

The origin and evolution of maize in the American Southwest

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Maize offers an ideal system through which to demonstrate the potential of ancient population genomic techniques for reconstructing the evolution and spread of domesticates. The diffusion of maize from Mexico into the North American Southwest (SW) remains contentious with the available evidence being restricted to morphological studies of ancient maize plant material. We captured 1 Mb of nuclear DNA from 32 archaeological maize samples spanning 6000 years and compared them with modern landraces including those from the Mexican West coast and highlands. We found that the initial diffusion of domesticated maize into the SW is likely to have occurred through a highland route. However, by 2000 years ago a Pacific coastal corridor was also being used. Furthermore, we could distinguish between genes that were selected for early during domestication (such as *zgf1* involved in shattering) from genes that changed in the SW context (e.g. related to sugar content and adaptation to drought) likely as a response to the local arid environment and new cultural uses of maize.

Maize was domesticated from the wild grass teosinte in southern Mexico more than 6000 years ago¹⁻³. One of the key stepping stones in maize's spread and evolution was its entry into the area today encompassed by the United States (USA) via the SW around 4100 years before present⁴. In the SW, maize adapted to the local arid environment and to new cultural uses⁴, with changes reflected in evolving cob morphology. We have used genetic information from archaeological SW maize, together with that of ancient maize samples from Chile and Mexico (including the third oldest maize macrofossil ever dated, ca. cal. 5910 BP, from Tehuacan), to investigate the route of maize diffusion into the SW, and to identify genes presenting evidence of strong directional selection by farming societies of this region.

Using a hybridization-target capture approach⁵ we generated sequence data spanning predominantly the exons of 348 genes (selection criteria in⁶; Supplementary Tables 8 and 9) in i) 3 ancient Mexican samples dating to ca. cal. 5910 BP, 5280 BP and 1410 BP, ii) 25 temporally distributed ancient SW maize samples dating to ca. cal. 4000-3000, 2000 and 750

BP (SW3K, SW2K and SW750 hereafter), and iii) 4 ancient Chilean samples dating to less than 1500 BP (Table 1). Additional sequence data from a previous study ⁵ was added to the dataset (one ancient sample from Mexico, Supplementary Table 7).

The sequence data display typical ancient DNA (aDNA) damage patterns (Figs. S1 and S2). Error rates for the SW750 and SW2K individuals were below 0.2% per base after removal of C to T and G to A transitions (potentially resulting from aDNA damage; Supplementary Fig. 3). Comparisons were made with sequence data from modern maize landraces and the wild teosintes *Zea mays* ssp. *parviglumis* (henceforth *parviglumis*, the grass from which maize was domesticated) and *Z. m.* ssp. *mexicana* (henceforth *mexicana*) from maize HapMap2 ⁷ and shotgun data from an open-pollinated Mexican highland individual ⁶ (Supplementary Fig. 4 and Supplementary Tables 4 and 5).

The origin of maize in the SW has long remained contentious. For many years the only available archaeological samples of maize were from higher altitude sites, suggesting a high elevation Sierra Madre Occidental corridor of diffusion ^{8,9}. More recently discovered samples from lower elevation river valleys ^{10,11} and the morphological similarities between archaeological samples and extant Mexican landraces ^{12,13}, however, suggest diffusion along a low-elevation Pacific coastal route. Results from four different analytical approaches indicate that while maize originally entered the SW through a highland route, it subsequently received genetic contributions arriving via the Pacific coastal corridor by 2000 BP: i) Treemix results using ancient and extant samples show gene flow from the Pacific coast Chapalote landrace into the SW2K maize (Fig. 1a); ii) admixture analysis ^{6,14} of ancient and extant samples for k=5 groups shows evidence for an increase in admixture with coastal lowland maize by 2000 BP (Fig.1B, Supplementary Fig. 5), iii) explicit comparison of the different chronological SW groups using the D-statistics ¹⁵ with ancient and extant samples provides strong support for significantly more shared derived variants between SW individuals and the Pacific coast Mexican lowland

