bioRxiv preprint doi: https://doi.org/10.1101/063560; this version posted July 18, 2016. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

Phylogenetic scale in ecology and evolution

Catherine H. Graham [1], Antonin Machac [2,3,4], David Storch [3,4]

[1] Department of Ecology and Evolution, 650 Life Sciences Bldg, Stony Brook University, Stony Brook, NY 11794, USA – [2] Center for Macroecology, Evolution, and Climate, Natural History Museum of Denmark, Universitetsparken 15, DK 2100 Copenhagen – [3] Department of Ecology, Vinicna 7, 12844 Prague 2, Czech Republic – [4] Center for Theoretical Study, Jilska 1, 11000 Prague 1, Czech Republic.

Email contacts: CHG: catherine.graham@stonybrook.edu, AM: A.Machac@email.cz, DS: storch@cts.cuni.cz. Author for correspondence: Catherine H. Graham, telephone (US 1-631-632-1962), fax (US 1-631-632-7626). Statement of authorship: The authors conceived and wrote the manuscript together, contributing equally.

Running title: Phylogenetic scale in ecology and evolution. Keywords: biogeography, community structure, diversification, domains of scale, extent, grain, macroecology, macroevolution, spatial scale. The text consists of 5200 words, 2 boxes (~420 words each), 1 figure, 1 table, 100 references

bioRxiv preprint doi: https://doi.org/10.1101/063560; this version posted July 18, 2016. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

SUMMARY It has been widely acknowledged that many phenomena in ecology and evolution depend on spatial and temporal scale. However, important patterns and processes may vary also across the phylogeny and depend on phylogenetic scale. Though phylogenetic scale has been implicitly considered in some previous studies, it has never been formally conceptualized and its potential remains unexplored. Here, we develop the concept of phylogenetic scale and, building on previous work in the field, we introduce phylogenetic grain and extent, phylogenetic scaling and the domains of phylogenetic scale. We use examples from published research to demonstrate how phylogenetic scale has been considered so far and illustrate how it can inform, and possibly resolve, some of the longstanding controversies in evolutionary biology, community ecology, biogeography and macroecology. To promote the concept of phylogenetic scale empirically, we propose methodological guidelines for its treatment.

bioRxiv preprint doi: https://doi.org/10.1101/063560; this version posted July 18, 2016. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

INTRODUCTION

Numerous patterns in ecology and evolution vary across the phylogenetic hierarchy (Fig. 1). Species diversity declines with latitude across higher taxa but not necessarily across their constituent families and genera (Buckley *et al.* 2010) that also often conform to different diversification dynamics (Phillimore & Price 2008; Morlon *et al.* 2010; Rabosky *et al.* 2012). Phylogenetic delimitation of species pools determines our inferences about community assembly (Cavender-Bares *et al.* 2009). Many other, similar examples can further illustrate that patterns in ecology and evolution often depend on phylogenetic scale (Fig. 1). Yet, unlike the extensively developed and widely recognized concepts of spatial and temporal scale (e.g. Wiens 1989; Levin 1992; Schneider 2001), phylogenetic scale remains largely unnoticed and its importance has only recently been emerging. Here, we formalize and develop the concept of phylogenetic scale, summarize how it has been considered across disciplines, and suggest further research directions.

Inspired by the concept of spatial scale (e.g. Wiens 1989; Levin 1992; Schneider 2001), we define phylogenetic scale in terms of phylogenetic grain and phylogenetic extent. Phylogenetic grain refers to the elementary unit of analysis, defined in terms of tree depth, taxonomic rank, clade age, or clade size. Phylogenetic extent refers to the entire phylogeny encompassing all these units (Box 1). The grain and extent of biological patterns might inform us about the mechanisms that produced these patterns. For example, the number of families in the fossil record seems to stay constant while the number of genera seems to increase continually over geological time, suggesting that different mechanisms produce genus-level and family-level diversity (Benton & Emerson 2007). In community ecology, clade-wide analyses typically suggest that communities have been shaped by environmental filters (Parra *et al.* 2011) while focused analyses of narrowly defined clades often uncover a suite of additional mechanisms (e.g. competition, mutualisms, dispersal limitation) (Parra *et al.* 2011; McGuire *et al.* 2014). Different patterns, and by extension different inferences about the underlying mechanisms, might therefore emerge across the continuum of phylogenetic scales.

The concept of phylogenetic scale seems particularly pertinent, given the growing body of research and statistical methods to explore the increasingly accurate and ever more complete phylogenetic data (e.g. phylogenetic comparative methods, community phylogenetics,

bioRxiv preprint doi: https://doi.org/10.1101/063560; this version posted July 18, 2016. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

diversification analysis). Yet, few studies have extended the explorative strategies to systematically investigate phylogenetic scale-dependence (e.g. upscaling, downscaling), delimit phylogenetic domains of ecological theories (e.g. niche conservatism, competition and filtering, equilibrial diversification), or evaluate the universal laws of ecology (e.g. species-abundance distributions, latitudinal gradients). We therefore contend that the full potential of the phylogenetic data and the methods at hand has not yet been realized, and further progress might be precipitated by a more focused and formalized treatment of phylogenetic scale, akin to the one commonly applied across temporal and spatial scales (e.g. Wiens 1989; Levin 1992; Schneider 2001).

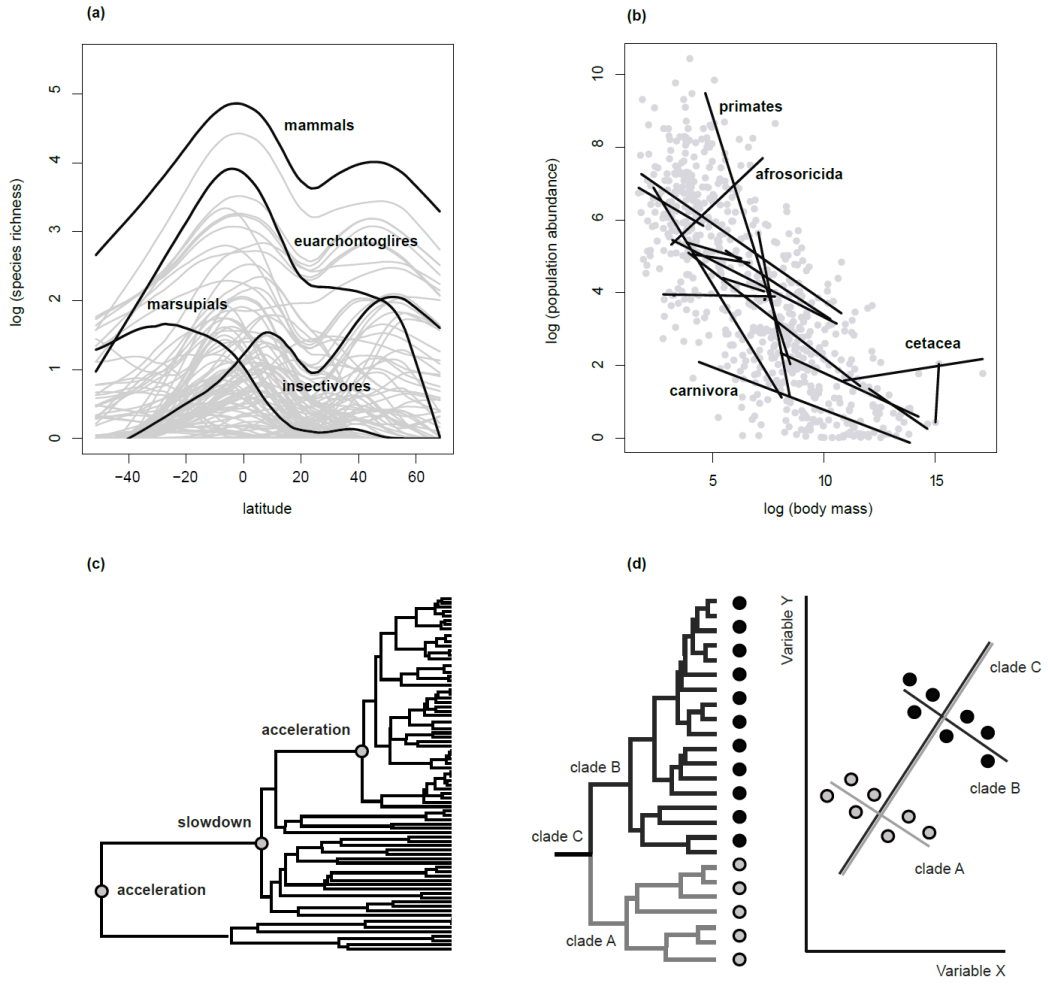
Here, we summarize the variety of ways in which different disciplines have considered phylogenetic scale, highlighting their respective benefits and pitfalls. We further propose how these efforts might be consolidated under one conceptual and empirical framework that would provide the common ground for cross-disciplinary discussion. In particular, we define the concept of phylogenetic scale, distinguish between phylogenetic grain and extent, scale-dependence, phylogenetic scaling and the domains of scale. We also provide practical guidelines for the treatment of phylogenetic scale across empirical studies, using the data and statistical methods currently available. We hope this will inspire further discussion, draw more focused attention to the subject, and advance the notion of phylogenetic scale in ecology and evolution.

