COGNITIVE DOMAINS FUNCTION COMPLEMENTATION

BY *NTNG* GENE PARALOGS

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ABSTRACT

Gene duplication was proposed by S.Ohno (1) as a key mechanism of a novel gene function

evolution. A pair of gene paralogs, *NTNG1* and *NTNG2*, sharing identical gene and protein

structures and encoding similar proteins, forms a functional complement subfunctionalising

(SF) within cognitive domains and forming cognitive endophenotypes, as detected by

Intellectual Quotient (IQ) tests (2). Both *NTNG* paralogs are associated with autism spectrum

disorder (ASD), bipolar disorder (BD) and schizophrenia (SCZ), with unique non-

overlapping segregation among the other 15 cognitive disorders (CD), emphasizing an

evolutionary gain-dependent link between advanced cognitive functions and concomitant

neurocognitive pathologies. Complementary expression and human brain transcriptome

composition of the paralogs explains the observed phenomena of their functional

complementarity. The lowest identity among *NTNGs* is found in a middle of encoded by

them proteins designated as uknown (Ukd) domain. *NTNG1* contains anthropoid-specific

constrained regions, and both genes contain non-coding conserved sequences underwent

accelerated evolution in human. *NTNG* paralogs SF perturbates "structure drives function"

concept at protein and gene levels. The paralogs function diversification forms a so-called

"Cognitive Complement (CC)", a product of gene duplication and subsequent cognitive

subfunction bifurcation among the *NTNG* gene duplicates.

INTRODUCTION

Complex behaviors arise from a combination of simpler genetic modules that either have

evolved separately or co-evolved. Many genes and proteins they encode have been found to

be involved in cognitive information processing with a single variant or a single gene

generally accounting for only a partial phenotypic variation of a complex trait, such as

cognition. Cognition as a quintessence of brain functioning can be viewed as a product of

intricately interlinked networks generated by deeply embedded into it gene-nodes with

specific or partially overlapping functions. The robustness of the cognitive processing

towards its single elements genetic eliminations (to study their function) and its simultaneous

fragility expressed in the multiple forms of neurological disorders manifest the existence of

cognitive domains interlocked but SF within a unit of cognition formed upon these domains

interaction. Previously, we have described a function of a pair of gene paralogs, *NTNG1* and

NTNG2, involved in human IQ tests performance, and underwent hominin-specific

evolutionary changes (2). Hereby, we report on these gene paralogs features focusing on

underlying mechanisms of their function segregation and complementation within the

cognitive domains.

RESULTS

The previously observed phenomena of functional complementation among the

NTNG paralogs within cognitive domains (2) is also manifested in *NTNG*-associated human

pathologies diagnosed in most cases (if only not in all) by a cognitive decline (Figure 1A-1

and A-2). Both genes are associated with BD and SCZ – devastating disorders sharing similar

etiology (3) with genetic correlation by multivariate analysis of 0.590 (4), linked to human

creativity (5), and characterized by impulsiveness as a common diagnostic feature (6).

Recently found associations of both paralogs with ASD (7) supports the reported genetic

correlation of 0.194 ASD/SCZ pair (4) and shared module eigengenes detected by PC1

among these two disorders (8). 12 *NTNG1*-linked CDs, ranging from AD to TS, span a broad

spectrum of clinical features frequently involving reduced processing speed (PS) and verbal

comprehension (VC, Figure 1A-1). As for *NTNG2*, working memory (WM) deficit and

inability "to bind" events (perceptual organization, PO) are the most prominent diagnostic

traits for the SLE and TLE patients (Figure 1A-2), with PN also being characterised by

indolent behavior in 90% of the cases (9). Interestingly to note that association of the

synapse-expressed *NTNG2* with both SCZ and autoimmune pathology (SLE) correlates with

a recent finding that human complement component C4 is involved in the synapse

elimination and SCZ development (10). Thus, both *NTNG* paralogs are associated with a

variety of CDs and mostly in a non-overlapping manner, except for ASD, BD and SCZ

characterized by shared and wide spectrum of cognitive abnormalities. The clinical etiology

of the aforementioned diseases supports the deduced by IQ functional complementation

existing among the *NTNG* paralogs (2) with (VC/PS) and (WM/PO) endophenotypic deficits

being uniquely segregated within the associated cognitive pathologies.

Since both gene paralogs are expected to have identical gene exon/intron

compositions but different in their intron lengths (11) we have reconstructed both paralogs

transcriptomes by re-processing the publicly available RNA-seq dataset (12) from healthy

and SCZ human subjects superior temporal gyrus (STG) post-mortem brain tissue

(Supplementary Table 1a=ST1a). A difference is noted instantly at the total expression levels

(genes, exons, individual RNA transcripts) when two gene paralogs are compared (Figure

1B-1 and B-2). *NTNG2* amount (as a whole gene) is 5 times larger comparing to *NTNG1*;

exons (2-5) are 3 times, exons (8-9) are 18 times and exon 10 is 4 times higher expressed for

NTNG2 than for *NTNG1*. The only two exons outlaying the prevailing amount rule for the

NTNG2 mRNAs are exons 6 and 7, expressed nearly at the same absolute level as for the

NTNG1 exon paralogs, making them highly underrepresented within the *NTNG2*

transcriptome. Next, distinct non-alternating splicing modules are formed by exons (2-5) for

NTNG1 (Figure 1B-1), while exons (4-5) and exons (8-9) for *NTNG2* (Figure 1B-2). Two

structurally identical RNA transcript paralogs (*NTNG1a* = G1a and *NTNG2a* = G2a) have

been found to exist in both *NTNG* transcriptomes with G2a being expressed at 8-9 times

higher level than G1a. *NTNG1* is uniformly presented across the all analysed 16 human

samples by 2 more protein coding RNAs (G1c and G1d, detected previously in mice brain,

Nakashiba et al., 2000) and by 2 non-coding intron (9-10) derived transcripts (Figure 1B-1).