race Chapalote than between those SW individuals and the highland landrace Palomero de Jalisco, except for the SW3K group (Fig. 1c), and iv) additional analyses using a panel of 2310 modern maize landraces and teosinte shows that all of SW maize landraces (across elevations) share ancestry with landraces from highland Mexico, highland Guatemala and, to a lesser extent, with mexicana (Supplementary Fig. 6). The admixture analysis (Fig. 1b, Supplementary Fig. 5) also shows evidence of teosinte admixture in all ancient SW maize. Teosinte has no history of occurrence in the SW and was likely absent during the timeframe of the introduction of maize¹⁶. Instead, admixture with teosinte almost certainly predated the introduction of maize into the SW, and is evident in the oldest Mexican samples sequenced (Fig. 1b, Supplementary Fig. 5). Furthermore, we interpret the lack of observed admixture with teosinte or Mexican maize in the extant SW Santo Domingo landrace (USA17) to be a result of recent extensive gene flow with other American landraces (Supplementary Fig. 5).

The SW2K and SW750 sample sets present several features that allowed us to identify the targets and timing of prehistoric selection: i) the error rate for these individuals was below 0.2% per base, ii) they correspond to distinct occupations of the same cave site with sample sizes ranging from 10 (SW2K) to 12 (SW750) individuals, which allowed us to calculate various population summary statistics such as F_{ST} and Tajima's D to time the onset of early genetic changes^{17,18}, iii) the median F_{ST} across all captured genes was 3%, indicating low differentiation. We used the population branch statistic, PBS¹⁹, to detect genes that were likely early targets of maize domestication, potentially before arriving in the SW, as they present the highest dissimilarity between parviglumis (HapMap2 individuals, Supplementary Table 4) and ancient SW landraces (Fig. 2a). Seven of the top 10 genes (Supplementary Table 1) are located in regions that were detected as being under selection in an unrelated study by Hufford *et al*²⁰ (the function of the other 3 genes is not well characterized). The top gene, *zag1*, corresponds to a MADS-box transcription factor associated with shattering, a key domestication feature strongly selected for by human harvesting^{21–23}. The other genes are also well known from their

roles in domestication traits: i) *ba1* has a major role in the architecture of maize ²⁴, ii) *zcn1* and *ZmG1* are associated with the regulation of flowering ^{23,25}, and iii) *tga1* controls the change from encased to exposed kernels ²⁶. To provide further evidence of selection we calculated Tajima's D for each gene ²⁷. The median D for the SW750 population is more positive than that of the SW2K population (Supplementary Fig. 8 and Supplementary Table 2), consistent with reduced estimates of effective population size (Supplementary Fig. 9), but many domestication-related genes show the opposite trend, suggesting the influence of recent strong selection (Fig. 2a).

We have also identified a number of candidate loci that appear to have been targeted by selection starting sometime between 2000 and 750 years BP in the context of the SW. The top PBS outlier for SW750 is a dehydration-responsive element-binding protein shown to be up-regulated as much as 50-fold in maize roots under drought conditions ²⁸, perhaps a signature of adaptation to arid SW conditions (Supplementary Fig. 10). Furthermore, *ae1* shows reduced diversity in both the SW2K and SW750 populations, consistent with early selection (Fig. 3). Mutations in both *sugary1* locus (*su1*) and *ae1* (Fig. 3) affect the structure of amylopectin ²⁹, which is involved in the pasting properties of maize tortillas and porridge ³⁰. While analysis of individual ancient samples suggests the presence of modern maize haplotypes at the *su1* as early as 4400 BP ¹⁷, our data show a substantial (61%) reduction of diversity at this locus only after 2000 BP (Fig. 3), a signature that correlates with a shift toward larger cobs and floury kernel endosperm ¹¹ in archaeological maize around AD 800-1000. The *su1* mutation with the highest allele frequency difference between SW and modern individuals is known to cause the partial replacement of starch by sugar in sweet corn ³¹. Several Native American tribes grew sweet corn before the arrival of Europeans and the high frequency of a *su1* mutation in SW maize could help explain the early appearance and maintenance of sweet corn varieties by Native Americans.

The study of domestication and early crop evolution has largely been limited to the identification of key phenotypic and morphological changes between extant crops and their wild

relatives. As demonstrated here, the application of new paleogenomic approaches to well-documented temporal sequences of archaeological assemblages opens a new chapter in the study of domestication: it is now possible to move beyond a simple distinction of "wild" vs. "domesticated"³² and track sequence changes in a wide range of genes over the course of thousands of years.

