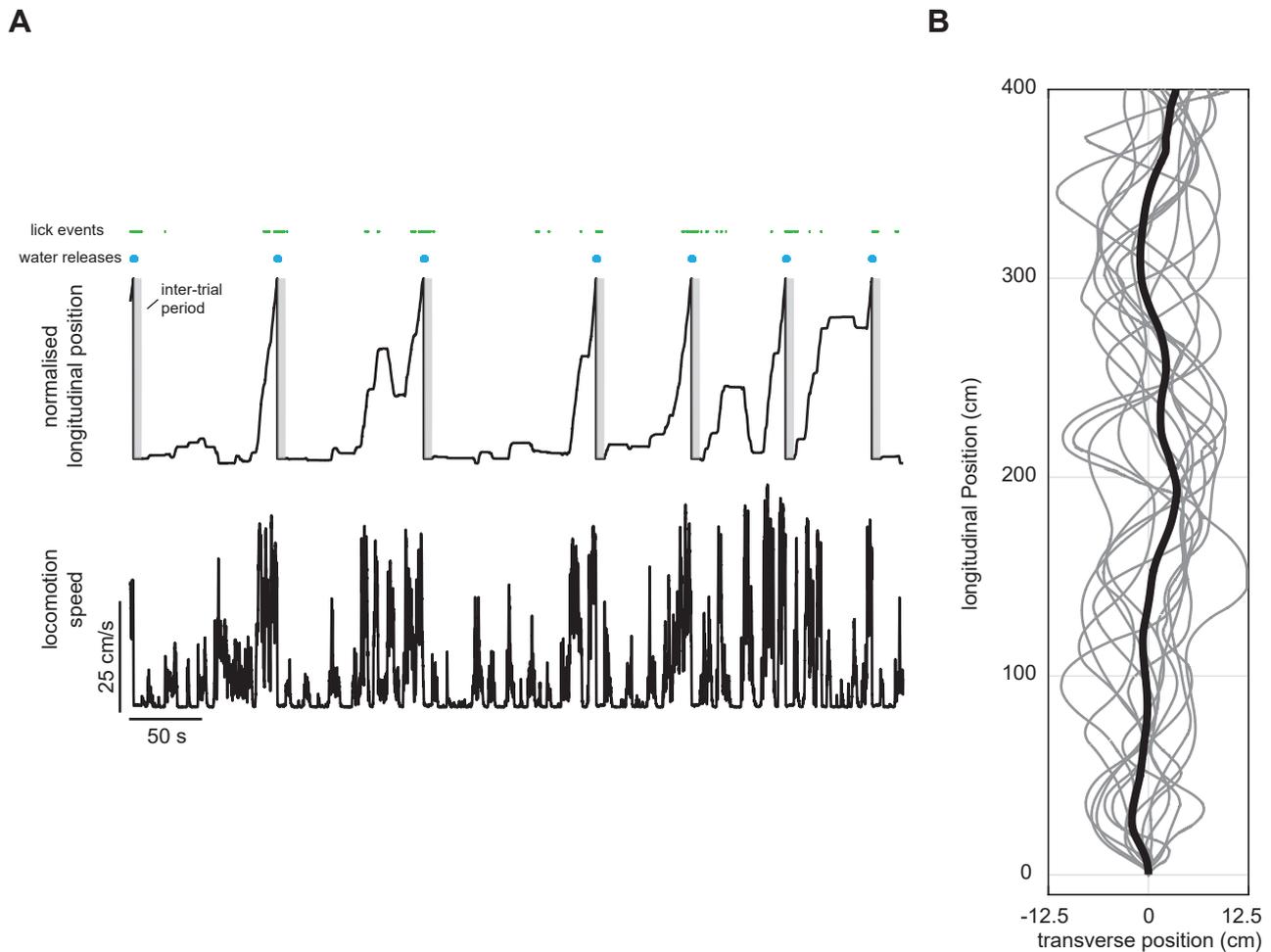
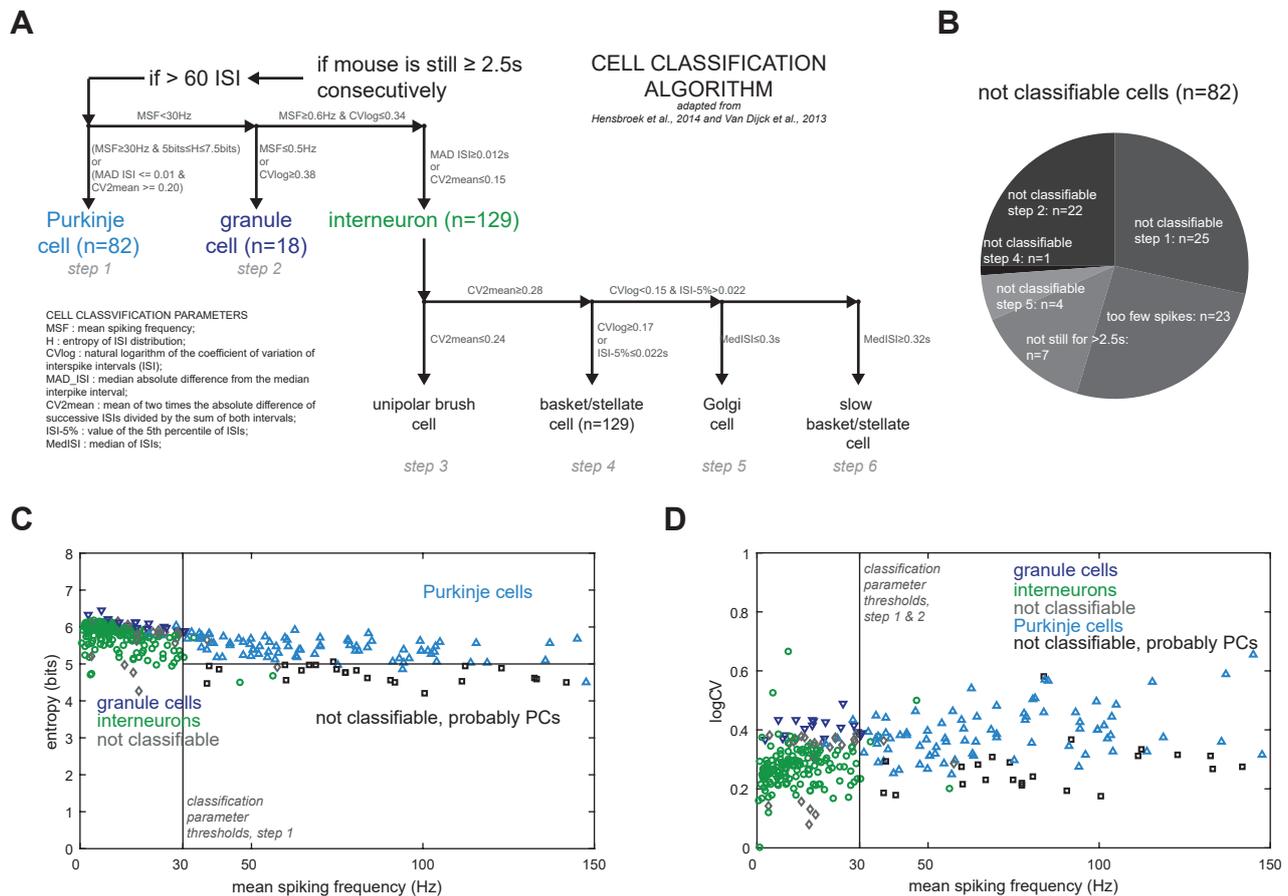


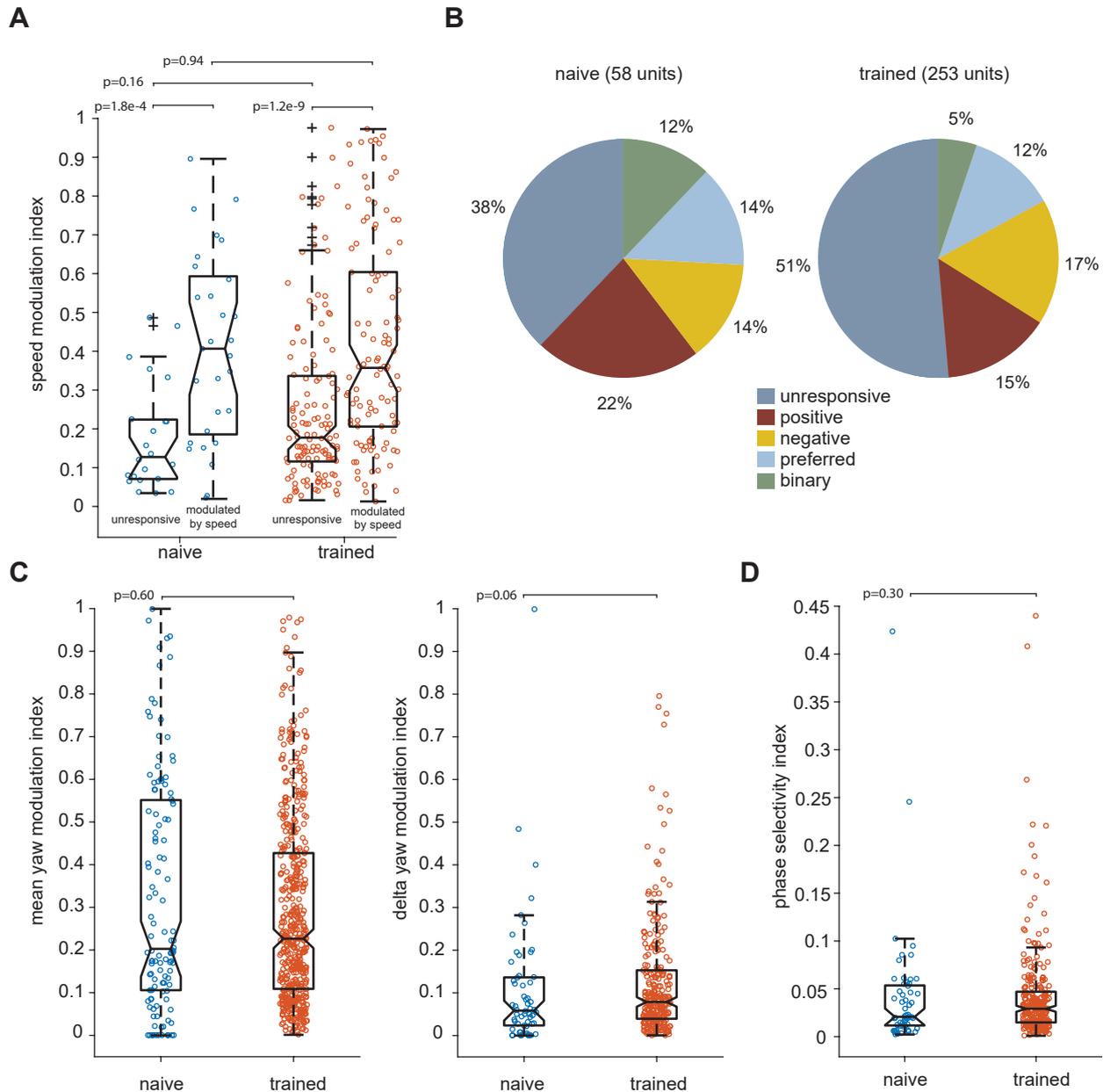
Figure 7: Population decoding of cerebellar neurons reconstruct speed. **(A)** Diagrammatic representation of the optimal (least MSE) linear decoder used to reconstruct locomotion speed showing one example experiment with 15 units. Top: Time course of original and reconstructed locomotor speed signal. Bottom, normalised firing rate of all units used for the decoder. Right: firing rates of all units are weighted and linearly summed with the result then being half-wave rectified (diode symbol). **(B)** Reconstruction quality as a function of the number of the units used for the decoder of the same example experiment shown in A. Box plots show first and third quartile with middle horizontal line indicating the median of  $2N$  combinations of populations' accuracy. **(C)** Mean reconstruction quality as a function of the number of the units for 6 different experiments. The average of the most correlated ensembles is plot together with the mean of the randomly formed ensembles.



