

Low-cost HPV screening and the prevalence of cervical infection in asymptomatic populations in Guatemala

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Running Title: Low cost HPV screening in Guatemala

ABSTRACT

Background: A low cost and reliable method for detecting high-risk (HR) HPV is important to permit HPV screening for cervical cancer prevention. We validated a low-cost commercially available HPV method (H13, HybriBio, Hong Kong) and determined the distribution of HPV infections in over 1717 cancer-free women in Guatemala.

Methods: H13 results were compared with two more established HPV tests: (Xpert™ (Cepheid) and SPF10-LIPA₂₅™ (DDL)). HR-HPV was detected in cervical samples from 1717 cancer-free women receiving Pap smears using the HybriBio™ real-time PCR assay of 13 HR types. HPV positive samples were sequenced to determine viral type.

Results: The HybriBio H13 Assay showed 93% agreement with Xpert, and a similar result with SPF10-LIPA₂₅ (kappa=0.78 and 0.76). A total of 13% (226/1717) of women tested HPV+. The highest prevalence was found in younger women (<30 years, 22 %) and older ones (≥60 years, 15%). The six most common HR-HPV types among the 148 HPV+ typed were HPV16 (22%), HPV18 (11%), HPV39 (11%), HPV58 (10%), HPV52 (8%), and HPV45 (8%).

Conclusions: The HybriBio Assay was low cost, and reliable in screening for HR-HPV infection. As in most places, HPV16 was the most prevalent HR type in Guatemala and the age-specific prevalence curve peaked in younger ages with a secondary peak possibly representing immune senescence in older women.

Highlights

1. A low cost and valid method, HybriBio Assay, could be used for CC screening in low income regions.
2. A total of 13% of cancer-free women were HPV+ and positivity was associated with younger age (<30 years old) in Guatemala.
3. HPV16 was the major prevalent type.

Keywords HPV; cervical cancer; Guatemala, prevalence, screening, real-time PCR

Introduction

In Guatemala, cervical cancer (CC) is a leading cause of cancer in women (1530 cases/year, Age-standardized rate (ASR) 31/100,000) resulting in an estimated 717 deaths) (Globocan 2012). In the Instituto de Cancerología (INCAN), the main adult oncology hospital in Guatemala, over 40% of women diagnosed with malignancies have advanced CC, requiring costly management that often has a poor outcome (1). Therefore, a focus on prevention is important.

There is overwhelming evidence that persistent infection with specific types of HPV is the main cause of CC (2, 3). HPV types that are classified as established carcinogens are HPV16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, and possibly 68 (4). While prophylactic vaccination of adolescents and possibly young adults is the ultimate preventive strategy, screening will remain important for decades to come. Cytology-based screening has been associated with a major reduction in the incidence and mortality of the disease in developed countries (5). However, cytology is either unavailable or poorly conducted in most low-income countries (6).

In the general population, HPV prevalence reaches its peak of 25-30% in women in their early 20s (7-9), and typically declines to 8-15% from age 30-45 (10). A second peak is often observed in postmenopausal women (11, 12)]. HPV DNA testing has been proposed as an alternative to cytology in women older than 25-30 years (13-15). However, commercially available tests are typically expensive and require sophisticated equipment (16, 17). The use of HPV assays targeting lower-resource settings would be useful for CC prevention in such settings, which contribute most the world CC burden.

This study sought to assess the concordance of a new low-cost HPV assay (H13, HybriBio Ltd, Hong Kong) with established commercial tests, GeneXpert (Xpert, Cepheid, California, USA) and SPF10-LIPA₂₅ (DDL, Netherlands) assays (18-20). In addition, we determined the prevalence of HPV infection in the general female population in Guatemala.