References and Notes:

1. Ranere, A. J., Piperno, D. R., Holst, I., Dickau, R. & Iriarte, J. The cultural and chronological context of early Holocene maize and squash domestication in the Central Balsas River Valley, Mexico. *Proc. Natl. Acad. Sci. U. S. A.* **106**, 5014–8 (2009).
2. Piperno, D. R. & Flannery, K. V. The earliest archaeological maize (*Zea mays* L.) from highland Mexico: new accelerator mass spectrometry dates and their implications. *Proc. Natl. Acad. Sci. U. S. A.* **98**, 2101–3 (2001).
3. Matsuoka, Y. *et al.* A single domestication for maize shown by multilocus microsatellite genotyping. *Proc. Natl. Acad. Sci. U. S. A.* **99**, 6080–4 (2002).
4. Merrill, W. L. *et al.* The diffusion of maize to the southwestern United States and its impact. *Proc. Natl. Acad. Sci. U. S. A.* **106**, 21019–26 (2009).
5. Avila-Arcos, M. C. *et al.* Application and comparison of large-scale solution-based DNA capture-enrichment methods on ancient DNA. *Sci. Rep.* **1**, (2011).
6. Supplementary materials.
7. Chia, J., Song, C., Bradbury, P. & Costich, D. Maize HapMap2 identifies extant variation from a genome in flux. *Nat. Genet.* **44**, 803–U238 (2012).
8. Haury, E. in *Courses Toward Urban Life* (Braidwood, R. & Willey, G.) 106–131 (Aldine, 1962).
9. Ford, R. in *Prehist. Food Prod. North Am. Museum Anthropol. Anthropol. Pap.* (Ford, R.) 341–364 (University of Michigan, 1985).
10. Gregory, D. *Excavations in the Santa Cruz River Floodplain.* (1999).
11. Huckell, L. in *Hist. Maize* (Staller, J., Tykot, R. & Benz, B. F.) 97–106 (Elsevier, 2006).
12. Cutler, H. in *Mogollon Cult. Contin. Chang. Stratigr. Anal. Tularosa Cordova Caves* (Martin, P., Rinaldo, J., Bluhm, E., Cutler, H. & Grange, R.) 461–479 (Chicago Natural History Museum, 1952).
13. González, J. in *Corn Cult. Prehist. New World* (Johannessen, S. & Hastorf, C.) 135–157 (Westview Press, 1994).
14. Skotte, L., Korneliussen, T. S. & Albrechtsen, A. Estimating Individual Admixture Proportions from Next Generation Sequencing Data. *Genetics* genetics.113.154138– (2013). doi:10.1534/genetics.113.154138

