Variation in rural African gut microbiomes is strongly shaped by parasitism and diet

Elise Morton[1], Joshua Lynch[1], Alain Froment[2], Sophie Lafosse[2], Evelyne Heyer[2], Molly Przeworski[3], Ran Blekhman[#,1] and Laure Ségurel[#,2]

Co-supervised the work

1) Department of Genetics, Cell Biology, and Development, University of Minnesota, Minneapolis, MN, 55455
2) Eco-anthropology and ethnobiology, UMR 7206, CNRS-MNHN-University Paris 7 Diderot
3) Department of Biological Sciences, Columbia University, New York, NY 10027

Correspondence and requests for materials should be addressed to E.M. (email: emorton@umn.edu), R.B. (email: blekhman@umn.edu), or L.S. (email: lsegurel@mnhn.fr).

Abstract

The human gut microbiome is influenced by its host's nutrition and health status, and represents an interesting adaptive phenotype under the influence of metabolic and immune constraints. Previous studies contrasting rural populations in developing countries to urban industrialized ones have shown that geography is an important factor associated with the gut microbiome; however, studies have yet to disentangle the effects of factors such as climate, diet, host genetics, hygiene and parasitism. Here, we focus on fine-scale comparisons of African rural populations in order to (i) contrast the gut microbiomes of populations that inhabit similar environments but have different traditional subsistence modes and (ii) evaluate the effect of parasitism on microbiome composition and structure. We sampled rural Pygmy hunter-gatherers

as well as Bantu individuals from both farming and fishing populations in Southwest Cameroon and found that the presence of *Entamoeba* is strongly correlated with microbial composition and diversity. Using a random forest classifier model, we showed that an individual's infection status can be predicted with 79% accuracy based on his/her gut microbiome composition. We identified multiple taxa that differ significantly in frequency between infected and uninfected individuals, and found that alpha diversity is significantly higher in infected individuals, while beta-diversity is reduced. Another factor associated with microbial composition in our data is subsistence mode, notably with some taxa previously shown to differ between Hadza Eastern African hunter-gatherers and Italians also discriminating Pygmy hunter-gatherers from Cameroon from neighboring farming or fishing populations. In conclusion, our results stress the importance of taking into account an individual's parasitism status in studies of the microbiome, and highlight how sensitive the microbial ecosystem is to subtle changes in host nutrition. Our fine-scale analysis allowed us to identify microbial features that are specific to hunter-gatherers versus ones shared by all rural African populations, increasing our understanding of the influence of subsistence mode and lifestyle on gut microbiome composition.

Introduction

The human gut microbiota, the community of microorganisms inhabiting our gastrointestinal tract, is involved in a number of metabolic and immune functions and is now considered to be an essential determinant of human health[1-3]. Notably, these microbes enable an individual to recover key nutrients from food, and can prevent host infection by opportunistic pathogens[4]. Despite considerable plasticity in the structure and composition of an individual's gut microbiota[5], there are significant correlations between characteristics of the microbiome, dietary composition, host genotype, and patterns of disease[3,6,7]. These relationships suggest that composition of the microbiome represents a potentially adaptive phenotype that can be targeted by natural selection, especially in regard to adaptation to diet and exposure to pathogens.

Since the Neolithic revolution about 10,000 years ago, human populations have started to diversify their dietary regimes, resulting in the contrasted subsistence modes known today. This major cultural transition has also created new pathogenic pressures due to the proximity of livestock and the increased density of populations. Cultural and environmental differences among populations have resulted in physiological adaptations that can be detected in our genome[8-10]. Additionally, these factors have likely affected the community dynamics of our gut microbial ecosystem. Dietary changes have been shown to facilitate rapid changes in gut microbiota; however, the roles of habituation (over a lifetime) versus host adaptation (across generations) in these broader patterns are unclear. Understanding the long-term interaction that took place between the dietary specialization of populations and their gut microbiomes is therefore of great interest, notably to understand and predict the effect of recent and rapid changes in lifestyle and food on human health.

Nevertheless, to date, microbiome studies have mostly focused on industrialized populations. The few studies that have included a more diverse array of populations contrasted urban populations in highly industrialized countries to populations in developing countries and found that, outside of age, the factor that most strongly influences the gut microbiome is geography[11-16]. Such designs do not allow the respective influences of factors such as diet, climate, hygiene, parasitism and host genetics on population differences in the microbiome to be disentangled. While some specific changes in microbial communities have clearly been linked to components of human dietary regimes[17] and the use of antibiotics[18], the effect of other environmental or host-related factors is not clear. Notably, we do not know the extent to which the observed loss of microbial diversity of the human gut microbiome in urban industrialized populations[11-14] is attributable to their dietary specialization, changes in sanitation/hygiene practices, or other

factors. This loss of microbial biodiversity is a public health concern, as it may reflect a perturbed ecosystem associated with multiple diseases[19,20].

In addition to the loss of microbial diversity, developed countries nearly ubiquitously present a marked decrease in the prevalence of human gut parasites[21]. Although it is estimated that 3.5 billion people worldwide are infected with some parasite (protozoan or helminth)[22], little is known about their role in shaping the gut microbiome. Yet, throughout evolution, gut microbes and gut-dwelling parasites have co-inhabited the human gastrointestinal tract[23], and community dynamics are likely determined by current and past interactions (both during an individual's lifespan and throughout evolutionary history) between microbiota, protozoa, helminths, and the host immune response[24]. For example, it has been shown that direct competition by commensal microbes can provide protection from invading pathogens, and a disturbance to the natural microflora can effectively result in increased susceptibility to pathogens and/or parasites[25,26]. There is also substantial evidence that these interactions are essential for the development of a healthy immune system, and that the underlying cause of the increased incidence of autoimmune disorders in industrialized countries is the absence of exposure to pathogens and parasites early in life (the "hygiene hypothesis")[44]. In this context, it is important to evaluate the potential role of parasitism in shaping gut microbiome composition and structure.

Here, we focus on fine-scale comparisons of African rural populations with contrasting modes of subsistence and unequal access to medicine. Our objective is to understand the influence of current and ancestral diet, host genetics, and parasitism on the composition of the human gut microbiome. We focus on populations from Cameroon for which a large diversity of subsistence modes coexist in a restricted geographical area. We include individuals from Pygmy foraging (hunter-gatherer) populations, Bantu farming populations and Bantu fishing populations, all living in a similar rural environment. These populations are almost entirely self-sufficient in food;

104 their primary source of energy comes from cassava (*Manihot esculenta*), and fish or meat
105 provides the main source of protein. Animal food production for these populations has been
106 estimated to be high compared to elsewhere in Cameroon or Africa[27]. To account for recent
107 changes in diet, we evaluated current dietary regimes using dietary surveys. We also assessed
108 parasitism status by direct observations of fecal samples under the microscope. The focus on
109 populations living in the tropical rainforest is complementary to previous African populations
110 sampled: the Hadza hunter-gatherers and a population from Burkina Faso, living in the East and
111 West African tropical savanna, respectively[11,14]; and a population from Malawi living in a
112 relatively dry subtropical area of East Africa[12]. To the best of our knowledge, this study
113 represents the first comparison of the gut microbiome of human populations with limited
114 geographic separation and contrasting subsistence modes, as well as the first characterization
115 of the relationship between microbial communities and intestinal parasites.
116
117 **Results**
118 **Description of the samples: host genetics, diet and parasites**
119 *Host genetics*
120 We analyzed 64 individuals in seven different villages in Southwest Cameroon (average age 50
121 years, ranging from 26 to 78 years) corresponding to 20 hunter-gatherers, 24 farmers and 20
122 individuals from a fishing population (see Fig. 1 and Supplementary Table 1). The Pygmy
123 hunter-gatherers diverged from the other Bantu populations about 60,000 years ago[28,29] and the
124 farming subsistence mode likely started over the last 5,000 years[30]. The sampled populations
125 therefore not only have contrasted subsistence modes, but also have different genetic
126 backgrounds.
127
128
129

130 *Diet*

131 We chose these populations because previous work done in 1984-1985, based on nutritional
132 questionnaires and isotopes analyses, showed they had distinct diets[27,31]. We performed new
133 nutritional frequency surveys to assess how diet had changed during the past 30 years (see
134 Supplementary Table 1). Interestingly, the amount of meat in the hunter-gatherers' diet has
135 substantially decreased, reflecting the lower abundance of wild game in the forest reserve and
136 the hunting ban applied for some species. In contrast, the consumption of fish has increased in
137 inland populations (especially in farmers), due to the construction of new roads connecting the
138 coastal and inland populations. Similar to the results from 1984-1985, the farmers eat less
139 starchy foods (cassava) than hunter-gatherers and individuals from the fishing population
140 (Wilcox Rank Sum test: $p = 0.005$ and 0.017, respectively). A principal component analysis on
141 all dietary components revealed roughly three clusters corresponding to the three dietary
142 regimes, with the first axis distinguishing hunter-gatherers from the others, and the second axis
143 separating the farming and fishing populations (see Fig. 1b). The one exception to this pattern
144 concerns farmers from the North (living along the same road as the hunter-gatherers), which
145 cluster with the hunter-gatherers. In the following analyses, we therefore consider this
146 population separately from the farmers living in the South.