Materials and Methods

Study populations

To validate the H13 test against other clinical tests, a convenience sample of 40 mostly positive subjects undergoing screening at the Curacao Medical Lab was employed. In addition, a total of 1717 samples were obtained from the general population of asymptomatic sexually active women undergoing routine screening with Pap smear at hospital-based screening clinics in Guatemala, after obtaining informed consent. A questionnaire on reproductive and socio-demographic characteristics was administered by trained personnel on the Guatemalan subjects (21). The samples were collected by a medical practitioner using a Dacron swab placed in a tube containing 3 ml of Scope mouthwash, maintained at 4° C and transported at room temperature (22). The study was approved by an Institutional Review Board (IRB) and testing in the US laboratory was judged exempt by the NIH Office of Human Studies Research.

HPV testing

The detection of HPV was performed using a HybriBio Assay that detects 13 High-risk HPV types by Real time PCR (HybriBio Biochem Co. Ltd. China). A cell lysate was prepared per the manufacturer's instruction. To determine HPV type, HPV positive samples were subjected to Touchdown PCR and DNA sequencing (Fig. 2). Samples in which the internal control (IC) did not amplify and those that remained negative on retesting were excluded.

The sensitivity of the Touchdown PCR was determined by a series of 10-fold dilutions of DNA from HPV+ and HPV- cell lines using Broad-Spectrum (BS) GP5+ and GP6+ Primers (BSGP5+/6+) (150bp) (Supplementary Fig. 1) (23). For the HybriBio Assay, each 96-well plate included four HPV+ controls: CC cell line DNA from HeLa (HPV18), CaSki (HPV16), MS751 (HPV45) DNA and plasmid DNA from the HybriBio Assay kit; HPV- controls included C33A

DNA and water. The HPV+ samples were amplified by using 400 nM BSGP5+/6+. Briefly, 10 min denaturation step at 95°C was followed by 40 cycles of amplification. Each cycle included denaturation at 94°C for 20 s, annealing at 38°C for 30 s, and elongation at 71° C for 80 s. The final elongation step was 5 min. The ramping rates were adjusted as described (24); 1.8° C/cycle from 74° C to 38° C in first 20 cycles. Each experiment included HPV+ and HPV- controls and a sample lacking template DNA (Supplementary Fig. 1 and 2). The PCR products were subjected to Sanger Sequencing on an ABI3730XL. Sequences were analyzed by assembly and trimming in SeqMan (DNASTAR, Madison, WI) followed by BLAST search (NCBI). Samples with inconclusive Sanger sequence were repeated with a next-generation sequencing method (S. Wagner, and J. Boland, manuscript in preparation).

Statistical analyses

We compared the detection of HPV by the HybriBio Assay to Xpert and SPF10-LIPA₂₅ in 40 cervical cytology samples collected in a clinical laboratory in Curacao and calculated percent agreement and kappa. Statistical analyses on the Guatemalan samples were performed to determine age-specific HPV prevalence, comparing the age groups with the Pearson Chi-square test including Fisher's exact test using GraphPad Prism version 7 for Windows. $P < 0.05$ was regarded as statistically significant. We performed analyses of association between HPV infection and other risk factors (age, age of menarche, age of first birth, smoking history, oral contraceptives and cooking method as a measure of socioeconomic status).

Results

To evaluate the HybriBio H13 test as a potential low cost assay for HR-HPV screening we compared it against two commercial assays, Xpert and DDL. Table 1 shows that high levels of agreement were seen between HybriBio Assay and the other tests (93%, kappa=0.78 and 0.76, respectively). Also, 89% agreement (kappa=0.67) was observed between SPF10-LIPA₂₅ and Xpert (**Table 1**). We also evaluated the sensitivity and required assay volume of the H13 test and determined that a 10ul real-time PCR volume gave equivalent results (Supplementary Table 1). In a separate study, we determined that the H13 test is xxx. Therefore, H13 is a low-cost and reliable assay for HPV screening.

To apply the H13 test to samples from a low and middle income country, we used the test to determine the prevalence and distribution of HPV types in Guatemala. We recruited asymptomatic women from the general population at hospital clinics performing cytology screening. The women sampled ranged in age from 17 to 79 years attending clinics in Guatemala City and the city of Puerto Barrios. The data on age of menarche, reproductive history, smoking, oral contraceptive use, and wood smoke exposure is shown in **Table 2**.

