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FinisherSC : A repeat-aware tool for upgrading de-novo assembly using long reads

Ka-Kit Lam [1], Kurt LaButti [2], Asif Khalak [3]*and David Tse [1,4]†

[1]Department of Electrical Engineering and Computer Sciences, UC Berkeley
[2]U.S. Department of Energy Joint Genome Institute, Walnut Creek, CA, USA
[3]Pacific Biosciences, Menlo Park, CA, USA
[4]Department of Electrical Engineering, Stanford University

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ABSTRACT

We introduce FinisherSC, a repeat-aware and scalable tool for upgrading de-novo assembly using long reads. Experiments with real data suggest that FinisherSC can provide longer and higher quality contigs than existing tools while maintaining high concordance.

Availability: The tool and data are available and will be maintained at http://kakitone.github.io/finishingTool/

Contact: dntse@stanford.edu

1 INTRODUCTION

In de-novo assembly pipelines for long reads, reads are often trimmed or thrown away. Moreover, there is no evidence that state-of-the-art assembly pipelines are data-efficient. In this work, we ask whether state-of-the-art assembly pipelines for long reads have already used up all the available information from raw reads to construct assembly of the highest possible quality. To answer this question, we first collect output contigs from the HGAP (Chin et al., 2013) pipeline and the associated raw reads. Then, we pass them into our tool FinisherSC (Lam et al., 2014b) to see if higher quality assemblies can be consistently obtained after post-processing.

2 METHODS

2.1 Usage and pipeline

FinisherSC is designed to upgrade de-novo assembly using long reads (e.g. PacBio® reads). It is especially suitable for data consisting of a single long reads library. Input to FinisherSC are contigs (contigs.fasta) constructed by an assembler and all the raw reads with adaptors removed (raw_reads.fasta). Output of FinisherSC are upgraded contigs (improved3.fasta) which are expected to be of higher quality than its input (e.g. longer N50, longer longest contigs, fewer number of contigs, high percentage match with reference, high genome fraction, etc). In Fig 1, we show an example pipeline in which FinisherSC can fit. As shown in Fig 1, FinisherSC can be readily incorporated into state-of-the-art assembly pipelines (e.g. PacBio® HGAP).

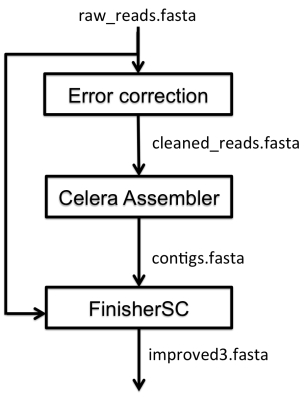

Fig. 1: Pipeline where FinisherSC can fit in

2.2 Algorithm and features

The algorithm of FinisherSC is summarized in Alg 1. Detailed description of the algorithm is in the supplementary materials. We summarize the key features of FinisherSC as follows.

- Repeat-aware: FinisherSC uses a repeat-aware rule to define overlap. It uses string graphs to capture overlap information and to handle repeats so that FinisherSC can robustly merge contigs. There is an optional component, X-phaser (Lam et al., 2014a), that can resolve long approximate repeats with two copies by using the polymorphisms between them. There is also an optional component, T-solver, that can resolve tandem repeat by using the copy count information.
- Data-efficient: FinisherSC utilizes all the raw reads to perform re-layout. This can fill gaps and improve robustness in handling repeats.
- Scalable: FinisherSC streams raw reads to identify relevant reads for re-layout and refined analysis. MUMMER (Kurtz et al., 2004) does the core of the sequence alignment. Although MUMMER is single threaded, we provide an option to segment the files and run multiple MUMMER jobs in parallel. These techniques allow FinisherSC to be easily scalable to high volume of data.

*Current affiliation: Samsung SSIC

†To whom correspondence should be addressed

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Algorithm 1: Main flow of FinisherSC

Input : contigs.fasta, raw_reads.fasta
Output: improved3.fasta

1. Filter completely embedded contigs
2. Form a string graph with the BEST successors/predecessors as edges
3. Condense the string graph by contracting edges with both in-degree and out-degree being 1
4. Use raw reads to declare potential successors/predecessors of dangling contigs
5. Merge contigs (with gaps filled by reads) when they respectively only have 1 successor/1 predecessor
6. Form a string graph with ALL successors/predecessors as edges
7. Merge contigs with only 1 predecessor or 1 successor and each has no more than two competing edges

3 RESULTS AND DISCUSSION

3.1 Experimental evaluation on bacterial genomes

We evaluated the performance of FinisherSC as follows. Raw reads were processed according to the pipeline in Fig 1. They were first error corrected and then assembled into contigs by an existing pipeline (i.e. HGAP). Contigs were upgraded using FinisherSC and evaluated for quality with Quast (Gurevich et al., 2013). The data used for assessment are real PacBio® reads. These include data recently produced at JGI and data available online supporting the HGAP publication. We compared the assembly quality of the contigs coming out from the Celera assembler (Myers et al., 2000) of HGAP pipeline, the upgraded contigs by FinisherSC and the upgraded contigs by PBJelly (English et al., 2012). A summary of the evaluation is shown in Fig 2. More details can be found in the supplementary materials. We find that FinisherSC can upgrade the assembly from HGAP without sacrifice on accuracy on these test cases. Moreover, the upgraded contigs by FinisherSC are generally of higher quality than those upgraded by PBJelly. This suggests that there is extra information from the reads that is not fully utilized by state-of-the-art assembly pipelines for long reads.

3.2 Experiments on scalability

We tested the scalability of FinisherSC by applying it to handle larger genomes. The data used are the benchmark data available on PacBio Devnet. We run FinisherSC with the option of using 20 threads (-par 20) on a server computer. The server computer is equipped with 64 cores of CPU at clock rate of 2.4-3.3GHz and 512GB of RAM. The running time is tabulated in Table 1.

3.3 Discussion

Although FinisherSC was originally designed to improve de-novo assembly by long reads, it can also be used to scaffold long contigs (formed by short reads) using long reads. For that use case, we note that the contigs formed by short reads can sometimes have length shorter than the average length of long reads. Therefore, we suggest users to filter out those short contigs before passing them into FinisherSC.