147

148 *BMI*

149 In addition to dietary questionnaires, we assessed the nutritional status of individuals by
150 measuring their BMI (Body Mass Index) (see Supplementary Table 1). Twenty percent of the
151 Pygmy hunter-gatherers were underweight (BMI<18) whereas 12%, 0%, and 4% of the South
152 farmers, North farmers, and individuals from the fishing population were, respectively.
153 Conversely, 0% of hunter-gatherers were overweight (BMI>25) while 12%, 14% and 26% of
154 individuals in the other groups were, respectively. This likely reflects the difference in socio-
155 economical status and access to medicine between these populations.

156

157 *Parasitism*

158 We assessed the intestinal parasitism of individuals by direct observation of their fecal samples
159 under the microscope. In more than one individual, we detected the presence of *Entamoeba*, as
160 well as eggs of *Ascaris*, *Trichuris*, and *Ancylostoma* (see Supplementary Fig. 1 and
161 Supplementary Table 1). Overall, 89% of hunter-gatherers, 76% of farmers from the South,
162 100% of farmers from the North, and 58% of individuals from the fishing population were
163 infected by at least one of these parasites. Regarding *Entamoeba*, 37%, 41%, 57% and 16% of
164 individuals were infected in each population, respectively. The reduced rate of parasitism in the
165 fishing population most likely reflects their higher level of hygiene and increased access to
166 medicine.

167

168 **Characterization of microbiome composition**

169 The fecal microbiota of 69 samples (including 5 biological replicates) were characterized by
170 sequencing of the V5-V6 region of the bacterial 16S ribosomal RNA with the Illumina MiSeq
171 technology. We obtained a total of 12.65 million high-quality reads, resulting in an average of
172 175,784 reads per sample (+/- 72,822). The average percent of mapped reads per individual
173 was 83% (SD = 7.5%) and did not vary significantly between populations (Welch's t-test, $p >$
174 0.2). The dataset was then rarefied to 50,000 reads/sample (see Supplementary Fig. 2), and
175 reads were clustered into 5039 operational taxonomic units (OTU) at 97% identity.

176

177 The five biological replicates (sampling of the same individual few days apart, see
178 Supplementary Table 1) allowed us to compare the microbial differences within individuals to
179 those between individuals. We calculated the UniFrac distance, a phylogenetic based distance
180 metric, which when weighted, accounts for relative abundance of taxa[32]. Because both weighted
181 and unweighted metrics capture different aspects of microbial diversity[32], we included both

182 types of analyses in the manuscript. We found that the average UniFrac distance between
183 replicates of the same individual was lower than between individuals (although only statistically
184 significant for the unweighted distances, one-sided Wilcox Rank Sum test: p = 0.003, see
185 Supplementary Fig. 3).
186
187 To test which variables were significantly associated with overall microbiome composition, we
188 performed a PERMANOVA analysis on the microbial abundance data. Intergroup comparisons
189 for various variables revealed that infection by the amoeba parasite *Entamoeba*, location,
190 subsistence mode and ancestry (the latter three being nested) were significantly associated with
191 variation in microbiome composition (p = 0.0001, 0.01, 0.003 and 0.01, respectively;
192 Supplementary Table 2). To further characterize patterns of variation that account for
193 phylogenetic relationships of community taxa, we also performed a PERMANOVA analysis on
194 both weighted and unweighted UniFrac distance matrices. Congruent with our previous
195 analysis, we found that infection by *Entamoeba* was the most significant variable for both
196 weighted and unweighted UniFrac distances (p = 0.007 and 0.0001, respectively;
197 Supplementary Table 2). *Entamoeba* infection also provided clear separation along the primary
198 axis of variation of the multidimensional scaling plots (Fig. 2a and Supplemental Fig. 4a).
199 Subsistence and location were both determined to be highly significant based on unweighted
200 UniFrac distances (p = 0.0003 and p = 0.002, respectively), but not weighted (p = 0.14 and p =
201 0.29, respectively). Because unweighted UniFrac distances assign increased value to rare taxa,
202 this suggests that less abundant taxa are more important in describing differences between the
203 microbiomes across subsistence modes and locations. Furthermore, subsistence provided only
204 weak visual separation along the first two axes of variation for both metrics (Supplementary Fig.
205 4b-c).
206
207

208 **Influence of parasitism on the microbiome**

209 Because of the significant relationship between *Entamoeba* infection status and patterns of
210 variation in the gut microbial communities found in all populations, we further investigated the
211 relationship between infection by this parasite and composition of the microbiome (Fig. 2). As it
212 is difficult to distinguish between the opportunistic pathogenic species (E. *histolytica*) and the
213 strict commensal (E. *dispar*) by microscopy alone, we were unable to characterize this parasite
214 at the species level. However, fewer than 10% of infected individuals were suffering from
215 diarrhea, suggesting that they were not experiencing symptomatic amebiasis[33].

216

217 At the phylum level, we found that 8 of the 13 phyla represented are significantly different
218 between *Entamoeba* infected (*Ent*+) and uninfected (*Ent*-) individuals, with most phyla (except
219 Bacteroidetes and Lentisphaerae) occurring at a higher relative abundance in *Ent*+ individuals
220 (see Table 1). When looking at individual taxa, based on a linear regression model, we also
221 identified a number of notable differences between *Ent*+ and *Ent*- individuals (Fig. 2b-c,
222 Supplementary Table 3-4), and we found that eighteen of the 93 most abundant taxa (present at
223 ≥ 0.1% in at least 4 individuals) differed significantly in their relative abundance between *Ent*+
224 and *Ent*- individuals ($q < 0.05$).

225

226 These taxonomic signatures for *Entamoeba* infection are so strong that an individual's infection
227 status can be predicted with 79% accuracy using a Random Forests Classifier (RFC) model
228 based on gut microbiome composition ($p < 0.001$; See Supplementary Fig. 5). Of the ten taxa
229 identified as being the most important in their predictive power, all but *Prevotella stercorea* were
230 significant in our linear regression model (of which all are in higher abundance in *Ent*+
231 individuals except *Prevotella copri*). The reason for the association between *Entamoeba* and
232 these microbes have yet to be identified, but it is noteworthy that the two most important taxa
233 identified in the RFC model, Elusimicrobiaceae unclassified (uncl) and Ruminococcaceae uncl,