To determine the prevalence of HPV infection, 1717 subjects were tested (Fig. 2). The overall prevalence of HPV infection was 13% (226/1717) (Table 3). HPV typing was successful in 148 HPV+ samples and the 13 HR-HPV types 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 68 were detected in 143 subjects (Table 3.) (143/226). Of the 13 HR types, HPV16 (22%, 32/148), HPV18 (11%, 16/148) accounted for 32% (48/148) and HPV39, 58, 45, 52 combined 37%, 55/148). Types of unknown risk (Unk), HPV67 and 74 were detected in 1% (Table 3).

To understand the age specific prevalence of HPV infection, the women were divided into 6 age groups (Table 1). HPV prevalence ranged from 22% in the <30 to 8% to 14% in the 30-59 age groups, and 15% in the ≥ 60 -year group. Age specific prevalence for HPV was significantly higher in the younger age groups (<30) ($P=0.0022$). Interestingly, the oldest age group (≥ 60) showed an increased prevalence as compared to age groups from 30-59 (Fig. 1 and Table 2).

Discussion

Numerous studies support HPV testing as the most sensitive primary screening method for CC. However, there are few HPV tests that are affordable for LMICs. We performed a small comparison study of the HybriBio H13 test with two other commercial assays, GeneXpert and DDL and demonstrate a high rate of concordance. Commercial tests in Guatemala cost between \$100-210, out of the range of practical use. Mexico carried out a large screening program with the hybrid capture (HC2, Qiagen) at an approximate cost of \$11 per test, but many poor and rural areas remain unscreened. A method has been developed with support from medical foundations, called careHPV (Qiagen). This system has been rigorously tested in China, India and in pilot programs in other areas (25) and is under further evaluation in several Latin American countries. However, there is still considerable discussion on the most effective strategies for managing HPV+ women in different economic and cultural settings (26-28).

We sought to establish a method that would be cost effective, and use only equipment available in a standard molecular biology laboratory. We employed a validated storage buffer, mouthwash containing 15% ethanol, which costs \$0.01 per sample with fewer shipping requirements than methanol based buffers (22). The HybriBio Assay under the conditions we employed (10ul reaction volume), costs about \$3/assay (Table S1). While HybriBio requires a real-time PCR instrument, we have purchased used ABI7000 instruments for under \$1000, and found that some Latin American hospitals and clinical laboratories have real-time PCR instrumentation.

In this study, the HybriBio Assay was compared for detection of HPV to two clinically validated tests (Xpert and SPF10-LIPA₂₅). Both HybriBio Assay and Xpert test target the E6 and E7 of 13 and 14 HR-HPV types, respectively. HybriBio Assay has a pooled probe for HR-HPV

types, 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 68; whereas Xpert test detects HR-HPV16, 18/45 in separate detection channels, with 11 other HR types detected in 3 additional channels. HR types detected are the same as the HybriBio Assay except HPV66 is included [19, 20]. The SPF10-LIPA₂₅ test targets the conserved L1 region and detects 14 HR types and 11 LR types (18). In a separate study, we further validate H13 against HC2 (Qiagen) and Onclarity (BD Diagnostics) (Fokom Domgue et al, manuscript in press).

Few studies of HPV prevalence have been described in Guatemala, even though this country has one of the highest incidences of HPV related diseases and mortality in Latin America (1, 29). However, our study has the advantage of collecting samples from healthy women using a low-cost HPV screening method. The overall prevalence of HR HPV infection in this study was 13% for Guatemala, similar to rural Costa Rica (30) and lower than studies from Northern Spain (29%) (7) and other countries (25% to 29%) (31, 32). The difference is likely due to differences in age between the populations.

