The promise of disease gene discovery in South Asia

Nathan Nakatsuka[1,2], Priya Moorjani[3,6], Niraj Rai[4], Biswanath Sarkar[5], Arti Tandon[1,6], Nick Patterson[6], Gandham SriLakshmi Bhavani[7], Katta Mohan Girisha[7], Mohammed S Mustak[8], Sudha Srinivasan[9], Amit Kaushik[10], Saadi Abdul Vahab[11], Sujatha M. Jagadeesh[12], Kapaettu Satyamoorthy[11], Lalji Singh[4,13], David Reich[1,5,14,*], Kumarasamy Thangaraj[4,*]

[1]Department of Genetics, Harvard Medical School, New Research Building, 77 Ave. Louis Pasteur, Boston, MA 02115, USA
[2]Harvard-MIT Division of Health Sciences and Technology, Harvard Medical School, Boston, MA 02115, USA
[3]Department of Biological Sciences, Columbia University, 600 Fairchild Center, New York, NY 10027, USA
[4]CSIR-Centre for Cellular and Molecular Biology, Habsiguda, Hyderabad, Telangana 500007, India
[5]Superintending Anthropologist (Physical) (Rtd.), Anthropological Survey of India, 27 Jawaharlal Nehru Road, Kolkata 700016, India
[6]Broad Institute of Harvard and Massachusetts Institute of Technology, Cambridge, MA 02141, USA
[7]Department of Medical Genetics, Kasturba Medical College, Manipal University, Manipal, India
[8]Department of Applied Zoology, Mangalore University, Mangalagangothri 574199, Mangalore, Karnataka, India
[9]Centre for Human Genetics, Biotech Park, Electronics City (Phase I), Bangalore 560100, India
[10]Amity Institute of Biotechnology, Amity University, Sector125, Noida 201303, India
[11]School of Life Sciences, Manipal University, Manipal 576104, India
[12]Fetal Care Research Foundation, 197 Dr. Natesan Road, Chennai 600004, India
[13]Present address: Genome Foundation, Hyderabad 500076, India
[14]Howard Hughes Medical Institute, Harvard Medical School, Boston, MA 02115, USA
*co-senior authors

The more than 1.5 billion people who live in South Asia are correctly viewed not as a single large ethnic group, but as many small endogamous groups. We assembled genome-wide data from over 2,800 individuals from over 275 distinct South Asian groups. We document shared ancestry across groups that correlates with geography, language, and religious affiliation. We characterize the strength of the founder events that gave rise to many of the groups, and identify 14 groups with census sizes of more than a million that descend from founder events significantly stronger than those in Ashkenazi Jews and Finns, both of which have high rates of recessive disease due to founder events. These results highlight a major and under-appreciated opportunity for reducing disease burden among South Asians through the discovery of and testing for recessive disease genes. As a proof-of-principle, we show that it is possible to localize genes for progressive pseudorheumatoid dysplasia and mucopolysaccharidosis type IVA, two recessive diseases prevalent in South India, using the founder event disease gene mapping approach introduced here.

South Asia is a region of extraordinary cultural, linguistic, and genetic diversity, with a conservative estimate of over 4,600 anthropologically well-defined groups, many of which are endogamous communities with significant barriers to gene flow due to sociological and cultural factors that restrict marriage between groups[1]. Of the small fraction of South Asian groups that have been characterized using genome-wide data, many exhibit large allele frequency differences from geographically close neighbors[2-4], indicating that they have experienced strong founder events, whereby a small number of ancestors gave rise to many descendants today[4]. The pervasive founder events in South Asia present a major opportunity for reducing disease burden in South Asia. The promise is highlighted by studies of founder groups of European ancestry – including Ashkenazi Jews, Finns, Amish, Hutterites, Sardinians, and French Canadians – which have resulted in the discovery of dozens of recessive disease causing mutations in each group. Prenatal testing for these mutations has substantially reduced recessive disease burden in all of these communities[5,6].

To characterize the medically relevant founder events in South Asia, we carried out new genotyping of 1,663 samples from 230 endogamous groups in South Asia on the Affymetrix Human Origins single nucleotide polymorphism (SNP) array[7]. We developed an algorithm to measure the strength of an endogamous group's founder event, aiming to genotype at least five samples per group as our power calculations indicated that this was sufficient to detect a founder event stronger than that in Finns with high confidence (Supplementary Figure 1 and Supplementary Table 1). We combined the new data we collected with previously reported data, leading to four datasets (Figure 1a). The Affymetrix Human Origins SNP array data comprised 1,955 individuals from 249 groups in South Asia, to which we added 7 Ashkenazi Jews. The Affymetrix 6.0 SNP array data comprised 383 individuals from 52 groups in South Asia[4,8]. The Illumina SNP array data comprised 188 individuals from 21 groups in South Asia[9] and 21 Ashkenazi Jews[9,10]. The Illumina Omni SNP array data comprised 367 individuals from 20 groups in South Asia[11]. We merged 1000