Genome name	Genome size (Mbp)	Size of reads (Gbp)	Running time (hours)
Caenorhabditis elegans	104	7.65	23
Drosophila	138	2.27	9.4
Saccharomyces cerevisiae	12.4	1.40	0.66

Table 1. Summary of running time for the experiments on scalability

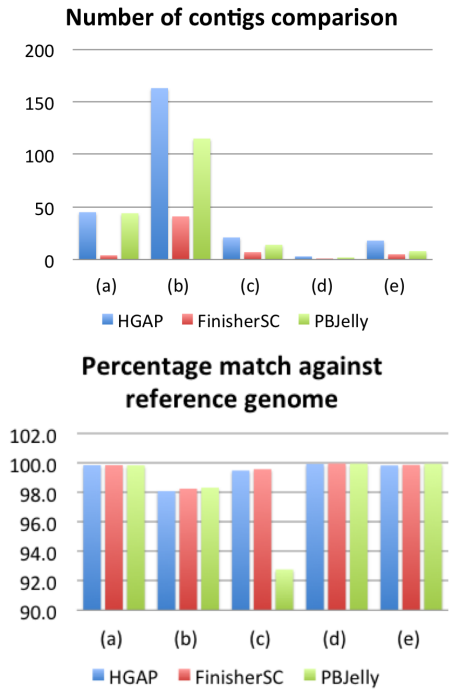

Fig. 2: Experimental evaluation on bacterial genomes. (a,b) : Pedobacter heparinus DSM 2366 (PacBio® long reads from JGI) (c, d, e) : Escherichia coli MG 1655, Meiothermus ruber DSM 1279, Pedobacter heparinus DSM 2366 (PacBio® long reads supporting the HGAP publication).

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4 SUPPLEMENTARY MATERIALS

4.1 Summary of dataflow in FinisherSC and key experimental results

We visualize in Figure 3 the flow of FinisherSC. Moreover, we summarize the key experimental results in Table 2.

4.2 Typical use cases

In this section, we describe example use cases of FinisherSC. Below are several scenarios that FinisherSC is helpful to you.

4.2.1 Low coverage data There are many reasons that you end up having low coverage reads. You may want to save chemicals, the genome may be too long, some parts of the experimental setup may just malfunction or you do not want to overwhelm the assembler with huge amount of data. In any of these situations, you want to utilize as much information from the reads as possible because of the scarcity of read data.

4.2.2 Simple setup for assemblers There are normally a lot of parameters that can be tuned for modern assemblers. It is also often not clear what parameters work best for your data. However, you do not want to waste time in repeatedly running the assembler by varying different combinations of parameters/setting. In this case, you need a tool that can efficiently and automatically improve your assemblies from the raw reads without rerunning the assembler.

4.2.3 Scaffolding You may have long contigs prepared from one library and long reads prepared from the other. In this case, you want to robustly and seamlessly combine data from two libraries through scaffolding.

4.3 Instructions on using FinisherSC

Our software, FinisherSC, is helpful for the use cases discussed above. It processes long contigs with long reads. You only need to supply the input data files and issue a one-line command as follows to perform the processing. Let us assume that mumP is the path to your MUMMER and destP is the location where the input and output files stay.

- Input : raw_reads.fasta, contigs.fasta
- Output : improved3.fasta
- Command :
  ```
  python finisherSC.py destP/ mumP/
  ```

We provide a sandbox example linked in our webpage. Besides the standard usage, there are extra options with details in our webpage.Specifically, we note that users can run FinisherSC in parallel by using the option of [-par numberOfThreads].

4.4 Detailed description of the algorithm

We adopt the terminology in Lam et al. (2014a). Random flanking region refers to the neighborhood of a repeat interior. A copy of a repeat being bridged means that some reads cover the copy into the random flanking region. Subroutine 1 removes embedded contigs that would otherwise confuse the later string graph operations. Subroutines 2, 3, 6, 7 are designed to handle repeats. Subroutines 2, 3 resolve repeats whose copies are all bridged by some reads. Subroutines 6, 7 resolve two-copies repeats of which only one copy is bridged. Subroutines 4, 5 utilize neglected information from raw reads. They define merges at locations which are not parts of any long repeats. [1]

Algorithm 2: Subroutine 1: Filter completely embedded contigs

Input : contigs.fasta
Output: noEmbed.fasta

1. Obtain alignment among contigs from contigs.fasta
2. For any (x,y) contig pair, if x is completely embedded in y, then we add x to removeList
3. Remove all contigs specified in removetList from contigs.fasta. The resultant set of contigs are outputted as noEmbed.fasta

Algorithm 3: Subroutine 2: Form a string graph with the BEST successors/predecessors as edges

Input : noEmbed.fasta
Output: String_graph_1

1. Initialize the nodes of G to be contigs from noEmbed.fasta
2. Obtain alignment among contigs from noEmbed.fasta
3. **for** *each contig x* **do**
   Find predecessor y and successor z with the largest overlap with x
   if *such y exists* **then**
   add an edge $y \rightarrow x$ to G
   if *such z exists* **then**
   add an edge $x \rightarrow z$ to G
4. Output G as String_graph_1

Algorithm 4: Subroutine 3: Condense the string graph by contracting edges with both in-degree and out-degree being 1

Input : String_graph_1, noEmbed.fasta
Output: improved1.fasta

1. **for** *each edge $u \rightarrow v$ in String_graph_1,* **do**
   if *out-deg (u) = in-deg (v) =1* **then**
   condense (u,v) into a single node and concatenate the node labels
2. **for** *each node x in the transformed String_graph_1* **do**
   output the concatenation of contigs associated with node x to be a merged contig
3. Output all the merged contigs as improved1.fasta

[1] To simplify discussion, the subroutines described are based on the assumption that reads are extracted from a single-stranded DNA. However, we remark that we have implemented FinisherSC by taking into account that reads are extracted from both forward and reverse strands.

