bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

Understanding stem cell states: naïve to primed pluripotency in rodents and humans

Leehee Weinberger, Muneef Ayyash, Noa Novershtern & Jacob H. Hanna

The Department of Molecular Genetics, Weizmann Institute of Science, Rehovot 7610001, Israel.
Correspondence to: J.H.H (Jacob.hanna@weizmann.ac.il) and N.N (noa.novershtern@weizman.ac.il).

Preface
The molecular mechanisms and signalling pathways that regulate the *in vitro* preservation of distinct pluripotent stem cell configurations, and their induction in somatic cells via direct reprogramming approaches, continue to constitute a highly exciting area of research. In this Review, we provide an integrative synthesis on recent discoveries related to isolating unique naïve and primed pluripotent stem cell states with altered functional and molecular characteristics, and from different species. We overview pathways underlying pluripotent state transitions and interconversion *in vitro* and *in vivo*. We conclude by highlighting unresolved key questions, future directions and potential novel applications of such pluripotent cell states.

Introduction

Pluripotency describes cells that have the potential to give rise to cells from all three embryonic germ-layers and primordial germ cells (PGCs), but not extra-embryonic tissues[1]. While pluripotency is a transient cell state *in vivo*, pluripotent cells can be derived from different stages of early embryonic development and indefinitely maintained in an artificially induced self-renewal state *in vitro,* by supplementing exogenous cues[2]. Thus, it is important to stress that self-renewal is not a defining feature of pluripotency and is only transiently assembled during early development. Pluripotency is highly dynamic and evolves at different stages of pre- and post-implantation stages[3]. However, the self-renewal aspect is a highly useful *in vitro* artificial "engineering trick"[4] that has brought pluripotent stem cells to the front stage as a potential tool for tissue replacement, disease modelling and animal engineering technologies[5].

There are multiple pluripotent stem cell types that can be isolated from vertebrates, including rodents and human, typically annotated based on their donor cell of origin **(Fig. 1)**. Embryonic stem cells (ESCs) are isolated from the inner cell mass (ICM) of developing pre-implantation mouse or human blastocysts[6,7]. Epiblast stem cells (EpiSCs) are isolated from mouse post-implantation epiblasts[8,9], however equivalent derivations have not been attempted with human embryos due to ethical complexities. Early rodent migrating PGCs can be converted *in vitro* into pluripotent ESC-like cells termed embryonic germ cells[10]. Mouse neonatal and adult spermatogonial stem cells can be reverted toward pluripotency and generate male germ stem cells (GSCs)[11-14]. The latter have the disadvantage of retaining only male imprint signature, which compromises their safety for clinical use[13]. Intriguingly, indefinitely stable and validated EGs and GSCs have not been isolated from primates thus far[15,16] (**Fig. 1**).

Somatic cell reprogramming provides alternative routes for isolating pluripotent cell types. Human and rodent somatic cells can be artificially reprogrammed into ESC-like cells following reprogramming via nuclear transfer, termed NT-ESCs[17-19]. Ten years ago, Yamanaka established direct *in vitro* reprogramming of somatic cells to pluripotency via ectopic expression of defined factors[20], that yield induced pluripotent stem cells (iPSCs) without the need for oocytes or

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

embryos[21-24] **(Fig. 1)**. NT-ESCs and iPSCs offer the advantage for being able to generate patient specific pluripotent cells with genomic DNA identical to the donor somatic cell, however mitochondrial DNA in NT-ESCs is non-isogenic and provided by the a-nucleated donor oocytes[25]. The latter can be an advantageous in settings aiming at correcting maternally inherited mitochondrial diseases[26-28].

While the above overview pertains to classify different pluripotent cell types based on their tissue derivation source, the growth conditions used to expand such cells dictate the pluripotent state they attain (ICM-like, ESC-like, EpiSC-like state etc.)[29,30]. iPSCs generated in classical mouse ESC growth conditions yield ESC-like iPSCs; while those reprogrammed in EpiSC growth conditions yield EpiSC-like iPSCs[29,31]. The same analogy applies to explanting rodent ICM cells in ESC or EpiSC growth conditions[29,32]. In comparison to developmentally restricted mouse EpiSCs, ESCs are highly competent in generating high-contribution chimeric mice after microinjection into host-blastocysts, retain a pre x-inactivation state in female cell lines and reduced expression of lineage commitment factors[29,30]. Such attributes influence the utility of pluripotent cells in cell differentiation, animal transgenics and molecular disease modelling. Thus it is of critical importance to understand and define different pluripotent states and configurations, and their implications on pluripotent cell differences across different species[4,33].

In this review we provide an integrative perspective on recent breakthroughs in understanding the diversity and complexity of pluripotent state regulation *in vitro*. This includes advances on preserving naïve pluripotency from non-rodent species and alternative EpiSC pluripotent states. We highlight unresolved issues, key questions and future directions in this exciting front of stem cell research.

Murine pluripotent states

Mouse ESCs were thoroughly shown to reside in an ICM-like state[29], now referred to as the naïve state of pluripotency[30], since they retain many of the molecular characteristics of ICM. In 2007, Mckay and Vallier groups derived a novel type of pluripotent cells termed EpiSCs from post-implantation rodent epiblasts[8,9]. In comparison to naïve ESCs, EpiSCs retain an alternative pluripotency configuration, commonly referred to a primed pluripotency[30]. Dramatic molecular and functional differences exist between different pluripotent cells, which subsequently influence their characteristics, function and safety.

Growth conditions for naïve pluripotency

To fully grasp the biology of mouse naïve ESCs and their developmental context, it is of relevance to review the evolution of growth conditions devised to isolate such cells over the course of the last thirty years. Martin and Evans derived ESCs from 129-mouse strain[7,34], by utilizing mitotically inactive embryonic fibroblast cells (MEFs) as feeder cells and foetal bovine serum (FBS). Leukaemia Inhibitory Factor (LIF) that activates the STAT3/JAK pathway was later identified as a key ingredient that allowed expansion of mouse ESCs in FBS/LIF conditions without MEFs[35,36] **(Fig. 2)**. Such naïve mouse ESCs express hallmark pluripotency factors (e.g. Oct4, Nanog, Esrrb) and retain a pre-x inactivation state in female cell lines[37] **(Fig. 3)**. Functionally, ESCs can populate the host pre-implantation mouse ICM upon microinjection, and generate high-contribution chimeras with colonization of the germ-line niche[38,39].

Smith group described the first serum- and feeder-free defined conditions for expanding mouse ESCs by combining low dose BMP4 with LIF[40]. Addition of small molecule inhibitors for MEK signalling, increased ESC derivation efficiency and stability[41]. Developing different defined conditions, all involving MEK inhibitors, which can be used to isolate murine ESCs, extended the latter observations. A cocktail combing 3 inhibitors termed '3i conditions', was shown to

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

stabilize pluripotent cells without LIF, indicating existence of redundant pathways to isolate ESCs *in vitro* that can compensate for lack of LIF/Stat3 signalling[42]. Notably, such cell configuration was labelled as "ground state pluripotency" since the cells in 3i were reported to grow independent of any exogenous signalling stimuli. However, this term is challenged by the fact that growth in these conditions heavily relies GSK3 inhibition that mimics WNT stimulation; and on exogenous insulin that activates PI3K/AKT signalling[42]. Further, 2i/LIF conditions were adopted as an enhanced mean to expand murine ESCs, and the reduced proliferation in 3i conditions indicates a role for autocrine secreted FGF4 in promoting naïve ESC growth[43] **(Fig. 2)**.

"Alternative 2i" conditions, involving small molecule inhibitors for Gsk3 and Src pathways, have been shown to yield germ-line competent ESCs[44] **(Fig. 2)**. Paul and colleagues identified atypical PKC small molecule inhibitor Go6983 (aPKCi) as another stimulator for isolating murine ESCs together with LIF and/or MEK inhibitors[45]. Single cell RNA-seq analysis has proven equivalent global heterogeneity between different naïve conditions, however the difference exists in the identity of genes that underlie heterogeneity in each condition[46].

Enriched conditions were important for deriving ESCs from other mouse strains that have until recently been considered "non-permissive" for deriving naïve ESCs. While 129-mouse strain derived ESCs could be expanded in FBS/LIF conditions, for other mouse strains like non-obese-diabetic ICR mice (NOD), supplementation of 2i or GSK3 inhibitors is essential for both derivation and maintenance[29]. 3i and 2i/LIF conditions yielded rat ESCs, however these conditions are suboptimal[47,48]. LIF/MEKi/aPKCi have been recently described as a more robust method to support rat ESCs in feeder free conditions[49] **(Figure 2,3)**.

The above discoveries underscore the relevance of analysing rodent ESCs expanded in different naïve conditions and from different genetic backgrounds. Further they emphasize the importance of other signalling pathways that remain to be deeply characterized in the context of pluripotency **(Fig. 2)**. NF-kB inhibition has been implicated as a downstream effector underlying aPKC inhibition mediated support of naïve pluripotency[45], however other pathways, like Mbd3/NuRD repressor complex, can be neutralized by aPKCi (unpublished observations, J.H.H). SRC functions as a downstream target of ERK and Calcineurin-NFAT signalling to promote ESC differentiation[50], and its inhibition promotes naïve pluripotency[44,50].

The Hippo signalling pathway has been uncovered as cardinal regulator acting at epiblast vs. trophoblast segregation in late mouse morulas. The Hippo pathway is highly active in pluripotent epiblast cells, leading to exclusion of Yap/Taz effectors from the nucleus[51,52]. Piccolo and colleagues indicated that Yap/Taz depletion in mouse naïve ESCs expanded in 2i/LIF prevents their differentiation[52], while Lian et al. have indicated Yap/Taz as essential regulators for stability of naïve ESCs expanded in FBS/LIF conditions [53]. Systematic analysis of these findings in different defined conditions might resolve these seemingly opposing results (**Fig. 3**).

It should be noted that signalling pathways are often pleiotropic and may simultaneously have positive and negative effects on naïve pluripotency. For instance, nuclear β-catenin stabilization following Gsk3 inhibition, promotes naïve pluripotency via neutralizing the repressive activity of Tcf3 on its bound target genes in the nucleus[54]. Cytoplasmic β-catenin promotes naïve pluripotency via increasing E-cadherin membrane stability[55] (**Fig. 2**). However, nuclear β-catenin can induce mesodermal gene expression through its Lef co-effectors[47]. Yet, such differentiation priming effects are outweighed by naïve pluripotency promoting functions of β-catenin under optimized conditions. LIF has also been shown to promote primitive endoderm

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

specification in naïve pluripotency growth conditions[56]. Such "non-purist" effects should be kept in mind when dissecting the role of signalling pathways on pluripotency.

The ample conditions to grow naïve murine ESCs have been important for better understanding and revisiting the roles of several classical pluripotency regulators. While Nanog was first purported to be absolutely essential and irreplaceable for establishing naive pluripotency through iPSC reprogramming, cell fusion or EpiSC reversion[57], multiple conditions enable reprogramming of Nanog null donor cells *in vitro*[58,59]. Still however, Nanog null ESCs cannot be derived from mouse ICMs, indicating that while Nanog is indispensible for establishing pluripotency *in vivo*, it is dispensable during *in vitro* induction and maintenance by alternative pluripotency regulators like Esrrb and enriched growth conditions[60]. The latter example highlights that *in vitro* pluripotency maintenance and induction can not be considered "authentic", as some *in vitro* conditions can clearly potentiate the robustness of the naïve pluripotency program and compensate for deficiencies that are not sustainable *in vivo*. Similarly, Klf2 knockout embryos do not present lethality at the pre-implantation stage, and naïve ESC in FBS/LIF conditions can tolerate Klf2 ablation[61]. However ESCs in 2i only conditions, can no longer tolerate loss of Klf2[61], and LIF can compensate for the lack of Klf2.

Another emerging regulatory principle is that not all factors expressed in the ICM or ESCs necessarily promote naïve pluripotency, and some of them in fact are promoting its dissolution. However, they are tolerated by ESCs *in vitro* due to the optimized and enriched *in vitro* growth conditions used. For instance, Tcf3 binding represses the expression of its naïve pluripotency promoting target genes, leading to their partial repression, is tolerated in serum/LIF naïve conditions. However, Tcf3 neutralization by adding GSK3i boosts naïve pluripotency[54]. In a similar manner, mouse ESCs tolerate Mbd3/NuRD complex expression although it partially represses naïve pluripotency targets like Esrrb and Tfcp2l1[62]. However, genetic ablation of Mbd3 leads to upregulation of master regulators of naive pluripotency and allows LIF independent growth[62]. Consistently, derivation of Mbd3 KO ESCs from null ICMs is uncompromised in 2i/LIF conditions[63]. In summary, both Tcf3 and Mbd3 are expressed in the ICM and ESCs likely to set the stage for terminating the naïve pluripotency program. Thus the molecular characteristics of a pluripotent state fixated in a certain condition represent the net outcome of conflicting stabilizing and destabilizing factors simultaneously residing and conflicting in that state[64].