89   Genomes Phase 3 data[12] (2,504 individuals from 26 different groups including 99
90   Finns) with each of these datasets. We performed quality control to remove SNPs
91   and individuals with a high proportion of missing genotypes or that were outliers in
92   Principal Component Analysis (PCA).
93

We performed PCA on each of the three different datasets along with European
Americans (CEU), Han Chinese (CHB), and West Africans (YRI), and found that the
Siddi are strong outliers as previously reported (Supplementary Figure 2)[4,13,14]. We
next removed YRI, Siddi and indigenous Andamanese (another known outlier) from
the datasets and repeated PCA (Figure 1b, Supplementary Figure 3). Similar to past
studies, the PCA documents three broad genetic groupings[4,8,11]. First, almost all
South Asian groups speaking Indo-European and Dravidian languages lie along the
"Indian Cline," with different proportions of Ancestral North Indian (ANI) ancestry
related to Europeans, Central Asians, and Near Easterners, and Ancestral South
Indian (ASI) ancestry that is as different from ANI as Europeans and East Asians are
from each other[4]. The second major cluster includes groups that speak Austroasiatic
languages, as well as some non-Austroasiatic speaking groups that have similar
ancestry possibly due to gene flow with Austroasiatic speaking neighbors or due to
a history of language shift. This set of groups cluster together near the ASI end of the
Indian cline, likely reflecting a large proportion of ASI-like ancestry as well as a
distinct ancestry that has some affinity to East Asians. The Tibeto-Burman speaking
groups and other groups with high proportions of East Asian related ancestry such
as the Bengali and Austroasiatic speaking Khasi form a gradient of ancestry relating
them to East Asian groups such as Han Chinese. These groupings are also evident in
a neighbor-joining tree based on F_{ST} (Supplementary Figure 4). We confirmed the
East Asian related mixture in some groups by observing significantly negative
f_3(Test; Mala, Chinese) statistics[7] (Supplementary Table 2).
116

We devised an algorithm to quantify the strength of the founder events in each
group based on Identity-by-Descent (IBD) segments, large stretches of DNA shared
from a common founder in the last approximately one hundred generations (Figure
2). We computed an "IBD score" as a measure for the strength of the founder event
in each group's history: the average length of IBD segments between 3-20
centimorgans (cM) shared between two genomes normalized to sample size. Since
we are interested here in recessive diseases that do not owe their origin to
consanguineous marriages of close relatives, we filtered the data to minimize this
effect by computing IBD between all pairs of individuals in each group and removing
one individual from the pairs with outlying numbers of IBD segments. We validated
the effectiveness of this procedure for removing close relatives by simulation
(Supplementary Table 2; Online Methods).
129

We expressed IBD scores for each group as a fraction of the IBD scores of the 1000
Genomes Project Finns merged into each respective dataset. Due to the fact that all
the arrays we analyzed included more SNPs ascertained in Europeans than in South
Asians, the sensitivity of our methods to founder events is expected to be greater in
Europeans than in South Asians, and thus, our estimates of founder event strengths

in South Asian groups is likely to be a conservative underestimate relative to that in
Europeans (Supplementary Figure 5 demonstrates this effect empirically and shows
that it is less of a bias for the strong founder events that are the focus of this study).
We computed standard errors for these ratios by a weighted Block Jackknife across
chromosomes and declared significance where the 95% confidence intervals did not
overlap with 1. Our simulations suggest that for sample sizes of 4, the algorithm's
sensitivity was greater than 95% for determining that a group with two times the
bottleneck strength as Finns would have an IBD score significantly greater than that
of Finns, while specificity was perfect in all the simulations we performed
(Supplementary Figure 1 and Supplementary Table 1). We also used two other
methods for measuring founder events that did not require phasing or IBD
detection. First, we computed F_{ST} between each group and every other group with
similar ancestry sources. Second, for groups on the Indian Cline we fit a model of
population history using *qpGraph*[7] and measuring the founder event as the group-
drift after admixture (Supplementary Figure 6 and Online Data Table 1). The results
of both methods were highly correlated to that of the IBD-based method for all cases
where a comparison was possible (Pearson correlation r=0.82-0.98).
152
The IBD analyses suggest that 29% of the South Asian groups we analyzed (96 out
of 327) have significantly stronger founder events than those in both Finns and
Ashkenazi Jews (Figure 3). The South Asian groups with evidence of strong founder
events include diverse tribe, caste, and religious groups, and our analysis identifies
14 groups with strong founder events census sizes of over a million (Figure 3; Table
1). The groups with smaller census sizes are also medically significant. Study of
small census size groups with extremely strong founder events such as Amish,
Hutterites, and the people of the Saguenay Lac-St. Jean region have led to the
discovery of dozens of novel disease variants specific to each group[6], which
highlights the potential of similar studies in South Asian groups. In addition to these
analyses, we measured IBD across groups – searching for cases in which the across-
group IBD score is at least a third of the within-group IBD score of Ashkenazi Jews –
and found many cases of relatedness, which typically follow geography, religious
affiliation (e.g. Catholic Brahmins), or linguistic grouping (particularly Austroasiatic
speakers) (Supplementary Table 3).
168
Our documentation that medically significant founder events affect a large fraction
of South Asian groups presents an opportunity for decreasing disease burden. This
source of risk for recessive diseases is very different from that due to marriages
among close relatives, which is also a major cause of recessive disease, especially in
southern India. In the case of recessive diseases arising due to founder events, there
are mutations that occur recurrently across members of a group (due to deeply
shared founders), and these can be mapped at a group level and easily tested.
176
177 As proof-of-principle, we highlight two examples. The first concerns the Vysya, a
178 group with a census size of more than 3 million that we identified as having a
179 founder event about 1.2-fold stronger than that in Finns (Figure 3). The Vysya have
180 a 100-fold higher rate of butyrylcholinesterase deficiency than other Indian groups,