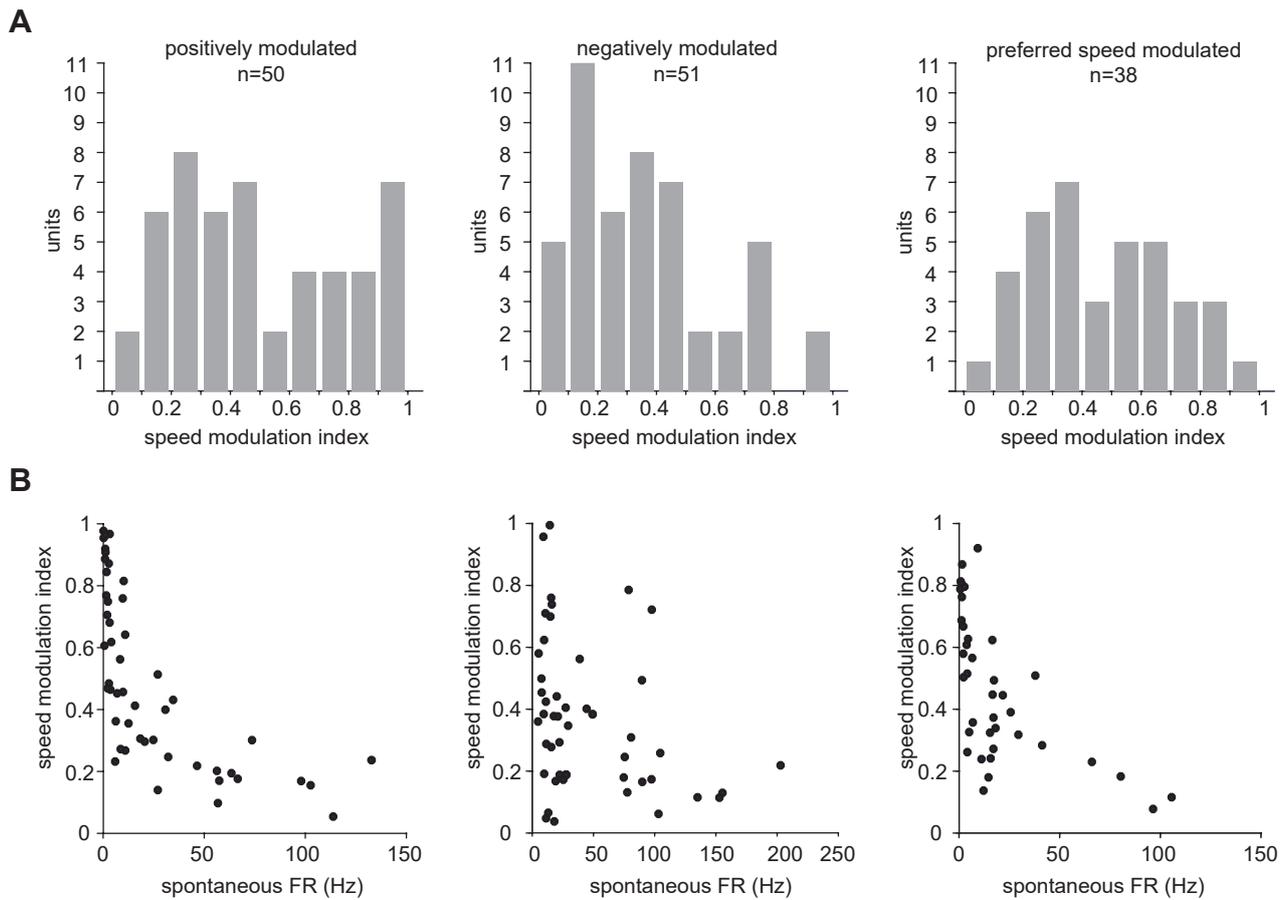
**Figure 1 - supplement 1.** Behaviour in virtual reality environment. (A) Top: normalised longitudinal position of a mouse in the virtual corridor with respect to time during a few example trials with lick events and water pump activations. The mouse starts licking when it gets closer to the end as it expects the liquid reward for completing the task. Gray areas indicate inter-trial periods (duration = 6 seconds) during which there is no virtual reality projection and a black screen stimulus is presented. Bottom: speed signal as recorded by the computer mice positioned on the spherical treadmill. (B) A few example traces from the same experiment in A showing the trajectories in the virtual corridor. Please note the different scales along the transverse and longitudinal directions. Black line indicate the mean trajectory of the trials shown.



**Figure 1 - supplement 2.