Transition from environmental to partial genetic sex determination in *Daphnia* through the evolution of a female-determining incipient W chromosome.

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Running title: Partial genetic sex determination in *D. magna*.

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Abstract

Sex chromosomes can evolve during the evolution of genetic sex determination (GSD) from environmental sex determination (ESD). Despite theoretical attention, early mechanisms involved in the transition from ESD to GSD have yet to be studied in nature. No mixed ESD-GSD animal species have been reported, except for some species of *Daphnia*, small freshwater crustaceans in which sex is usually determined solely by the environment, but in which a dominant female sex-determining locus is present in some populations. This locus follows Mendelian single-locus inheritance, but has otherwise not been characterized genetically. We now show that the sex-determining genomic region maps to the same low-recombining peri-centromeric region of linkage group 3 (LG3) in three highly divergent populations of *D. magna*, and spans 3.6 Mb. Despite low levels of recombination, the associated region contains signs of historical recombination, suggesting a role for selection acting on several genes thereby maintaining linkage disequilibrium among the 36 associated SNPs. The region carries numerous genes involved in sex differentiation in other taxa, including *transformer2* and *sox9*. Taken together, the region determining the NMP phenotype shows characteristics of a sex-related supergene, suggesting that LG3 is potentially an incipient W chromosome despite the lack of significant additional restriction of recombination between Z and W. The occurrence of the female-determining locus in a pre-existing low recombining region illustrates one possible form of recombination suppression in sex chromosomes. *D. magna* is a promising model for studying the evolutionary transitions from ESD to GSD and early sex chromosome evolution.

21 # Introduction

22 Sex chromosomes have evolved independently multiple times in many taxa (Miura et al.
23 2008; Pokorná and Kratochvíl 2009; Stöck et al. 2011; Cheng et al. 2013; Tree of Sex
24 Consortium 2014). Two evolutionary routes are thought to lead to the evolution of genetic sex
25 determination, and sometimes of sex chromosomes. First, separate sexes may evolve from
26 hermaphroditism, most likely through a male-sterility (i.e. female determining) mutation creating
27 a breeding system called gynodioecy, with genetic females co-occurring with functional
28 hermaphrodites. An initial male-determining mutation is also possible, but is less likely
29 (Charlesworth and Charlesworth 1978). The sex-determining mutation can be favoured through
30 causing obligate outcrossing, if inbreeding depression is severe, or if there is a fitness
31 disadvantage to investing resources in both male and female functions compared to investing in
32 only one sex (Charlesworth and Charlesworth 1978; Innes and Dunbrack 1993). Additional
33 mutations in genes linked to the sex-determining locus may then be favoured if their effects are
34 sexually antagonistic (i.e. have opposite effects in the two sexes; Rice 1987; Ellegren 2011). In
35 the case of a female sex-determining locus, additional mutations include ones benefiting males,
36 and deleterious in females (including female-sterility mutations creating males), which will often
37 be eliminated unless they are linked to the femaleness-determining gene. Closer linkage will then
38 be favored in this genomic region (Bull, 2006), which may lead to suppression of recombination.
39 Eventually, this may result in a system with "proto-sex chromosomes" carrying linked genes that
40 determine both sexes genetically (with male and female determining mutations on opposite
41 homologs).

42 The second route towards evolving GSD and sex chromosomes may start from
43 environmental sex determination (ESD), which could be the ancestral state in several major

animal groups (Ohno 1967; Pokorná and Kratochvíl 2016). Although transition from ESD to GSD may be gradual, involving shifting genotype-specific thresholds for male vs. female development under fluctuating environmental conditions (Van Dooren and Leimar 2003), a scenario similar to that of the transition from hermaphroditism to separate sexes is also plausible: a female-determining mutation in a population with pure ESD could lead to a system in which females are genetically determined, while other individuals have ESD (the route through an initial male-determining mutation is also possible). Such a sex-determining mutation can be favoured if it restores a 1:1 sex ratio of the population, for example, after environmental conditions change (Edwards 1998), or through other advantages like those outlined for the first pathway above.