Collectively, it is important when analysing function of pluripotency regulators to systematically compare different naïve growth conditions, genetic backgrounds and *in vivo* context[65]. Such integrative analysis will likely unravel additional layers of underappreciated complexity and may resolve some conflicting results that stem for conducting analysis on pluripotent states with different molecular flavours[52,53].

Growth conditions for primed pluripotency

Primed EpiSCs were derived from post-implantation epiblasts of a variety of rodent strains in FGF2/Activin A conditions[8,9] (**Fig. 1**). EpiSCs are capable of differentiating into cells of all three germ layers *in vitro* or in teratoma assay, and thus are pluripotent. However, they are inefficient in yielding chimeric animals once injected in pre-implantation epiblasts (**Fig. 3**), likely because they have altered molecular characteristics and correspond to a more advanced developmental stage in comparison to the host pre-implantation environment[8,9].

While EpiSCs maintain Oct4 and Sox2 expression, they down-regulate most of the other pluripotency factors including Nanog, Esrrb, Klf2 and Klf4[3]. EpiSCs have not undergone differentiation, but they upregulate lineage commitment factors like Otx2, Brachyury and Zic2[66].

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

Epigenetically, EpiSCs retain distinct characteristics from naïve ESCs: they inactivate X chromosome in females, upregulate global DNA methylation levels and acquire H3K27me3 at developmental regulators[67,68]. Enhancer landscape and activity is rewired between naive and primed states[66], and developmental regulator gene associated "seed enhancers" convert from a dormant to an active state in EpiSCs, thus pre-marking differentiation tendency of primed PSCs[69]. Summary of divergent signalling and molecule characteristics between murine primed and naïve cells are highlighted in **Fig. 2-3**.

At the regulatory level, naïve and primed pluripotent cells have been shown to retain opposing dependence on epigenetic repressors[65] (**Fig. 4**). Naïve ESCs tolerate loss of epigenetic repressors like Dnmt1, Dicer, Dgcr8, Eed, Mbd3 and Mettl3[65], and in fact renders these cells "hyper–naïve" and resistant to differentiation[65,70,71] (**Fig. 4**). On the contrary, murine primed pluripotency maintenance and viability depends on these regulators, and their ablation destabilizes the murine primed pluripotent state[65] (**Fig. 4**). Defining how the depletion of each of these repressors precisely destabilizes the primed configuration is of future interest.

Alternative growth conditions to expand murine EpiSCs have begun to emerge. Elegant work by Ying and colleagues SHOWED THAT simultaneous use of a GSK3 inhibitor (that induces β-catenin stabilization), together with a Tankyrase small molecule inhibitor IWR1 (that up regulates Axin1/2 levels thus leading to retention of β-catenin in the cytoplasm) maintain novel primed EpiSCs without exogenous Fgf2/Activin A supplementation[72] (**Fig. 3**). Removal of IWR1 leads to increased nuclear β-catenin shuttling and EpiSC differentiation[72]. The mechanisms by which cytoplasmic β-catenin prevents EpiSC differentiation remains to be uncovered[72]. It is tempting to speculate whether the recently described ability of cytoplasmic APC/Axin/β-catenin destruction complex to act as a sequestration "sink" for Yap/Taz and prevent their nuclear shuttling[52], is involved in the ability of GSK3i/IWR1 conditions to maintain EpiSCs. Notably, the latter alternative EpiSC state is different from EpiSCs expanded in classical Fgf2/Activin A conditions, and retains higher expression of naïve markers like Dppa2, 4, 5[72], and are thus relatively "less primed" (**Fig. 3**).

Recent studies indicate that different primed conditions used can endow EpiSCs with region specific characteristics of the post-implantation epiblasts. Tam and colleagues showed that EpiSCs expanded in Fgf2/Activin A correspond transcriptionally and functionally to anterior late-gastrula primitive streak cells[73]. Belmonte and colleagues showed that alternative FGF2/IWR1 conditions generate murine EpiSCs corresponding rather to posterior-proximal epiblasts[74]. Further, even in classical Fgf2/Activin A conditions distinct subpopulation of EpiSCs can co-exist, each representing different stages of post-implantation embryonic development[75].

Finally, the time length at which pluripotent cells are maintained under primed conditions greatly influences their characteristics and functionality[76]. Counter intuitively; while murine PGCs are specified from the post-implantation epiblast *in vivo*[77], EpiSCs maintained *in vitro* for more than 7 days in FGF2/Activin A, lose competence to generate PGCs in response to Bmp4[76]. Starting with naïve cells and inducing priming for no longer than 2-4 days, yields distinct primed cells highly competent for generating PGCLCs, termed EpiLCs[76]. The latter are transcriptionally more similar to *in vivo* post-implantation epiblast than EpiSCs[76]. Thus, the above paradigm indicates another aspect of artificial features that can be acquired by pluripotent cells once expanded indefinitely *in vitro,* in contrast to their *in vivo* "counterparts" that transiently exist during development.

Studies involving clonal lines and single cell analysis will be key for deeper understanding of features of region specific EpiSCs and shortly after *in vitro* induction from a naïve state in

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

different priming conditions[73]. This may help understand how lineage priming is established at the single cell level during these key early developmental transitions[73,74] and might be relevant for optimizing other differentiation protocols and predicting PSC behaviour.

Interconversion between naïve and primed states

As somatic cells can be reprogrammed into a naïve ESC-like state via combined overexpression of pluripotency factors together with LIF, primed EpiSCs can also be reverted to naive iPSCs. Overexpression of Klf4 or Myc in EpiSCs, under LIF containing conditions, generates naïve ESCs[29,78]. FBS/LIF signalling alone can be sufficient to induce such conversion from permissive mouse genetic backgrounds (i.e. 129 strains)[79], but not from "non-permissive" ones like NOD, where supplementation of small molecules like 2i is necessary[29]. Other factors like Nanog, Prdm14 and Esrrb have been shown to synergistically induce and boost the efficiency of this process[80,81]. Explanting post-implantation E5.5-E7.5 epiblasts in naïve conditions also reverts them into naïve PSCs[29,79]. The opposite conversion is attainable from *in vitro* and *in vivo* isolated naïve cells, as expanding murine naïve PSCs or ICMs in primed conditions leads them to gradually adapt an EpiSC state[29,78,82].

Studies focusing on *in vitro* molecular changes accompanying naïve to primed pluripotency conversion have unravelled key events in understanding mechanisms of reprogramming[67]. Naïve ESCs expanded in 2i/LIF retain global hypo-methylated levels in both promoters and gene bodies, highly similar to those measured in ICMs[83,84]. When transferred into LIF/FBS naïve conditions, this is accompanied by an increase in global DNA methylation levels, however promoter and enhancer regulatory regions remain protected from invasion by DNA methylation[85]. Only after transfer into primed Fgf2/Activin A EpiSC inducing conditions, DNA methylation accumulates over enhancer and promoter regulatory elements[85].

Transitioning primed EpiSCs or naïve FBS/LIF PSCs into 2i/LIF conditions leads initially to dramatic changes in Oct4, Sox2 and Nanog pluripotency factor occupancy[86]. Changes in H3K27me3 deposition and enhancer landscape follows only later, likely in response to the rewiring in transcription factor binding[86]. Downregulation in DNA methylation follows next, which has mainly been attributed to downregulation in *de novo* DNA methyltransferase enzymes[83]. It should be noted however, that ablation of Dnmt3a/b in ESCs in FBS/LIF condition does not lead to such rapid loss of DNA methylation[87], and other yet to be identified events might be involved in this rapid 2i induced epigenetic response. MEK/ERK inhibition influences polycomb interactions and leads to decreased occupancy of PRC2 and RNA PolII at developmental regulatory genes[88], leading to loss of H3K27me3 and increased PolII pausing at bivalent developmental regulatory genes[67].

Analysis of other molecular changes in naïve to primed transitions is likely to increase our knowledge of pluripotency regulation and lineage priming. Analysing other defined naïve pluripotency growth conditions (2i/LIF/PKCi, alternative 2i etc.) and in other rodents, will be important for discerning redundancies and specificities of different singling pathways and how they cross-talk with chromatin organization.

Human conventional pluripotent cells

Thomson group first isolated human ESCs in 1998 from bastocysts[6]. Surprisingly, they were drastically different from murine ESCs in their characteristics and tissue culture requirements. FGF2 and TGFβ1, but not LIF, signalling are at the core signalling modules maintaining such conventional human ESCs derived from the ICM, or iPSCs obtained via direct *in vitro* reprogramming[89].

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

A primed pluripotent state

Differences between conventional human and mouse ESCs had been initially attributed only to unknown species genetic differences, since human ESCs were also derived from the ICM and not from post-implantation stages. However studies on different mouse strain derived stem cells have discerned a scenario where ICM cells can adapt *in vitro* into a primed state if naïve conditions that match the requirements of the particular genetic background of donor embryos used, are not devised[29]. Specifically, NOD mice are relatively "less-permissive" than 129 mice to yield naïve ESCs/iPSCs, as LIF alone is not sufficient to maintain NOD naïve pluripotency and 2i/LIF are permanently required to stabilize and maintain this state *in vitro* in NOD PSCs[29]. Further, ICMs from both 129 and NOD strain expanded in primed conditions yield EpiSCs that are indistinguishable from EpiSCs derived from E6.5 embryos[29] or *in vitro* from already established ESCs[29].

The relevance of the latter *in vitro* priming scenario to dictating conventional human ESC identity is supported by the fact that human conventional ESCs/iPSCs retain a great milieu of primed pluripotency features. This includes low expression of naïve pluripotency markers (e.g. KLF5, TFCP2L1, DPPA3), deposition of H3K27me3 over developmental genes, lack of exclusive nuclear localization of TFE3, loss of pluripotency upon inhibition of MEK/ERK pathway, lack of global hypomethylation as seen in ICM cells, lack of a pre x-inactivation state in most conventional female PSCs lines[68,90,91]. Further, human primed ESCs do not tolerate complete loss of DNMT1[87], similarly to what has been shown for mouse EpiSCs[65] (**Fig. 4**). Complete human KO ESCs have not been obtained thus far for DICER, MBD3 or METTL3[63,65] (**Fig. 3**).

Less primed than murine EpiSCs

In spite of the above, it is of critical importance to realize that human conventional/primed ESCs are not identical to murine EpiSCs, and can be considered relatively "less primed". For instance, human ESCs do not upregulate FGF5 and N-CADHERIN as seen in murine EpiSCs, and express high-levels of E-CADHERIN as detected in mouse naïve ESCs[68]. Human ESCs express high levels of some naïve markers like NANOG, PRDM14 and REX1 that are not expressed in murine EpiSCs[92]. Moreover, human primed ESCs are functionally dependent on NANOG and PRDM14 and their ablation induces differentiation[92]. DNA methylation distribution in human ESCs shows they rather correspond to murine naïve ESCs expanded in FBS/LIF conditions, rather than mouse FGF2/Activin A expanded mouse EpiSCs, as their promoters are protected from invasion by repressive DNA methylation[85,93]. Further, while murine EpiSCs demonstrate exclusive TFE3 cytoplasmic localization and naïve 2i/LIF ESCs show exclusive nuclear TFE3 localization[94], human primed ESCs show an intermediate configuration where TFE3 is present in both the cytoplasm and the nucleus[68].

Human naive pluripotent cells

The metastability of naïve and primed pluripotent state depending on the growth conditions applied[29], and the stringency in requirement for exogenous naïve pluripotency promoting factors to isolated naïve PSCs from previously "non-permissive" rodent strains[29,30], have underscored a scenario of whether unique and more stringent conditions can be applied to isolated previously unidentified alternative naïve-like pluripotent states in humans.