As seen in most countries, HPV16 is the most common type found in women with or without cancer (33, 34). The combined prevalence of HPV16, and HPV18 was highest in the youngest age groups in this country. A similar prevalence of HPV16 and 18 was reported in other studies (35) as well as in Guatemalans with CC (21). In addition to HPV16 and 18, 15 other HPV types were observed frequently in our study, most notably HR-HPV 39, 58, 45, and 52.

Women <30 years of age had the highest prevalence, while the HPV prevalence decreased markedly with increasing age, up to age 60. This trend has been observed in Costa Rica and other studies (7-9). In Guatemala, there is a second peak in women ≥ 60 . Similar results

were observed in Guangdong China (11) and Russia (12) and may be associated with reduced clearance or reactivation of HR-HPV infection at later age.

Our study has several limitations that might affect our conclusions. We used a HybriBio Assay that detects only 13 HR types. In addition, we attempted to sequence all positive samples to determine type. In a small portion of samples, we had a failure to detect the IC indicating a failure in sample collection, preservation or storage. Most of these samples were negative using both HybriBio and a touchdown PCR method (data not shown). This could indicate a limitation to using Scope mouthwash as a preservative. We have limited cytology data on the women and have not demonstrated that HybriBio Assay is effective in CC prevention. However, the manufacturer reports data on a comparison with Qiagen (with >95% agreement with and FDA approved test), the test passed two WHO proficiency trials and has been used in a study of 48,559 women in China (11).

In conclusion, we have established the HybriBio H13 test as an affordable alternative for HPV screening. HPV infection was detected in 13% of asymptomatic women in Guatemala. The distribution of HPV types is typical of other countries and the highest HPV prevalence is in youngest age groups (<30). This low-cost approach to detect HPV could be employed in other countries to reduce the burden of CC.

Conflict of interest statement

None declared

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Tables

Table 1. Comparison of the HybriBio Assay to the Xpert and SPF10-LIPA₂₅ assay

					HR HPV, No.	
	Pos/Pos	Pos/Neg	Neg/Pos	Neg/Neg	Overall Agreement (%)	Kappa
HybriBio Assay/Xpert	30	2	1	7	93%	0.78
HybriBio Assay/SPF10-LIPA₂₅*	22	0	2	4	93%	0.76
SPF10-LIPA₂₅/Xpert	21	3	0	4	89%	0.67
Data shown for the 13 HR types in common to the three assays. HR, high risk; HPV, human papillomavirus; Pos, positive; Neg, negative.			0	4	89%	0.67

Table 2. Crude and multivariate analysis for risk factors related to 13HR HPV detection by HybriBio assay

Age group	Total No.	HPV+ No.	HPV+ (%)	Crude OR for 13HR HPV	95% CI	p	Age adjusted OR for 13HR HPV	95% CI	p
<30	17	5	29						
30-34	209	26	12	0.34	0.12 - 1.1	0.060	0.34	0.12 - 1.1	0.060
35-39	293	31	11	0.28	0.098 - 0.94	0.025	0.28	0.098 - 0.94	0.026
40-49	496	48	10	0.26	0.091 - 0.84	0.014	0.26	0.091 - 0.84	0.014
50-60	370	36	10	0.26	0.090 - 0.85	0.015	0.26	0.090 - 0.85	0.016
>60	7	2	29	0.96	0.11 - 6.4	0.970	0.96	0.11 - 6.4	0.97
Age of menarche									
<13	772	84	11						
≥13	579	58	10	1.1	0.59 - 2.00	0.69	1.2	0.61 - 2.1	0.62
Age of first birth									
<20	760	95	13						
≥20	582	51	9	0.67	0.47 - 0.96	0.030	0.67	0.47 - 0.96	0.032
Smoking history									
Yes	73	7	10	0.89	0.36 - 1.8	0.77	0.91	0.37 - 1.9	0.82
No	1319	141	11						
Oral contraceptives									
Yes	300	33	11	1.00	0.65 - 1.5	0.99	0.95	0.60 - 1.5	0.81
No	952	105	11						
Cooking method									
Wood & Gas	970	105	11	0.90	0.61 - 1.3	0.58	0.91	0.62 - 1.3	0.64
Gas only	417	41	10						

