Leaves as composites of latent developmental and evolutionary shapes

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Abstract

Across plants, leaves exhibit profound diversity in shape. As a single leaf expands, its shape is in constant flux. Additionally, plants may also produce leaves with different shapes at successive nodes. Because leaf shape can vary in many different ways, theoretically the effects of distinct processes are separable, even within the shape of a single leaf. Here, we measure the shapes of >3,200 leaves representing >270 vines from wild relatives of domesticated grape. We isolate latent shapes embedded within the overall shape of leaves that can predict developmental stages independent from species identity and vice versa. Shapes predictive of development are then used to stage leaves from 1,200 varieties of domesticated grape, revealing that changes in timing underlie leaf shape diversity. Our results indicate distinct latent shapes combine to produce a composite morphology in leaves, and that developmental and evolutionary contributions to shape vary independently from each other.

Significance Statement

The factors influencing leaf shape are multifaceted, including differences between species, different shapes of leaves a single plant produces during its lifetime, and the constant changes in shape that occur as an individual leaf expands. Here, we show that these processes affect distinct shape features of leaves independently from each other, allowing leaf development to be predicted separately from species identity. Our results have implications for taxonomy and assessments of biodiversity, isolating specifically those features most defining species from conserved, developmental shapes that evolve more slowly. Specific shape features can define outputs from distinct molecular pathways that change leaf morphology over evolutionary versus developmental timescales. Within a single leaf are distinct components of shape that are affected by distinct processes.

Leaf morphology represents a beautiful and tangible example of the infinite phenotypic possibilities in nature. Venation networks form an endless range of topologies and the contours of a leaf are defined by a boundless set of continuous points. Leaf shape varies not only across evolutionary timescales, but during development as well. Two distinct temporal processes regulate leaf shape during development. 1) The shape of individual leaves is in constant flux as local regions within the leaf expand at different rates. This phenomenon, allometric expansion, was explored as early as Hales' *Vegetable Staticks* (1727) (1). Using a grid of pins, regularly spaced puncture points in fig leaves were tracked to determine if their relative spacing changed during development. The same experiment can be microscopically studied using fluorescent particles today (2, 3). 2) The leaves that emerge at successive nodes differ in their shape, as the shoot apical meristem from which they derive transitions from a juvenile to adult stage of development. This process, heteroblasty, can affect other features of leaves besides shape, such as cuticle and trichome patterning (4-8).

The developmental stage of a leaf and the position of the node from which it arises (leaf number) are distinct temporal factors affecting leaf shape. Genetic changes in the timing of either process (e.g., protracted development of individual leaves or precociously adult leaf morphology in early nodes) between species can lead to evolutionary differences in leaf shape, a process known as heterochrony. The timing of these processes can be changed non-genetically as well, through responses to environmental changes during the lifetime of a plant, known as plasticity (9, 10). Distinguishing effects of developmental stage from leaf number provides mechanistic insights into how genetic changes during evolution or plastic changes in response to the environment are achieved (11, 12). For example, recent work has linked molecular pathways regulating the timing of shape changes throughout the shoot (miR156/172 and their targets) with leaf morphology (CUC-induced serrations) through a mediator (TCPs) (13).

If distinct molecular pathways affect different traits within the infinite features defining the architecture of a leaf, then the outline and venation topology of leaves should theoretically be decomposable into latent shapes (14). For example, shape differences defining species, regardless of developmental context, may vary distinctly from those features that define developmental context, regardless of species. These shapes are latent because they globally affect leaf shape in different ways, and each of the features comprising the latent shapes alone do not necessarily discriminate the effects of genetics or development. From this perspective, the shape of a given leaf—from any species, from any time point during development or any position in the plant—would result from the confluence of latent shapes regulated by these processes. The single organ that we call a leaf would actually be a composite of latent features that vary by genetic versus developmental effects, independently from each other.

Grapevine (*Vitis* spp.) leaves exhibit a breathtaking range of variation in leaf shape, making this genus ideally suited to explore evolutionary and developmental influences on form (**Fig. 1**). Variation in leaf lobing and leaf margins have been used by taxonomists to delimit the nearly 60 species in the genus (15, 16). Leaf shape is also important in assessing intraspecific variation in the European grapevine (*Vitis vinifera* ssp. *vinifera*), which is grown around the world for wine-making and table grapes. Unique among crops, variation in grape leaf shape (together with other vine features) was once, and is still today, used by viticulturists to quantitatively classify grape varieties, a field known as ampelography (αμπελος, "vine" and γραφος, 'writing') (17). Grape leaves have several homologous points amenable to landmark-based analyses (18, 19), increasing the biological interpretation of morphometric data compared to species with stochastic venation topologies or limited homology (such as *Arabidopsis*, tomato, *Antirrhinum*, etc.). Like Hales' grid of pins (1), naturally homologous points in grape leaves allow developmental stage and leaf number effects to be quantitatively tracked, potentially revealing separable latent processes contributing to the composite morphology known as a leaf.

155 **Results**

156

157 *Developmental stage versus leaf number*

158

159 To determine the effects of developmental stage, leaf number, and evolutionary
160 lineage (taxonomic identity) on leaf shape we scanned >3,200 leaves from wild
161 relatives of domesticated grape held by the USDA germplasm repository in Geneva,
162 NY. Like many living collections of perennial crops, the Geneva repository houses
163 multiple genotypes of different *Vitis* species in common conditions. In addition to an
164 extensive collection of assorted *V. vinifera* hybrids, the repository includes wild-
165 collected accessions of at least 19 North American and Asian *Vitis* species. From this
166 collection we sampled >270 vines representing 12 *Vitis* species, 4 *V. vinifera* hybrids,
167 and 3 species from the related genus *Ampelopsis.* Each sampled vine represents a
168 unique genetic accession. The number of vines sampled for each species varied
169 widely (**Fig. S1**), but in this study we focus on 10 species for which ≥5 accessions
170 were sampled: *Vitis riparia* (71 vines), *V. labrusca* (40 vines), *V. cinerea* (37 vines), *V.*
171 *rupestris* (28 vines), *V. acerifolia* (16 vines), *V. amurensis* (16 vines), *V. vulpina* (13
172 vines), *V. aestivalis* (8 vines), *V. coignetiae* (5 vines), and *Vitis palmata* (5 vines). All
173 *Vitis* leaves possess a midvein, distal and proximal veins, a petiolar vein, as well as
174 proximal and distal lobes and sinuses, and wide variation in the width of the petiolar
175 sinus (**Fig. 1A**). We leverage these points and others to measure 17 homologous
176 landmarks in all leaves (**Dataset S1**). Later, we compare the leaves from the wild
177 relatives of grape described above to previously published data on 1,200 varieties of
178 domesticated grape (19), which we describe in subsequent sections.

179

180 For each vine accession, a representative shoot was selected and the shoot position
181 of each leaf recorded (**Fig. 1B-C**). Developmental stage is measured by counting
182 from the youngest, first measureable leaf at the shoot tip. The time between
183 successively initiated leaves is a plastochron, and the youngest initiated leaf
184 primordium at the shoot apical meristem is denoted plastochron 1, P1 to indicate
185 this. But because we begin not with P1 (which is microns in size) but with the first

186 measureable leaf (~1 cm in size), we use S1 (for "stage") to denote the youngest
187 measured leaf at the shoot tip counting numerically upwards (S2 . . . Sn) in a
188 basipetal direction towards the shoot base. Contrastingly, leaf number begins with
189 the first initiated leaf (L1) found at the shoot base and counts numerically upward
190 (L2 . . . Ln) in an acropetal direction towards the shoot tip (**Fig. 1C**). These two
191 metrics are used to differentiate the effects of developmental stage (Sn) from leaf
192 number (Ln) (**Fig. 1C**).
193
194 The effects of developmental stage are expected to be strongest in young leaves at
195 the shoot tip (with low S numbers and high L numbers; **Fig. 1C**), as their shape is in
196 flux during their expansion. Older leaves at the shoot base (with low L numbers and
197 high S numbers) are more influenced by leaf number (i.e., their node position),
198 because they reflect changes in mature leaf shape at successive nodes. The vast
199 majority of the vines we sampled possess leaves corresponding to S1-S10 and L1-
200 L10 (86% represent developmental stages up to P10 and 88% represent leaf
201 numbers up to L10, **Fig. S1**) and we restrict our analyses to these positions (**Fig. 1C**).
202
203 *Morphospace of leaves in the genus Vitis*
204
205 Landmarks are aligned (accounting for translation, rotation, and scaling) using a
206 Generalized Procrustes Analysis (GPA). A Principal Component Analysis (PCA) is
207 then performed to visualize the major sources of shape variance among leaves. The
208 first four PCs explain 73.2% of shape variance among leaves from *Vitis* species,
209 developmental stages, and leaf numbers (**Fig. 2A**). More than half of the shape
210 variance, represented by PC1 and PC2, is influenced by lobing and the width of the
211 petiolar sinus. Low PC1 and high PC2 values readily distinguish highly dissected
212 *Vitis* species and some members of the related genus *Ampelopsis* (**Fig. 2B**).
213
214 In order to examine the variability of developmental stage and leaf number in
215 morphospace, we visualize each PC as a locally weighted scatterplot smoothing
216 (LOWESS) curve plotted against stage and leaf number. Developmental stage and

leaf number vary less by PC1 than by PC2 and PC3 (i.e., PC1 values are relatively developmentally-invariant compared to PC2 and PC3 values) (**Fig. 2C-D**). PC2 and PC3 values are largely indistinguishable between species across developmental stages (**Fig. 2C**) and leaf number (**Fig. 2D**), suggesting strongly conserved morphological features in developing leaves across species for these shape attributes, particularly for early developmental stages (**Fig. 2C**). Because PCs are orthogonal (i.e., uncorrelated), the fact that species, developmental stage, and leaf number correlate differentially with PCs is suggestive that each might be represented by independent shape attributes present within leaves. Re-visualizing the morphospace by developmental stage (**Fig. 2E**) and leaf number (**Fig. 2F**) demonstrates these factors mostly vary by PC2 and PC3, whereas species shape differences traverse along morphospace paths defined by PC1, PC2, and PC3 (**Fig. 2B**).