Transgene-dependent generation

2i/LIF conditions are not sufficient to maintain naïve human ESCs or iPSCs[95]. However, additional transgene expression can induce an artificial transgene dependent state that may be of considerable interest. Continued exogenous OCT4/KLF4 or KLF2/KLF4 transgene expression

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

can maintain human ESCs/iPSCs in a unique pluripotent state in 2i/LIF conditions[95]. Recently, these observations were extended by optimizing over-expression of KLF2/NANOG transgenes, allowing expansion of human naïve iPSCs in 2i/LIF[96]. Smith and colleagues overexpressed KLF2 and NANOG transgenes in primed ESCs and expanded them in 2i/LIF/aPKCi conditions [97]. These cells exhibited more extensive DNA demethylation, and strong upregulation of naïve markers like TFCP2L1, KLF2, and KLF4. However, as KLF2 is not expressed in the human ICM[98], and as 2i/LIF/aPKCi are insufficient to convert primed ESCs without exogenous transgene induction[97], and as transgene-free cells remain to be validated in 2i/LIF/aPKCi conditions, it is unclear whether this state is indefinitely stable without retaining leaky transgenes and/or MEFs as claimed by the authors. Further, independent examination of DNA methylation landscape in reset cells indicates aberrant global loss of imprinting, excessive hypomethylation of endogenous retroviral genes[90,99]. Finally, while these growth conditions do not contain exogenous FGF or TGFβ1/Activin A cytokines, applying short-term inhibition of FGFR/TGFR signalling is not sufficient evidence to validate FGF/TGF/Activin A signalling independence[97] (**Fig. 2**).

While the field has shifted to study transgene independent conditions as detailed below, it should be noted that such transgene dependent states might be important, since it may be possible that robust naïve pluripotency currently obtained in mouse ESCs is a rodent specific phenomenon. Capturing human naive PSCs identical to those obtained from mice might still involve genetic modifications. Nevertheless, the latter studies provide evidence for the possibility to generate naïve-like pluripotent states in humans and other species[29,95].

Transgene independent generation

Our team was the first to describe naïve conditions, designated as NHSM conditions, which entail complete ablation of MEK/ERK signalling and are compatible with indefinitely expanding genetically unmodified human PSCs both in MEF- containing and -free conditions[68]. These naïve MAPK independent pluripotent cell lines could be derived from human pre-implantation embryos, through *de novo* iPSC generation, or from previously established primed ESCs/iPSCs[68]. NHSM conditions contain 2i/LIF together with P38i, JNKi, aPKCi, ROCKi, low doses of FGF2 and TGFβ1 (or Activin A), and render human PSCs more similar, but not identical, to murine naïve PSCs[68] (**Fig. 2,3**). In fact these cells have features of so called naïve 2i/LIF "ground state" pluripotency, which are not found even in naïve mouse ESCs expanded in FBS/LIF. This includes exclusive nuclear localization of TFE3, and cleansing of H3K27me3 over developmental genes[67,68,94]. Transcriptionally, these cells down regulated expression of lineage commitment markers like OTX2, ZIC2 and CD24 and moderately upregulated pluripotency genes (more prominently on MEFs)[68,100]. Enhancer rewiring has been attained in these human naive PSCs as seen with mouse cells[69]. The cells exhibited downregulation in DNMT3B[101] and a mild global decrease in global DNA methylation levels, while maintaining imprinting integrity and chromosomal stability[68]. While conducting chimeric analysis with human PSCs and using human embryos as hosts is ethically and legally forbidden, these human naïve PSCs cells showed better integration upon microinjection into host mouse morulas and were able to contribute at low-grade levels in mouse embryos up to E17.5[68] (**Fig. 3**).

Important publications describing alternative conditions that yield human ERK independent naïve pluripotent cells emerged soon after, each producing cells with different enhanced molecular properties (**Figure 3**). A combination of 2i/LIF, ROCKi, BMPRi, high doses of FGF2 and TGFB1 were able to maintain human PSCs only in the presence of MEFs[102]. These cells demonstrated transcriptional upregulation of pluripotency markers like STELLA and KLF5. Jaenisch team described conditions[96] that adopted most of components found in NHSM[68] (i.e. 2i/LIF, ROCKi, Activin A (instead of TGFβ1) - with or without FGF2 and JNKi) and

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

supplemented inhibitors for BRAF and SRC pathways (conditions termed 5i/LA-MEF with optional inclusion of JNKi or FGF2). In comparison to the previous two studies, cells in 5i/LA-MEF conditions demonstrated a more impressive upregulation of naïve pluripotency markers. However, the cells did not down regulate DNMT3B, maintained an inactive X chromosome state in female cell lines, and demonstrated an unusual expression pattern of early pre-ICM genes[96]. Intriguingly, the process of converting primed cells back to naïve state in 5i/LA-MEF conditions is inefficient, taking two weeks to isolate initial clones that retain a very slow growth rate[96]. Further, these conditions exclusively yield chromosomally abnormal cell lines[96]. Thus, it remains to be determined whether such chromosomal abnormalities are in fact inherent to 5i/LA-MEF cells and dictate the properties described for this state[96], and are being selected for during this inefficient conversion process. Finally, DNA methylation profiling and whether these cells maintain epigenetic imprinting integrity is another important aspect that remains to be evaluated.

It is clear from the above summaries that none of the many already published conditions generate human naïve PSCs that are identical to mouse ESCs or human ICM[98,103,104]. However, these studies implicate new signalling pathways and pave avenues for further optimization and characterization of such novel PSCs (**Fig. 2**). Mechanistically it will be interesting to test whether there is a connection between RAF and aPKC inhibition. Further, RHO signalling has been shown to promote YAP/TAZ nuclear localization in primed human ESCs and sustain their pluripotency[105]. Thus it remains to be defined whether ROCKi influencing naïve pluripotency characteristics[68] via YAP/TAZ modulation.

The role of FGF2, ACTIVIN A, TGFβ1 signalling, either autocrine or exogenously provided at low doses, remains to be understood in human naïve PSCs. The latter demonstrate upregulation of Activin like ligand GDF3[97], and human, but not mouse, ICM cells abundantly express Activin receptors[98]. Thus, it is tempting to speculate that primed human ESCs are relatively less primed than murine EpiSC due to differences in response to Activin-like ligands, where it might promote some naïve features in human[68,96], but not in mouse. To conclude, systematic analysis of the response of pluripotent states from different species to a variety of TGF ligand family members is of importance (**Fig. 2**), while the possibility to generate human PSCs that are entirely independent of FGF and/or TGF signalling cannot be excluded.

Differences between mouse and human epiblasts

Recent studies focusing on single cell RNA-seq of human pre-implantation embryos are starting to provide answers to some of the questions highlighted above. While human and mouse blastocyst do not display morphological differences, they retain profound molecular differences at the cellular level[98] (**Fig. 5a,b**). Human ICM epiblast cells do not express genes that are considered important pluripotency factors in mouse, such as KLF2 and ESRRB. Instead, KLF17 might have a human-specific role in the ICM[98]. ERAS, an ESC specific form of RAS that is constitutively active, is not expressed in humans, as it became a pseudo-gene[106]. Eras null mouse ESCs propagate slowly in FBS/LIF conditions[106], and whether they can be expanded in 2i/LIF conditions remains to be defined.

At the post implantation stage, dramatic differences exist between rodent and human embryos (**Fig. 5a,b**). Rodents are rather unusual as their post-implantation epiblast assumes egg-like cylinder shape, while in humans the post-implantation epiblast assumes a flat disc shape, like in most other mammals[107]. While it might be impossible to conduct single cell analysis on early human post-implantation epiblasts, non-human primates might provide some relevant insights. Collectively, these species differences might directly influence the distinct pluripotent characteristics observed in PSCs from different species *in vitro* and their distinct growth

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

requirements. Further, they are of relevance for understanding the developmental context of human *in vitro* isolated pluripotent cells.

A framework for classification of pluripotent states

The advent of different conditions to isolate human naïve PSCs with distinct characteristics, and the limitations in conducting chimeric analysis in humans, simulate discussions and perspectives on classifying pluripotent states. It is often claimed that ability to derive ESCs for human ICMs in a newly devised growth condition constitutes "a gold standard" for proving naivety[108]. However, it should be kept in mind that the pluripotent state identity is eventually dictated by the derivation growth condition and not by whether their source was from the pre- or post-implantation epiblast[29,82]. Utilization of OCT4 distal vs. proximal enhancer element as a binary distinguishing marker can be also misinterpreted[96]. Both OCT4 distal and proximal enhancer elements are active in naïve and primed states, both in humans and mice[109,110]. The difference rather emerges from their relative activity levels (high/low) and dominance.

Relying on a single attribute marker or functional test is limiting and must be accompanied by systematic analysis of ever increasing characteristics that continue to be uncovered for different pluripotent states (**Fig. 3,4**). Nevertheless, in our opinion, a molecular and functional characteristic that can be considered as a major divider between naïve vs. primed pluripotent states, is the response of a certain pluripotency circuitry to inhibition of MEK/ERK signalling (**Fig. 5c**). Human conventional ESCs and mouse EpiSCs rapidly collapse upon MEK/ERK inhibition, while naïve pluripotent cells rather tolerate and consolidate their naivety following this challenge[95]. Emphasizing this molecular and functional quality is supported by the ability of MER/ERK inhibition to expand murine epiblast in ICMs and signifies consolidation of naïve pluripotency *in vivo*[111].

Within the domains of the naïve and primed ground states of pluripotency, it is clear that if one considers a list of many naïve and primed pluripotency features originally described for mouse naive 2i/LIF and primed FGF2/Activin A, different pluripotency growth conditions can simultaneously endow a mix of primed and naïve properties in the same cell type (**Fig. 3**). As such, pluripotent states can be classified as "more naïve" or "more primed" by having more of such properties (**Fig. 6**). Human primed ESCs have a number of naïve pluripotency features (e.g. protection of promoter regions from hypermethylation, dependence on Nanog). Murine naïve ESCs expanded in FBS/LIF conditions can give rise to "all-ESC" chimeric embryos and tolerate Dnmt1/Mettl3 ablation, however, they are globally hypermethylated and acquire H3K27me3 over developmental genes as seen in EpiSCs (**Fig. 3,5c**).

Other functional tests that can be used to assess the stringency and extent of naivety in different primate naïve PSCs, is whether the cells can tolerate complete ablation of epigenetic repressors like METTL3, DNMT1, DGCR8 and MBD3[4,65] (**Fig. 3,4**). Further, such tests might be useful for optimizing conditions that close the gap between mouse and human naive pluripotent cells isolated thus far (**Fig. 4, Box 1**). Collectively, it will be informative to systematically annotate different naïve and primed sates from many different species isolated thus far according to such criteria (**Fig. 3**).

Implications and future directions

Yamanaka's breakthrough of reprogramming somatic cells to pluripotency has provided the foundation for deeper sleuthing of pluripotent states and the understanding that pluripotent

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

configurations can be reconfigured. The latter, feeds back and bares direct influence on issues associated with current hurdles and limitations related to human iPSC quality and characteristics (**Box 2**).

Perhaps one of the most fascinating questions related to the naïve to primed pluripotency continuum is "why do these divergent pluripotent configurations actually exist? The latter is often accompanied by the reductionist question of "Which cells are better to work with - naïve or primed?" In our opinion, as this phenomenon is deeply rooted in early embryonic development *in vivo*, it is likely that both configurations constitute essential and integral parts for safeguarding optimality and maximizing the benefits of multi-potency and lineage specification simultaneously. We hypothesize that naive pluripotency emerged as an epigenetic erasure state that renders pluripotent cells free from lineage and epigenetic restriction, while simultaneously making these cells relatively less responsive to signalling pathways that might interfere with establishing such a lineage neutral state. After resetting, the induction of specification by morphogens may not be efficiently forced without a short "delay period". As such, the naïve pluripotency network is gradually resolved and becomes more receptive to inductive cues at the post-implantation stage, and PSCs get differentially patterned/primed according to their spatial localization, before overt somatic differentiation occurs.

At the functional level, it remains to be thoroughly established whether using human naïve PSCs as a starting material, with or without a brief priming, would resolve hurdles currently faced in human *in vitro* PSC differentiation protocols: 1) Will human naïve PSCs yield increased consistency in differentiation outcome among independent iPSC lines[112]? 2) Can naïve PSC conditions yield better quality cells in different differentiation protocols when used as a starting material? 3) Can human naïve PSC facilitate the success of differentiation protocols that have not been conductive with conventional human PSCs? Encouraging support for the latter has recently been provided by showing the enhanced ability of human PSCs expanded in NHSM conditions (even in the absence of aPKCi) for undergoing *in vitro* differentiation into PGCs, a protocol that was inefficient from primed human PSCs[113]. The molecular rationale for evaluating the potential benefits highlighted above is that naïve pluripotency is more associated with cleansing of epigenetic repressive marks over regulatory regions, compared to primed cells[68,97]. This might enable more adequate activation of lineage specifiers during differentiation. Further, lineage biases in human primed PSCs are heavily associated with localized accumulation of repressive marks like DNA methylation[114].