181 and Vysya ancestry is a known counter-indication for the use of muscle relaxants
182 such as succinylcholine or mivacurium that are given prior to surgery[15].

184 The second proof-of-principle, we newly genotyped 18 patients from India, 12 of
185 whom had progressive pseudorheumatoid dysplasia (PPD), a disease known to be
186 caused by mutations in the gene *WISP3*[16,17], and 6 of whom had
187 mucopolysaccharidosis type IVA, known to be caused by mutations in the gene
188 *GALNS*[18]. Though we lacked ethnic group information for most of the 18 patients, 6
189 of the PPD patients carried Cys78Tyr mutations, and 5 of these 6 were from non-
190 consanguineous marriages. We found a much higher fraction of IBD at the disease
191 mutation site than in the rest of the genome (Supplementary Figure 7a). Thus,
192 Cys78Tyr in PPD is a mutation that owes its origin to a founder event[16,17]. The 6
193 other PPD patients carried Cys337Tyr mutations, and 6 of 6 were from
194 consanguineous marriages, while the 6 patients with MPS carried Cys79Arg
195 mutations, and 4 of 6 were from consanguineous marriages. These patients did not
196 have IBD at the disease mutation site that was detectable using our conservative
197 settings, but we were still able to map the disease locus using homozygosity
198 mapping (Supplementary Figures 7b and 7c) similar to methods used by others[19,20].
199 When we examined the haplotypes at the disease loci, we found that each mutation
200 group had high sharing of unique haplotypes (Supplementary Figure 8), but the PPD
201 Cys337Tyr and MPS Cys79Arg haplotypes were smaller than the PPD Cys78Tyr
202 haplotypes. This suggests that these 2 mutations are at high frequency due to older
203 founder events than the one that occurred for the PPD Cys78Tyr mutations, which
204 could explain why they were not discovered by IBD (which is most sensitive for
205 young founder events) and also why they are present primarily in individuals
206 descending from consanguineous marriages (because they may be sufficiently rare
207 that they do not come together at an appreciable rate except in the context of a
208 consanguineous marriage). Beyond the new genotyping we performed here, another
209 study this year demonstrated that an Indian founder mutation in *ISCA1* mutation
210 causes predisposition to a severe mitochondrial dysfunction syndrome.[21]

212 These observations highlight how systematic studies of South Asian founder groups
213 are likely to be an effective approach for discovering mutations that cause recessive
214 disease. Identification of pathogenic mutations responsible for such founder
215 diseases is straightforward. All that is required is collection of DNA samples from a
216 small number of affected individuals and their families, usually followed by whole-
217 exome sequencing to discover the causal changes. Once group specific founder
218 event disease mutations are discovered, they can be tested for prenatally. Mapping
219 of recessive disease mutations may be particularly important in traditional
220 communities practicing arranged marriages, which is common in India. An example
221 of the power of this approach from outside India is given by *Dor Yeshorim*, a
222 community genetic testing program among religious Ashkenazi Jews[22], which visits
223 schools, screens students for common recessive disease causing mutations
224 previously identified to be segregating at a higher frequency in the target group, and
225 enters the results into a confidential database. Match-makers query the database
226 prior to making suggestions to the families and receive feedback about whether the