Table 3. Prevalence of HR-HPV types by HybriBio assay in cancer free women in Guatemala

HPV types	No.	Specific HPV type in 13HR (%)
13 HR types*	32	22%
16	16	11%
18	4	3%
31	2	1%
33	6	4%
35	16	11%
39	12	8%
45	5	3%
51	12	8%
52	6	4%
56	15	10%
58	10	7%
59	7	5%
68	1	1%
Other types	4	1%
67 [#]	1	1%
70 ^{**}	1	1%
73 [*]	1	1%
74 [#]	2	1%
with HPV types No.148		
Total HPV+ No.	226	
Total HPV- No.	1491	
Total No.	1717	

Overall HPV+ frequency **13%**

***:** **HR**

****:** **LR**

#: **Unknown**

Table S1. Comparison of the HybriBio Assay with 10 μ l to 20 μ l volume

Sample	Input DNA (5ng/ μ l)		13HR HPV Types (FAM)	Internal Control (JOE)	
	Total Volume (μ l)	Volume Input DNA (μ l)	(ng)	Ct	
HPV positive controls					
CaSki (HPV16)	20	2	10	15.74	27.26
	10	1	5	16.15	28.61
HeLa (HPV18)	20	2	10	22.26	26.23
	10	1	5	23.03	26.26
MS751 (HPV45)	20	2	10	26.90	29.79
	10	1	5	27.96	29.91
ME180 (HPV39/18)	20	2	10	27.61	25.99
	10	1	5	27.72	25.96
Unknown samples from cell lysate					
S1 DD015959	20	2		ND*	24.05
	10	1		ND	24.80
S2 DD015960	20	2		17.60	27.59
	10	1		18.02	28.10
S3 DD015963	20	2		16.85	20.78
	10	1		17.20	21.30
S6 DD015964	20	2		16.01	22.05
	10	1		16.80	22.70

HPV negative control (C33A)	20	2	10	ND	25.66
	10	1	5	ND	25.04
Controls from kit					
Positive control (Kit)	20	2		23.88	23.39
	10	1		25.07	25.68
Negative control (Kit)**	20	2		ND	ND
	10	1		ND	ND

***:** Not detected

****:** Dnase-free distilled water from kit

Figure Legends

Figure 1. Age specific prevalence of HPV by age group in Guatemala. The prevalence of HPV in asymptomatic women in Guatemala is displayed by age group. A Pearson test of all six groups is statistically significant ($P < 0.0022$). The number of total women in each age group (n) is shown.

Figure 2. Flow chart of participants and associated HPV test outcomes.