The recent advances in generating human naïve PSCs will continue to boost attempts to generate naïve-like PSCs from other species, and test same-species and inter-species embryo chimerism assays. Cynomolgus monkey naïve ESCs have been derived in NHSM conditions supplemented with Vitamin C, and gave rise to the first chimera competent ESCs, which following ICM-microinjection, yielded chimeric monkey foetuses[115]. Developmentally advanced mouse embryos (E10.5-E17.5) with low chimerism levels were obtained following injection of naive human[68] or monkey iPSCs[116]. These observations raise a variety of exciting challenges relating to defining what are the frequency, lineage preference and developmental quality of such integrated primate iPSC derived cells. Systematic efforts will be key to conclude whether humanized animal models might become relevant for disease modelling, studying human development or generate transplantable human organs.

Continued breakthroughs in single cell technologies and applying them on different pluripotent cell types and embryonic samples will facilitate defining properties that are relevant for adequate functionality of PSCs. This will help set standards for desired optimal starting material for stem cell based therapeutics and research (**Box 1**). It is expected that during this journey aiming at

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

allowing scientists to better control cell fate by deconstructing this previously underappreciated complexity of pluripotency, proposed criteria and standards will likely be debated and revised.

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

Box 1 | A 'dark side' of naïve pluripotency?

With the development of naïve conditions and efforts to endow these cells with more features of naivety, one question that emerges is "how much naivety is needed and is there a dark side for permanently maintaining PSCs under naive conditions?"

Rodent ESCs expanded in 2i/LIF have an increased tendency for acquiring genomic abnormalities[43], and it remains unclear whether they occur as a by-product of non-specific activity of small molecule inhibitors used[96,117], or as a direct result of intrinsic molecular features of naïve pluripotency (e.g. increased ERV activity, reduction in epigenetic repressive marks). One can envision a scenario where such features can be tolerated *in vivo* where this configuration exists for only 1-2 days, while prolonged *in vitro* expansion of this state increases the occurrence frequency of such unwanted damaging events.

The latter concern may also relate to safeguarding the integrity of DNA methylation and imprinting in naïve PSCs expanded *in vitro* over an extended period of time. Studies focusing on loss of DNA methylation upon transfer of mouse PSCs into 2i/LIF conditions have quantified methylation after 10-24 days of transfer, and have documented rapid global loss of DNA methylation accompanied with relative resistance of retrotransposons and imprinting regions to such demethylation[84]. However, it is unclear whether that latter described methylation state represents a final plateau that naive cells achieve, or whether carrying out 2i/LIF cultures further eventually leads to erosion in the relative resistance of such regions to demethylation. Indeed, methylation over imprinted genes and retrotransposons are partially, yet significantly, reduced in 2i/LIF conditions[84].

If such effects are overwhelmingly frequent, researchers will have to re-evaluate how to optimize applying naive conditions. One scenario might involve decreasing inhibitor levels as a mean to avoid excessive hypomethylation or other unwanted effects. An alternative scenario is to maintain cells in primed and transfer them into naive conditions only for short time before initiation of differentiation.

Box 2 | Potential implications on iPSC reprogramming.

Recent studies have implicated how certain epigenetic regulators in fact have opposing effects on naïve and primed murine PSC maintenance[65]. These findings might be relevant when comparing induction of pluripotency mechanisms in human vs. mouse, as human, but not mouse iPSCs, are typically reprogrammed in conventional/primed pluripotency conditions[118]. Consequently, some of the differences observed between human and mouse iPSC regulators might not be related to species differences but rather to the fact that distinct pluripotent states are being induced. Therefore, it would be imperative to expand human iPSC reprogramming regulator screens, and include different pluripotency conditions, as they might yield different outcomes.

Another implication of pluripotent state characteristics on reprogramming is whether naïve conditions might improve the quality of obtained iPSCs. It should be noted that, residual epigenetic memory involving DNA methylation in human iPSCs appears to be rather permanent even after extended cell passaging, while in naïve murine iPSCs, epigenetic memory was more reminiscent of incomplete reprogramming and faded after a few passages *in vitro*[119]. Thus, it should be tested whether the use of adequate naïve conditions might resolve such epigenetic memory in human iPSCs. Similarly, subtle epigenetic differences in DNA methylation found between NT-ESCs and iPSCs generated from the same human donor cells[120] might also be neutralized when deriving iPSCs in naïve conditions that mimic more closely the epigenetic features of ICM.

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

Display items *(all will be subject to graphic editing by NRMCB after review)*

Figure 1 | A**vailable and potential routes for deriving different mouse and human pluripotent cell types.** A variety of pluripotent cell types can be derived from different embryonic cells harvested at various stages of mouse or human development. Alternatively, somatic cells can be reprogrammed via somatic cell nuclear transfer (SCNT) or *in vitro* reprogramming via exogenous transcription factors to generate iPSCs. All pluripotent cell types listed have been derived from both mouse and human donor cells, except for germ cell lineage derived pluripotent cells (EG and GSCs), which have not been stably derived yet in human or other primates. For therapeutic purposes, iPSCs and NT-ESCs have the advantage of being genetically identical to donor original somatic cells. However, NT-ESCs will retain mitochondria of the a-nucleated donor female oocyte. Spermatogonial germ stem cells are generated from spermatogonial stem cells that have already established exclusive male/paternal imprinting pattern, and so do GSCs derived from them. Such maternal imprint-free characteristics likely limits future therapeutic potential of GSCs even if eventually established from adult human males in the future.

Figure 2 | **Signalling pathways and their influence on naïve and primed pluripotent states.** Figure delineates different signalling pathways and their ability to positively or negatively regulate naïve and primed murine pluripotent cells. Please note how that the majority of signalling pathways delineated have opposing effects on murine naïve vs. primed pluripotent state stability (e.g. Lif/Stat3, Fgf2/Erk). It is important to highlight that other pathways not included in this scheme, are likely to be crucially involved in such regulation and will like y be further characterized over the next years. This may include Hippo, Rho, Notch and NFkB signalling. Purple dotted boxes highlight signalling pathways that may be acting differently in mouse vs. human PSC regulation. More specifically, it remains to be fully understood whether low dose of FGF2, TGFB/ACTIVIN/NODAL and/or BMP4 influence human naïve pluripotency differently than previously observed in rodent naïve ESCs. Dotted arrow indicates potential links that remain to be established. (Image is reused and modified from our Nature Reviews Molecular Cell Biology Poster (http://www.nature.com/nrm/posters/pluripotency/index.html)).

Figure 3 | **Naïve and primed pluripotent cell properties in different isolated PSCs.** The scheme delineates in the first column on the left a list of different properties that distinguish between murine ESCs expanded in 2i/LIF (naïve) and murine EpiSCs expanded in Fgf2/Activin A (primed). The latter two states are used as a reference to annotate a variety of other naïve and primed conditions devised for mouse, rat, human or monkey PSCs. For each condition and for each stem cell property, we indicate whether it retains naïve-like (orange circle) or primed-like pattern (blue circle). Empty boxes indicate lack of characterization. This list of features is likely to expand with time and can be used to systematically annotate new pluripotent states isolated in unique conditions and from different species. (Image is reused and modified from our Nature Reviews Molecular Cell Biology Poster (http://www.nature.com/nrm/posters/pluripotency/index.html)).

Figure 4 | **Opposing influence for repressors on murine naïve and primed pluripotent cells.** Murine naïve and primed pluripotent cells do not only differ in their dependence on distinct signalling pathways or in their epigenetic profile, but also in their lineage decision-making. Murine naïve ESCs expanded either in 2i/LIF and FBS/LIF conditions, tolerate complete loss of epigenetic and mRNA repressors such as Dnmt1, Mbd3, Dicer, Dgcr8, Mettl3, Eed, Ezh2. Further, the latter modifications strengthens the equilibrium in favour of pluripotency promoting factors and generates "hyper-naïve" pluripotent cells that are relatively more resistant to differentiation and can tolerate withdrawal of LIF cytokine. Primed EpiSCs actually respond in

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

an opposite manner to the complete ablation of such repressors. Established murine primed EpiSCs naturally down regulate pluripotency factors and upregulate lineage priming factors. Ablation of repressors at this stage tips the balance toward differentiation and/or compromises cell survival. It should be noted that ablation of two different repressors (e.g. Mettl3 vs. Dgcr8) overall negatively influences primed pluripotency stability, this does not necessarily mean that the downstream events leading to the state collapse are identical, and thus should be thoroughly dissected *in vitro* and *in vivo*. (Figure 4 is modified from Geula et al. Science 2015 (Supplementary Figure S34)).

Figure 5 | **Key differences between mouse and human pre and post-implantation development *in vivo*. (a)** Scheme delineates similarities and differences in mouse and human early pre- and post-implantation development. While mouse and human embryos are not morphologically different up to the blastocyst stage, there are striking transcriptional differences as summarized in (**b**). Further, at the postimplantation stage, morphological differences in human vs. mouse embryo shape become striking including (i) differences in epiblast shape and (ii) extra-embryonic structures, as delineated in (a). In mouse, naïve ESCs (orange arrows) can be derived from ICM or post-implantation epiblast when naïve growth conditions are applied. Primed EpiSCs can be derived from post-implantation epiblast or ICM when primed culture conditions are used. Thus, growth condition rather than source dictates pluripotent state configuration acquired *in vitro*. A similar scenario applies for derivation of naïve and primed cells from human ICM depending on the growth conditions used. PSC derivations or molecular analysis on human post-implantation embryos cannot be conducted due to ethical issues. PE-Primitive Endoderm; TE – Trophectoderm; ICM- Inner Cell Mass. (**c**) A model to explain the relativity of naivety within the naive to primed pluripotency spectrum. One major molecular and functional criterion that can be considered for separating naïve and primed pluripotent cells is their ability to maintain and stabilize their pluripotent state in the absence of MEK-ERK1/2 activity (dashed black line). Within the naïve or primed pluripotent states, it is difficult to describe the pluripotent state of the cells in absolute terms, as naïve cells can have, to some extent, primed pluripotency features. Similarly, within the primed pluripotency spectrum, primed PSCs expanded in different conditions have different features and varying degrees of naivety (**Figure 3**). Finally, it is possible that supplementation of 2i/LIF conditions with small molecules such as aPKCi, FGFRi or NOTCHi will further consolidate naïve pluripotency features, particularly from other rodents like rats whose stability in 2i/LIF feeder free conditions should be further improved. Full annotation of different human pluripotent states will allow charting an equivalent landscape for human and monkey PSCs. (Images in **a-b** are reused and modified from our Nature Reviews Molecular Cell Biology Poster (http://www.nature.com/nrm/posters/pluripotency/index.html). Image in **c** is reused and modified from our Figure 3 in Manor et al. Curr. Opin. in Genetics & Development).

Copyright permissions

- Figure 2,3,5a-b: Graphic figures are reused from our Nature Reviews Molecular Cell Biology Poster (http://www.nature.com/nrm/posters/pluripotency/index.html) as indicated in figure legends**.**

- Figure 4 is modified from Geula et al. Science 2015 (Supplementary Figure S34).

- Figure 5c is reused and modified from our Figure 3 in Manor et al. Curr. Opin. In Genetics & development

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

Acknowledgements

J.H.H is supported by a generous gift from Ilana and Pascal Mantoux; the New York Stem Cell Foundation (NYSCF), Flight Attendant Medical Research Institute (FAMRI), the Kimmel Innovator Research Award, the ERC-StG (StG-2011-281906), Moross Cancer Institute, Israel Science Foundation – NFSC program, Morasha Biomed program, ICORE program, the ICRF Foundation, MINERVA fund, Helen and Martin Kimmel Institute for Stem Cell research, the Benoziyo Endowment fund, David and Fela Shapell Family Foundation INCPM Fund for Preclinical Studies, an HFSPO research grant. J.H.H. is a New York Stem Cell Foundation - Robertson Investigator. We thank W. Greenleaf and members of the Hanna lab for discussions. We apologize to those whose work could not be covered or directly cited due to space limitations.

Further information

Online Web link sites:

Addgene plasmid repository: https://www.addgene.org/

ENCODE project: http://www.nature.com/encode/

Epigenome Roadmap project: http://www.nature.com/collections/vbqgtr

Mouse ES cell ChIP compendium: http://bioinformatics.cscr.cam.ac.uk/ES_Cell_ChIP-seq_compendium.html

CRISPR/CAS9 genome wide screen resource: http://genome-engineering.org/gecko/

Online poster by Nature Reviews Molecular Cell Biology on Pluripotent States: http://www.nature.com/nrm/posters/pluripotency/index.html

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

Glossary

Primordial germ cells (PGCs) – embryonic progenitor cells that give rise to germ cells in the gonads (sperm and oocytes).

