Benzoate and Salicylate Tolerant Strains Lose Antibiotic Resistance during Laboratory Evolution of *Escherichia coli* K-12

by

Kaitlin E. Creamer,[1*] Frederick Ditmars,[1*] Preston Basting,[1*] Santiago Acero,[1] Karina Kunka,[1] Issam Hamdallah,[1] Sean P. Bush,[1] Zachary Scott,[1] Amanda He,[1] Stephanie Penix,[1] Alexandra Gonzales,[1] Elizabeth K. Eder,[1] Dominic Camperchioli,[1] Adama Berndt,[1] Michelle W. Clark,[1] Kerry Rouhier,[2] and Joan L. Slonczewski[1]

Corresponding Author:

Joan L. Slonczewski

Robert A. Oden, Jr. Professor of Biology

Higley Hall, 202 N. College Road

Kenyon College

Gambier, OH 43022

http://biology.kenyon.edu/slonc/slonc.htm

slonczewski@kenyon.edu

Phone: 740-427-5397

Affiliations:

1. Department of Biology, Kenyon College, Gambier, OH, USA

2. Department of Chemistry, Kenyon College, Gambier, OH, USA

*These authors contributed equally to the work.

ABSTRACT

Escherichia coli K-12 W3110 grows in the presence of membrane-permeant organic acids that can depress cytoplasmic pH and accumulate in the cytoplasm. We conducted laboratory evolution by daily dilution in increasing concentrations of benzoic acid (from 5 to 20 mM) buffered at external pH 6.5, a pH at which permeant acids concentrate in the cytoplasm. By 2,000 generations, clones isolated from the evolving populations showed change in phenotype from benzoate-sensitive to benzoate-tolerant but sensitive to chloramphenicol and tetracycline. Sixteen clones isolated at 2,000 generations grew to stationary phase in 20 mM benzoate, whereas the ancestral strain W3110 peaked and declined. Similar growth profiles were seen in 10 mM salicylate. The strains showed growth profiles indistinguishable from W3110 in the absence of benzoate; in media buffered at pH 4.8, pH 7.0, or pH 9.0; or in 20 mM acetate or sorbate at pH 6.5. The genomes of 16 strains revealed over 100 mutations including SNPs, large deletions, and insertion sequence knockouts. Most strains acquired deletions in the benzoate-induced multiple antibiotic resistance (Mar) regulon or associated regulators such as *rob* and *cpx*, as well as MDR efflux pumps *emrA*, *emrY*, and *mdtA*. Strains also lost or down-regulated the Gad acid fitness regulon. In 5 mM benzoate, or in 2 mM salicylate, most strains showed increased sensitivity to the antibiotic chloramphenicol, some more sensitive than a *marA* knockout. Thus, the benzoate-evolved strains may reveal additional unknown drug resistance components.

IMPORTANCE

Benzoate is a common food preservative, and salicylate is the primary active metabolite of aspirin. In the gut microbiome, genetic adaptation to salicylate may involve

loss or downregulation of inducible multidrug resistance systems. This discovery implies that aspirin therapy may modulate the human gut microbiome to favor salicylate tolerance at the expense of drug resistance.

INTRODUCTION

Pathogenic and commensal enteric bacteria maintain cytoplasmic pH homeostasis in the face of extreme external acid (pH 2-4 in the stomach) and the high concentrations of membrane-permeant organic acids in the colon (70-140 mM) (1–6). Many studies have focused on the response and recovery of *E. coli* to external pH stress (1–5), while relatively few studies have focused on the genetic response to membrane-permeant organic acids (permeant acids) (7–9) despite the importance of permeant acids as food preservatives (10). Permeant acids depress cytoplasmic pH and decrease the proton motive force (PMF), while their anion accumulates in the cytoplasm (5, 11, 12). Permeant acids include aromatic molecules such as benzoic acid (benzoate), a food preservative found in soft drinks and acidic foods (13). The related molecule salicylic acid (salicylate) is a plant defense regulator (14, 15) as well as the primary active metabolite of acetylsalicylate (aspirin) (16–18). Salicylates enter the human diet from fruits and vegetables, leading to circulating plasma levels as high as 0.1-0.2 µM (19). Aspirin therapy for cardio protection and other metabolic conditions (20, 21) may generate plasma levels of 0.2-0.5 mM (17, 22, 23). Yet despite the important metabolic effects of aspirin, salicylate and benzoate on plants and animals, there is surprisingly little research on their effects on the host microbiomes. In one study, aspirin inhibits the growth of *Helicobacter pylori* and enhances the pathogen's sensitivity to antibiotics (24).

Aromatic permeant acids such as salicylate and benzoate induce a large number of low-level multidrug efflux systems, governed by the Mar operon (*marRAB*) as well as additional unidentified mechanisms (25). Thus, in natural environments, aromatic acids may serve as early warning systems for the presence of antibiotic-producing competitors.

Benzoate and salicylate upregulate numerous genes of commensals and pathogens (26–29) including *acrAB*, *tolC*, and transport complexes that expel drugs across both the cytoplasmic and outer membrane. Genomic evidence indicates widespread existence of Mar-family systems in bacteria (30).

In *E. coli*, MarR represses expression of *marRAB*; repression is relieved when MarR binds salicylate (31) or one of several less potent inducers such as benzoate or 2,4-dinitrophenol. The upregulated MarA is an AraC-type global regulator that differentially regulates approximately 60 genes (28, 32). Another AraC-type regulator, Rob, activates *marRAB* (27, 33). MarA downregulates the acid-inducible Gad acid fitness island (34); Gad includes glutamate decarboxylase (*gadA*) for extreme-acid survival (33–35), as well as periplasmic chaperones *hdeA* and *hdeB* (36), and MDR loci *mdtE*, *mdtF* (37). Besides Mar, short-term benzoate exposure up-regulate amino-acid decarboxylases (*cadA*, *adiY*, *gadA*), succinate dehydrogenase (*sdhABCD*), biofilm-associated genes (*bdm, gatAB, and ymgABC*), and the Gad, Fur and Rcs regulons (38).

Given the high energy cost of aromatic acid-inducible gene expression and PMF consumption by efflux pumps, *E. coli* bacteria incur a tradeoff between inducible Mar drug resistance and the toxicity of the drugs involved (39). One would expect a high selective pressure for regulator alleles that shift expression based on environmental factors. In fact, selection screens based on *lac* fusions readily pick up mutations in *marR* and in MarR-regulated genes (8, 12). Selective growth under antibiotic pressure leads to upregulation of *marRAB* (40).

A powerful tool for dissecting long-term response to environmental stresses is experimental evolution (41, 42). Other experimental evolution procedures with *E. coli* have

included the adaption to high temperatures (43), freeze-thaw cycles (44), high ethanol concentrations (45), and extreme acid survival (46, 47). We developed a microplate dilution cycle in order to generate evolving populations buffered at low pH (48). The advantage of our microplate dilution cycle is that we propagate a number of populations directly in the microplate, eliminating the intermediate stage of culture in flasks or tubes. Compared to flasks, the semiaerobic condition of the closed plate more closely resembles that of the enteric habitat.

For the present study, we conducted an experimental evolution of *E. coli* K-12 W3110 in microplate well populations containing buffered at pH 6.5 and supplemented with increasing concentrations of benzoate (from 5 mM initially to 20 mM at 2,000 generations). We sequenced genomes of selected isolates, then identified genetic variants using the *breseq* pipeline (48–50). The *breseq* pipeline assembles a reference-based alignment to predict mutations compared to a previously sequenced genome (NCBI GenBank accession number NC_007779.1, *E. coli* K-12 W3110). *breseq* is now able to predict structural variations including large deletions, mobile element insertions, and gene duplications—all of which account for much of the genetic diversity in evolved clones (49–51).

Our analysis unexpectedly shows that genetic adaptation to benzoate is associated with a loss of benzoate- and salicylate-inducible genes including those that encode multidrug resistance systems. The results have implications for evolution of the gut microbiome during aspirin therapy.

MATERIALS AND METHODS

Bacterial strains and media. *Escherichia coli* K-12 W3110 (52) was the ancestral strain of all benzoate-adapted populations. Additional strains derived from *E. coli* K-12 W3110 were isolated during the course of the evolution experiment. Strains are listed in Supplemental File 1 (Table S1).

Bacteria were cultured in LBK (10 g/L tryptone, 5 g/L yeast extract, 7.45 g/L KCl) (53). Culture media were buffered with either 100mM piperazine-N,N'-bis(ethanesulfonic acid) (PIPES; pKa= 6.8), 100 mM, 2-(N-morpholino)ethanesulfonic acid (MES; pKa= 5.96), 100 mM 3-morpholinopropane-1-sulfonic acid (MOPS; pKa= 7.20), 100 mM homopiperazine-N,N'-bis-2-(ethanesulfonic acid) (HOMOPIPES; pKa = 4.55, 8.12), or 150 mM N-Tris(hydroxymethyl)methyl-3-aminopropanesulfonic acid (TAPS; pKa= 8.4). The pH of the medium was adjusted as necessary with either 5 M HCl or 5 M KOH. Potassium benzoate (referred to as benzoate), sodium salicylate (salicylate), potassium acetate, potassium sorbate, chloramphenicol, tetracycline or streptomycin was added before filter sterilization for LBK media requiring various concentrations of acids or antibiotics. Temperature of incubation was 37°C unless noted otherwise.

Experimental Evolution. Experimental evolution was conducted according to the procedure of our low-pH laboratory evolution experiment (48) with modifications. Briefly, 24 cultures derived from the same ancestral strain (W3110, freezer stock D13) were cultured continuously in increasing concentrations of benzoate for 2,000 generations (Supplemental File 2, Figure S1). An overnight culture of ancestral *Escherichia coli* K-12 W3110 was diluted 1:100 in LBK, pH 6.5 100 mM PIPES, 5 mM potassium benzoate. Growth was

recorded over 22 h in a SpectraMax Plus384 MicroPlate reader (Molecular Devices). Every 15 minutes, the microplate was shaken for 3 seconds and the OD_{450} of each culture was recorded. The cultures were re-diluted 1:100 into fresh benzoate growth medium at the end of the daily cycle. 100 µl glycerol (50% glycerol, 100 mM MES pH 6.5) was added to each well, after which the microplate was frozen at -80°F (48). After 60 generations, the concentration of potassium benzoate was increased to 6 mM; 10 mM after 90 generations; 12 mM after 540 generations; 15 mM after 1,020 generations; 18 mM after 1,210 generations; 20 mM after 1,580 generations to the conclusion of the experiment with a cumulative 3,000 generations of growth. If the strains had to be restarted from a frozen microplate, the frozen cultures were thawed and diluted 1:50 into fresh potassium benzoate growth media.

A generation rate of the exposed cells was calculated based on the 1:100 daily dilution, resulting in a 100-fold daily growth to achieve 6.64 generations of binary-fission (54). In the course of the 22-hour cycle, all bacterial populations attained stationary phase densities. After 2,000 generations, eight clones with increased growth rate in 20 mM benzoate were chosen for genome sequencing. An additional eight clones were chosen based on sensitivity to chloramphenicol (8 µg/ml). Microplates were taken from the freezer and samples from specific wells were spread on LBK agar plates. Two clones from each chosen well were streaked three times and stored as freezer stocks (Table S1; Table S2).