PHYLOGENETIC SCALE IN ECOLOGY AND EVOLUTION

Phylogenetic scale has been considered to varying degrees in ecology and evolution, from being largely neglected to being relatively well-developed. In this section, we describe previous research that has implicitly or explicitly considered phylogenetic scale but also how the core disciplines in the field might further benefit from this concept (e.g. macroevolution, community ecology, biogeography, macroecology).

bioRxiv preprint doi: https://doi.org/10.1101/063560; this version posted July 18, 2016. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

Figure 1. Examples of patterns that vary across phylogenetic scales. (a) The latitudinal diversity gradient. Mammal diversity decreases with latitude across large clades but many other patterns emerge across small clades, including inverse ones (select clades depicted in black). (b) The dependence of population abundance on body mass. The dependence is negative across large phylogenetic scales (mammals depicted in grey) but varies substantially across small scales (select orders depicted in black). (c) Diversification dynamics. Slowdowns detected over some phylogenetic scales might be accompanied by accelerations over both larger and smaller scales. (d) Statistical correlations. Even though the depicted variables are negatively correlated within each of the two subclades, the correlation becomes positive when the subclades are studied together. The data (a, b) were taken from the IUCN (2016) and PanTHERIA (2016).



bioRxiv preprint doi: https://doi.org/10.1101/063560; this version posted July 18, 2016. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

BOX 1: The concept of phylogenetic scale

The concept of scale is based on the fact that some entities can be ordered, or placed on a scale (*scala* means *ladder* in Latin). For example, continents contain biomes, ecoregions, and localities, giving rise to spatial hierarchy. Similarly, large clades contain small clades, creating phylogenetic hierarchy which defines phylogenetic scale. However, clades are not always nested, in which case standard measures might be needed to order the clades along the scale continuum. These measures might include time (clade age) but also clade size (number of species within a clade) or the degree of molecular, phenotypic, or ecological divergence within a clade. These measures will be inherently correlated across mutually nested clades but might become decoupled across non-nested clades (e.g. old clades will not necessarily be most diverse) (Box 2).

In the concept of spatial scale, **grain** and **extent** are usually distinguished. Grain refers to the area of the basic unit analyzed (e.g. ecoregion) while extent refers to the total area analyzed (e.g. continent). Phylogenetic scale can be defined analogically, such that phylogenetic grain refers to the basic unit of analysis (e.g. species, genera, families) and phylogenetic extent to the total phylogeny that would encompass all the units analyzed (e.g. class, phylum).

Even though taxonomic ranks are commonly used to define phylogenetic scale, they are not always comparable (e.g. genera in mammals are not comparable to genera in insects), and standard measures might be better suited to define phylogenetic scales across distant taxa (e.g. taxon age, taxon size), but even these measures might not ensure entirely that the analyzed taxa are fully comparable. For example, clade age might reflect the degree of phenotypic divergence across clades, but some clades might be more diverged than others despite being of similar age. The same limitations apply to the measures of spatial scale because spatial grains of standardized sizes might not ensure comparability across species of dramatically different geographic and home range sizes (Wiens 1989). Therefore, the most suitable measure and definition of the phylogenetic scale should be dictated by the biological properties of the organismal system (e.g. generation time, reproductive output, population size) and the question under study (e.g. phenotypic divergence, diversification dynamics, diversity patterns).

In some cases, it may be useful to work with non-standardized grains which represent more natural units of analysis (e.g. islands in spatial scaling or island faunas in phylogenetic scaling).

bioRxiv preprint doi: https://doi.org/10.1101/063560; this version posted July 18, 2016. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

The extents will then be defined correspondingly, so as to cover all of the units analyzed (e.g. all islands or the entire biotas across islands). Finally, grain and extent are defined only in relation to each other. The grain from one study can therefore act as an extent in another study, or vice versa.

Evolution and diversification

Evolutionary diversification and disparification are known to vary across phylogenetic scales but have rarely been thoroughly studied in this context. Even though a suite of methods are commonly used to explore these processes across the phylogeny (e.g. Rabosky *et al.* 2012; Alfaro *et al.* 2009) (see Table 1), most studies report the recovered patterns without a focused examination of their scale-dependence. Focused examination of patterns across scales may precipitate the resolution of several outstanding controversies in the field.

One such controversy revolves around the dynamics of diversity and diversification. It has been debated whether the dynamics are expansionary, such that regional and clade diversity accumulate constantly over time (Benton & Emerson 2007; Wiens 2011; Harmon & Harrison 2015), or whether the dynamics are ecologically limited, such that diversity tends toward an equilibrium (Rabosky 2009; Rabosky & Hurlbert 2015). Empirical evidence suggests that genera with dozens of species often expand in terms of their diversity (McPeek 2008; Morlon *et al.* 2010) whereas higher taxa with thousands of species are mostly saturated at their equilibrium diversity (Vamosi & Vamosi 2010; Rabosky *et al.* 2012). Island radiations and fossil evidence also indicate that clades often expand, seemingly without bounds, during the initial phases of their diversification but eventually reach an equilibrium and saturate (Alroy 1996; Benton & Emerson 2007; Glor 2010; Quental & Marshall 2013). It is therefore possible that diversification varies systematically across phylogenetic scales such that seemingly contradictory dynamics (i.e. expansionary and equilibrial) might be detected across different segments of the same phylogeny. If that were the case, the debate as to whether the dynamics are expansionary or equilibrial might not prove particularly productive and should perhaps be reframed in terms of phylogenetic scale (e.g. phylogenetic scales over which the different dynamics prevail, scale-related factors that

bioRxiv preprint doi: https://doi.org/10.1101/063560; this version posted July 18, 2016. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

determine the shift between the dynamics, how the dynamics combine across scales and across nested clades of different ages and sizes).