To help qualitatively understand the different ways grape leaves differ among species and developmental contexts, we compared average shapes (**Fig. 3**). One trend among species is that related members of Moore's Series Ripariae (*V. acerifolia*, *V. riparia*, and *V. rupestris*) (15, 20) are defined by a shallow petiolar sinus, especially *V. rupestris* (**Fig. 3A**). Outside of the Ripariae, *V. vulpina* and *V. cinerea* also exhibit shallow petiolar sinuses but to a lesser extent, while the petiolar sinus of the remaining species (*V. aestivalis, V. labrusca, V. palmata, V. coignetiae, V. amurensis*) is more acute. *Vitis palmata* exhibits especially deep distal lobing relative to other species. Each species' leaves also vary by developmental stage (**Fig. 3B**) and leaf number (**Fig. 3C**), usually by the length of the leaf tip and the shallowness of the petiolar sinus. In the next section, we determine the extent that shape attributes varying by species, developmental stage, and leaf number are separable from each other.

Latent shapes independently predict species, developmental stage, and leaf number

We used a Linear Discriminant Analysis (LDA) to maximize the separation of leaf

248 attributes (whether species, developmental stage, or leaf number) from each other
249 using all measured shape information. The resulting linear discriminants can be
250 used to predict the apparent class of a leaf and are useful for comparing actual
251 versus apparent leaf identities as confusion matrices (**Fig. 4**).
252
253 Linear discriminants separating species, without regard to developmental stage or
254 leaf number, can be used to predict species identity (**Fig. 4A-C**). For most taxa, the
255 largest proportion of predicted leaves corresponds to the taxonomic identity
256 assigned to that species (**Fig. 4B**, the diagonal indicates the proportion of correctly
257 assigned leaves). Incorrectly assigned leaves are most often confused for other
258 species that are phylogenetically related to the assigned species. For example, a
259 majority of *V. acerifolia* leaves are confused for its close relative *V. riparia*; similarly,
260 the largest group of *V. aestivalis* leaves are confused for its relative *V. labrusca*. In
261 contrast, the Asian *V. coigenetiae* leaves are confused for more distant relatives
262 native to North America, including *V. labrusca* and *V. cinerea* and sometimes even *V.*
263 *riparia,* perhaps indicating convergent evolution, germplasm misidentification, or
264 segregating leaf shape differences among these accessions. Interestingly*, Vitis x*
265 *andersonii,* described as a hybrid of *V. coignetiae* and either *V. vulpina* or *V. riparia*, is
266 strongly mistaken for *V. riparia*, providing circumstantial evidence of parentage. As
267 has been suggested by ampelographers and taxonomists (15, 17), our results
268 demonstrate that leaf shape is often sufficient to identify species, even in the
269 absence of developmental information. Instances where leaves of one species are
270 assigned to another offer a valuable opportunity to develop hypotheses about
271 phylogenetic history and convergent evolution that can be tested with additional
272 evolutionary and ecological analyses.
273
274 Reciprocally, we wondered whether developmental stage and leaf number,
275 regardless of species identity, might be similarly informative about developmental
276 context. Indeed, linear discriminants trained on developmental stage or leaf number
277 can predict the developmental context of a leaf for either measure (**Fig. 4D-4I**).
278 Developmental stage can be predicted with a Spearman's correlation coefficient of

279 rho = 0.70 (**Fig. 4D**), and leaf number with rho = 0.61 (**Fig. 4G**). Prediction of both
280 developmental stage and leaf number is more accurate at the beginning of their
281 respective series, where the effects of each are anticipated to be the strongest (**Fig.**
282 **1C**). Based on these observations we conclude that developmental context,
283 measured by either developmental stage or leaf number, can be predicted
284 independently of genotypic information. These results suggest that the effects of
285 development on leaf shape are conserved across genotypes and species, and have
286 implications for isolating distinct molecular pathways that affect leaf shape over
287 evolutionary versus developmental time scales.
288
289 The ability to predict developmental context separate from species identity provides
290 a method to quantify differences between species attributable to changes in
291 developmental timing, also known as heterochrony. For each leaf, we calculate
292 relative developmental stage and relative leaf number as the apparent value minus
293 the actual value (e.g., an S5 leaf predicted to be S7 would have a relative stage value
294 of +S2, and an L4 leaf predicted to be L2 would have a relative value of -2L). Relative
295 values indicate, for a given leaf, how many nodes ahead or behind (developmentally
296 speaking) that leaf appears to be. Averaging the relative developmental stage and
297 leaf number values for each vine, we can detect species that are precociously ahead
298 or lagging behind from expected developmental stage (**Fig. 4E**) and leaf number
299 values (**Fig. 4H**). For example, the mean relative developmental stage of *V. rupestris*
300 vines is about +1S ahead of their actual developmental stage (**Fig. 4E**) and about -2L
301 behind actual leaf number (**Fig. 4H**). This accelerated development (Sn) and
302 protracted display of juvenile leaf types typically found at the base of the shoot (Ln)
303 contributes towards the unique shape of *V. rupestris* leaves compared to other *Vitis*
304 spp. (**Fig. 3**). *Vitis rupestris* leaves (**Fig. 3A**), older leaves (high Sn values, **Fig. 3B**),
305 and juvenile leaves (low Ln values, **Fig. 3C**) all share a characteristic wideness
306 conferred by a diminished leaf tip.
307
308 Within *Vitis*, the identity of inflorescences, which appear opposite leaves, transform
309 from clusters at the shoot base to tendrils at the shoot tip (21-24) (**Fig. 1C**). Both

310 changes in leaf shape at successive nodes and the transformation of inflorescence
311 identity indicate temporal changes in the development of the shoot apical meristem,
312 known as heteroblasty. The cluster-to-tendril transformation serves as a discrete,
313 binary indication of the heteroblastic transition. If heteroblasty regulates both the
314 latent shapes predicting leaf number and the cluster-to-tendril transition, we would
315 assume they would be correlated. Indeed, there is a significant negative correlation
316 between relative leaf number and the first node a tendril is observed: that is, in
317 vines with precocious, adult leaves (i.e., higher relative leaf numbers, typical of
318 leaves found more towards the shoot tip), tendrils appear at earlier nodes closer to
319 the base of the shoot, linking two different measures of premature heteroblastic
320 change (**Fig. 4I**). Moreover, this correlation is not observed for relative
321 developmental stage (**Fig. 4F**). Taken together, these results indicate that latent
322 shapes predictive of developmental stage and leaf number are functionally distinct,
323 reflecting the developmental progression of leaves (Sn) and their heteroblastic
324 transitions across successive nodes of the shoot (Ln) separately.
325
326 Average leaf shapes indicate that developmental stage and leaf number vary by the
327 prominence of the leaf tip and the shallowness of the petiolar sinus (**Fig. 4J-L**). The
328 shape changes for these two factors are inversely related, but obviously
329 distinguishable from each other, given their differential correlation with tendril
330 position (**Fig. 4F**, **4I**). Upon closer inspection, the distal lobe tips and the branch
331 point of the petiolar vein distinguish shape changes attributable to developmental
332 stage and leaf number (**Fig. 4L**), isolating the shape attributes unique to these
333 functionally distinct processes.
334
335 *Morphological differences between domesticated grape and wild relatives*
336
337 Previously, we measured >9,500 leaves from *V. vinifera* subsp. *vinifera*
338 (domesticated grape) and its wild progenitor *V. vinifera* subsp. *sylvestris*. These
339 leaves, collected from the USDA germplasm repository in Winters, CA, represent
340 over 2,300 vines and 1,200 varieties (19). These leaves were sampled to measure

