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Symmetric assembly and disassembly of an ecological network

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Abstract

The position of species in interaction networks can predict their extinction

probability. However, the processes whereby these network roles emerge and

persist, then decay during ecosystem development are unknown. Here we study

networks of plant and arbuscular mycorrhizal fungal (AMF) communities along a

120,000 yr soil chronosequence, with two distinct phases: progressive (i.e. assembly,

where plant richness and biomass increase) and retrogressive (i.e. disassembly,

where plant richness and biomass decline with declining nutrients). We find that the

order of interaction formation during progression mirrors that of interaction

extinction during retrogression, and that interactions in sites moving forwards or

backwards from the (12,000 yr) site of peak biomass were nested subsets of the

interactions at that site. Network assembly and disassembly were symmetrical, self-

reinforcing processes that together generated key attributes of network architecture.

Plant species that had high AMF-partner overlap with others in the community

(measured as 'closeness centrality') were best able to attract and retain new AMF

partners, and AMF species with high partner overlap were better able to retain their

interactions with plants. In contrast, interaction generalism ('node degree') per se

was a poor predictor of partner attraction during assembly or retention during

disassembly. These results could be used to predict extinction sequences based on

community assembly or network roles, identify focal points for invasions, and

suggest trajectories for ecosystem restoration.

Keywords: community assembly, succession, ecosystem development, retrogression,

mutualist network, mycorrhizal symbiosis, preferential attachment

Introduction

The arrangement of species interactions in ecological networks can be critical for the

stability and functioning of ecosystems (1-3). However the processes that generate

and maintain network structure during ecosystem development are unresolved. The

generally accepted hypothesis for complex network assembly is that new nodes (e.g.,

species) preferentially connect with those that are linked to many immediate

neighbors (4) (i.e. generalist nodes of high 'degree'). This 'preferential attachment'

was predicted (5), then shown empirically (6), to occur during the assembly of

pollination networks within a season. In contrast, a study of long-term changes to

pollination networks found that less-connected species preferentially attracted new

interactions (7), such that the importance of preferential attachment based on node

degree remains unclear for ecological networks. Although studies on the generation

of network architecture have largely focused on node attachment during network

assembly, the opposing process of preferential detachment can theoretically

produce the same architecture as preferential attachment (8, 9). This possibility is

strengthened by empirical findings that pollination networks show preferential loss

of less-connected species and their interactions (7, 10). Finally, it is possible that

network architecture may be generated by preferential attachment during

community assembly, and maintained by preferential detachment during

disassembly. This would imply that the same rules may operate during both the

assembly and disassembly of ecological networks (11), but this hypothesis remains

untested.

Here we study interaction network assembly and disassembly in a community of

plants and arbuscular mycorrhizal fungi (AMF)(12) throughout 120,000 years of

ecosystem development. Arbuscular mycorrhizal fungi are essential mutualists of

most terrestrial plants (13), and have typical within-site diversity comparable to

plant communities (14). Moreover, the high importance of species interactions for

determining AMF community composition (15, 16) demands a focus on interactions

(17) for these systems, within the context of soil nutrients which can also be

important drivers of community structure (18). First, we test whether network

assembly and disassembly processes are random or related to species' position in

the network. Second, we test the hypothesis (11) that attachment and detachment

are symmetrical, such that the process and order of disassembly mirror those during

assembly. Finally, we use simulations to determine how non-random assembly and

disassembly processes impact upon key aspects of network architecture.

The first requirement for an interaction to take place is an encounter, and if co-

occurrence rates between AMF propagules and plants were merely stochastic (19),

we would expect newly-arriving species to interact preferentially with the most

abundant species in the site. An established AMF species that has used its

associations with many plant species to develop an extensive mycelium network

could also be a beneficial partner, due to a reduction in initial construction costs and

immediate access to a large soil volume (20, 21). In this sense, the local abundance

of a species, along with the number of species with which it interacts, may partly

indicate its quality as a mutualist. If plants or AMF exhibit selectivity in their

interaction partners (22), we might therefore expect a general pattern of newly-

arriving species having a greater probability of interacting with generalist species, i.e.

traditional preferential attachment based on species degree (4, 6). This pattern

could also occur simply if generalists are least selective in the partners they accept.

Moreover, high AMF diversity can reduce competition among plant species and

provide insurance against variability in soil conditions, such that plant species that

interact with many AMF experience increased biomass (23). This greater biomass

could make AMF-generalist plants more attractive partners because they have the

capacity to provide more carbon across a wider range of conditions.

However, the benefits of interacting with generalists may depend in part on whether

they share their interaction partners with other species. Although AMF typically

interact with many plant species, there is growing evidence that their composition

(15) and fitness can differ across plant hosts (24). Host-plant selectivity may not

necessarily be species specific, but rather groups of AMF can associate with

ecological groups of plants (25). Closely-related AMF also tend to co-occur in a given

location (26), and closely-related organisms tend to share interaction partners (27).

We may therefore expect large interacting consortia of ecologically-similar, and in

some cases phylogenetically related, plants and fungi to form at a given successional

stage, with the dominant consortium potentially differing between early-

successional generalist species and later-successional forest species (14, 25). Thus, at

each stage of ecosystem development, newly-arriving fungi may have a higher

probability of interacting with plants in the dominant ecological group at that stage,

which will already interact with species from the dominant fungal group. These plant

species would not only (or even necessarily) be the most generalist individually, but

they would have high AMF-partner overlap with other plant species in the dominant

group at that stage (i.e. with the majority of species in the site). Species that have

high interaction-partner overlap with others in the network may also receive fitness

benefits from positive indirect effects (1), making them more reliable interaction

partners. We therefore hypothesise that the attractiveness of a species to new

arrivals may depend on its sharing of interaction partners with other species (see Fig.

S1 for our approach to measuring partner overlap).

To test these hypotheses, we used a dataset (15) on plant-AMF networks that were

sampled at different sites along a long-term (>120 kyr) chronosequence comprising

all stages of ecosystem development, including retrogression (28). Chronosequences

represent a powerful tool for understanding long-term co-ordinated changes

amongst species and their interactions, resource availability and ecosystem

processes (28). Strong gradients from initial N-limitation to eventual strong P-

limitation of ecosystem processes drive turnover of plant and AMF species (15, 29).

We treated ecosystem progression (during which plant species richness and biomass

increase (29)) as the assembly phase, and ecosystem retrogression (during which

strong P-limitation causes a loss of plant diversity and biomass (29)) as the network

disassembly phase. We defined attachment as the first ecosystem stage at which a

plant or AMF species was observed along the chronosequence, and detachment as

the final ecosystem stage in which a species was observed.

Results

In total our samples yielded 33 operational taxonomic units (OTUs, hereafter

referred to as species) of AMF and 53 species of plant, among which we observed

399 pairwise interactions (i.e. links, defined as the colonisation of a plant root by an

AMF species) (Fig. 1). There were an average of 16 AMF (SD = 3.33) and 10.8 (SD =

3.77) plant species, with 137.5 (SD = 24.14) interactions per site (Table S1). The AMF

community changes and turnover along the chronosequence have been described in

detail elsewhere (15).

We first tested whether preferential attachment or detachment occurs to existing

species that have many partners or high partner overlap. Newly arriving AMF species

were significantly more likely to interact with plant species that shared fungal

partners with many others (Fig. 2A, $P = 1.4 \times 10^{-8}$), but not those with many partners

(i.e. high 'degree'; $P = 0.401$) (Table S2). In contrast, plants showed no preferential

attachment based on partner overlap (Fig. 2B) or degree of AMF ($P > 0.4$ in both

cases, Table S2). However, preferential detachment occurred in both taxa, such that

plants and AMF were significantly less likely to be lost from the network during

ecosystem retrogression if they interacted with species that had high partner

overlap with others in the community ($P < 1.8 \times 10^{-5}$ in both cases; Fig. 2C,D; Table

S2).

As hypothesized previously (11), the disassembly process mirrored the assembly

process; the last interactions to form during assembly were the first to be lost during

ecosystem retrogression (Fig. 3, $P = 1.64 \times 10^{-9}$). Moreover, the interactions present

in sites moving forwards or backwards from the site of peak biomass were nested

subsets of those present at that site ($P < 0.001$ in both cases), such that interactions

progressively accumulated during progression but were sequentially lost during

retrogression. Thus, symmetrical assembly and disassembly processes, either of

which could theoretically generate complex network architecture (1, 4, 8, 9), both

operated to rapidly generate architecture that did not change significantly

throughout ecosystem development (SI Results, Table S3, Fig. S2).

Although degree and partner overlap were correlated in our networks (Fig. S3), we

found no evidence of preferential detachment of plants from AMF of low degree (P =

0.577), and AMF taxa even had *increased* detachment probability when they

associated with nodes of high degree (P = 0.040), as has been observed in a long-

term study of pollination networks (7), and this relation became stronger when

partner overlap was also included in the model (Table S2). Moreover, in all our

models, partner overlap predicted attachment or detachment probability better

than did equivalent models with only degree as the predictor, and its effect

remained qualitatively unchanged when degree was also included in the model

(Table S2, S4).

Our findings suggest that both attachment and detachment are key processes in

network development and, when combined, the loss or gain of links becomes self-

reinforcing (9). We simulated the processes observed here to explore the

consequences for network architecture (SI Materials and Methods) and found that

both preferential attachment and detachment processes were important for

determining and maintaining the nestedness of the network (Fig. S4), a key element

of ecological network architecture (1, 12) whereby specialists interact with species

that also interact with generalists. Preferential attachment (based on degree or

partner overlap) generated networks that were considerably more nested than

random, and preferential detachment maintained this nestedness in the face of

species extinctions (Fig. S4).