234 include established endosymbionts of protists and common inhabitants of the termite gut[34].
235 Spirochaetaceae *Treponema*, the third most important taxon, include species that are
236 established human pathogens and others that have been reported to inhabit the cow rumen, the
237 pig gastrointestinal tract, and the guts of termites[35]. *Christensenellaceae*, the fourth most
238 important taxon, was recently identified as being the most heritable taxon in an analysis of twins
239 from the UK[3]. Two taxa in the order Bacteroidales, *Prevotella stercorea* and *Prevotella copri*, the
240 seventh and eighth most important taxa, are the only ones occurring at significantly reduced
241 abundance in infected individuals; *Prevotella* is an important genus of gut bacteria and is
242 underrepresented in Western versus African microbiomes[11]. While members of the Clostridia
243 and Gammaproteobacteria are more abundant in infected individuals, the pattern for
244 Bacteroidales is the opposite (see Fig. 2b. *Oscillospira uncl* and Parabacteroides uncl, the ninth
245 and tenth most important taxa, are associated with the rumen and human intestine, respectively.
246
247 Furthermore, when looking at the microbial diversity of *Ent*+ versus *Ent*- individuals, we found
248 that the presence of *Entamoeba* is associated with a significant increase in alpha (intra-host)
249 diversity using the Phylogenetic Distance Whole Tree metric (Welch's t-test: $p < 0.0001$, Fig.
250 3a), as well as using the Shannon and Simpson indices (Welch's t-test: $p = 0.001$ and $p =$
251 0.025, respectively; Supplementary Fig. 6). Interestingly, although the alpha (intra-host) diversity
252 of *Ent*+ individuals is significantly higher than *Ent*- individuals, the beta (inter-host) diversity (as
253 estimated by both UniFrac distance metrics) reveals that gut communities across *Ent*+
254 individuals are more similar than across *Ent*- individuals (Welch's t-test: weighted and
255 unweighted, $p < 0.0001$; Fig. 3b and Supplementary Fig. 7). This could suggest that, as alpha
256 diversity increases, there are fewer potential stable states for individual gut communities, or that
257 infection by *Entamoeba* drives changes in the microbiome that are dominant over other factors.
258
259

260 *Relationship between specific taxa and microbial community diversity*

261 Because of the striking relationship between *Entamoeba* infection status and alpha diversity, we
262 sought to identify any phyla for which abundance was significantly correlated with community
263 diversity. To account for the effect of *Entamoeba* infection, we added infection status as a binary
264 covariate to our linear model and identified 11 phyla that are significantly correlated with alpha
265 diversity ($q < 0.05$; see Supplementary Fig. 8). Although, as expected, the majority of these taxa
266 increase in abundance with higher diversity, Bacteroidetes and Proteobacteria exhibit a
267 decrease in relative abundance as alpha diversity increases. This negative relationship
268 suggests that these taxa might be more competitive than others and drive down diversity.

269

270 *Predicted metagenome*

271 We used the KEGG (Kyoto Encyclopedia of Genes and Genomes) database[36] and the PICRUSt
272 (Phylogenetic Investigation of Communities by Reconstruction of Unobserved States) pipeline[37]
273 to predict abundances of pathways across individuals (see Supplementary Fig. 9). Considering
274 the 220 most abundant KEGG pathways (comprising ≥ 0.01% of all assigned reads in at least 4
275 individuals), we identified 19 pathways with significant differences in abundance between *Ent*+
276 and *Ent*- individuals (linear regression: $q < 0.05$, see Supplementary Table 6 and Fig. 2d). Of
277 these 19, of particular interest are an increase in amoebiasis ($q = 0.001$), biosynthesis of the
278 antibiotic tetracycline ($q = 0.03$), and yeast MAPK signaling pathways ($q = 0.01$) in *Ent*+
279 individuals. These changes are largely attributed to Clostridiales and Ruminococcaceae, which
280 occur at significantly greater abundance in *Ent*+ individuals (6.53% vs. 4.53%, $q = 0.044$; and
281 29.58% vs. 16.34%, $q < 0.0001$, respectively, Fig. 2d). Interestingly, the Cellular Antigens
282 pathway, potentially involved in host-microbe and microbe-microbe interactions, is more
283 represented in the predicted metagenomes of *Ent*- individuals (linear regression: $q = 0.01$). This
284 pathway is predominantly attributed to members of the Enterobacteriaceae family, which was
285 found to be twice as abundant in individuals lacking the parasite.

286

287 Finally, we found that there was no statistically significant relationships between infection status
288 for this parasite and most of the covariates tested (including sex, BMI, subsistence, location, or
289 dietary components). The only significant covariate was adult age, with *Ent*+ individuals being
290 significantly older than *Ent*- individuals (See Supplementary Fig. 10; Welch's t-test: $p = 0.018$).
291 Despite this relationship, age is not a significant predictor of the gut microbial patterns across
292 populations (PERMANOVA for weighted and unweighted UniFrac distance matrices, $p > 0.06$;
293 Supplementary Table 2). Outside of an association between *Ancylostoma* and Bacteroidales
294 uncl ($q = 0.019$), none of the other parasites tested (*Ascaris* and *Trichuris*) exhibited a
295 significant association with any taxon, whether individually or as the number of all non-
296 *Entamoeba* parasite types present. However, the overall composition seems to shift with the
297 number of parasites (see Supplementary Fig. 11), and there is a significant increase in alpha
298 diversity when three parasite species are present (Supplementary Fig. 12).

299

300 **Influence of subsistence on the microbiome**

301 *Microbial community patterns across subsistence*

302 To investigate the effect of subsistence on the composition of the gut microbiome, we
303 summarized microbial taxonomic composition across the four subsistence groups and their
304 geographic locations (Fig 4a, Supplementary Fig. 13). At the phylum level, we found a
305 significant difference in the relative contribution of Proteobacteria across subsistence (linear
306 regression: $p = 0.003$, Table 1), with hunter-gatherers having a higher frequency than the fishing
307 population, farmers from the South and the North (23% versus 12.4%, 7.2% and 8.3%,
308 respectively), mirroring the higher frequency in the Hadza hunter-gatherers compared to
309 Italians[14]. Based on a linear regression, we also found that 8 of the most abundant taxa differed
310 significantly across subsistence modes (see Fig. 4b, Supplementary Tables 3-5). Of particular
311 interest is the genus *Bifidobacterium*, both *B. uncl* and *B. adolescentis*, which were found at

312 higher abundance in the fishing population (means 0.30% and 0.51%, respectively) relative to
313 all other populations (≤ to 0.11% and 0.07%, respectively; linear regression: q = 0.0003 and q =
314 0.008). This genus is associated with a higher consumption of dairy products, a pattern already
315 observed in a comparison of Italians to Hadza hunter-gatherers[14] and consistent with the
316 occasional consumption of yogurt in the fishing population. We also found Bacteroidales uncl to
317 occur at significantly lower relative abundance in the fishing population relative to the other
318 three populations (0.7% vs. ≥ 2.4%; linear regression: q = 0.003), a order of bacteria also
319 identified as being less abundant in the Italians versus the Hadza[14]. In contrast with other
320 Firmicutes genera that tend to be in lower frequency in hunter-gatherers, we found the genus
321 *Sarcina,* a synthesizer of microbial cellulose, to be only present in the hunter-gatherers (means
322 of 0.69% compared to ≤ 0.07% in the other subsistence groups, linear regression: q = 0.007).
323 Finally, we found three members of the Lachnospiraceae family to be significantly different
324 among populations, with *Ruminococcus uncl* and *Ruminococcus gnavus* being in lower
325 frequency in hunter-gatherers (0.34% and 0.19%, respectively) compared to other populations
326 (0.46-0.86% and 0.41-0.99%, respectively; linear regression: q = 0.030 and 0.006). This family
327 has been linked to obesity[39] in addition to protection from colon cancer attributable to their
328 production of butyric acid[40].
329
330 A random forest classifier (RFC) model for microbiome composition revealed an overall
331 accuracy of 59% ($p < 0.001$) for predicting the four subsistence groups but varied widely across
332 populations (see Supplementary Table 7 and Supplementary Fig. 14a). The hunter-gatherer
333 population was the most distinguishable such that the correct subsistence group was accurately
334 predicted 85% of the time (versus 31% if predictions had been made by chance alone).
335 Individuals of the fishing and South farming populations were predicted with 65% and 47%
336 accuracy, respectively (versus 31% and 27%, by chance), and the North farming population was
337 never predicted correctly (versus 11%, by chance). Furthermore, incorrect assignments for