Growth assays. Growth curves were measured in the microplate reader at 37°C for 22 hours under various conditions of organic acids, pH, and antibiotics. Strains were cultured overnight in LBK pH 5.5 buffered with 100 mM MES; LBK pH 6.5 buffered with 100 mM PIPES; LBK pH 7.0 buffered with 100 mM MOPS; or LBK pH 8.5 buffered with 150 mM TAPS. Supplements included benzoate, salicylate, acetate, or sorbate, as stated in figures.

Overnight cultures were diluted 1:100 (1:200 for the antibiotic growth assays) into the exposure media which included LBK pH 6.5 buffered with 100 m M PIPES; LBK pH 4.8, 100 mM HOMOPIPES; LBK pH 7.0, 100 mM MOPS; LBK pH 9.0, 150 mM TAPS; or LBK pH 7.0, 100 mM MOPS. Every 15 minutes, the plate was shaken for 3 seconds and an OD_{600} measurement was recorded. The growth rate *k* of each culture was calculated over the period of 1-3 h, approximately the log phase of growth (35). The cell density *E* of each culture was measured at 16 h unless stated otherwise.

Genomic DNA extraction and sequencing. Genomic DNA from benzoate-evolved clones and from the ancestral wild type strain W3110 (freezer stock D13) was extracted using the DNeasy DNA extraction kit (Qiagen) and the MasterPure Complete DNA and RNA Purification Kit (Epicentre). The DNA purity was confirmed by measuring the 260nm/280nm and 260nm/230nm absorbance ratios using a NanoDrop 2000 spectrophotometer (Thermo Fisher Scientific) and the concentration of the DNA was measured using both the NanoDrop 2000 spectrophotometer (Thermo Fisher Scientific) and a Qubit 3.0 Fluorometer (Thermo Fisher Scientific), according to manufacturer instructions.

The genomic DNA was sequenced by Michigan State University Research Technology Support Facility Genomics Core. For Illumina MiSeq sequencing, libraries were prepared using the Illumina TruSeq Nano DNA Library Preparation Kit. After library validation and quantitation, they were pooled and loaded on an Illumina MiSeq flow cell. Sequencing was done in a 2x250bp paired-end format using an Illumina 500 cycle V2 reagent cartridge. Base calling was performed by Illumina Real Time Analysis (RTA) v1.18.54 and output of RTA was demultiplexed and converted to FastQ format with Illumina Bcl2fastq v1.8.4.

Nucleotide sequence accession number. Sequence data have been deposited in the NCBI Sequence Read Archive (SRA) under accession number SRP074501.

Sequence assembly and analysis using *breseq* computational pipeline. The computational pipeline *breseq* version 0.27.1 was used to assemble and annotate the resulting reads of the evolved strains (49–51). The reads were mapped to the *E. coli* K-12 W3110 reference sequence (NCBI GenBank accession number NC_007779.1) (55). Mutations were predicted by *breseq* by comparing the sequences of the evolved isolates to that of the ancestral strain W3110, lab stock D13 (51). In order to visualize the assembly and annotations of our evolved isolate sequences mapped to the reference *E. coli* K-12 W3110 genome, we used Integrative Genomics Viewer (IGV) from the Broad Institute at MIT (56). Sequence identity of clones was confirmed by PCR amplification of selected mutations.

P1 phage transduction and strain construction. P1 phage transduction was conducted by standard procedures to replace an evolved mutated gene with the ancestral non-mutated genes, as well to create a *marA*::kanR knockout strain (48). Strains with desired deletions or functional insertions were ordered from the Keio collection (Coli Genetic Stock Center, Yale University) (57) and were introduced into the evolved strain of choice or the ancestral wild type strain. We used a *marR*::kanR strain provided by Frederick R. Blattner, University of Wisconsin-Madison. Constructs were confirmed by PCR amplification and Sanger sequencing of key alleles of donor and recipient.

MIC assays. For assays of minimum inhibitory concentration (MIC) of antibiotics, the strains were cultured in a microplate for 22 h in LBK 100 mM MOPS pH 7.0, 2 mM salicylate. The medium contained a range of antibiotic concentration (µg/ml): 0, 1, 2, 4, 6, 8, 12, 16, 24. Measurement of $OD_{600} \geq 0.05$ was defined as positive for growth. MIC was

reported as the median value of 8 replicates. For each antibiotic, three sets of 8 replicates were performed.

GABA assays. The procedure for measuring GABA production via glutamate decarboxylase was modified from that of Ref. (58). Strains were cultured overnight in LB medium (10 g/L tryptone, 5 g/L yeast extract, 100 mM NaCl) buffered with 100 mM MES pH 5.5. 10 mM glutamine was included, which the bacteria convert to glutamate, the substrate of glutamate decarboxylase (59). For anaerobic culture, closed 9-ml screwcap tubes nearly full of medium were incubated for 18 hours at 37°C. The pH of each sample was lowered with HCl to pH 2.0 for extreme-acid stress (60) and incubated 2 h with rotation. Cell density (OD_{600}) was measured in microplate wells, in a SpectraMax plate reader. 1 ml of each culture was pelleted in a microfuge. The supernatant was filtered and prepared for GC-MS by EZfaast amino acid derivatization (Phenomenex, 2005). GABA concentration was calculated using a standard solution prepared at a concentration of 200 nm/ml. GABA and other compounds from the culture fluid were identified using NIST library analysis.

RESULTS

Experimental evolution of benzoate-tolerant strains. We conducted experimental evolution of *Escherichia coli* K-12 W3110 exposed to increasing concentrations of benzoic acid, as described under Methods (**Figure S1**). Benzoate tolerance was tested over the course of the experiment. Clones were sampled from microplate populations frozen at intervals over the course of zero to 2,900 generations (**Figure 1**). The clones were cultured in microplate wells in media containing 20 mM benzoate at pH 6.5, and the endpoint cell density was measured at 16 h. Over 1,400-2,900 generations the population growth levels increased significantly compared to that of the ancestor (**Fig. 1A**). A similar increase was observed for cell density during growth with 10 mM salicylate (**Fig. 1B**). Thus overall, tolerance to benzoate and salicylate increased over generations of exposure.

Since benzoate and salicylate induce multidrug resistance via the Mar regulon (25), it was of interest to test drug resistance of the evolved clones. We measured growth in chloramphenicol, an antibiotic that is effluxed by the MarA-dependent pump AcrA-AcrB-TolC (28), which confers low-level resistance. For our experiment, the same set of clones observed for growth in benzoate and salicylate were cultured in media containing 8 µg/µl chloramphenicol (**Fig. 1C, 1D**). The media were adjusted to pH 7.0 for maximal growth and contained a low concentration of benzoate or salicylate for induction of Mar regulon. Later generations (1,900-2,900 gen) reached significantly lower cell density compared to that of the ancestor W3110.

Our results suggested that populations evolving with benzoate experienced a tradeoff between benzoate-salicylate tolerance and inducible chloramphenicol resistance. This tradeoff is confirmed by the plot of benzoate or salicylate tolerance versus growth in

chloramphenicol (**Fig. 1E, 1F**). Strain W3110 and early generation clones showed little growth in 20 mM benzoate, but growth in chloramphenicol was variable. Later generations showed increased growth in 20 mM benzoate but little if any growth in chloramphenicol. Note however that outliers appeared under all conditions, as expected under selection pressure (61).

Genome resequencing showed numerous SNPs, deletions, and IS5 insertion mutations. After 2,000 generations, eight clones showing benzoate tolerance were selected for genome sequencing (described under Methods). An additional eight clones were selected for chloramphenicol sensitivity; all were benzoate-tolerant. The 16 clones were streaked for isolation and established as strains (**Table S1**). We used the *breseq* computational pipeline to analyze the mutation predictions of our re-sequenced genomes compared to the reference *E. coli* W3110 reference genome (55). More than 100 mutations were detected across the sixteen sequenced genomes (**Table S2**). Mutations found in our resequenced ancestor W3110 (lab stock D13, **Table S3**) were filtered from the results. The types of mutations that accumulated across the sixteen strains included SNPs, small indels, insertion sequences (IS), and large IS-mediated deletions. Both coding changes and intergenic mutations were frequent. A large number of insertion knockouts were mediated by IS5 (62) or other mobile insertion sequence elements. An example, *gadX*::IS5 found in clone A5-1 is shown in **Fig. 2**. The inserted sequence, including *insH* plus IS5 flanking regions, was identical to those of 11 known IS5 inserts in the standard W3110 sequence; there was also a four-base duplication of the target site. Insertion sequence mobility is a major source of evolutionary change in *E. coli* (63).

The 2,000-generation benzoate-adapted strains were grouped in six clades based on shared mutations (**Table S2**); a representative member of each clade is shown in **Table 1**. Some of the shared mutations originated within a population, as in the case of the two strains taken from the A5, E1, and C3 populations. Other shared mutations could have originated in the shared founder culture, or from inadvertent cross-transfer between microplate wells. One population (G5) included a strain G5-2 that shares mutation with the strains from the H1, H3 and G3 populations while sharing no mutations with strain G5-1, from the G5 population. Evolving populations commonly show diverse independent genetic adaptations to a common stress condition (61).

Mutations appeared in Mar and other multidrug efflux systems. Five of the six clades showed mutations affecting the Mar regulon, as well as other MDR genes (**Table 1**). Strain A5-1 had a 6,115-bp deletion including *marRAB* (*ydeA*, *marRAB*, *eamA*, *ydeEH*). Regulators of Mar showed point mutations in strain G5-2 (*mar* paralog *rob*, Ref. (27)) and in strain A1-1 (two-component activator *cpxA*, Ref. (64)). Mutations appeared in other multidrug efflux systems: *emrA*, *emrY* (33, 65), *mdtA*, deletions covering *mdtEF* (65), and *yeaS* (*leuE*) leucine export (66).

The benzoate-evolved strains were tested by MIC assay for sensitivity to antibiotics chloramphenicol and tetracycline (**Table 2**). The assay medium included 2 mM salicylate for inducible resistance, such as MarA-activated resistance to chloramphenicol. For chloramphenicol, strains A1-1, A5-1, C3-1, and G5-2 showed MIC levels half that of ancestor W3110. These lowered MIC levels were comparable to that of a *marA* mutant, and only E1-1 showed chloramphenicol resistance equivalent to that of W3110. For tetracycline, our *marA* knockout strain showed no loss of resistance; this may be due to induction of non-

Mar salicylate-dependent resistance (25). Nevertheless, tetracycline resistance was decreased for all of our benzoate-evolved strains with the exception of strain E1-1.

Mutations appeared in Gad acid resistance, RNAP, and fimbriae. Strikingly, five of the six clades of the 16 sequenced strains showed a mutation in the Gad acid resistance regulon (36), which undergoes regulation intertwined with that of MDR systems. Strain E1-1 had a 14,000-bp deletion mediated by the insertion sequence *insH* flanking the *gad* gene region (*gadXW*, *mdtFE*, *gadE*, *hdeDAB*, *yhiDF*, *slp*, *insH*, *yhiS*). Similarly, strain A1-1 showed a 10,738-bp deletion covering most of the *gad* region. A1-1 also had an insertion in the *ariR* (*ymgB*) biofilm-dependent activator of Gad (38, 67). The *mdtFE* genes encode components of the efflux pump MdtF-MdeE-TolC, which confers resistance to chloramphenicol, as well as fluoroquinolones and other drugs (68). Thus, *gad* deletion might explain chloramphenicol sensitivity of strain A1-1; but not E1-1, which was chloramphenicol resistant despite the deletion.