15. Patterson, N. J. *et al.* Ancient Admixture in Human History. *Genetics* (2012). doi:10.1534/genetics.112.145037
16. Hufford, M. B., Martínez-Meyer, E., Gaut, B. S., Eguiarte, L. E. & Tenailon, M. I. Inferences from the historical distribution of wild and domesticated maize provide ecological and evolutionary insight. *PLoS One* **7**, e47659 (2012).
17. Jaenicke-Despres, V. *et al.* Early Allelic Selection in Maize as Revealed by Ancient DNA. *Science* (80-.). **302**, 1206–1208 (2003).
18. Svensson, E. M. *et al.* Tracing genetic change over time using nuclear SNPs in ancient and modern cattle. *Anim. Genet.* **38**, 378–383 (2007).
19. Li, Y. *et al.* Resequencing of 200 human exomes identifies an excess of low-frequency non-synonymous coding variants. *Nat. Genet.* **42**, 969–72 (2010).
20. Hufford, M. B. *et al.* Comparative population genomics of maize domestication and improvement. *Nat. Genet.* **44**, 808–U118 (2012).
21. Tenailon, M. I., U'Ren, J., Tenailon, O. & Gaut, B. S. Selection versus demography: a multilocus investigation of the domestication process in maize. *Mol. Biol. Evol.* **21**, 1214–25 (2004).
22. Zhao, Q., Weber, A. L., McMullen, M. D., Guill, K. & Doebley, J. MADS-box genes of maize: frequent targets of selection during domestication. *Genet. Res. (Camb)*. **93**, 65–75 (2011).
23. Weber, A. L. *et al.* The genetic architecture of complex traits in teosinte (*Zea mays* ssp. *parviglumis*): new evidence from association mapping. *Genetics* **180**, 1221–32 (2008).
24. Gallavotti, A. *et al.* The role of barren stalk1 in the architecture of maize. *Nature* **432**, 630–635 (2004).
25. Weber, A. *et al.* Major regulatory genes in maize contribute to standing variation in teosinte (*Zea mays* ssp. *parviglumis*). *Genetics* **177**, 2349–59 (2007).
26. Wang, H. *et al.* The origin of the naked grains of maize. *Nature* **436**, 714–9 (2005).
27. Korneliussen, T. S., Moltke, I., Albrechtsen, A. & Nielsen, R. Calculation of Tajima's D and other neutrality test statistics from low depth next-generation sequencing data. *BMC Bioinformatics* **14**, 289 (2013).
28. Liu, S. *et al.* Genome-wide analysis of ZmDREB genes and their association with natural variation in drought tolerance at seedling stage of *Zea mays* L. *PLoS Genet.* **9**, e1003790 (2013).
29. Wilson, L. M. *et al.* Dissection of maize kernel composition and starch production by candidate gene association. *Plant Cell* **16**, 2719–33 (2004).
30. Schultz, J. A. & Juvik, J. A. Current models for starch synthesis and the sugary enhancer1 (*se1*) mutation in *Zea mays*. *Plant Physiol. Biochem.* **42**, 457–464 (2004).
31. Dinges, J. R., Colleoni, C., Myers, A. M. & James, M. G. Molecular Structure of Three Mutations at the Maize sugary1 Locus and Their Allele-Specific Phenotypic Effects. *PLANT Physiol.* **125**, 1406–1418 (2001).
32. Zeder, M. A., Emshwiller, E., Smith, B. D. & Bradley, D. G. Documenting domestication: the intersection of genetics and archaeology. *Trends Genet.* **22**, 139–55 (2006).

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Figure legends

Figure 1. Origins of the SW ancient maize samples. SW3K, SW2K and SW750 correspond to SW maize from ~3000, ~2000 and ~750 BP. The ancient Mexican sample dates to 4,500 BP (TH564). The Mex prefix indicates modern Mexican samples that also include the Reventador, Chapalote and Highland landraces. More details in Table 1 and Supplementary Tables 4 and 5.

a) Treemix maximum likelihood tree depicting the expected signal of gene flow from *Z. m. Mexicana* into the highland landraces (also Supplementary Fig. 12) and gene flow from the coastal Chapalote into the SW2K. **b)** Subset of the population structure plot determined by NGSadmix with $K=5$ ⁶ (full plot in Supplementary Fig. 5); each individual is represented by a stacked column of the five proportions. **c)** The results from allele frequency-based *D*-tests²⁰ suggest an initial highland diffusion route from Mexico to the SW of the USA followed by extensive gene flow from the Pacific coast Chapalote race (Supplementary Table 6, Supplementary Fig. 12); positive values of *D* indicate gene flow from the coastal varieties into the SW maize; thick and thin bars correspond to 2 and 3 standard errors, respectively.

Figure 2. Potential targets of selection during domestication. **a)** Tajima's *D* for the two SW populations dated to ~2000 (colored dots) and ~750 BP (white triangles) plotted against the PBS distance for parviglumis. *zag1* shows the highest dissimilarity between parviglumis and the ancient Southwest landraces, i.e. the largest PBS(parviglumis). The gene with the lowest Tajima's *D* value for the SW750 population is also *zag1*. Genes with major roles in domestication traits are depicted in trapezoids. **b)** Gene trees built using PBS distances. *dehyd1A* is the top outlier for PBS(SW750) (Supplementary Fig. 10) and *su1* displayed the highest decrease in nucleotide diversity between the SW2K and the SW750 populations.

Figure 3. Timing of selective pressures on genes involved in the starch metabolism.