At the same time, *NTNG2* transcriptome is comprised of one extra potentially coding RNA

(G2a-like with exon 2 spliced out but in-frame coding preserved) and 2 assumed to be non-

coding RNAs with exons 6 and 7 retained along with preceding and following them introns.

Quite interesting that these two latter transcripts are the only RNA species with *NTNG2* exon

6 and 7 retained (Figure 1B-2). Two more coding (G1f and G1n) and 4 more non-coding for

NTNG1 and 9 extra non-coding for *NTNG2* RNA species have been also assembled from the

available reads but due to inconsistency in their appearance across all 16 STG samples they

are not presented on the figure but summarized in the table (Figure 1C, for details refer to

ST1d). Summarising above, quantitative and qualitative complementary differences is a

prominent feature characterising the brain RNA transcriptome of human *NTNG* paralogs.

However, no significant changes at the transcription level of neither whole genes, nor

individual exons, nor reconstructed RNA transcripts have been found when SCZ and healthy

subjects samples are compared.

Upon calling the presence of IQ-affecting SNPs (2) across all STG samples (ST1c) it

has been revealed that 15 out of 16 subjects were positive for the T-allele of rs2149171 (exon

4-nested), shown above to attenuate the WM score in SCZ patients, and making a comparison

among the allele carrier *vs* non-carrier impossible. Four healthy and three SCZ samples carry

a T-allele of rs3824574 (exon 3-nested, non-affecting IQ), and 1 healthy and 1 SCZ sample

each contains a C-allele of rs4915045 (exon10, non-coding part-nested, and non-affecting IQ).

Thus, among the eleven cognitive endophenotype-associated SNPs only 3 were possible to

call out of the available *NTNG* transcriptome.

Distinctly complementary nature of the *NTNG* paralogs segregation within

neurological disorders and RNA transcriptome usage in STG (Figure 1) has prompted us to

analyse both genes expression across the entire human brain. We have reconstructed both

genes expression profiles in the human brain areas over the life span from conception (pcw =

post-conception week) to mature age (30-40 yrs old) using the RNA-seq data from BrainSpan

(www.brainspan.org). Similarities and differences are easily noted when the age-dependent

phases of *NTNG1* and *NTNG2* expression profiles are matched (Figure 2). Based on the

visual inputs three distinct classifiers have been elaborated: 1. predominantly synchronous

(Figure 2A(1-4)), characteristic mostly for the cortical areas; 2. predominantly mixed and

asynchronous (Figure 2B), characteristic for the cerebellar cortex and subcortical formations;

and 3. anti-phasic (complementary, Figure 2C), characteristic for the MD of thalamus and

hippocampus. All analysed brain areas demonstrate an elevated level of *NTNG2* expression in

comparison to *NTNG1* except for thalamus (Figure 2C) with the largest difference observed

at the time of birth (35-37 pcw) or soon after (4 mo) for the synchronous classifiers (Figure

2A), oscillating increment values across the life span for the mixed (Figure 2B) and anti-

phasic (Figure 2C) classifiers. It is quite intriguing to note that essentially all brain areas

show a trend towards the expression difference being negated between the paralogs by

reaching the mature age of 30-40 yrs old (nearly or above the mean age used for the IQ

testing), except MD where the expression discrepancy is increased. Thus, the observed

functional complementation among the *NTNG* paralogs is supported by the anatomical

distribution of the genes in human brain and their expression pattern modality over the

human subjects lifetime.

A direct comparison of the *NTNG* paralogs shows not only identical intron-exon gene

structure (Figure 1B-1, 2B-2) but also closely matched exon sizes (Figure 3A). There are

three exons of identical sizes (exons 4, 8 and 9), another three exons differed by one encoded

amino acid (exons 3, 5 and 6) and there are exons of different sizes (exons 2, 7 and 10). In

terms of size the largest difference among the genes is visually presented by the introns:

intron (9-10) of *NTNG1* is 52.7 times larger its *NTNG2* paralogous intron with intron (6-7) of

NTNG1 being only 1.43-times larger pointing towards non-equilibria process of non-coding

elements elaborations as the process of gene paralogs SF proceeded. Nevertheless, it can be

generalised that in average all *NTNG1* introns are several times larger their *NTNG2* analogs

(Figure 3A). We have shown previously that exons 6 and 7 are differentially used within the

brain *NTNG* transcriptome (Figure 1B-1 and B-2) and to explore their potential contribution

into the paralogs SF we have built identity matrices with these exons being excluded and

included (but still producing in-frame existing transcripts, Figure 3B-1 left and right panels,

respectively). Exclusion of both exons from the full-lengths transcripts (thus converting

NTNG1m to *NTNG1a* and *NTNG2b* to *NTNG2a*, respectively) increases the identity of DNA

on 2% (a relatively large effect since both exons together represent only 7.22 and 9.69% of

the total coding part of the full-length RNA transcripts, *NTNG1m* and *NTNG2b*, respectively).