Other strains showed mutations in the *gadX* activator: A5-1 (IS5 insertion), G5-1 (missense L199F), and G5-2 (78-bp deletion). G5-1 also showed an *hfq* point substitution, at a position known to affect function of RpoS (69), which activates Gad (2). The *gad* mutants showed different levels of GABA production by glutamate decarboxylase (GadA) during extreme-acid exposure (incubation at pH 2) (**Fig. 3**). The two strains with full Gad deletions (A1-1 and E1-1) produced no GABA, whereas strains with *gadX* mutations (A5-1, G5-1, and G5-2) produced significantly less GABA than did the ancestor W3110 (Friedman/Conover test). Only one representative strain, C3-1, showed no Gad-related mutation; this strain produced GABA in amounts comparable to that of W3110.

Each benzoate-adapted strain also showed a mutation in an RNAP subunit (*rpoB*, *rpoA*), a sigma factor (*rpoD*, *rpoS*) or an RNAP-associated helicase (*hepA*). These mutations in the transcription apparatus are comparable to those we find under low-pH evolution (48). Four of the six clades had mutations in fimbria subunit *fimA* or in regulators *fimB*, *fimE*. Thus, benzoate exposure could select for loss of fimbriae synthesis. Other interesting mutations affected cell division (*ftsZ*), cell wall biosynthesis (*mrdA*), and envelope functions (*ecpD*, *lptD*, *ybbP*, *yejM*, *yfhM*, *yqiGH*, and *rfaY*). The envelope mutations suggest responses to benzoate effects on the outer membrane and periplasm.

Benzoate-evolved strains showed increased growth rate and stationary phase cell density. We investigated the phases of growth for each benzoate-evolved strain, in order to characterize the focus of selection pressure with respect to early growth rate, stationary-phase cell density, and death phase. Observing the entire growth curve provides more information than an endpoint MIC. Growth curves were conducted in microplate wells for each of the six representative 2,000-generation benzoate-adapted strains. For each strain, eight replicate wells of the microplate were inoculated alongside eight replicate wells of strain W3110, as seen in the example for strain G5-2 (**Fig. 4A**).

In the example shown, strain G5-2 maintained log-phase growth for more than five hours in the presence of 20 mM benzoate (0.42±0.5 gen/h, measured over times 1-3 h). Strain G5-2 eventually reached a stationary-phase OD_{600} of approximately 1.0. By contrast, ancestral strain W3110 grew more slowly (0.18±0.01 gen/h) and peaked at OD_{600}=0.5-0.7 by about 8 h. After 8 h, W3110 entered a death phase as the cell density declined. The presence of chloramphenicol, however, reversed the relative fitness of the two strains (**Fig. 4B**). The benzoate-evolved strain barely grew, and 7 of 8 replicates entered death phase by 3-4 h. By

contrast, the ancestor grew steadily to an OD_{600} of 0.5-0.6, a level that was sustained for several hours.

The effect of various permeant acids was tested, in order to determine the specificity of acid tolerance (**Fig. 5**). For all growth curves, statistical comparison was performed using cell density values at 16 h. Each panel shows a curve with median cell density (at 16 h) for a benzoate-evolved strain, as well as for strain W3110. Both benzoate and salicylate conditions showed a marked fitness advantage for all six benzoate-evolved strains (**Fig. 5A, B**). Five of the strains showed log-phase growth rates equivalent to each other, whereas G5-1 grew significantly more slowly (Friedman, Conover tests; $p \leq 0.05$). All benzoate-evolved strains grew faster than strain W3110. All six benzoate-evolved strains reached equivalent plateau cell densities (OD_{600} values of approximately 1.0, with 20 mM benzoate; 0.9, with 10 mM salicylate).

In the presence of aliphatic acids acetate or sorbate (**Fig. 5B, C**) no significant difference was seen between growth of the ancestral strain W3110 and that of the benzoate-evolved strains. Thus, the evolved fitness advantage is unlikely to result from cytoplasmic pH depression but appears specific to the presence of aromatic acids benzoate or salicylate. Further testing with 40 µM carbonyl cyanide m-chlorophenyl hydrazone (the uncoupler CCCP) showed no difference in growth rate or stationary-phase cell density among the strains (data not shown). Thus, while decrease of proton motive force may be one factor it cannot be the sole cause of the fitness advantage of our strains.

We also tested whether the benzoate-evolved strains showed any fitness advantage with respect to pH stress. The strains were cultured in media buffered at pH7.0 (**Fig. 6A**) and at pH 4.8 or pH 9.0 (**Fig. S3, S4**). All of the benzoate-evolved strains grew similarly to the

ancestor at external pH values across the full range permitting growth. Thus, the fitness advantage of the evolved strains was specific to the presence of benzoate or salicylate.

Chloramphenicol inhibits growth of benzoate-evolved clones, despite benzoate fitness advantage. Since each of the six clades showed a mutation in an MDR gene or regulator (discussed above), we characterized the growth profiles of all strains in the presence of chloramphenicol (**Fig. 6BCD**). In the absence of benzoate or salicylate inducer (**Fig. 6B**), all strains showed growth curves equivalent to that of W3110 or W3110 *marA*::kanR (which lacks the MarA activator of MDR efflux). Only the *marR*::*kanR* strains (constitutive for activator *marA*) showed resistance. In the presence of benzoate (**Fig. 6C**) or salicylate (**Fig. 6D**), the various benzoate-evolved strains showed distinct degrees of resistance to chloramphenicol. (Panels presenting all replicates of each strain are presented in supplemental **Figures S3** and **S4**). Strain E1-1 consistently grew at a rate comparable to W3110, whereas strains A5-1 and G5-1 generally grew to a lower density, comparable to the *marA*::kanR strain. Strains C3-1 and G5-2 showed hypersensitivity, with cell densities significantly below that of *marA*::kanR. The sensitivity of strain C3-1 is noteworthy given the absence of Mar or Gad mutations. Another strain, G5-2, shows chloramphenicol sensitivity greater than the level that would be predicted from loss of Rob activating MarA (26). Thus, the C3-1 and G5-2 genomes may reveal defects in previously unknown benzoate-inducible MDR genes.

Reversions of *rob*, *gadX*, and *cpxA* do not affect benzoate tolerance or chloramphenicol sensitivity. We tested whether reversion of mutant alleles of Mar activator *rob* (S34P) and Gad activator *gadX* (Δ78bp) affect either the benzoate fitness or chloramphenicol sensitivity of strains G5-2 or A1-1 (**Fig. 7**). The ancestral alleles of *rob* and

of *gadX* were each moved into G5-2 by cotransduction with linked markers *yjjX790*::*kanR* and *treF774*::*kanR* respectively. Strain G5-2 constructs with either *rob*$^+$, *gadX*$^+$, or *rob*$^+$ *gadX*$^+$ showed no significant difference in benzoate tolerance compared to the parental strain G5-2 (data not shown). The constructs also showed no change in chloramphenicol sensitivity, in the presence of 5 mM benzoate (**Fig. 7**). Nevertheless, a *gadX*::*kanR* knockout in the W3110 background showed chloramphenicol sensitivity comparable to that of W3110 *marA*::*kanR*. Thus it is possible that GadX has some uncharacterized effect upon chloramphenicol efflux, outside the Mar regulon, such as activation of *mdtE*, *mdtF* (37).

We also moved the parental allele of MDR regulator *cpxA* into A1-1, replacing *cpxA*(N107S) with a marker *fdhD*::*kanR* linked to *cpxA*$^+$. No effect of *cpxA*$^+$ was seen for the A1-1 benzoate tolerance and chloramphenicol sensitivity (data not shown). Thus, strain A1-1 may contain previously unknown MDR genes.

DISCUSSION

In order to identify candidate genes for benzoate stress response, we sequenced the genomes of experimentally evolved strains (41, 42, 47, 48, 70). We observed over 100 distinct mutations fixed in the sequenced isolates after 2,000 generations, including a surprising number of knockout alleles due to mobile elements (62, 63) such as the IS5 insert of *ariR* (38). The *E. coli* genome contains many IS-elements, including eight copies of IS1, five copies of IS2, and copies of other less well-studied IS types where the most prevalent are IS1 and IS5 (55, 71, 72). Transposition of IS5 may be induced by environmental factors such as motility conditions, which induce IS5 insertion upstream of motility regulator *flhD* (73). Nonetheless, finding an additional 33 insertion sequences under benzoate selection is remarkable. We have preliminary evidence that benzoate stress increases transcription of the IS5 *insH* transposase (J. Moore and J. L. Slonczewski, unpublished).

Benzoate exposure decreases the cell's PMF while simultaneously upregulating several regulons, including many involved in drug resistance. Our data suggests that benzoate exposure selects for genetic changes in *E. coli* that result in, over time, the loss of energetically costly systems such as Mar and other MDR regulons, as well as Gad acid-inducible extreme-acid regulon. MarA is a potent transcriptional factor in *E. coli*, upregulating numerous efflux pumps and virulence factors (8, 26, 28). The transcription and translation of so many gene products would result in a considerable energy strain on the individual cell. Furthermore, many of the products are efflux pumps that spend PMF, which is diminished by the presence of a permeant acid. The decrease of energy expense could explain the benzoate fitness advantage of strains that have broad-spectrum downregulation of Mar gene products, such as we see in A5-1 and G5-2.

A similar energy load may occur under benzoate-depressed cytoplasmic pH, where the Gad regulon is induced. Gad includes expression of numerous gene products such as glutamate decarboxylase, whose activity enhances fitness only in extreme acid (pH 2) (36) and which breaks down valuable amino acids. Thus the deletion of the *gad* region (seen in strains A1-1 and E1-1) could eliminate a fruitless energy drain. Likewise, the possible loss of fimbriae synthesis (strains A5-1, C3-1, E1-1, G5-1) could save energy.

The progressive loss of antibiotic resistance is a remarkable consequence of benzoate selection, evident as early as 1,500 generations (**Fig. 1**). Several observations point to the existence of inducible MDR systems yet to be discovered. Strains C3-1 and G5-2 show hypersensitivity to chloramphenicol, beyond the level of sensitivity seen in a *marA* knockout (**Fig. 6C, D**). Furthermore, the reversion of mutant alleles of *rob*, *gadX*, and *cpxA* do not diminish the phenotypes of the 2,000-generation strains. It is likely that these alleles conferred a fitness advantage early on (61) but have since been superseded by further mutations in as yet unidentified players in drug resistance.

The fitness tradeoff between drug resistance and benzoate/salicylate exposure has implications for the human gut microbiome. Mar and homologs such as Mex are reported in numerous bacteria, including proteobacteria and *Bacteroides fragilis* (74, 75). Salicylate is a plant defense molecule commonly obtained via human diets rich in fruits and vegetables. Aspirin is deacetylated in the liver and stomach, forming salicylic acid (76); and as a membrane-permeant acid, salicylic acid permeates human tissues. Both food-related and aspirin-derived salicylates come in contact with enteric gut bacteria, where they would be expected to activate Mar-like antibiotic resistance systems. Commonly prescribed for cardiac health, aspirin releases salicylate at plasma levels of approximately 0.2 mM (17, 20, 21, 23).