Discussion

Obviously species do not have information about the architecture of their interaction

networks, so partner overlap must be associated with ecological characteristics that

make species more likely to attract and retain interaction partners. There are several

potential, non-exclusive explanations for this attractiveness. The simplest would be if

abundant species shared many partners due to a high probability of random

encounters. In our dataset, there was a weak correlation between a species'

abundance and partner overlap within sites. Moreover, the core of interacting

generalists (i.e. the species involved in the majority of interactions in the network,

and whose partners overlapped considerably with the rest of the community) was

present throughout much of ecosystem development (see dark colors in top left of

matrices in Fig. 1), and this ubiquity likely made them a reliable target for newly

arriving species. However, causality cannot be inferred from these correlations in our

dataset, as attraction of diverse fungal mutualists could equally cause longer-term

persistence of plants (30). Most importantly, the relation between plant partner

overlap and AMF attachment and detachment probability remained significant after

controlling for abundance (SI Results, Table S4). In contrast, the effect of degree was

eliminated or even reversed when we controlled for abundance, and the preferential

detachment of plants from AMF species with low partner overlap became non-

significant when controlling for abundance (SI Results, Table S4).

Therefore, although abundance likely plays some role, the attractiveness of plants

with high partner overlap to AMF taxa reflects additional benefits to associating with

those species. Plants may benefit from the AMF communities generated by other

species (31), and plant-plant competition can be reduced (and plant biomass

increased) by high AMF diversity (23). Thus, consortia of plants that are able to share

diverse AMF communities with one another may grow better, and therefore have

the ability to provide more carbon to AMF symbionts. These groupings may also

reflect habitat preference of the species involved, with habitat generalist plant and

AMF species interacting tightly during early succession and forest specialists during

late succession (25), an hypothesis congruent with findings of partner specificity

occurring at the level of ecological groups (14).

The potential benefits of interacting with high-partner-overlap species may be

partially offset in newly-arriving plants because seedling performance can be

impaired by the AMF network of large heterospecific plants, which compete for

AMF-mediated resources and deplete the nutrient pool (32, 33). Also, the high

generality of AMF may mean that there is less variation across species in the extent

to which they share plant hosts (evidenced by the range of x-axis values for AMF

species being less than half that of plants Fig. 2A,B). Combined, these factors may

explain the non-significant effect of AMF degree and partner overlap on plant

attachment probability (Fig. 2B).

However, both plant and AMF species were less likely to be lost during retrogression

if they interacted with species that had high partner overlap with others in their

community (Fig. 2C,D). We hypothesized (based on the benefits described above)

that mutually beneficial interactions will involve species with high partner overlap,

and we observed that species that preferentially interacted with these core species

were significantly more likely to persist during retrogression. The decline in soil P

during ecosystem development likely makes plants increasingly dependent on AMF-

provided P. Likewise, declining plant photosynthetic rates along the chronosequence

(34) could increase competition among AMF for plant-derived carbon. Thus, the

need to develop and maintain mutualists and avoid symbiotic 'cheaters' will intensify

along the sequence, particularly during retrogression. Over time, both plants and

AMF can reduce their resource allocation to less-beneficial partners (22), particularly

when resources are scarce (35). The interactions that persist during retrogression

may therefore be those that are most mutually beneficial. Previous work has found

that species in late-successional plant-AMF networks tend to interact more

frequently with a subset of their total range of partners (36), and a viable strategy

may thus be to test a number of partners initially, then gradually restrict resource

allocation to a subset of these that provide the greatest benefit (35) as resources

become limiting. Congruent with this potential strategy, the most generalist plants

tended to lose many AMF partners during retrogression (AMF detachment

probability increased with host-plant degree). This process could easily occur at the

level of plant individuals, but be reflected as a loss of links at the species level.

Finally, if our findings are found to generalize to other ecological networks, they

would have several potential applications. First the symmetry of assembly and

disassembly processes (Fig. 3) throughout long-term ecosystem development

suggests that knowledge of assembly order for a given system could be used to

predict the interactions most at risk of extinction, such as in using network

information for conservation (37). Species reintroductions during restoration could

also follow the reverse order of extinction and replace extinct species with others

that fulfill the same network role, or focus on species that share interaction partners

with others, and will then attract interactions with any new species that colonize.

Similarly, non-native species should interact preferentially with high-partner-overlap

species in the network, which could therefore be a focus for biosecurity monitoring

to detect invasions early.

It is impossible to fully understand the architecture of complex networks in isolation

from the dynamic processes that generate and maintain them (4). We have shown

empirically that both preferential attachment and detachment underpinned the

development of an ecological network under changing abiotic conditions. These

symmetrical, self-reinforcing processes can generate and maintain important

features of interaction network architecture (1, 4, 8, 9), and thereby link species

colonization and extinction to emergent and potentially stabilizing (1, 2) ecosystem

properties.

Materials and Methods

Dataset

We generated networks of interactions between vascular plants and arbuscular

mycorrhizal fungi (AMF) using data from a study that examined AMF beta diversity

and the importance of soil age vs. plant host for explaining AMF community

structure (15). Sampling and sequencing methods can be found in that study or in

the SI Materials and Methods, but we summarise the key points here. Sampling was

conducted along the Franz Josef soil chronosequence, on the southern west coast of

the South Island, New Zealand. An important feature of the Franz Josef

chronosequence, which led us to select this site, is that strong soil nutrient gradients

(SI Materials and Methods) are associated with pronounced shifts in ecological

community composition, structure and function, such that ecosystem development

exhibits a clear progressive phase up to the 12,000 year old surface at which peak

tree biomass occurs (we treated this as the assembly phase), and a retrogressive

phase thereafter in which tree basal area (biomass) declines about three-fold (29,

38) (the disassembly phase). We sampled fungal communities on roots at ten

surfaces of the following ages (in years): <5, 15, 70, 290, 500, 1000, 5000, 12,000,

60,000, 120,000 (SI Materials and Methods; Table S1).

At each site we sampled 50 root fragments, which then underwent a molecular

analysis to identify the plant species, as well as any AMF OTUs present inside the

root (see (15) or SI Materials and Methods). We did not assign weights to these links,

as the hypotheses we test here relate to the initial formation of interactions, rather

than their strength or frequency once formed.

<u>Data</u> <u>analysis</u>

We hypothesized that species entering the network for the first time during the

progressive phase would associate non-randomly with other existing species based

on their position within the network (i.e. preferential attachment (4)). We also

hypothesized that the same would occur during the disassembly of the network,

such that the probability of species going locally extinct from the network would

depend on the network position of species with which they associate (i.e.

preferential detachment (8, 10)). We tested for preferential attachment during the

progressive phase (5 - 12,000 year-old) and preferential detachment during the

retrogressive phase (12,000 – 120,000 year-old), representing respectively the

assembly and disassembly of the network. We then examined the appearance

(attachment) of new species in the network during assembly, and disappearance

(detachment) of species during disassembly. A species was deemed to have attached

during the progressive phase when it first appeared in the chronosequence. During

the progressive phase, once a species had attached (i.e. a link was formed), it was

deemed to remain part of the network for the remainder of ecosystem progression.

This ensured that any species that appeared, disappeared then reappeared during

the progressive phase was not counted twice, which would give any species-specific

interaction preferences of these species a disproportionate weighting in analyses.

Finally, it prevented underestimation of degree or partner overlap due to imperfect

sampling of the network in any given stage. Similarly, detachment was deemed to

have occurred during the retrogressive phase when a species that had been present

during the progressive phase was absent from a given site and all remaining sites

along the sequence. Note that, because by definition attachment and detachment

processes must occur during the interval between two chronosequence stages (e.g.,

an interaction was present in site B that was not present in A), we treated the 12,000

year-old site (which represented the community peak in diversity and biomass) as

the 'peak' community, comprising the last stage of assembly and the first stage from

which disassembly occurred.

Preferential attachment and detachment

We analyzed preferential attachment and detachment processes using generalized

linear mixed effects models with binomial errors and the canonical logit link function,

conducted in the lme4 package (39) in R (40). We used separate models to test for

attachment processes during progression vs. detachment processes during

retrogression, and for plants vs. AMF. The response variable in each model was

binary, whereby each existing species in the network for each site was coded with a

value of 0 (a newly-arriving species did not attach to it, or a species did not detach

from it, in that site) or 1 (a new species attached to or detached from the species in

that site). Site (surface along the chronosequence) was included as a random factor,

such that preferential attachment was tested according to the relative network roles

of the various species within a site, rather than comparing species across sites, which

could be influenced by differences in the sizes of networks. The fixed predictor

variable was either degree or centrality (defined below).

To measure overlap in the use of interaction partners among species at a given

trophic level, we used closeness centrality (hereafter 'centrality', defined in Fig. S1)

in the unipartite projection of the bipartite interaction network (41). Centrality is

often correlated with degree (42), but carries additional information by measuring

the number of links from a focal species to all others in the network (not just

immediate neighbors), and has been used as a measure of species' importance (42)

or functional specialization (43) in mutualistic networks. In the unipartite projection,

species at a given trophic level are linked if they share an interaction partner (Fig. S1),

so high centrality indicates that a species shares many partners with other species

that also share partners with many others. In contrast, species with low centrality

interact with partners that are not used by the dominant consortium of tightly-

interacting plants and AMF. Although centrality may be implicitly used to infer the

nature of flows through a network (e.g. paths vs. walks, replication vs. spread), we

do not make any assumption about the nature of flows, if any, in the plant-AMF

network. Rather, we use closeness centrality simply as a measure of the extent to

which different species share interaction partners within a trophic level. Centrality

was calculated in the 'sna' package (44) in R, scaled within a given network (i.e.

within each site).