Nucleotide diversity variation for two key elements of the starch metabolism pathway. There is a steep decrease in nucleotide diversity before 2,000 BP for *ae1*, whereas the reduction in π for *sugary1* to less than half occurred after 2K and before 750 BP.

Table 1. Identification and summary statistics of the ancient samples sequenced in this study.

Age group	Type of analyses*	Ids	Intercept of radiocarbon age with calibration curve years BP	Cob morphology (P: pineapple; C:cylinder)	Site	Retained nucleotides	Average depth (targets)
SW3K	a,d	SW443	2,780		McEuen, USA	8,230,593	13
	t,a,d	SW4Ba	3,390		Bat Cave, USA	17,621,611	9
SW2K	a,d,s	SW207	1,860	P, 12 row, 2.0 cm	Tularosa, USA	5,870,362	11
	s	SW256	ca. 1,850-1,750	P, 12 row, 2.5 cm		3,768,546	9
	s	SW261	ca. 1,850-1,750	P, 10 row, 1.9 cm		4,613,152	9
	a,d,s	SW264	1,820	P, 12 row 1.8 cm		15,134,398	14
	s	SW278	ca. 1,850-1,750	P, 12 row, 2.2 cm		3,431,137	10
	a,d,s	SW280	ca. 1,850-1,750	P, 10 row, 2.1 cm		5,209,183	6
	s	SW283	1,860; 1,850; 1,830	P, 12 row, 2.2 cm		5,642,954	3
	s	SW288	ca. 1,850-1,750	P, 12 row, 2.3 cm		148,791	1
	s	SW296	ca. 1,850-1,750	P, 10 row, 1.9 cm		2,072,254	5
	t,a,d,s	SW298	1,770; 1,760; 1,740	P, 12 row, 2.0 cm		80,568,726	10
SW750	s	SW105	670	C, 10 row, 1.5 cm	Tularosa, USA	2,058,626	4
	a,d,s	SW107	ca. 700-900	C, 8 row, 1.3cm		34,929,483	20
	a,d,s	SW109	ca. 700-900	C, 8 row, 1.3 cm		12,364,145	15
	a,d,s	SW110	ca. 700-900	C, 8 row, 1.6 cm		35,088,565	17
	a,d,s	SW111	ca. 700-900	C, 8 row, 1.5 cm		29,640,515	19
	a,d,s	SW112	ca. 700-900	C, 8 row, 1.4 cm		22,887,209	16
	s	SW118	ca. 700-900	C, 8 row, 1.2 cm		3,855,808	4
	a,d,s	SW121	790	C, 10 row, 1.5 cm		29,736,402	7
	a,d,s	SW124	ca. 700-900	C, 8 row, 1.3 cm		33,518,448	18
	a,d,s	SW132	740	C, 8 row, 1.4 cm		17,131,288	18
	t,a,d,s	SW146	690	C, 8 row, 1.3 cm		111,329,149	12
	s	SW1b9	740	C, 8 row, 1.5 cm		686,34	2
	a	SW1AX				Turkey House, USA	59,526,622
TH	a	TH563	5,910	4 Ranks, 8 Rows, 1.2 cm	Tehuacan, Mexico	9,544,881	3
	t,a	TH564	5,280; 5,160; 5,140; 5,100	4 Ranks, 8 Rows, 1.1 cm		10,791,297	5
	a	TH157	1,410	8 Rows, 1.5 cm		18,126,654	2
AR	a	AR14B			Arica, Chile	5,328,366	16
	a	AR1A9				11,261,584	11
	a	AR1A8				286,639,854	24
	a	AR171				159,400,189	21

*t: Treemix (Fig. 1A), a: NGSadmix (Fig. 1b, Supplementary Fig. 5), d: D-statistics (Fig. 1c, Supplementary Fig. 12), s: selection tests (Fig. 2, Fig. 3, Supplementary Fig. 10)