Aspirin therapy is known to prevent clotting by inactivation of cyclooxygenase, leading to suppression of prostaglandins. There is little attention, however, to the possible effects of aspirin on bacteria. Bacteria are found in arterial plaques and associated with heart attacks (77). Aspirin also prevents colon cancer, by some unknown mechanism (17, 21). Colon cancer depends on colonic bacteria and the formation of biofilms (78, 79).

Long-term salicylate exposure via aspirin therapy may select a microbiome that is salicylate-tolerant but drug-sensitive. A salicylate-adapted microbiome may confer the benefit of excluding drug-resistant pathogens that lack salicylate tolerance. In blood plasma, salicylate levels might help exclude bacteria from arterial plaques. An adverse consideration, however, is that the salicylate-adapted microbiome of the colon may be more vulnerable to high dose antibiotic therapy. These speculative possibilities should be tested in host microbial models.

Acknowledgements

We thank Zachary Blount, Jeff Barrick, Michael Harden, Rohan Maddamsetti, and Bradley Hartlaub for valuable discussions. This work was funded by grant MCB-1329815 from the National Science Foundation, and by the Kenyon College Summer Science Scholars.

REFERENCES

1. **Kanjee U**, **Houry WA**. 2013. Mechanisms of acid resistance in *Escherichia coli*. Annu Rev Microbiol **67**:65–81.

2. **Foster JW**. 2004. *Escherichia coli* acid resistance: Tales of an amateur acidophile. Nat Rev Microbiol **2**:898–907.

3. **Audia JP**, **Webb CC**, **Foster JW**. 2001. Breaking through the acid barrier: An orchestrated response to proton stress by enteric bacteria. Int J Med Microbiol **291**:97–106.

4. **Krulwich TA**, **Sachs G**, **Padan E**. 2011. Molecular aspects of bacterial pH sensing and homeostasis. Nat Rev Microbiol **9**:330–43.

5. **Slonczewski JL**, **Fujisawa M**, **Dopson M**, **Krulwich TA**. 2009. Cytoplasmic pH measurement and homeostasis in bacteria and archaea.Adv Microb Physiol.

6. **Qin J**, **Li R**, **et al.** 2010. A human gut microbial gene catalogue established by metagenomic sequencing. Nature **464**:59–67.

7. **Slonczewski JL**, **Macnab RM**, **Alger JR**, **Castle AM**. 1982. Effects of pH and repellent tactic stimuli on protein methylation levels in *Escherichia coli*. J Bacteriol **152**:384–399.

8. **Rosner JL**, **Slonczewski JL**. 1994. Dual regulation of inaA by the multiple antibiotic resistance (Mar) and superoxide (SoxRS) stress response systems of *Escherichia coli*. J Bacteriol **176**:6262–6269.

9. **Lee CH**, **Oon JSH**, **Lee KC**, **Ling MHT**. 2012. *Escherichia coli* ATCC 8739 adapts to the presence of sodium chloride, monosodium glutamate, and benzoic acid after extended culture. ISRN Microbiol **2012**.

10. **Beales N**. 2004. Adaptation of microorganisms to cold temperatures, weak acid preservatives, low pH, and osmotic stress: A review. Compr Rev Food Sci Food Saf **3**:1–20.

11. **Wilks JC**, **Slonczewski JL**. 2007. pH of the cytoplasm and periplasm of *Escherichia coli*: Rapid measurement by green fluorescent protein fluorimetry. J Bacteriol **189**:5601–5607.

12. **Slonczewski JL**, **Macnab RM**, **Alger JR**, **Castle AM**. 1982. Effects of pH and repellent tactic stimuli on protein methylation levels in Escherichia coli. J Bacteriol **152**:384–399.

13. **Lennerz B**, **Vafai SB**, **Delaney NF**, **Clish CB**, **Deik AA**, **Pierce KA**, **Ludwig D s.**, **Mootha VK**. 2015. Effects of sodium benzoate, a widely used food preservative, on glucose homeostatsis and metabolic profiles in humans. Mol Genet Metab **114**:73–79.

14. **An C**, **Mou Z**. 2011. Salicylic acid and its function in plant immunity. J Integr Plant Biol **53**:412–428.

15. **Du L**, **Ali GS**, **Simons KA**, **Hou J**, **Yang T**, **Reddy ASN**, **Poovaiah BW**. 2009. Ca2+/calmodulin regulates salicylic-acid-mediated plant immunity. Nature **457**:1154–1158.

16. **Hawley S a**, **Fullerton MD**, **Ross F a**, **Schertzer JD**, **Chevtzoff C**, **Walker KJ**, **Peggie MW**, **Zibrova D**, **Green K a**, **Mustard KJ**, **Kemp BE**, **Sakamoto K**, **Steinberg GR**, **Hardie DG**. 2012. The ancient drug salicylate directly activates AMP-activated protein kinase. Science (80-) **336**:918–922.

17. **Paterson JR**, **Lawrence JR**. 2001. Salicylic acid: a link between aspirin, diet and the prevention of colorectal cancer. QJM **94**:445–448.

18. **Paterson JR**, **Baxter G**, **Dreyer JS**, **Halket JM**, **Flynn R**, **Lawrence JR**. 2008. Salicylic acid sans aspirin in animals and man : Persistence in fasting and biosynthesis from benzoic acid. J Agric Food Chem **56**:11648–11652.

19. **Spadafranca a**, **Bertoli S**, **Fiorillo G**, **Testolin G**, **Battezzati A**. 2007. Circulating salicylic acid is related to fruit and vegetable consumption in healthy subjects. Br J Nutr **98**:802–806.

20. **Patrono C**, **García Rodríguez LA**, **Landolfi R**, **Baigent C**. 2005. Low-dose aspirin for the prevention of atherothrombosis. N Engl J Med **353**:2373–2383.

21. **Patrono C**. 2013. Low-dose aspirin in primary prevention: cardioprotection, chemoprevention, both, or neither? Eur Hear J **34**:3403–3411.

22. **Hundal RS**, **Petersen KF**, **Mayerson AB**, **Randhawa PS**, **Inzucchi S**, **Shoelson SE**, **Shulman GI**. 2002. Mechanism by which high-dose aspirin improves glucose metabolism in type 2 diabetes. J Clin Invest **109**:1321–1326.

23. **Ahmed N**, **Meek J**, **Davies GJ**. 2010. Plasma salicylate level and aspirin resistance in survivors of myocardial infarction. J Thromb Thrombolysis **29**:416–420.

24. **Wang WH**, **Wong WM**, **Dailidiene D**, **Berg DE**, **Gu Q**, **Lai KC**, **Lam SK**, **Wong BCY**. 2003. Aspirin inhibits the growth of *Helicobacter pylori* and enhances its susceptibility of antimicrobial agents. Gut **52**:490–495.

25. **Cohen SP**, **Levy SB**, **Foulds J**, **Rosner JL**. 1993. Salicylate induction of antibiotic resistance in *Escherichia coli*: Activation of the *mar* operon and a *mar*-independent pathway. J Bacteriol **175**:7856–7862.

26. **Duval V**, **Lister IM**. 2014. MarA, SoxS and Rob of *Escherichia coli* – Global regulators of multidrug resistance, virulence and stress response. Int J Biotechnol Wellness Ind **2**:101–124.

27. **Chubiz LM**, **Rao C V.** 2010. Aromatic acid metabolites of *Escherichia coli* K-12 can induce the *marRAB* operon. J Bacteriol **192**:4786–4789.

28. **Ruiz C**, **Levy SB**. 2010. Many chromosomal genes modulate MarA-mediated multidrug resistance in *Escherichia coli*. Antimicrob Agents Chemother **54**:2125–2134.

29. **Critzner FJ**, **Dsouza DH**, **Golden DA**. 2008. Transcription analysis of *stx1*, *marA*, and *eaeA* genes in *Escherichia coli* O157:H7 treated with sodium benzoate. J Food Prot **71**:1469–74.

30. **Perera IC**, **Grove A**. 2010. Molecular mechanisms of ligand-mediated attenuation of DNA binding by MarR family transcriptional regulators. J Mol Cell Biol **2**:243–254.

31. **Alekshun MN**, **Levy SB**, **Mealy TR**, **Seaton BA**, **Head JF**. 2001. The crystal structure of MarR, a regulator of multiple antibiotic resistance, at 2.3 Å resolution. Nat

Struct Mol Biol **8**:710–714.

32. **Barbosa TM**, **Levy SB**. 2000. Differential expression of over 60 chromosomal genes in *Escherichia coli* by constitutive expression of MarA. J Bacteriol **182**:3467–3474.

33. **Grkovic S**, **Brown MH**, **Skurray RA**, **Repressor A**, **Subtilis B**. 2002. Regulation of bacterial drug export systems. Microbiolgy Mol Biol Rev **66**:671–701.

34. **Ruiz C**, **McMurry LM**, **Levy SB**. 2008. Role of the multidrug resistance regulator MarA in global regulation of the *hdeAB* acid resistance operon in *Escherichia coli*. J Bacteriol **190**:1290–1297.

35. **Deininger KNW**, **Horikawa A**, **Kitko RD**, **Tatsumi R**, **Rosner JL**, **Wachi M**, **Slonczewski JL**. 2011. A requirement of tolc and MDR efflux pumps for acid adaptation and GadAB induction in *Escherichia coli*. PLoS One **6**:1–7.

36. **Mates AK**, **Sayed AK**, **Foster JW**. 2007. Products of the *Escherichia coli* acid fitness island attenuate metabolite stress at extremely low pH and mediate a cell density-dependent acid resistance. J Bacteriol **189**:2759–2768.

37. **Nishino K**, **Senda Y**, **Yamaguchi A**. 2008. The AraC-family regulator GadX enhances multidrug resistance in *Escherichia coli* by activating expression of *mdtEF* multidrug efflux genes. J Infect Chemother **14**:23–29.

38. **Kannan G**, **Wilks JC**, **Fitzgerald DM**, **Jones BD**, **Bondurant SS**, **Slonczewski JL**. 2008. Rapid acid treatment of *Escherichia coli*: transcriptomic response and recovery. BMC Microbiol **8**.

39. **Wood KB**, **Cluzel P**. 2012. Trade-offs between drug toxicity and benefit in the multi-antibiotic resistance system underlie optimal growth of *E. coli*. BMC Syst Biol **6**:48.

40. **Weiss SJ**, **Mansell TJ**, **Mortazavi P**, **Knight R**, **Gill RT**. 2016. Parallel mapping of antibiotic resistance alleles in *Escherichia coli*. PLoS One **11**:1–17.

41. **Blount ZD**, **Borland CZ**, **Lenski RE**. 2008. Historical contingency and the evolution of a key innovation in an experimental population of *Escherichia coli*. Proc Natl Acad Sci U S A **105**:7899–7906.

42. **Blount ZD**, **Barrick JE**, **Davidson CJ**, **Lenski RE**. 2012. Genomic analysis of a key innovation in an experimental *Escherichia coli* population. Nature **489**:513–518.

43. **Riehle MM**, **Bennett AF**, **Lenski RE**, **Long AD**. 2003. Evolutionary changes in heat-inducible gene expression in lines of *Escherichia coli* adapted to high temperature. Physiol Genomics **14**:47–58.

44. **Sleight SC**, **Lenski RE**. 2007. Evolutionary adaptation to freeze-thaw-growth cycles in *Escherichia coli*. Physiol Biochem Zool **80**:370–385.