individuals of the hunter-gatherer, farmers from the South and fishing populations were distributed evenly across all other subsistence groups, with the exception of farmers from the North, to which no individual was predicted to belong. Only five of the top ten taxa identified in the RFC model were determined to be significant in the linear regression (see Supplementary Fig. 14b). This suggests that rather than an individual signature taxon, it is the pattern of abundances of multiple taxa that is important for predicting subsistence. In agreement with our linear regression, the taxon identified as being the most important in distinguishing subsistence groups was *Bifidobacterium uncl* (see Supplementary Fig. 14b and Fig. 4b), occurring at significantly higher frequency in the fishing population (q = 0.0003, Supplementary Table 5). *Ruminococcus bromii*, important for degradation of resistant starch[38], was the second most important taxon, occurring at 0.01%, 0.01%, 0.15%, and 0.12% in the fishing population, farmers from the North, the South, and hunter-gatherers, respectively (q < 0.0001) (see Supplementary Fig. 14c). The third, fourth, fifth and eighth most important taxa include members of the Lachnospiraceae family, two of which were found to be significant in the linear model (see above). When grouped together, taxa in this family are less abundant in the hunter-gatherers relative to other subsistence groups (11.3% vs. 15.6-19.6%, respectively), a difference significant only when comparing hunter-gatherers to both farmer populations. Finally, two species of the family Succinivibrionaceae family, *Succinivibrio sp.* and *Ruminobacter sp.*, were also identified as being important taxa in the model, both of which were more abundant in the hunter-gatherers at 9.7% and 3.7%, respectively, vs. less than 5.7% and less than 0.1% for the other three subsistence modes (q = 0.068 and 0.057, respectively; see Supplementary Fig. 14c). Both of these taxa, associated with the bovine rumen, were also found in higher frequency in the Hadza hunter-gatherers[14]. Finally, only five of the top ten taxa identified in the random forest classifier model were determined to be significant in the linear model (see Supplementary Fig. 14b). This suggests that rather than an individual signature taxon, it is the pattern of abundances of multiple taxa that is important for predicting subsistence.

364

365 *Diet and gut microbial diversity*

366 We found the alpha (intra-host) diversity to be significantly lower in the fishing population than in
367 farmers from the South and the North for the phylogenetic distance whole tree metric (Welch's t-
368 test: $p = 0.021$ and $p = 0.008$, respectively) and only compared to farmers from the North for the
369 Shannon and Simpson metrics (Welch's t-test: $p = 0.017$, and 0.021, respectively) (see Fig. 5a,
370 Supplementary Fig. 15-17). Interestingly, the pattern of beta diversity across subsistence modes
371 using both unweighted and weighted Unifrac distance metrics also distinguishes the fishing
372 population from both farmers, such that the within-group variation is significantly higher in the
373 fishing and hunter-gatherer populations compared to both farmers (Welch's t-test: $p < 0.001$ for
374 all relevant pairwise comparisons) (see Fig. 5b and Supplementary Fig. 17a-b).

375

376 Community differences between subsistence groups based on weighted and unweighted
377 Unifrac distance metrics are the greatest between the fishing population and the other three
378 subsistence groups (Fig. 5c, Supplementary Fig. 17c). Beta diversity is the highest between the
379 fishing population and hunter-gatherers, the two groups for which there is the largest number of
380 significantly differentially abundant taxa (94% of significant taxa overall; Supplementary Table
381 5). This finding might be expected, given that these populations differ not only in terms of diet,
382 but also in their genetic ancestry and access to medicine. Both farmer populations are slightly
383 more different from the fishing population than from the hunter-gatherers (Fig. 5c,
384 Supplementary Fig. 17c), and present a higher number of significantly differentially abundant
385 taxa relative to the other populations (60% and 50% of the significant OTUs differ between the
386 fishing population and the farmers from the South and the North, respectively, versus 44% and
387 38% that differ from the hunter-gatherers; Supplementary Table 5). This could suggest that
388 differences in access to medicine, or the occasional consumption of processed food in the

389 fishing population, has considerable influence on their gut microbiomes, but this requires further
390 investigation.
391
392 According to the taxonomy-based predicted metagenome for each subject's gut microbiota, we
393 found that only one pathway, bacterial invasion of epithelial cells, differed significantly across all
394 subsistence types; represented at the highest relative abundance in the hunter-gatherers and
395 lowest in the farmers (linear regression: q = 0.03, Supplementary Fig. 18 and Supplementary
396 Table 6). This pathway includes proteins expressed by pathogenic bacteria that are important
397 for entry into host cells. The importance of this difference is unclear, but could be indicative of
398 an increased abundance of pathogens in the microbiomes of hunter-gatherers.
399
400 **Discussion**
401 We have conducted the first investigation of the relationship between intestinal parasitism and
402 the human gut microbiome, and found that infection by the amoeboid parasite, *Entamoeba,*
403 outcompetes diet, geographic location and ancestry in predicting composition and structure of
404 the gut microbiome. Furthermore, we have conducted the first analysis assessing the role of
405 subsistence, location and genetic ancestry in shaping the gut microbiota at a local scale. We
406 showed that there is striking variation amongst different rural African populations, indicating that
407 there are multiple signatures of rural, unindustrialized microbiomes.
408
409 *Influence of parasitism*
410 The importance of gastrointestinal parasites in human disease is well established, both as
411 infectious agents and in shaping immunity[22,41], and infection by helminths has notably been
412 found to be a major force underlying the evolution of interleukin genes in humans[23]. It has also
413 been demonstrated that loss of helminth exposure removes the enhanced T helper cell 2 (Th2)
414 and regulatory immune response imparted by these organisms, which is correlated with the

415 development of an array of immune-mediated diseases[42,43]. This relationship is the basis of the
416 hygiene hypothesis, which proposes that the underlying cause of the high incidence of
417 autoimmune diseases, unique to industrialized countries, is the absence of childhood exposure
418 to infectious agents[44]. Recent research supporting this hypothesis shows that mild and
419 controlled infection by internal parasites can activate an immune response and reduce
420 symptoms of a range of autoimmune diseases[45]. Likewise, the relationship between
421 gastrointestinal microbiota and host immune response has been well established[46,47]. Despite
422 these clear host-parasite and host-microbiome interactions, and the fact that gut parasites and
423 microbes share the same environment, there have been no studies assessing the relationship
424 between these organisms. In this context, it is unclear what the observed differences in gut
425 microbiome composition and structure between *Entamoeba* positive and negative individuals
426 mean in terms of host health, but it could be that *Entamoeba* infection indirectly triggers a host
427 immune response that differentially affects the success of different microbes. Alternatively, the
428 effects of *Entamoeba* infection could be direct, whereby, for example, amoeba feed on certain
429 species of bacteria, allowing others to proliferate. For instance, members of the phylum
430 Bacteroidetes are negatively correlated with alpha diversity. The decrease in these taxa in *Ent+*
431 individuals could therefore result in the observed increase in other taxa and overall increase in
432 alpha diversity. Interestingly, Clostridiales Ruminococcaceae, the second most important taxon
433 in the RFC model, significantly more abundant in *Ent+* individuals, has been found to be
434 underrepresented in individuals suffering from Crohn's Disease and Ulcerative Colitis[19].
435 Likewise, a decreased prevalence of *Prevotella copri* and Fusobacteria*,* as observed in *Ent+*
436 individuals, was recently shown to be negatively correlated with Rheumatoid arthritis[48] and
437 incidence of colorectal cancer[49], respectively, two conditions for which immunity also plays an
438 important role. This suggests that the link between parasite removal and inflammation related
439 human disease could in part be mediated by the gastrointestinal microbiome.
440

441 *Influence of subsistence and genetic ancestry*

442 In addition to identifying *Entamoeba* infection as an important predictor of gut microbiome
443 composition and structure across populations, we were also able to examine the relative
444 influence of other factors. First, we compared the gut microbiome composition of individuals
445 from the same subsistence mode and genetic ancestry, but coming from different villages.
446 Within the hunter-gatherers, we saw clear differences in composition as well as in diversity
447 between individuals living in Bandevouri versus those living in Makouré and Bidou, although this
448 difference was only significant using the Shannon Index for alpha diversity ($p < 0.05$; Welch's t-
449 test). Based on the data we have available, we found that these groups do not differ in terms of
450 diet or parasitism, suggesting a role for other unexplored environmental factors (e.g., water
451 source).

