The importance of litter quality for the role of arbuscular mycorrhizal

fungi in plant litter decomposition (short communication)

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Abstract

Arbuscular mycorrhizal fungi (AMF) have been shown to positively and negatively affect plant litter

decomposition. The use of litter types with different quality and different observation periods might be

responsible for these contradictory results. Therefore, we performed a 10-week laboratory experiment

with 7 litter types differing in their C:N ratio, and tested for effects of litter quality and the presence of

AMF on litter decomposition. We found that decomposition of plant litter with higher C:N ratios was only

beginning and was stimulated by AMF, whereas decomposition of plant litter with lower C:N ratios had

already progressed and was decreased by AMF. With this study we show that not only litter quality is

important for effects of AMF on litter decomposition, but also the stage of litter decomposition.

Keywords: Litter decomposition; Arbuscular mycorrhizal fungi; Litter quality

More than half of the earth's carbon (C) is stored in terrestrial ecosystems (Le Quéré et al., 2015). Our knowledge of the processes of soil C cycling and potential stabilization of soil carbon is crucial for understanding and predicting global warming (Jobbagy and Jackson, 2000). In this context there is increasing interest in belowground ecosystem processes such as the decomposition of plant litter, which can be strongly influenced by the soil microbial community composition (Averill et al., 2014; Cheng et al., 2012; Cotrufo et al., 2013; Fernandez and Kennedy, 2016).

Arbuscular mycorrhizal fungi (AMF) are widespread organisms that can associate with up to 80 % of the land plant species (Smith and Read, 2008). They have been shown to positively and negatively affect plant litter decomposition: Several studies observed an increase in (easy to decompose) litter decomposition in the presence of AMF, especially under conditions of elevated CO₂ and nitrogen (N) concentrations (Cheng et al., 2012; Hodge et al., 2001; Koller et al., 2013). The main hypothesis arising from these experiments was that AMF promoted litter decomposition by stimulating the activity of other soil biota, e.g. bacteria and protozoa. However, AMF have also been shown to reduce the decomposition of woody plant litter and of litter in later stages of decomposition, respectively (Leifheit et al., 2015; Verbruggen et al., 2016). Interestingly, studies that observed increased decomposition in the presence of AMF predominantly used easy to decompose materials ('high quality' litter or finely ground litter) and studied short-term decomposition, while studies that observed decreasing decomposition in the presence of AMF used either difficult to decompose materials ('low quality' litter) or observed longer-term decomposition. Hence, litter quality and the decomposition stage are of crucial importance when studying AMF effects on litter decomposition. In AMF dominated ecosystems both low and high quality litter can be important for nutrient cycling and for the contribution to soil OM, e.g. in AMF-dominated forests and in the case of woody plants and perennials that develop lignified roots in grasslands.

To test the effect of AMF on the decomposition of litter types with different quality we performed a 10-week laboratory experiment with 7 different litter types ranging from herbaceous leaf material to woody fragments. As this was a short-term study, we expected that AMF have a stimulating effect on the decomposition of the added plant litter, especially for the 'high quality' substrates.

We used 6 different plant litter types from an experimental field site in Berlin (52° 46′ 71N, 13° 29′ 96E, Germany) and commercially available wood sticks (Meyer & Weigand GmbH, Germany). The OM was analyzed for total C and N concentration and subsequently categorized into three groups of C:N ratios: herbaceous plant leaves were categorized as 'low' (C:N ratios 10 - 22), leaves of woody plants were categorized as 'medium' (35 – 70) and twigs/pieces of woody plants as 'high' (121 - 430) (see Table 1).

Table 1: Plant litter types used in the experiment

Plant litter type	Plant part	C:N ratio	C:N ratio group used in statistical analysis
Taraxacum officinale	Leaves	10	Low
Plantago lanceolata	Leaves	22	Low
Ligustrum vulgare	Leaves	35	Medium
Mespilus germanica	Leaves	50	Medium
Acer campestre	Leaves	70	Medium
Symphoricarpos albus	Twigs	121	High
Tilia sp.	Stem pieces	430	High

150 mg of chopped litter were enclosed in a mesh bag (30 µm pore size) and buried in a conetainer with 375 g of loamy sand, originating from the same field site as the plant litter. For the AMF treatment *Plantago lanceolata* seedlings were inoculated by dipping the roots into a (premixed) powder of a rock flour material (attapulgit clay based powder) containing sterile spores of *Rhizophagus irregularis* (15500 spores g⁻¹; Symplanta GmbH & Co. KG) and grown for 10 weeks. Roots of control plants received blank carrier material. Each treatment was replicated 6 times, giving a total number of 84 pots. At harvest plant shoots and mesh bags were removed and dried at 40°C. Plant litter was removed from the mesh bags, cleaned with a brush and the weight loss determined gravimetrically. We analyzed the weight loss of the plant litter using linear regression models and ANOVA with AMF and the C:N ratio group as factors and tested for their interaction in the software R version 3.0.2 (R Core Team, 2013).