Evolutionary disparification may also vary across the phylogeny because traits (phenotypic, behavioral, but also molecular) diverge at different rates and therefore are conserved over different phylogenetic scales (Freckleton *et al.* 2002; Blomberg *et al.* 2003; Wiens & Graham 2005). Even though the dynamics of trait divergence and niche conservatism have been the subject of much research (e.g. Blomberg *et al.* 2003; Freckleton *et al.* 2002), clear generalizations about their scale-dependence have not yet emerged. In most cases, physiological traits that largely determine the extent of species distributions seem conserved over extensive phylogenetic scales (Freckleton *et al.* 2002; Blomberg *et al.* 2003) while habitat- and diet-related traits that mediate species coexistence locally seem generally labile and conserved over small scales (Ackerly *et al.* 2006; Buckley *et al.* 2010). However, the opposite pattern has also been observed where physiological tolerances were conserved over small scales while habitat, diet, body size, and feeding method remained unchanged for most of a clade's history (Kennedy *et al.* 2012; Price *et al.* 2014).

These mixed results suggest that temporal scale may be insufficient to fully capture the variance in niche conservatism. Phylogenetic scale, in contrast, captures the fact niches and traits may evolve at different rates even across closely related clades (e.g. due to clade-specific selection regimes, genetic architecture, pleiotropy) that span similar temporal scales but different phylogenetic scales (e.g. one clade has undergone an explosive radiation on an island while the other has accumulated only limited morphological, ecological, and species diversity on the mainland). In these cases, time will not capture the phylogenetic hierarchy as well as phylogenetic scale would (e.g. phylogenetic domains defined to reflect clade size, phenotypic and ecological divergence) (see below). The concept of phylogenetic scale may therefore encourage a more realistic and potentially more accurate way of thinking about trait evolution and niche conservatism.

Community ecology

Patterns of community phylogenetic structure, and hence the inferred processes that shape communities, can vary with phylogenetic scale (Webb *et al.* 2002; Cavender-Bares *et al.* 2009;

bioRxiv preprint doi: https://doi.org/10.1101/063560; this version posted July 18, 2016. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

Vamosi *et al.* 2009; Münkemüller *et al.* 2014). Even though community phylogeneticists have long been aware of this fact (Webb *et al.* 2002; Cavender-Bares *et al.* 2009), most studies routinely do not recognize the influence of phylogenetic scale on their results.

To study the phylogenetic structure of a community, researchers calculate standardized community metrics (e.g. the net relatedness index, NRI) based on null models which assemble random communities from the regional pool of species. Phylogenetic delimitation of the species pool can influence these metrics profoundly and inform us about the mechanisms that mediate local coexistence of differently delimited suites of species (Cavender-Bares *et al.* 2009; Lessard *et al.* 2012; Cornell & Harrison 2014). For example, communities of mutually unrelated species that have been selected from a broadly defined species pool are typically shaped by environmental filters (Parra *et al.* 2011; Cavendar-Bares *et al*. 2009) while narrowly defined pools often uncover signatures of competition, mutualism, or dispersal limitation, among the closely related and locally coexisting species (Swenson *et al.* 2007; Cavender-Bares *et al.* 2009; Parra *et al.* 2011).

The interpretation of community structure has been under increasing scrutiny lately because different processes can produce similarly-structured communities (e.g. Mayfield & Levine 2010; Gerhold *et al*. 2015) and a single metric may not capture community structure sufficiently to identify the processes that produced the community (Gerhold *et al.* 2015). We argue that using multiple metrics across phylogenetic scales, as illustrated by some recently developed statistical approaches (Leibold *et al.* 2010; Pavoine *et al.* 2010; Borregaard *et al.* 2014), could be a particularly powerful strategy to capture community structure more completely and disentangle the interplay of processes behind it. Moreover, we would also recommend that the results are complemented by experimental work whenever possible (Cadotte *et al.* 2013; Godoy *et al.* 2014). Community ecology represents one of the disciplines where patterns and processes have already been analyzed in relation to phylogenetic scale, illustrating the theoretical and empirical potential of the concept. Further advances on this front are certainly possible (e.g. multiple metrics across phylogenetic scales) and hold the promise of a more conclusive interpretation of community-level patterns and the processes behind them.

bioRxiv preprint doi: https://doi.org/10.1101/063560; this version posted July 18, 2016. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

Biogeography and niche conservatism

Biogeographic patterns, such as species distributions and diversity gradients, are largely shaped by the conservatism of the ecological niche (Wiens & Graham 2005; Wiens *et al.* 2010), which varies across the different dimensions of the niche and across phylogenetic scales (Freckleton *et al.* 2002; Wiens & Graham 2005). Instead of studying whether niches are conserved or not (Freckleton *et al.* 2002; Wiens & Graham 2005; Losos 2008; Wiens 2008), we should perhaps identify the scales over which they are conserved and study the imprints of these phylogenetic scales in biogeographic patterns.

Diversity gradients vary dramatically across taxa (Fig. 1), presumably because the taxa's climatic niches are conserved over different phylogenetic scales (Wiens & Donoghue 2004; Buckley *et al.* 2010). In mammals, many of the ancient lineages have failed to colonize high latitudes (e.g. treeshrews, sloths, armadillos) presumably because their physiological tolerances have been conserved over larger phylogenetic scales than those of the lineages that have not only invaded high latitudes but also accumulated most of their diversity there (e.g. rabbits and hares) (Buckley *et al.* 2010), producing reverse latitudinal gradients (e.g. Cook 1969; Owen & Owen 1974; Buckley *et al.* 2003; reviewed in Kindlman *et al.* 2007). The occasional breakdowns of niche conservatism, which typically span only a short period in the history of a clade and limited phylogenetic scales, sometimes precipitate diversification episodes that significantly enrich the diversity of the regional biota (e.g. African cichlids, Madagascan vangas, or ray-finned fishes and angiosperm plants) (Gavrilets & Losos 2009; Glor 2010; Jonsson *et al.* 2012; Rabosky *et al.* 2013). The phylogenetic scale over which niches are conserved, or break away from the conservatism, may consequently contribute to the formation of diversity patterns.

Diversity patterns may be further influenced by the effects of niche conservatism on regional extinctions (Cahill *et al.* 2012). Many genera whose climatic niches were conserved over phylogenetic scales that extended beyond the timeframe of the climatic changes during Pleistocene were wiped out by these changes (e.g. North American trees, European megafauna) (Stuart 1991; Jackson & Weng 1999). Yet, Pleistocene changes in climate have exterminated only few families, perhaps because climatic niches are less conserved at the family-level than at the genus-level (Freckleton *et al.* 2002; Blomberg *et al.* 2003). The extinction footprint of climate change therefore likely depends on the phylogenetic scale at which climatic niches are conserved.

bioRxiv preprint doi: https://doi.org/10.1101/063560; this version posted July 18, 2016. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

Evaluating scale-dependent vulnerability to extinction seems particularly relevant in the face of the on-going worldwide changes in climate and land use, and the results might provide insights into the patterns of loss of phylogenetic diversity. Even though it has long been recognized that niches are conserved to varying degrees, few studies have systematically investigated this variation across phylogenetic scales despite the promise that such an investigation could advance our understanding of diversity patterns.

Macroecology

Macroecologists, concerned mostly with statistical patterns across large spatial and temporal scales, rarely consider phylogenetic scale in their research. Yet, cross-scale comparisons can identify statistical patterns (e.g. latitudinal diversity gradient, body size distributions, species-area relationship, species-abundance distributions) that are truly universal and those that disintegrate over certain phylogenetic scales (Marquet *et al.* 2004; Storch & Šizling 2008). Phylogenetic scale may therefore inform us about the generality of statistical patterns in ecology and about the mechanisms (e.g. mathematical, geometric, random sampling, or biological) that likely produced them (Marquet *et al.* 2004; McGill 2008).