This effect becomes even stronger when the encoded by these transcripts proteins are also

compared (Figure 3B-2). The spliced out Ukd protein domains (encoded by the exons 6 and

7) increases the proteins identity on 3.8% thus making the middle of both genes (and encoded

proteins) substantially more different among the both gene paralogs. To corroborate this

observation and to explore the importance of other protein parts we have directly compared

the sequences encoded by the full-length transcripts and producing Netrin-G1m and Netrin-

G2b (Figure 3C). Similarly to what has been shown on Figure 3B-1 and 3B-2, the lowest

identity (17.5%) is represented by the Ukd domain (encoded by the exons 6 and 7) and by the

preceding it exon 5 (a 3'-part of the LE1 domain). Two other areas also show a substantially

low identity, namely the N-terminus (it includes the protein secretory signal indicated by an

arrow) and the outmost C-terminus responsible for the unique feature of Netrin-Gs – the GPI

attachment. Thus, based on the percent identity comparisons among the Netrin-G paralogs it

can be predicted that there are several potential protein parts contributing to the paralogs SF.

As it has been reported by Seiradake et al. (13), identical gene and protein domain

compositions result in the identical structural motif with differences only in the spatial

arrangement of the loops facing the post-synaptic Netrin-G's interacting partners, NGL-1 and

NGL-2, respectively (Figure 3D). Loop I binding surfaces alignment (Figure 3C, blue color)

shows a high level of conservation (with at least 5 amino acids 100% conserved) among the

Netrin-G paralogs, indicating that it is unlikely to be responsible for the cognate ligand

binding specificity. Neither Loop II (Figure 5C, yellow color) nor Loop III (Figure 5C,

orange color) display a single conserved amino acid shared among the paralogous binding

interfaces (as it has originally been described in (13)). Thus the complementary pattern of the

pre-postsynaptic interactions mediated via specific Netrin-G/NGL pairs is reflected in the

reciprocally different sizes of the loops binding interfaces representing another element of the

NTNG-encoded protein paralogs SF.

DISCUSSION

Complementary contribution of *NTNG* paralogs into human cognitive pathologies.

Involvement of the pre-synaptically expressed axon-localised *NTNGs* in SCZ diagnosis

supports the established view of SCZ as a product of distorted trans-synaptic signaling (14),

with a recent study also proving that axonal connectivity-associated genes form a functional

network visualisable by fMRI (15), and that brain connectivity predicts the level of fluid

intelligence (16, 17). Both *NTNGs* have been found to participate in the brain functional

connectivity found by the parcellated connectome reconstruction (18). Most of the reported

disease associations link *NTNG1* to SCZ with a variety of other neurologic pathologies (15 in

total, Figure 1A-1), while *NTNG2* pathologic associations (6 in total, Figure 1A-2) are quite

limited to those affecting WM or PO. Among them is SLE frequently characterized by WM

deficit (19) and also known to represent schizoid-type abnormalities characteristic for

autoimmune pathologies (20, 21). Immune activation is known to lead to altered pre-pulse

inhibition (a key diagnostic trait for SCZ) reversed by antipsychotics (22). The three diseases

associated with both paralogs (ASD, BD and SCZ) are also a primary focus of the recently

initiated PsychENCODE project (23). It is also worth to mention the resemblance of the

reported disease associations with the behavioral phenotypes of *Ntng1* and *Ntng2* gene

knockout mice (24).

A gene content associated with attenuated IQ score often relates to numerous diseases,

such as SCZ, ASD, depression, and others (see (25) for ref.; 26) with several of them also

undergone positive selection following the human brain evolution (27). Despite the fact that

global network properties of the brain transcriptome are highly conserved, among the species

there are robust human-specific disease-associated modules (28) and human accelerated

regions (HARs) - highly conserved parts of genome that underwent accelerated evolution in

humans (29). HARs can serve as genomic markers for human-specific traits underlying a

recent acquisition of modern human cognitive abilities by brain (30) but that also "might

have led to an increase in structural instability… resulted in a higher risk for

neurodegeneration in the aging brain" (31), rendering our intellectual abilities genetically

fragile (32) and resulting in a variety of CDs. The role genomic context, epistasis (33), plays

in the evolution and pathology is manifested by frequently found disease-causing alleles

present in animals without obvious pathological symptoms for the host (34). Any CD is

characterized by general intellectual disability (GID) plus psychiatric symptoms. A genetic

perturbation-exerted behavioral cognitive deficit (BCD) in an animal model organism is a

poor match to a human CD *per se* due to very poor contextual resemblance between the

human GID and animal BCD together with the absence of interpretable psychiatric symptoms.