Successful root colonization of AM fungal hyphae was confirmed with staining and microscopic magnification using the gridline intersect method (McGonigle et al., 1990; Vierheilig et al., 1998). Hyphal length in the AMF treatments was increased by an average of 1.64 m g⁻¹ soil compared to the control (staining and analysis according to Jakobsen et al. (1992)).

The C:N ratio group of the material was of crucial importance for decomposition, and effects were significantly influenced by the presence of AM fungal hyphae (see Fig. 1 and Table 2). We could confirm these findings with an additional ANOVA for the effect of AMF and the C:N ratio as continuous variable (p < 0.0001 for C:N ratio and p < 0.03 for the interaction term of AMF and C:N ratio). Within the C:N ratio groups plant litter with 'low' and 'medium' C:N ratios generally had higher weight loss, but weight loss was reduced in the presence of AMF compared to OM with 'high' C:N ratios, where weight loss was increased in the presence of AMF (suggested by the results of the linear model regression). We did not find the expected effect of AMF stimulating the decomposition of 'high quality' litter. However, the overall

weight loss in the 'low' and 'medium' group was between 52 – 77 % and we therefore assume that these litters already were in a later stage of decomposition. In later stages of decomposition competition for nutrients or easily accessible energy sources can be increased and via this mechanism AMF might have been able to decrease the activity of decomposing organisms and thus reduced decomposition of litters with 'low' and 'medium' C:N ratio ('high quality' litters). Accordingly, litters with 'high' C:N ratios probably still were in an earlier stage of decomposition at the end of the experiment, given that their weight loss was only 25.8 % on average. Here, C sources and easily available nutrients might not yet have been limiting and AMF could have stimulated the activity of other microorganisms through various mechanisms (see Leifheit et al., 2015) and thus increased the decomposition of 'low quality' litter.

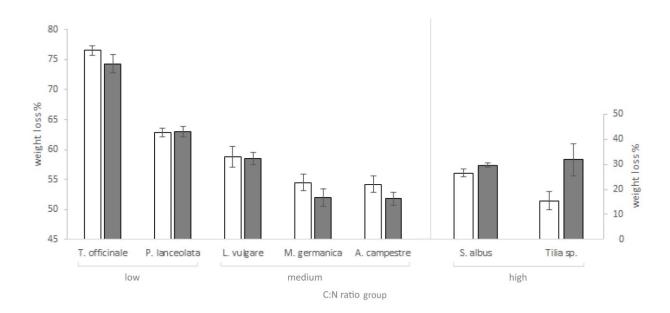


Fig. 1: Weight loss (%) of the added plant litter for the different C:N ratio groups (low = 10-22; medium = 35-70; high = 121-430); open bars = without AMF, grey bars = with AMF

Table 2: Results of the ANOVA for the presence/absence of AMF and the effect of the C:N ratio groups ('low', 'medium' and 'high', see Experiment design)

	p-value
AMF	0.77832
C:N ratio group	< 0.0001
AMF:C:N ratio group	< 0.01

However, higher litter mass loss or initial faster decomposition in the presence of AMF does not necessarily lead to higher soil C losses. The fate of C in the soil depends – amongst others - on the carbon use efficiency, i.e. the amount of metabolized litter-carbon that is fixed within microbial products in the soil matrix, which could lead to C accumulation in the long-term (Cotrufo et al., 2013; Verbruggen et al., 2013). Furthermore, AMF can contribute to soil C storage through the input of plant photosynthates in combination with stabilization of C within soil aggregates (Rillig and Mummey, 2006; Six et al., 2004; Talbot et al., 2008; Wilson et al., 2009).

With this study we showed that litter quality and the decomposition stage of plant litter is of crucial importance when studying effects of AMF on litter decomposition. Our results confirm previous findings in the literature that AMF can stimulate decomposition in early stages of plant litter decomposition and that they can diminish decomposition in later stages of plant litter decomposition. For future research on AMF effects on litter decomposition it is essential to study the fate of C in the soil, for instance by using labeling techniques.

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References

Averill, C., Turner, B.L., Finzi, A.C., 2014. Mycorrhiza-mediated competition between plants and decomposers drives soil carbon storage. Nature 505, 543-545.

Cheng, L., Booker, F.L., Tu, C., Burkey, K.O., Zhou, L., Shew, H.D., Rufty, T.W., Hu, S., 2012. Arbuscular mycorrhizal fungi increase organic carbon decomposition under elevated CO2. Science 337, 1084-1087.

Cotrufo, M.F., Wallenstein, M.D., Boot, C.M., Denef, K., Paul, E., 2013. The Microbial Efficiency-Matrix Stabilization (MEMS) framework integrates plant litter decomposition with soil organic matter stabilization: do labile plant inputs form stable soil organic matter? Global Change Biology 19, 988-995.