potential couple is "incompatible" in the sense of both being carriers for a recessive
mutation at the same gene. Given that approximately 95% of community members
whose marriages are arranged participate in this program, recessive diseases like
Tay-Sachs have virtually disappeared in these communities. A similar approach
should work as well in Indian communities where arranged marriages are common.
Given the potential for saving lives, this or similar kinds of research could serve as
an important investment for future generations[23].
234
This study of more than 275 distinct groups represents the first systematic survey
for founder events in South Asia, and to our knowledge also presents the richest
dataset of genome-wide data from anthropologically well-documented groups
available from any region in the world. Despite the breadth of this data, the groups
surveyed here represent only about 5% of the well-documented endogamous
groups in South Asia, and extensions of the survey to all groups would make it
possible to identify large numbers of additional founder groups susceptible to
recessive diseases. To take advantage of the unique population structure of South
Asia to improve health, we propose to reverse the standard approach to recessive
disease gene discovery. Instead of focusing entirely on collecting cases in tertiary
medical centers and mapping diseases in a group of individuals found to have the
same phenotype while blinded to information about their caste or tribal status, we
propose that medical geneticists should adopt a parallel strategy of working with
community doctors, medical workers, and social workers to identify recessive
diseases that occur at a high rate in endogamous groups with founder events, and
then go out into the community to identify cases. Once a small number of cases are
sampled, it is straightforward to map the causal variants. This approach was
pioneered in 1950s by the work of Victor McKusick and his colleagues studying the
Old Order Amish in Pennsylvania U.S.A., a founder population of approximately
100,000 individuals in whom many dozens of recessive diseases were mapped, a
research program that was crucial to founding modern medical genetics and was of
extraordinary health benefit to that community[24]. Our study suggests that the
potential for disease gene mapping in India would be orders of magnitude greater.
258

Supplementary Data:

Supplementary Data include an excel spreadsheet detailing all groups and their scores on the IBD, F_{ST}, and group-specific drift analyses. Also included are 8 supplementary figures and 5 supplementary tables.

Acknowledgements:

We are thankful to the many Indian, Pakistani, Bangladeshi, and Nepalese individuals who contributed the DNA samples analyzed here including in particular the Indian PPD and MPS patients. We are grateful to Analabha Basu and Partha P. Majumder for sharing data. Funding was provided by an NIGMS (GM007753) fellowship to NN, a Translational Seed Fund grant from the Dean's Office of Harvard Medical School to DR, CSIR network project GENESIS (BSC0121) to KT, and an NIGMS grant 115006 to OPM. SS and SMJ acknowledge the funding from the Department of Biotechnology (BT/PR4224/MED/97/60/2011) and Department of Science and Technology (SR/WOS-A/LS-83/2011), Government of India. DR is an Investigator of the Howard Hughes Medical Institute. Genotyping data for the samples collected for this study will be made available upon request from the corresponding authors.

References:

1. Mastana, S.S. Unity in diversity: an overview of the genomic anthropology of India. *Ann Hum Biol* **41**, 287-99 (2014).
2. Bamshad, M.J. *et al.* Female gene flow stratifies Hindu castes. *Nature* **395**, 651-2 (1998).
3. Basu, A. *et al.* Ethnic India: a genomic view, with special reference to peopling and structure. *Genome Res* **13**, 2277-90 (2003).
4. Reich, D., Thangaraj, K., Patterson, N., Price, A.L. & Singh, L. Reconstructing Indian population history. *Nature* **461**, 489-94 (2009).
5. Lim, E.T. *et al.* Distribution and medical impact of loss-of-function variants in the Finnish founder population. *PLoS Genet* **10**, e1004494 (2014).
6. Arcos-Burgos, M. & Muenke, M. Genetics of population isolates. *Clin Genet* **61**, 233-47 (2002).
7. Patterson, N. *et al.* Ancient admixture in human history. *Genetics* **192**, 1065-93 (2012).
8. Moorjani, P. *et al.* Genetic evidence for recent population mixture in India. *Am J Hum Genet* **93**, 422-38 (2013).
9. Metspalu, M. *et al.* Shared and unique components of human population structure and genome-wide signals of positive selection in South Asia. *Am J Hum Genet* **89**, 731-44 (2011).
10. Behar, D.M. *et al.* The genome-wide structure of the Jewish people. *Nature* **466**, 238-42 (2010).
11. Basu, A., Sarkar-Roy, N. & Majumder, P.P. Genomic reconstruction of the history of extant populations of India reveals five distinct ancestral components and a complex structure. *Proc Natl Acad Sci U S A* (2016).
12. Genomes Project, C. *et al.* A global reference for human genetic variation. *Nature* **526**, 68-74 (2015).
13. Narang, A. *et al.* Recent admixture in an Indian population of African ancestry. *Am J Hum Genet* **89**, 111-20 (2011).
14. Shah, A.M. *et al.* Indian Siddis: African descendants with Indian admixture. *Am J Hum Genet* **89**, 154-61 (2011).
15. Manoharan, I., Wieseler, S., Layer, P.G., Lockridge, O. & Boopathy, R. Naturally occurring mutation Leu307Pro of human butyrylcholinesterase in the Vysya community of India. *Pharmacogenet Genomics* **16**, 461-8 (2006).
16. Dalal, A. *et al.* Analysis of the WISP3 gene in Indian families with progressive pseudorheumatoid dysplasia. *Am J Med Genet A* **158A**, 2820-8 (2012).
17. Bhavani, G.S. *et al.* Novel and recurrent mutations in WISP3 and an atypical phenotype. *Am J Med Genet A* **167A**, 2481-4 (2015).
18. Bidchol, A.M. *et al.* GALNS mutations in Indian patients with mucopolysaccharidosis IVA. *Am J Med Genet A* **164A**, 2793-801 (2014).
19. Hildebrandt, F. *et al.* A systematic approach to mapping recessive disease genes in individuals from outbred populations. *PLoS Genet* **5**, e1000353 (2009).