Usefulness of animals as psychiatric models is also compromised by the fact that

transcriptome differences within species tissues is smaller than among the homologous

tissues of different species (35, 36). No wonder that the compounds that "cure" mice models

consistently fail in human trials (discussed in (37)).

NTNG paralogs brain transcriptome intrinsic complementarity and possible mechanism for

the IQ-affecting mutation alleles effect. There is no global change at the mRNA level

between healthy subjects and SCZ patients (Figure 1B). This conclusion is supported by

previously published works stating that globally altered mRNA expression of *NTNG1* or

NTNG2 is unlikely to confer disease susceptibility, at least in the temporal lobe (38), and

Brodmann's area (39). However, the original paper-source of the STG samples RNA-seq

along with many other genes (>1,000) found that *NTNG1* (but not *NTNG2*) falls within the

group of genes with significant alternative promoter usage ((12): ST6, $p<9.05E-10$ at FDR

<0.5) and *NTNG2* (but not *NTNG1*) clusters with genes (>700) with significant alternative

splicing change ((12): ST7, $p<6.15E-12$ at FDR<0.5) when SCZ and controls are compared.

Such GWAS observation adds an extra layer of complementary regulation to both *NTNG*

paralogs on a top of the described in the results section complementary usage rule for the

exons, formed by unspliced splicing modules, resulting transcripts and comprising them

exons (Figure 1B). Based on the available RNA-seq dataset it was almost impossible to

detect RNA with the matched position of *NTNG* SNPs used for the IQ testing (ST2c) except

for two coding exons located (rs2149171 and rs3824574) and exon 10 non-coding area

located but transcribed rs4915045 (in 2 out of 16 samples). This fact points towards indirect

effect of the IQ-affecting mutation alleles potentially associated with shorter (secretable)

isoforms generation (Prosselkov et al., unpublished) lacking two of the most prominent

NTNG features: GPI-link and the Ukd domain through an aberrant splicing factor binding.

The GPI-link is a hallmark of Netrin-G family members (40, 41) and lacking it the aberrant

Netrin-G isoforms are likely to mimic the action of their releasable ancestry molecules -

netrins, still being able to bind to their cognate postsynaptic ligand – NGL but without

forming an axonal-postsynaptic contact and potentially dominant negative consequences. The

Ukd domain of Netrin-G1, despite its so-far unknown function, is involved in lateral binding

to the pre-synaptically localised LAR modulating the binding strength between NGL-1 and

Netrin-G1 (42). Work is currently underway in search for a similar lateral interaction partner

for the Netrin-G2 Ukd domain. Inclusion of the Ukd-encoding domain exons 6 and 7 is

regulated by the Nova splicing factor (43) affecting the cortex Netrin-G1 exon 7 but not exon

6, and, simultaneously, Netrin-G2 paralog exons exhibiting an opposite pattern. In general, it

is tempting to speculate that deregulation of *NTNG* transcripts processing may have a role in

the brain-controlled cognitive abilities and associated CDs. Supporting such notion, a

decreased level of Netrin-G1c mRNA (exons 6-9 excluded, Figure 1B-1) has been reported

for BD and SCZ (38) with Netrin-G1d (exons 6 and 7 included but 8-9 excluded, Figure 1B-

1) and Netrin-G1f (a secretable short isoform consisted of domain VI only and lacking the

Ukd and GPI-link) being increased in BD, but not in SCZ, in anterior cingulated cortex (44).

Higher Netrin-G1d mRNA expression in fetal brain but low for the Netrin-G1c isoform in the

human adult (38) indicates different functionality of these two splice variants joggling with

the Ukd domain inclusion/exclusion pattern. And, according to our unpublished data, if

Netrin-G1 Ukd-containing isoforms are the dominant isoforms in adult mouse brain, Netrin-

G2 Ukd-containing isoforms are present only at the trace level (Prosselkov et al.,

forthcoming), resembling the human STG samples transcriptome pattern (Figure 1B-1 and B-

2). A similar "dynamic microexon regulation" associated with the protein interactome

misregulation has been reported to be linked to ASD (45).

Synchronous and complementary expression of *NTNG* paralogs in the human brain supports

the IQ-associated cognitive endophenotypes. Influential parieto-frontal integration theory (P-

FIT, (46)) states that general intelligence ("g") is dependent on multiple brain cortical areas

such as dlPFC, Broca's and Wernicke's areas, somatosensory and visual cortices (47). Despite

"g" is widely accepted as the only correlate of the intelligence, its unitary nature was

challenged by (48) claiming had indentified two independent brain networks (for memory

and for reasoning) responsible for the task performance, the idea later criticised for the

employed data processing approach (49). Higher IQ scores (a composite surrogate of "g")

have been reportedly associated with the fronto-parietal network (FPN) connectivity (50, 51).