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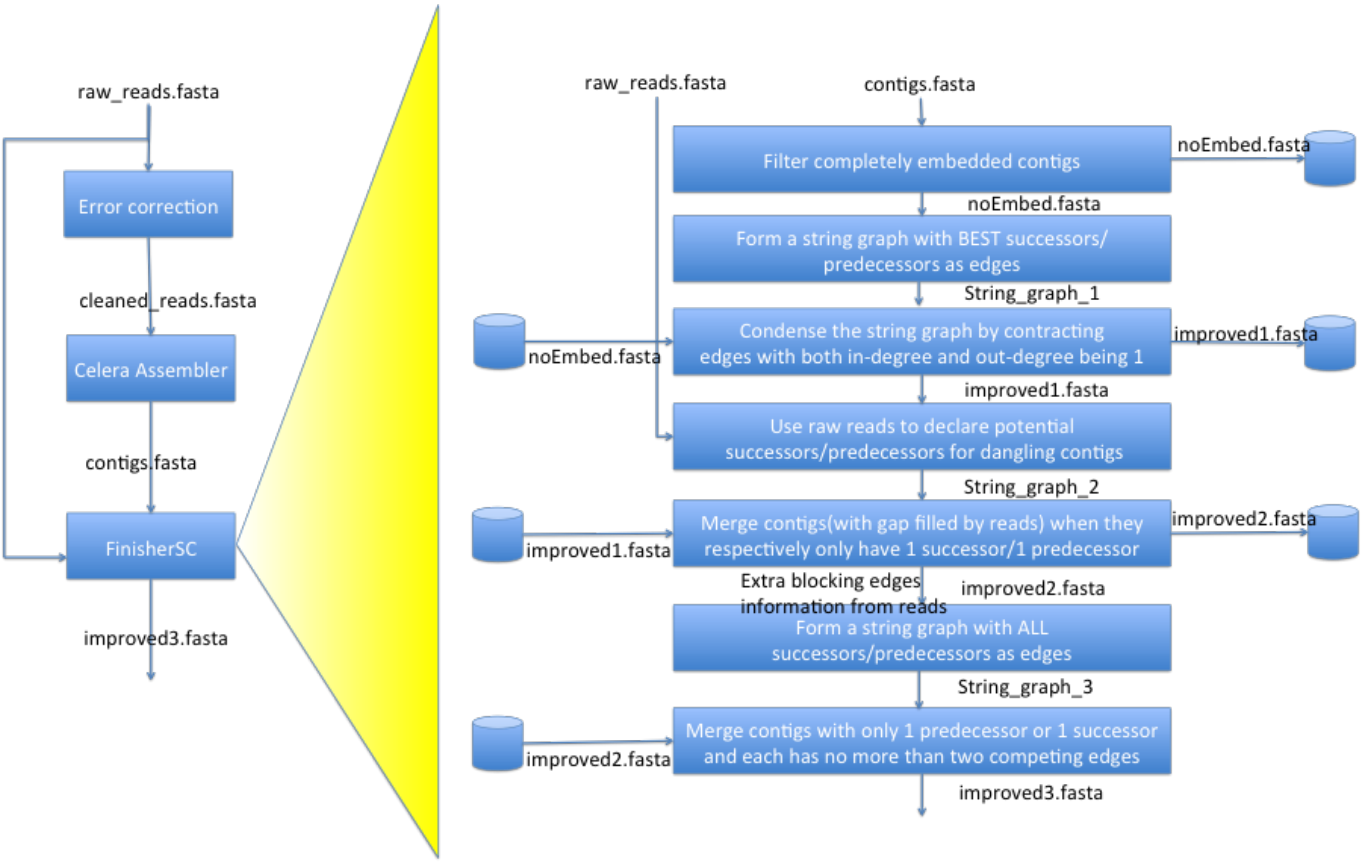

Fig. 3: Summary of data flow of FinisherSC

Genome name	(a)	(b)	(c)	(d)	(e)
Genome size	5167383	5167383	4639221	3097457	5167383
Coverage of raw reads	50	30	50	55	53.1
Coverage of corrected reads	32.93	17.74	10.1	11.3	9.5
Coverage of input to Celera	32.93	17.74	10.1	11.3	9.5
N50 of HGAP output	4097401	89239	392114	1053479	1403814
N50 of FinisherSC upgraded output	5168551	215810	1525398	3099349	2913716
N50 of PBJelly upgraded output	4099674	145441	1200847	1715191	3343452
Number of contigs of HGAP output	45	163	21	3	18
Number of contigs of FinisherSC upgraded output	4	41	7	1	5
Number of contigs of PBJelly upgraded output	44	115	14	2	8
Length of the longest contig of HGAP output	4097401	254277	1241016	1390744	2103385
Length of the longest contig of FinisherSC upgraded output	5168551	637485	2044060	3099349	2913716
Length of the longest contig of PBJelly upgraded output	4099674	495596	1958341	1715191	3343452
Percentage match of HGAP output against reference	99.87	98.11	99.51	99.96	99.85
Percentage match of FinisherSC upgraded output against reference	99.87	98.27	99.60	99.99	99.89
Percentage match of PBJelly upgraded output against reference	99.86	98.34	92.77	99.98	99.97
Total length of HGAP output	5340498	5536634	4689701	3102769	5184825
Total length of FinisherSC upgraded output	5212355	5139491	4660679	3099349	5167414
Total length of PBJelly upgraded output	5383836	5821106	4718818	3106774	5210862

Table 2. Experimental evaluation results. (a,b) : Pedobacter heparinus DSM 2366 (recent real long reads from JGI) (c, d, e) : Escherichia coli MG 1655, Meiothermus ruber DSM 1279, Pedobacter heparinus DSM 2366 (real long reads supporting the HGAP publication). Detailed analysis by Quast is shown in the supplementary material.

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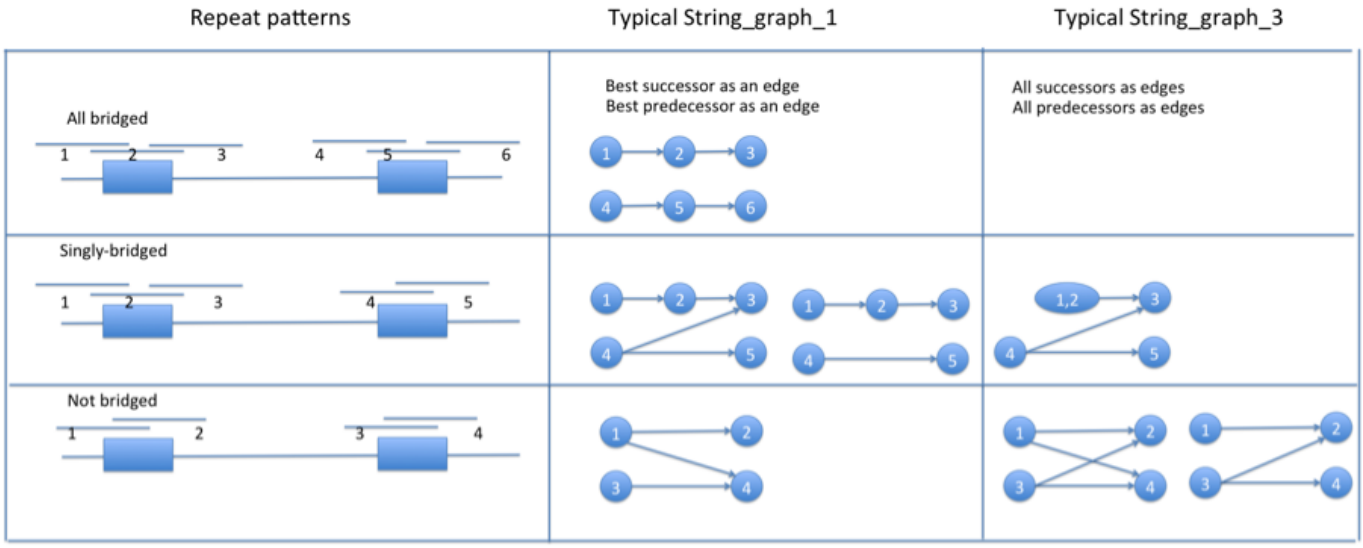

Fig. 4: Repeat patterns, typical String_graph_1, typical String_graph_3

Algorithm 5: Subroutine 4: Use raw reads to declare potential successors/predecessors of dangling contigs