We tested whether attachment to a node during assembly, or detachment from a

node during disassembly, depended on that node's centrality (Fig. S1) or degree (the

number of partners with which the species interacts, normalized within networks of

a given site age). We used the centrality/degree of a node in the chronosequence

stage immediately prior to that in which the new node attached or an existing node

detached. This allowed us to compare the fit of models with centrality vs. degree as

the predictor using the Akaike Information Criterion (AIC). Thus, the predictor

variable in each model was either the degree or centrality of each existing species in

the site, to which newly-arriving species either attached vs. did not, or detached vs.

did not. For completeness, we also ran models with both of these predictors, to

determine if any potential collinearity qualitatively altered their separate effect.

None of our models showed any signs of overdispersion. We repeated these

analyses controlling for species abundances in case sampling effort generated any

spurious effects (see Controlling for the effect of species abundances in SI Materials

and Methods).

We omitted from this analysis any links representing new species attaching to other

newly-arriving species, because we could not ascribe a value of network position for

those species to use as the predictor variable. However, these pairings among two

newly-arriving species were rare, and the species involved typically also interacted

with other species already present in the network. Across all sites, 20 newly-arriving

plant species were associated with newly-arriving AMF, and of these, 16 species also

attached to AMF species that were already present in the network, for which

network position could be calculated. Similarly, of the 15 newly-arriving AMF species

that attached to newly-arriving plants, 8 also attached to existing species.

Arrival and extinction order

We tested whether the arrival and disappearance (extinction) order of links in the

ecological network through time occurred at random, or whether links that

appeared first during progression were last to be lost from the network during

retrogression. To test this, we analyzed the arrival and extinction order of each link

(i.e. each association between a mycorrhizal species and a plant species). This

analysis included both links of newly-arriving species (as in the preferential

attachment analysis above) and links that formed among existing species. Arrival

order was defined as the rank age of the site at which the interaction was first

observed; e.g. an interaction observed at the earliest (< 5 year-old) site was given a

value of 1, whereas an interaction first observed at the 1,000 year old site (6th along

the sequence) was given a value of 6. We treated the network at the site of peak

plant biomass (29) (12,000 years old) as the end point of the assembly phase, then

examined the order in which interactions were lost during the retrogressive phase

following this peak (i.e. the extinction order). Extinction order was defined, in the

same way as arrival order, as the first site following the peak biomass site in which

an interaction was not observed. For example, an interaction that was present at the

peak biomass site but not the latter two sites was given an extinction order of 1, an

interaction that was present at the peak biomass site and both of the remaining sites

was given an extinction order of 3, etc. For the analysis we used only those

interactions that were present in the peak biomass site, in order to be conservative;

had we simply used the arrival order and extinction order (last site in which an

interaction occurred) of all interactions across the sequence, this would have biased

us towards finding a significant result, because any interactions that only appeared

once along the sequence, or appeared early and were gone by the site of peak

biomass, would have generated an automatic correlation between their arrival and

extinction orders.

Thus, our analysis focused on the network of the peak biomass site and tested

whether the order in which interactions came into the network was correlated with

the order in which they were lost from the network. We tested for a relationship

between arrival and extinction order using a simple regression, with extinction order

as the response variable and arrival order as the predictor variable, and each

interaction (plant-mycorrhizal combination) was a replicate. There remained the

possibility that any observed relationship could be driven by interactions that

appeared only in the site of peak biomass, and hence would be last to arrive and first

to go extinct. To account for this, we re-ran the analysis after removing interactions

that were only present in the site of peak biomass.

A negative correlation between arrival and extinction order of interactions would

suggest a gradual accumulation of interactions in the network up until the site of

peak biomass, followed by a progressive loss of interactions. This would be expected

to generate a pattern whereby the peak biomass site has a set of interactions, of

which progressively older or younger sites have decreasing subsets. To test for such

a pattern, we generated two matrices in which we ranked the sites according to their

age (rows), either becoming progressively older or younger than the site of peak

biomass, which was in the top row. Cells of the matrices depicted the presence or

absence of each pairwise plant-AMF association (columns). We tested each matrix

for nestedness, using the nestedness metric based on overlap and decreasing fill

(NODF), with 999 permutations and a swap algorithm, using the oecosimu function

in the vegan (45) package for R (40).

<u>Simulations</u>

To explore the consequences of preferential attachment and detachment based on

centrality vs. degree, we conducted a set of simulations of network assembly and

disassembly scenarios (more detail provided in SI Materials and Methods). At each

time step, newly-arriving plant or AMF species would interact with one existing

species, with a probability proportional to the existing species' degree (i.e. the

Barabasi and Albert model (4)) or partner overlap (closeness centrality). We also ran

a third scenario whereby attachment was random (equally probable attachment to

any existing species), to provide a null point of comparison.

This assembly phase continued until networks contained 50 species of plant and 50

of AMF. Subsequently, each network was subjected to a disassembly phase, using

the same scenario (degree-based, centrality-based or random) as was used during

that network's assembly. Alternating plant and AMF species were removed (i.e. went

extinct) with a probability that was inversely proportional to the degree or closeness

centrality of the species with which they interacted, such that species that interacted

with species of high degree or high centrality were less likely to go extinct. As with

the assembly phase, we ran a scenario of the disassembly phase whereby extinction

was random.

Each scenario of the assembly and disassembly phases was run for 1000 replicates

(each comprising multiple time steps). At each time step within each replicate, after

species arrived or went extinct, we recorded the nestedness of the network using

NODF, calculated using the nestednodf function in the vegan package (45) for R.

Nestedness is a common feature of mutualistic networks (46), which has also been

observed recently in plant-mycorrhizal networks (12) and has been shown to

increase the persistence of networks (1, 2). We scaled the observed NODF by the

distribution of NODF values obtained by randomizing the adjacency matrix using a

null model (SI Materials and Methods).

References

1.   Bastolla U*, et al.* (2009) The architecture of mutualistic networks minimizes
competition and increases biodiversity. *Nature* 458(7241):1018-U1091.
2.   Thebault E & Fontaine C (2010) Stability of Ecological Communities and the
Architecture of Mutualistic and Trophic Networks. *Science* 329(5993):853-856.
3.   Thompson RM*, et al.* (2012) Food webs: reconciling the structure and
function of biodiversity. *Trends in ecology & evolution*.
4.   Barabási A-L & Albert R (1999) Emergence of scaling in random networks.
Science 286(5439):509-512.
5.   Jordano P, Bascompte J, & Olesen JM (2003) Invariant properties in
coevolutionary networks of plant-animal interactions. *Ecology letters* 6(1):69-
81.
6.   Olesen JM, Bascompte J, Elberling H, & Jordano P (2008) Temporal dynamics
in a pollination network. *Ecology* 89(6):1573-1582.
7.   Burkle LA, Marlin JC, & Knight TM (2013) Plant-pollinator interactions over
120 years: loss of species, co-occurrence, and function. *Science*
339(6127):1611-1615.
8.   Salathé M, May RM, & Bonhoeffer S (2005) The evolution of network
topology by selective removal. *Journal of the Royal Society Interface* 2(5):533-
536.
9.   König M, Tessone C, & Zenou Y (2012) Nestedness in Networks: A Theoretical
Model and Someapplications. *SIEPR Discussion Papers* 11(005):1-61.
10.   Aizen MA, Sabatino M, & Tylianakis JM (2012) Specialization and Rarity
Predict Nonrandom Loss of Interactions from Mutualist Networks. *Science*
335(6075):1486-1489.
11.   Bascompte J & Stouffer DB (2009) The assembly and disassembly of
ecological networks. *Philosophical Transactions of the Royal Society B-*
Biological Sciences 364(1524):1781-1787.
12.   Montesinos-Navarro A, Segarra-Moragues JG, Valiente-Banuet A, & Verdu M
(2012) The network structure of plant-arbuscular mycorrhizal fungi. *New*
Phytologist 194(2):536-547.
13.   Brundrett MC (2009) Mycorrhizal associations and other means of nutrition
of vascular plants: understanding the global diversity of host plants by
resolving conflicting information and developing reliable means of diagnosis.
Plant and Soil 320(1-2):37-77.
14.   Öpik M, Metsis M, Daniell T, Zobel M, & Moora M (2009) Large‐scale
parallel 454 sequencing reveals host ecological group specificity of arbuscular
mycorrhizal fungi in a boreonemoral forest. *New Phytologist* 184(2):424-437.
15.   Martínez‐García LB, Richardson SJ, Tylianakis JM, Peltzer DA, & Dickie IA
(2015) Host identity is a dominant driver of mycorrhizal fungal community
composition during ecosystem development. *New Phytologist* 205(4):1565-
1576.
16.   Vályi K, Rillig MC, & Hempel S (2015) Land‐use intensity and host plant
identity interactively shape communities of arbuscular mycorrhizal fungi in
roots of grassland plants. *New Phytologist* 205(4):1577-1586.