452

453 In term of genetic ancestry, we found that, despite a genetic divergence as old as 60,000 years,
454 the gut microbiome of the Pygmy hunter-gatherers is not strikingly different from that of the
455 Bantu populations. The UniFrac distances and the number of significant taxa are indeed lower
456 between hunter-gatherers and farmers than between the farmers and fishing population, two
457 Bantu groups that share the same genetic ancestry. In agreement with previous studies
458 showing that effects of host genetics are outweighed by diet and only affect specific taxa[3,50], this
459 suggests that host genetics alone is not a dominant factor in determining overall shifts in gut
460 microbiome composition.

461

462 Nonetheless, in our study, we found key differences distinguishing the microbiota of hunter-
463 gatherers from those of the farming and fishing populations, likely reflecting the influence of their
464 long-term diet. The hunter-gatherers were correctly assigned to their subsistence mode with
465 higher accuracy (85%) relative to the other populations. Furthermore, some of our findings
466 mirror patterns previously observed in the Hadza from Tanzania[14] (see Table 1b), suggesting

467 this ancestral subsistence mode might carry a specific microbial signature. Notably, we found a
468 higher frequency of Proteobacteria in hunter-gatherers compared to the other Cameroonian
469 populations, similar to the relationship between the Hadza and Italians[14]. Lachnospiraceae uncl,
470 identified as the third important in the RFC model with the tendency to be lower in the Pygmy
471 hunter-gatherers (5.7% versus 7.7-11.3% in other populations, q = 0.075), was also found to be
472 in lower frequency in the Hadza compared to Italians[14]. Finally, *Succinivibrio* and *Ruminobacter*
473 species, found enriched in Hadza, were also identified as important taxa in the RFC model, and
474 occur at higher frequencies in the Pygmy hunter-gatherers (see Table 1b). Thus, all these taxa
475 seem to be a specificity of hunter-gatherer populations, rather than reflecting a difference
476 between industrialized European and rural African populations. *Succinivibrio* is considered to be
477 an opportunistic pathogen, which could mean that hunter-gatherer populations have more
478 opportunistic pathogens than other populations, as proposed by Schnorr et al[14]. However, while
479 the opportunistic *Treponema* was also found enriched in the Hadza, we found it at a very low
480 frequency in all the populations studied here (< 3.5%, Table 1b), and not differing among
481 subsistence. When looking at other opportunistic genera in the Enterobacteriaceae family, we
482 found that *Shigella* and *Escherichia,* both previously found only in Italian children and not in
483 children from Burkina Faso[11], occur at extremely low abundances in all four subsistence groups
484 (< 0.1%, see Table 1b). As for *Klebsiella* and *Salmonella*, neither taxon differed significantly
485 amongst our groups (Table 1b). Thus, there does not seem to be any clear trend for
486 opportunistic pathogens in hunter-gatherers populations compared to others.
487
488 *Influence of geography and industrialized lifestyle*
489 Amongst the four populations included in this study, the fishing population is the most urbanized
490 due to increased consumption of processed food and access to medicine (resulting in a higher
491 BMI and lower prevalence of intestinal parasites). As such, the characteristics distinguishing the
492 gut microbiomes of the fishing population from the farmers and hunter-gatherers that also differ

493 between rural populations in developing countries and urban populations in industrialized
494 countries[12,14] might correspond to signature patterns of a more industrialized lifestyle. In
495 particular, within the phylum Bacteroidetes, we found a lower overall abundance of
496 Bacteroidales uncl in the fishing population relative to the other three populations (Table 1), an
497 order also depleted in Italians compared to Hadza[14]. High abundance of *Prevotella* and
498 *Bacteroides* have also been shown to represent signatures of the microbiomes for people in
499 developing and industrialized countries, respectively[11,12,14]. Higher abundances of *Prevotella* are
500 indeed often correlated with increased consumption of carbohydrates and simple sugars,
501 whereas an elevated proportion of *Bacteroides* is associated with a diet richer in protein and fat.
502 Although differences between the populations studied here was not statistically significant, the
503 fishing population harbored the highest abundance of *Prevotella sp*. (30.8%), while the farmers
504 from the North and the hunter-gatherers harbored the lowest (19.2% and 20.2%, respectively).
505 This abundance of *Prevotella sp.* is high relative to other genera in this order across all
506 populations (Table 1b). Species of *Prevotella* were the most reduced in individuals infected with
507 *Entamoeba,* a decrease that correlates significantly with higher alpha diversity.
508
509 In conclusion, our study suggests an important role for eukaryotic gut inhabitants and the
510 potential for feedbacks between helminths, protozoa, microbes, and the host immune response,
511 one that has been largely overlooked in studies of the microbiome. Prior analyses of the African
512 gut microbiome have found an enrichment of *Treponema,* Bacteriodetes *a*nd *Prevotella* as
513 compared to European populations. Although this enrichment was proposed to be related to
514 diet, our observations indicate that some of these trends could be related to infection by
515 *Entamoeba* (or other parasites). In addition, our results highlight the substantial variability in gut
516 microbiome composition among relatively closely related populations. Using a single population
517 as a representative of a diet or geographical region may therefore be overlooking important fine-

518 scale patterns in microbiome diversity. Hence, comparative population studies of the human
519 microbiome stand to benefit tremendously from considering variation within a geographic region.
520

521 **MATERIALS AND METHODS**

522 **Sample collection.** We sampled 80 volunteers (34 women and 46 men) in seven rural villages
523 (Bidou, Makouré, Bandevouri, Ndtoua, Afan Essokié, Akak and Ebodié) in Southern Cameroon,
524 after obtaining their informed consent for this research project. For each participant, we
525 collected information about their age, gender, anthropometric traits, health status, ethno-
526 linguistic and quantitative nutritional questionnaires. We also collected saliva and fecal samples.
527 The fecal sample was self-collected in the morning and stored in a plastic bag at most 3-4h
528 before further handling. It was then split in two separate samples; one was used to perform the
529 parasitological analysis at a local hospital (fresh or covered with formol) and the other was
530 stored to run the sequencing analyses. This latter sample was handled following previous
531 methods[51]: the sample was first submerged with pure ethanol for about 24h at room
532 temperature, then the ethanol was poured out of the container and the sample was wrapped in
533 a sterile gauze and deposited on silica gel. The silica gel was then replaced by new gel when it
534 changed colors from orange to yellow, i.e. when it could not absorb further humidity. The
535 samples were then transported back to France and stored at -80°C until they were shipped to
536 Minnesota, USA, on dry ice, and stored there at -80°C until further use. For five individuals, we
537 were able to collect replicate fecal samples at two different time points: four individuals 7 days
538 apart, and one individual 1 day apart.
539

540 **DNA extraction and 16S rDNA amplification from the fecal sample.** Total DNA was
541 extracted directly from approximately 50 mg of each fecal sample using the MO BIO
542 PowerFecal™ DNA Isolation Kit (MO BIO Laboratories, Carlsbad, CA, USA) according to the
543 manufacturer's protocol. DNA isolated from fecal samples was quantified using a NanoDrop