Input : improved1.fasta, raw_reads.fasta
Output: String_graph_2

1. Initialize nodes of G to be contigs from improved1.fasta
2. Divide raw_reads into batches B_S
3. Stream the data in B_S.
   for $b \in B_S$ **do**
   i) align b with contigs from improved1.fasta
   ii) record the overlap information in I
4. **for** *each pair of nodes* u, v *in G* **do**
   if $u \rightarrow v$ *is a predecessor-successor pair* **then**
   Add an edge $u \rightarrow v$ to G
   if *there exists read R such that* (u, R) *and* (R, v) *are predecessor-successor pairs according to* I **then**
   Add an edge $u \rightarrow v$ to G
5. Output graph G as String_graph_2

Algorithm 6: Subroutine 5: Merge contigs (with gaps filled by reads) when they respectively only have 1 successor/1 predecessor

Input : improved1.fasta, String_graph_2
Output: improved2.fasta, connectivity_info

1. **for** *each edge* $u \rightarrow v$ *of String_graph_2* **do**
   if *out-deg (u) = in-deg (v) =1* **then**
   condense (u,v) into a single node and concatenate the node labels
2. **for** *each node in the transformed String_graph_2* **do**
   output concatenated contigs as new contigs (with reads filling the gaps) and connectivity information to connectivity_info

Algorithm 7: Subroutine 6: Form a string graph with ALL successors/predecessors as edges

Input : improved2.fasta, connectivity_info
Output: String_graph_3

1. Use connectivity_info to form a graph G with nodes from improved2.fasta. All predecessor-successor pairs are edges in G.
2. Output the corresponding graph as String_graph_3

4.5 Justification of the algorithm

4.5.1 Big picture There are two main parts of the algorithm underlying FinisherSC. They are

1. Gap filling
2. Repeat resolution

With uniform sampling assumption, the gaps are unlikely to land on the few long repeats on bacterial genomes. Therefore, subroutines 4, 5 can close most of the gaps. For repeat resolution, subroutines 1, 2, 3, 6, 7 robustly define merges using various transformations of string graphs. Detailed discussion is in the coming section.

4.5.2 Detailed justification on repeat resolution We focus the discussion on a long repeat with two copies. To simplify discussion, we further assume that each base of the genome is covered by some reads and the read length is fixed. The goal here is to correctly merge as many reads as possible in the presence of that repeat. The claim is that Subroutines 2, 3, 6, 7 collectively can achieve this goal. In the

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Algorithm 8: Subroutine 7: Merge contigs with only 1 predecessor or 1 successor and each has no more than two competing edges

Input : improved2.fasta, String_graph_3
Output: improved3.fasta

1. Traverse the String_graph_3 for pattern of $u1 \rightarrow u3$, $u2 \rightarrow u3$, $u2 \rightarrow u4$ and that out-deg (u1) =1, out-deg (u2) = 2, in-deg (u3) =2, in-deg (u4) =1, if found, then,
   a) Delete the edge $u2 \rightarrow u3$
   b) Condense the graph
   c) Continue until the whole graph is traversed
2. Output the merged contigs as improved3.fasta

case of one repeat, we only need consider the reads either bridging the repeat copies/ reads at the interior of repeats/ touching the repeat copies of that repeat. We separate the discussion on each of the cases depicted in the rows of Fig 4. They are listed as follows.

1. Both copies are bridged
2. Only one copy is bridged
3. Both copies are not bridged

Let us first clarify some terminologies before proceeding. A read y is called a successor of another read x if they have significant head-to-tail overlap in the order of $x \rightarrow y$. y is called the best successor of x if the overlap length is the largest among all the successors of x. y is called the true immediate successor of x if y is the closest read to x's right side in the sequencing process. Similarly, we can also define the corresponding notion for predecessors.

In the first case, without loss of generality, let us consider any read R emerging from the left flanking region of the left copy. It will get merged with its best successor when condensing String_graph_1. Moreover, the best successor is also the true immediate successor. It is because reads from the other copy of the repeat either have smaller overlap length or are not successors.

Now, let us move to the second case. Since there is a bridging read, there are no reads completely embedded in the interior of the repeat. Without loss of generality, we consider the case that the left copy is bridged and the right copy is not. Now we label R2 as the bridging read, R1/R3 respectively as the true immediate predecessor/successor of the bridging read, R4/R5 as the most penetrating reads into the second copy of the repeat. For all other reads, they get merged with their true immediate successors/predecessors when condensing in String_graph_1. For the remaining five items of interest, the main question is whether there is an edge between R4 and R5 in String_graph_1 (i.e. whether the best successor of R4 is R3). If not, then condensing in String_graph_1 will merge R4 with R5, which is the true immediate successor. If such an edge exists, then we end up with the pattern shown in Fig 4 for String_graph_3. This means that only R1 is merged to R2 when condensing String_graph_1. However, given the existence of the Z-shape pattern, graph operations on String_graph_3, the subroutine 7 will merge R2 and R3, and also will merge R4 and R5.

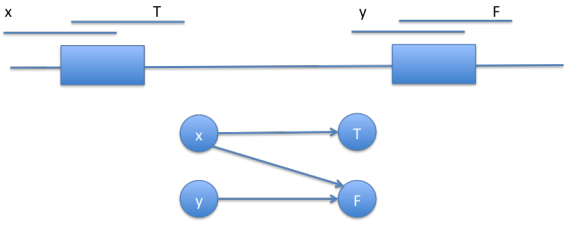

Fig. 5: Z-pattern in string graph

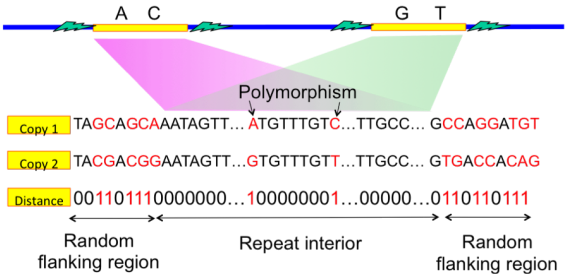

Fig. 6: Approximate repeat with two copies

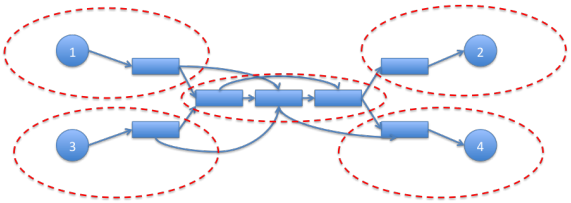

Fig. 7: Using string graph to define repeat interiors and flanking regions

Finally, consider the third case, when both repeat copies are not bridged. For reads that are not closest to the repeat copies, they get merged correctly when condensing String_graph_1. Without loss of generality, we consider a read x closest to the left flanking region of the left copy of the repeat. An illustration of this situation in String_graph_1 is shown in Fig 5. Let its true immediate successor be T. We are going to show that it will not get merged with the wrong read in String_graph_1 through a proof by contradiction. If x got merged with some wrong F, then $x \rightarrow F$ would be an edge. Let y be the read closest the left flanking region of the right copy of the repeat. Then, $y \rightarrow F$ is also an edge. Therefore, there should be no merges of $x \rightarrow F$, which results in contradiction. Now we consider String_graph_3, if x has only 1 successor, then it should be T. Otherwise, it is connected to both T and some F. Then, we consider the y coming from the left flanking region of the right copy. There must be an edge from y to F. If there is also an edge from y to T, then both x and y are not merged in String_graph_3. However, if there is no edge from y to T, then x is merged with T and y with F correctly.