341 purely genetic effects and minimize the influence of development by collecting four
342 successive leaves from the midpoint of the shoot. That species identity can be
343 predicted independently from developmental context in wild *Vitis* spp. (**Fig. 4A-C**)
344 with disparate leaf shapes lends credence to the assumption that developmental
345 context can be ignored, which is almost always invoked when mapping traits for
346 quantitative genetic purposes or taxonomically defining species.
347
348 We wanted to compare leaves from domesticated grape and its wild progenitor to
349 the other *Vitis* species described above. To do so, we restricted our analysis *in silico*
350 to leaves at the same shoot positions we had originally collected in domesticated
351 grape (the four leaves closest to the midpoint of the shoot). In the combined
352 morphospace, PC1 and PC2 describe nearly 60% of shape variance (**Fig. 5A**). Similar
353 to the wild *Vitis* species-only morphospace (**Fig. 2A**), lobing and the petiolar sinus
354 define the first PCs. Petiolar veins in domesticated grape varieties can be so angled
355 they cross each other, and because of this the closure of the petiolar sinus
356 represented by high PC1 values is particularly strong (**Fig. 5A**). Leaf dissection
357 defined by high PC1 and high PC2 values clearly delineates less lobed wild *Vitis*
358 species from more acutely lobed domesticated grape varieties (**Fig. 5B**). Highly
359 dissected species such as *Ampelopsis acontifolia*, *V. piasezkii*, *V. thunbergii*, and *V.*
360 *vinifera* var. Ciotat form a distinct morphological grouping.
361
362 While there are clear patterns in leaf shape related to development and species
363 identity, correspondence between the morphospace of wild and domesticated *Vitis*
364 species and evolutionary relationships among *Vitis* species is complex. Some, but
365 not all, well-known phylogenetic relationships are reflected in leaf morphology.
366 (**Fig. 5C**) (20, 25). Notably, clustering averaged shapes from different genotypes
367 reveals that *V. vinifera* subsp. *vinifera* is morphologically sister to *V. vinifera* subsp.
368 *sylvestris*, well-known to be the wild progenitor of the domesticated grape (**Fig. 5C**,
369 Group II) (25, 26). Also within Group II are two Asian species, *V. amurensis* and *V.*
370 *coignetiae*, which together with *V. vinifera* subsp. *vinifera* and *V. vinifera* subsp.
371 *sylvestris* represent members of a Eurasian clade. Within Group III are found

372 members of Series Ripariae (*V. acerifolia*, *V. riparia*, *V. rupestris*). Leaf shapes of *Vitis*
373 hybrids cluster based on parental lineages, with *V. x champinii* and *V. x doaniana*
374 clustering in Group III presumably because of their *V. rupestris* and *V. acerifolia*
375 heritages (respectively) and *V. x andersonii* and *V. x novae-angliae* clustering closer
376 with their *V. coignetiae* and *V. labrusca* parents (respectively). These observations
377 suggest similarities in leaf shape may reflect recent evolutionary events,
378 domestication, or contemporary interspecific gene flow.
379
380 At broader levels of phylogenetic scale, however, patterns of leaf shape similarity
381 do not appear to correspond with known evolutionary relationships. For example,
382 together with *V. vinifera* subsp. *vinifera*, *V. vinifera* subsp. *sylvestris*, and two Asian
383 species found Group II (discussed above) are other North American species (*V.*
384 *labrusca, V. palmata*), several hybrids, and *Ampelopsis brevipedunculata*, taxa that
385 are not known to be closely related to one another. A previously identified clade
386 based on molecular data (*V. aestivalis*, *V. cinerea*, *V. labrusca*, and *V. vulpina)* here is
387 divided between Groups II and III. Although *Vitis* is well known to be a monophyletic
388 genus (27, 28), based on leaf shape three species of *Ampelopsis* cluster in three
389 different groups, together with *Vitis* species. These data suggest that while leaf
390 shape may bear signatures of recent evolutionary events, leaf shape does not appear
391 to track with phylogeny at larger scales; distantly related species, even from distinct
392 genera, resemble each other in leaf shape through evolutionary convergence.
393
394 *Changes in developmental timing underlie morphological diversity in domesticated*
395 *grape leaves*
396
397 Even though leaves from equivalent positions in the shoot clearly separate species
398 and varieties by genetic effects (**Fig. 5**), such effects may still be developmental in
399 nature, an example of heterochrony. If changes in the relative timing of
400 developmental stage (Sn) or leaf number (Ln) have occurred, they would contribute
401 to morphological differences between species, as described above for wild *Vitis* spp.
402 (**Fig. 4E, 4H**). The conserved latent shapes defining developmental stage and leaf

403 number in morphologically disparate *Vitis* species can be used predictively to detect
404 such changes in timing.
405
406 We used wild species leaves, from S1-S10 and L1-L10 (where developmental stage
407 and leaf number effects are the strongest, respectively) as a training set, to predict
408 the stage (Sn) and leaf number (Ln) values of domesticated grape leaves (**Fig. 6**).
409 When comparing leaf averages across predicted developmental stages (**Fig. 6A**) and
410 leaf numbers (**Fig. 6B**), it is apparent that variation in petiolar sinus depth is the
411 major predictor of developmental timing in domesticated grape leaves (**Fig. 6C**).
412 Variation in the petiolar sinus is a recurrent theme in morphology associated with
413 both developmental stage and leaf number (**Figs. 3B-C; 4J-L**). Indeed, the predicted
414 developmental stages and leaf numbers of domesticated grape leaves are
415 significantly correlated with PC1 and PC2 (**Fig. 6D-E, 6G-H**; **Dataset S2**), which
416 together explain near 60% of total shape variance for measured *Vitis* genotypes (**Fig.
417 5A**). From this, we conclude that latent shape attributes modulating developmental
418 stage and leaf number explain large amounts of shape variance in domesticated
419 grape, indicating changes in developmental timing (heterochrony) underlie the
420 morphological evolution of grape varieties.
421
422 *The relationship of leaf shape and timing to other traits in domesticated grape*
423
424 Previously, numerous other traits, from timing of bud burst and flowering to berry
425 ripening and titratable acids and sugar content, had been measured on the
426 domesticated grapevines described above (19). We were curious about the
427 correlational context of overall leaf shape (represented by PCs) and developmental
428 timing (predicted developmental stage and leaf number) to these other traits. One
429 trait, "PETSINMALF 2009", is a qualitative, 0-8 rating of petiolar sinus depth, and is
430 tightly correlated to PC1, PC2, and predicted developmental stage and leaf number,
431 confirming that our quantitative measures reflect intuitive perceptions of leaf shape.
432 (**Fig. 6F, 6I-J**; **Dataset S2**). Leaf morphology traits, including predicted
433 developmental timing, are largely separate from bud burst and bloom dates

434 ("BUDBURST DATE", "BLOOMDATE", "LEAFDATE"), indicating that these timing
435 features are separate from the overall transition to reproductive development in
436 grapevine (**Fig. 6J**). Further, leaf shape is distinct from overall size of leaves ("LEAF
437 SIZE"), demonstrating that grape leaf shape is not merely a developmental
438 constraint correlated with overall size (i.e., allometry) (**Fig. 6J**). One group of traits
439 significantly correlated and clustering with leaf shape is hirsuteness, both in leaves
440 and the shoot ("LEAF HAIR", "SHOOT HAIR"), which we had observed previously in
441 domesticated grape (19). Leaf hirsuteness varies by both developmental stage, as
442 trichomes become less dense as the leaf expands, and also by leaf number. It is
443 interesting to think about how thermoregulation, disease resistance, or other
444 trichome mediated functionality may vary as a developmental constraint as
445 developmental timing changed over the evolution and domestication of grapevines.
446
447 **Discussion**
448
449 Many descriptions of complex traits, from faces to voices, and even the evolution of
450 cultural products, such as violins (29-32) potentially stand to benefit from isolating
451 features uniquely regulated by distinct pathways. Like a face, leaf shape is
452 modulated in complicated ways by genetics, development, and the environment.
453 The latent shapes we describe here, describing developmental stage and leaf
454 number, embedded within the overall shape of a leaf, and conserved across the
455 genus *Vitis,* fit the definition of a "cryptotype". Recently, we had suggested the use of
456 this term in biology, which we borrowed from linguistics, which uses both
457 "phenotype" and "cryptotype" parallel to the biological definitions (14, 33).
458
459 In linguistics, overtly marked categories are called phenotypes. An example of an
460 overt category is English plurals, denoted in the vast majority of cases by a suffix "-s"
461 or a vowel change. Contrastingly, covert categories are called cryptotypes, and are
462 defined by latent combinations of words used more frequently than others. For
463 example, whereas most people would say "small, yellow house," few would use the
464 equivalent phrase "yellow, small house." The adjective-noun pairs "strong coffee"

465 and “powerful computer” are another example of cryptotype: although “powerful
466 coffee” and “strong computer” might technically be synonymous to the previously
467 mentioned pairs, they are rarely used, and convey their meaning much weaker than
468 their favored counterparts (33). Similar to a linguistic cryptotype, different factors
469 (genotype, developmental stage, and leaf number) act on the multivariate features
470 of a leaf (in this study, 17 homologous landmarks) combinatorially, enhancing their
471 effects on shape, not unlike favored combinations of words. These combinative
472 shapes are latent (a covert category) only because they are not immediately
473 recognizable when each of their component landmarks is considered alone; the
474 effects of genetics and development affect the entirety of the leaf. The multivariate
475 cryptotypes of genotype, developmental stage, and leaf number stand in contrast to
476 traditional univariate phenotypes, such as height or biomass.
477
478 Defining such combinative, latent shapes is critical to understand the mechanisms
479 by which morphological diversity manifests in plants. The unequal expansion of
480 leaves (allometry) and the different types of leaves displayed at nodes (resulting
481 from heteroblasty) have been studied for centuries (1, 4, 34-35), but the inability to
482 distinguish the effects at a morphological level, within the shared morphology of
483 single leaves where these processes act, has led to confusion. Early hypotheses that
484 shade prolongs juvenility through a heteroblastic mechanism were later refuted by
485 carefully studying the morphology of leaf primordia (12). And similarly, the degree
486 that changes in the development of leaves or the heteroblastic series influences
487 evolutionary changes—and plasticity—remains an open question.
488
489 Recent studies have uncovered a transcriptomic basis underlying heteroblasty (36),
490 and similarly dramatic transitions to reproductive fates (37), within meristems. An
491 understanding of molecular clocks, regulating phase change through small RNAs
492 (38-39), and their signals, such as sugar (40-41), has come into focus. Of the many
493 gene regulatory networks regulating leaf shape, specific pathways have been
494 identified mediating heteroblastic shape changes (13). Similar molecular
495 descriptions of leaf development exist (42), and the crosstalk between these two

processes is large. Strangely, even though molecular correlates of leaf development and heteroblasty have been uncovered, a phenotypic basis for the morphological effects of temporal patterning remains obscure, except in a qualitative sense. Decomposing complex morphologies into latent morphs regulated by distinct pathways is required to precisely describe the mechanisms that, together, produce the composite shape of a single leaf.