544 (ThermoScienctific), and the V5-V6 regions of the 16S rRNA gene were PCR amplified with the
545 addition of barcodes for multiplexing. The forward and reverse primers were the V5F and V6R
546 sets[52]. The barcoded amplicons were pooled and Illumina adapters were ligated to the reads. A
547 single lane on an Illumina MiSeq instrument was used (250 cycles, paired-end) to generate 16S
548 rRNA gene sequences yielding 175,784 Pass Filter (PF) reads per fecal sample (SD = 72,822)
549 and ~12.65 million total PF reads (4.9Gb of data).
550
551 **16S rDNA analysis**. Raw Illumina sequences were demultiplexed and filtered using cutadapt
552 1.7.1 (Martin, 2011) to remove adaptor sequences (Read
553 1:CTGTCTCTTATACACATCTCCGAGCCCACGAGAC, Read
554 2:CTCTCTCTTATACACATCTGCCGCTGCCGACGA), sequences containing ambiguous bases
555 and low quality reads (Phred quality scores < 20). Read pairs were resynced using RISS-UTIL
556 and matching paired-end sequences were merged using FLASH[53]. Merged sequences over 250
557 bp in length (the maximum length of the V5-V6 region) were removed. The remaining merged
558 sequences were analyzed using the open-source software package QIIME 1.7.0 (Quantitative
559 Insights Into Microbial Ecology)[54]. We performed closed-reference Operational Taxonomic Unit
560 (OTU) picking at 97% identity against the May 2013 Greengenes database[55] such that OTUs
561 were assigned based on taxonomic assignment of the reference sequence with a 97% similarity
562 cutoff. Reads which failed to hit a reference sequence with at least 97% identity were discarded.
563 The average number of reads that were assigned to an OTU per population was 82%. All
564 summaries of the taxonomic distributions ranging from phylum to species were generated from
565 the non-rarefied OTU table generated from this analysis.
566
567 **Diversity analyses**
568 To characterize diversity across individuals, rarefaction plots were generated for each sample
569 using the phylogenetic distance metric for diversity[56]. Samples were rarefied to 50,000 reads,

570 the maximum depth permitted to retain all samples in the dataset. All diversity analyses were
571 conducted on the rarefied OTU table containing 50,000 sequences per sample. Measurements
572 are based on the mean values calculated from 100 iterations using a rarefaction of 10,000
573 sequences per sample (20% of the total 50,000). Alpha-diversity was calculated for each
574 sample based on phylogenetic diversity, Shannon's index[57] and the Simpson index[58]. Beta-
575 diversity was assessed based on both unweighted and weighted UniFrac distance metrics[32]
576 using the Greengenes phylogenetic tree[55]. Principal Coordinate Analysis (PCoA) was carried
577 out on the distance matrices. P-values were calculated using the Welch's t-test. To determine if
578 the UniFrac distances were on average significantly different for groups of samples, we
579 conducted Principal Coordinates Analysis (PCA) to reduce raw gastrointestinal microbial
580 community data into axes of variation. We assessed the significance of each covariate by
581 performing a permutational multivariate analysis of variance (PERMANOVA)[59], a non-
582 parametric test, on both weighted and unweighted UniFrac distance matrices using the "adonis"
583 function from the *vegan* package in R[60]. This test compares the intragroup and intergroup
584 distances using a permutation scheme to calculates a p-value. For all PERMANOVA tests we
585 used 10,000 randomizations,
586
587 **Multivariate analysis of composition data**
588 Intergroup differences in microbiome composition for subsistence, location, population, BMI,
589 sex, language, age, dietary factors, and parasitism were assessed by PERMANOVA[59]
590 implemented using the "adonis" function of the *vegan* package in R[60]. Then, to identify taxa
591 significantly associated with each covariate of interest, we used a generalized linear model,
592 FDR corrected for the number of OTUs. For the linear model, OTUs with identical taxonomic
593 identifiers were merged. In parallel, we also restricted the merging only to OTUs names defined
594 at the family, genus or species level. Both results are reported in Supplementary Material
595 ("merged OTUs" versus "partially merged OTUs"). For analyses of both merged and partially

596 merged OTUs, the resulting taxa were filtered to include only those that occurred at least 0.1%
597 in at least 4 individuals.
598
599 **Random forest classifier model**
600 A random forest classifier with 2000 decision trees was trained on the taxa abundance table
601 consisting of 93 OTUs with 5-fold cross-validation using scikit-learn[61]. Mean accuracy (the ration
602 of the number of correct predictions relative to the total number of predictions) over the 5 folds
603 was 0.79 (standard deviation 0.09) with $p < 0.001$ (estimated using 1000 permutation tests with
604 5-fold cross-validation). The most discriminating taxa were identified by random forest
605 importance values (in scikit-learn random forest importance values are calculated as mean
606 decrease in node impurity). We report the top ten median importance values and 95%
607 confidence intervals from 1000 random forests.
608
609 **Metagenomic predictions**
610 We used PICRUSt v1.0.0 (Phylogenetic Investigation of Communities by Reconstruction of
611 Unobserved States) to generate taxonomy-based predicted metagenomes for each sample[37].
612 Counts from the rarefied OTU Table (50,000 OTUs per sample) were normalized by the
613 predicted 16S rRNA gene abundances and functional predictions of Kyoto Encyclopedia of
614 Genes and Genomes (KEGG)[36] pathways were determined using pre-computed files for the
615 May 2013 Greengenes database[55]. Relative abundances of the functional predictions were
616 calculated. We also compared the predicted metagenomes of individuals to determine which
617 functions were enriched or depleted across covariates (subsistence, population, BMI, age, sex,
618 dietary components, and parasitism phenotypes) for abundant (≥ 0.1% in at least 4 individuals)
619 and rare (<0.1% in at least 4 individuals) pathways. A linear mixed model was used to
620 determine which predicted pathways were significant ($q < 0.05$) for all each covariate.

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Figure 1

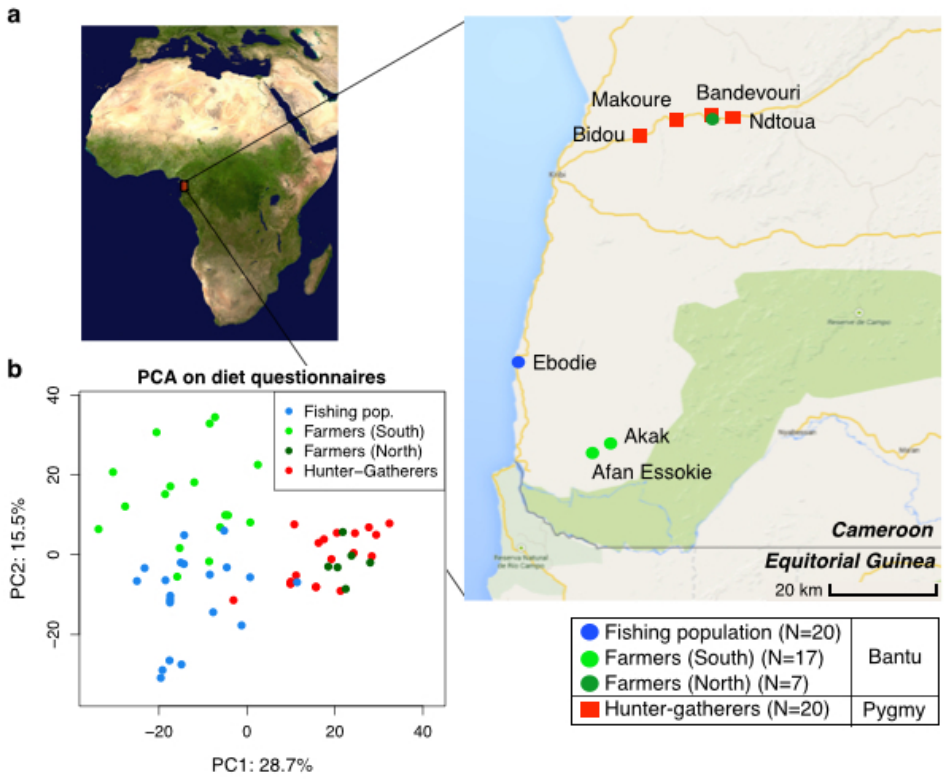

Fig. 1. (a) Map showing the geographic locations of the villages sampled in Southwest Cameroon, the number of samples (N) collected for each subsistence group (the fishing population, farmers from the South, farmers from the North, and hunter-gatherers), and their genetic ancestry (Bantu or Pygmy). (b) Principle Components Analysis based on dietary questionnaires for all 64 individuals. The first two principal components (PC1 and PC2) are shown, with the amount of variation explained reported for each axis.