Some of the patterns originally considered to be universal have later been reported to disintegrate across phylogenetic scales. The latitudinal diversity gradient provides a very intuitive example, where the pattern holds across most higher taxa (e.g. mammals, birds, amphibians, reptiles, plants) but often breaks down across their constituent lower taxa that encompass limited phylogenetic scales (e.g. penguins, hares, aphids, ichneumonids, Proteacea) and produce a variety of diversity gradients, including reverse ones (e.g. Cook 1969; Owen & Owen 1974; Buckley *et al.* 2003; reviewed in Kindlman *et al.* 2007) (Fig. 1a). Likewise, species abundance and body mass are negatively correlated across birds and mammals (Damuth 1981), but the correlation disappears across narrowly defined taxa (Isaac *et al.* 2011) and becomes even positive in some tribes of birds (Cotgreave 1994) (Fig. 1b). These changes in correlation coefficients across phylogenetic scales implicate the mechanisms behind the correlation. Within large phylogenetic extents, small-bodied species are locally abundant because their low metabolic requirements raise the carrying capacities of their populations (Gaston and Blackburn 1997). However, within restricted extents, local abundance becomes constrained by competition

bioRxiv preprint doi: https://doi.org/10.1101/063560; this version posted July 18, 2016. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

between closely related species, and large-bodied species become locally abundant because of their competitive superiority, thus reversing the directionality of the correlation between body size and population abundance across phylogenetic scales (Cotgreave 1994) (Fig. 1b).

Moreover, the species-area relationship (SAR) and species-abundance distribution (SAD) were traditionally believed to universally conform to certain mathematical forms (the power-law function and the lognormal distribution, respectively) (Preston 1948; Rosenzweig 1995). However, if two sister clades follow power-law SARs and lognormal SADs which differ in their parameters, it can be proven mathematically that the clade containing both sister taxa cannot follow either the power-law SAR or the lognormal SAD (Storch & Šizling 2008; Šizling *et al.* 2009). Even though some of these patterns represent classic examples of ecological laws, cross-scale analyses indicate that they are not truly universal and sometimes provide insights into the biological mechanisms behind them.

The fact that some statistical patterns disintegrate across phylogenetic scales implies that the theories to explain these patterns, based on the universal principles of geometry and mathematics, might be fundamentally ill-founded (Storch & Šizling 2008). It is also possible that the theories pertain to certain phylogenetic scales only. This would suggest that phylogenetic scales form phylogenetic domains (Box 2) within which the processes hypothesized by our theories operate. However, the boundaries of these phylogenetic domains remain largely unexplored, and their empirical delimitation might further inform the theory (see Box 2).

bioRxiv preprint doi: https://doi.org/10.1101/063560; this version posted July 18, 2016. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

BOX 2: Research across phylogenetic scales

Many attributes, such as diversification rate, niche conservatism, or community structure, vary across phylogenetic scales (Table 1). They may vary in three different ways:

(a) Scale dependence refers to the situation when the studied attribute varies across phylogenetic scales without any obvious trend. In this case, the suitable scale of investigation should be defined a priori, based on the objective of the study. The results from one scale will be difficult to extrapolate to other scales.

(b) Scaling occurs when the attribute of interest varies systematically along the scale axis. The interpretation of scaling is at least threefold, depending on the underlying mechanism (note that only one of the interpretations is biological):

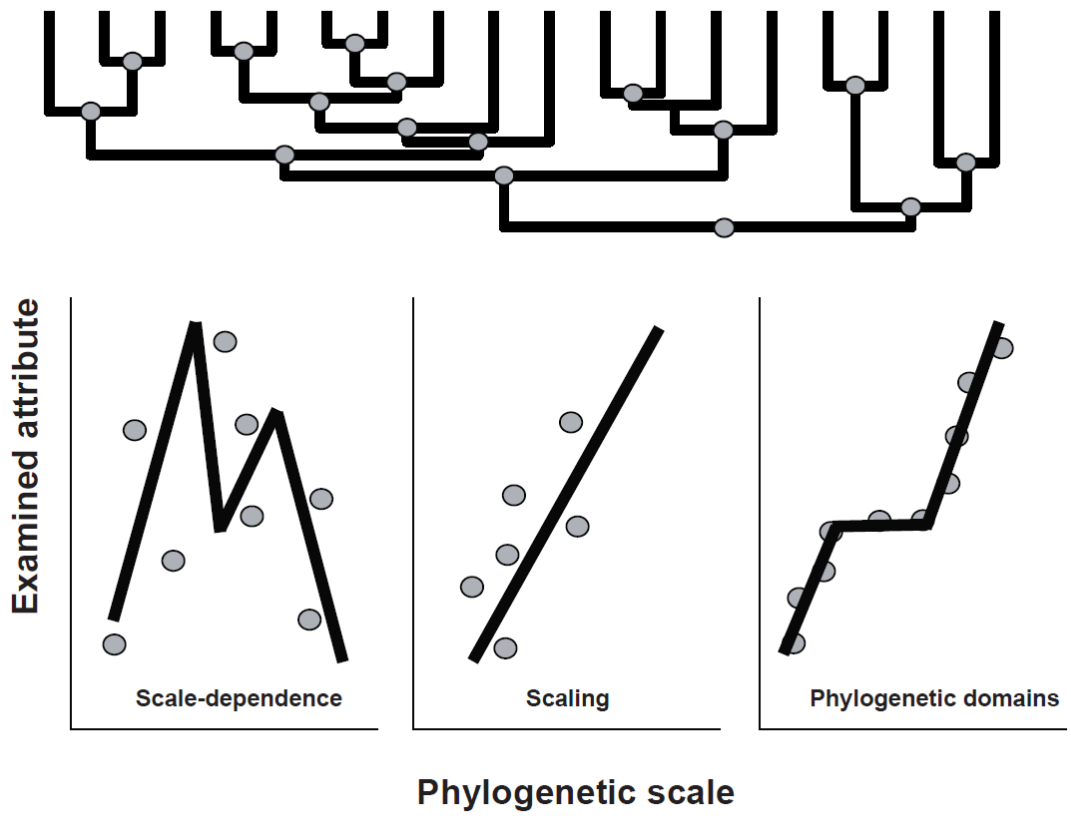
1. **Statistical scaling** is a sample-size effect whereby the statistical power of the analysis increases with clade size. Consequently, the attribute under study appears to change systematically from small clades to large clades (e.g. Machac et al. 2013). While the inferred values of the attribute itself may be technically correct, their systematic variation across scales is biologically irrelevant.
2. **Methodological artifacts** result when a statistical analysis becomes increasingly misleading toward the deep nodes of the phylogeny, resulting in incorrect and potentially biased estimates for the attribute of interest (e.g. ancestral reconstructions under dispersal-vicariance models tend to suggest that the ancestor occupied all of the regions examined) (Ronquist 1997). Methodological artifacts can be mitigated under various statistical corrections or when the results are validated using supplementary data, such as fossils.
3. **Phylogenetic scaling in the strict sense** occurs when the studied attribute changes across scales because the underlying biological process changes. True scaling can therefore inform us about the processes which generate the patterns observed across scales. If the scaling can be described mathematically, it allows to extrapolate across scales, even those not included in the original study, i.e. *downscale* or *upscale* the patterns under study.

(c) Domains of scale refer to the segments of the phylogeny (e.g. taxonomic units, tree depth, distinct clades) within which the attribute of interest appears relatively unchanged. The attribute might change abruptly between domains, indicating changes in the underlying biological

bioRxiv preprint doi: https://doi.org/10.1101/063560; this version posted July 18, 2016. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

processes. Therefore, it should be possible to extrapolate across phylogenetic scales within domains, but not across scales between them.

FIGURE (BOX 2): Numerous attributes can be studied across phylogenetic scales. These may include diversification measures, statistical relationships between ecological variables, parameters of frequency distributions, metrics that describe community phylogenetic structure, or measures of niche conservatism (see Table 1). Phylogenetic scale can be defined in terms of clade age, clade size, taxonomic rank, the degree of molecular or phenotypic divergence, etc., depending on the question under study.



bioRxiv preprint doi: https://doi.org/10.1101/063560; this version posted July 18, 2016. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

Table 1. Ecological and evolutionary attributes that vary across phylogenetic scales. Examples of methods to evaluate these attributes are also indicated.