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Materials and Methods

Germplasm, sample collection, and scanning. Over 270 vines in the USDA *Vitis* germplasm collection in Geneva, NY were sampled in June, 2013. Using an automatic label maker, vine identification number and the identity of organs (or lack thereof) at each node, beginning with the first sampled leaf at the tip, was recorded in the field. Leaves, beginning with the first leaf at the tip that could be flattened and scanned (~1 cm in length), were collected in order, shoot tip to base, as a stack and placed into a Ziploc bag to which the label was affixed. A single shoot was collected per vine, and bags were placed into a cooler until scanning. A description of the number of vines (genotypes) collected for each species and hybrid, as well as the number of leaves representing different developmental stages (Sn) and leaf numbers (Ln) can be found in supplemental information (**Fig. S1**).

Leaves were arranged on a scanner (Mustek A3 1200S), in the collected order from the shoot, and next to each leaf was placed a small label indicating nodes (as measured by developmental stage) and organ identity opposite the node ("C", cluster; "T", tendril; "N", no organ). The abaxial side of leaves was imaged. The file name of the image indicates the vine ID, and the appended letter indicates which

527 image in the series the file represents for each vine. The raw scans are publically
528 available at the following link: http://thedata.harvard.edu/dvn/dv/vitis_leaves/
529
530 **Landmarking.** For both wild *Vitis* species and reanalyzed *V. vinifera* data, 17
531 landmarks were placed, in order, for each leaf using the ImageJ (43) point tool.
532 Landmarks and their order were as follows: 1) petiolar junction, 2) midvein tip, 3)
533 left distal sinus, 4) right distal sinus, 5) left distal lobe tip, 6) right distal lobe tip, 7)
534 left proximal sinus, 8) right proximal sinus, 9) left proximal lobe tip, 10) right
535 proximal lobe tip, 11) left terminus petiolar vein, 12) right terminus petiolar vein,
536 13) branch point midvein, 14) branch point left distal vein, 15) branch point right
537 distal vein, 16) branch point left proximal vein, 17) branch point right proximal vein.
538 Using ggplot2 (44) in R (45), graphs for landmarks from each image were visually
539 checked for errors. If errors were detected, the landmarking was redone for those
540 particular samples.
541
542 **Morphometric analysis and visualization.** Once a quality landmarked dataset was
543 created, a Generalized Procrustes Analysis (GPA) was undertaken using the R
544 package shapes (46). For 17 landmarks in two dimensions (x,y coordinates) for
545 3,292 leaves of wild *Vitis* species and 9,548 *V. vinifera* leaves, GPA was performed
546 using the procGPA function, reflect=TRUE. Separate Procrustes analyses were
547 performed for 1) all wild *Vitis* species leaves at all shoot positions and 2) *V. vinifera*
548 subsp. *vinifera* and subsp. *sylvestris* leaves and wild *Vitis* species leaves selected for
549 the corresponding shoot position as the domesticated grape dataset. Eigenleaves
550 were visualized using the shapepca function and PC scores, percent variance
551 explained by each PC, and Procrustes-adjusted coordinates were obtained from
552 procGPA object values. To test for correlation between PCs and developmental stage
553 or leaf number, Spearman's rho was calculated, whereas for variability of PC values
554 across species a Kruskal-Wallis test was used.
555
556 Linear Discriminant Analysis (LDA) on Procrustes-adjusted coordinates was
557 performed using the lda function from the MASS package (47). Species,

558 developmental stage, and leaf number were all analyzed independent of each other.
559 The predict function (stats package) and table function (base package) were used
560 (dependent on MASS) to reallocate leaves (whether by species, developmental stage,
561 or leaf number) using the linear discriminants. For the case of predicting *V. vinifera*
562 subsp. *vinifera* developmental stage and leaf number, the wild *Vitis* species data was
563 used as a training set to predict values for domesticated grape leaves. For the wild
564 *Vitis* species data, for each leaf there is actual versus apparent species,
565 developmental stage, and leaf number identities. Relative developmental stage and
566 relative leaf number are calculated as apparent value – actual value. Mean relative
567 developmental stage and relative leaf number values for each vine were used to
568 determine significant deviation of species relative values from zero using a one
569 sample, two tailed t-test. Correlation between relative values and first tendril node
570 was analyzed using Spearman's rho.
571
572 As previously described (19), Germplasm Resources Information Network (GRIN)
573 trait values, averaged on a per accession basis, for *V. vinifera* vines were correlated
574 with each other and morphometric and predicted temporal data using the rcorr
575 function from Hmisc (48) using Spearman's rho and false-discovery rate controlled
576 using the Benjami-Hochberg procedure (49). Hierarchical clustering for *V. vinifera*
577 subsp. *vinifera* traits, and for averaged Procrustes-adjusted coordinates for species
578 throughout the genus *Vitis*, was carried out using the hclust function on a distance
579 matrix calculated from correlation performed on complete, pairwise observations
580 and visualized using the as.phylo function from the package ape (50).
581
582 Visualization was performed in ggplot2 (44) using geom_bar, geom_boxplot,
583 geom_point, geom_segment, geom_tile, and stat_smooth functions, among others,
584 and color schemes derived from colorbrewer2.org.
585
586 **References**
587

1. Hales S (1727) *Vegetable Staticks or, an Account of Some Statistical Experiments on the Sap in Vegetables.* (London: W. and J. Innys.).

2. Remmler L, Rolland-Lagan AG (2012) Computational method for quantifying growth patterns at the adaxial leaf surface in three dimensions. *Plant Physiol* 159(1):27-39.

3. Rolland-Lagan AG, Remmler L, Girard-Bock C (2014) Quantifying shape changes and tissue deformation in leaf development. *Plant Physiol* 165(2):496-505.

4. Goebel K (1900) *Organography of Plants. I. General Organography.* (New York: Hafner Publishing Co.).

5. Ashby E (1948) Studies in the morphogenesis of leaves. I. An essay on leaf shape. *New Phytol* 47:152-176.

6. Poethig RS (1990) Phase change and the regulation of shoot morphogenesis in plants. *Science* 250:923-930.

7. Kerstetter RA, Poethig RS (1998) The specification of leaf identity during shoot development. *Annu. Rev Cell Dev Biol* 14:373-398.

8. Poethig RS (2010) The past, present, and future of vegetative phase change. *Plant Physiol* 154:541-544.

9. Allsopp A (1954) Juvenile stages of plants and the nutritional status of the shoot apex. *Nature* 173:1032-1035.

10. Diggle PK (2002) A developmental morphologist's perspective on plasticity. *Evolutionary Ecology* 16:267-283.

11. Jones CS (1993) Heterochrony and heteroblastic leaf development in two subspecies of *Curbita argyrosperma* (Cucurbitaceae). *American Journal of Botany* 80(7):778-795.

12. Jones CS (1995) Does shade prolong juvenile development? A morphological analysis of leaf shape changes in Cucurbita argyrosperma Subsp. Sororia (Cucurbitaceae). *American Journal of Botany* 82(3):346-359.

13. Rubio-Somoza I, Zhou CM, Confraria A, Martinho C, von Born P, Baena-Gonzalez E, Wang JW, Weigel D (2014) Temporal control of leaf complexity by miRNA-regulated licensing of protein complexes. *Curr Biol* 24(22):2714-9.

14. Chitwood DH, Topp CN (2015) Revealing plant cryptotypes: defining meaningful phenotypes among infinite traits. *Curr Opin Plant Biol* 24C:54-60.