45. **Goodarzi H**, **Bennett BD**, **Amini S**, **Reaves ML**, **Hottes AK**, **Rabinowitz JD**, **Tavazoie S**. 2010. Regulatory and metabolic rewiring during laboratory evolution of ethanol tolerance in *E. coli*. Mol Syst Biol **6**:378.

46. **Hughes BS**, **Cullum AJ**, **Bennett AF**. 2007. Evolutionary adaptation to environmental pH in experimental lineages of *Escherichia coli*. Evolution (N Y) **61**:1725–1734.

47. **Dragosits M**, **Mattanovich D**. 2013. Adaptive laboratory evolution – principles and applications for biotechnology. Microb Cell Fact **12**.

48. **Harden MM**, **He A**, **Creamer K**, **Clark MW**, **Hamdallah I**, **Martinez K a.**, **Kresslein RL**, **Bush SP**, **Slonczewski JL**. 2015. Acid-Adapted Strains of *Escherichia coli* K-12 Obtained by Experimental Evolution. Appl Environ Microbiol **81**:1932–1941.

49. **Barrick JE**, **Colburn G**, **Deatherage DE**, **Traverse CC**, **Strand MD**, **Borges JJ**, **Knoester DB**, **Reba A**, **Meyer AG**. 2014. Identifying structural variation in haploid microbial genomes from short-read resequencing data using *breseq*. BMC Genomics **15**:1039.

50. **Deatherage DE**, **Traverse CC**, **Wolf LN**, **Barrick JE**. 2015. Detecting rare structural variation in evolving microbial populations from new sequence junctions using *breseq*. Front Genet **5**:1–16.

51. **Deatherage DE**, **Barrick JE**. 2014. Identification of mutations in laboratory-evolved microbes from next-generation sequencing data using *breseq*. Methods Mol Biol **1151**:165–188.

52. **Smith MW**, **Neidhardt FC**. 1983. Proteins induced by aerobiosis in *Escherichia coli*. J Bacteriol **154**:344–350.

53. **Maurer LM**, **Yohannes E**, **Bondurant SS**, **Radmacher M**, **Slonczewski JL**. 2005. pH regulates genes for flagellar motility, catabolism, and oxidative stress in *Escherichia coli* K-12. J Bacteriol **187**:304 – 319.

54. **Lenski RE**, **Rose MR**, **Simpson SC**, **Tadler SC**. 1991. Long-term Experimental Evolution in *Escherichia coli* I. Adaptation and Divergence During 2,000 Generations. Am Nat **138**:1315–1341.

55. **Hayashi K**, **Morooka N**, **Yamamoto Y**, **Fujita K**, **Isono K**, **Choi S**, **Ohtsubo E**, **Baba T**, **Wanner BL**, **Mori H**, **Horiuchi T**. 2006. Highly accurate genome sequences of *Escherichia coli* K-12 strains MG1655 and W3110. Mol Syst Biol **2**:2006.0007.

56. **Thorvaldsdóttir H**, **Robinson JT**, **Mesirov JP**, **Thorvaldsdottir H**, **Robinson JT**, **Mesirov JP**. 2013. Integrative genomics viewer (IGV): High-performance genomics data visualization and exploration. Brief Bioinform **14**:178–192.

57. **Baba T**, **Ara T**, **Hasegawa M**, **Takai Y**, **Okumura Y**, **Baba M**, **Datsenko KA**, **Tomita M**, **Wanner BL**, **Mori H**. 2006. Construction of *Escherichia coli* K-12 in-frame, single-gene knockout mutants: the Keio collection. Mol Syst Biol **2**.

58. **Ma Z**, **Gong S**, **Richard H**, **Tucker DL**, **Conway T**, **Foster JW**. 2003. GadE (YhiE) activates glutamate decarboxylase-dependent acid resistance in *Escherichia coli* K-12. Mol Microbiol **49**:1309–1320.

59. **Lu P**, **Ma D**, **Chen Y**, **Guo Y**, **Chen G-Q**, **Deng H**, **Shi Y**. 2013. L-glutamine provides acid resistance for *Escherichia coli* through enzymatic release of ammonia. Cell Res **23**:635–44.

60. **Lin J**, **Smith MP**, **Chapin KC**, **Baik HS**, **Bennett GN**, **Foster JW**. 1996. Mechanisms of acid resistance in enterohemorrhagic *Escherichia coli*. Appl Environ Microbiol **62**:3094–3100.

61. **Maddamsetti R**, **Lenski RE**, **Barrick JE**. 2015. Adaptation, clonal interference, and frequency-dependent interactions in a long-term evolution experiment with *Escherichia coli*. Genetics **200**:619–631.

62. **Schnetz K**, **Rak B**. 1992. IS5: a mobile enhancer of transcription in *Escherichia coli*. Proc Natl Acad Sci U S A **89**:1244–1248.

63. **Schneider D**, **Lenski RE**. 2004. Dynamics of insertion sequence elements during experimental evolution of bacteria. Res Microbiol **155**:319–327.

64. **Weatherspoon-Griffin N**, **Yang D**, **Kong W**, **Hua Z**, **Shi Y**. 2014. The CpxR/CpxA two-component regulatory system up-regulates the multidrug resistance cascade to facilitate *Escherichia coli* resistance to a model antimicrobial peptide. J Biol Chem **289**:32571–32582.

65. **Li X-Z**, **Plésiat P**, **Nikaido H**. 2015. The challenge of efflux-mediated antibiotic resistance in gram-negative bacteria. Clin Microbiol Rev **28**:337–418.

66. **Kutukova EA**, **Livshits VA**, **Altman IP**, **Ptitsyn LR**, **Zyiatdinov MH**, **Tokmakova IL**, **Zakataeva NP**. 2005. The yeaS (leuE) gene of *Escherichia coli* encodes an exporter of leucine, and the Lrp protein regulates its expression. FEBS Lett **579**:4629–4634.

67. **Lee J**, **Page R**, **García-Contreras R**, **Palermino JM**, **Zhang XS**, **Doshi O**, **Wood TK**, **Peti W**. 2007. Structure and function of the *Escherichia coli* Protein YmgB: A protein critical for biofilm formation and acid-resistance. J Mol Biol **373**:11–26.

68. **Trittler R**, **Kern W V**, **Bohnert A**, **Schuster S**, **Fa E**. 2007. Altered spectrum of multidrug resistance associated with a single point mutation in the *Escherichia coli* RND-type MDR efflux pump YhiV (MdtF). J Antimicrob Chemother **59**:1216–1222.

69. **Ziolkowska K**, **Derreumaux P**, **Folichon M**, **Pellegrini O**, **Boni I V**, **Hajnsdorf E**. 2006. Hfq variant with altered RNA binding functions. Nucleic Acids Res **34**:709–720.

70. **Barrick JE**, **Lenski RE**. 2013. Genome dynamics during experimental evolution. Nat Rev Genet **14**:827–839.

71. **Herring CD**, **Raghunathan A**, **Honisch C**, **Patel T**, **Applebee MK**, **Joyce AR**, **Albert TJ**, **Blattner FR**, **van den Boom D**, **Cantor CR**, **Palsson BØ**. 2006. Comparative genome sequencing of *Escherichia coli* allows observation of bacterial evolution on a laboratory timescale. Nat Genet **38**:1406–1412.

TABLES

Table 1. Mutations in representative benzoate-evolved genomes compared to the genome of *E. coli* W3110.[1]

JLS	K0001	K0022	K0014	K0006	K0030	K0031			
Position	A1-1	A5-1	C3-1	E1-1	G5-1	G5-2	Mutation	Annotation	Gene
56,273							G→A	L279L (CTC→CTT)	imp (lptD) ←
62,682							+G	coding (583/2907 nt)	hepA ←
105,411							G→A	G36D (GGT→GAT)	ftsZ →
156,056							T→G	N49T (AAT→ACT)	ecpD ←
444,525							T→G	intergenic (+127/+1)	yajQ → / ← yajR
490,544							Δ6 bp	intergenic (+61/-87)	ybaN → / → apt
490,998							Δ1 bp	coding (363/552 nt)	apt →
520,989							G→T	P450P (CCG→CCT)	ybbP →
556,778							A→C	F63V (TTC→GTC)	folD ←
573,671							T→A	intergenic (+109/+289)	ybcQ → / ← insH
666,783							A→T	D619E (GAT→GAA)	mrdA ←
683,143						91.8%	Δ5,116 bp	insH-3 IS5-mediated	[hscC]–gltI
755,210					98.8%		IS5 +8bp	intergenic (+147/-374)	gltA ← / sdhC →
909,411							G→A	P102P (CCC→CCT)	ltaE ←
991,031							IS5 (–) +4 bp	intergenic (-253/-10)	pncB ← / → pepN
1,213,665							(C)8→9	intergenic (-85/+615)	elbA ← / ← ycgX
1,218,024							IS5 (+) +4 bp	coding (79-82/267 nt)	ariR (ymgB) →
1,337,160							G→A	intergenic (+617/-385)	cysB → / → acnA
1,349,606	97.8%						IS5 +5 bp	coding (1021/1935 nt)	rnb ←
1,372,264							A→G	E112G (GAG→GGG)	ycjM →
1,459,205							G→A	G354G (GGG→GGA)	paaE →
1,485,978							C→A	R383S (CGT→AGT)	hrpA →
1,549,542							IS5 (–) +4 bp	coding (428-431/3048 nt)	fdnG →
1,553,926							T→C	intergenic (+221/+186)	fdnI → / ← yddM
1,565,001							A→G	intergenic (-211/+47)	ddpX ← / ← dos
1,574,188							IS5 (+) +4 bp	coding (2726-2729/2796 nt)	pqqL ←
1,592,479							C→A	intergenic (-229/+89)	yneL ← / ← hipA
1,618,943							Δ6,115 bp	coding inclusive deletion	ydeA–marCRAB-ydeH
1,704,037							Δ1 bp	coding (91/1002 nt)	add →
1,772,079			97.9%				IS5 +4 bp	intergenic (+179/-250)	ydiK → / ydiL →
1,822,769		98.8%					IS5 +4 bp	coding (160/1359 nt)	chbC ←
1,881,543							IS186/IS421 (+) +6 bp	coding (115-120/360 nt)	yeaR ←
1,882,210							G→A	L88L (CTC→CTT)	yeaS ←
1,908,956							IS5 (–) +4 bp	coding (191-194/210 nt)	cspC ←
1,909,258							IS1 (+) +9 bp	coding (40-48/144 nt)	yobF ←
1,932,183							Δ1 bp	coding (279/291 nt)	yebG ←
2,093,073							G→A	E249K (GAA→AAA)	hisG →
2,156,723							C→A	L191M (CTG→ATG)	mdtA →
2,434,645							A→T	V347E (GTG→GAG)	purF ←
2,447,095							C→T	intergenic (-44/-115)	fabB ← / → trmC
2,487,190							IS5 (+) +4 bp	coding (430-433/1539 nt)	emrY ←
2,646,569							C→A	E1459* (GAG→TAG)	yfhM ←
2,739,952							C→A	intergenic (-146/-64)	aroF ← / → yfiL
2,810,717							IS2 (–) +5 bp	coding (635-639/1173 nt)	emrA →
2,865,825							A→T	I89N (ATC→AAC)	rpoS ←
2,931,775							C→A	P190P (CCG→CCT)	fucA ←
3,104,794							G→A	G142D (GGC→GAC)	nupG →
3,169,126							A→C	T215P (ACA→CCA)	qseB →
3,187,655							IS5 (–) +4 bp	coding (1600-1603/2466 nt)	yqiG →
3,188,360							IS5 (–) +4 bp	coding (2305-2308/2466 nt)	yqiG →
3,189,138							IS5 (+) +4 bp	coding (602-605/750 nt)	yqiH →
3,212,340							A→C	D213A (GAC→GCC)	rpoD →
3,241,721							G→T	L84M (CTG→ATG)	uxaA ←
3,277,113		98.8%					INDEL +5 bp	coding (257/336 nt)	prlF →
3,277,128							(TTCAACA)2→3	coding (272/336 nt)	sohA →
3,454,320							C→T	G373S (GGC→AGC)	rpoB ←
3,532,025							A→G	N107S (AAC→AGC)	cpxA →
3,840,032							IS5 (–) +4 bp	coding (583-586/699 nt)	rfaY →
3,909,304							IS5 (+) +4 bp	intergenic (-20/-343)	xylF ← / → xylA
3,948,766							G→A	R320H (CGT→CAT)	bcsB →
3,974,240							Δ14,146 bp	insH IS5-mediated	gadXW–mdtFE-hdeDAB-yhiS
3,974,646				Δ			Δ78 bp	coding (42-119/825 nt)	gadX →
3,975,201				Δ			G→T	L199F (TTG→TTT)	gadX →
3,975,230				Δ			IS5 (+) +4 bp	coding (626-629/825 nt)	gadX →
3,976,435				Δ			Δ10,738 bp	insH-mediated	gadW–slp
3,986,969	Δ			Δ			Δ204 bp	insH-mediated	slp ← / → insH
4,114,118							C→A	intergenic (-171/-149)	yrfF ← / → nudE
4,136,677							G→T	V191V (GTC→GTA)	frlD ←