References

1. Valles X, Murga GB, Hernandez G, Sabido M, Chuy A, Lloveras B, et al. High prevalence of human papillomavirus infection in the female population of Guatemala. *Int J Cancer* 2009;125(5):1161-7.
2. Ngelangel C, Munoz N, Bosch FX, Limson GM, Festin MR, Deacon J, et al. Causes of cervical cancer in the Philippines: a case-control study. *J Natl Cancer Inst* 1998;90(1):43-9.
3. Smith JS, Lindsay L, Hoots B, Keys J, Franceschi S, Winer R, et al. Human papillomavirus type distribution in invasive cervical cancer and high-grade cervical lesions: a meta-analysis update. *Int J Cancer* 2007;121(3):621-32.
4. Bouvard V, Baan R, Straif K, Grosse Y, Secretan B, El Ghissassi F, et al. A review of human carcinogens--Part B: biological agents. *Lancet Oncol* 2009;10(4):321-2.
5. Arbyn M, Raifu AO, Weiderpass E, Bray F, Anttila A. Trends of cervical cancer mortality in the member states of the European Union. *Eur J Cancer* 2009;45(15):2640-8.
6. Gakidou E, Nordhagen S, Obermeyer Z. Coverage of cervical cancer screening in 57 countries: low average levels and large inequalities. *PLoS Med* 2008;5(6):e132.
7. de Ona M, Alvarez-Arguelles ME, Torrents M, Villa L, Rodriguez-Feijoo A, Palacio A, et al. Prevalence, evolution, and features of infection with human papillomavirus: a 15-year longitudinal study of routine screening of a women population in the north of Spain. *J Med Virol* 2010;82(4):597-604.
8. Leinonen MK, Anttila A, Malila N, Dillner J, Forslund O, Nieminen P. Type- and age-specific distribution of human papillomavirus in women attending cervical cancer screening in Finland. *Br J Cancer* 2013;109(11):2941-50.
9. Ley C, Bauer HM, Reingold A, Schiffman MH, Chambers JC, Tashiro CJ, et al. Determinants of genital human papillomavirus infection in young women. *J Natl Cancer Inst* 1991;83(14):997-1003.
10. Schiffman M, Doorbar J, Wentzensen N, de Sanjose S, Fakhry C, Monk BJ, et al. Carcinogenic human papillomavirus infection. *Nat Rev Dis Primers* 2016;2:16086.
11. Chen Q, Xie LX, Qing ZR, Li LJ, Luo ZY, Lin M, et al. Epidemiologic characterization of human papillomavirus infection in rural Chaozhou, eastern Guangdong Province of China. *PLoS One* 2012;7(2):e32149.
12. Syrjanen K. New concepts on risk factors of HPV and novel screening strategies for cervical cancer precursors. *Eur J Gynaecol Oncol* 2008;29(3):205-21.
13. Mayrand MH, Duarte-Franco E, Rodrigues I, Walter SD, Hanley J, Ferenczy A, et al. Human papillomavirus DNA versus Papanicolaou screening tests for cervical cancer. *N Engl J Med* 2007;357(16):1579-88.
14. Naucler P, Ryd W, Tornberg S, Strand A, Wadell G, Elfgren K, et al. Human papillomavirus and Papanicolaou tests to screen for cervical cancer. *N Engl J Med* 2007;357(16):1589-97.
15. Rijkaart DC, Berkhof J, Rozendaal L, van Kemenade FJ, Bulkman NW, Heideman DA, et al. Human papillomavirus testing for the detection of high-grade cervical intraepithelial neoplasia and cancer: final results of the POBASCAM randomised controlled trial. *Lancet Oncol* 2012;13(1):78-88.
16. Barski A, Cuddapah S, Cui K, Roh TY, Schones DE, Wang Z, et al. High-resolution profiling of histone methylations in the human genome. *Cell* 2007;129(4):823-37.