20. Hagiwara, K. *et al.* Homozygosity mapping on homozygosity haplotype
analysis to detect recessive disease-causing genes from a small number of
unrelated, outbred patients. *PLoS One* **6**, e25059 (2011).
21. Anju Shukla, M.H., Anshika Srivastava, Rajagopal Kadavigere, Priyanka
Upadhyai, Anil Kanthi, Oliver Brandau, Stephanie Bielas, Katta Girisha.
Homozygous c.259G>A variant in ISCA1 is associated with a new multiple
mitochondrial dysfunctions syndrome. *bioRxiv* (2016).
22. Raz, A.E. Can population-based carrier screening be left to the community? *J*
Genet Couns **18**, 114-8 (2009).
23. Rajasimha, H.K. *et al.* Organization for rare diseases India (ORDI) -
addressing the challenges and opportunities for the Indian rare diseases'
community. *Genet Res (Camb)* **96**, e009 (2014).
24. Francomano, K.R.D.a.C.A. Victor A. McKusick and the History of Medical
Genetics. *Springer Science & Business Media*, 119-130 (2012).
25. Sudmant, P.H. *et al.* Global diversity, population stratification, and selection
of human copy-number variation. *Science* **349**, aab3761 (2015).
26. Mallick, S. *et al.* The Simons Genome Diversity Project: 300 genomes from
142 diverse populations. *Nature* **538**, 201-206 (2016).
27. Sudmant, P.H. *et al.* An integrated map of structural variation in 2,504 human
genomes. *Nature* **526**, 75-81 (2015).
28. Patterson, N., Price, A.L. & Reich, D. Population structure and eigenanalysis.
PLoS Genet **2**, e190 (2006).
29. Gusev, A. *et al.* Whole population, genome-wide mapping of hidden
relatedness. *Genome Res* **19**, 318-26 (2009).
30. Hoaglin, B.I.a.D. *How to Detect and Handle Outliers*, (1993).
31. Palamara, P.F. ARGON: fast, whole-genome simulation of the discrete time
Wright-fisher process. *Bioinformatics* (2016).
32. Felsenstein, J. PHYLIP - Phylogeny Inference Package (Version 3.2). *Cladistics*
5, 164-166 (1989).
33. Letunic, I. & Bork, P. Interactive Tree Of Life v2: online annotation and
display of phylogenetic trees made easy. *Nucleic Acids Res* **39**, W475-8
(2011).
34. Browning, S.R. & Browning, B.L. Rapid and accurate haplotype phasing and
missing-data inference for whole-genome association studies by use of
localized haplotype clustering. *Am J Hum Genet* **81**, 1084-97 (2007).
35. Durand, E.Y., Eriksson, N. & McLean, C.Y. Reducing pervasive false-positive
identical-by-descent segments detected by large-scale pedigree analysis. *Mol*
Biol Evol **31**, 2212-22 (2014).
36. Browning, B.L. & Browning, S.R. Improving the accuracy and efficiency of
identity-by-descent detection in population data. *Genetics* **194**, 459-71
(2013).
364

Group	Sample Size	IBD Score	IBD Rank	F_{ST} Rank	Drift Rank	Census Size	Location
Gujjar	5	11.6	19	20	46	1,078,719	Jammu and Kashmir
Baniyas	7	9.6	24	24	18	4,200,000	Uttar Pradesh
Pattapu_Kapu	4	9.5	25	24	21	13,697,000	Andhra Pradesh
Vadde	3	9.2	26	34	26	3,695,000	Andhra Pradesh
Yadav	12	4.4	48	102	67	1,124,864	Puducherry
Kshatriya_Aqnikula	4	2.4	75	154	NA	12,809,000	Andhra Pradesh
Naga	4	2.3	76	NA	NA	1,834,483	Nagaland
Kumhar	27	2.3	77	222	197	3,144,000	Uttar Pradesh
Reddy	7	2.0	84	133	106	22,500,000	Telangana
Brahmin_Nepal	4	1.9	86	95	141	4,206,235	Nepal
Kallar	27	1.7	94	95	73	2,426,929	Tamil Nadu
Brahmin_Manipuri	17	1.6	99	NA	NA	1,544,296	Manipur
Arunthathiyar	18	1.3	108	133	81	1,192,578	Tamil Nadu
Vysya	39	1.2	110	55	35	3,200,000	Telangana

Table 1. South Asian groups with census sizes over 1 million and IBD scores greater than those of Ashkenazi Jews
and Finns. Fourteen South Asian groups with IBD scores significantly higher than that of Finns, census sizes over 1
million, and sample sizes of at least 3 that are of particularly high interest for founder event disease gene mapping studies.
For reference, Finns and Ashkenazi Jews (on Human Origins) would have IBD scores of 1.0 and 0.9, IBD ranks of 121 and
135, and F_{ST} ranks of 133 and 154, respectively (the group-specific drift is difficult to compare for groups with
significantly different histories, so they were not calculated for Finns or Ashkenazi Jews).