Field	Examined attribute	Examples of methods for evaluation
Evolution and diversification	diversification mode	coalescent inference to distinguish between accelerations, slowdowns, and saturation (Morlon *et al.* 2010)
	diversification rate	product-moment estimators (Magallon & Sanderson 2001), equal-splits measures (Jetz *et al.* 2012)
	slowdown strength	gamma statistic (Pybus & Harvey 2000)
Community ecology	community structure	phylometrics (NRI, NTI, MNND, MPD) (Webb *et al.* 2002)
	phylogenetic diversity	Faith's PD (Faith 1992)
Biogeography and macroecology	relationship form	linear, polynomial, exponential, lognormal functions
	relationship strength	Pearson's correlation, Spearman's correlation, regression slope
Niche conservatism	phylogenetic signal	Pagel's lambda (Freckleton *et al.* 2002), Blomberg's K (Blomberg *et al.* 2003)
	evolutionary rates	Brownian motion model (Felsenstein 1985), Ornstein-Uhlenbeck model (Hansen 1997) of trait evolution

bioRxiv preprint doi: https://doi.org/10.1101/063560; this version posted July 18, 2016. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

PHYLOGENETIC SCALE IN PRACTICE

The above overview demonstrates that the consideration of phylogenetic scale varies across fields, both in terms of the approach and the vocabulary. Therefore, it seems of value to find a common language to discuss and study phylogenetic scale across disciplines. There are two general approaches with which phylogenetic scale can be considered in ecological and evolutionary research. One is exploratory, where patterns are identified across a range of phylogenetic scales and then explained in the light of specific events or mechanisms. The other approach relies on testing a priori hypotheses, which are based on mechanisms that presumably take place at a given phylogenetic scale. Both approaches have their strengths and either may be appropriate, depending on the objective of a given study; however, we advocate the hypotheses testing approach for most questions.

To study the effects of phylogenetic scale, one can evaluate how a specific attribute of interest (such as diversification rate, niche conservatism, geographic distribution, statistical relationships) changes with phylogenetic scale. These attributes may vary randomly or systematically across the phylogeny, be more prevalent at particular scales, or stay unchanged across a discrete set of mutually nested clades (Box 2). We refer to the latter as a domain of phylogenetic scale which, in analogy to spatial domains (Wiens 1989), corresponds to a segment of phylogeny that reveals homogeneity in the attribute of interest. In this section, we consider conceptual and methodological approaches to explore patterns which are phylogenetic scale-dependent.

Choice of phylogenetic scale

While most researchers are aware that the choice of scale can influence inferences about patterns or processes, all too often the choice of scale, be it spatial, temporal or phylogenetic, is influenced by data availability or other logistical concerns. Instead, the scale of an investigation should be chosen based on a specific objective or question whenever possible. For example, phylogenies of higher taxa may not be appropriate for evaluating the processes of community assembly that typically take place at small phylogenetic scales. To test the hypothesis that competition reduces species coexistence, for example, small phylogenetic scales (e.g. genera, or generally clades where species can reasonably compete) should be preferred to large scales where most species are

bioRxiv preprint doi: https://doi.org/10.1101/063560; this version posted July 18, 2016. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

highly unlikely to compete (e.g. the entire classes, such as birds and mammals). However, even with a specific question at hand, it can be difficult to choose a single most appropriate phylogenetic scale. Therefore, evaluating multiple phylogenetic extents or grains should be considered.

Multiple phylogenetic scales

Simultaneous consideration of multiple phylogenetic scales may be particularly important in large phylogenies because different clades within such phylogenies may show different patterns with respect to the attribute of interest (e.g. diversification rate, the strength of niche conservatism, patterns of community phylogenetic structure) (Fig. 1). For example, Cetacean systematists had long been perplexed as to why there is little correspondence between diversification dynamics estimated from the fossil record and phylogenetic trees (Quental & Marshall 2010; Slater *et al.* 2010; Morlon *et al.* 2011). The correspondence between the two datasets emerged only when diversification dynamics were evaluated independently for clades within cetaceans (whales, dolphins, and porpoises) as opposed to cetaceans as a whole. In this case, each clade appeared to have its own dynamics which were obscured when the entire tree was evaluated (Morlon *et al.* 2011).

In some cases, it may be difficult or even undesirable to specify, a priori, a specific set of scales. It might be instead more illuminating to study how the attribute of interest varies across an inclusive range of scales. There are several approaches, originating in community phylogenetics, which allow for such cross-scale analyses and return results for each node of the phylogenetic tree (Leibold *et al.* 2010; Pavoine *et al.* 2010; Borregaard *et al.* 2014). For example, the method developed by Borregaard *et al.* (2014) identifies nodes whose descendant clades underwent conspicuous geographic, phenotypic, or ecological shifts. In evolutionary research, evaluation of all nodes is not uncommon, and multiple tools have been developed to identify node-specific shifts in diversification rates and clades with conspicuously fast or slow diversification (Alfaro *et al.* 2009; Rabosky 2014) (Table 1). However, statistical analyses that would include all nodes of the phylogeny remain relatively scarce (e.g. Machac et al. 2012; Machac et al. 2013), and most studies analyze select clades only, despite the often cited concerns that the

bioRxiv preprint doi: https://doi.org/10.1101/063560; this version posted July 18, 2016. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

selection of clades is rarely random, reflects our prior biases, and might influence the analysis profoundly (e.g. Phillimore & Price 2008; Cusimano & Renner 2010; Pennell *et al.* 2012).

Two potential issues associated with the evaluation of all nodes are data non-independence and nestedness. Non-independence can be readily accommodated by the widely used comparative methods (e.g. PIC, PGLS) (Hurlbert 1984; Felsenstein 1985; Freckleton *et al.* 2002; Rohlf 2006). These methods typically estimate the same parameters as their conventional counterparts (e.g. intercepts, regression slopes, group means) but adjust the confidence intervals for these parameters based on the inferred degree of phylogenetic correlation in the data (Hurlbert 1984; Felsenstein 1985; Freckleton *et al.* 2002; Rohlf 2006). The nestedness of the data is more difficult to accommodate. For example, the diversification rate of a clade is inherently determined by the rate values across its constituent subclades. Nestedness therefore extends beyond the phylogenetic correlation of rate values and reflects how the value for a clade is produced by the subclade values. This information cannot be readily accommodated under the currently available comparative methods whose phylogenetic corrections consequently cannot guarantee proper estimates of statistical significance across nested data. For these reasons, we argue that parameter estimates can be extracted, compared, and analyzed across nested clades, but their significance needs to be interpreted cautiously. New theory that would illuminate how different attributes of interest (e.g. diversification rates, regression slopes, phylogenetic signal) combine and compound across nested hierarchies, as well as the methods to capture these correlations, are clearly needed.

Phylogenetic scaling

Statistical methods that evaluate all clades (nodes) in a given phylogeny (Leibold *et al.* 2010; Borregaard *et al.* 2014; Rabosky 2014) can be used to explore phylogenetic scaling. Scaling is a systematic trend along the scale axis in the attribute of interest. For example, diversification rate or net relatedness index (NRI) (Webb *et al.* 2002) may change systematically with increasing phylogenetic scale (Cavender-Bares *et al.* 2009; Vamosi *et al.* 2009).