Figure 2

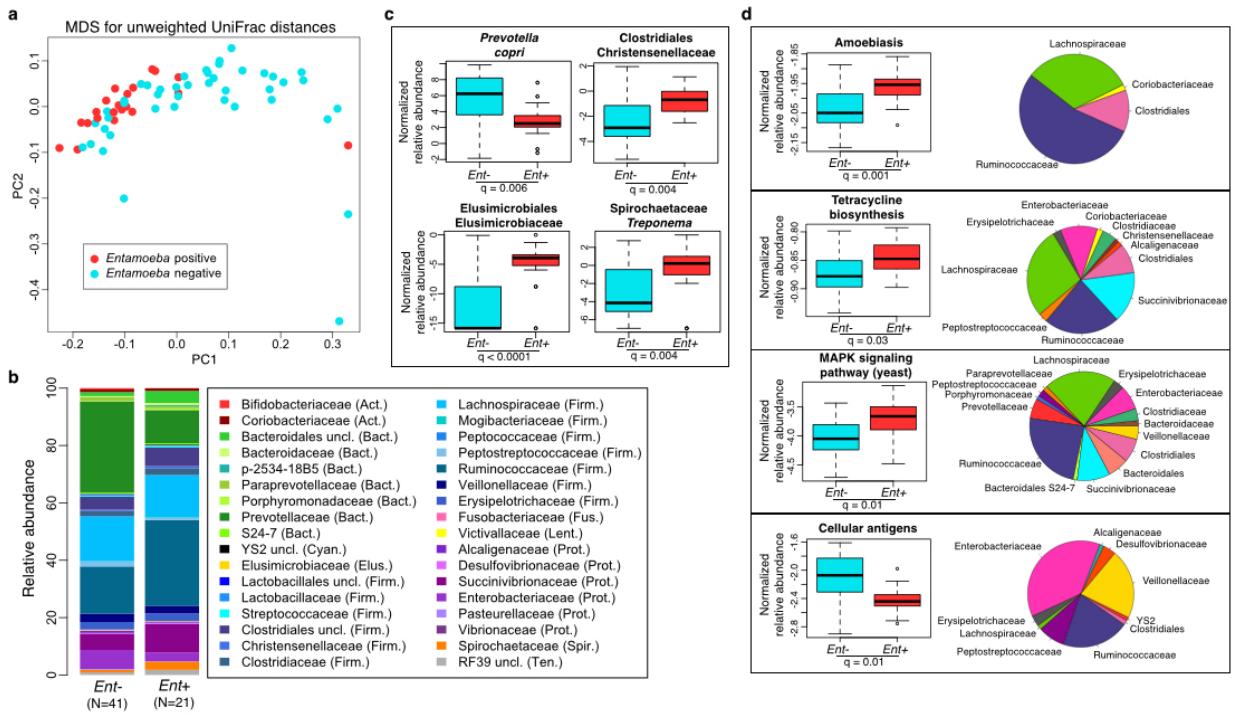

Fig. 2. Relationship between *Entamoeba* infection (*Ent-* or *Ent+*) and fecal microbiome composition. (a) Multidimensional Scaling plot of unweighted UniFrac distances colored by *Entamoeba* infection. The first two principal components (PC1 and PC2 are shown. (b) Summary of the relative abundance of taxa (>= 0.1% in at least 4 individuals) for *Ent-* and *Ent+* individuals color coded by phylum (Acinobacteria (Act.) = red, Bacteroidetes (Bact.) = green, Cyanobacteria (Cyan.) = black, Elusimicrobia (Elus.) = gold, Firmicutes (Firm.) = blue, Fusobacteria (Fus.) = pink, Lentisphaerae (Lent.) = yellow, Proteobacteria (Prot.) = purple, Spirochaetes (Spir.) = orange, and Tenericutes (Ten.) = gray). The number of individuals (N) in each population is indicated below the bars. (c) Normalized relative abundance of four taxa significantly associated with *Entamoeba* infection status in the linear regression as well as in the Random Forest Classifier model (q < 0.05). (d) Normalized relative abundance of four KEGG metabolic pathways significantly associated with *Entamoeba* infection status in the linear regression (q<0.05 using the most abundant (>= 0.4% in at least one group) KEGG (Level 3)

pathways) (left panel) and the relative contributions of each taxon for each pathway (right panel).

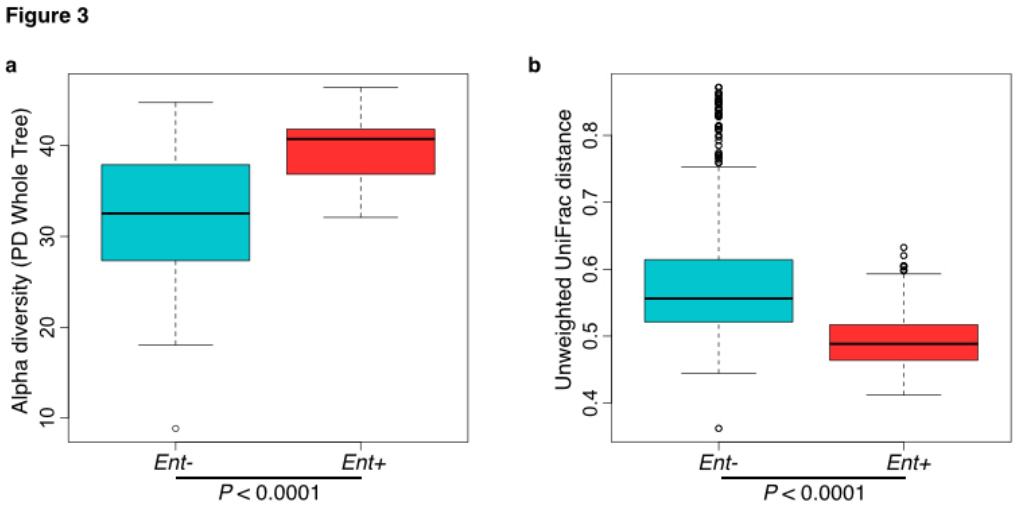

Fig. 3. (a) Comparison of alpha diversity for *Entamoeba* negative (*Ent*-) and positive (*Ent*+) individuals using the phylogenetic distance whole tree metric. (b) Comparison of beta diversity within *Ent*-, within *Ent*+, and between *Ent*- and *Ent*+ individuals based on unweighted UniFrac distances. P-values are based on Welch's t-test.