17.   Powell JR*, et al.* (2015) Deterministic processes vary during community
assembly for ecologically dissimilar taxa. *Nature communications* 6.
18.   Krüger M*, et al.* (2015) The rise and fall of arbuscular mycorrhizal fungal
diversity during ecosystem retrogression. *Mol. Ecol.* 24(19):4912-4930.
19.   Encinas‐Viso F, Alonso D, Klironomos JN, Etienne RS, & Chang ER (2015)
Plant‐mycorrhizal fungus co‐occurrence network lacks substantial structure.
Oikos.
20.   Newman EI (1988) Mycorrhizal links between plants: their functioning and
ecological significance. *Advances in Ecological Research* 18:243-270.
21.   Simard SW & Durall DM (2004) Mycorrhizal networks: a review of their extent,
function, and importance. *Canadian Journal of Botany* 82(8):1140-1165.
22.   Kiers ET*, et al.* (2011) Reciprocal rewards stabilize cooperation in the
mycorrhizal symbiosis. *science* 333(6044):880-882.
23.   Wagg C, Jansa J, Stadler M, Schmid B, & Van Der Heijden MGA (2011)
Mycorrhizal fungal identity and diversity relaxes plant-plant competition.
Ecology 92(6):1303-1313.
24.   Ehinger M, Koch AM, & Sanders IR (2009) Changes in arbuscular mycorrhizal
fungal phenotypes and genotypes in response to plant species identity and
phosphorus concentration. *New Phytologist* 184(2):412-423.
25.   Davison J, Öpik M, Daniell TJ, Moora M, & Zobel M (2011) Arbuscular
mycorrhizal fungal communities in plant roots are not random assemblages.
FEMS Microbiology Ecology 78(1):103-115.
26.   Horn S, Caruso T, Verbruggen E, Rillig MC, & Hempel S (2014) Arbuscular
mycorrhizal fungal communities are phylogenetically clustered at small scales.
The ISME journal 8(11):2231-2242.
27.   Gómez JM, Verdú M, & Perfectti F (2010) Ecological interactions are
evolutionarily conserved across the entire tree of life. *Nature* 465(7300):918-
921.
28.   Peltzer DA*, et al.* (2010) Understanding ecosystem retrogression. *Ecological
Monographs* 80(4):509-529.
29.   Richardson SJ, Peltzer DA, Allen RB, McGlone MS, & Parfitt RL (2004) Rapid
development of phosphorus limitation in temperate rainforest along the
Franz Josef soil chronosequence. *Oecologia* 139(2):267-276.
30.   Scheublin TR, Van Logtestijn RSP, & Van Der Heijden MGA (2007) Presence
and identity of arbuscular mycorrhizal fungi influence competitive
interactions between plant species. *Journal of Ecology* 95(4):631-638.
31.   Bever JD (2002) Negative feedback within a mutualism: host–specific growth
of mycorrhizal fungi reduces plant benefit. *Proceedings of the Royal Society of
London B: Biological Sciences* 269(1509):2595-2601.
32.   Kytöviita MM, Vestberg M, & Tuomi J (2003) A test of mutual aid in common
mycorrhizal networks: established vegetation negates benefit in seedlings.
Ecology 84(4):898-906.
33.   Janoušková M, Rydlová J, Püschel D, Száková J, & Vosátka M (2011)
Extraradical mycelium of arbuscular mycorrhizal fungi radiating from large
plants depresses the growth of nearby seedlings in a nutrient deficient
substrate. *Mycorrhiza* 21(7):641-650.

34. Whitehead D*, et al.* (2005) Photosynthesis and reflectance indices for
rainforest species in ecosystems undergoing progression and retrogression
along a soil fertility chronosequence in New Zealand. *Oecologia* 144(2):233-
244.
35. Bever JD (2015) Preferential allocation, physio‐evolutionary feedbacks, and
the stability and environmental patterns of mutualism between plants and
their root symbionts. *New Phytologist* 205(4):1503-1514.
36. Bennett AE*, et al.* (2013) Arbuscular mycorrhizal fungal networks vary
throughout the growing season and between successional stages. *PloS one*
8(12):e83241.
37. Tylianakis JM, Laliberté E, Nielsen A, & Bascompte J (2010) Conservation of
species interaction networks. *Biological Conservation* 143:2270-2279.
38. Wardle D, Bardgett R, Walker L, Peltzer D, & Lagerström A (2008) The
response of plant diversity to ecosystem retrogression: evidence from
contrasting long-term chronosequences. *Oikos* 117:93 -103.
39. Bates D, Maechler M, Bolker B, & Walker S (2014) lme4: Linear mixed-effects
models using Eigen and S4), R package version 1.0-6.
40. Team RC (2013) R: A language and environment for statistical computing. (R
Foundation for Statistical Computing, Vienna Austria), 3.0.1.
41. Gómez JM & Perfectti F (2012) Fitness consequences of centrality in
mutualistic individual-based networks. *Proceedings of the Royal Society B:*
Biological Sciences 279(1734):1754-1760.
42. Martín González AM, Dalsgaard B, & Olesen JM (2010) Centrality measures
and the importance of generalist species in pollination networks. *Ecological*
Complexity 7(1):36-43.
43. Dalsgaard B*, et al.* (2008) Pollination networks and functional specialization: a
test using Lesser Antillean plant–hummingbird assemblages. *Oikos*
117(5):789-793.
44. Butts CT (2014) sna: Tools for Social Network Analysis.), R package version
2.3-2.
45. Oksanen J*, et al.* (2008) vegan: Community Ecology Package ([http://cran.r-](http://cran.r-project.org/)
[project.org/)](http://cran.r-project.org/).
46. Bascompte J, Jordano P, Melian CJ, & Olesen JM (2003) The nested assembly
of plant-animal mutualistic networks. *Proceedings of the National Academy*
of Sciences of the United States of America 100(16):9383-9387.
47. Stevens PR (1968) A chronosequence of soils near the Franz Josef Glacier.
PhD thesis (University of Canterbury, New Zealand).
48. Walker T & Syers J (1976) The fate of phosphorus during pedogenesis.
Geoderma 15(1):1-19.
49. Allison V, Condron L, Peltzer D, Richardson S, & Turner B (2007) Changes in
enzyme activities and soil microbial community composition along carbon
and nutrient gradients at the Franz Josef chronosequence, New Zealand. *Soil*
Biology and Biochemistry 39(7):1770-1781.
50. Jangid K, Whitman WB, Condron LM, Turner BL, & Williams MA (2013)
Progressive and retrogressive ecosystem development coincide with soil
bacterial community change in a dune system under lowland temperate
rainforest in New Zealand. *Plant and soil* 367(1-2):235-247.

51. Dickie IA*, et al.* (2013) Mycorrhizas and mycorrhizal fungal communities
throughout ecosystem development. *Plant and Soil* 367(1-2):11-39.
52. Lambers H, Raven JA, Shaver GR, & Smith SE (2008) Plant nutrient-acquisition
strategies change with soil age. *Trends in Ecology & Evolution* 23(2):95-103.
53. Turner S*, et al.* (2014) Mineralogical impact on long-term patterns of soil
nitrogen and phosphorus enzyme activities. *Soil Biology and Biochemistry*
68:31-43.
54. Richardson SJ, Peltzer DA, Allen RB, & McGlone MS (2010) Declining soil
fertility does not increase leaf lifespan within species: evidence from the
Franz Josef chronosequence, New Zealand. *New Zealand Journal of Ecology*
34(3):306-310.
55. Holdaway RJ, Richardson SJ, Dickie IA, Peltzer DA, & Coomes DA (2011)
Species- and community-level patterns in fine root traits along a 120 000-
year soil chronosequence in temperate rain forest. *Journal of Ecology*
99(4):954-963.
56. Wardle DA, Walker LR, & Bardgett RD (2004) Ecosystem properties and forest
decline in contrasting long-term chronosequences. *Science* 305:509-513.
57. Doblas-Miranda E, Wardle DA, Peltzer DA, & Yeates GW (2008) Changes in
the community structure and diversity of soil invertebrates across the Franz
Josef Glacier chronosequence. *Soil Biology and Biochemistry* 40(5):1069-1081.
58. Wardle P (1991) *Vegetation of New Zealand* (CUP Archive).
59. Öpik M & Moora M (2012) Missing nodes and links in mycorrhizal networks.
New Phytologist 194(2):304-306.
60. Simon L, Lalonde M, & Bruns T (1992) Specific amplification of 18S fungal
ribosomal genes from vesicular-arbuscular endomycorrhizal fungi colonizing
roots. *Applied and Environmental Microbiology* 58(1):291-295.
61. Lee J, Lee S, & Young JPW (2008) Improved PCR primers for the detection and
identification of arbuscular mycorrhizal fungi. *FEMS Microbiology Ecology*
65(2):339-349.
62. Dickie I & FitzJohn R (2007) Using terminal restriction fragment length
polymorphism (T-RFLP) to identify mycorrhizal fungi: a methods review.
Mycorrhiza 17(4):259-270.
63. van Dorst J*, et al.* (2014) Community fingerprinting in a sequencing world.
FEMS microbiology ecology 89(2):316-330.
64. White TJ, Bruns T, Lee S, & Taylor J (1990) Amplification and direct
sequencing of fungal ribosomal RNA genes for phylogenetics. *PCR protocols:
a guide to methods and applications* 18:315-322.
65. Taberlet P, Gielly L, Pautou G, & Bouvet J (1991) Universal primers for
amplification of three non-coding regions of chloroplast DNA. *Plant molecular
biology* 17(5):1105-1109.
66. Trewick S*, et al.* (2002) Polyploidy, phylogeography and Pleistocene refugia of
the rockfern Asplenium ceterach: evidence from chloroplast DNA. *Mol. Ecol.*
11(10):2003-2012.
67. Wirta HK*, et al.* (2014) Complementary molecular information changes our
perception of food web structure. *Proceedings of the National Academy of
Sciences* 111(5):1885-1890.