Phylogenetic scaling should be most prevalent across mutually nested clades because the patterns associated with larger clades are determined by the patterns of clades nested within them (or vice versa). For example, diversification rate of a clade is determined by the rate values of its

bioRxiv preprint doi: https://doi.org/10.1101/063560; this version posted July 18, 2016. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

subclades, similarly as species richness of a spatial plot is determined by the richness of its subplots. Consequently, it should be possible to predict the value of an attribute (e.g. diversification rate, regression slopes, phylogenetic signal) at a particular phylogenetic scale from the knowledge of those values across other scales, much like it is possible to estimate species richness within large geographic areas, based on the knowledge of richness within small areas (Harte *et al.* 2009; Storch *et al.* 2012). When characterized mathematically, phylogenetic scaling should allow for predictions across phylogenetic scales not covered by the phylogeny at hand (i.e. upscaling or downscaling).

Domains of phylogenetic scale

When moving along the scale axis, the values of an attribute might sometimes change abruptly. Such discontinuities provide the opportunity to delimit the domains of phylogenetic scale (Box 2). Domains are discrete segments of a phylogeny, such as monophyletic clades, taxonomic ranks, or tree depth, which show homogeneity in the attribute of interest (i.e. diversification rate, statistical correlation, or phylogenetic signal). By definition, the attribute stays largely unchanged within a domain but varies substantially between domains. Phylogenetic domains may therefore provide insights into the processes which operate over different segments of a phylogenetic tree.

Traditionally, phylogenetic domains were delimited by taxonomists whose objective was to organize species into biologically meaningful units, such as families, orders, or classes. These units are based mostly on morphological and ecological attributes. However, phylogenetic domains can also consist of clades that show diversification homogeneity, similar rates of morphological evolution, or similar life-history trade-offs. Therefore, the domains may be delimited based on key innovations, episodes of historical dispersals, or extinction events, but also statistically, using quantitative methods without the prior knowledge of the evolutionary history of a clade. While the statistical approach may be more transparent and reproducible, the resulting domains may be harder to interpret biologically. Nonetheless, statistically delimited domains may reveal otherwise unnoticed evolutionary events and potentially important breaks in the clade's history that may have shaped its extant diversity.

Phylogenetic domains may further facilitate statistical inference, given that most comparative methods assume that the analyzed attributes are largely homogeneous (e.g.

bioRxiv preprint doi: https://doi.org/10.1101/063560; this version posted July 18, 2016. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

regression slopes do not vary across genera within the analyzed family, diversification is homogenous across the analyzed lineages) and return spurious results when applied to clades that show a mixture of patterns and processes (Morlon *et al.* 2011; O'Meara 2012; Rabosky & Goldberg 2015). Phylogenetic domains may therefore identify when comparative methods report reasonably reliable results and when their conclusions must be interpreted with caution because the results span different domains and the underlying assumptions have been violated.

CONCLUSION

It is well established that different processes dominate over different spatial and temporal scales. Phylogenetic scale, however, has received limited attention although much research in ecology and evolution relies on molecular phylogenies (Table 1). Explicit consideration of phylogenetic scale, scale dependence, phylogenetic scaling, and the domains of phylogenetic scale can therefore inform multiple disciplines in the field (e.g. diversification analysis, community ecology, biogeography and macroecology).

We have discussed phylogenetic scale largely in isolation from spatial and temporal scales, but these types of scale will often be related. For instance, competitive exclusion may be prominent among closely related species within local communities over short time periods (Cavender-Bares *et al.* 2009). In contrast, plate tectonics might influence deeper nodes in a phylogeny and operate over broad geographic and temporal scales (Ricklefs 1985; Willis & Whittaker 2002). In some notable cases, however, the scales may not be related. Diversity anomalies, such as New Caledonia or Madagascar, represent examples of decoupling where rich biotas that encompass extensive phylogenetic scales diversified in a relatively small region (Warren *et al.* 2010; Espeland & Murienne 2011). In contrast, recent radiations within grasses and rodents have had a large geographic footprint but encompass only limited phylogenetic scales (Edwards et al. 2010). Evaluating when different types of scale are coupled (or decoupled) may yield new insights into the evolutionary history of different clades and regions (Willis & Whittaker 2002).

We hope that the perspective presented here will spur further theoretical, empirical, and methodological research. Explicit consideration of phylogenetic scale may turn our focus away from the importance of particular mechanisms (diversification, trait evolution, niche

bioRxiv preprint doi: https://doi.org/10.1101/063560; this version posted July 18, 2016. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

conservatism) toward the appreciation for the interplay of multiple processes which together, but over different phylogenetic scales, shape the diversity of life.

ACKNOWLEDGEMENTS

Funding was provided by the NSF program Dimensions of Biodiversity (DEB-1136586) and by the Grant Agency of the Czech Republic (14-36098G).

REFERENCES

Ackerly, D. D., D. W. Schwilk, and C. O. Webb. 2006. Niche evolution and adaptive radiation: testing the order of trait divergence. Ecology 87:50–61.

Alfaro, M. E., F. Santini, C. Brock, H. Alamillo, A. Dornburg, D. L. Rabosky, G. Carnevale, et al. 2009. Nine exceptional radiations plus high turnover explain species diversity in jawed vertebrates. Proceedings of the National Academy of Sciences of the United States of America 106:13410–13414.

Benton, M. J., and B. C. Emerson. 2007. How did life become so diverse? The dynamics of diversification according to the fossil record and molecular phylogenetics. Palaeontology 50:23–40.

Blomberg, S. P., T. Garland, and A. R. Ives. 2003. Testing for phylogenetic signal in comparative data: behavioral traits are more labile. Evolution 57:717–745.

Borregaard, M. K., C. Rahbek, J. Fjeldså, J. L. Parra, R. J. Whittaker, and C. H. Graham. 2014. Node-based analysis of species distributions. Methods in Ecology and Evolution 5:1225–1235.

Buckley, H. L., T. E. Miller, A. M. Ellison, and N. J. Gotelli. 2003. Reverse latitudinal trends in species richness of pitcher-plant food webs. Ecology Letters 6:825–829.

Buckley, L. B., T. J. Davies, D. D. Ackerly, N. J. B. Kraft, S. P. Harrison, B. L. Anacker, H. V. Cornell, et al. 2010. Phylogeny, niche conservatism and the latitudinal diversity gradient in mammals. Proceedings of the Royal Society of London B: Biological Sciences: rspb20100179.

Cadotte, M., C. H. Albert, and S. C. Walker. 2013. The ecology of differences: assessing community assembly with trait and evolutionary distances. Ecology Letters 16:1234–1244.

Cahill, A. E., M. E. Aiello-Lammens, M. C. Fisher-Reid, X. Hua, C. J. Karanewsky, H. Y. Ryu, G. C. Sbeglia, et al. 2012. How does climate change cause extinction? Proceedings of the Royal Society B: Biological Sciences: rspb20121890.

Cavender-Bares, J., A. Keen, and B. Miles. 2006. Phylogenetic structure of Floridian plant communities depends on taxonomic and spatial scale. Ecology 87:S109–122.

Cavender-Bares, J., K. H. Kozak, P. V. A. Fine, and S. W. Kembel. 2009. The merging of community ecology and phylogenetic biology. Ecology Letters 12:693–715.

Cook, R. E. 1969. Variation in species density of North American birds. Systematic Zoology 18:63–84.

Cornell, H. V., and S. P. Harrison. 2014. What are species pools and when are they important? Annual Review of

bioRxiv preprint doi: https://doi.org/10.1101/063560; this version posted July 18, 2016. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

Ecology, Evolution, and Systematics 45:45–67.

Cotgreave, P., and P. Stockley. 1994. Body size, insectivory and abundance in assemblages of small mammals. Oikos 71:89–96.

Damuth, J. 1981. Population density and body size in mammals. Nature 290:699–700.

Daniel L Rabosky, G. J. S. 2012. Clade age and species richness are decoupled across the Eukaryotic Tree of Life. PLoS biology 10:e1001381.