634 15. Moore M (1991) Classification and systematics of eastern North American Vitis
635 L. (Vitaceae) North of Mexico. *SIDA Contrib Bot* 14:339-367.
636
637 16. Ren H and Wen J (2007) Vitis. *Flora of China* 12:210-222.
638
639 17. Galet P (1952) Précis d'Ampélographie Pratique. (Montpellier, France: Impr. P.
640 Déhan).
641
642 18. Galet P (1979) A Practical Ampelography: Grapevine Identification. Translated
643 by L Morton. (Ithaca, NY: Cornell University Press)
644
645 19. Chitwood DH, Ranjan A, Martinez CC, Headland LR, Thiem T, Kumar R,
646 Convington MF, Hatcher T, Naylor DT, Zimmerman S, Downs N, Raymundo N,
647 Buckler ES, Maloof JN, Aradhya M, Prins B, Li L, Myles S, Sinha NR (2014) A modern
648 ampelography: a genetic basis for leaf shape and venation patterning in grape. *Plant*
649 *Physiol* 164(1):259-72.
650
651 20. Miller AJ, Matasci N, Schwaninger H, Aradhya MK, Prins B, Zhong G-Y, Simon C,
652 Buckler ES, Myles S (2013) Vitis Phylogenomics: hybridization intensities from a
653 SNP array outperform genotype calls. *PLOS ONE* 8(11):e78680.
654
655 21. Srinivasan C and Mullins MG (1981) Physiology of flowering in the grapevine—a
656 review. *Am J Enol Vitic* 32(1):47-63.
657
658 22. Gerrath JM (1988) Morphological and anatomical development in the Vitaceae. I.
659 Vegetative development in *Vitis riparia*. *Can J Bot* 66:209-224.
660
661 23. Gerrath JM (1993) Developmental morphology and anatomy of grape flowers.
662 *Hortic Rev* 13:315-337.
663
664 24. Boss PK, Thomas MR (2002) Association of dwarfism and floral induction with a
665 grape 'green revolution' mutation. *Nature* 416:847-850.
666
667 25. Zecca G, Abbott JR, Sun WB, Spada A, Sala F, Grassi F (2012) The timing and the
668 mode of evolution of wild grapes (Vitis). *Mol Phylogenet Evol* 62(2):736-47.
669
670 26. Myles S, Boyko AR, Owens CL, Brown PJ, Grassi F, Aradhya MK, Prins B, Reynolds
671 A, Chia J-M, Ware D, Bustamante CD, Buckler ES (2011) Genetic structure and
672 domestication history of the grape. *Proc Natl Acad Sci USA* 108(9):3530-3535.
673
674 27. Wen J, Nie Z-L, Soejima A, Meng Y (2007) Phylogeny of Vitaceae based on the
675 nuclear GAi1 gene sequences. Canadian Journal of Botany 85: 731 – 745.
676
677 28. Wen J, Xiong Z, Nie Z-L, Mao L, Zhu Y, Kan X-Z, Ickert-Bond SM, Gerrath J, Zimmer
678 EA, Fang X-D (2013) Transcriptome sequences resolve deep relationships of the
679 grape family. PLOS ONE 8(9): e74394.

680
681 29. Zhang J, Yan Y, Lades M (1997) Face recognition: eigenface, elastic matching, and
682 neural nets. *Proceedings of the IEEE* 85(9):1423-1435.
683
684 30. Kuhn R, Junqua CJ, Nguyen P, Niedzielski N (2000) Rapid speaker adaptation in
685 eigenvoice space. *Speech and Audio Processing, IEEE Transactions on* 8(6):695-707.
686
687 31. Claes P, Liberon DK, Daniels K, Rosana KM, Quillen EE, Pearson LN, McEvoy B,
688 Bauchet M, Zaidi AA, Yao W et al. (2014) Modeling 3D facial shape from DNA. *PLOS*
689 *Genet* 10(3):e1004224
690
691 32. Chitwood DH (2014) Imitation, genetic lineages, and time influenced the
692 morphological evolution of the violin. *PLOS ONE* 9(10):e109229.
693
694 33. Whorf BJ (1945) Grammatical categories. *Linguistic Soc Am* 21:1-11.
695
696 34. Goethe JW (1817) Italienishe Reise. Goethe's Werk. (Stuttgart, Germany:
697 Dreizehnter Band).
698
699 35. Goethe JW (1952). Botanical Writings. Trans. B Mueller, introduction C.J. Engard.
700 (Honolulu, HI: University of Hawaii Press).
701
702 36. Efroni I, Blum E, Goldshmidt A, Eshed Y (2008) A protracted and dynamic
703 maturation schedule underlies Arabidopsis leaf development. *Plant Cell* 20(9):2293-
704 2306.
705
706 37. Park SJ, Jiang K, Schatz MC, Lippman ZB (2012) Rate of meristem maturation
707 determines inflorescence architecture in tomato. *Proc Natl Acad Sci USA* 109(2):639-
708 44.
709
710 38. Chuck G, Cigan AM, Saeteum K, Hake S (2007) The heteroblastic maize mutant
711 Corngrass1 results from overexpression of a tandem microRNA. *Nat Genet*
712 39(4):544-549.
713
714 39. Wu G, Park MY, Conway SR, Wang JW, Weigel D, Poethig RS (2009) The
715 sequential action of miR156 and miR172 regulates developmental timing in
716 Arabidopsis. *Cell* 138(4):750-9.
717
718 40. Yang L, Xu M, Koo Y, He J. Poethig RS (2013) Sugar promotes vegetative phase
719 change in Arabidopsis thaliana by repressing the expression of MIR156A and
720 MIR156C. *eLIFE* 2:e00260.
721
722 41. Yu S, Cao L, Zhou C-M, Zhang T-Q, Lian H, Sun Y, Wu J, Huang J, Wang G, Wang J-
723 W (2013) Sugar is an endogenous cue for juvenile-to-adult phase transition in
724 plants. *eLIFE* 2:e00269.
725

42. Ichihashi Y, Aguilar-Martinez JA, Farhi M, Chitwood DH, Kumar R, Milon LV, Peng J, Maloof JN, Sinha NR (2014) Evolutionary developmental transcriptomics reveals a gene network module regulating interspecific diversity in plant leaf shape. *Proc Natl Acad Sci USA* 111(25):E2616-21.

43. Abramoff MD Magalhaes PJ, Ram SJ (2004) Image processing with ImageJ. *Biophotonics International* 11(7):36-43.

44. Wickham H (2009) ggplot2: elegant graphics for data analysis. (Springer Science & Business Media).

45. R Core Team (2014) R: A language and environment for statistical computing. (Vienna, Austria: R Foundation for Statistical Computing).

46. Dryden IL (2013) shapes: Statistical shape analysis. R package version 1.1-9, http://CRAN.R-project.org/package=shapes

47. Venables WN, Ripley BD (2002) Modern applied statistics with S. Fourth Edition (New York: Springer).

48. Harell FE (2013) Hmisc: Harrell Miscellaneous. R package version 3.10-1.1, http://CRAN.R-project.org/package=Hmisc

49. Benjamini Y, Hochberg Y (1995) Controlling the false discovery rate: a practical and powerful approach to multiple testing. *Journal of the Royal Statistical Society, B* 57(1):289-300.

50. Paradis E, Claude J, Strimmer K (2004) APE: analyses of phylogenetics and evolution in R languages. *Bioinformatics* 20:289-290.

Figure Legends

Fig. 1: Morphological features defining the temporal development of *Vitis* leaves. (*A*) All *Vitis* leaves possess distal (yellow), proximal (orange), and petiolar (red) veins, as well as distal and proximal lobes and sinuses and a petiolar sinus. 17 homologous landmarks (black circles) were used in this study. Morphologically diverse species are shown. (*B*) Leaf series, scaled by the length between distal lobe tips, showing developmental stage and leaf number shape variance. (*C*) Unscaled *V. riparia* leaves showing changes in size at the shoot tip and base. Developmental stage (Sn) is measured starting from the shoot tip and leaf number is measured

766 from the shoot base (Ln). Inflorescences, opposite leaves, skipping every third node,
767 transform from clusters into tendrils from the shoot base to tip.
768
769 **Fig. 2: Morphospace of leaves in the genus *Vitis*.** (*A*) "Eigenleaves" showing leaf
770 morphs represented by Principal Components (PCs) at -/+ 3 standard deviations
771 (s.d.) and shape variance explained by each. (*B*) Select species (indicated by color)
772 representing morphological diversity in the dataset projected onto the
773 morphospace. 95% confidence ellipses are drawn. (*C* and *D*) Locally Weighted
774 Scatterplot Smoothing (LOWESS) showing the relationship between PCs 1-3 and (*C*)
775 developmental stage and (*D*) leaf number. Phylogenetic relationships between
776 species are indicated by color. (*E*) Developmental stage and (*F)* leaf number
777 projected onto the morphospace.
778
779 **Fig. 3: Shapes represented among *Vitis* species, developmental stage, and leaf**
780 **number.** (*A*) Outlines representing the average shape of each pairwise comparison
781 of species, indicated by magenta and green. Phylogenetic relationships are indicated.
782 (*B* and *C*) Comparison of average shapes across (*B*) developmental stages and (*C*)
783 leaf number for each species indicated by color.
784
785 **Fig. 4: Species, developmental stage, and leaf number can be predicted**
786 **independently of each other.** (*A*) Confusion matrix resulting from prediction of
787 species identity using a Linear Discriminant Analysis (LDA). The panel should be
788 read left to right: for each species indicated on the left, leaves appeared to be
789 derived from which apparent species (indicated on bottom)? Developmental stage
790 and leaf number were not considered in the prediction. (*B*) Same as (*A*), except the
791 apparent species assigned to each actual species have been converted into a
792 proportion. Proportion totals equal one adding left to right. (*C*) The proportion of
793 correctly assigned leaves for each species. (*D*) Similar to (*A*), actual developmental
794 stage (left) and apparent developmental stage (bottom), predicted without species
795 or leaf number information. (*E*) Relative developmental stage values (apparent –
796 actual developmental stage) averaged for vines from each species. Significant