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Fig. 8: Pattern of back-to-back tandem repeat

Fig. 9: Pattern of string graph that corresponds to a typical back-to-back tandem repeat

4.6 Approximate repeat phasing option

FinisherSC provides an optional component, X-phaser to resolve approximate repeat with two copies, which cannot be bridged by any reads. An example of such an approximate repeat is shown in Fig 6. The algorithm behind X-phaser involves two main parts.

1. Identify repeat interior and its flanking regions
2. Merge contigs by phasing the polymorphisms within the repeat

Algorithm 9 achieves the first part by performing various operations on a string graph. The nodes of the string graph are either contigs or reads near the end points of the contigs. An illustration of a typical string graph is shown in Fig 7. The contigs are indicated by solid circles and reads are indicated by rectangles. The dotted circles specify the random flanking region and repeat interior that we want to infer through Algorithm 9. The X-phasing step in Lam et al. (2014a) achieves the second part. It utilizes the polymorphisms within the repeat interior to help distinguish the repeat copies. Interested readers may refer to Xphased-Multibridging in Lam et al. (2014a) for more details. In FinisherSC, we use the implementation of the Xphasing step in Lam et al. (2014a).

4.7 Tandem repeat resolution option

The optional tandem repeat resolution step can resolve back-to-back tandem repeat. A back-to-back tandem repeat refers to repeat whose copies directly follow one another. An example is given in Fig 8. FinisherSC provides a component, T-solver, to resolve that. The key idea is to detect cycles in the string graph that join the reads and contigs. An illustration of such a graph is shown in Fig 9, where the circles are contigs and rectangles are reads associated with the end points of the contigs. The algorithm behind T-solver is in Alg 10.

4.8 Detailed experimental results on bacterial genomes

In this section, we provide the detailed Quast report for the results described in Table 2. Moreover, we compare in Fig 10 the memory consumption and running time of FinisherSC with those of PBJelly. The computing experiments for this section were performed on the genepool cluster at JGI. Below are commands used to run PBJelly.

```
Jelly.py setup Protocol.xml -x "--minGap=1"
Jelly.py mapping Protocol.xml
Jelly.py support Protocol.xml -x " --debug"
Jelly.py extraction Protocol.xml
Jelly.py assembly Protocol.xml -x "--nproc=16"
Jelly.py output Protocol.xml
```

The BLASR configuration in Protocol.xml is

```
-minMatch 8 -minPctIdentity 70 -bestn 8
-nCandidates 30 -maxScore -500 -nproc 16
-noSplitSubreads
```

Algorithm 9: Repeat phasing option

Input : improved3.fasta, raw_read.fasta
Output: improved4.fasta

1. Stream reads to identify reads that are associated with end points of contigs
2. Form a 2-color graph with black nodes as reads and red nodes as contigs. Name it as G
3. **for** *each red node in G* **do**
   Perform graph search to determine other red nodes that are reachable by it through path consisting of black nodes only
4. Form a bipartite graph $B = (B_L, B_R)$ with nodes on left/right side representing left/right ends of the contigs. An edge $L \rightarrow R$ exists, where $L \in B_L, R \in B_R$, if there exists a path of black nodes in G such that R is reachable from L
5. Find connected components in B
6. **for** *each connected component at* $b \in B$ **do**
   if $|b \cap B_L| = 2$ *and* $|b \cap B_R| = 2$ **then**
   define b as a two-copies repeat
7. **for** *each two-copies repeat[with associated nodes (L1,L2; R1, R2)]* **do**
   a) Go back to graph G, and label nodes reachable from each of L1/L2 and to each of R1/R2 through black paths.
   b) For each black node in G connected to all L1/L2/R1/R2, name it as inside nodes. If it only misses one of the four, then label it as miss_x node where x is the missed item
   c) Define start node S as an inside node connected to some miss_L1 nodes and miss_L2 nodes. Similarly, define end node E as an inside node connected to some miss_R1 nodes and miss_R2 nodes
   d) Define repeat interior and flanking region
   i) Find a black path between S and E and label it as a repeat path (this is the repeat interior)
   ii) Find paths from L1 to S, L2 to S , E to R1, E to R2 respectively (these are the flanking regions)
   e) For each black node involved with this repeat,
   i) If it is connected to nodes on paths L1 to S, add it to L1 to S read set. Similarly, do it for L2 to S, E to R1 and E to R2 read sets
   ii) If it is connected to inside nodes only, then add it to repeat read set
   iii) Use the separated read sets to perform repeat phasing as described in Lam et al. (2014a)
   iv) Declare merging of contigs based on the phasing results

4.9 Details of the scalability experiments

We run the scalability experiments on a server computer, which is equipped with 64 cores of CPU at clock rate of 2.4-3.3GHz and 512GB of RAM. We also note that, for even larger contig or read

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data with genomes of higher repeat content, one may be interested in the following options. They are [-f True] for fast alignment and [-l True] for breaking down large contig file. As a reference, we also attach the Quast analysis results on all the intermediate output for the scalability test in Table 8, 9 and 10. We note that the misassembly count in the Quast analysis for these genomes should only be used as a reference because there is a lack of high quality reference and reference genomes may be from different strains.

5 ACKNOWLEDGEMENT

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Algorithm 10: Tandem repeat resolution option

Input : improved3.fasta, raw_read.fasta
Output: tandem_resolved.fasta

1. Classify contigs into two clusters in which one contains long contigs (S_L) and the other contains short contigs(S_S). We only focus on the treatment on S_L.
2. Form a 2-color read-contig string graph G with contigs from S_L and raw reads associated to the endpoints of these contigs. This step is similar to step 2 in Alg 9
3. **repeat**
   a) Identify local graph pattern in G consisting of 1 incoming contig, 1 outgoing contig and cycles linking intermediate reads. Declare that as a tandem repeat target t.
   b) For such target t, find the repetitive pattern of the tandem repeat through finding a cycle on the intermediate reads.
   c) Estimate the copy count of the repetitive pattern by using overall coverage and amount of reads associated with t.
   d) Connect the contigs associated with t with appropriate tandem repeat copies filled in.
   until *there is no back-to-back tandem repeat pattern detected in G*;