Inner cell mass (ICM) - the mass of cells inside the pre-implantation blastocyst that will subsequently give rise to the definitive structures of the fetus.

Embryonic stem cells (ESCs) – *in vitro* expanded pluripotent cells that originate from the ICM.

Epiblast stem cells (EpiSCs)- *in vitro* expanded pluripotent cells that originate from the rodent post-implantation epiblast.

Embryonic germ cells – *in vitro* expanded pluripotent cells that are derived from embryonic PGCs.

Germ stem cells (GSCs) – *in vitro* expanded pluripotent stem cells that originate from neonatal or adult testis derived spermatogonial stem cells.

Nuclear transfer – cloning of somatic cell derived nucleus and its introduction into a-nucleated host oocyte.

Induced pluripotent stem cells (iPSCs) – *in vitro* generated pluripotent cells derived via ectopic expression of defined exogenous factors in somatic cells.

Naïve pluripotency – pluripotent state that resembles pre-implantation pluripotent configuration(s).

Primed pluripotency – pluripotent state that corresponds to post-implantation embryonic configuration(s).

Ground state pluripotency – originally described as a state of pluripotency that is independent of exogenous activator signalling input or stimulation.

X inactivation – dosage compensation of X chromosome in female, where one of the X chromosomes gets epigenetically silenced.

Seed enhancers - subgroup of enhancers that are dormant in naive cells but become more active in primed pluripotent and somatic cells.

3i – Defined naïve pluripotency growth conditions combing 3 inhibitors (i) for MEK, FGF and GSK3 signalling.

2i/LIF – Defined naïve pluripotency growth conditions containing 2 inhibitors (i) for MEK and GSK3 together with LIF cytokine.

“Alternative 2i” – Defined naïve pluripotency growth conditions composed of 2 small molecule inhibitors for GSK3 and SRC pathways

LIF/MEKi/aPKCi - Defined naïve pluripotency growth conditions containing 2 inhibitors (i) for MEK and atypical PKC signalling, together with LIF cytokine.

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

FGF2/ACTIVIN A – Defined primed pluripotency growth conditions for mouse EpiSCs composed of recombinant FGF2 and ACTIVIN A cytokines.

FGF2/IWR1 - Defined primed pluripotency growth conditions for mouse EpiSCs composed of recombinant FGF2 and Tankyrase small molecule inhibitor, IWR1.

GSK3i/IWR1 - Defined primed pluripotency growth conditions for mouse EpiSCs composed of GSK3 pathway inhibitor and Tankyrase small molecule inhibitor, IWR1.

Table of contents summary

Hanna and colleagues review recent advances on molecular underpinnings of alternative primed- and naïve-like pluripotent states isolated in rodents and in man. They highlight potential benefits and identify key unanswered challenges in this rapidly evolving fundamental topic.

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

Author biographies

Leehee Weinberger M.Sc., has been a PhD student with Jacob H. Hanna at the Department of Molecular Genetics at the Weizmann Institute of Science, Rehovot, Israel for 4 years. She carried out her Masters work with Naama Barkai at the the same department. Her doctoral research focused on regulation and differentiation of human naïve pluripotent cells.

Muneef Ayyash Ph.D., has been a staff researcher in Jacob H. Hanna lab at the Department of Molecular Genetics at the Weizmann Institute of Science Rehovot, Israel for 1 year. He carried docotral work with David Wallach at the Weizmann Institute of Science, Rehovot, Israel and did postdoctoral studies with Yinon Ben-Neriah at the Hebrew University, Jeruslem, Israel. His research currently focuses on comparative analysis of distinct human naïve pluriptoent states.

Noa Novershtern Ph.D., has been a staff scientist and senior bioinformatician in Jacob H. Hanna lab at the Department of Molecular Genetics at the Weizmann Institute of Science, Rehovot, Israel for 5 years. She carried out her doctoral work with Nir Friedman at the Hebrew University, Jerusalem, Israel and Aviv Regev at the Broad Institute, Cambridge, USA. She focuses on dissecting epigenetic dynamics in pluriptoency and reprogramming.

Jacob H. Hanna M.D. Ph.D., has been on the Faculty of the Department of Molecular Genetics at the Weizmann Institute of Science, Rehovot, Israel for 5 years, and is also a Robertson Investigator by the New York Stem Cell Foundation (NYSCF). He carried out his doctoral work with Ofer Mandelobim at the Hebrew University, Jerusalem, Israel and then did his postdoctoral studies with Rudolf Jaenisch at the Whitehead Institute for Biomedical Research - Massachusetts Institute of Technology, Cambridge, USA. His laboratory studies molceular mechanisms of stem cell reprogramming and differentiation. Hanna lab website: http://hannalabweb.weizmann.ac.il/

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

References

1 Hanna, J. H., Saha, K. & Jaenisch, R. Pluripotency and cellular reprogramming: facts, hypotheses, unresolved issues. Cell 143, 508-525 (2010).

2 Nichols, J. & Smith, A. Pluripotency in the embryo and in culture. Cold Spring Harb Perspect Biol 4, (2012).

3 Hackett, J. A. & Surani, M. A. Regulatory principles of pluripotency: from the ground state up. Cell Stem Cell 15, 416-430 (2014).

4 Manor, Y. S., Massarwa, R. & Hanna, J. H. Establishing the human naive pluripotent state. Curr Opin Genet Dev 34, 35-45 (2015).

5 Inoue, H., Nagata, N., Kurokawa, H. & Yamanaka, S. iPS cells: a game changer for future medicine. EMBO J 33, 409-417 (2014).

**6 Thomson, J. A. et al. Embryonic stem cell lines derived from human blastocysts. Science 282, 1145-1147 (1998).
First study to derive primed ESCs from human blastocysts.

7 Evans, M. J. & Kaufman, M. H. Establishment in culture of pluripotential cells from mouse embryos. Nature 292, 154-156 (1981).

**8 Tesar, P. J. et al. New cell lines from mouse epiblast share defining features with human embryonic stem cells. Nature 448, 196-199 (2007).
One of two studies that derived primed EpiSC lines from rodent post-implantation epiblasts.

**9 Brons, I. G. et al. Derivation of pluripotent epiblast stem cells from mammalian embryos. Nature 448, 191-195 (2007).
One of two studies that derived primed EpiSC lines from rodent post-implantation epiblasts.

10 Leitch, H. G. et al. Embryonic germ cells from mice and rats exhibit properties consistent with a generic pluripotent ground state. Development 137, 2279-2287 (2010).

*11 Kanatsu-Shinohara, M. et al. Generation of pluripotent stem cells from neonatal mouse testis. Cell 119, 1001-1012 (2004).
First study to desribe generation of pluripotent ES-like cells from mouse spermatogonial stem cells.

12 Tanaka, T., Kanatsu-Shinohara, M., Hirose, M., Ogura, A. & Shinohara, T. Pluripotent cell derivation from male germline cells by suppression of Dmrt1 and Trp53. J Reprod Dev (2015).

13 Ko, K. et al. Induction of pluripotency in adult unipotent germline stem cells. Cell Stem Cell 5, 87-96 (2009).

14 Seandel, M. et al. Generation of functional multipotent adult stem cells from GPR125+ germline progenitors. Nature 449, 346-350 (2007).

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

15 Ko, K. et al. Human adult germline stem cells in question. Nature 465, E1; discussion E3 (2010).

16 Shamblott, M. J. et al. Derivation of pluripotent stem cells from cultured human primordial germ cells. Proc Natl Acad Sci U S A 95, 13726-13731 (1998).

**17 Yamada, M. et al. Human oocytes reprogram adult somatic nuclei of a type 1 diabetic to diploid pluripotent stem cells. Nature 510, 533-536 (2014).
First study to generate validated human NT-ESCs from adult patient specific fibroblasts.

**18 Tachibana, M. et al. Human embryonic stem cells derived by somatic cell nuclear transfer. Cell 153, 1228-1238 (2013).
First study to generate validated human NT-ESCs from embryonic fibroblasts.

*19 Wakayama, T. et al. Differentiation of embryonic stem cell lines generated from adult somatic cells by nuclear transfer. Science 292, 740-743 (2001).
Study demonstrating feasibility of SCNT in mice.

**20 Takahashi, K. & Yamanaka, S. Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. Cell 126, 663-676 (2006).
First study that invented direct in vitro reprogramming of somati cells into iPSCs with defined transcription factors.

21 Takahashi, K. et al. Induction of pluripotent stem cells from adult human fibroblasts by defined factors. Cell 131, 861-872 (2007).

22 Wernig, M. et al. In vitro reprogramming of fibroblasts into a pluripotent ES-cell-like state. Nature 448, 318-324 (2007).

23 Yu, J. et al. Human induced pluripotent stem cells free of vector and transgene sequences. Science 324, 797-801 (2009).

24 Yu, J. et al. Induced pluripotent stem cell lines derived from human somatic cells. Science 318, 1917-1920 (2007).

25 Deuse, T. et al. SCNT-derived ESCs with mismatched mitochondria trigger an immune response in allogeneic hosts. Cell Stem Cell 16, 33-38 (2015).

26 Ma, H. et al. Metabolic rescue in pluripotent cells from patients with mtDNA disease. Nature 524, 234-238 (2015).

27 Tachibana, M. et al. Mitochondrial gene replacement in primate offspring and embryonic stem cells. Nature 461, 367-372 (2009).

28 Paull, D. et al. Nuclear genome transfer in human oocytes eliminates mitochondrial DNA variants. Nature 493, 632-637 (2013).

**29 Hanna, J. et al. Metastable pluripotent states in NOD-mouse-derived ESCs. Cell Stem Cell 4, 513-524 (2009).

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

One of two studies first showing interconversion ebtween murine naïve and primed pluriptoent cells, and in different mouse strains.

30 Nichols, J. & Smith, A. Naive and primed pluripotent states. Cell Stem Cell 4, 487-492 (2009).

31 Han, D. W. et al. Direct reprogramming of fibroblasts into epiblast stem cells. Nat Cell Biol 13, 66-71 (2011).

32 Factor, D. C., Najm, F. J. & Tesar, P. J. Generation and characterization of epiblast stem cells from blastocyst-stage mouse embryos. Methods Mol Biol 1074, 1-13 (2013).

33 Davidson, K. C., Mason, E. A. & Pera, M. F. The pluripotent state in mouse and human. Development 142, 3090-3099 (2015).

34 Martin, G. R. Isolation of a pluripotent cell line from early mouse embryos cultured in medium conditioned by teratocarcinoma stem cells. Proc Natl Acad Sci U S A 78, 7634-7638 (1981).

35 Smith, A. G. et al. Inhibition of pluripotential embryonic stem cell differentiation by purified polypeptides. Nature 336, 688-690 (1988).

36 Williams, R. L. et al. Myeloid leukaemia inhibitory factor maintains the developmental potential of embryonic stem cells. Nature 336, 684-687 (1988).

37 Orkin, S. H. & Hochedlinger, K. Chromatin connections to pluripotency and cellular reprogramming. Cell 145, 835-850 (2011).

38 Jaenisch, R. & Young, R. Stem cells, the molecular circuitry of pluripotency and nuclear reprogramming. Cell 132, 567-582 (2008).

39 Tanaka, M., Hadjantonakis, A. K., Vintersten, K. & Nagy, A. Aggregation chimeras: combining ES cells, diploid, and tetraploid embryos. Methods Mol Biol 530, 287-309 (2009).

**40 Ying, Q. L., Nichols, J., Chambers, I. & Smith, A. BMP induction of Id proteins suppresses differentiation and sustains embryonic stem cell self-renewal in collaboration with STAT3. Cell 115, 281-292 (2003).
Study describing defined 3i naïve conditions capable of generating Lif/Stat3 independent mouse ESCs.

41 Burdon, T., Stracey, C., Chambers, I., Nichols, J. & Smith, A. Suppression of SHP-2 and ERK signalling promotes self-renewal of mouse embryonic stem cells. Dev Biol 210, 30-43 (1999).

42 Ying, Q. L. et al. The ground state of embryonic stem cell self-renewal. Nature 453, 519-523 (2008).

43 Buehr, M. et al. Capture of authentic embryonic stem cells from rat blastocysts. Cell 135, 1287-1298 (2008).

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

44 Shimizu, T. et al. Dual inhibition of Src and GSK3 maintains mouse embryonic stem cells, whose differentiation is mechanically regulated by Src signaling. Stem Cells 30, 1394-1404 (2012).