797 deviations from 0 indicated by * ($p<0.05$), ** ($p<0.01$), and *** ($p<0.001$). (*F*)
798 Correlation between mean relative developmental stage of vines and first tendril
799 node. Fitted linear model is drawn. (*G-I*) same as (*D-F*) but for leaf number. Note the
800 significant correlation between first tendril node and relative leaf number (*I*) not
801 seen for developmental stage (*F*). (*J* and *K*) Latent shape attributes defining
802 developmental stage (*J*) and leaf number (*K*) indicated with average leaf shapes. (*L*)
803 Comparison of changes in latent shape attributes across developmental stage
804 (orange, outline S1; purple, arrow to S10) and leaf number (grey, outline L1; black,
805 arrow to L10). Note non-inverse relationship between developmental stage and leaf
806 number changes in the distal lobe tips and petiolar vein branch points (indicated).
807
808 **Fig. 5: Morphospace of wild *Vitis* species and domesticated grape.** (*A*)
809 "Eigenleaves" showing leaf morphs represented by PCs at -/+ 3 s.d. and shape
810 variance explained by each. Lobing and petiolar sinus variation indicated. (*B*)
811 Comparison of wild *Vitis* species leaf shape (red, black) with *V. vinifera* subsp.
812 *vinifera* and *V. vinifera* subsp. *sylvestris* (green, dark green, respectively), and highly-
813 lobed species (blue). Strong shape variation in lobing and petiolar sinus depth
814 indicated. (*C*) Hierarchical clustering based on morphology. Phylogenetic groupings
815 indicated in light gray. Groups mentioned in text indicated with Roman numerals.
816
817 **Fig. 6: Evolution in developmental timing underlies leaf shape diversity in**
818 **domesticated grape.** (*A* and *B*) Latent shape attributes defining developmental
819 stage (*A*) and leaf number (*B*) in domesticated grape indicated with average leaf
820 shapes. (*C*) Comparison of changes in latent shape attributes across predicted
821 developmental stage (orange, outline S1; purple, arrow to S10) and predicted leaf
822 number (grey, outline L1; black, arrow to L10). (*D* and *E*) Predicted developmental
823 stage (*D*) and predicted leaf number (*E*) projected onto the *V. vinifera* subsp. *vinifera*
824 morphospace demonstrates changes in developmental timing (heterochrony)
825 underlie leaf shape diversity in domesticated grape. (*G – I*) Correlation between PC1
826 and PC2 with predicted developmental stage (*G*), predicted leaf number (*H*), and a
827 qualitative measure of petiolar sinus depth, PETSINMALF 2009 (*I*). (*J*) Hierarchical

clustering of measured traits in grape. Morphological and predicted temporal traits (yellow), hirsuteness (green), reproductive transitions (blue), and overall leaf size (grey) indicated.

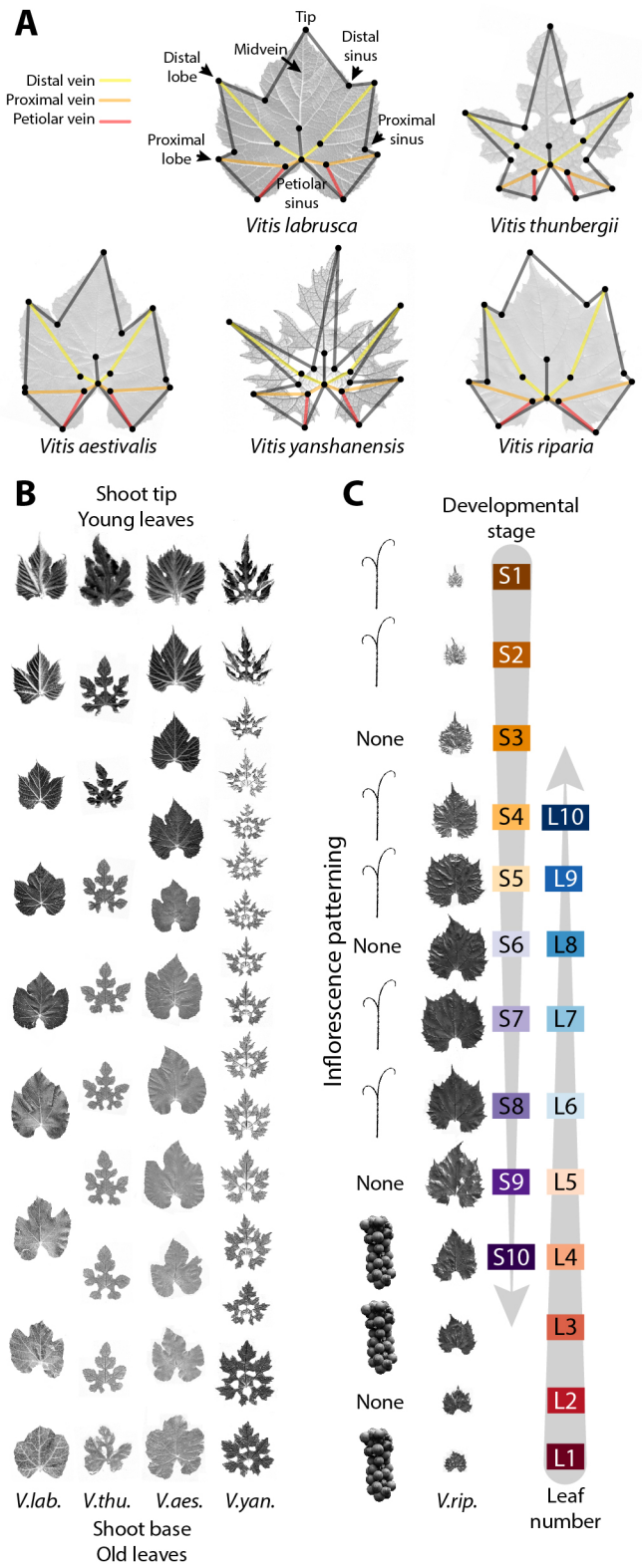
Supporting Information

Dataset S1: Procrustes-aligned coordinates. Wild *Vitis* species Procrustes-aligned coordinates can be found under the tab "wild_vitis." "id" indicates vine identification information, "species" the species, "file" the file from which the coordinate information was extracted, "image" which file in the series the image from which the leaf was extracted represents, "stage" the developmental stage of the leaf, "leaf_number" the position in the heteroblastic series of the leaf, "total_nodes" the total number of nodes found on the shoot from which the leaf was derived, "organ" the organ opposite the leaf at that particular node, and "x1" and "y1" through "x17" and "y17" indicate Procrustes-aligned coordinates for each leaf. Procrustes-aligned coordinates for *V. vinifera* subsp. *vinifera* data can be found under the tab "vitis_vinifera." "id" indicates vine identification information, "genotype" the genotype, and "x1" and "y1" through "x17" and "y17" indicate Procrustes-aligned coordinates for each leaf.

Dataset S2: Correlation between traits. P values ("p_values"), Benjamini-Hochberg adjusted p values ("BH"), and Spearman's rho ("rho") provided for the correlation between each pair of traits ("Trait1" and "Trait2").

Figure S1: Numbers of species, vines, leaves, and shoot positions sampled. (*A*) Leaves per species, arranged in descending order. (*B*) Vines sampled per species. For some analyses in the paper, only those *Vitis* species with ≥5 vines were analyzed (indicated by red line). (*C*) The average number of leaves sampled per vine for each species. (*D*) Number of leaves represented at each developmental stage. For some analyses, only developmental stage ≤ S10 were analyzed (indicated by red line). (*E*) Number of leaves represented at each leaf number position. For some analyses, only

859 leaf numbers ≤ L10 were analyzed (indicated by red line).