JLS	K0001	K0022	K0014	K0006	K0030	K0031			
Position	A1-1	A5-1	C3-1	E1-1	G5-1	G5-2	Mutation	Annotation	Gene
3,975,230				Δ			IS5 (+) +4 bp	coding (626-629/825 nt)	gadX →
3,976,435				Δ			Δ10,738 bp	insH-mediated	gadW–slp
3,986,969	Δ			Δ			Δ204 bp	insH-mediated	slp ← / → insH
4,114,118							C→A	intergenic (-171/-149)	yrfF ← / → nudE
4,136,677							G→T	V191V (GTC→GTA)	frlD ←
4,200,197							A→C	K271Q (AAA→CAA)	rpoA →
4,218,986							IS5 (–) +4 bp :: Δ4	intergenic (+187/-79)	metA → / → aceB
4,221,755							G→A	A353T (GCA→ACA)	aceA →
4,297,865							+T	intergenic (+136/-206)	nrfG → / → gltP
4,397,133							C→T	intergenic (+18/-19)	glyV → / → glyX
4,397,136							A→G	intergenic (+21/-16)	glyV → / → glyX
4,405,094							G→A	V43M (GTG→ATG)	hfq →
4,485,284							IS5 (–) +4 bp	coding (875-878/1197 nt)	yjgN →
4,495,464							IS5 (+) +4 bp	coding (353-356/999 nt)	idnR ←
4,546,700							IS5 (–) +4 bp	intergenic (+461/-14)	fimB → / → fimE
4,546,841							IS5 (–) +4 bp	coding (125-128/597 nt)	fimE →
4,547,128							IS5 (–) +4 bp	coding (412-415/597 nt)	fimE →
4,547,650							Δ1 bp	intergenic (+337/-145)	fimE → / → fimA
4,547,860							(TCCCTCAGTTCTACAGCGGCTCTG)1→2	coding (66/549 nt)	fimA →
4,626,165							C→G	C201W (TGC→TGG)	deoD →
4,639,891							A→G	S34P (TCC→CCC)	rob ←

[1] NCBI reference strain NCBI NC_007779.1. "Δ" indicates mutation site is absent within a larger deleted region. Percentage scores indicate *breseq* calls less than 100%. Annotations: Red = changed base pairs; green = synonymous mutation; blue = missense mutation; purple = IS-mediated deletion; and * = nonsense mutation. Mutations present in our laboratory stock strain W3110-D13 compared to the NCBI reference strain (**Table S3**) are omitted from Table 1.

Table 2. MIC (minimum inhibitory concentration) of benzoate-evolved strains in the antibiotics chloramphenicol, tetracycline, and streptomycin.*

	Chloramphenicol	Tetracycline
W3110	16	4
marA	8	4
A1-1	8	2
A5-1	8	1
C3-1	8	3
E1-1	16	4
G5-1	12	2
G5-2	8	2

*Cultured in LBK 100 mM MOPS pH 7.0, 2 mM salicylate, for 22 h. Positive for growth was defined as cell density at 22 h (OD600 $\geq$ 0.05). Concentration (µg/ml) represents median value of 8 replicates.

FIGURES

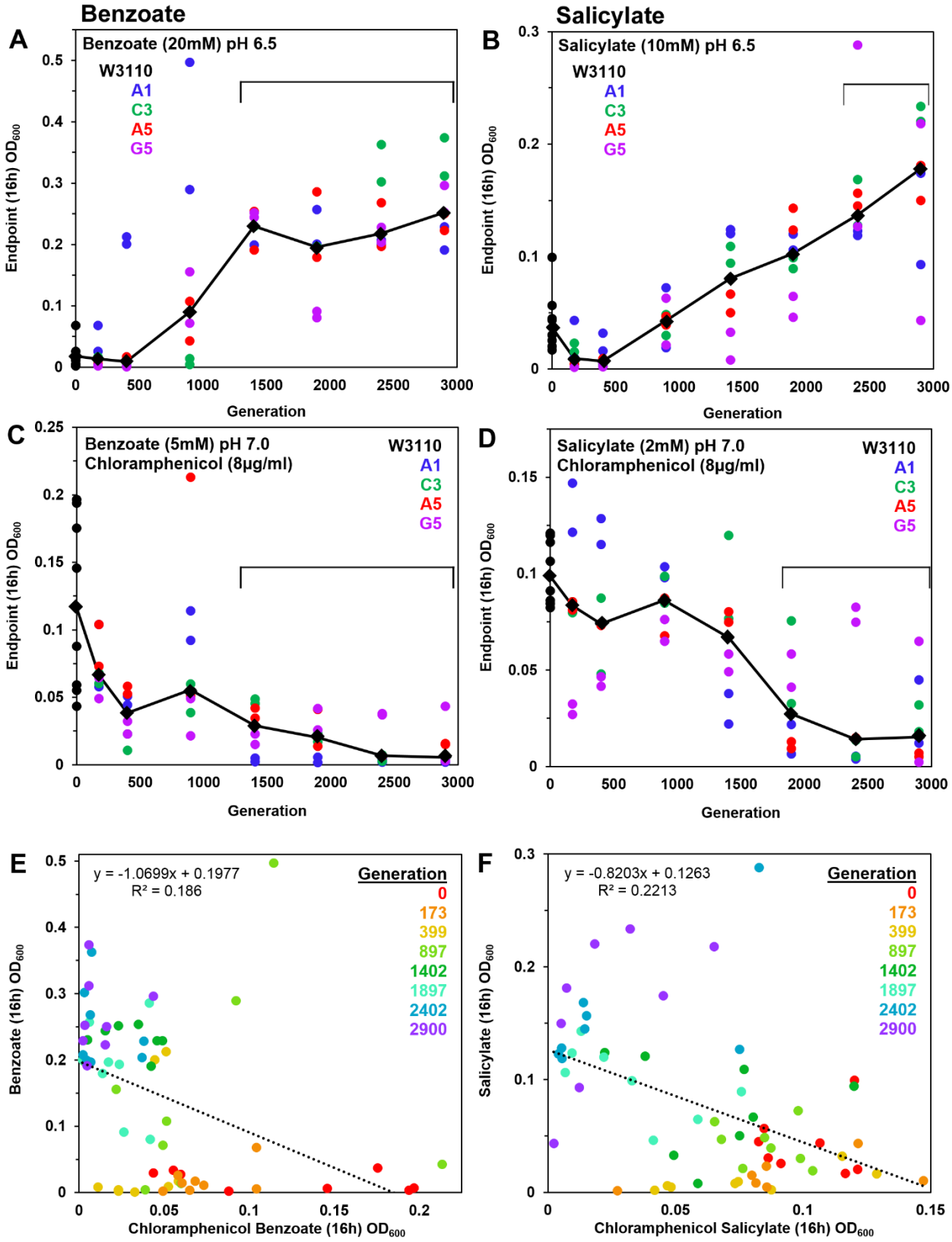
Benzoate
Salicylate
A
Benzoate (20mM) pH 6.5
W3110
A1
C3
A5
G5
Endpoint (16h) OD600
Generation
B
Salicylate (10mM) pH 6.5
C
Benzoate (5mM) pH 7.0
Chloramphenicol (8μg/ml)
D
Salicylate (2mM) pH 7.0
E
y = -1.0699x + 0.1977
R² = 0.186
Generation
0
173
399
897
1402
1897
2402
2900
Benzoate (16h) OD600
Chloramphenicol Benzoate (16h) OD600
F
y = -0.8203x + 0.1263
R² = 0.2213
Salicylate (16h) OD600
Chloramphenicol Salicylate (16h) OD600

FIG 1 Growth measured after generations of repeated dilution and culture in 5-20 mM benzoate. Benzoate-evolved strains were obtained from frozen microplates, selecting 2 different clones from each of 4 populations. Clones from each plate generation were cultured at 37ºC in a column of microplate wells; cell density "E" values (OD_{600}) were obtained at 16 h. Diamonds indicate median cell density for each generation tested. Bracket indicates generations for which the 16-h cell density differed significantly from that of the ancestral strain W3110, in 2 of 3 trials (Friedman test; post-hoc Conover pairwise comparisons with Holm-Bonferroni adjusted p-values). LBK media contained: **A.** 100 mM PIPES pH 6.5 with 20 mM benzoate (diluted 1:200 from overnight cultures with 5mM benzoate). **B.** 100 mM PIPES pH 6.5 with 10 mM salicylate (diluted 1:200 from overnight cultures in 2 mM salicylate). **C.** 100 mM MOPS pH 7.0 with 5 mM benzoate, 8 µg/ml chloramphenicol (diluted 1:200 from overnight cultures without chloramphenicol). **D.** 100 mM MOPS pH 7.0 with 2 mM salicylate, 8 µg/ml chloramphenicol (diluted 1:200 from overnight cultures without chloramphenicol). **E.** Plot with linear regression of 16-h cell-density values for 20 mM benzoate and for 5 mM benzoate, 8 µg/ml chloramphenicol exposures. **F.** Plot of 16-h cell-density values for 10 mM salicylate and for 2 mM salicylate, 8 µg/ml chloramphenicol exposures.