*45 Dutta, D. et al. Self-renewal versus lineage commitment of embryonic stem cells: protein kinase C signaling shifts the balance. Stem Cells 29, 618-628 (2011).
First study utulizing aPKCi to boost naïve murine iPSCs and ESCs generation.

46 Kolodziejczyk, A. A. et al. Single Cell RNA-Sequencing of Pluripotent States Unlocks Modular Transcriptional Variation. Cell Stem Cell 17, 471-485 (2015).

47 Chen, Y., Blair, K. & Smith, A. Robust self-renewal of rat embryonic stem cells requires fine-tuning of glycogen synthase kinase-3 inhibition. Stem Cell Reports 1, 209-217 (2013).

48 Meek, S. et al. Tuning of beta-catenin activity is required to stabilize self-renewal of rat embryonic stem cells. Stem Cells 31, 2104-2115 (2013).

49 Rajendran, G. et al. Inhibition of protein kinase C signaling maintains rat embryonic stem cell pluripotency. J Biol Chem 288, 24351-24362 (2013).

*50 Li, X. et al. Calcineurin-NFAT signaling critically regulates early lineage specification in mouse embryonic stem cells and embryos. Cell Stem Cell 8, 46-58 (2011).
First study to show that Src inhibition promotes murine naïve pluriptoency.

**51 Nishioka, N. et al. The Hippo signaling pathway components Lats and Yap pattern Tead4 activity to distinguish mouse trophectoderm from inner cell mass. Dev Cell 16, 398-410 (2009).
Study identifies Hippo signaling pathway role in epiblast vs. trophoblast specification in pre-implantaioon mouse embryos.

52 Azzolin, L. et al. YAP/TAZ incorporation in the beta-catenin destruction complex orchestrates the Wnt response. Cell 158, 157-170 (2014).

53 Lian, I. et al. The role of YAP transcription coactivator in regulating stem cell self-renewal and differentiation. Genes Dev 24, 1106-1118 (2010).

54 Wray, J. et al. Inhibition of glycogen synthase kinase-3 alleviates Tcf3 repression of the pluripotency network and increases embryonic stem cell resistance to differentiation. Nat Cell Biol 13, 838-845 (2011).

55 Faunes, F. et al. A membrane-associated beta-catenin/Oct4 complex correlates with ground-state pluripotency in mouse embryonic stem cells. Development 140, 1171-1183 (2013).

56 Morgani, S. M. & Brickman, J. M. LIF supports primitive endoderm expansion during pre-implantation development. Development (2015).

57 Silva, J. et al. Nanog is the gateway to the pluripotent ground state. Cell 138, 722-737 (2009).

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

58 Maza, I. et al. Transient acquisition of pluripotency during somatic cell transdifferentiation with iPSC reprogramming factors. Nat Biotechnol 33, 769-774 (2015).

59 Carter, A. C., Davis-Dusenbery, B. N., Koszka, K., Ichida, J. K. & Eggan, K. Nanog-independent reprogramming to iPSCs with canonical factors. Stem Cell Reports 2, 119-126 (2014).

60 Festuccia, N. et al. Esrrb is a direct Nanog target gene that can substitute for Nanog function in pluripotent cells. Cell Stem Cell 11, 477-490 (2012).

61 Yeo, J. C. et al. Klf2 is an essential factor that sustains ground state pluripotency. Cell Stem Cell 14, 864-872 (2014).

*62 Reynolds, N. et al. NuRD suppresses pluripotency gene expression to promote transcriptional heterogeneity and lineage commitment. Cell Stem Cell 10, 583-594 (2012).
Study establishess Mbd3/NuRD as a repressor of naïve pluriptoency promoting genes in mouse ESCs.

63 Rais, Y. et al. Deterministic direct reprogramming of somatic cells to pluripotency. Nature 502, 65-70 (2013).

64 Loh, K. M. & Lim, B. A precarious balance: pluripotency factors as lineage specifiers. Cell Stem Cell 8, 363-369 (2011).

*65 Geula, S. et al. Stem cells. m6A mRNA methylation facilitates resolution of naive pluripotency toward differentiation. Science 347, 1002-1006 (2015).
First study proving opposing dependence on epigentic repressors between mouse naïve and primed PSCs.

66 Buecker, C. et al. Reorganization of enhancer patterns in transition from naive to primed pluripotency. Cell Stem Cell 14, 838-853 (2014).

67 Marks, H. et al. The transcriptional and epigenomic foundations of ground state pluripotency. Cell 149, 590-604 (2012).

**68 Gafni, O. et al. Derivation of novel human ground state naive pluripotent stem cells. Nature 504, 282-286 (2013).
First study to generate geneticaly unmodified and indefinitely stable human MEK independent naive PSCs, capable of generating advanced cross species mosue-human chimeric embryos.

69 Factor, D. C. et al. Epigenomic comparison reveals activation of "seed" enhancers during transition from naive to primed pluripotency. Cell Stem Cell 14, 854-863 (2014).

70 Wang, Y., Medvid, R., Melton, C., Jaenisch, R. & Blelloch, R. DGCR8 is essential for microRNA biogenesis and silencing of embryonic stem cell self-renewal. Nat Genet 39, 380-385 (2007).

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

71 Kumar, R. M. et al. Deconstructing transcriptional heterogeneity in pluripotent stem cells. Nature 516, 56-61 (2014).

72 Kim, H. et al. Modulation of beta-catenin function maintains mouse epiblast stem cell and human embryonic stem cell self-renewal. Nat Commun 4, 2403 (2013).

*73 Kojima, Y. et al. The transcriptional and functional properties of mouse epiblast stem cells resemble the anterior primitive streak. Cell Stem Cell 14, 107-120 (2014).
One of first two studies descrining region-specific features of in vitro expanded mouse EpiSCs.

*74 Wu, J. et al. An alternative pluripotent state confers interspecies chimaeric competency. Nature 521, 316-321 (2015).
One of first two studies descrining region-specific features of in vitro expanded mouse EpiSCs.

75 Han, D. W. et al. Epiblast stem cell subpopulations represent mouse embryos of distinct pregastrulation stages. Cell 143, 617-627 (2010).

*76 Hayashi, K., Ohta, H., Kurimoto, K., Aramaki, S. & Saitou, M. Reconstitution of the mouse germ cell specification pathway in culture by pluripotent stem cells. Cell 146, 519-532 (2011).
First study to establish mouse PGCLC induction from murine ERK independent naïve PSCs.

77 Magnusdottir, E. & Surani, M. A. How to make a primordial germ cell. Development 141, 245-252 (2014).

**78 Guo, G. et al. Klf4 reverts developmentally programmed restriction of ground state pluripotency. Development 136, 1063-1069 (2009).
One of two studies first showing interconversion between murine naïve and primed plurippotent cells.

79 Bao, S. et al. Epigenetic reversion of post-implantation epiblast to pluripotent embryonic stem cells. Nature 461, 1292-1295 (2009).

80 Gillich, A. et al. Epiblast stem cell-based system reveals reprogramming synergy of germline factors. Cell Stem Cell 10, 425-439 (2012).

81 Greber, B. et al. Conserved and divergent roles of FGF signaling in mouse epiblast stem cells and human embryonic stem cells. Cell Stem Cell 6, 215-226 (2010).

82 Najm, F. J. et al. Isolation of epiblast stem cells from preimplantation mouse embryos. Cell Stem Cell 8, 318-325 (2011).

83 Leitch, H. G. et al. Naive pluripotency is associated with global DNA hypomethylation. Nat Struct Mol Biol 20, 311-316 (2013).

84 Ficz, G. et al. FGF signaling inhibition in ESCs drives rapid genome-wide demethylation to the epigenetic ground state of pluripotency. Cell Stem Cell 13, 351-359 (2013).

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

85 Hackett, J. A. et al. Synergistic mechanisms of DNA demethylation during transition to ground-state pluripotency. Stem Cell Reports 1, 518-531 (2013).

86 Galonska, C., Ziller, M. J., Karnik, R. & Meissner, A. Ground State Conditions Induce Rapid Reorganization of Core Pluripotency Factor Binding before Global Epigenetic Reprogramming. Cell Stem Cell (2015).

87 Liao, J. et al. Targeted disruption of DNMT1, DNMT3A and DNMT3B in human embryonic stem cells. Nat Genet 47, 469-478 (2015).

88 Tee, W. W., Shen, S. S., Oksuz, O., Narendra, V. & Reinberg, D. Erk1/2 activity promotes chromatin features and RNAPII phosphorylation at developmental promoters in mouse ESCs. Cell 156, 678-690 (2014).

89 Bertero, A. et al. Activin/nodal signaling and NANOG orchestrate human embryonic stem cell fate decisions by controlling the H3K4me3 chromatin mark. Genes Dev 29, 702-717 (2015).

90 Smith, Z. D. et al. DNA methylation dynamics of the human preimplantation embryo. Nature 511, 611-615 (2014).

91 Mekhoubad, S. et al. Erosion of dosage compensation impacts human iPSC disease modeling. Cell Stem Cell 10, 595-609 (2012).

92 Chia, N. Y. et al. A genome-wide RNAi screen reveals determinants of human embryonic stem cell identity. Nature 468, 316-320 (2010).

93 Shipony, Z. et al. Dynamic and static maintenance of epigenetic memory in pluripotent and somatic cells. Nature 513, 115-119 (2014).

94 Betschinger, J. et al. Exit from pluripotency is gated by intracellular redistribution of the bHLH transcription factor Tfe3. Cell 153, 335-347 (2013).

*95 Hanna, J. et al. Human embryonic stem cells with biological and epigenetic characteristics similar to those of mouse ESCs. Proc Natl Acad Sci U S A 107, 9222-9227 (2010).
Study provided first evidence for alternative transgene dependent human PSCs that can be expanded in 2i/LIF conditions.

96 Theunissen, T. W. et al. Systematic identification of culture conditions for induction and maintenance of naive human pluripotency. Cell Stem Cell 15, 471-487 (2014).

97 Takashima, Y. et al. Resetting transcription factor control circuitry toward ground-state pluripotency in human. Cell 158, 1254-1269 (2014).

98 Blakeley, P. et al. Defining the three cell lineages of the human blastocyst by single-cell RNA-seq. Development 142, 3151-3165 (2015).

99 Gkountela, S. et al. DNA Demethylation Dynamics in the Human Prenatal Germline. Cell 161, 1425-1436 (2015).

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

100 Shakiba, N. et al. CD24 tracks divergent pluripotent states in mouse and human cells. Nat Commun 6, 7329 (2015).

101 Barakat, T. S. et al. Stable X chromosome reactivation in female human induced pluripotent stem cells. Stem Cell Reports 4, 199-208 (2015).

102 Chan, Y. S. et al. Induction of a human pluripotent state with distinct regulatory circuitry that resembles preimplantation epiblast. Cell Stem Cell 13, 663-675 (2013).

103 Duggal, G. et al. Alternative Routes to Induce Naive Pluripotency in Human Embryonic Stem Cells. Stem Cells 33, 2686-2698 (2015).

104 Chen, H. et al. Reinforcement of STAT3 activity reprogrammes human embryonic stem cells to naive-like pluripotency. Nat Commun 6, 7095 (2015).

*105 Ohgushi, M., Minaguchi, M. & Sasai, Y. Rho-Signaling-Directed YAP/TAZ Activity Underlies the Long-Term Survival and Expansion of Human Embryonic Stem Cells. Cell Stem Cell (2015).
Study connecting RHO and HIPPO signalling pathways in maintenance of primed human pluripotency.

106 Kameda, T. & Thomson, J. A. Human ERas gene has an upstream premature polyadenylation signal that results in a truncated, noncoding transcript. Stem Cells 23, 1535-1540 (2005).

107 Surani, M. A. Human Germline: A New Research Frontier. Stem Cell Reports 4, 955-960 (2015).

108 De Los Angeles, A. et al. Hallmarks of pluripotency. Nature 525, 469-478 (2015).

109 Buecker, C. & Wysocka, J. Enhancers as information integration hubs in development: lessons from genomics. Trends Genet 28, 276-284 (2012).

110 Karwacki-Neisius, V. et al. Reduced Oct4 expression directs a robust pluripotent state with distinct signaling activity and increased enhancer occupancy by Oct4 and Nanog. Cell Stem Cell 12, 531-545 (2013).

111 Boroviak, T., Loos, R., Bertone, P., Smith, A. & Nichols, J. The ability of inner-cell-mass cells to self-renew as embryonic stem cells is acquired following epiblast specification. Nat Cell Biol 16, 516-528 (2014).