17. Green RE, Krause J, Ptak SE, Briggs AW, Ronan MT, Simons JF, et al. Analysis of one million base pairs of Neanderthal DNA. *Nature* 2006;444(7117):330-6.
18. Castle PE, Porras C, Quint WG, Rodriguez AC, Schiffman M, Gravitt PE, et al. Comparison of two PCR-based human papillomavirus genotyping methods. *J Clin Microbiol* 2008;46(10):3437-45.
19. Cuschieri K, Geraets D, Cuzick J, Cadman L, Moore C, Vanden Broeck D, et al. Performance of a Cartridge-Based Assay for Detection of Clinically Significant Human Papillomavirus (HPV) Infection: Lessons from VALGENT (Validation of HPV Genotyping Tests). *J Clin Microbiol* 2016;54(9):2337-42.
20. Einstein MH, Smith KM, Davis TE, Schmeler KM, Ferris DG, Savage AH, et al. Clinical evaluation of the cartridge-based GeneXpert human papillomavirus assay in women referred for colposcopy. *J Clin Microbiol* 2014;52(6):2089-95.
21. Lou H, Villagran G, Boland JF, Im KM, Polo S, Zhou W, et al. Genome Analysis of Latin American Cervical Cancer: Frequent Activation of the PIK3CA Pathway. *Clin Cancer Res* 2015;21(23):5360-70.
22. Castle PE, Sadorra M, Garcia FA, Cullen AP, Lorincz AT, Mitchell AL, et al. Mouthwash as a low-cost and safe specimen transport medium for human papillomavirus DNA testing of cervicovaginal specimens. *Cancer Epidemiol Biomarkers Prev* 2007;16(4):840-3.
23. Schmitt M, Dondog B, Waterboer T, Pawlita M. Homogeneous amplification of genital human alpha papillomaviruses by PCR using novel broad-spectrum GP5+ and GP6+ primers. *J Clin Microbiol* 2008;46(3):1050-9.
24. Snijders PJ, van den Brule AJ, Jacobs MV, Pol RP, Meijer CJ. HPV DNA detection and typing in cervical scrapes. *Methods Mol Med* 2005;119:101-14.
25. Trope LA, Chumworathayi B, Blumenthal PD. Preventing cervical cancer: stakeholder attitudes toward CareHPV-focused screening programs in Roi-et Province, Thailand. *Int J Gynecol Cancer* 2009;19(8):1432-8.
26. Gottschlich A, Rivera-Andrade A, Grajeda E, Alvarez C, Mendoza Montano, Meza R. Acceptability of Human Papillomavirus Self-Sampling for Cervical Cancer Screening in an Indigenous Community in Guatemala. *Journal of Global Oncology* 2017;0(0):JGO.2016.005629.
27. Jeronimo J, Holme F, Slavkovsky R, Camel C. Implementation of HPV testing in Latin America. *J Clin Virol* 2016;76 Suppl 1:S69-73.
28. Schiffman M, Wentzensen N, Wacholder S, Kinney W, Gage JC, Castle PE. Human papillomavirus testing in the prevention of cervical cancer. *J Natl Cancer Inst* 2011;103(5):368-83.
29. Nunez-Troconis J, Delgado M, Gonzalez J, Mindiola R, Velasquez J, Conde B, et al. Prevalence and risk factors of human papillomavirus infection in asymptomatic women in a Venezuelan urban area. *Invest Clin* 2009;50(2):203-12.
30. Herrero R, Hildesheim A, Bratti C, Sherman ME, Hutchinson M, Morales J, et al. Population-based study of human papillomavirus infection and cervical neoplasia in rural Costa Rica. *J Natl Cancer Inst* 2000;92(6):464-74.
31. de Sanjose S, Diaz M, Castellsague X, Clifford G, Bruni L, Munoz N, et al. Worldwide prevalence and genotype distribution of cervical human papillomavirus DNA in women with normal cytology: a meta-analysis. *Lancet Infect Dis* 2007;7(7):453-9.
32. Dunne EF, Unger ER, Sternberg M, McQuillan G, Swan DC, Patel SS, et al. Prevalence of HPV infection among females in the United States. *Jama* 2007;297(8):813-9.

33. Alibegashvili T, Clifford GM, Vaccarella S, Baidoshvili A, Gogiashvili L, Tsagareli Z, et al. Human papillomavirus infection in women with and without cervical cancer in Tbilisi, Georgia. *Cancer Epidemiol* 2011;35(5):465-70.
34. Zhao R, Zhang WY, Wu MH, Zhang SW, Pan J, Zhu L, et al. Human papillomavirus infection in Beijing, People's Republic of China: a population-based study. *Br J Cancer* 2009;101(9):1635-40.
35. Haguenoer K, Giraudeau B, Gaudy-Graffin C, de Pinieux I, Dubois F, Trignol-Viguier N, et al. Accuracy of dry vaginal self-sampling for detecting high-risk human papillomavirus infection in cervical cancer screening: a cross-sectional study. *Gynecol Oncol* 2014;134(2):302-8.

Supplementary Figure Legends

Figure S1. Amplification of HPV positive and negative cell line genomic DNA by Touchdown PCR method using BS GP5+/6+ Primers.