Figure 4

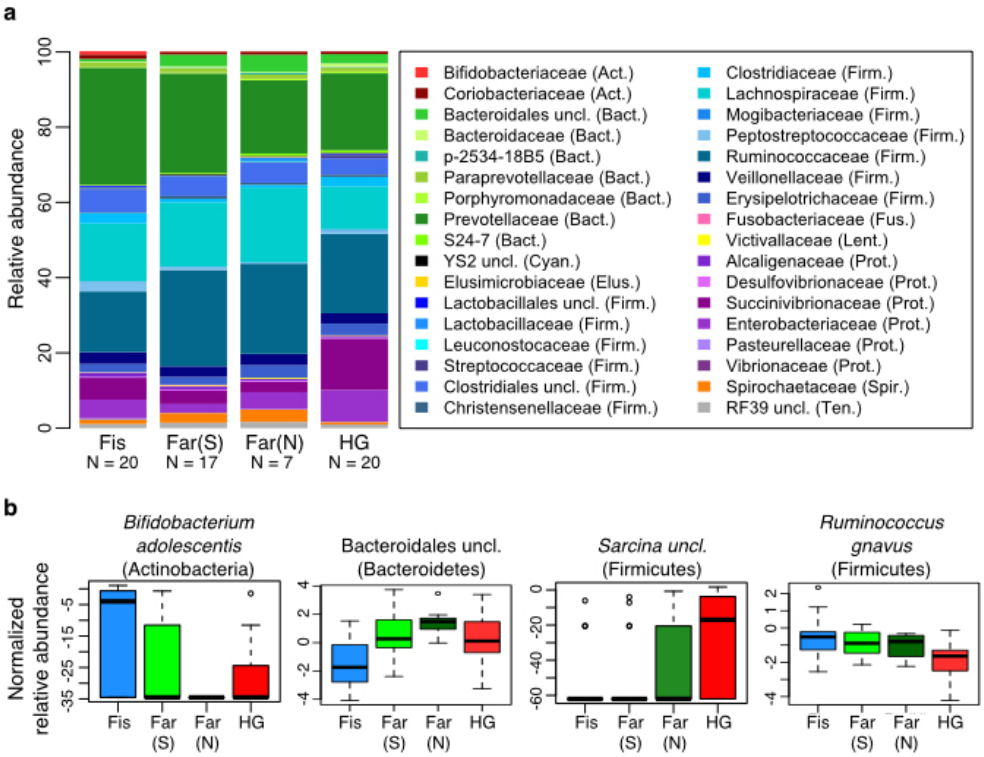

Fig. 4. Relationship between subsistence modes and fecal microbiome composition. (a) Summary of the relative abundance of taxa (occurring at >= 0.1% in at least 4 individuals) for individuals across subsistence. Taxa are colored by phylum (Actinobacteria (Act.) = red, Bacteroidetes (Bact.) = green, Cyanobacteria (Cyan.) = black, Elusimicrobia (Elus.) = gold, Firmicutes (Firm.) = blue, Fusobacteria (Fus.) = pink, Lentisphaerae (Lent.) = yellow, Proteobacteria (Prot.) = purple, Spirochaetes (Spir.) = orange, and Tenericutes (Ten.) = gray). The number of individuals (N) in each population is indicated below the bars. (b) Relative abundance of four taxa significantly associated with subsistence based on a linear regression model, $q < 0.05$. Fis = Fishing population; Far(S) = Farmers from the South; Far(N) = Farmers from the North; HG = Hunter-gatherers.

Figure 5

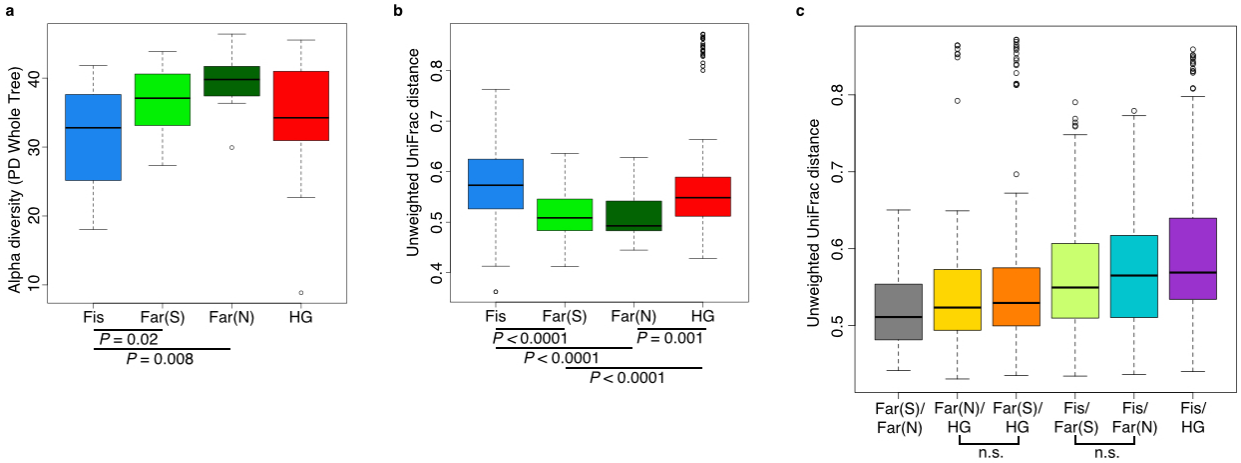

Fig. 5. Comparison of the diversity of gut microbiomes of individuals across subsistence (a) Alpha diversity based on the phylogenetic metric, phylogenetic distance (PD) whole tree. (b) Beta diversity within each subsistence group based on unweighted UniFrac distances. (c) Beta diversity for pairs of subsistence groups based on unweighted UniFrac distances. For interpair comparisons, all are significant ($P < 0.05$) unless specified (n.s.). All p-values are based on Welch's t-tests. Fis = Fishing population; Far(S) = Farmers from the South; Far(N) = Farmers from the North; HG = Hunter-gatherers.

Table 1

a

Phylum (>=0.1% in at least 4 ind)	Effect of *Entamoeba*			Effect of subsistence					Previous results from the literature[11,14]
	Ent -	*Ent* +	p-val	Fis	Far(S)	Far(N)	HG	p-val	
Actinobacteria	1.4	1.0	0.634	2.1	0.9	0.9	0.8	0.056	I > H
Bacteroidetes	35.3	18.6	0.003	33.4	31.7	26.7	26.0	0.708	H > I
Cyanobacteria	0.17	0.24	0.006	0.10	0.26	0.18	0.24	0.089	
Elusimicrobia	0.03	0.10	1E-07	0.01	0.08	0.23	0.05	0.206	
Euryarchaeota	0.03	0.09	4E-06	0.02	0.04	0.08	0.07	0.247	
Firmicutes	47.0	60.8	0.008	49.5	55.3	58.0	47.8	0.518	I > H
Fusobacteria	0.15	0.02	0.169	0.13	0.21	0.01	0.01	0.376	
Lentisphaerae	0.19	0.15	0.012	0.09	0.26	0.29	0.14	0.189	
Proteobacteria	13.8	14.1	0.915	12.4	7.2	8.3	23.0	0.003	H > I
Spirochaetes	0.9	2.9	1E-04	1.1	2.6	3.2	0.7	0.574	H > I
Tenericutes	1.0	1.9	0.001	1.1	1.5	1.9	1.0	0.238	
Verrucomicrobia	0.04	0.16	0.074	0.02	0.04	0.23	0.12	0.078	

b

Specific taxa of interest	Fis	Far(S)	Far(N)	HG	p-val	Previous results from the literature[11,14]
(Act.) *Bifidobacterium all species*	0.86	0.15	0.11	0.06	4E-06	I > H
(Act.) *Bifidobacterium adolescentis*	0.51	0.07	0.00	0.01	0.001	
(Bact.) *(Prevotellaceae) Prevotella all species*	30.8	26.2	19.2	20.2	0.409	H > I
(Bact.) *Bacteroides all species*	0.22	0.38	0.25	0.84	0.586	I > H
(Bact.) *Bacteroidales unclassified*	0.67	3.14	4.56	2.44	8E-05	H > I
(Firm.) *Lachnospiraceae unclassified*	7.7	9.8	11.3	5.7	0.015	I > H
(Firm.) *Ruminococcaceae Ruminococcus all species*	0.81	1.20	1.14	1.29	0.109	I > H
(Firm.) *Veillonellaceae unclassified*	0.22	0.35	0.67	0.21	0.008	H > I
(Prot.) *Succinivibrio all species*	5.7	3.3	2.8	9.7	0.012	H > I
(Prot.) *Ruminobacter all species*	0.07	0.10	0.02	3.74	0.007	H > I
(Prot.) *Klebsiella all species*	0.55	0.36	0.47	0.24	0.754	I > BF
(Prot.) *Salmonella all species*	0.04	0.01	0.02	0.09	0.501	I > BF
(Spir.) *Treponema all species*	1.1	2.6	3.1	0.7	0.806	H > I

Table 1. (a) Frequency (in %) of phyla for *Entamoeba* negative (*Ent*-) and positive (*Ent*+) individuals and for the four subsistence groups (Fis = Fishing population; Far(S) = Farmers from the South; Far(N) = Farmers from the North; HG = Hunter-gatherers). (b) Frequency (in %) of specific taxa of interest previously associated with geography in the four subsistence groups. P-values are based on a linear regression model. The last column indicates whether previous studies[11,14] found an enrichment of each phylum in Hadza adults (H) versus Italians adults (I), or in Burkina Faso children (BF) versus Italians children (I). The first letters in parenthesis indicate to which phylum each taxa belongs (Act. = Actinobacteria, Bact. = Bacteroidetes, Firm. = Firmicutes, Prot. = Proteobacteria, and Spir. = Spirochaetes).