Fernandez, C.W., Kennedy, P.G., 2016. Revisiting the 'Gadgil effect': do interguild fungal interactions control carbon cycling in forest soils? New Phytologist 209, 1382-1394.

Hodge, A., Campbell, C.D., Fitter, A.H., 2001. An arbuscular mycorrhizal fungus accelerates decomposition and acquires nitrogen directly from organic material. Nature 413, 297-299.

Jakobsen, I., Abbott, L.K., Robson, A.D., 1992. External hyphae of Vesicular-Arbuscular Mycorrhizal Fungi associated with Trifolium subterraneum L. 1. Spread of hyphae and Phosphorus inflow into roots. New Phytologist 120, 371-380.

Jobbagy, E.G., Jackson, R.B., 2000. The vertical distribution of soil organic carbon and its relation to climate and vegetation. Ecological Applications 10, 423-436.

Koller, R., Rodriguez, A., Robin, C., Scheu, S., Bonkowski, M., 2013. Protozoa enhance foraging efficiency of arbuscular mycorrhizal fungi for mineral nitrogen from organic matter in soil to the benefit of host plants. New Phytologist 199, 203-211.

Le Quéré, C., Moriarty, R., Andrew, R.M., Peters, G.P., Ciais, P., Friedlingstein, P., Jones, S.D., Sitch, S., Tans, P., Arneth, A., Boden, T.A., Bopp, L., Bozec, Y., Canadell, J.G., Chini, L.P., Chevallier, F., Cosca, C.E., Harris, I., Hoppema, M., Houghton, R.A., House, J.I., Jain, A.K., Johannessen, T., Kato, E., Keeling, R.F., Kitidis, V., Klein Goldewijk, K., Koven, C., Landa, C.S., Landschuetzer, P., Lenton, A., Lima, I.D., Marland, G., Mathis, J.T., Metzl, N., Nojiri, Y., Olsen, A., Ono, T., Peng, S., Peters, W., Pfeil, B., Poulter, B., Raupach, M.R., Regnier, P., Roedenbeck, C., Saito, S., Salisbury, J.E., Schuster, U., Schwinger, J., Seferian, R., Segschneider, J., Steinhoff, T., Stocker, B.D., Sutton, A.J., Takahashi, T., Tilbrook, B., van der Werf, G.R., Viovy, N., Wang, Y.P., Wanninkhof, R., Wiltshire, A., Zeng, N., 2015. Global carbon budget 2014. Earth Syst. Sci. Data 7, 47-85.

Leifheit, E.F., Verbruggen, E., Rillig, M.C., 2015. Arbuscular mycorrhizal fungi reduce decomposition of woody plant litter while increasing soil aggregation. Soil Biology & Biochemistry 81, 323-328.

McGonigle, T.P., Miller, M.H., Evans, D.G., Fairchild, G.L., Swan, J.A., 1990. A new method which gives an objective measure of colonization of roots by vesicular-arbuscular mycorrhizal fungi. New Phytologist 115, 495-501.

R Core Team, 2013. R statistical functions, Part of R 3.0.2.

Rillig, M.C., Mummey, D.L., 2006. Mycorrhizas and soil structure. New Phytologist 171, 41-53.

Six, J., Bossuyt, H., Degryze, S., Denef, K., 2004. A history of research on the link between (micro)aggregates, soil biota, and soil organic matter dynamics. Soil & Tillage Research 79, 7-31.

Smith, S.E., Read, D.J., 2008. Mycorrhizal Symbiosis, 2nd ed. Academic Press, San Diego and London, 605 pp.

Talbot, J.M., Allison, S.D., Treseder, K.K., 2008. Decomposers in disguise: mycorrhizal fungi as regulators of soil C dynamics in ecosystems under global change. Functional Ecology 22, 955-963.

Verbruggen, E., Jansa, J., Hammer, E.C., Rillig, M.C., 2016. Do arbuscular mycorrhizal fungi stabilize litter-derived carbon in soil? Journal of Ecology 104, 261-269.

Verbruggen, E., Veresoglou, S.D., Anderson, I.C., Caruso, T., Hammer, E.C., Kohler, J., Rillig, M.C., 2013. Arbuscular mycorrhizal fungi - short-term liability but long-term benefits for soil carbon storage? New Phytologist 197, 366-368.

Vierheilig, H., Coughlan, A.P., Wyss, U., Piche, Y., 1998. Ink and vinegar, a simple staining technique for arbuscular-mycorrhizal fungi. Applied and Environmental Microbiology 64, 5004-5007.

Wilson, G.W.T., Rice, C.W., Rillig, M.C., Springer, A., Hartnett, D.C., 2009. Soil aggregation and carbon sequestration are tightly correlated with the abundance of arbuscular mycorrhizal fungi: results from long-term field experiments. Ecology Letters 12, 452-461.