68. Toju H, Guimarães PR, Olesen JM, & Thompson JN (2014) Assembly of
complex plant–fungus networks. *Nature communications* 5.
69. Caruso T, Rillig MC, & Garlaschelli D (2012) On the application of network
theory to arbuscular mycorrhizal fungi–plant interactions: the importance of
basic assumptions. *New Phytologist* 194(4):891-894.
70. Graham J & Eissenstat D (1994) Host genotype and the formation and
function of VA mycorrhizae. *Plant and Soil* 159(1):179-185.
71. Nguyen NH, Smith D, Peay K, & Kennedy P (2014) Parsing ecological signal
from noise in next generation amplicon sequencing. *New Phytologist*.
72. Hurst J & Allen R (2007) *A permanent plot method for monitoring indigenous
forests – Field protocols* (Landcare Research, Lincoln, New Zealand),
(Research L).
73. Albrecht M, Riesen M, & Schmid B (2010) Plant-pollinator network assembly
along the chronosequence of a glacier foreland. *Oikos* 119(10):1610-1624.
74. Fahimipour AK & Hein AM (2014) The dynamics of assembling food webs.
Ecology Letters 17(5):606-613.
75. Kuznetsova A, Brockhoff P, & Christensen R (2013) lmerTest: Tests for
random and fixed effects for linear mixed effect models (lmer objects of lme4
package). *R package version*:2.0-0.
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Author contributions

J.M.T. and I.A.D. designed the study, in discussion with all other authors. L.B.M.-G.

and I.A.D. conducted molecular analyses. J.M.T. conducted statistical analyses. S.J.R.

and D.A.P. collected plant abundance data. All authors contributed to field sampling

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Author information

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Figure 1: Plant-mycorrhizal association matrices during assembly (N = 8 sites) and

disassembly (i.e. retrogressive, N = 3 sites) phases of ecosystem development.

Darker colours indicate that the plant-mycorrhizal association was present in a

greater number of sites (i.e. it formed early during assembly or persisted during

retrogression). The network is significantly nested (see SI Results), whereby

specialists interact with proper subsets of the species that interact with generalists.

Figure 2: Probability (left vertical axis) that a new A,C) arbuscular mycorrhizal fungus

(AMF), or B,D) plant entering the network will interact with an existing species

during assembly (top), or detach from a species during disassembly (bottom) as a

function of that species interaction-partner overlap with others of its trophic level

(measured as closeness centrality of the unipartite projection, see Fig. S1).

Histograms top and bottom of each graph represent the frequencies (right vertical

axis) of ones and zeroes respectively in the raw data. Trend lines are based on

inverse-linked coefficients of a binomial linear mixed effects model with

chronosequence stage (plant-mycorrhizal network) or year (arms trade network) as a

random effect. Solid lines were statistically significant (P < 0.005 in all cases), dashed

line was not significant at alpha = 0.05.

Figure 3: The relationship between the order in which plant-AMF interactions

(network 'links') were formed (arrival order) and that in which they were lost

(extinction order). Larger circles indicate more overlapping data points. There was a

significant negative association between the formation and disappearance order of

interactions ($F_{1,269}$ = 39, P < 0.0001), such that the last interactions to form during

network assembly were the first to be lost during disassembly. This analysis only

included interactions that were present at the site of peak biomass (arrival order 8),

so was not confounded by interactions that appeared in only one site (i.e. arrived

and went 'extinct' instantly). The relationship was also not simply driven by

interactions that appeared only in the site of peak biomass, as it remained significant

even when these data points were removed ($F_{1,245}$ = 5.38, P < 0.022).

 SI Materials and Methods

 Study site

 We sampled interactions between vascular plants and arbuscular mycorrhizal fungi

 (AMF) along the Franz Josef soil chronosequence, on the southern west coast of the

 South Island, New Zealand. The sites are schist outwash surfaces that have been

 exposed during glacial retreat, and span a range of ages (since exposure) from more

 than 120,000 years to the present. An important feature of the Franz Josef

 chronosequence, which led us to select this site, is that strong soil nutrient

 gradients) are associated with pronounced shifts in ecological community

 composition, structure and function, such that ecosystem development exhibits a

 clear progressive phase up to the 12,000 year old surface at which peak tree biomass

 occurs (we treated this as the assembly phase), and a retrogressive phase thereafter

 in which tree basal area (biomass) declines about three-fold (29, 38) (the

 disassembly phase). We sampled fungal communities on roots at ten surfaces of the

 following ages (in years): <5, 15, 70, 290, 500, 1000, 5000, 12,000, 60,000, 120,000

 (Table S1).

 The sampling sites occur between the terminus of the Franz Josef glacier (43.45° S,

 170.17° E) and the coast of central western South Island (43.25° S,170.19° E) (29, 47).

 The current climate is highly oceanic and mean annual temperature (1926–1975) at

 the valley mouth is 10.8ºC . Precipitation is ca. 6.5m for first seven sites of the

 sequence, which are within the glacier valley, and ca. 3.5m for the last three sites

 that lie between the valley mouth and the coast. Precipitation falls evenly

 throughout the year.

 Inorganic phosphorus (P) is initially high (> 800 mg/kg), but declines to ca. 100 mg/kg

 by the oldest stage (29). Declining phosphorus is associated with a shift from plant-

 available mineral forms to organic forms. Mineral soil nitrogen and carbon

 concentrations are initially negligible (<0.1%) but increase rapidly during the first 100

 years of ecosystem development in association with nitrogen-fixing species and

 accumulating tree biomass; both reach peak concentrations at 500 years and decline

 slowly thereafter (29, 38).

We sampled fungal communities on roots at ten surfaces of the following ages (in

years): <5, 15, 70, 290, 500, 1000, 5000, 12,000, 60,000, 120,000. These sites are

based on those used by Richardson et al. (29), with three differences: the addition of

a <5-year old surface at the glacier forefront and a 1000-year old site, and a 12,000

year old site that was used in the original description (48) of the chronosequence, in

lieu of Richardson et al's (29) 12 kyr. site. Our sites ranged from initial post-glacial

primary succession (gravel bed with sparse grasses and subshrubs, highly nitrogen

limited with abundant phosphorus) through the assembly of a temperate rainforest

(soils ca. 12,000 years old with maximal aboveground biomass limited primarily by P)

and ultimately retrogressive phases (the two sites ≥60 kyr old having severe nutrient

depletion through pedogenic processes) (29).

Forests along the chronosequence, and in the region generally, are formed by

associations of broadleaved evergreen angiosperm and long-lived coniferous

(arbuscular mycorrhizal Podocarpaceae) tree species with abundant tree ferns,

lianas and ground ferns (29). Endemic evergreen angiosperms dominate the

youngest sites (from 5 to 290 yr age), and thereafter are replaced by major canopy

dominant tree species, primarily Podocarpaceae; these dominate the plant

community throughout the remainder of the chronosequence. Tree ferns occur

across all the sites except the oldest (120,000 yr) (29); grasses and ericaceous shrubs

are almost absent from the chronosequence, but do form a minor component of the

vegetation on the 15 yr and oldest sites respectively.

Strong shifts also occur in belowground properties and biota along this sequence,

mirroring the co-ordinated shifts in nutrients, community composition and ecological

processes throughout ecosystem development. Soil microbial communities decline

in diversity and richness along the sequence, and compositionally shift from

bacterial-dominated to fungal-dominated (49). In addition, some bacterial phyla are

common throughout the sequence (e.g., Actinomycetes and alpha-Proteobacteria),

whereas other phyla decline over time (e.g., Bacilli, beta-Proteobacteria, and

Bacteriodetes (50)). Similarly, mycorrhizal fungi are dominated by arbuscular species

throughout. Orchid mycorrhizas are present throughout the sequence, but are

extremely rare; ericoid mycorrhizas are present primarily on the oldest, retrogressive

stage (51, 52) (although the ericaceous shrub *Gaultheria* is also present at the 15

year old site); and a plant species capable of forming ectomycorrhizas

(*Leptospermum scoparium*) is present at the oldest site, but no ectomycorrhizal

roots have been observed. In addition, soil microbes and fungi are an increasingly

important pool of P whereby the majority of available (non-occluded) P is retained in

the soil microbial biomass during ecosystem retrogression (53). Activities of the soil

microbial biomass are coordinated such that investment in phosphorous acquisition

increases along the sequence, as demonstrated by a four-fold increase in the major

enzymes used to process phosphorus (phosphomonoesterase and phophodiesterase

(49, 53)).

Plant functional leaf and root traits show a co-ordinated shift predictably towards an

increasingly conservative resource-use strategy with increasing surface age, both

within widespread species and through species turnover (29, 54, 55). The sites in the

early (progressive) phase represent the stages typical of ecosystem succession,

whereas the two oldest sites are considered retrogressive (28, 29, 56) due to soil

nutrient limitation.

Changes in the diversity, abundance and composition of litter and soil-dwelling

invertebrates are less well understood. The abundance of microbial-feeding and

omnivorous nematodes and copepods show unimodal responses along the sequence

in soil, but more complex responses in the litter layer (57). Richness and diversity of

both nematodes and macroinvertebrates differ significantly among stages or sites,

but do not shift consistently throughout ecosystem development. Acari abundance is

biomodal, peaking in early and mid stages of the sequence, whereas diversity

increases throughout. In contrast, Collembola abundance and diversity are both

bimodal, peaking at the earliest and maximal biomass stages. These results suggest

that soil microbial community composition and activity are coordinated along the

sequence, but that other soil biota, such as invertebrates, have more complex

responses; these are likely driven by the relative strength and importance of top-

down vs. bottom-up regulation of these taxa (56), but these processes have not yet

been investigated.