The evolution of suppressed recombination between sex chromosomes and more generally, the early stages of sex chromosome evolution remain poorly understood (Wright et al. 2016). Because of their intermediate position between hermaphroditic and dioecious species, gynodioecious plants might have been good candidates to study these early stages. However, they proved not to be very informative in this respect, because gynodioecy is often controlled by cyto-nuclear interactions between mitochondrial male sterility mutations and nuclear "restorer" genes (McCauley and Bailey 2009; Beukeboom and Perrin 2014). Only a few species appear to have pure nuclear control and may therefore conform to the above theory of sex chromosome evolution (Kohn 1988; Connor and Charlesworth 1989; Weller and Sakai 1991; Spigler et al. 2008). Species in transition from ESD to GSD are therefore of particular interest, although the transition may be rapid (Pokorná and Kratochvíl 2009), so that few systems are available for study.

In some populations of *Daphnia*, pure ESD individuals co-occur with genetically determined females (Innes and Dunbrack 1993, Tessier and Caceres 2004, Galimov et al. 2011).

68 *Daphnia* are cyclical parthenogens, in which clonal reproduction with live born offspring is
69 interspersed with sexual reproduction phases producing diapause stages. Sex determination is
70 usually environmental (Hobaek and Larson 1990, Zhang and Baer 2000, LeBlanc and Medlock
71 2015), with cues differing among populations (Roulin et al. 2013). In nature, male development
72 of the clonal offspring present in the ovaries may be elicited by the mother emitting a juvenile
73 hormone (JH) or a JH pathway-related molecule (Olmstead and Leblanc 2002). Male production
74 can also be artificially induced by adding hormone analogs to the culture medium (Olmstead and
75 Leblanc 2002). However, some *Daphnia* individuals never produce males, neither under natural
76 conditions nor when artificially exposed to hormone analogs (Innes and Dunbrack 1993; Tessier
77 and Caceres 2004; Galimov et al. 2011). This non-male-producer "NMP" trait segregates as a
78 single Mendelian locus (or single region) with a dominant female-determining allele called "W".
79 Heterozygous genotypes ("ZW") are genetically determined females called NMP individuals,
80 which take part in sexual diapause stage production exclusively as females. Homozygous
81 genotypes ("ZZ") are cyclical parthenogens with ESD. They produce diapause stages either
82 through male or female function, and are called male producers ("MP" individuals; Galimov et
83 al. 2011).

84 *Daphnia* populations harboring NMP as well as MP individuals offer an opportunity to
85 study the transition from ESD to GSD, and potentially the early stages of sex chromosome
86 evolution. At present, female is the only genetically determined sex in the populations, so that the
87 situation resembles gynodioecy, with no genetically determined males present. Nonetheless, the
88 early steps of sex chromosome evolution may have taken place on both homologs. For instance,
89 sexually antagonistic mutations may have occurred in genes linked to the female-determining
90 locus, and suppressed recombination may have evolved in response to such two-locus
91 polymorphisms in the region.

We used genetic linkage mapping, association mapping, and comparative genomics methods to genetically characterize the sex-determining locus and its surrounding region in *D. magna*. Specifically, we first investigated whether the sex-determining locus maps to the same genomic location in crosses involving NMP females from populations with highly divergent mitochondrial lineages (Galimov et al. 2011, Svendsen et al. 2015). This is relevant because the W allele is expected to get co-transmitted with the mitochondrial haplotype (as both have female-limited transmission). Hence the existence of NMP in these divergent lineages might be due to recurrent evolution of a W allele, or, alternatively, might hint at long-term maintenance of the polymorphism within this species. The recombinant frequencies among the offspring of the experimental crosses were also used to investigate the hypothesis of reduced recombination between W and Z. Second, we used association mapping within a single population to identify female-associated SNPs and test if the association among SNPs is maintained purely through physical linkage or also by selection. For this, we estimated levels of linkage disequilibrium and searched for traces of historical recombination between different associated SNPs in the region around the sex-determining locus. Finally, we screened the NMP associated region of *D. magna* for genes involved in sex-determination in this or other species to identify potential candidate female determining genes. The overall aim of these approaches was to gain a genetic understanding of the transition from ESD to GSD and to assess the possible evolution of an incipient ZW sex chromosome system in *D. magna*.

112 # Results

113 Genetic linkage mapping of the NMP-determining region using microsatellites markers

114 We performed three NMPxMP crosses involving NMP females from three populations
115 with highly divergent mitochondrial haplotypes. In each cross, F1 lines were phenotyped (MP vs.
116 NMP) and genotyped at a total of 81 microsatellite loci to perform linkage mapping of the
117 genomic regions underlying the NMP phenotype. The resulting maps were compared to the
118 *Daphnia* genetic map (Dukič et al. in press), which is based on an F2 panel of an MPxMP cross.