Higher levels of *NTNG* paralogs expression within the cognition intensively loaded areas of

the brain and the distinct patterns of expression profiles (synchronous, asynchronous/mixed,

and complementary, Figure 2A) support associations of *NTNG1* and *NTNG2* with the

recorded cognitive endophenotypes (2). Based on the expression patterning over the human

life-span, among the total 16 analysed brain areas we found two falling under the same "anti-

phasic (complementary)" classifier (Figure 2C): HIP and MD. Adding more to that, MD is

the only brain area (out of the 16 presented) where *NTNG1* expression level exceeds that of

NTNG2 making it a promising candidate for the phenomena of *NTNGs* SF explanation. Two

other brain areas classified by a synchronous paralogs expression deserve a special attention,

dlPFC and mPFC (Figure 2A-4). PFC circuitry has been known as a "hub of the brain's WM

system" (52, 53), which acts through direct HIP afferents (54) and has many connections

with other cortical and subcortical areas (55). mPFC may function as an intelligence-control

switchboard and lPFC, part of the FPN global connectivity, predicts the WM performance

and fluid intelligence (56). Interactions of the auditory recognition information fed by the

vPFC stream with the sequence processing by the dorsal stream are crucial for the human

language articulation (57; 58). The fact that both *NTNG* paralogs are extensively expressed

across PFC (Figure 2A-2 and A-4) pinpoints this area as a key for future molecular studies of

NTNGs and the human-unique symbolic communications. And PFC is not only implicated in

many psychiatric disorders, including SCZ ((59); see also (55) for ref.), but is also the only

brain structure unique to primates without known homologs in the animal kingdom (60).

Evolution of the protein paralogs encoded by the *NTNGs*. Forkhead box P2 (FOXP2) – a

ubiquitously expressed transcription factor that has been reportedly linked to the evolution of

human language through T303N, N325S substitutions when compared to a primate ortholog

(61), is 100% identical to Nea protein (62). FOXP2 regulates expression of multiple genes in

human and chimpanzee (63), and among them is an M3 gene brain module representative

responsible for general fluid cognitive abilities (26), *LRRC4C*, a gene encoding NGL-1, a

post-synaptic target of Netrin-G1. Similarly to FOXP2, Netrin-G1 is a 100% conserved

protein among the hominins with only 1 mutation found in chimpanzee which is absent in

marmoset (and other primates) and mice proteins (2). On the other hand, extinct hominins'

Netrin-G2 relative to modern human contains T346A point mutation (as per current version

of hg19), also found in primates and mouse and known as rs4962173 (dbSNP missense

mutation) representing an ancient substitution from Neandethal genomes found in modern

humans and reflecting a recent acquisition of the novel allele around 5,300 yrs BC. Nothing

is known regarding the functional significance of this mutation allele but biochemically a

substitution of alanine (A) on a polar threonine (T) could bring an extra point of post-

translational modification, e.g. a phosphorylation or glycosylation (NetPhos2.0 (64) assigns a

low probability score for the T346 to be phosphorylated but NetOGlyc4.0 (65) robustly

predicts it to be glycosylated, see SM). Another mutation, S371A/V, reflects a selective

sweep in Netrin-G2 protein from primates to hominins within a similar to T346A functional

context when a hydrophobic alanine (in chimpanzee, A)/valine (in marmoset, V) is replaced

by a polar serine (S) with a strong positive predictions for glycosylation but not

phosphorylation (see SM). This poses a question whether these two human-specific protein

substitutions associate with advanced cognitive traits as they may represent a hidden layer of

poorly studied so far protein glycosylation-associated regulatome known to affect the brain

function and diseases (66, 67). Adding more to this, T346 is nested on exon 5 just 20 nu away

from the affecting WM score rs2274855 mutation allele (2), and, together with S371A/V,

they are both located within the lowest percent identity area (exons (5-7)) of Netrin-Gs

(Figure 3C) and, proposedly, contributing to the *NTNG* duplicates SF. There are at least three

more protein parts potentially contributing to the gene paralogs specialised function

subdivision (based on the low identity scores, Figure 3C): the secretory peptide, the GPI-link,

and the outmost structurally elaborated unstructured loops (I-III) responsible for the

reciprocal binding of Netrin-Gs to their post-synaptic cognate partners, NGL-1 or NGL-2,

both containing a C-terminal PDZ-binding domain (68). An interesting finding was reported

in (69) reported a presence of SH3(PSD95) domain binding site (required for the

phosphatidylinositol-3-kinase recruitment) in mice Netrin-G2 (100% identical to human) but

not in Netrin-G1. The detected SH3 binding site overlaps with the Netrin-G2-loop III

responsible for the binding specificity to NGL-2 (12, 70, 71). A plausible working hypothesis

would be that while internalised (and being GPI-link naïve/immature) the pre-synaptic

Netrin-G2 is bound to SH3-PSD95 via loop III but as soon as being secreted extracellularly

(and being attached to the membrane) it is bound to post-synaptic NGL-2. Corroborating this,

in the absence of Netrin-G2 in the KO mice NGL-2 is unstable on the post-synaptic surface

and gets quickly internalised (24). We can only speculate regarding the potential importance

of PSD-95(SH3)-Netrin-G2-NGL-2 scaffolding loop interaction/competition but the ability

for Netrin-G1 to bind to SH3 has not been reported. Following this logic, Netrin-G1 should

have a similar binding partner via loop II.