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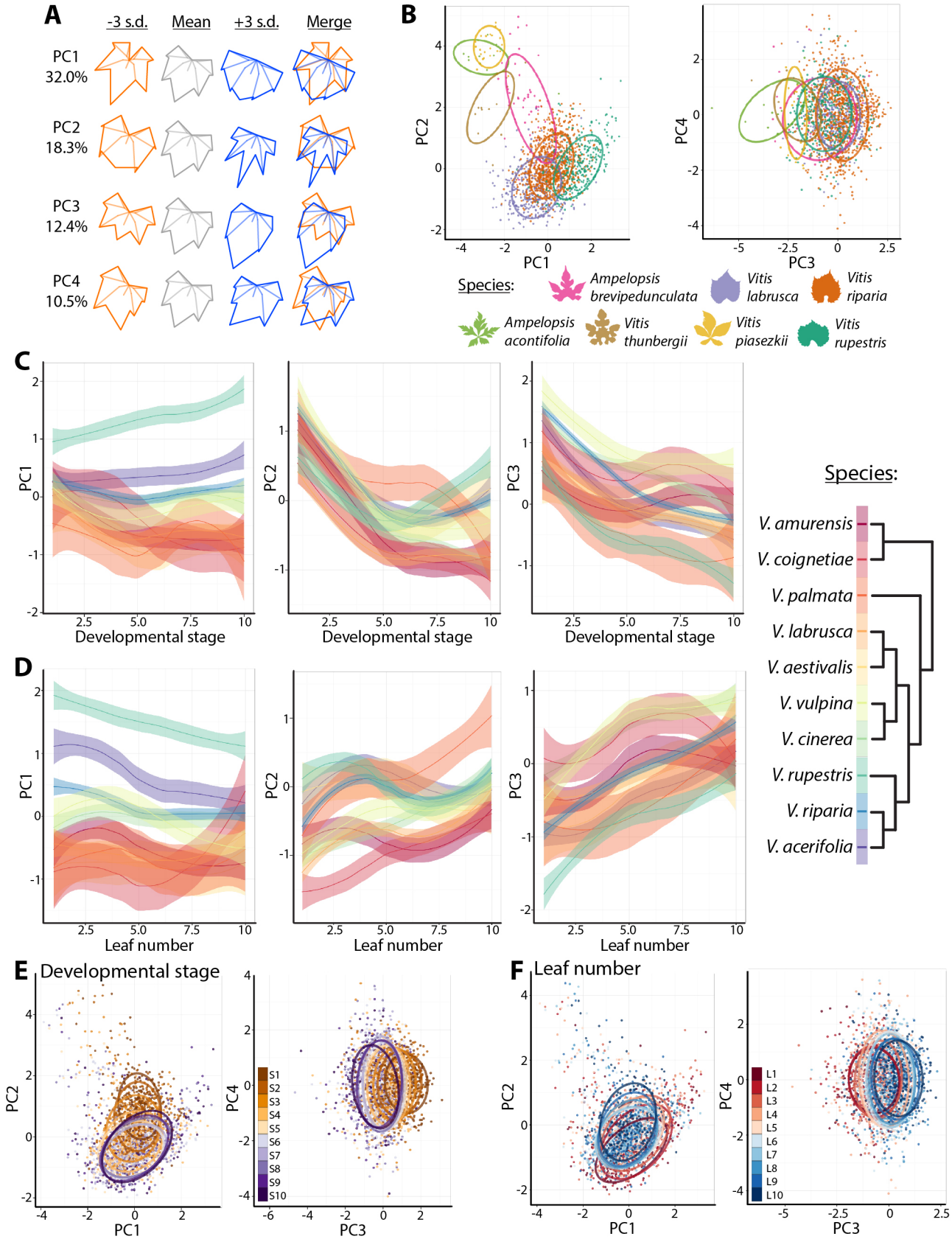

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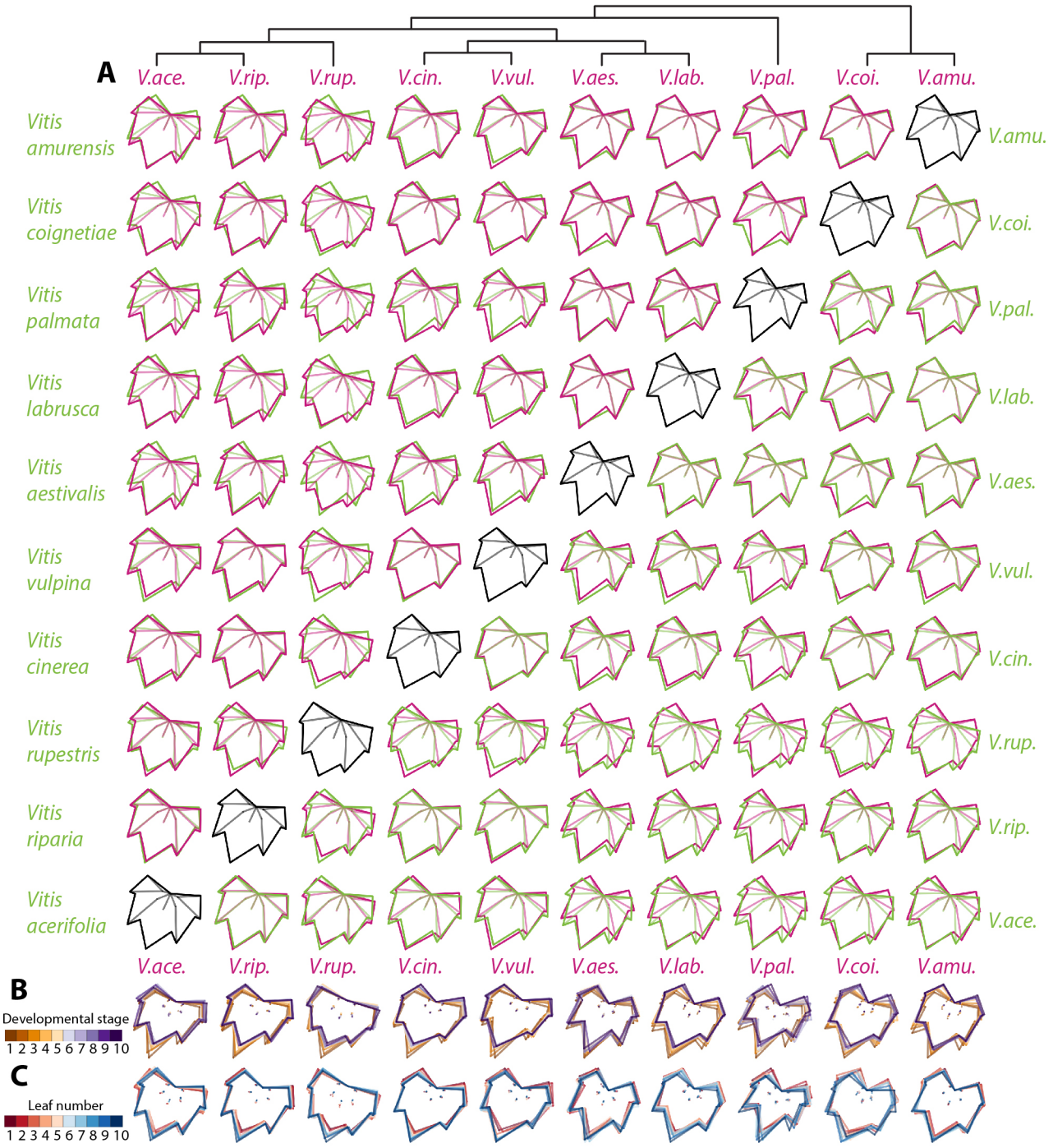
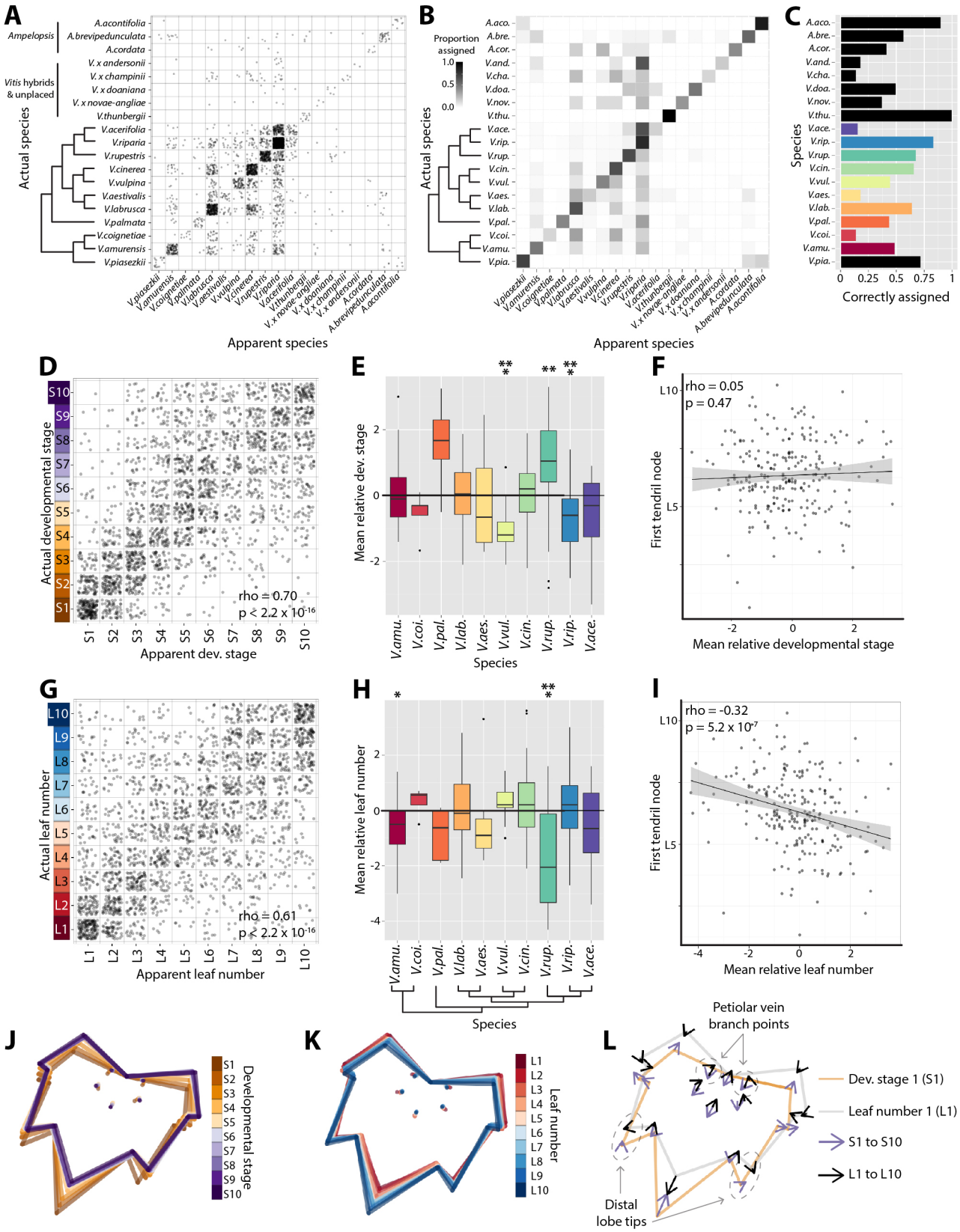


Fig. 3: Shapes represented among Vitis species, developmental stage, and leaf number. (A) Outlines representing the average shape of each pairwise comparison of species, indicated by magenta and green. Phylogenetic relationships are indicated. (B and C) Comparison of average shapes across (B) developmental stages and (C) leaf number for each species indicated by color.