Edwards, E. J., C. P. Osborne, C. A. E. Strömberg, S. A. Smith, C4 Grasses Consortium, W. J. Bond, P.-A. Christin, et al. 2010. The origins of C4 grasslands: integrating evolutionary and ecosystem science. Science 328:587–591.

Espeland, M., and J. Murienne. 2011. Diversity dynamics in New Caledonia: towards the end of the museum model? BMC Evolutionary Biology 11:254.

Felsenstein, J. 1985. Phylogenies and the comparative method. American Naturalist 125:1–15.

Freckleton, R. P., P. H. Harvey, and M. Pagel. 2002. Phylogenetic analysis and comparative data: a test and review of evidence. American Naturalist 160:712–726.

Gaston, K. J., T. M. Blackburn, and J. H. Lawton. 1997. Interspecific Abundance-Range Size Relationships: An Appraisal of Mechanisms. Journal of Animal Ecology 66:579–601.

Gavrilets, S., and J. B. Losos. 2009. Adaptive radiation: Contrasting theory with data. Science 323:732–737.

Gerhold, P., J. F. Cahill, M. Winter, I. V. Bartish, and A. Prinzing. 2015. Phylogenetic patterns are not proxies of community assembly mechanisms (they are far better). Functional Ecology 29:600–614.

Glor, R. E. 2010. Phylogenetic insights on adaptive radiation. Annual Review of Ecology, Evolution, and Systematics 41:251–270.

Godoy, O., N. J. B. Kraft, and J. M. Levine. 2014. Phylogenetic relatedness and the determinants of competitive outcomes. Ecology Letters 17:836–844.

Gould SJ. 2002. The Structure of evolutionary theory. Harvard University Press, Cambridge, MA.

Hansen, T. F. 1997. Stabilizing selection and the comparative analysis of adaptation. Evolution 51:1341–1351.

Harmon, L. J., and S. Harrison. 2015. Species diversity is dynamic and unbounded at local and continental scales. American Naturalist 185:584–593.

Harte, J., A. B. Smith, and D. Storch. 2009. Biodiversity scales from plots to biomes with a universal species–area curve. Ecology Letters 12:789–797.

Heath, T. A., J. P. Huelsenbeck, and T. Stadler. 2014. The fossilized birth–death process for coherent calibration of divergence-time estimates. Proceedings of the National Academy of Sciences 111:2957–2966.

Hurlbert, S. H. 1984. Pseudoreplication and the design of ecological field experiments. Ecological Monographs 54:187–211.

Isaac, N. J. B., D. Storch, and C. Carbone. 2011. Taxonomic variation in size–density relationships challenges the notion of energy equivalence. Biology Letters 7:615–618.

IUCN 2016. The IUCN Red List of Threatened Species.

Jackson, S. T., and C. Weng. 1999. Late Quaternary extinction of a tree species in eastern North America. Proceedings

bioRxiv preprint doi: https://doi.org/10.1101/063560; this version posted July 18, 2016. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

of the National Academy of Sciences of the United States of America 96:13847–13852.

Lessard, J. P. 2012. Inferring local ecological processes amid species pool influences. Trends in Ecology & Evolution 27:600–607.

Jones, K. E., J. Bielby, M. Cardillo, S. A. Fritz, J. O'Dell, C. D. L. Orme, K. Safi, et al. 2009. PanTHERIA: a species-level database of life history, ecology, and geography of extant and recently extinct mammals. Ecology 90:2648–2648.

Jønsson, K. A., P.-H. Fabre, S. A. Fritz, R. S. Etienne, R. E. Ricklefs, T. B. Jørgensen, J. Fjeldså, et al. 2012. Ecological and evolutionary determinants for the adaptive radiation of the Madagascan vangas. Proceedings of the National Academy of Sciences of the United States of America 109:6620–6625.

Kennedy, J. D., J. T. Weir, D. M. Hooper, D. T. Tietze, J. Martens, and T. D. Price. 2012. Ecological limits on diversification of the Himalayan Core Corvoidea. Evolution 66:2599–2613.

Kindlman, P., Schodelbauerova, I. & Dixon, A.F.G. (2007). Inverse latitudinal gradients in species diversity. In: Scaling Biodiversity, eds. Storch, D., Marquet, P.A. & Brown, J. H. Cambridge University Press, Cambridge.

Kindlman, Schodelbauerova, and Dixon. 2007. Inverse latitudinal gradients in species diversity. In: Scaling Biodiversity, ed. David Storch, Pablo A. Marquet and James H. Brown. Cambridge University Press.

Kozak, K. H., and J. J. Wiens. 2010. Accelerated rates of climatic-niche evolution underlie rapid species diversification. Ecology Letters 13:1378–1389.

Laland, K., T. Uller, M. Feldman, K. Sterelny, G. B. Müller, A. Moczek, E. Jablonka, et al. 2014. Does evolutionary theory need a rethink? Nature 514:161–164.

Leibold, M. A., E. P. Economo, and P. Peres-Neto. 2010. Metacommunity phylogenetics: separating the roles of environmental filters and historical biogeography. Ecology Letters 13:1290–1299.

Levin, S. A. 1992. The Problem of Pattern and Scale in Ecology: The Robert H. MacArthur Award Lecture. Ecology 73:1943–1967.

Losos, J. B. 2008. Phylogenetic niche conservatism, phylogenetic signal and the relationship between phylogenetic relatedness and ecological similarity among species. Ecology Letters 11:995–1003.

Machac, A., Graham, C.H. & Storch, D. Ecological controls of mammalian diversification vary with phylogenetic scale, in review.

Machac A., Storch D., Wiens J. J. (2013) Ecological causes of decelerating diversification in carnivoran mammals. Evolution 67: 2423-2433.

Machac A., Storch D., Zrzavy J., Smrckova J. (2012) Temperature dependence of evolutionary diversification: differences between two contrasting model taxa support the metabolic theory of ecology. Journal of Evolutionary Biology 25: 2449-2456.

Marquet, P.A., M. Fernández, S.A. Navarrete & C. Valdovinos (2004) Diversity emerging: Towards a deconstruction of biodiversity patterns. In: Frontiers of Biogeography: New Directions in the Geography of Nature. Cambridge University Press, Cambridge.

May, R. M. 1974 Patterns of species abundance and diversity. Harvard University Press.

Mayr E. 1942. Systematics and the origin of species. Dover, New York.

bioRxiv preprint doi: https://doi.org/10.1101/063560; this version posted July 18, 2016. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

McGill, B. J., A. E. K. Roy, and E. M. C. Whitlock. 2008. Exploring Predictions of Abundance from Body Mass Using Hierarchical Comparative Approaches. American Naturalist 172:88–101.

McGuire, J. A., C. C. Witt, J. V. Remsen Jr., A. Corl, D. L. Rabosky, D. L. Altshuler, and R. Dudley. 2014. Molecular phylogenetics and the diversification of hummingbirds. Current Biology 24:910–916.

McPeek, M. A. 2008. The ecological dynamics of clade diversification and community assembly. American Naturalist 172:270–284.

Morlon, H., T. L. Parsons, and J. B. Plotkin. 2011. Reconciling molecular phylogenies with the fossil record. Proceedings of the National Academy of Sciences 108:16327–16332.

Morlon, H., M. D. Potts, and J. B. Plotkin. 2010. Inferring the dynamics of diversification: a coalescent approach. PLoS biology 8.

Münkemüller, T., L. Gallien, S. Lavergne, J. Renaud, C. Roquet, S. Abdulhak, S. Dullinger, et al. 2014. Scale decisions can reverse conclusions on community assembly processes. Global ecology and biogeography 23:620–632.