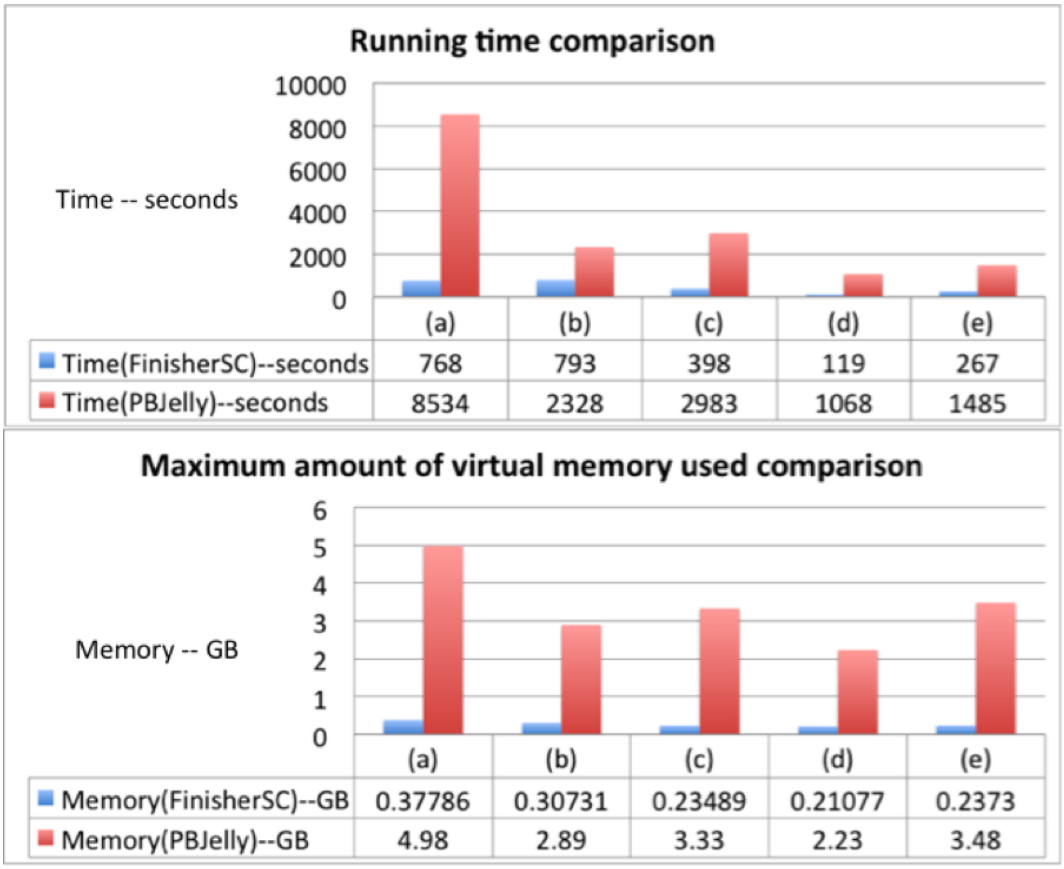

Fig. 10: Running time and memory consumption comparison of FinisherSC and PBJelly . (a) to (e) are the corresponding data sets in Table 2.

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Table 3. (a) in Table 2. All statistics are based on contigs of size ≥ 500 bp, unless otherwise noted (e.g., "# contigs (≥ 0 bp)" and "Total length (≥ 0 bp)" include all contigs).

Assembly	HGAP	FinisherSC	PBJelly
# contigs (≥ 0 bp)	45	**4**	44
# contigs (≥ 1000 bp)	45	**4**	44
Total length (≥ 0 bp)	5340498	5212355	**5383836**
Total length (≥ 1000 bp)	5340498	5212355	**5383836**
# contigs	45	**4**	44
Largest contig	4097401	**5168551**	4099674
Total length	5340498	5212355	**5383836**
Reference length	5167383	5167383	5167383
GC (%)	42.16	42.06	42.19
Reference GC (%)	42.05	42.05	42.05
N50	4097401	**5168551**	4099674
NG50	4097401	**5168551**	4099674
N75	4097401	**5168551**	4099674
NG75	4097401	**5168551**	4099674
L50	1	1	1
LG50	1	1	1
L75	1	1	1
LG75	1	1	1
# misassemblies	**1**	**1**	3
# misassembled contigs	**1**	**1**	2
Misassembled contigs length	**9679**	**9679**	4117533
# local misassemblies	**2**	3	4
# unaligned contigs	39 + 0 part	**1 + 0 part**	39 + 0 part
Unaligned length	135514	**17453**	163702
Genome fraction (%)	100.000	100.000	100.000
Duplication ratio	1.007	**1.005**	1.010
# N's per 100 kbp	4.25	**0.48**	4.46
# mismatches per 100 kbp	9.56	**1.30**	10.14
# indels per 100 kbp	92.62	**54.90**	94.75
Largest alignment	4097400	**5168480**	4098915
NA50	4097400	**5168480**	4098915
NGA50	4097400	**5168480**	4098915
NA75	4097400	**5168480**	4098915
NGA75	4097400	**5168480**	4098915
LA50	1	1	1
LGA50	1	1	1
LA75	1	1	1
LGA75	1	1	1

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Table 4. (b) in Table 2. All statistics are based on contigs of size ≥ 500 bp, unless otherwise noted (e.g., "# contigs (≥ 0 bp)" and "Total length (≥ 0 bp)" include all contigs).

Assembly	HGAP	FinisherSC	PBJelly
# contigs (≥ 0 bp)	163	**41**	115
# contigs (≥ 1000 bp)	163	**41**	115
Total length (≥ 0 bp)	5536634	5139491	**5821106**
Total length (≥ 1000 bp)	5536634	5139491	**5821106**
# contigs	163	**41**	115
Largest contig	254277	**637485**	495596
Total length	5536634	5139491	**5821106**
Reference length	5167383	5167383	5167383
GC (%)	41.98	41.96	42.01
Reference GC (%)	42.05	42.05	42.05
N50	89239	**215810**	145441
NG50	94672	**215810**	161517
N75	44568	**117879**	98297
NG75	53723	**117879**	116800
L50	20	**9**	14
LG50	18	**9**	12
L75	42	**17**	26
LG75	36	**17**	21
# misassemblies	**0**	**0**	12
# misassembled contigs	**0**	**0**	10
Misassembled contigs length	**0**	**0**	439591
# local misassemblies	**0**	3	3
# unaligned contigs	46 + 1 part	**1 + 0 part**	43 + 22 part
Unaligned length	200727	**11862**	302804
Genome fraction (%)	98.727	98.957	**99.964**
Duplication ratio	1.046	**1.003**	1.068
# N's per 100 kbp	15.64	**6.17**	9.50
# mismatches per 100 kbp	**63.00**	67.78	68.92
# indels per 100 kbp	**577.98**	589.72	597.99
Largest alignment	254274	**637485**	495589
NA50	89239	**215810**	145441
NGA50	94672	**215810**	161490
NA75	44567	**117879**	98293
NGA75	50860	**117879**	115834
LA50	20	**9**	14
LGA50	18	**9**	12
LA75	42	**17**	26
LGA75	36	**17**	21

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Table 5. (c) in Table 2. All statistics are based on contigs of size ≥ 500 bp, unless otherwise noted (e.g., "# contigs (≥ 0 bp)" and "Total length (≥ 0 bp)" include all contigs).