375
376

378 **Figure 1. Dataset overview.** (a) Sampling locations for all analyzed groups. Each
379 point indicates a distinct group (random jitter was added to help in visualization at
380 locations where there are many groups). (b) PCA of Human Origins dataset along
381 with European Americans (CEU) and Han Chinese (CHB).

A)

B)

C)

D)

Figure 2. Example histograms of IBD segments to illustrate the differences between groups with founder events of different magnitudes: These histograms provide visual illustrations of differences between groups with different IBD scores. As a ratio relative to Finns (FIN; black), these groups (red) have IBD scores of: (A) ~26 in Ulladan, (B) ~3 in Birhor, (C) ~0.9 in Ashkenazi Jews, and (D) ~0.1 in Mahadeo_Koli. In each plot, we also show European Americans (CEU) with a negligible founder event in blue. Quantification of these founder events is shown in Figure 3 and Online Table 1. The IBD histograms were normalized for sample size by dividing their frequency by $\left\{\binom{2n}{2} - n\right\}$, where n is the number of individuals in the sample. All data for the figure are based on the Human Origins dataset.

Figure 3. IBD scores relative to Finns (FIN). Histogram ordered by IBD score,
399
which is roughly proportional to the per-individual risk for recessive disease due to
the founder event. (These results are also given quantitatively for each group in
Online Table 1.) We restrict to groups with at least two samples, combining data
from all four genotyping platforms onto one plot. Data from Ashkenazi Jews and
Finns are highlighted in red, and from Indian groups with significantly stronger
founder events than Finns and census sizes of more than a million in brown. Error
bars for each IBD score are standard errors calculated by weighted block jackknife
over each chromosome. YRI=Yoruba (West African); CEU=Northern European.

Online Methods:

Data Sets:

408
409
410
411
We assembled a dataset of 1,955 individuals from 249 groups genotyped on the
Affymetrix Human Origins array, of which data from 1,663 individuals from 230
groups are newly reported here (Figure 1a). We merged these data with the dataset
published in Moorjani et al.[8], which consisted of 332 individuals from 52 groups
genotyped on the Affymetrix 6.0 array. We also merged it with two additional
datasets published in Metspalu et al.[9], consisting of 151 individuals from 21 groups
genotyped on Illumina 650K arrays as well as a dataset published in Basu et al.[11],
consisting of 367 individuals from 20 groups generated on Illumina Omni 1-Quad
arrays. These groups come from India, Pakistan, Nepal, Sri Lanka, and Bangladesh.
All samples were collected under the supervision of ethical review boards in India
with informed consent obtained from all subjects.
423
424 We analyzed two different Ashkenazi Jewish datasets, one consisting of 21
425 individuals genotyped on Illumina 610K and 660K bead arrays[10] and one consisting
426 of 7 individuals genotyped on Affymetrix Human Origins arrays.
427
Our "Affymetrix 6.0" dataset consists of 332 individuals genotyped on 329,261 SNPs,
and our "Illumina_Omni" dataset consists of 367 individuals genotyped on 750,919
SNPs. We merged the South Asia and Jewish data generated by the other Illumina
arrays to create an "Illumina" dataset which consists of 172 individuals genotyped
on 500,640 SNPs. Finally, we merged the data from the Affymetrix Human Origins
arrays with the Ashkenazi Jewish data and data from the Simons Genome Diversity
Project[25,26] to create a dataset with 4,402 individuals genotyped on 512,615 SNPs.
We analyzed the four datasets separately due to the small intersection of SNPs
between them and possible systematic differences across genotyping platforms. We
merged in the 1000 Genomes Phase 3 data[27] (2504 individuals from 26 different
groups; notably, including 99 Finnish individuals) into all of the datasets. We used
genome reference sequence coordinates (hg19) for analyses.
440

Quality Control:

441
442
443 We filtered the data at both the SNP and individual level. On the SNP level, we
444 required at least 95% genotyping completeness for each SNP (across all
445 individuals). On the individual level, we required at least 95% genotyping
446 completeness for each individual (across all SNPs).
447
To test for batch effects due to samples from the same group being genotyped on
different array plates, we studied instances where samples from the same group A
were genotyped on both plates 1 and 2 and computed an allele frequency difference
at each SNP, $Diff_A^i = \left(Freq_{PopA,Plate1}^i - Freq_{PopA,Plate2}^i\right)$. We then computed the
product of these allele frequencies averaged over all SNPs for two groups A and B
genotyped on the same plates, $\frac{1}{n}\sum_{i=1}^{n}\left(Diff_A^i\right)\left(Diff_B^i\right)$, as well as a standard error