** Identity classification of units. (A) Schematic of the classification algorithm based on the decision tree algorithm from Van Dijck et al., 2013 and Hensbroek et al., 2014. Purkinje, granule and interneuron cells are identified in the first three steps by using the parameters indicated on the figure. (B) Breakdown of the units that were not classifiable with reason and/or steps in which did not pass parameters thresholds. (C) Entropy and mean spontaneous spiking frequency of all units. Black lines show parameter thresholds for the first step of the classification algorithm. (D) Logarithm of the coefficient of variation of two consecutive ISIs and mean spontaneous spiking frequency of all units. Black line shows the parameter threshold for first classification step. Panel C and D are produced for comparison purposes with data shown by Van Dijck et al., 2013 in their research article.

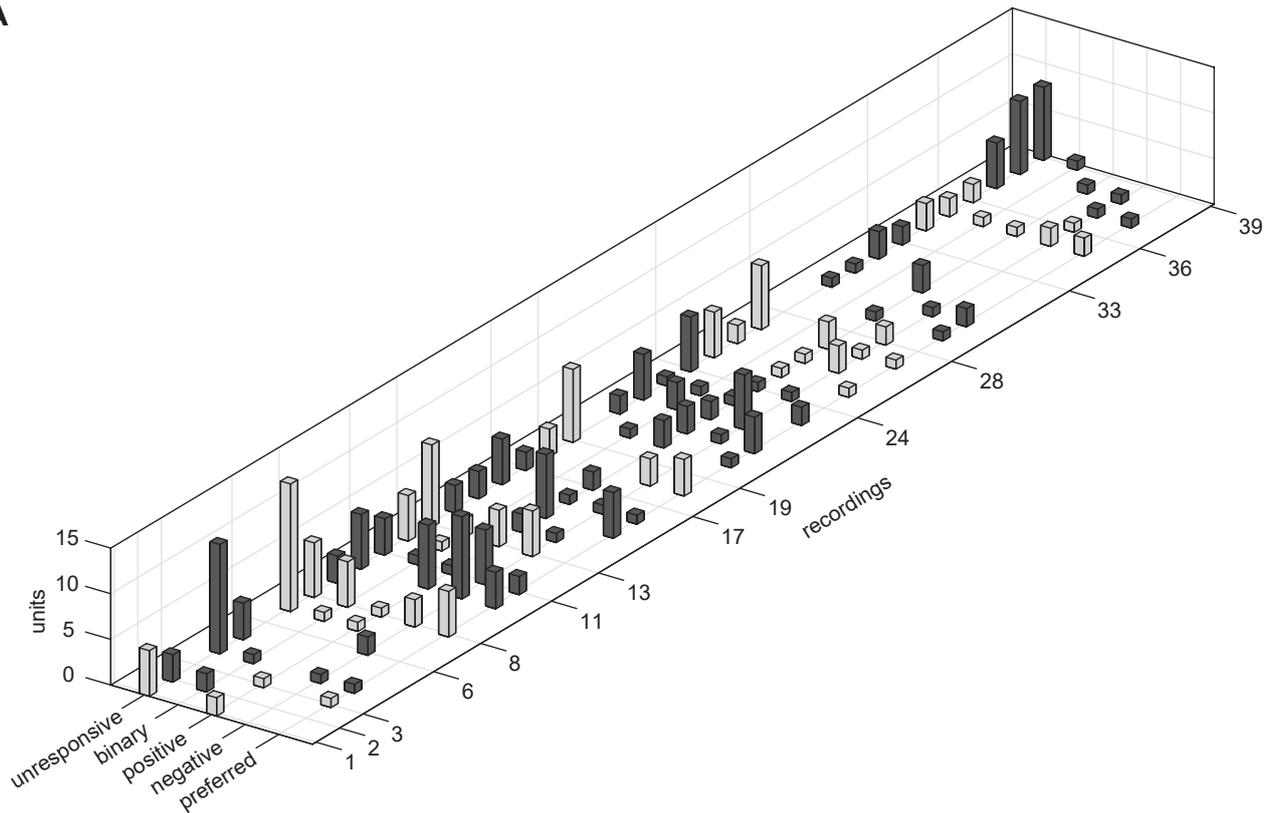


**Figure 2 - figure supplement 1.