The overall identical structural scaffold among the Netrin-G paralogs (Figure 3D) is

likely to represent an anciently preserved one of the primordial protein (encoded by a single

gene in the primitive urochordate *C.intestinalis*) and its contribution to the process of SF

among the *NTNG* paralogs goes against the "structure drives function" concept. It looks like

that it is not the "structure" but rather the "evolution" itself that drives a selection for the best

structural (or unstructural in our case) fit out of the available frameworks provided by the

gene duplicates to fulfill the emerged functional demand in a new ecological niche. The

intricate variability of phenotype is grounded by the conserved nature of genotype and

constrained by the "structure-function" limitations of the coding DNA and is only possible

due to permissive evolutionary continuing elaborations of non-coding areas able to absorb the

most recently acquired elements (having a potential to become regulatory at some point, e.g.

like HAR5 (30)) and carried over by the neutral drift. At the same time, the multiple protein

substitutions coinciding with the SF labor segregation phenomena among the Netrin-G

paralogs question their neutral nature. Both of them undergo a purifying selection from mice

to human through the reduction in size of non-coding DNA (introns) and encoded proteins

(the mice Netrin-G2 is 2 aa longer its human ortholog) further contributing to the host-

specific SF. Thus while the non-coding sequences are used to explore the evolutionary space

in time, the restrictive boundaries of the paralogs SF are determined by the protein

(unstructured) elements.

Molecular evolution of the Cognitive Complement (CC). Appearance of the neural crest (72),

an event that "affected the chordate evolution in the unprecedented manner" (73), multipotent

progenitor cells (74), and neurogenic placodes (suggesting a chemosensory and

neurosecretory activities (75)) in the first primitive urochordates/tunicates coincides with the

presence of *Ntng* precursor gene (ENSCING00000024925) later undergoing two rounds of

duplication events in fish and found to affect human cognitive abilities (2). *NTNG* paralogs

are expressed in the human neural crest-forming cells with *NTNG2* 10 times stronger than

NTNG1 (76), both are differentially expressed in human comparing to chimpanzee and rhesus

monkey with *NTNG2* expression model showing stronger probability than *NTNG1* (77), and

both are stronger expressed in human telencephalon comparing to chimpanzee and macaque

(78). *NTNG1* has been classified as a brain module hub gene "whose pattern fundamentally

shifted between species" (18). Belonging to distinct modules of brain expression regulation

(78, 79), *NTNGs* are classified as "genes with human-specific expression profiles" (79). The

nearby gene ~260 kbp upstream of *NTNG2* is *MED27* (mediator of RNA polymerase II) has

been proposed to be associated with the evolution of human-specific traits (80). *NTNG1* has

also been reported among the "adaptive plasticity genes" (81) potentiating rapid adaptive

evolution in guppies (*NTNG2* was not found in the input RNA).

Complementarity among the *NTNG* paralogs and encoded by them proteins has been

reported previously: brain expression complementary pattern (in almost self-exclusive

manner) defined by the 5'-UTR-localised *cis*-regulatory elements (82); complementary

distribution within the hippocampal laminar structures (83); axon-dendrite synaptic ending

resulting in differential control over the neuronal circuit plasticity (84); mutually-exclusive

binding pattern to post-synaptic partners, NGL-1 and NGL-2, dictated by the protein

unstructural elements (13); alternative promoter usage *vs* alternative mRNA splicing (12) and

increased coefficient of variation (CV, ST1d) for *NTNG1* expression but not *NTNG2* in SCZ

patients (also reported in (85)); KO mice behavioral phenotypes and subcellular signaling

partners complementarity (24); "differential stability" brain modules expression (*NTNG1* is

expressed in the dorsal thalamus (M11) as a hub gene (Pearson's 0.92) while *NTNG2* is in

neocortex and claustrum module (M6, Pearson's 0.65)) (18); hypocretin neurons-specific

expression of *NTNG1* (but not *NTNG2*) as a sleep modulator (86); top-down *vs* bottom-up

information flows gating in mice and differential modality (Prosselkov et al., forthcoming),

and human IQ-compiling cognitive domains complementation (2). The current study reports

on *NTNGs* complementarity association with the CDs (Figure 1A); mRNA splicing pattern

complementary at the quantitative and qualitative levels via differential use of the middle-

located exons (Figure 1B); brain complementary oscillatory expression over the human life

span observed in the intensive cognitively loaded brain areas (Figure 2); AE of the paralogs-

segregated unique non-coding elements (Figure 3A); complementary pattern of the protein

orthologs (mice-to-human) protein sequence evolution. Such multi-level complementation is

likely to reflect a shared evolutionary origin from a single gene in a primitive vertebrate

organism 700 mln yrs ago and its subsequent functional segregation among the evolution-

generated gene duplicates in jawless fish, such as lamprey.

Occupying independent but intercalating functional niches, *NTNG1* and *NTNG2* do

not compensate but complement each other's function forming a "functional complement" of

genes. Half a billion yrs ago the doubled gene dosage led to the gradual SF and manifested in

a function complementation within the cognitive domains, at least in human. We would like

to coin such gene pair as a Cognitive Complement (CC).