from a weighted Block Jackknife across chromosomes. This quantity should be
consistent with zero within a few standard errors if there are no batch effects that
cause systematic differences across the plates, as allele frequency differences
between two samples of the same group should be random fluctuations that have
nothing to do with the array plates on which they are genotyped. This analysis
found strong batch effects associated with one array plate, and we removed these
samples from further analysis.
461
We used EIGENSOFT 5.0.1 smartpca[28] on each group to detect PCA outliers. We also
developed a procedure to distinguish recent relatedness from founder events so
that we could remove recently related individuals. We first identified all duplicates
or obvious close relatives by using Plink "genome" and GERMLINE[29] to compute IBD
(described in more detail below) and removed one individual from all pairs with a
PI_HAT score greater than 0.45 and the presence of at least 1 IBD fragment greater
than 30cM. We then used an iterative procedure to identify additional recently
related individuals. For sample sizes above 5, we identified any pairs within each
group that had both total IBD and total long IBD (>20cM) that were greater than 2.5
SDs and 1 SD, respectively, from the group mean. For sample sizes 5 or below, we
used modified Z scores of $0.6745*(IBD\_score - median(score))/MAD$, where MAD is
the median absolute deviation, and identified all pairs with modified Z scores
greater than 3.5 for both total IBD and total long IBD as suggested by Iglewicz and
Hoaglin[30]. After each round, we repeated the process if the new IBD score was at
least 30% lower than the prior IBD score. Simulations showed that we were always
able to remove a first or second cousin in the dataset using this method
(Supplementary Table 5). Together these analyses removed 53 individuals from the
Affymetrix 6.0 dataset, 21 individuals from the Illumina dataset, 43 individuals from
the Illumina Omni dataset, and 225 individuals from the Human Origins dataset.
481
482 After data quality control and merging with the 1000 Genomes Project data, the
483 Affymetrix 6.0 dataset included 2,842 individuals genotyped on 326,181 SNPs, the
484 Illumina dataset included 2,662 individuals genotyped on 484,293 SNPs, the
485 Illumina Omni dataset included 2,828 individuals genotyped on 750,919 SNPs, and
486 the Human Origins dataset included 4,177 individuals genotyped at 499,158 SNPs.
487
488 **Simulations to Test Relatedness Filtering and IBD Analyses**
489
We used ARGON[31] to simulate groups with different bottleneck strengths to test the
IBD analyses, relatedness filtering, and founder event dating algorithms. We used
ARGON's default settings, including mutation rate of $1.65*10^{-8}$ per base pair (bp) per
generation and a recombination rate of $1*10^{-8}$ per bp per generation and simulated
22 chromosomes of size 130 Mb each. We pruned the output by randomly removing
SNPs until there were 22,730 SNPs per chromosome to simulate the approximate
number of positions in the Affymetrix Human Origins array. For the IBD analyses,
we simulated groups to have descended from an ancestral group 1,800 years ago
with N_e=50,000 and to have formed two groups with N_e=25,000. These groups
continued separately until 100 generations ago when they combined in equal

proportions to form a group with N_e=50,000. The group then split into 3 separate
groups 72 generations ago that have bottlenecks leading to N_e of either 400, 800, or
1600. The 3 groups then exponentially expanded to a present size of N_e=50,000.
We designed these simulations to capture important features of demographic
history typical of Indian groups as detailed by Moorjani *et al.*[8] and Reich *et al.*[4] We
chose the bottleneck sizes because they represent founder events with
approximately the strength of Finns (the bottleneck to 800), and twice as strong
(400) and half as strong (1600) as that group. We then performed the IBD analyses
described below with 99 individuals from the group with bottleneck strength
similar to that of Finns (198 haploid individuals were simulated and merged to
produce 99 diploid individuals) and different number of individuals from the other
groups. These analyses demonstrate that with only 4-5 individuals we can
accurately assess the strength of founder events in groups with strong founder
events (Supplementary Figure 1 and Supplementary Table 1). Weaker founder
events are more difficult to assess, but these groups are of less interest for founder
event disease mapping, so we aimed to sample ~5 individuals per group.
516
We wrote custom R scripts to carry out forward simulations for creating first and
second cousins. We took individuals from the bottleneck of size 800 and performed
"matings" by taking 2 individuals and recombining their haploid chromosomes
assuming a rate of $1*10^{-8}$ per bp per generation across the chromosome and
combining one chromosome from each of these individuals to form a new diploid
offspring. The matings were performed to achieve first and second cousins. We then
placed these back into the group with group of size 800, and ran the relatedness
filtering algorithms to evaluate whether they would identify these individuals.
525

526
527 **Distance-Based Phylogenetic Tree:**
528 We calculated genetic differentiation (F_{ST}) between all pairs of groups using
529 EIGENSOFT *smartpca* and created a neighbor-joining tree using PHYLIP[32] with
530 Yoruba as the outgroup. We used Itol[33] to display the tree.
531