Sampling

At each site we sampled 50 root fragments of 10 to 20 cm length from the organic or

mineral layer at a depth of 10-30 cm, where the majority of fine roots occur (p.43 in

(58)), with each sample spaced 2 m apart along two parallel 50 m transects. This

depth allowed consistent sampling across all sites (all sites had roots at this depth)

and high replication with minimal site disturbance, but we note that the root and

fungal community below 30 cm was not included. Thus, the frequency with which

plant species appeared in our samples should reflect their relative abundance (or at

least belowground biomass from 10-30 cm) in the site (59), congruent with transect

samples frequently used to sample pollination or host-parasitoid networks (6, 10).

Although species density per unit space (and therefore sampling probability of each

species) may change along the sequence, our analyses do not compare sites with

each other. Rather, we make comparisons within sites of attachment and

detachment probability based on the network role of species. We took the same

number of samples from each depth (10, 20 or 30 cm) at each site, randomly

positioned along each transect, to ensure coverage of different root zones. Roots

were thoroughly washed with tap water to remove any attached soil, then freeze-

dried overnight. Visual inspection suggested that tap water washing was effective at

removing all soil. Additionally, at each site, leaves from the most representative

plants were identified, collected and frozen for molecular analysis. During this

process both root and leaf samples were kept cold (5°C). Root samples then

underwent a molecular analysis to identify the plant species, as well as any AMF

OTUs present inside the root (see Justification of our Sequencing Approach below).

We did not assign weights to the links, as the hypotheses we test here relate to the

initial formation of interactions, rather than their strength or frequency once formed.

Molecular analysis

Arbuscular mycorrhizal fungal (Glomeromycota) DNA was amplified using the

universal eukaryotic primer NS31 (60) with the fluorochrome label VIC and the

arbuscular mycorrhizal fungal specific primer AML2 (61) labeled with 6FAM, cleaned,

and analyzed using terminal restriction fragment length polymorphism (T-RFLP (62)).

Network analysis of plant-fungal interactions across gradients requires a high level of

replication of small individual samples (< 0.2 mg root) within sites. As an effective

way to achieve this within limited budgets we used T-RFLP on individual roots (62).

At the time the samples were run it was not possible to achieve cost-effect next-

generation sequencing of highly replicated samples (n > 500), however we did pool

samples for next-generation sequencing (454-pyrosequencing) at the level of site (n

= 10). This comparison showed that the T-RFLP results captured similar patterns in

community composition and beta-diversity as next-generation sequencing (15), and

recent work suggests that these approaches can be as effective as deep sequencing

for detecting ecological patterns in microbial communities (63).

A subsample of 0.15 mg of each root was ground in a bead beater, then extracted

with Power Soil DNA Isolation Kit (Mo-Bio Labs, Carlsbad, CA, USA). Polymerase

chain reactions (PCRs) were carried out in a final volume of 20 µl, with 1µl of DNA

extract, 0.4µM of each primer, 0.2mM dNTP's, 2mM $MgCl_2$, 0.2 mg/ml BSA (New

England BioLabs, Ipswich, MA, USA) and 0.05U/µl Fast Taq DNA polymerase (Roche

Diagnostics N.Z., Ltd., Auckland, New Zealand). Thermal cycling for the PCRs started

with an initial denaturing step at 94°C for 3 min, followed by 35 cycles of 30 seconds

at 94°C, 1 min at 58°C and 1 min at 72°C, and ended with a final extension step of

72°C for 10 min. Five per cent of the PCRs did not amplify. For those samples, a

nested PCR was used combining a first PCR with NS1 and NS4 (universal fungal

primers (64)) as above, except that primer concentration was 0.1 µM and annealing

temperature 40°C, followed by NS31-VIC and AML2-6FAM. PCR products were

purified using DNA Clean & Concentrator Kit (Zymo Research Corporation, Irvine, CA,

USA).

T-RFLP restriction enzymes were chosen by running virtual digestions of 189

commercial restriction enzymes on the 10 most abundant sequences of a

pyrosequencing library generated from the pooled PCR products. Using virtual

digestions of 189 commercial restriction enzymes run on the 10 most abundant

arbuscular mycorrhizal sequences resulting from pyrosequencing, we selected the

enzymes HinfI (Life Technologies New Zealand Ltd., Auckland, New Zealand) and MslI

(New England BioLab) as having the greatest ability to distinguish molecular

Operational Taxonomic Units (OTUs). Clean PCR products were digested for 6 h and

1 h respectively and denatured using HiDi formamide mixed with MapMarker 1000

ROX standard (BioVentures, Murfreesboro TN, USA). Terminal restriction fragments

(TRFs) were separated by capillary electrophoresis using a Prism 3100 Genetic

Analyzer (Applied Biosystems) at the University of Canterbury Sequencing Centre

(Christchurch, New Zealand).

The TRAMPR package (62) for R (40) was used to process TRFLP data. Peaks with a

height ≤20% the height of the largest peak were ignored as background noise. The

build.knowns function from the TRAMPR package (with default settings) was used to

define TRFLP-OTUs by the highest peak from each restriction enzyme direction

(forward and reverse). TRFLP gave a total of 34 OTUs. For brevity, we henceforth

refer to fungal OTUs as "species".

Plant amplification and sequencing

Plant DNA from leaf and root samples was extracted using the Intron Plant DNA

extraction kit (Intron Biotechnology, Gyeonggi-Do, Korea) and amplified using the

chloroplast trnL (UAA) gene primers c and d (65). These primers failed to amplify

DNA from fern species, hence specific fern primers trnL- f and trn- Fern (66) were

used to amplify the DNA in the root samples for which PCR failed using trnL-c-d

primers. Amplicons from both leaf and root samples were purified using ExoI and

Sap enzyme (Thermo Fisher Scientific, Auckland New Zealand) and sequenced at the

University of Canterbury Sequencing Centre (Christchurch, New Zealand). Plant

species were identified based on BLAST matches to GenBank at 98% similarity,

including sequences from our vouchered plant leaf collections (GenBank KF591217 -

KF591315; KF591316 - KF591341).

Justification of sequencing approach

DNA-based methods not only provide an efficient means of data collection, but they

also allow the identification of cryptic taxa or those that are difficult to culture, and

are thus being increasingly used to document interactions among species for

network studies (67, 68). By sampling root tissue, we isolated fungi that were

symbiotic with the plant, but to be conservative we avoid assigning any direction to

the interactions, as they may be mutualistic, commensalistic or antagonistic,

depending on the taxa involved or their environmental context (68-70). This is

analogous to other network types where, for example, the visitation of flowers does

not necessarily imply successful pollination and fruit feeding does not imply

successful seed dispersal. We used the colonization of plant species by AMF species

at each site to generate community plant-AMF interaction networks, as in previous

studies (12, 36, 68).

Controlling for the effect of species abundances in attachment and detachment

analyses

Abundant species have a greater probability of being observed engaging in

interactions with others, which could make them more connected and/or central in

the interaction network. Likewise, if newly-arriving species interact with others at

random, they have a greater likelihood of interacting with abundant species by

chance. In this way, the abundance of species could drive both their network

position and probability of being attached to or detached from. Although the

processes of preferential attachment and detachment would still be useful to

understand, even if they were driven by abundances, we tested explicitly whether

centrality and degree affected attachment and detachment probability, after

controlling for differences in the abundance of species.

We first tested whether the abundance of species determined their network position

(centrality and/or degree), then we tested whether the preferential attachment

results changed after controlling for species abundances. To test these possibilities,

it is necessary to have an independent measure of species abundances from the site.

Defining and measuring the abundance of individual mycorrhizal fungal taxa in the

field presents numerous difficulties (71). Nevertheless, to account for their sampling

effort (rather than their 'abundance' as it would be perceived by a plant), we used

their frequency in our samples (i.e. the number of plant root samples in which they

were detected). However, plant abundances can be measured easily aboveground,

and aboveground biomass/percentage cover has been shown to correlate with

belowground biomass, which would be more important for belowground

interactions.

All vascular plant species rooted within each plot were identified to species, and

percentage cover measured using a relevé plot method. Briefly, cover was assessed

visually as cover classes (<1%, 1–5%, 6–25%, 26–50%, 51–75%, 76–95% and >95%)

within each of four height tiers (0–0.3 m, 0.3–2 m, 2–5 m, 5–12 m) following

standard vegetation survey protocols (72). The midpoint for each cover class used to

calculate percentage cover. We first tested whether the percentage cover of a plant

species was a good predictor of its network centrality. We used a linear mixed

effects model with closeness centrality as the response variable and percent cover as

the predictor, with site age (chronosequence stage) as a random effect. We then ran

an analogous model to test for an effect on plant degree. Because we did not have

an independent measure of fungal abundance, we used their frequency in our

samples (i.e. the number of plant root samples in which they were detected) as a

surrogate for their abundance.

We first tested whether the percentage cover of a plant species was a good

predictor of its network centrality. We used a linear mixed effects model with

closeness centrality as the response variable and percent cover as the predictor, with

site age (chronosequence stage) as a random effect. We then ran an analogous

model to test for an effect on plant degree. We also re-ran our earlier models for

preferential attachment including abundance of each species as a covariate.