119 In the NMPxMP_1 and NMPxMP_2 crosses, partially overlapping sets of thirteen
120 markers were significantly linked to the NMP phenotype (or to other markers linked to it). In
121 each cross, eleven of these markers were successfully genotyped in >70% of offspring and were
122 thus used for mapping (see Supplementary Material S1). They all mapped to linkage group 3
123 (LG3) of the reference genetic map (Fig. 1). In the NMPxMP_1 cross, three markers
124 (dm_scf02569_310402, dm_scf00933_2550 and dm_scf00700_81490) were completely linked
125 with the NMP phenotype, as were five markers in the NMPxMP_2 cross (dm_scf02569_317703,
126 dm_scf01492_1407, dm_scf00933_2550, dm_scf03156_57375 and dm_scf00966_75426). Only
127 two of these markers were genotyped in the NMPxMP_3 cross, but they too showed complete
128 linkage with the NMP phenotype (a third tested marker was not polymorphic in this cross, see
129 Supplementary Material S1). In all three crosses, the fully linked markers mapped between cM
130 positions 87.8 cM and 94.0 cM of LG3 in the reference genetic map. This region also contains
131 the centromere (at 90.8 cM). We call this region the "NMP region" (Fig. 2). A Marey map of
132 LG3 (Dukič et al. in press) shows that the NMP-region corresponds to a large (~3 Mb) non-
133 recombining region around the centromere (Fig. 2). Non-recombining regions around the
134 centromeres are found on all linkage groups of *D. magna*, and are not a sign of reduced

135 recombination between sex chromosomes, since they also occur on all other linkage groups in the
136 MPxMP cross of the reference genetic map.

137 We investigated the possibility of additional recombination suppression in NMPxMP
138 crosses as an indication of reduced recombination between the Z and (putative) W chromosomes
139 compared to between two Z chromosomes. Indeed, genetic map distances between adjacent
140 markers in and around the NMP region were on average lower in the NMPxMP crosses than in
141 the reference genetic map (Fig. 1). However, this pattern is not statistically supported. The
142 reduction was significant only for one, non-identical interval in each of the two crosses.
143 Furthermore, the two crosses are not entirely independent. Both involved, at a different stage of
144 the experiment, outcrossing of an NMP-female (different in the two crosses) to a male from the
145 same distant population. In one cross this was done to create the actual mapping population, in
146 the other it was done one generation earlier to obtain a highly heterozygous, hybrid NMP female
147 with the intention to increase the number of markers available for mapping the W. Moreover, it is
148 possible that the slight reduction in recombination frequencies was not specific to the NMP
149 region (we did not have sufficient genotype data on linked markers in other genomic regions to
150 test for this possibility). Overall, our results are clearly inconsistent with strongly reduced
151 recombination across large parts of the putative incipient Z and incipient W chromosomes (Table
152 1), suggesting that if such a reduction has happened (compared to recombination in MPxMP
153 crosses), it concerns only a small portion of the chromosome.

154

155 Association mapping of SNPs in the NMP region

156 A genome-wide association analysis using RAD-sequencing of 72 individuals from a
157 single population revealed 43 SNPs significantly associated (FDR $<10^{-5}$) with the NMP

158 phenotype (Fig. 3a, Supplementary material S2). Of these, 36 were contained in a region of LG3
159 between 72.3 and 95.7 cM, which is only slightly larger than the NMP region defined above (Fig.
160 3b). Of the other seven significantly associate SNPs, five mapped to two scaffolds on LG1, and
161 one SNP to each of LG2 and LG4 (Table 2; Fig. 3). The number of associated SNPs per scaffold
162 did not correlate with the scaffold size (Pearson's correlation coefficient r^2= - 0.117). On the
163 physical map of LG3 (see Methods and Dukič et al. in press), the main associations were
164 distributed across 3.6 Mb, between positions 5.4 and 9.0 (Fig. 4.c; LG3 is 10.1Mb in total). The
165 36 highly associated SNPs are distributed across 15 non-contiguous scaffolds whose combined
166 length is 2.42 Mb (about a quarter of LG3's size). Thus, even though the physical map based on
167 LD mapping may be incorrect, there is strong evidence that significantly associated SNPs are
168 distributed across a large proportion of LG3. Interestingly, 25% of the significantly associated
169 SNPs occurred on just a single 400kb scaffold. This scaffold, scf02569, is therefore a good
170 candidate for the location of the initial, female-determining mutation.
171