532
Phasing, IBD Detection, and IBD Score Algorithm:
We phased all datasets using Beagle 3.3.2 with the settings *missing=0; lowmem=true;*
gprobs=false; verbose=true[34]. We left all other settings at default. We determined IBD
segments using GERMLINE[29] with the parameters *-bits 75 -err_hom 0 -err_het 0 -*
min_m 3. We used the genotype extension mode to minimize the effect of any
possible phasing heterogeneity amongst the different groups and used the
HaploScore algorithm to remove false positive IBD fragments with the
recommended genotype error and switch error parameters of 0.0075 and 0.003[35].
We chose a HaploScore threshold matrix based on calculations from Durand *et al*.
for a "mean overlap" of 0.8, which corresponds to a precision of approximately 0.9
for all genetic lengths from 2-10cM. In addition to the procedure we developed to
remove close relatives (Quality Control section), we also removed segments longer
than 20cM as simulations showed that this increased sensitivity of the analyses
(Supplementary Table 4). We computed "IBD score" as the total length of IBD

segments between 3-20cM divided by $\left\{ \binom{2n}{2} - n \right\}$ where n is the number of
individuals in each group to normalize for sample size. We then expressed each
group's score as a ratio of their IBD score to that of Finns and calculated standard
errors for this score using a weighted Block Jackknife over each chromosome with
95% confidence intervals defined as IBD score ±1.96*s.e.

We also repeated these analyses with FastIBD[36] for the Affymetrix 6.0 and Illumina
datasets and observed that the results were highly correlated (r>0.96) (data not
shown). We chose GERMLINE for our main analyses, however, because the FastIBD
algorithm required us to split the datasets into different groups, since it adapts to
the relationships between LD and genetic distance in the data, and these
relationships differ across groups. We used data from several different Jewish
groups and all twenty-six 1000 Genomes groups to improve phasing, but of these
groups we only included results for Ashkenazi Jews and two outbred groups (CEU
and YRI) in the final IBD score ranking.

563
<u>Disease patient analyses:</u>

We use Affymetrix Human Origins arrays to genotype 15 patients with progressive
pseudorheumatoid dysplasia (PPD) and 6 patients with mucopolysaccharidosis
(MPS) type IVA, all of which had disease mutations previously determined[16-18], and
3 of which (MPS patients) are newly reported here. After quality control, 6 of the
PPD patients with Cys78Tyr mutations, 6 of the PPD patients with Cys337Tyr
mutations, and 6 of the MPS patients with Cys78Arg mutations remained. We
measured IBD as described above and also detected homozygous segments within
each individual by using GERMLINE with the parameters *-bits 75 -err_hom 2 -err_het*
0 -min_m 0.5 -homoz-only.

Haplotype sharing was assessed by analyzing phased genotypes for each mutation
group. At each SNP, we counted the number of identical genotypes for each allele
and calculated the fraction by dividing by the total number of possible haplotypes (2
times the number of individuals), then took the larger value of the two possible
alleles (thus the fraction range was 0.5-1). We then averaged these values over
blocks of 10 or 25 SNPs and plotted the averages around the relevant mutation site.

582
<u>Between-Group IBD Calculations:</u>

We determined IBD using GERMLINE as above. We collapsed individuals into
respective groups and normalized for between-group IBD by dividing all IBD from
each group by $\left\{ \binom{2n}{2} \right\}$ where n is the number of individuals in each group. We
normalized for within-group IBD as described above. We defined groups with high
shared IBD as those with an IBD score greater than three times the founder event
strength of CEU (and ~1/3 the event strength of Ashkenazi Jews).

591
<u>f_3-statistics:</u>

We used the f_3-statistic[7] $f_3(Test; Ref_1, Ref_2)$ to determine if there was evidence that
the *Test* group was derived from admixture of groups related to Ref_1 and Ref_2. A
significantly negative statistic provides unambiguous evidence of mixture in the
Test group. We determined the significance of the f_3-statistic using a Block Jackknife
and a block size of 5 cM. We considered statistics over 3 standard errors below zero
to be significant.
599

600 **Calculating Group Specific Drift:**
601

We used ADMIXTUREGRAPH[7] to model each Indian group on the cline as a mixture
of ANI and ASI ancestry, using the model (YRI, (Indian group, (Georgians, ANI)),
[(ASI, Onge])) proposed by Moorjani *et al.*[8] This approach provides estimates for
post-admixture drift in each group (Supplementary Figure 6), which is reflective of
the strength of the founder event (high drift values imply stronger founder events).
We only included groups on the Indian cline in this analysis, and we removed all
groups with evidence of East Asian related admixture because this admixture is not
accommodated within the above model.
610

611 **PCA-Normalized F_{ST} Calculations:**
612

To account for intermarriage across groups, we used clusters based on PCA to
estimate the minimum F_{ST} for each South Asian group (Supplementary Figure 6).
Specifically, we computed the F_{ST} between each group and the rest of the individuals
in their respective cluster based on EIGENSOFT *smartpca*. For these analyses we
only included groups on the Indian Cline and those with Austroasiatic-related
genetic patterns (groups clustering near Austroasiatic speakers on the PCA). For
Ashkenazi Jews and Finns, we used the minimum F_{ST} to other European groups.
620

621 **Code Availability:**
622

623 Code for all calculations available upon request.