112 Osafune, K. et al. Marked differences in differentiation propensity among human embryonic stem cell lines. Nat Biotechnol 26, 313-315 (2008).

*113 Irie, N. et al. SOX17 is a critical specifier of human primordial germ cell fate. Cell 160, 253-268 (2015).
First study to generate human in vitro PGCLCs and demonstrate altered function of human ERK independent naïve PSCs.

114 Bock, C. et al. Reference Maps of human ES and iPS cell variation enable high-throughput characterization of pluripotent cell lines. Cell 144, 439-452 (2011).

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

**115 Chen, Y. et al. Generation of Cynomolgus Monkey Chimeric Fetuses using Embryonic Stem Cells. Cell Stem Cell 17, 116-124 (2015).
Study demonstrating frist ever geenrated chimeric monkey fetuses with naïve monkey ESCs eastablished in NHSM conditions supplemented with Vitamin C.

116 Fang, R. et al. Generation of naive induced pluripotent stem cells from rhesus monkey fibroblasts. Cell Stem Cell 15, 488-496 (2014).

117 Bock, A. S., Leigh, N. D. & Bryda, E. C. Effect of Gsk3 inhibitor CHIR99021 on aneuploidy levels in rat embryonic stem cells. In Vitro Cell Dev Biol Anim 50, 572-579 (2014).

118 Qin, H. et al. Systematic identification of barriers to human iPSC generation. Cell 158, 449-461 (2014).

119 Kim, K. et al. Donor cell type can influence the epigenome and differentiation potential of human induced pluripotent stem cells. Nat Biotechnol 29, 1117-1119 (2011).

120 Ma, H. et al. Abnormalities in human pluripotent cells due to reprogramming mechanisms. Nature 511, 177-183 (2014).

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

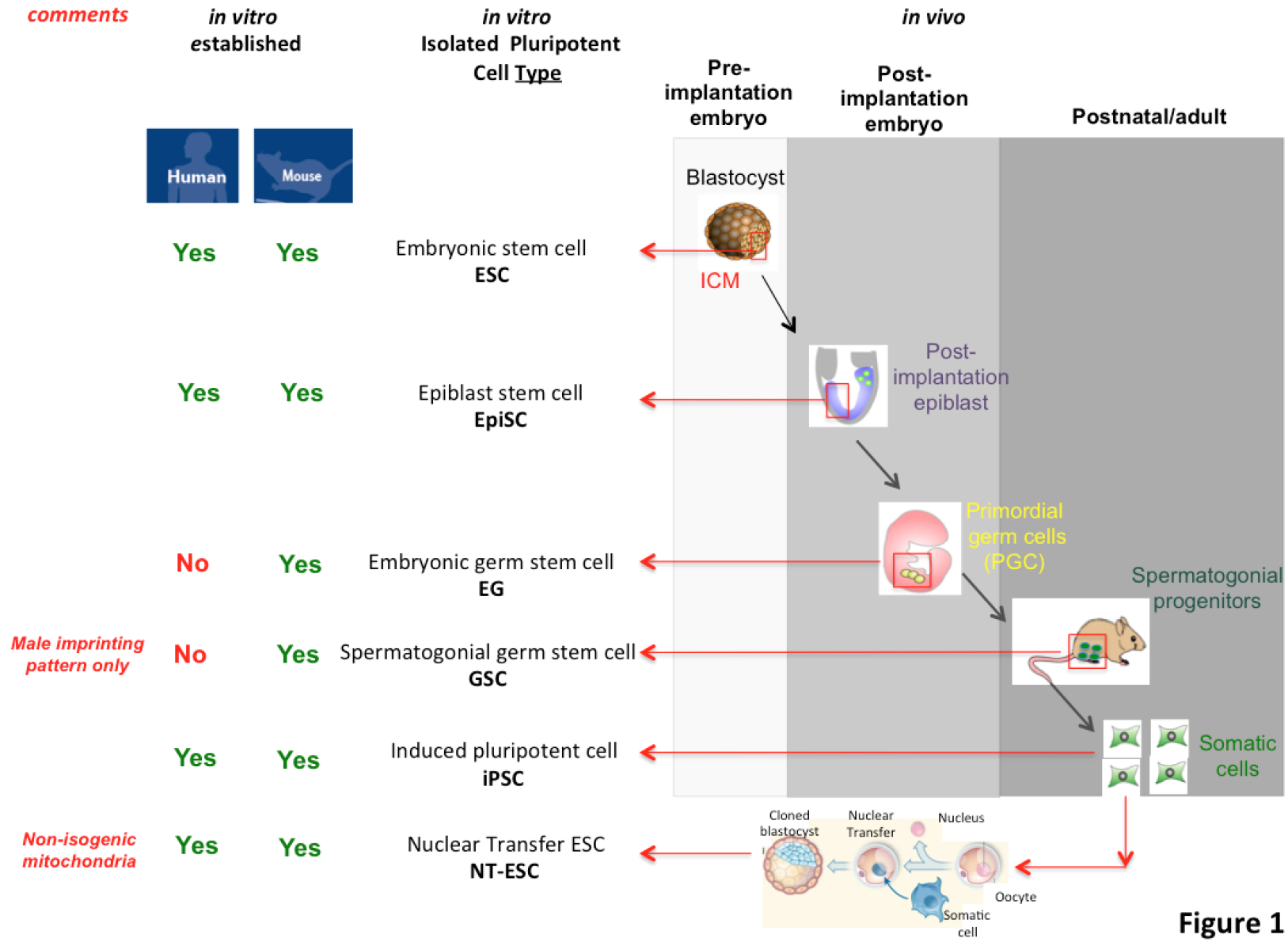


Figure 1

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

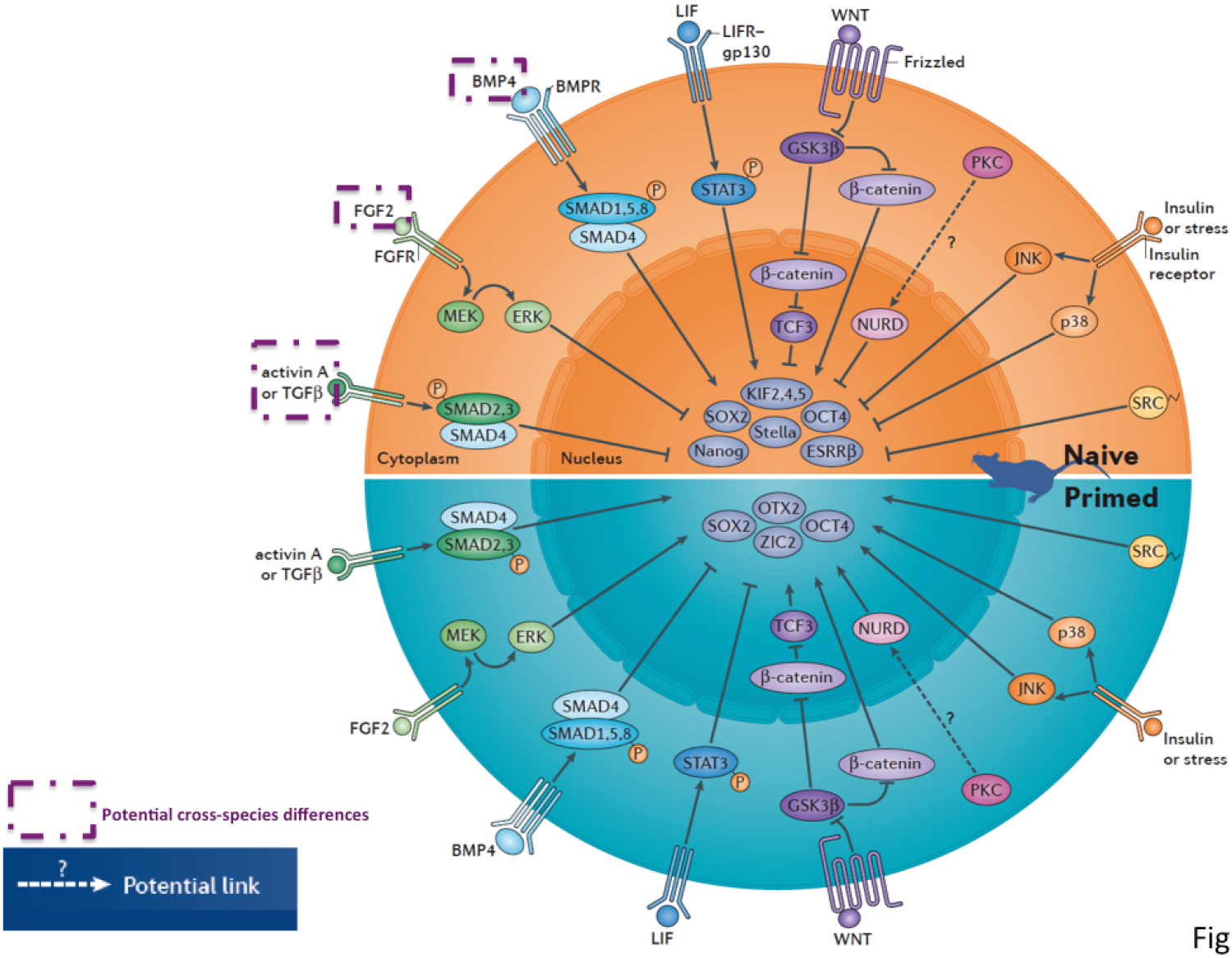


Figure 2

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

			Mouse						Rat				Human						Rhesus	
	Naive	Primed	2i, LIF	FBS, LIF	SRCi, GSK3βi	MEKi, LIF	BMP4, LIF	GSK3βi, AXINs	FGF2, activin A or FGF2, TGFβ	2i, LIF, MEF	LIF, MEKi, PKCi	FGF2, activin A or FGF2, TGFβ	OCT4, KLF2, KLF4, 2i, LIF	2i, LIF, p38i, JNKi, PKCi, ROCKi, FGF2, TGFβ	2i, LIF, PKCi, MEF (KLF2, NANOG)	2i, BRAFi, ROCKi, SRCi, LIF, activin A, MEF (FGF2, JNKi)	FGF2 TGFβ 2i, BMPi, LIF, MEF	FGF2, TGFβ or FGF2, activin A or FGF2, MEF	2i, LIF, JNKi, p38i, FGF2, MEF	FGF2, TGFβ or FGF2, activin A or FGF2, MEF
MEK–ERK dependence	No	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes
Long-term dependence on FGF2 signalling	No	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes		Yes			Yes	Yes	Yes	Yes
Long-term dependence on TGFβ–activin A signalling	No	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes		Yes		Yes	Yes	Yes		Yes
Dominant OCT4 enhancer	Distal	Proximal	Yes	Yes		Yes		Yes	Yes				Yes	Yes	Yes	Yes		Yes		
H3K27me3 on developmental regulators	Low	High	Yes	Yes					Yes					Yes		Yes	Yes	Yes		
Global DNA hypomethylation	Yes	No	Yes	Yes					Yes					Yes (Mild)	Yes (Strong)			Yes		Yes
X chromosome inactivation	No	Yes	Yes	Yes		Yes		Yes	Yes				Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes
Dependence on DNMT1, DICER, METTL3, MBD3	No	Yes	Yes	Yes					Yes									Yes		
Priming markers (OTX2, ZIC2)	↓	↑	Yes	Yes		Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes		Yes
Pluripotency markers (NANOG, KLFs, ESRRβ)	↑	↓	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes*	Yes (Mild*)	Yes (Strong*)	Yes (Strong*)	Yes (Mild*)	Yes*	Yes (Mild*)	Yes*
TFE3 nuclear localization	High	Low	Yes	Yes		Yes			Yes					Yes	Yes		Yes	Yes		
CD24/MHC class 1	Low/low	High/mod	Yes	Yes					Yes					Yes	Yes	Yes		Yes		
HERV-H and HERV-K expression	High	Low	Primate specific											Yes	Yes	Yes	Yes	Yes		
Expressed adhesion molecules	E-cadherin	N-cadherin	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes		Yes	Yes	Yes	Yes	Yes	Yes	Yes
Promotion of pluripotency maintenance via Nanog or Prdm14	Yes	No	Yes	Yes	Yes (Nanog only)	Yes	Yes	Yes (Nanog only)	Yes					Yes	Yes	Yes	Yes	Yes		
Metabolism	OxPhos, glycolytic	Glycolytic	Yes	Yes					Yes							Yes		Yes		
Competence as initial starting cells for PGCLC induction	High	Low	Yes	Yes					Yes					Yes				Yes		
Capacity of colonization of host pre-implantation ICM and contribution to chimaeras	High	Low	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes		Yes‡	Yes‡	Yes‡		Yes	Yes‡	Yes
Hypo-methylation of promoter and enhancer regions	Yes	No	Yes	Yes					Yes					Yes	Yes		Yes			
c-Kit	Yes	No	Yes	Yes					Yes								Yes			
Tolerance for absence of exogenous L-Glutamine	Yes	No	Yes	Yes					Yes											
Mitochondrial membrane activity and depolarization	High	Low	Yes						Yes						Yes		Yes			
Competence as initial starting cell for TSC induction	High	Low	Yes	Yes					Yes								Yes			