CONCLUSION

The emerged functional redundancy, as an outcome of gene duplication, leads to function

subdivision and its bifurcation among the gene paralogs resulting in the paralogs SF. A

functional compensation is known to exist among the evolutionary unrelated genes but has

not been reported among the gene paralogs, more frequently characterized by the function

complementation. Gene paralogs structural identity (at both, gene and protein levels) does not

provide a substrate for function compensation but rather for complementation, perturbating

"structure drives function" rule. A gene duplication event of a tunicate *NTNG* primordial

gene and the subsequent process of its function specialisation (driven by the new ecological

niches appearance and evolution) among the gene duplicates made them to SF into distinct

cognitive domains in a complementary manner forming a CC. In our forthcoming work we

are to describe how *Ntng* mice genes function resembles that of human orthologs (Prosselkov

et al., forthcoming).

MATERIALS AND METHODS

Human brain *NTNG* transcriptome reconstruction. Relates to Figure 1B and 1-C. The original

source of the dataset was produced by ((12): E-MATB-1030) and the downloaded .bam files

used for the re-processing are listed in ST1a. All reconstructed transcripts are presented in

ST1d standalone Excel file. Two samples were excluded from the analysis due to failed "per

base sequence quality" measure, and zero expression level for *NTNG1a* and *NTNG1int(9-10)*

otherwise consistently expressed throughout other samples (ST1b). SAMtools software was

used for the SNPs calling from the available RNA-seq datasets (ST1c). For details refer to

SM.

Human brain expression profiling for *NTNGs* across the life span. The original source of data

was www.brainspan.org. All available samples were initially included into the analysis but

two of them excluded at a later stage (MD for 12-13 pcw and mPFC for 16-19 pcw) due to

high deviation (6-7 times) from the mean for other replicas. The mean expression values per

each brain area as RPKM were plotted against the sampling age. Profiles classification was

done visually considering the trend over the all plotted points as an average.

NTNG1 (*NTNG1m*) and *NTNG2* (*NTNG2b*) full-length mRNA transcripts assembly. Relates

to Figure 3B. Human *NTNG1m* brain transcript has been reported previously (125) and we

have also confirmed its ortholog presence in the mice brain via full-length cloning (42). Since

NCBI contains only its partial CDS (AY764265), we used the RNA-seq-generated exons

(Figure 1B) to reconstruct its full-length and to generate an ORF of the encoded Netrin-G1m.

Similarly, human *NTNG2b* was reconstructed from the RNA-seq dataset and from Ensemble

as follows. Exon 5 sequence was deduced from ENST00000372179, other exons were from

ENST00000467453 (no longer available on the current version of Ensemble) except for exon

6 deduced by running three independent alignments against the human genomic DNA with

the mice 3'-intron (5-6), exon 6, and 5'-intron (6-7) concomitantly confirmed by the

generated full-length ORF for Netrin-G2b. The reconstructed protein was predicted to encode

587 amino acids, which is in a close proximity to the mice netrin-G2b ortholog of 589

residues (Prosselkov et al., forthcoming).

Full-lengths gene structures of *NTNG* paralogs reconstruction. Relates to Figure 3A. Both,

the obtained above from the STG brain samples RNA-seq and the reconstructed full-lengths

transcripts carrying all stably expressed exons were used to confirm the intron-exon junctions

positioning for *NTNG1* and *NTNG2*. Due to observed variability in the intron (1-2) and exon

10 sizes their boundaries were left unmarked.

SUPPLEMENTARY MATERIALS (SM)

Contain Supplementary Methods (RNA-seq of STG re-processing and SNPs detection) and

Supplementary Tables (ST1a-d, ST2) as a single compiled pdf file. Reconstructed RNA-seq

(.gtf) of the STG is presented as a standalone Excel file (ST1d). Also included: Netrin-G2b

predicted phosphorylation and *O*-glycosylation, Netrin-G1 *vs* Netrin-G2 Ukd alignment (87),

predicted secretory peptide cleavage and GPI attachments.

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COMPETING INTERESTS

Authors would like to express a lack of any competing interests associated with the work.

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Figure 1. *NTNG* paralogs complementation within neurological disorders and brain transcriptome. (A-1, A-2) Reported cognitive disorders associations for *NTNG1* and *NTNG2*. *denotes rather an indirect association via a direct interaction with the research target. (B-1, B-2) RNA-seq of the STG of healthy (circle) and SCZ (cross) human subjects. The original dataset was produced by **Wu et al. (12)**, accession number E-MTAB-1030 on ArrayExpress (**ST1a**) and reprocessed as described in **SM**. Five *NTNG1* and four *NTNG2* transcripts, consistently expressed across all 16 human samples are shown. Two samples (one healthy and one SCZ) have been omitted due to unsatisfactory quality of reads and expression profiling (**ST1b**). For the SNPs calling by SAMtools see **ST1c**. Data are presented as a mean RPKM±SEM. (C) Total number of the assembled transcripts across all samples for both paralogs (see **ST1d** for the completely reconstructed *NTNG* transcriptome). Dash-outlined are co-spliced exon clusters.