Assembly	HGAP	FinisherSC	PBJelly
# contigs (≥ 0 bp)	21	**7**	14
# contigs (≥ 1000 bp)	21	**7**	14
Total length (≥ 0 bp)	4689701	4660679	**4718818**
Total length (≥ 1000 bp)	4689701	4660679	**4718818**
# contigs	21	**7**	14
Largest contig	1241016	**2044060**	1958341
Total length	4689701	4660679	**4718818**
Reference length	4639221	4639221	4639221
GC (%)	50.87	50.85	50.85
Reference GC (%)	50.79	50.79	50.79
N50	392114	**1525398**	1200847
NG50	392114	**1525398**	1200847
N75	252384	**1525398**	275618
NG75	252384	**1525398**	321636
L50	3	**2**	**2**
LG50	3	**2**	**2**
L75	7	**2**	4
LG75	7	**2**	3
# misassemblies	**8**	**8**	12
# misassembled contigs	4	**3**	5
Misassembled contigs length	**2530799**	3584781	3672462
# local misassemblies	**3**	**3**	4
# unaligned contigs	0 + 1 part	**0 + 0 part**	0 + 3 part
Unaligned length	205	**0**	2605
Genome fraction (%)	99.583	99.656	**99.689**
Duplication ratio	1.015	**1.008**	1.021
# N's per 100 kbp	0.00	0.00	0.00
# mismatches per 100 kbp	**3.66**	4.61	4.11
# indels per 100 kbp	**8.36**	13.82	10.75
Largest alignment	683967	**1094192**	949307
NA50	339478	**860437**	685586
NGA50	339478	**860437**	685586
NA75	229039	**378942**	255377
NGA75	229039	**378942**	255377
LA50	5	**3**	**3**
LGA50	5	**3**	**3**
LA75	9	**5**	7
LGA75	9	**5**	7

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Table 6. (d) in Table 2. All statistics are based on contigs of size ≥ 500 bp, unless otherwise noted (e.g., "# contigs (≥ 0 bp)" and "Total length (≥ 0 bp)" include all contigs).

Assembly	HGAP	FinisherSC	PBJelly
# contigs (≥ 0 bp)	3	**1**	2
# contigs (≥ 1000 bp)	3	**1**	2
Total length (≥ 0 bp)	3102769	3099349	**3106774**
Total length (≥ 1000 bp)	3102769	3099349	**3106774**
# contigs	3	**1**	2
Largest contig	1390744	**3099349**	1715191
Total length	3102769	3099349	**3106774**
Reference length	3097457	3097457	3097457
GC (%)	63.38	63.39	63.38
Reference GC (%)	63.38	63.38	63.38
N50	1053479	**3099349**	1715191
NG50	1053479	**3099349**	1715191
N75	1053479	**3099349**	1391583
NG75	1053479	**3099349**	1391583
L50	2	**1**	**1**
LG50	2	**1**	**1**
L75	2	**1**	2
LG75	2	**1**	2
# misassemblies	0	0	0
# misassembled contigs	0	0	0
Misassembled contigs length	0	0	0
# local misassemblies	2	2	2
# unaligned contigs	0 + 0 part	0 + 0 part	0 + 0 part
Unaligned length	0	0	0
Genome fraction (%)	99.966	**99.986**	**99.986**
Duplication ratio	1.002	**1.001**	1.003
# N's per 100 kbp	0.00	0.00	0.00
# mismatches per 100 kbp	**0.03**	0.26	0.10
# indels per 100 kbp	**1.10**	3.87	1.32
Largest alignment	1390744	**3099004**	1713236
NA50	1053134	**3099004**	1713236
NGA50	1053134	**3099004**	1713236
NA75	1053134	**3099004**	1391558
NGA75	1053134	**3099004**	1391558
LA50	2	**1**	**1**
LGA50	2	**1**	**1**
LA75	2	**1**	2
LGA75	2	**1**	2

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Table 7. (e) in Table 2. All statistics are based on contigs of size ≥ 500 bp, unless otherwise noted (e.g., "# contigs (≥ 0 bp)" and "Total length (≥ 0 bp)" include all contigs).

Assembly	HGAP	FinisherSC	PBJelly
# contigs (≥ 0 bp)	18	**5**	8
# contigs (≥ 1000 bp)	18	**5**	8
Total length (≥ 0 bp)	5184825	5167414	**5210862**
Total length (≥ 1000 bp)	5184825	5167414	**5210862**
# contigs	18	**5**	8
Largest contig	2103385	2913716	**3343452**
Total length	5184825	5167414	**5210862**
Reference length	5167383	5167383	5167383
GC (%)	42.05	42.05	42.07
Reference GC (%)	42.05	42.05	42.05
N50	1403814	2913716	**3343452**
NG50	1403814	2913716	**3343452**
N75	790287	**2225895**	1814491
NG75	790287	**2225895**	1814491
L50	2	**1**	**1**
LG50	2	**1**	**1**
L75	3	**2**	**2**
LG75	3	**2**	**2**
# misassemblies	**1**	**1**	2
# misassembled contigs	**1**	**1**	2
Misassembled contigs length	**1403814**	2913716	1820739
# local misassemblies	**0**	**0**	1
# unaligned contigs	**0 + 0 part**	**0 + 0 part**	0 + 3 part
Unaligned length	**0**	**0**	13698
Genome fraction (%)	99.900	99.934	**99.954**
Duplication ratio	1.005	**1.001**	1.007
# N's per 100 kbp	0.00	0.00	0.00
# mismatches per 100 kbp	3.41	3.56	**2.28**
# indels per 100 kbp	**2.91**	5.31	5.21
Largest alignment	2103385	2225895	**3343452**
NA50	1259090	1656831	**3343452**
NGA50	1259090	1656831	**3343452**
NA75	790287	**1656831**	1270970
NGA75	790287	**1656831**	1270970
LA50	2	2	**1**
LGA50	2	2	**1**
LA75	3	**2**	**2**
LGA75	3	**2**	**2**

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Table 8. Quast analysis report of Caenorhabditis elegans