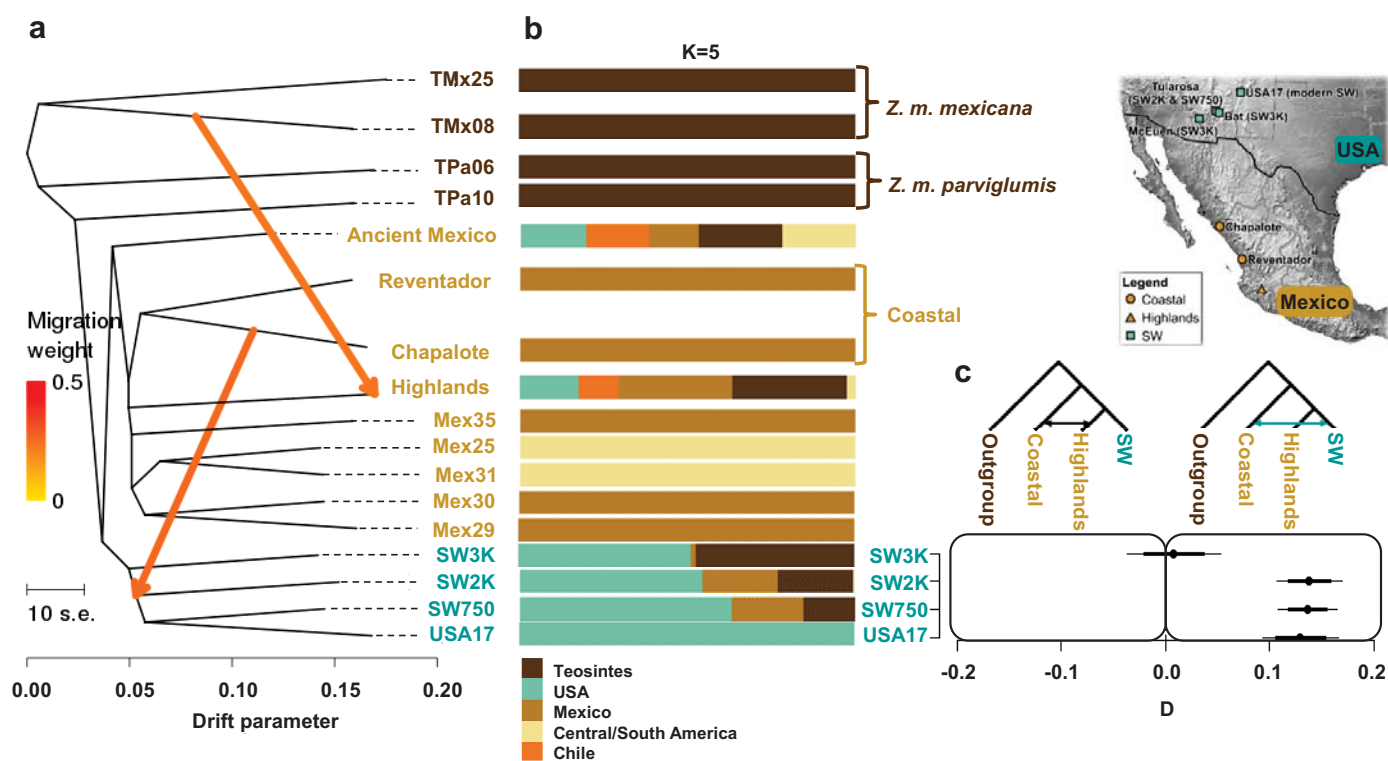


Figure 1

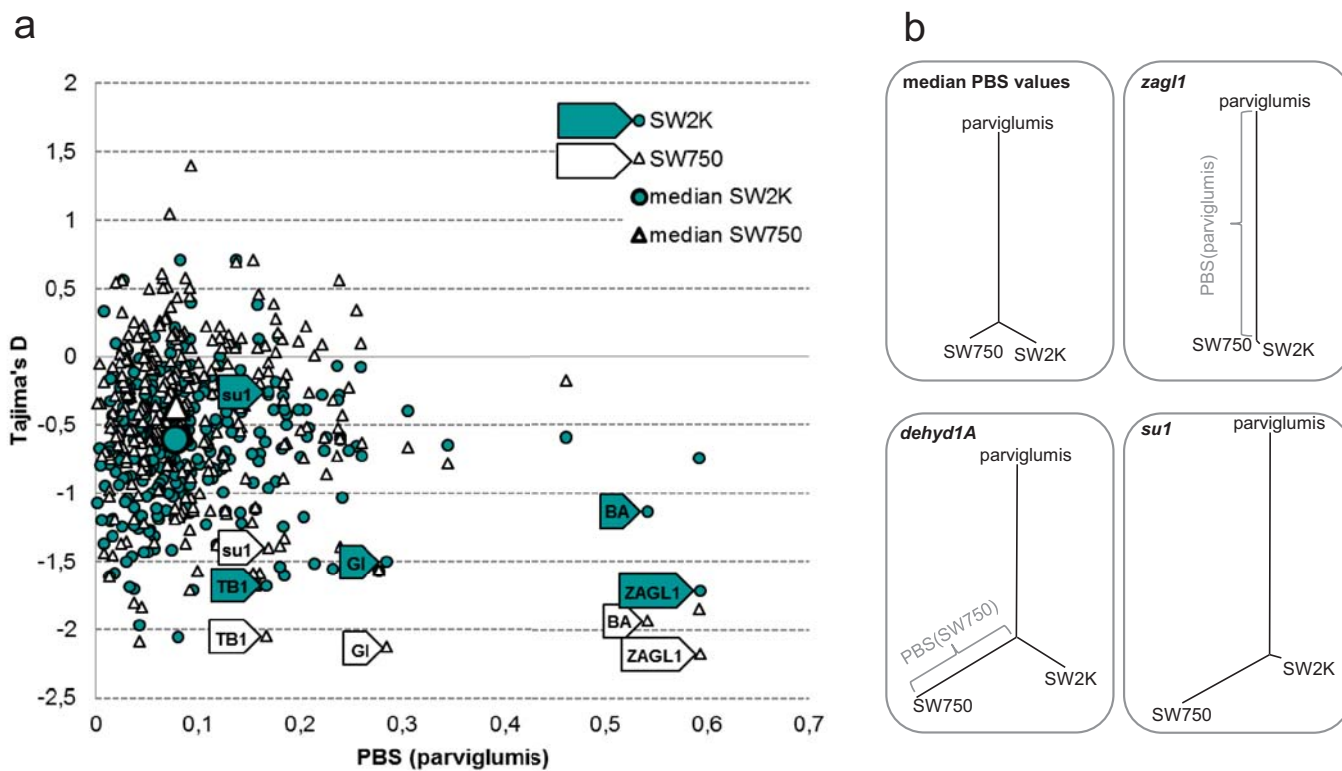