A-1

primary auditory cortex (core) (A1C)

no data

posterior (caudal) superior temporal cortex (area 22c) (STC)

inferolateral temporal cortex (area TEv, area 20) (ITC)

TCx/ITC

primary visual cortex (striate cortex, area V1/17) (V1C)

OCx

8-9 pcw, n=1-2
12-13 pcw, n=5-6
16-19 pcw, n=2-5
21-26 pcw, n=2-3
35-37 pcw, n=1
4 mo, n=2-3
10-12 mo, n=1-2
2-4 yrs, n=2-4
8 yrs, n=2
11-15 yrs, n=2-3
18-23 yrs, n=3-4
30-40 yrs, n=4

A-2

ventrolateral prefrontal cortex (vPFC)

orbital frontal cortex (OFC)

posteroventral (inferior) parietal cortex (IPC)

PCx

8-9 pcw, n=1-2
12-13 pcw, n=6
16-19 pcw, n=3-5
21-26 pcw, n=2
35-37 pcw, n=1-2
4 mo, n=1-2
10-12 mo, n=1-2
2-4 yrs, n=2-4
8 yrs, n=1-2
11-15 yrs, n=2-3
18-23 yrs, n=4
30-40 yrs, n=4

A(1-4). Predominantly synchronous paralogs expression classifiers

A-3

primary motor cortex (area M1, area 4) (M1C)

+S1C

primary somatosensory cortex (area S1, areas 3,1,2) (S1C)

+M1C

8-9 pcw, n=2
12-13 pcw, n=3-6
16-19 pcw, n=1-5
21-26 pcw, n=1-3
35-37 pcw, n=1
4 mo, n=1-2
10-12 mo, n=1-2
2-4 yrs, n=2-3
8 yrs, n=1-2
11-15 yrs, n=1-2
18-23 yrs, n=3-4
30-40 yrs, n=3-4

A-4

dorsolateral prefrontal cortex (dlPFC)

anterior (rostral) cingulate (medial prefrontal) cortex (mPFC)

B. Predominantly asynchronous and mixed paralogs expression classifier

amygdaloid complex (AMY)

striatum (STR)

LGE

cerebellar cortex (CBC)

URL   CB(C)

8-9 pcw, n=2
12-13 pcw, n=4-6
16-19 pcw, n=2-4
21-26 pcw, n=2-3
35-37 pcw, n=1-2
4 mo, n=2-3
10-12 mo, n=1-2
2-4 yrs, n=2-4
8 yrs, n=1-2
11-15 yrs, n=1-3
18-23 yrs, n=3-4
30-40 yrs, n=4

RPKM (Reads Per Kilobase of transcript per Million mapped reads)

C. Anti-phasic (complementary) paralogs expression classifier

hippocampus (hippocampal formation) (HIP)

mediodorsal nucleus of thalamus (MD)

DTH   DTH/MD

RPKM (Reads Per Kilobase of transcript per Million mapped reads)

Figure 2. *NTNG* paralogs expression dynamics classification (A-C) in the human brain across the life span. A (1-4): further subdivision of the classifier. RNA-seq data are from the BrainSpan (www.brainspan.org) presented as a mean±SEM. TCx = temporary neocortex; OCx = occipital neocortex; PCx = parietal neocortex; LGE = lateral ganglionic eminence; CB(C) = cerebellum(cortex); DTH = dorsal thalamus; URL = upper (rostral) rombic limb; pcw = post-conception week. Two data points (MD, 12-13 pcw, and mPFC, 16-19 pcw) for *NTNG1* expression were omitted as they were 6-7 times different from the mean of other replicas. All processed brain samples are listed in **ST2**.

Figure 3. Human *NTNG* paralogs DNA and protein sequence comparisons and "structure-function" rule incongruency. (A) Identical gene structures with different sizes of introns. RNA-seq data from **Figure 1B** were used to precisely deduce the exon/intron junction boundaries. The sizes of exons 1, 10 and introns (1-2) are not indicated due to observed among the splice transcripts lengths variability (see **ST1a** for details). Arrows indicate location of CNS = conserved non-coding sequences underwent accelerated evolution in human compare to mice (mCNS) and chimpanzee (chCNS), as per **(124)**; and ASC = anthropoid-specific constrained regions in human compare to marmoset (maASC), as per **(121)**. (B) Identical exonal composition of the longest *NTNG* encoded RNA paralog transcripts and corresponding proteins with relatively high percent of identity among them dependent on the included/excluded Ukd domain (**B-2**) encoded by the exons 6 and 7 (**B-1**). Notably, the protein sequence represents higher percent of the paralogs difference than encoded it DNA. The matrices were obtained by GeneJockey II (Biosoft). (C) Protein alignments for the longest human *NTNG* encoded proteins, Netrin-G1m and Netrin-G2b, with Loops I-III highlighting binding sites for their cognate post-synaptic binding partners NGL-1 (*Lrrc4c*) and NGL-2 (*Lrrc4*), respectively, as determined by **Seiradake et al. (13)**. Arrow indicates a putative secretory cleavage site location, as calculated by SignalIP **(122)**, the blue rectangle delineates the area of the lowest identity (3'-domain LE1+Ukd domain); ω – denotes a point of putative GPI-attachment, as predicted by Big-PI **(123)**. PSD-95 interaction site via the SH3-binding domain ((**69**), as determined for mice Netrin-G2) overlaps with the Loop III NGL-2 binding surface. Two stars indicate a modern human (T346A) and a hominin-specific (S371A/V) amino acid substitutions **(2)**. (D) Identical structural motif of the Netrin-G1/NGL1 and Netrin-G2/NGL2 complexes as per **(13)**. The figure's reproduction is covered by the Creative Commons license.