Figure S2. Touchdown PCR from swab cell lysate, (A): Lane 1.- Lane 4 HPV positive control with input DNA 1ng, 0.1ng, 0.01ng and 0.001ng; Lane 5. HPV negative control (cell line); Lane 6. no template control. Lane 7 to Lane 16 indicate unknown swab samples. M indicate the DNA ladder marker. Touchdown PCR from HPV positive swabs by HybriBio Assay (B): Lane 1. – Lane 18 are swab cell lysate; Lane 19. HPV positive control (cell line); Lane 20. HPV negative control (cell line); Lane 21. no template control. The samples were amplified with the BSGP5+/6+ primers.

Figure 1.

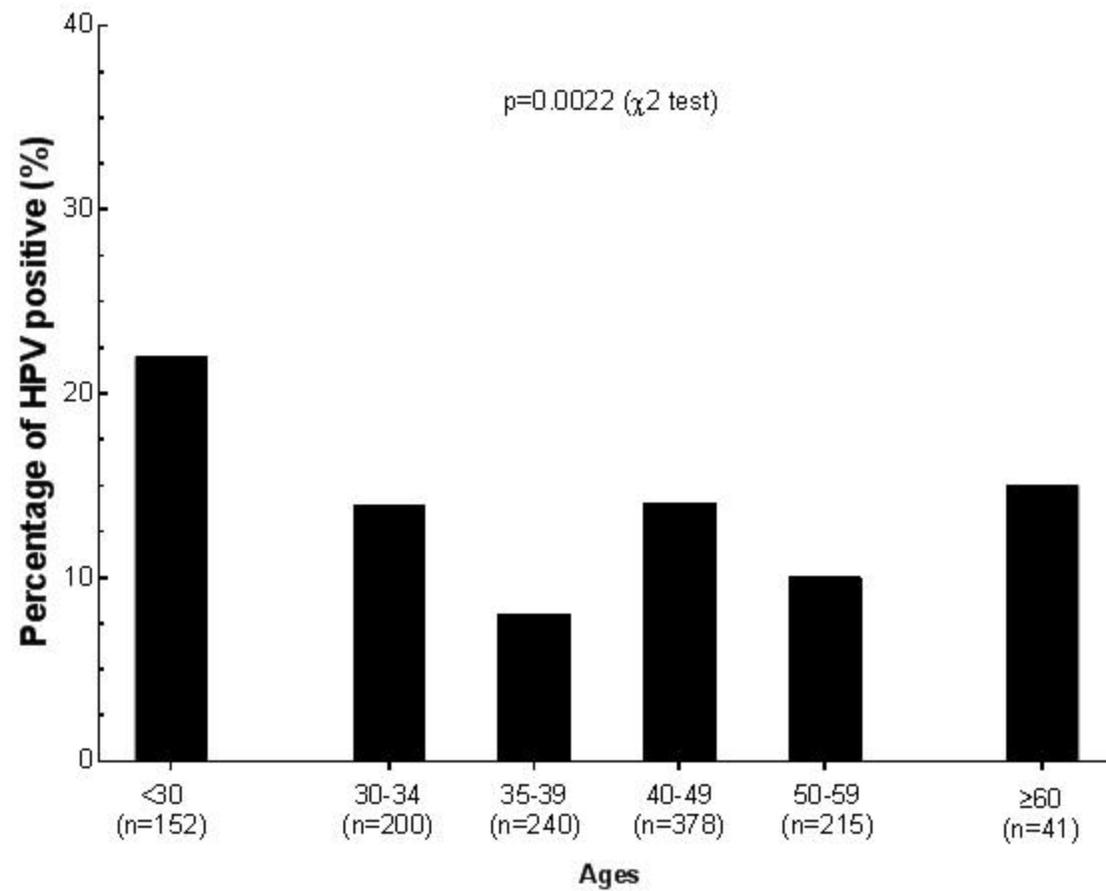


Figure 2.