Figure 2

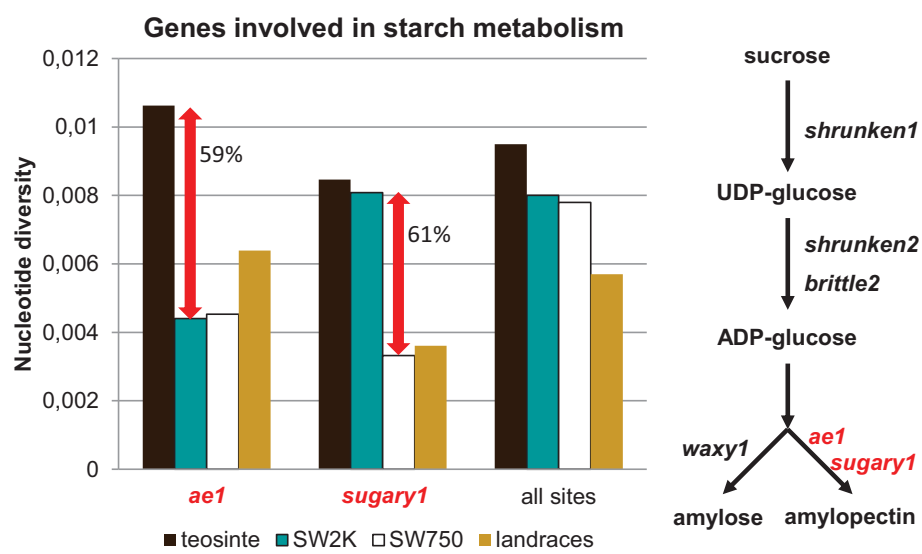


Figure 3