Assembly	contigs	noEmbed	improved	improved2	improved3
# contigs ($\geq$ 0 bp)	245	237	207	200	**196**
# contigs ($\geq$ 1000 bp)	245	237	207	200	**196**
Total length ($\geq$ 0 bp)	**104169699**	103975584	103804734	103826835	103822380
Total length ($\geq$ 1000 bp)	**104169699**	103975584	103804734	103826835	103822380
# contigs	245	237	207	200	**196**
Largest contig	3165643	3165643	**4217234**	**4217234**	**4217234**
Total length	**104169699**	103975584	103804734	103826835	103822380
Reference length	100286401	100286401	100286401	100286401	100286401
GC (%)	35.67	35.66	35.66	35.66	35.66
Reference GC (%)	35.44	35.44	35.44	35.44	35.44
N50	1613475	1613475	1773224	**1832604**	**1832604**
NG50	1619480	1619480	1832604	**1910174**	**1910174**
N75	834223	834223	947068	**959597**	**959597**
NG75	881109	881109	959597	**1031901**	**1031901**
L50	24	24	22	**21**	**21**
LG50	23	23	21	**20**	**20**
L75	48	48	43	**41**	**41**
LG75	44	44	40	**38**	**38**
# misassemblies	1358	**1334**	1411	1407	1403
# misassembled contigs	127	120	108	104	**102**
Misassembled contigs length	96744440	**96571278**	98809318	99454743	99479049
# local misassemblies	784	**783**	809	805	810
# unaligned contigs	70 + 20 part	70 + 20 part	70 + 11 part	68 + 11 part	**68 + 10 part**
Unaligned length	1862487	1862487	1743676	1745585	**1743404**
Genome fraction (%)	99.525	99.525	99.531	**99.535**	**99.535**
Duplication ratio	1.027	1.025	**1.024**	**1.024**	**1.024**
# N's per 100 kbp	0.00	0.00	0.00	0.00	0.00
# mismatches per 100 kbp	13.27	13.27	13.23	13.20	**13.13**
# indels per 100 kbp	20.89	20.89	**20.63**	20.80	20.86
Largest alignment	1362562	1362562	1362562	1362562	1362562
NA50	407833	407833	**407888**	402635	402635
NGA50	**421272**	**421272**	**421272**	412370	412370
NA75	213728	**217528**	213728	211961	211961
NGA75	**232043**	**232043**	229862	226413	226413
LA50	85	85	85	85	85
LGA50	80	80	80	80	80
LA75	169	**168**	**168**	169	169
LGA75	**156**	**156**	**156**	157	157

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Table 9. Quast analysis report of Drosophila

Assembly	contigs	noEmbed	improved	improved2	improved3
# contigs ($\geq$ 0 bp)	128	128	110	96	**93**
# contigs ($\geq$ 1000 bp)	128	128	110	96	**93**
Total length ($\geq$ 0 bp)	**138490501**	**138490501**	138082066	138131721	138096303
Total length ($\geq$ 1000 bp)	**138490501**	**138490501**	138082066	138131721	138096303
# contigs	128	128	110	96	**93**
Largest contig	24648237	24648237	**27967410**	**27967410**	**27967410**
Total length	**138490501**	**138490501**	138082066	138131721	138096303
Reference length	168717020	168717020	168717020	168717020	168717020
GC (%)	41.86	41.86	41.87	41.87	41.87
Reference GC (%)	41.74	41.74	41.74	41.74	41.74
N50	15305620	15305620	**21710673**	**21710673**	**21710673**
NG50	6168915	6168915	**15305620**	**15305620**	**15305620**
N75	920983	920983	1012145	**1448033**	**1448033**
NG75	291391	291391	308285	389196	**402690**
L50	4	4	**3**	**3**	**3**
LG50	6	6	**4**	**4**	**4**
L75	15	15	11	**10**	**10**
LG75	57	57	50	42	**41**
# misassemblies	8272	8272	**8057**	8071	8064
# misassembled contigs	98	98	84	76	**73**
Misassembled contigs length	**127872011**	**127872011**	130423069	130352009	130316591
# local misassemblies	290	290	**282**	283	284
# unaligned contigs	1 + 0 part	1 + 0 part	1 + 0 part	1 + 0 part	1 + 0 part
Unaligned length	61383	61383	61383	61383	61383
Genome fraction (%)	**76.376**	**76.376**	76.348	76.367	76.364
Duplication ratio	1.082	1.082	**1.079**	**1.079**	**1.079**
# N's per 100 kbp	0.00	0.00	0.00	0.00	0.00
# mismatches per 100 kbp	21.85	21.85	**21.34**	21.81	21.84
# indels per 100 kbp	20.88	20.88	**20.81**	20.94	20.99
Largest alignment	6494798	6494798	6494798	6494798	6494798
NA50	1316520	1316520	1316520	**1377072**	**1377072**
NGA50	820949	820949	821097	**829069**	**829069**
NA75	308622	308622	341648	**398564**	**398564**
NGA75	**6441**	**6441**	6389	6414	6384
LA50	26	26	26	26	26
LGA50	41	41	41	**40**	**40**
LA75	78	78	76	**72**	**72**
LGA75	532	532	533	**520**	522

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Table 10. Quast analysis report of Saccharomyces cerevisiae

Assembly	contigs	noEmbed	improved	improved2	improved3
# contigs ($\geq$ 0 bp)	30	27	23	22	**20**
# contigs ($\geq$ 1000 bp)	30	27	23	22	**20**
Total length ($\geq$ 0 bp)	**12370681**	12295446	12221471	12225040	12220764
Total length ($\geq$ 1000 bp)	**12370681**	12295446	12221471	12225040	12220764
# contigs	30	27	23	22	**20**
Largest contig	1538192	1538192	1538192	1538192	1538192
Total length	**12370681**	12295446	12221471	12225040	12220764
Reference length	12157105	12157105	12157105	12157105	12157105
GC (%)	38.21	38.18	38.17	38.17	38.17
Reference GC (%)	38.15	38.15	38.15	38.15	38.15
N50	777787	777787	777787	777787	777787
NG50	777787	777787	777787	777787	777787
N75	544615	544615	544615	544615	**583193**
NG75	544615	544615	544615	544615	**583193**
L50	6	6	6	6	6
LG50	6	6	6	6	6
L75	11	11	11	11	11
LG75	11	11	11	11	11
# misassemblies	112	109	**106**	**106**	107
# misassembled contigs	27	24	20	20	**18**
Misassembled contigs length	11049515	10974280	10900305	10900305	**10896029**
# local misassemblies	21	21	21	21	21
# unaligned contigs	0 + 0 part	0 + 0 part	0 + 0 part	0 + 0 part	0 + 0 part
Unaligned length	0	0	0	0	0
Genome fraction (%)	98.117	98.117	98.215	**98.243**	**98.243**
Duplication ratio	1.038	1.032	**1.024**	**1.024**	**1.024**
# N's per 100 kbp	0.00	0.00	0.00	0.00	0.00
# mismatches per 100 kbp	**76.68**	**76.68**	78.86	79.14	78.05
# indels per 100 kbp	11.71	11.71	**11.67**	13.58	13.50
Largest alignment	1027016	1027016	1027016	**1053518**	**1053518**
NA50	**377112**	**377112**	377016	377016	377016
NGA50	**377112**	**377112**	377016	377016	377016
NA75	181420	**198393**	**198393**	**198393**	**198393**
NGA75	198393	198393	198393	198393	198393
LA50	11	11	11	11	11
LGA50	11	11	11	11	11
LA75	23	**22**	**22**	**22**	**22**
LGA75	22	22	22	22	22