Munoz, F., B. R. Ramesh, and P. Couteron. 2014. How do habitat filtering and niche conservatism affect community composition at different taxonomic resolutions? Ecology 95:2179–2191.

Ndiribe, C., L. Pellissier, S. Antonelli, A. Dubuis, J. Pottier, P. Vittoz, A. Guisan, et al. 2013. Phylogenetic plant community structure along elevation is lineage specific. Ecology and Evolution 3:4925–4939.

Nilsson, M. A., U. Arnason, P. B. S. Spencer, and A. Janke. 2004. Marsupial relationships and a timeline for marsupial radiation in South Gondwana. Gene 340:189–196.

O'Meara, B. C. 2012. Evolutionary inferences from phylogenies: a review of methods. Annual Review of Ecology, Evolution, and Systematics 43:267–285.

Owen, D., and J. Owen. 1974. Species diversity in temperate and tropical Ichneumonidae. Nature 249:583–584.

PanTHERIA: A species-level database of life history, ecology, and geography of extant and recently extinct mammals. Ecology 90:2648

Parra, J. L., J. A. McGuire, and C. H. Graham. 2010. Incorporating clade identity in analyses of phylogenetic community structure: an example with hummingbirds. American Naturalist 176:573–587.

Parra, J. L., C. Rahbek, J. A. McGuire, and C. H. Graham. 2011. Contrasting patterns of phylogenetic assemblage structure along the elevational gradient for major hummingbird clades. Journal of Biogeography 38:2350–2361.

Pavoine, S., M. Baguette, and M. B. Bonsall. 2010. Decomposition of trait diversity among the nodes of a phylogenetic tree. Ecological Monographs 80:485–507.

Pigliucci, M. 2007. Do we need an extended evolutionary synthesis? Evolution 61:2743–2749.

Preston, F. W. 1948. The commonness, and rarity of species. Ecology 29:254–283.

Price, T. D., D. M. Hooper, C. D. Buchanan, U. S. Johansson, D. T. Tietze, P. Alström, U. Olsson, et al. 2014. Niche filling slows the diversification of Himalayan songbirds. Nature 509:222–225.

Quental, T. B., and C. R. Marshall. 2010. Diversity dynamics: molecular phylogenies need the fossil record. Trends in Ecology & Evolution 25:434–441.

———. 2013. How the Red Queen drives terrestrial mammals to extinction. Science 341:290–292.

bioRxiv preprint doi: https://doi.org/10.1101/063560; this version posted July 18, 2016. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

Rabosky, D. L. 2009. Ecological limits and diversification rate: alternative paradigms to explain the variation in species richness among clades and regions. Ecology Letters 12:735–743.

———. 2014. Automatic detection of key innovations, rate shifts, and diversity-dependence on phylogenetic trees. PLoS ONE 9:e89543.

Rabosky, D. L., and E. E. Goldberg. 2015. Model inadequacy and mistaken inferences of trait-dependent speciation. Systematic Biology syu131.

Rabosky, D. L., and A. H. Hurlbert. 2015. Species richness at continental scales is dominated by ecological limits. The American Naturalist 185:572–583.

Rabosky, D. L., and D. R. Matute. 2015. Macroevolutionary speciation rates are decoupled from the evolution of intrinsic reproductive isolation in Drosophila and birds.

Rabosky, D. L., F. Santini, J. Eastman, S. A. Smith, B. Sidlauskas, J. Chang, and M. E. Alfaro. 2013. Rates of speciation and morphological evolution are correlated across the largest vertebrate radiation. Nature Communications 4: 2958-2961.

Rabosky, D. L., G. J. Slater, and M. E. Alfaro. 2012. Clade age and species richness are decoupled across the eukaryotic tree of life. PLoS biology 10:e1001381.

Rohlf, F. J. 2001. Comparative methods for the analysis of continuous variables: geometric interpretations. Evolution 55:2143–2160.

Rohlf, F. J. 2006. A comment on phylogenetic correction. Evolution 60:1509–1515.

Ronquist, F. 1997. Dispersal-vicariance analysis: a new approach to the quantification of historical biogeography. Systematic Biology 46:195–203.

Rosenzweig. 1995. Species diversity in space and time. Cambridge University Press.

Scheiner, S. M. 2000. Species richness, species–area curves and Simpson's paradox. Evolutionary Ecology Research 2:791–802.

Schneider, D. C. 2001. The rise of the concept of scale in ecology the concept of scale is evolving from verbal expression to quantitative expression. BioScience 51:545–553.

Šizling, A. L., W. E. Kunin, E. Šizlingová, J. Reif, and D. Storch. 2011. Between geometry and biology: the problem of universality of the species-area relationship. American Naturalist 178:602–611.

Slater, G. J., S. A. Price, F. Santini, and M. E. Alfaro. 2010. Diversity versus disparity and the radiation of modern cetaceans. Proceedings of the Royal Society of B: Biological Sciences 277:3097–3104.

Storch, D., P. Keil, and W. Jetz. 2012. Universal species-area and endemics-area relationships at continental scales. Nature 488:78–81.

Storch, D., and A. L. Šizling. 2008. The concept of taxon invariance in ecology: Do diversity patterns vary with changes in taxonomic resolution? Folia Geobotanica 43:329–344.

Stuart, A. J. 1991. Mammalian extinctions in the late Pleistocene of northern Eurasia and North America. Biological Reviews of the Cambridge Philosophical Society 66:453–562.

Swenson, N. G., B. J. Enquist, J. Pither, J. Thompson, and J. K. Zimmerman. 2006. The problem and promise of scale

bioRxiv preprint doi: https://doi.org/10.1101/063560; this version posted July 18, 2016. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

dependency in community phylogenetics. Ecology 87:2418–2424.

Swenson, N. G., B. J. Enquist, J. Thompson, and J. K. Zimmerman. 2007. The influence of spatial and size scale on phylogenetic relatedness in tropical forest communities. Ecology 88:1770–1780.

Vamosi, J. C., and S. M. Vamosi. 2010. Key innovations within a geographical context in flowering plants: towards resolving Darwin's abominable mystery. Ecology Letters 13:1270–1279.

Vamosi, S. M., S. B. Heard, J. C. Vamosi, and C. O. Webb. 2009. Emerging patterns in the comparative analysis of phylogenetic community structure. Molecular Ecology 18:572–592.

Warren, B. H., D. Strasberg, J. H. Bruggemann, R. P. Prys-Jones, and C. Thébaud. 2010. Why does the biota of the Madagascar region have such a strong Asiatic flavour? Cladistics 26:526–538.

Webb, C. O., D. D. Ackerly, M. A. McPeek, and M. J. Donoghue. 2002. Phylogenies and community ecology. Annual Review of Ecology and Systematics 33:475–505.

Wiens, J. A. 1989. Spatial scaling in ecology. Functional Ecology 3:385–397.

Wiens, J. J. 2008. Commentary on Losos (2008): Niche conservatism deja vu. Ecology Letters 11:1004–1005.

———. 2011. The causes of species richness patterns across space, time, and clades and the role of ecological limits. The Quarterly Review of Biology 86:75–96.

Wiens, J. J., D. D. Ackerly, A. P. Allen, B. L. Anacker, L. B. Buckley, H. V. Cornell, E. I. Damschen, et al. 2010. Niche conservatism as an emerging principle in ecology and conservation biology. Ecology Letters 13:1310–1324.

Wiens, J. J., and M. J. Donoghue. 2004. Historical biogeography, ecology and species richness. Trends in Ecology & Evolution 19:639–644.

Wiens, J. J., and C. H. Graham. 2005. Niche conservatism: integrating evolution, ecology, and conservation biology. Annual Review of Ecology, Evolution, and Systematics 36:519–539.

Willis, K. J., and R. J. Whittaker. 2002. Species diversity - scale matters. Science 295:1245–1248.