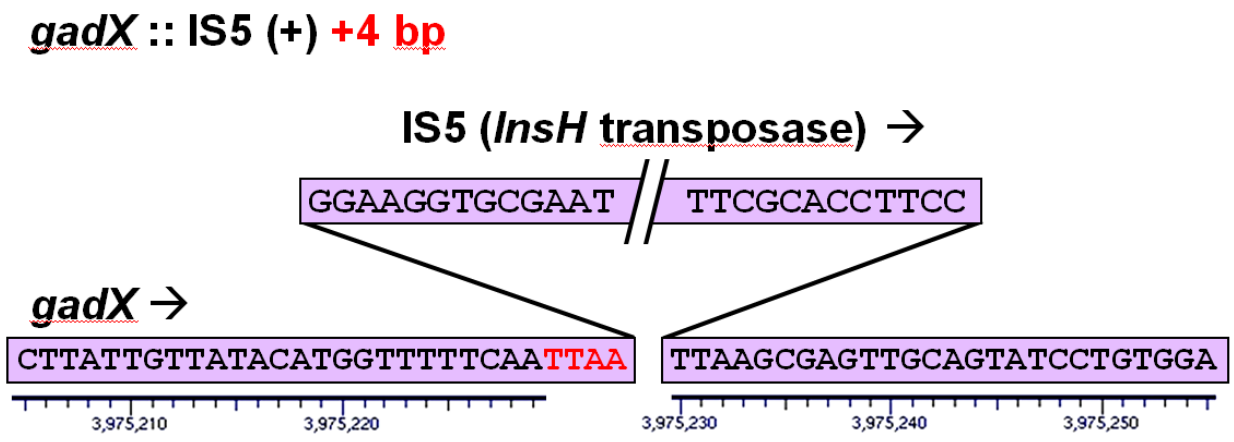

FIG 2 New insertion of IS5 within *gadX* including a 4-bp duplication of the target site, in the A5-1 genome at position 3,975,230.

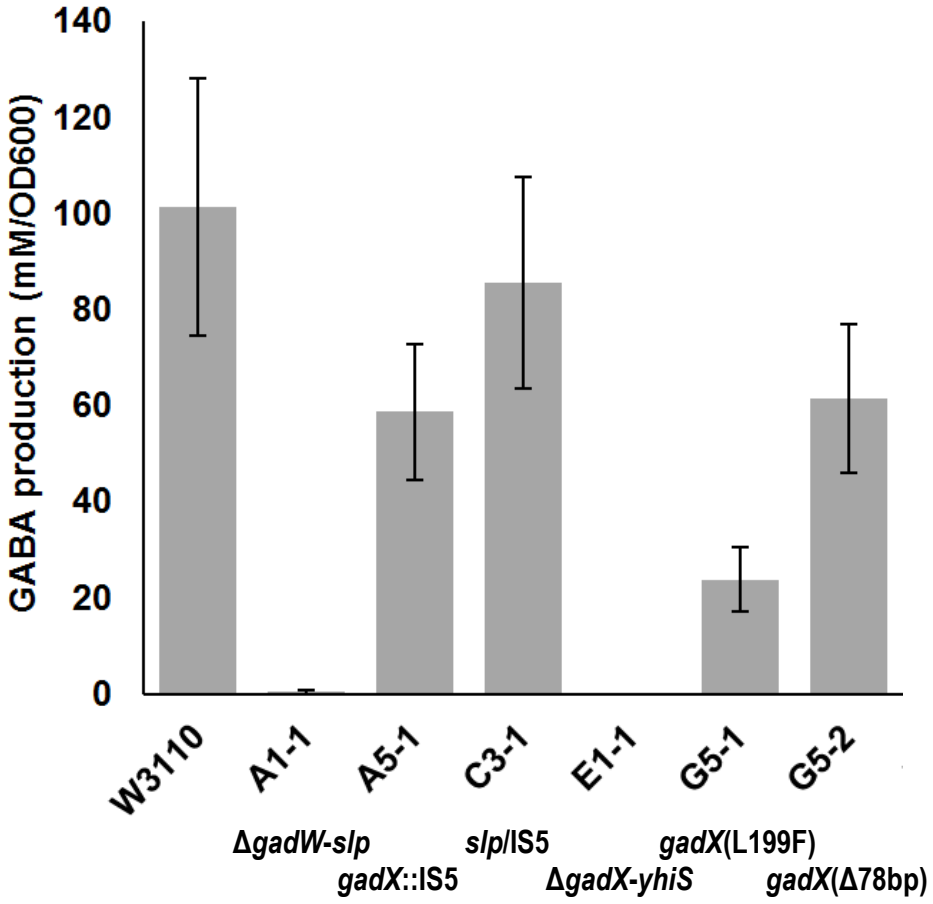

FIG 3 GABA produced by benzoate-evolved strains compared to W3110. Anaerobic overnight cultures in LB 10 mM glutamine, 100 mM MES pH 5.5 were adjusted with HCl to pH 2. After 2 h incubation, bacteria were pelleted and supernatant culture fluid was derivatized using EZ:faast (Phenomenex). GABA was quantified via GC/MS, and values were normalized to the cell density of the overnight culture. Error bars represent SEM (n=7 or 8). Genetic annotations are from Table 1.

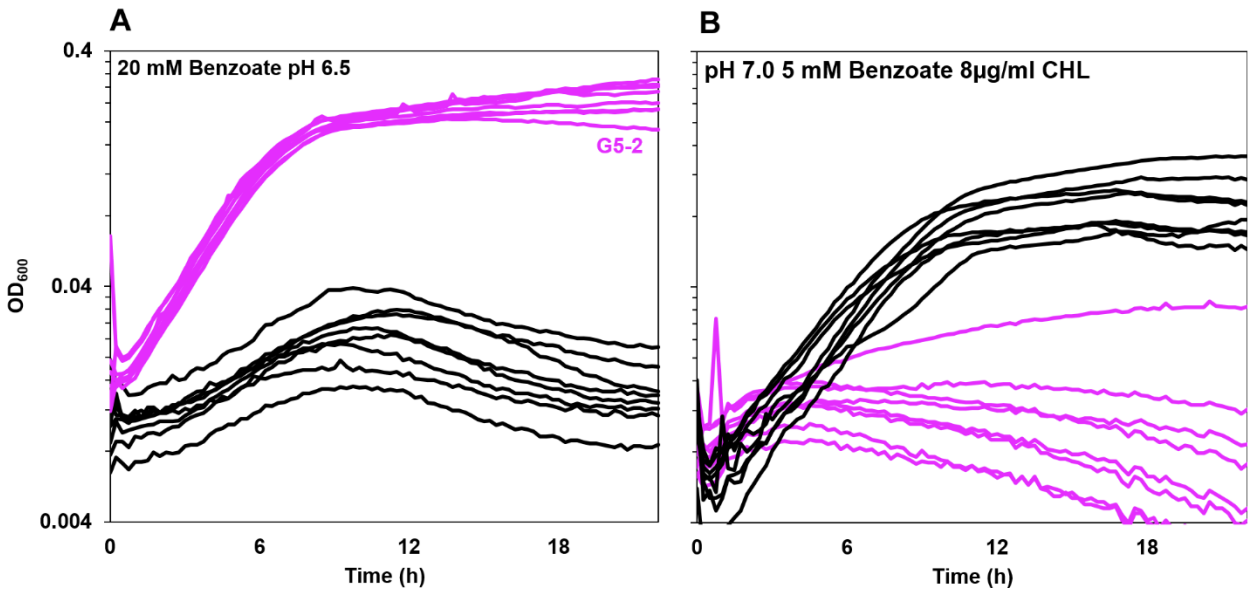

FIG 4 Benzoate-evolved strain G5-2 outgrows ancestor W3110 in presence of benzoate, but grows poorly in benzoate with chloramphenicol. Growth medium was LBK with (**A**) 100 mM PIPES, 20 mM benzoate pH 6.5, (**B**) 100 mM MOPS, 5 mM benzoate pH 7.0, 8 µg/ml chloramphenicol. For each strain, 8 replicate curves from microplate wells are shown. Cell density values post log-phase (OD_{600} at 16 h) were ranked and compared by Friedman test; post-hoc Conover pairwise comparisons with Holm-Bonferroni adjusted p-values.

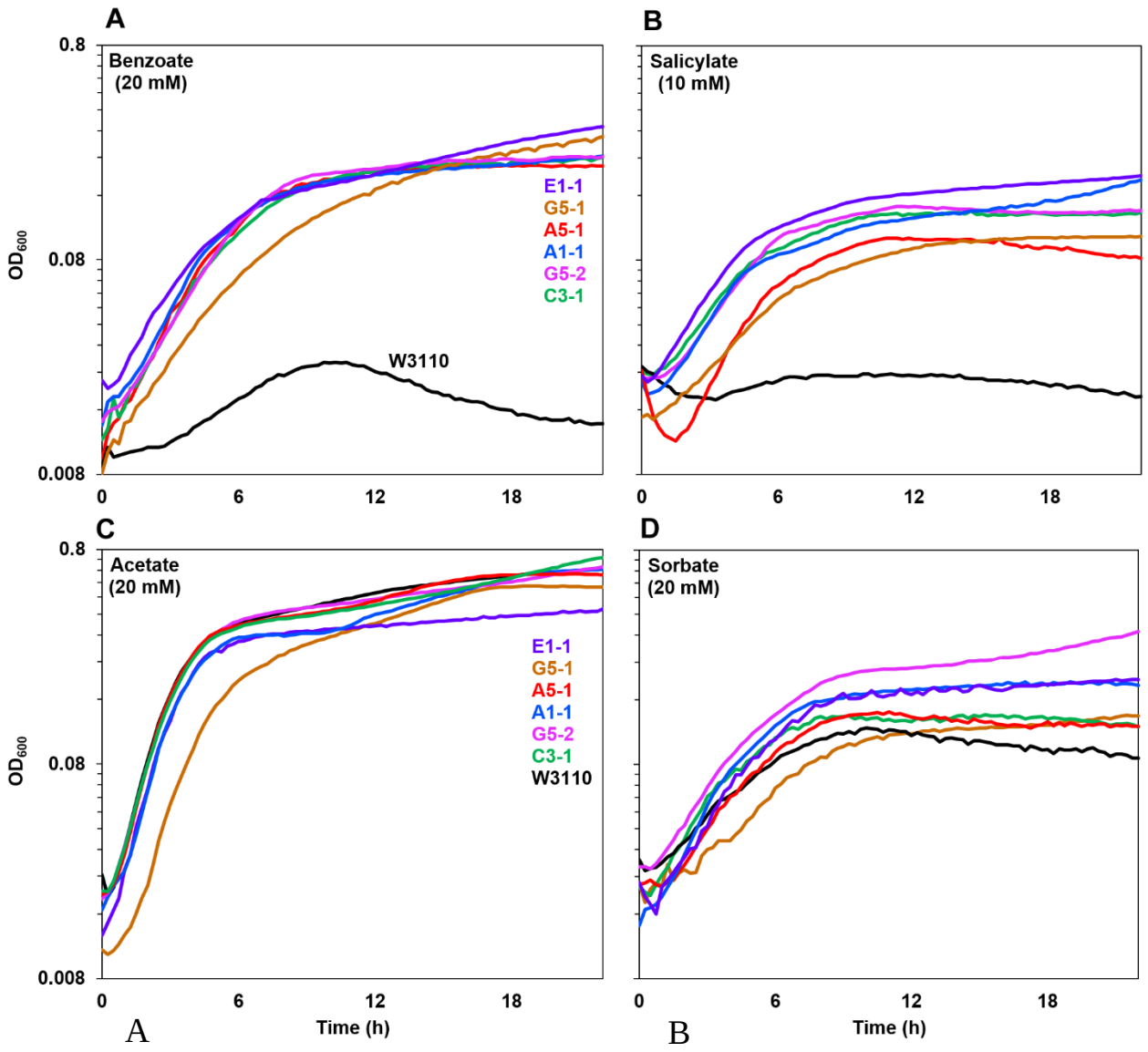

FIG 5 Benzoate-evolved strains all outgrow ancestor in benzoate or salicylate but not in acetate or sorbate. Growth curves of benzoate-evolved strains and ancestor in LBK 100 mM PIPES pH 6.5 with (A) 10 mM salicylate, (B) 20mM benzoate, (C) 20mM acetate, or (D) 20 mM sorbate. For each strain, a curve with median cell density at 16 h is shown. Panels A and B (but not C and D) showed significantly lower 16-h cell density for the ancestral W3110 strain (black curve) than for benzoate-evolved strains (Friedman test; post-hoc Conover pairwise comparisons with Holm-Bonferroni adjusted p-values).