Comments: Empty boxes – results have not been reported thus far

Figure 3

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

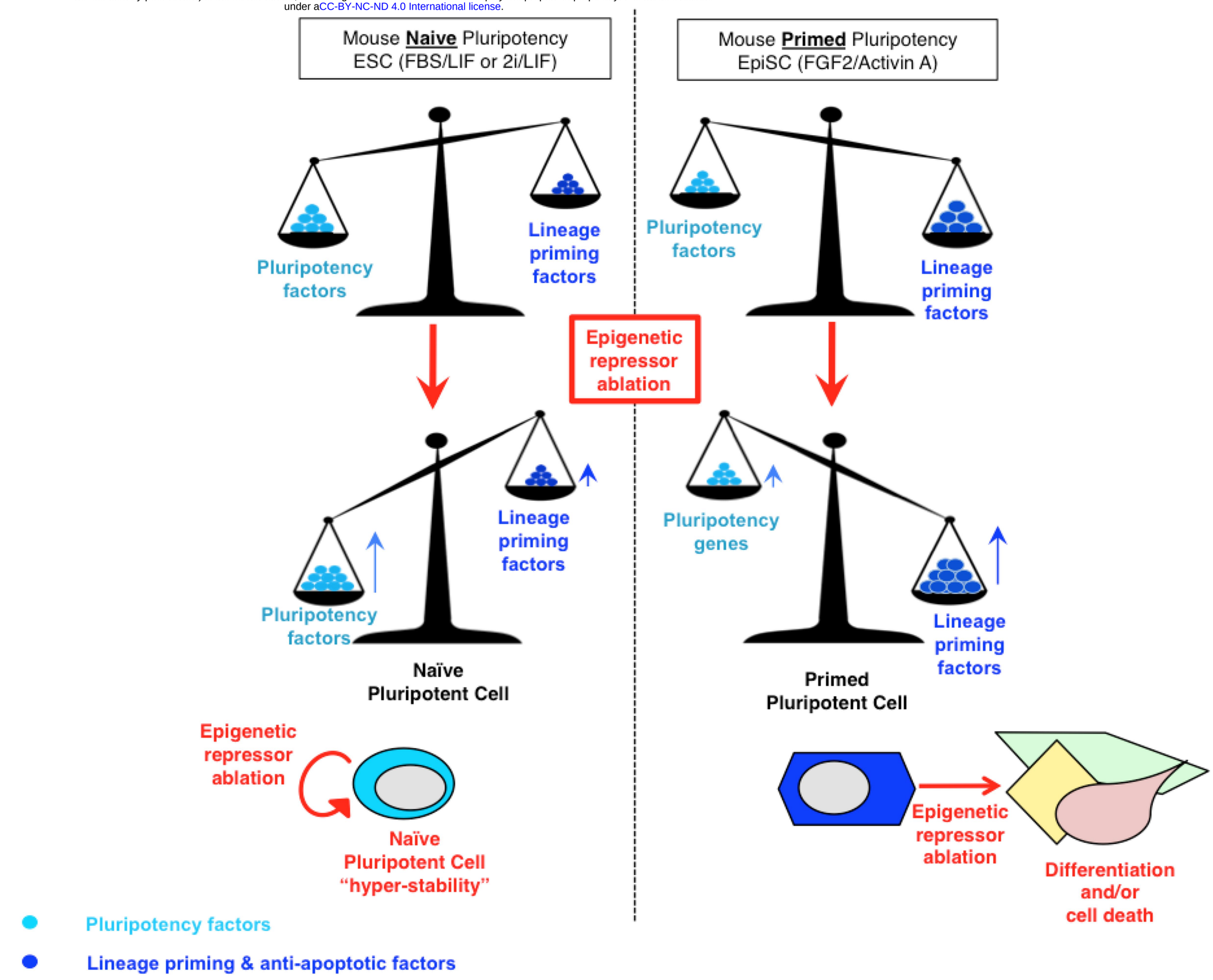
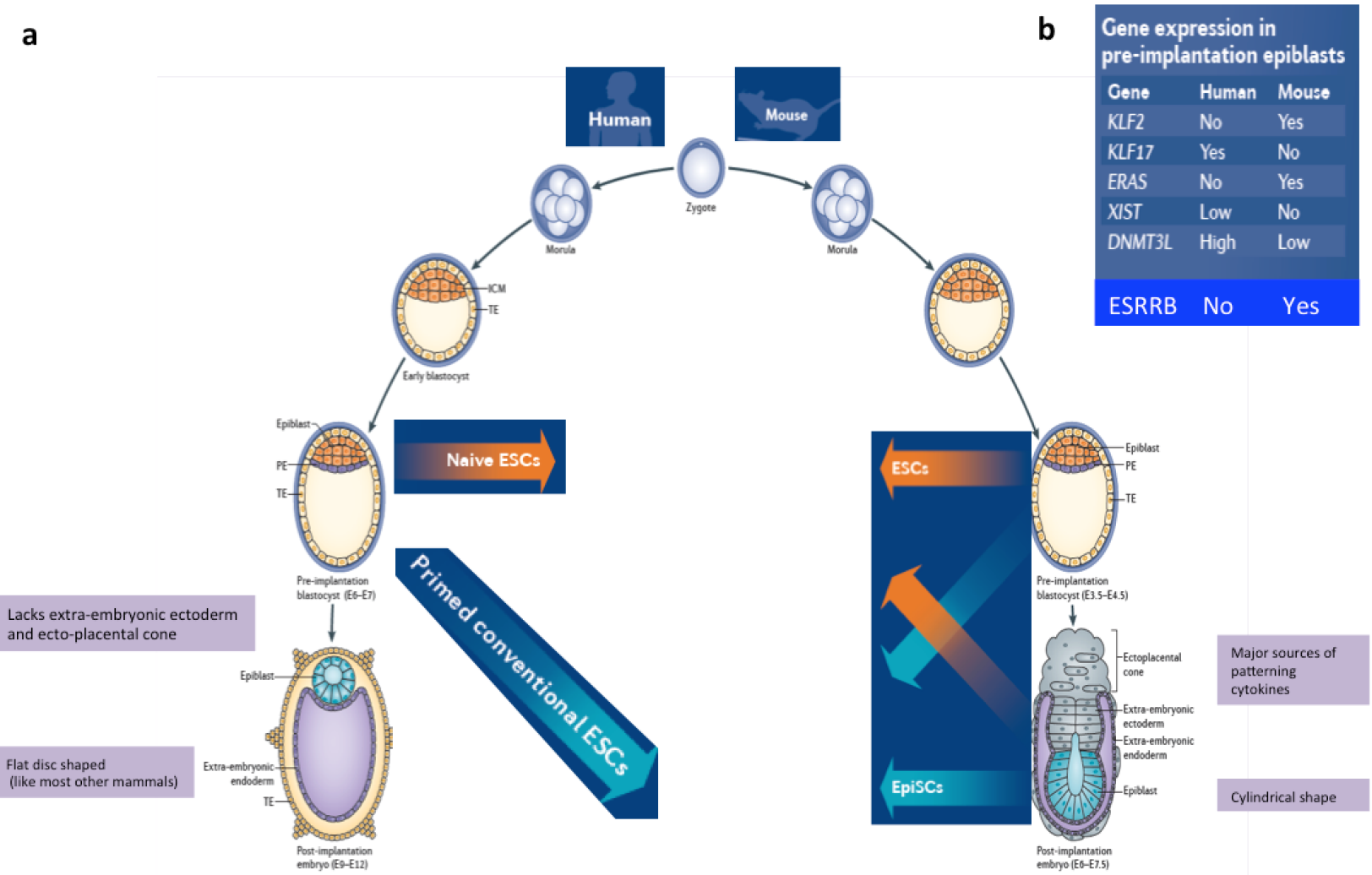


Figure 4

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.



Gene expression in pre-implantation epiblasts

Gene	Human	Mouse
KLF2	No	Yes
KLF17	Yes	No
ERAS	No	Yes
XIST	Low	No
DNMT3L	High	Low
ESRRB	No	Yes

Figure 5

bioRxiv preprint doi: https://doi.org/10.1101/030676; this version posted November 6, 2015. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

c

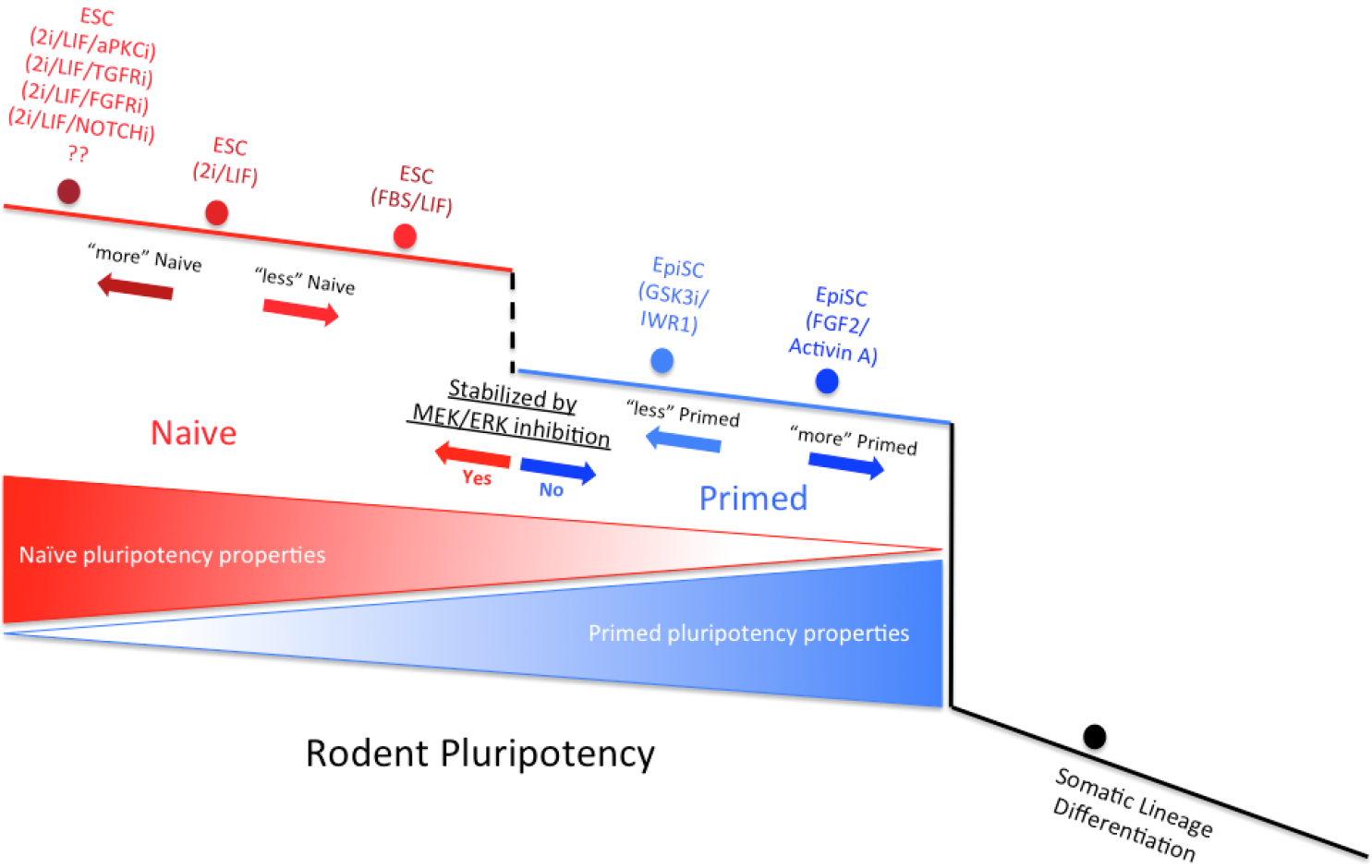


Figure 5