Simulations of preferential attachment and detachment

Our finding that preferential attachment was explained better by centrality than by

degree raises the question of how these differing non-random processes affect the

final outcome of the network produced. The overall plant-AMF network architecture

was significantly nested (P = 0.009, measured using NODF), as observed in previous

plant-mycorrhizal networks (12). We therefore assessed the extent to which

(linear) preferential attachment and detachment, based on centrality vs. degree,

would affect the nestedness of networks. To do this, we conducted a set of

simulations of network assembly and disassembly scenarios. In each replicate of the

assembly process, we began with a network comprising four species (two plants and

two arbuscular mycorrhizal fungi, AMF), with three interactions among them. One

species of plant and one AMF each had two interactions, and the remaining two

species had one interaction (this could be visualized as an upside-down 'L' in the

adjacency matrix). Note that if we began with a smaller matrix (e.g. only one

interaction between two species), a newly-arriving AMF species would have no

option but to interact with the single plant species. Then a newly-arriving plant

would choose between the two fungi with equal probability (they would have the

same degree and centrality), and this would inevitably produce the matrix with

which we began.

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From this starting network, we simulated the arrival of new AMF and plant species

(alternating between the trophic levels) through time, until the network contained

50 plants and 50 fungi. At each time step, the newly-arriving species would interact

with one existing species, with a probability proportional to the existing species'

degree (i.e. the Barabasi and Albert model (4)) or closeness centrality. We also ran a

third scenario whereby attachment was random (equally probable attachment to

any existing species), to provide a null point of comparison. In the random

attachment scenario, the lifetime probability of receiving new attachments (i.e.

'fitness' in network, but not Darwinian terminology) was proportionate to the arrival

order of species. Each newly-arriving species initially only interacted with one

existing species, but it could also potentially interact with other species that arrived

later.

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After this assembly phase, each network was subjected to a disassembly phase,

using the same scenario (degree-based, centrality-based or random) as was used

during that network's assembly. Alternating plant and AMF species were removed

(i.e. went extinct) with a probability that was inversely proportional to the degree or

closeness centrality of the species with which they interacted, such that species that

interacted with species of high degree or high centrality were less likely to go extinct.

If a species interacted with more than one other species, its detachment probability

was determined by the mean degree or centrality of those species. If, after a species

went extinct, other species were left unconnected (i.e. their only interaction partner

went extinct), these unconnected species were also deemed to have gone

secondarily extinct. As with the assembly phase, we ran a scenario of the

disassembly phase whereby extinction was random. The simulated extinction of

plant or AMF species proceeded until there remained only two species at that

trophic level or until the network became too small to accurately calculate centrality.

If only one trophic level met these criteria (e.g. if there remained only two plant

species, but more than two AMF species), then the other trophic level continued to

suffer extinctions until it also met the criteria (in Fig. S4, this results in primary

extinctions that begin as multiples of two then eventually include single extinctions).

Consequently, the number of primary extinctions during disassembly did not

consistently increase by two during every time step, unlike the assembly phase,

where two species attached in each time step.

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Each scenario of the assembly and disassembly phases was run for 1000 replicates

(each comprising multiple time steps). At each time step within each replicate, after

species arrived or went extinct, we recorded the nestedness of the network.

Nestedness is a network pattern whereby specialist species interact with a subset of

the species that interact with generalists. It is a common feature of mutualistic

networks (46), which has also been observed recently in plant-mycorrhizal networks

(12) and has been shown to increase the persistence of networks (1, 2).

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We calculated the nestedness of the network using NODF, calculated using the

nestednodf function in the vegan package (45) for R. Because this measure could be

influenced by network size, we scaled the observed NODF by the distribution of

NODF values obtained by randomizing the adjacency matrix using the null model

("Null model II") of Bascompte et al. (46). For each network at each time step, we

ran 99 iterations of the null model, each time calculating NODF. We subtracted the

mean NODF value of the resulting null distribution from the observed NODF of the

network, and divided the result by the standard deviation of the null distribution.

This provided a standardized measure of the extent to which the nestedness of the

network at each time step differed from null expectations for a network of that size.

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SI Results

There was no change in network structure along the chronosequence

Previous work has shown that some attributes of network structure can be rapidly detected even with small samples from networks (37). This suggests that overall network architecture, such as interaction nestedness, could emerge quickly during ecosystem development (i.e. even when the network is small), provided that preferential attachment and detachment processes occur from the outset of network formation.

We therefore analyzed how nestedness and other metrics of network structure changed along the chronosequence using general linear models (or Poisson generalized linear models in the case of richness) with the metric as the response variable, and log site age as the predictor. As metrics to describe network structure, we tested two measures of nestedness (nestedness temperature and NODF), connectance, both quantitative and binary versions of linkage density, quantitative interaction evenness, and species richness of plants and mycorrhizas. To allow for the possibility that the relationship between network metrics and site age was non-linear, we included a quadratic term for site age. We also included sample size (the number of root samples where both the plant species and at least one fungal species were identified) as a covariate to account for any potential sampling biases. We then simplified each of these models by removing variables and minimizing the Akaike Information Criterion (AIC).

In our networks, nestedness did not change significantly across the chronosequence (Table S3), as would be expected if the strength of preferential attachment was consistent across the chronosequence. In fact, no measure of structure changed with ecosystem development (Table S3, Fig. S2), congruent with previous studies of the assembly of pollination network and food web structures (6, 73, 74). In contrast, a comparison of early and late successional forests found two (connectance and specialization) out of six measures of plant-AMF network structure changed (36), though these changes could have been caused by changes in the diversity of potential interaction partners. Despite the macroscopically static structure we

1094   observed, there was high turnover of species and interactions, with new species

1095   joining the network and others no longer occurring at each stage along the sequence

1096   (Table S1).

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Effect of species abundances on results

The effect of plant percent cover on centrality was significant (t = 3.24, P = 0.002),

though the effect size was small (slope = 0.000217). For example, if a species'

percent cover changed by 11% (which would be equivalent to changing from the

lower quartile to the upper quartile), its fitted value for centrality would increase

only by 0.002 (which is an order of magnitude less than one standard deviation for

centrality, s.d. = 0.029). We note, however, that plant percent cover significantly

predicted plant normalized degree (t = 6.55, P = 2.46 x 10^{-9}), with a larger effect size

(slope for the effect of percent cover on normalized degree = 0.010). The abundance

of AMF taxa in samples significantly predicted their centrality (log transformed to

make relationship linear: t = 8.32, P < 0.0001) and normalized degree (t = 21.71, P <

0.0001).

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Thus, unsurprisingly, the network position of species was correlated with their

abundance. However despite this, the influence of plant centrality on attachment

and detachment probability of fungi remained significant (Table S4), even after

controlling for species abundances (i.e. including abundance as a covariate to test

whether centrality affected attachment or detachment probability at a given level of

abundance). We used aboveground cover as a measure of plant abundance, which

had the advantage of being measured independently from our plant-AMF

associations. However, in case aboveground cover was not correlated with root

abundance, we repeated the analyses with the abundance of each plant species in

the root samples as a covariate, and the effect of plant centrality on fungal

attachment remained significant (Table S4). In fact, plant abundance (using either

measure) was not significantly correlated with AMF attachment probability (Table

S4), though there was a non-significant tendency for AMF detachment probability to

increase with plant abundance. In contrast to this consistent relation between

attachment or detachment and centrality, the relation between normalized degree

and attachment or detachment probability of AMF became non-significant when

abundance (either aboveground cover or abundance in samples) was included as a

covariate (|t| < 1, P > 0.343 in both cases). This indicates that any of the variance in

attachment or detachment probability that was attributable to normalized degree

was also collinear with plant abundance, due to the strong correlation between

abundance and degree.

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Plants did not show any preferential attachment in the original analyses (Table S2),

and the addition of abundance as a covariate did not alter this (Table S4). However,

the formerly significant effect of AMF centrality on plant detachment probability

became non-significant when the abundance of AMF taxa in samples was added as a

covariate in the models. Moreover, the non-significant trend towards a positive

correlation between AMF degree and plant detachment probability became

significant when abundance was included in the model, in contrast with previous

studies showing that species with high-degree are more likely to retain interactions

and their partners have a reduced probability of extinction (7, 10).

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1146 **SI Figure legends**

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Fig. S1 Measurement of partner overlap. Illustration of a network of interactions

among plants and arbuscular mycorrhizal fungi (AMF). The network can be depicted

as a bipartite matrix, where black squares indicate the presence of an interaction

('link') in the network. A unipartite projection of this network can also be generated

for each trophic level (here depicted in red, only for AMF), whereby species are

linked when they share an interaction partner. This unipartite projection depicts

interaction-partner overlap among species. We then measured each species' partner

overlap as 'closeness centrality': the inverse of the mean path length (i.e. 'degrees of

separation') from the focal species to all others in the unipartite projection. This

distance has also been called 'functional specialization' (43). For example, species E

is connected directly to species C, D, F and G, has two steps to species B (via C) and

three steps to species A (via C and B). This gives species E a mean path length of 1.5

to any other species, which is the shortest mean path length and the highest

closeness centrality of any AMF species. Species A and G have the same number of

links in the unipartite projection (1 link), but G shares a plant partner with species E

that shares partners with many others in the network (mean path length of G = 2.3).

In contrast, A shares a partner with species B, which itself only shares partners with

A and C. Thus, A has a lower overall partner overlap (mean path length = 2.83)

because it does not share partners with plants in the largest cluster of interacting

species.

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1170 **Fig. S2 Plant-AMF networks along chronosequence.**

1171 Quantitative networks are shown (whereby the width of links indicates their relative

1172 frequency). However, binary (links unweighted) networks were used for analysis

1173 because frequency of association is not an unbiased measure of interaction

1174 frequency in plant-mycorrhizal associations (69).