** No differences in the neural populations recorded in naive or trained animals. **(A)** The speed modulation indexes are not significantly different between naive (total number of units=58, units modulated by speed=29) or trained animals (total number of units=253, units modulated by speed=110) for unresponsive or speed modulated units. **(B)** Classification of speed-response profiles in the naive versus trained conditions: the percentages of each type is similar in both conditions. **(C)** Both the mean and the absolute difference between the modulation indexes between CW and CCW yaw tuning curve (delta yaw modulation index) are not significantly different. **(D)** No significant difference is found in the phase selectivity index (PSI) in the naive versus the trained animals.

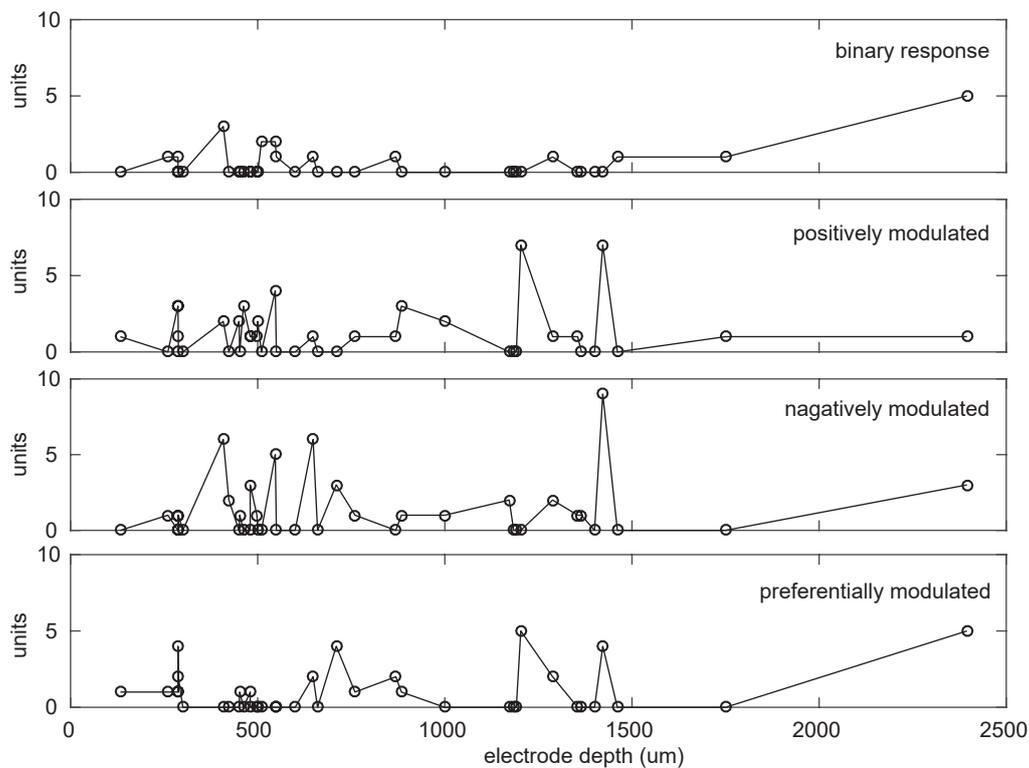


**Figure 2 - figure supplement 2.** Modulation of speed tuned units. **(A)** Modulation indexes of all units significantly modulated by speed, divided by response type. **(B)** Modulation indexes shown as a function of spontaneous firing rate (i.e. during stationary periods), divided by response type.