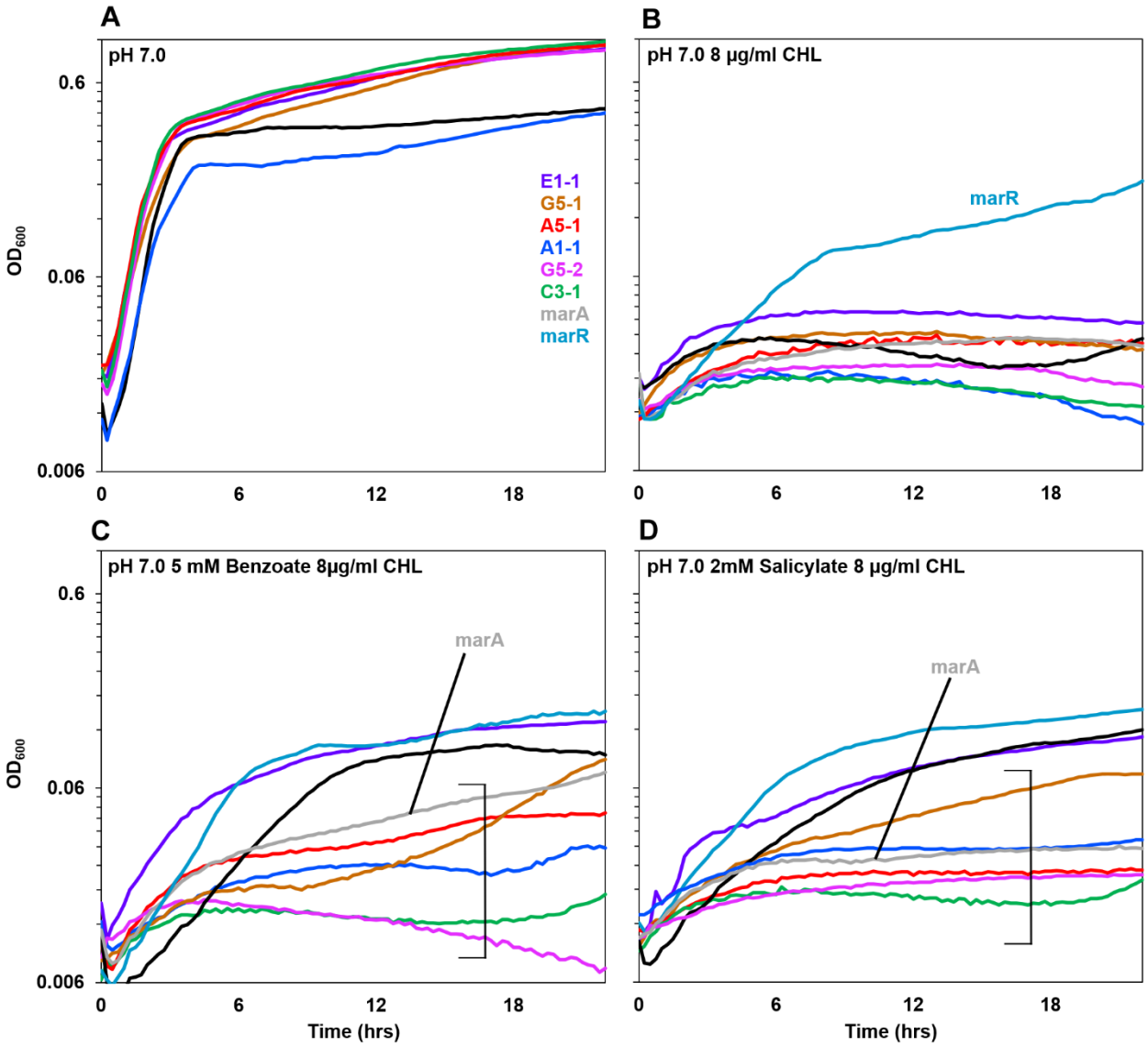
A
pH 7.0
0.6
0.06
0.006
OD600
0
6
12
18
E1-1
G5-1
A5-1
A1-1
G5-2
C3-1
marA
marR
B
pH 7.0 8 µg/ml CHL
marR
C
pH 7.0 5 mM Benzoate 8µg/ml CHL
marA
Time (hrs)
D
pH 7.0 2mM Salicylate 8 µg/ml CHL
marA
Time (hrs)

FIG 6 Benzoate-evolved strains are sensitive to chloramphenicol. Growth curves of benzoate-evolved strains and ancestor, compared to W3110 strains deleted for *marR* and for *marA*. Media contained LBK 100 mM MOPS pH 7.0 with (**A**) no supplements, (**B**) 8 µg/ml chloramphenicol, (**C**) 5 mM benzoate and 8 µg/ml chloramphenicol, (**D**) 2 mM salicylate and 8 µg/ml chloramphenicol. For each strain, a curve with median 16-h cell density is shown. Bracket indicates curves with 16-h cell density lower than that of ancestral strain W3110 (Friedman test; post-hoc Conover pairwise comparisons with Holm-Bonferroni adjusted p-values). For panels C and D, **Figure S3** shows all replicates of each strain plotted individually.

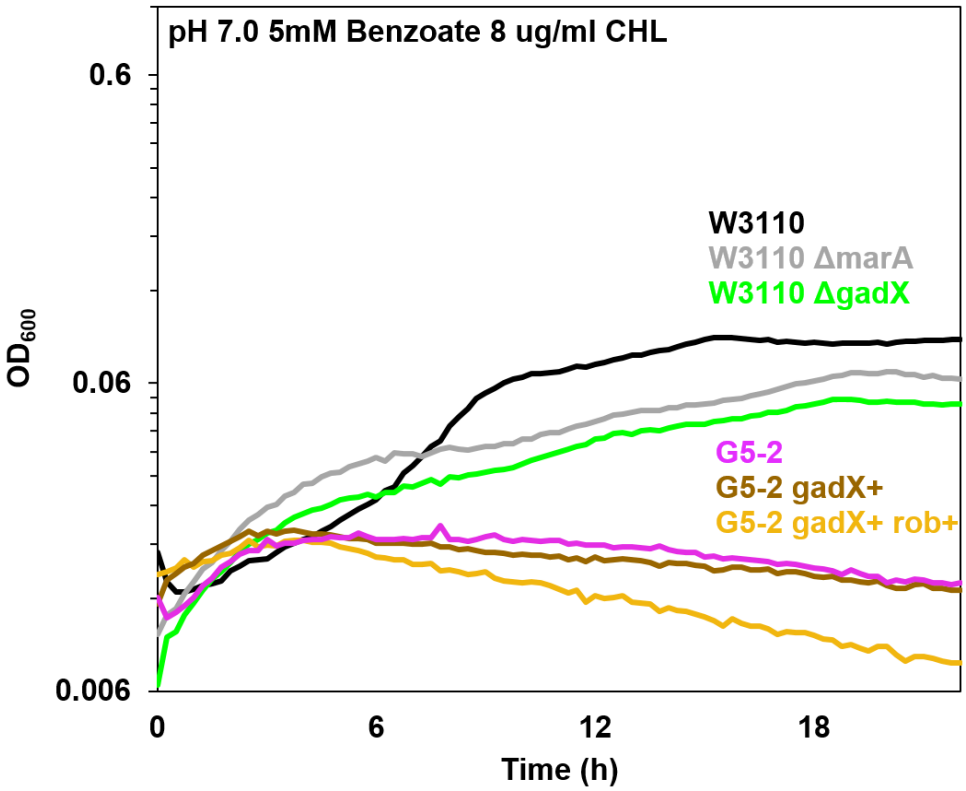

FIG 7 W3110 *gadX*::kanR shows chloramphenicol sensitivity similar to that of *marA*::kanR. Growth curves were conducted in LBK 100 mM PIPES pH 7.0 with 5 mM benzoate, as for Figure 6. The *gadX*::*kanR* knockout showed no significant difference in growth than the *marA*::*kanR* strain. At 16 h, the cell density of strain G5-2 showed no difference from those of G5-2 *gadX*+ or of G5-2 *gadX*+ *rob*+ (Friedman/Conover/Holm-Bonferroni).

72. **Blattner FR**, **Plunkett G**, **Bloch CA**, **Perna NT**, **Burland V**, **Riley M**, **Collado-Vides J**, **Glasner JD**, **Rode CK**, **Mayhew GF**, **Gregor J**, **Davis NW**, **Kirkpatrick HA**, **Goeden MA**, **Rose DJ**, **Mau B**, **Shao Y**. 1997. The complete genome sequence of *Escherichia coli* K-12. Science (80-) **277**:1453–1462.

73. **Wang X**, **Wood TK**. 2011. IS*5* inserts upstream of the master motility operon *flhDC* in a quasi-Lamarckian way. ISME J **5**:1517–1525.

74. **Pumbwe L**, **Skilbeck CA**, **Wexler HM**. 2007. Induction of multiple antibiotic resistance in *Bacteroides fragilis* by benzene and benzene-derived active compounds of commonly used analgesics, antiseptics and cleaning agents. J Antimicrob Chemother **60**:1288–1297.

75. **Ueda O**, **Wexler HM**, **Hirai K**, **Shibata Y**, **Yoshimura F**, **Fujimura S**. 2005. Sixteen homologs of the mex-type multidrug resistance efflux pump in *Bacteroides fragilis*. Antimicrob Agents Chemother **49**:2807–2815.

76. **Needs C**, **Brooks P**. 1985. Clinical pharmacokinetics of the salicylates. Clin Pharmacokinet **10**:164–177.

77. **Lanter BB**, **Sauer K**, **Davies DG**. 2014. Bacteria present in carotid arterial plaques are found as biofilm deposits which may contribute to enhanced risk of plaque rupture. MBio **5**:1–8.

78. **Sears CL**, **Garrett WS**. 2014. Microbes, microbiota, and colon cancer. Cell Host Microbe **15**:317–328.

79. **Johnson CH**, **Dejea CM**, **Sears CL**, **Johnson CH**, **Dejea CM**, **Edler D**, **Hoang LT**, **Santidrian AF**, **Felding BH**, **Casero RA**, **Pardoll DM**, **White JR**, **Patti GJ**, **Sears CL**, **Siuzdak G**. 2015. Metabolism links bacterial biofilms and colon carcinogenesis. Cell Metab **21**:891–897.