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Fig. S3 Relationship between normalized degree and centrality. Because we

compared preferential attachment based on interaction-partner overlap (defined as

'closeness centrality', Fig. S1) with a similar process based on degree, we tested

whether degree and centrality were correlated. We conducted Gaussian mixed

effects models with normalized degree (log transformed) as the predictor variable

and closeness centrality as the response. To account for the non-independence of

species from a given site, site age was included as a random effect. Mixed effects

models were conducted in the 'lme4' package (39) for R. P values for the models

were estimated using degrees of freedom calculated with the Satterthwaite method

of denominator synthesis, carried out in the 'lmerTest' package (75) for R. In the

figure, degree is log transformed and closeness centrality is presented as residuals

after removing variation due to random effects. All relationships were significantly

positive (mixed effects models: $t > 13$, $P < 0.0001$ in all cases).

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Fig. S4 Changes in interaction nestedness during simulated assembly and

disassembly phases. Nestedness (raw NODF top row, deviation from null

expectation bottom row) during simulated arrival (left column) and extinction (right

column) of plant and AMF species. Each scenario was run for 1000 replicates. Dark

horizontal lines show median values, boxes show upper and lower quartiles, and

whiskers show minima and maxima (with points signaling extreme values). The

scenarios were preferential attachment or detachment based on the degree (blue)

or closeness centrality (purple) of interaction partners (or inverse of degree and

centrality for detachment). For comparison, random extinctions are presented in

grey. Both assembly and disassembly phases had notable impacts on network

nestedness. During the assembly phase, the scenario based on preferential

attachment to high-degree nodes produced networks that were highly nested, and

considerably more nested than the other two scenarios. Nestedness tended to

increase with network size (through time), emphasizing the need to compare raw

nestedness against a null model. When compared against null expected nestedness,

only the degree-based scenario had increasing nestedness through time. In contrast,

the centrality-based and random assembly scenarios showed a slight decrease in

nestedness through time (as the addition of new nodes diluted the legacy of the

starting matrix, which was highly nested). Despite this, the centrality-based scenario

was consistently more nested than the random attachment scenario. Importantly,

the differences between the degree- and centrality-based scenarios were reversed

during the disassembly phase. As progressively more species were removed, the

degree-based scenario showed a rapid decline in nestedness, whereas the centrality-

based scenario became slightly more nested than null expectation (and more nested

than the centrality-based scenario).

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Table S1: Site details.
Details of site locations (Lat. = latitude, Long. = longitude), stage of ecosystem development, the
number of plant and arbuscular mycorrhizal fungal (AMF) taxa sampled and interactions (Links)
between them (with cumulative numbers up to, and including, that site in parentheses), and site
conditions. Additional details can be found in Richardson et al. (30) and updated in Holdaway et al.
(69).

Site age (years)	Stage	Lat.	Long.	Dominant vegetation	Plant taxa	AMF taxa	Links
<5	Progressive	-43.25	170.10	sparse herbfield	6	15	37
15	Progressive	-43.42	170.16	tall shrubland	20 (23)	24 (29)	113 (140)
70	Progressive	-43.42	170.16	early successional forest	9 (28)	13 (29)	57 (180)
290	Progressive	-43.41	170.18	broadleaved forest	13 (35)	19 (32)	73 (239)
500	Progressive	-43.41	170.17	broadleaved forest	11 (38)	15 (32)	55 (272)
1000	Progressive	-43.40	170.17	tall broadleaved forest	8 (41)	17 (32)	61 (307)
5000	Progressive	-43.40	170.17	tall broadleaved forest	9 (43)	15 (32)	61 (321)
12,000	Peak biomass	-43.33	170.21	tall broadleaved-podocarp forest	10 (46)	13 (32)	55 (344)
60,000	Retrogressive	-43.24	170.30	broadleaved-podocarp forest	11 (48)	15 (33)	64 (359)
120,000	Retrogressive	-43.25	170.19	short broadleaved-podocarp forest	11 (53)	14 (33)	61 (399)

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Table S2: Coefficients tables from attachment and detachment models for plants and arbuscular mycorrhizal fungi (AMF). Coefficients tables for fixed effects component of generalised linear mixed models testing for correlation between attachment and detachment probability and (closeness) centrality (our measure of interaction-partner overlap, see Fig. S1) or degree. Response variables are binary (attached/detached vs. did not attach/detach), and a binomial error distribution with a logit link function was used. All models had site included as a random factor to control for the non-independence of any new associations occurring at the same point along the chronosequence. Separate models are presented for (normalized) degree vs. centrality, as described in the main text. Additional models are presented with both centrality and degree to demonstrate that the significance and direction of effect for centrality does not change (though that of degree does in some cases). For each response variable, the best-fitting model (lowest AIC) is in bold.

Response variable		Estimate	Std. Error	Z	P	AIC
AMF attachment probabilty	(Intercept)	-8.651	3.938	-2.197	0.028	60.2
	Degree	-1.112	1.325	-0.839	0.401	
	(Intercept)	-5.862	0.886	-6.616	< 0.0001	58.1
	Centrality	31.720	5.591	5.674	< 0.0001	
	(Intercept)	**-5.733**	**1.131**	**-5.067**	**< 0.0001**	**53.8**
	Centrality	**44.248**	**8.899**	**4.972**	**< 0.0001**	
	Degree	**-3.973**	**1.633**	**-2.433**	**0.015**	
Plant attachment probabilty	(Intercept)	-0.384	0.444	-0.864	0.388	522.3
	Degree	-0.112	0.534	-0.209	0.835	
	(Intercept)	**-1.065**	**0.865**	**-1.231**	**0.218**	**522.0**
	Centrality	**15.972**	**19.968**	**0.800**	**0.424**	
	(Intercept)	-1.747	0.975	-1.793	0.073	523.1
	Centrality	40.984	27.932	1.467	0.142	
	Degree	-0.734	0.700	-1.048	0.295	
AMF detachment probabilty	(Intercept)	-3.259	1.256	-2.595	0.009	421.9
	Degree	1.820	0.885	2.057	0.040	
	(Intercept)	1.084	0.537	2.019	0.044	417.7
	Centrality	-95.425	22.235	-4.292	< 0.0001	
	(Intercept)	**0.834**	**0.632**	**1.320**	**0.187**	**411.4**
	Centrality	**-121.885**	**28.563**	**-4.267**	**< 0.0001**	
	Degree	**2.646**	**0.934**	**2.835**	**0.005**	
Plant detachment probabilty	(Intercept)	-0.256	0.589	-0.435	0.664	602.5
	Degree	0.247	0.442	0.558	0.577	
	(Intercept)	2.252	0.327	6.890	< 0.0001	593.8
	Centrality	-48.616	7.554	-6.436	< 0.0001	
	(Intercept)	**2.149**	**0.329**	**6.527**	**< 0.0001**	**593.2**
	Centrality	**-54.633**	**8.486**	**-6.438**	**< 0.0001**	
	Degree	**0.759**	**0.476**	**1.594**	**0.111**	

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Table S3: No significant change in network architecture along the chronosequence. Results of
best-fitting models for each metric of plant-mycorrhizal network structure (response variable) and any
predictor variables that were retained after model simplification. The maximal model contained sample
size (number of samples yielding an identifiable sequence) and site age and a quadratic site age term
(to allow for nonlinear relationships) as predictors. We compared all subsets of this maximal model and
selected the best-fitting model with the lowest AIC score. Raw P values are provided, though these are
non-conservative because multiple testing increases the probability of Type I error. Where no
predictors are given, the best-fitting model contained only an intercept. No measure of network
structure changed significantly along the chronosequence.
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Response	Predictor	D.F.	F	P
Nestedness (NODF)	Sample size	1,8	4.3	0.073
Nestedness temperature	Sample size	1,8	1.9	0.207
Connectance	< none >			
Binary linkage density	< none >			
Weighted linkage density	Sample size	1,6	1.7	0.234
	Log site age	1,6	0.0	0.931
	$(\text{Log site age})^2$	1,6	1.6	0.253
Interaction evenness	Sample size	1,8	4.1	0.077
Plant richness	< none >			
Mycorrhizal richness	Sample size	1,6	3.4	0.113
	Log site age	1,6	0.2	0.697
	$(\text{Log site age})^2$	1,6	3.0	0.133

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Response variable		Coefficient	Std. Error	Z	P
AMF attachment probability	(Intercept)	-5.925	1.366	-4.339	< 0.0001
	Percent cover	0.004	0.063	0.062	0.951
	Centrality	32.057	7.879	4.069	< 0.0001
	(Intercept)	-5.681	1.370	-4.147	< 0.0001
	Sample abundance	-0.020	0.119	-0.165	0.869
	Centrality	31.016	6.862	4.52	< 0.0001
Plant attachment probability	(Intercept)	-1.092	1.181	-0.925	0.355
	Sample abundance	-0.001	0.010	-0.067	0.946
	Centrality	16.921	30.149	0.561	0.575
AMF detachment probability	(Intercept)	0.305	0.254	1.203	0.229
	Percent cover	0.011	0.006	1.813	0.070
	Centrality	-72.357	9.615	-7.525	< 0.0001
	(Intercept)	0.275	0.267	1.028	0.304
	Sample abundance	0.024	0.014	1.772	0.076
	Centrality	-72.342	10.088	-7.171	< 0.0001
Plant detachment probability	(Intercept)	0.196	1.110	0.176	0.860
	Sample abundance	0.008	0.003	2.368	0.018
	Centrality	-9.550	21.276	-0.449	0.654

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