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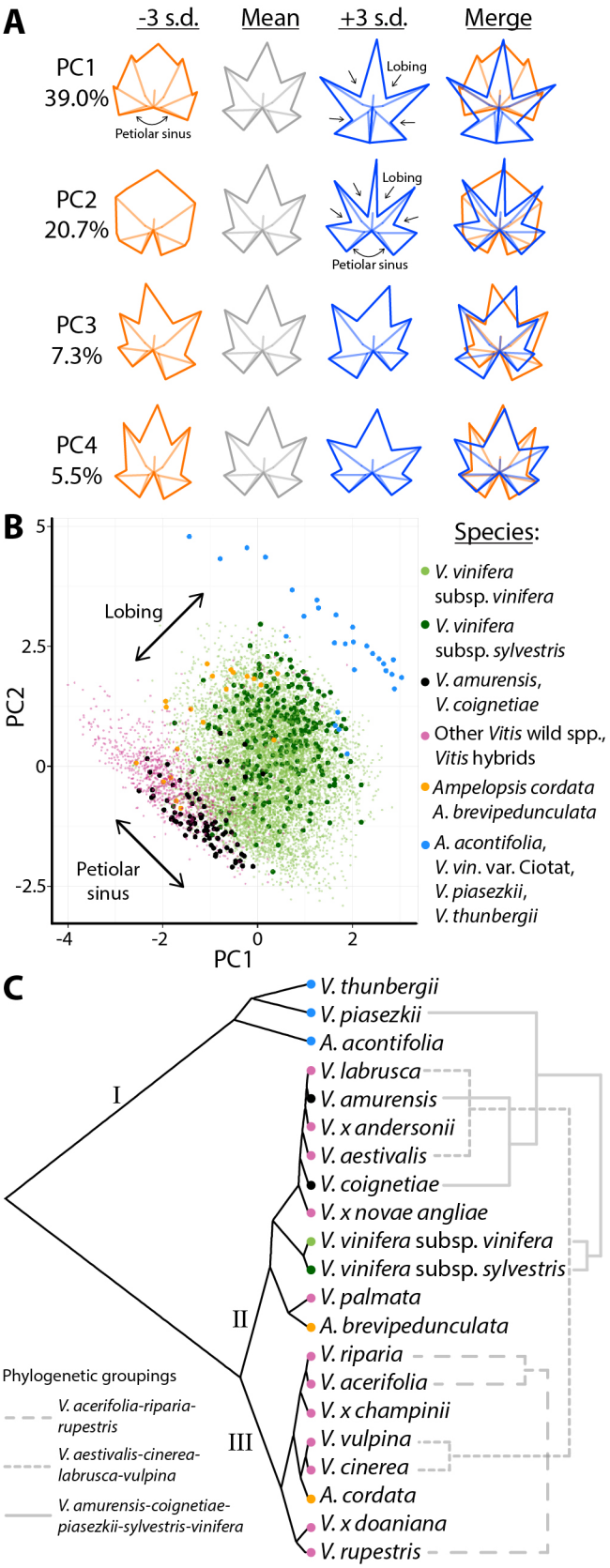

Fig. 5: Morphospace of wild Vitis species and domesticated grape. (A) "Eigenleaves" showing leaf morphs represented by PCs at -/+ 3 s.d. and shape variance explained by each. Lobing and petiolar sinus variation indicated. (B) Comparison of wild Vitis species leaf shape (red, black) with V. vinifera subsp. vinifera and V. vinifera subsp. sylvestris (green, dark green, respectively), and highly-lobed species (blue). Strong shape variation in lobing and petiolar sinus depth indicated. (C) Hierarchical clustering based on morphology. Phylogenetic groupings indicated in light gray. Groups mentioned in text indicated with Roman numerals.

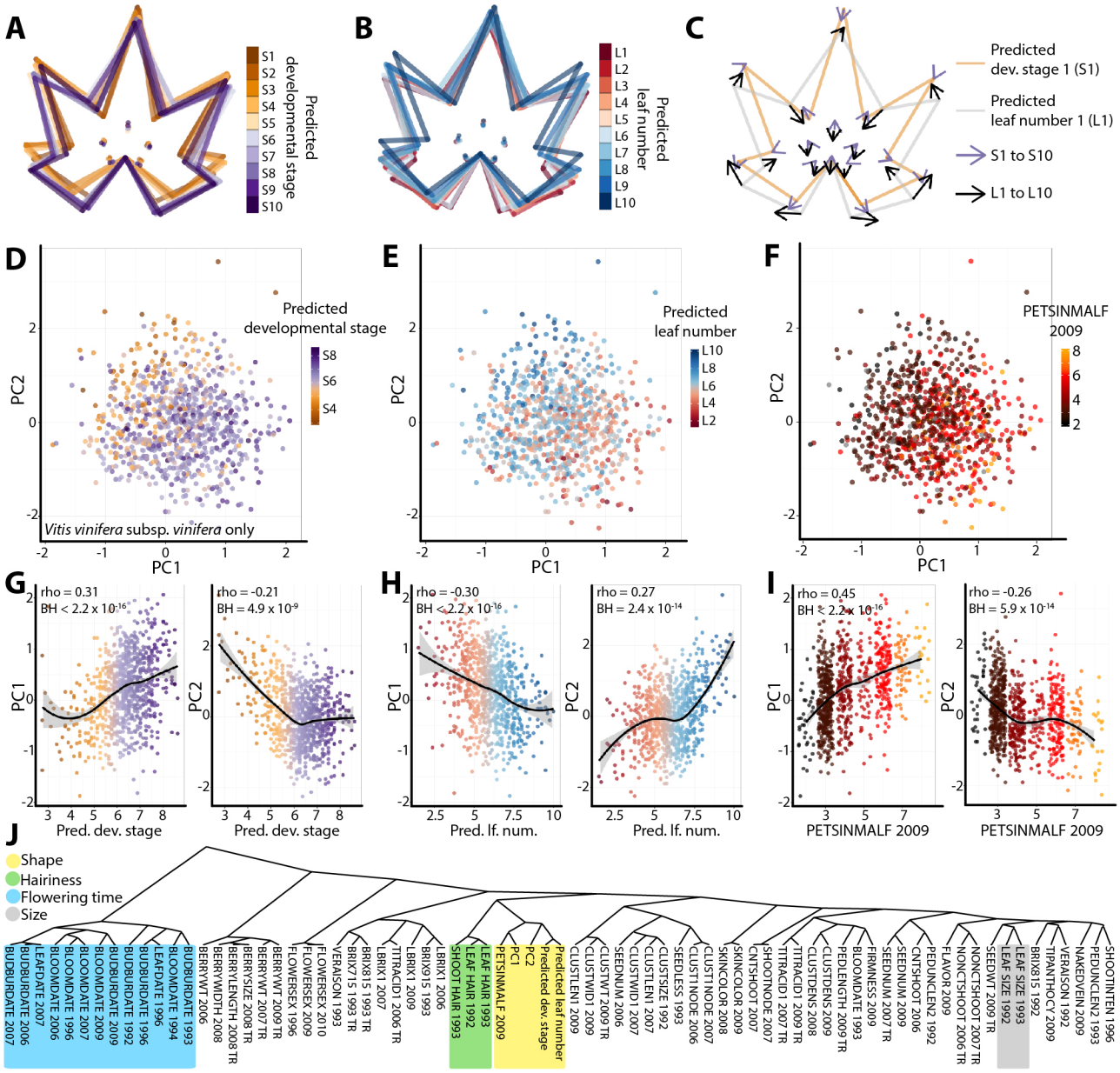

Fig. 6: Evolution in developmental timing underlies leaf shape diversity in domesticated grape. (A and B) Latent shape attributes defining developmental stage (A) and leaf number (B) in domesticated grape indicated with average leaf shapes. (C) Comparison of changes in latent shape attributes across predicted developmental stage (orange, outline S1; purple, arrow to S10) and predicted leaf number (grey, outline L1; black, arrow to L10). (D and E) Predicted developmental stage (D) and predicted leaf number (E) projected onto the V. vinifera subsp. vinifera morphospace demonstrates changes in developmental timing (heterochrony) underlie leaf shape diversity in domesticated grape. (G – I) Correlation between PC1 and PC2 with predicted developmental stage (G), predicted leaf number (H), and a qualitative measure of petiolar sinus depth, PETSINMALF 2009 (I). (J) Hierarchical clustering of measured traits in grape. Morphological and predicted temporal traits (yellow), hirsuteness (green), reproductive transitions (blue), and overall leaf size (grey) indicated.