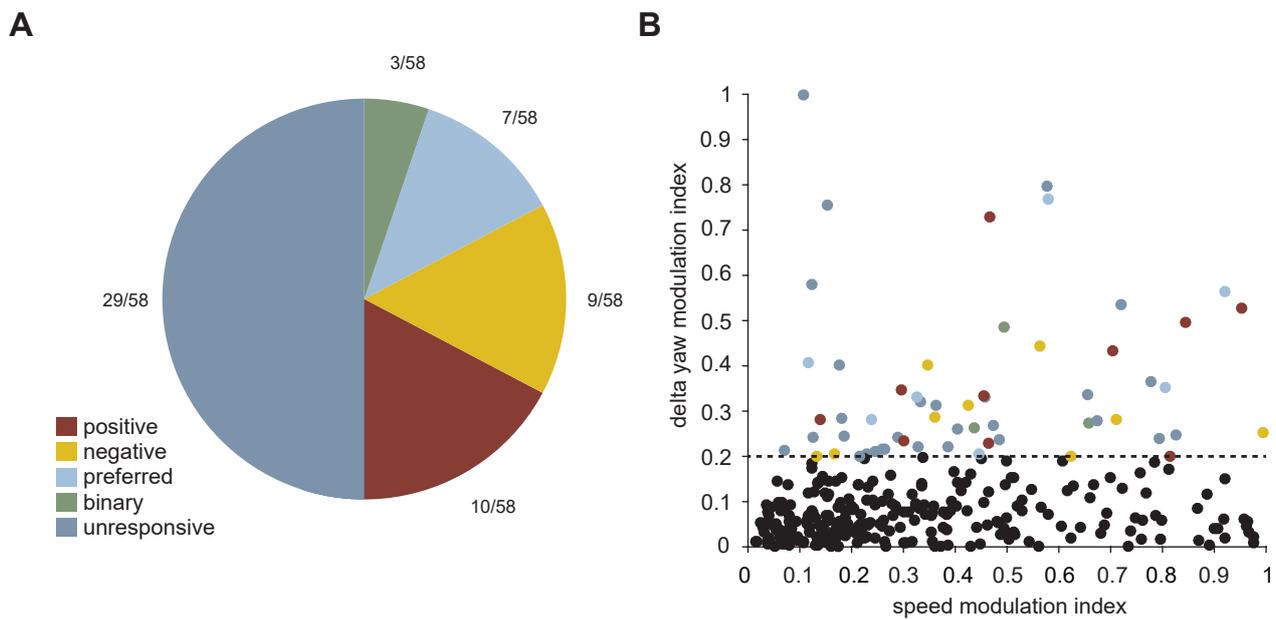
**A**



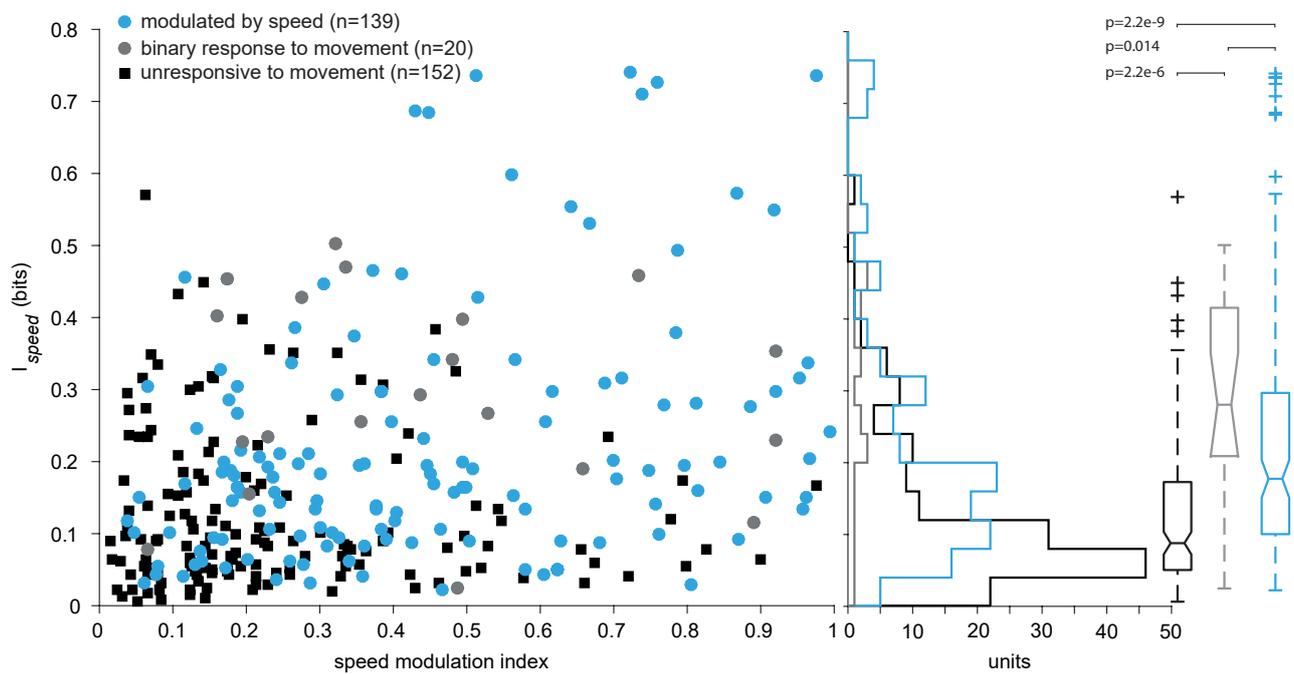
**B**



**Figure 2 - figure supplement 3.** Count of units per recording. (A) Count of units for every recording used in this study classified according to their response profile to locomotion speed. Black and white bars are used only to help distinguishing between consecutive animals; ticks on the recordings axes sign the switch to different animal. (B) Poll of units responsive to movement with respect to the depth of each recording.



**Figure 3 - figure supplement 1.** Yaw tuning responses with respect to speed tuning responses. **(A)** Out of the 58 units with an absolute difference in modulation indexes between CW and CCW yaw tuning curve (delta yaw modulation index) larger than 0.2, more than half are not modulated by speed, while the rest are evenly spread amongst the other response categories. **(B)** A large delta yaw modulation index is not related to the speed modulation index (n=311). The dashed line is the threshold that separates the 58 units represented in A.



**Figure 6 - figure supplement 1.** Mutual information versus speed modulation index. Mutual information of locomotion speed computed at zero lag as a function of the modulation index for speed modulated for all units. P values of Mann-Whitney U-test between the three distributions.