172 Differentiation, heterozygosity, linkage disequilibrium, and historical recombination in
173 the NMP region

174 To investigate the levels of differentiation between the NMP and the MP individuals in
175 the NMP region, we performed several correlated analyses (to test for consistency of the results):
176 we estimated $F_{ST,}$ compared levels of heterozygosity between both phenotypes, and performed a
177 linkage disequilibrium analysis (LD). The NMP region showed strong differentiation between
178 NMP and MP individuals for the associated SNPs, reflected in F_{ST}-values for individual markers
179 as high as 0.7 (Fig. 4.a). Equally, there was strong LD between the 36 associated SNPs (Fig. 5),
180 although interspersed with areas of low LD, among non-associated SNPs. In concordance with

181 the association results, we also found higher levels of heterozygosity in NMP individuals than in
182 MP individuals (average heterozygosity in the NMP region of 0.50 vs. 0.33, respectively, $P <$
183 0.0001; Figure 4.b). This difference in heterozygosity is specific to the NMP region: in the rest of
184 the genome, the NMP and MP individuals did not differ in heterozygosity (the means were 0.34
185 and 0.32 respectively, $P = 0.149$). However, heterozygosity of four, supposedly inbred MP
186 individuals was only half of the population average (Supplementary Material S3).

187 Second, to investigate whether the occurrence of multiple, highly associated SNPs
188 distributed across a large region can be explained by a lack of recombination, we tested for
189 historical recombination in the region using Hudson's four gamete tests (see Material and
190 Methods). These tests detected the presence of recombination in 37 pairs of adjacent polymorphic
191 sites in the NMP region. These historical recombination events were distributed across the entire
192 NMP region, and were found both between and within scaffolds (Fig. 4.d.). This implies that a
193 low level of recombination or gene conversion occurs between the incipient Z and the incipient
194 W chromosome, or has at least occurred historically.

195

196 <u>Identification of candidate genes involved in sex determination, and prediction of the</u>
197 <u>effect of SNP substitutions on amino acid identity.</u>

198 To search the NMP region for genes known to be involved in sex determination /
199 differentiation in other taxa, we analysed the 283 non annotated protein sequences that where
200 located in the NMP region. Of these, 184 returned a BLAST result, but 39 were described as
201 hypothetical proteins in the *D. pulex* draft transcriptome DAPPUDRAFT and did not reliably
202 match any proteins in the NCBI non-redundant protein database. Among the remaining 145
203 successfully annotated sequences, 121 (82%) had a top hit on *D. pulex* or *D. magna* sequences,

204 while the others matched various arthropods (15 sequences), and other invertebrates (ten
205 sequences). We compared the 145 annotated genes with the NCBI list of 601 genes known to be
206 involved in sex determination or sex differentiation in other invertebrates, and identified 14
207 candidate genes (Table 3). These include genes involved in sex-specific endocrine signaling
208 pathways, such as a membrane androgen receptors (*zip9*), a tissue specific modulator of the
209 ecdysone response in *Drosophila* (*Broad* complex; Karim et al. 1993), as well as a member of the
210 aldo-keto reductase family (Penning et al. 2000). Scf02569 also harbors a gene resembling
211 transcription factor *sox9*, which acts to inactivate the female differentiation pathway and promote
212 spermatogenesis in males in mammals. In addition, genes involved in chromatin remodeling
213 (lysine-specific histone demethylases, histone deacetylases) are also present in the region. The 14
214 genes are located on 6 different scaffolds, with scf02569 containing eight of these genes, whereas
215 other scaffolds harbor at most two of these genes (Table 3).

216 Nine of the 176 SNPs identified by RAD-sequencing and mapping in the NMP region
217 were located in genes within which the exon-intron structure could not be determined (very likely
218 because of errors in the assembly or gene structure definition), 69 were in intergenic regions, 17
219 in 5' UTR regions, 8 in 3' UTR regions, 19 in introns, and 53 within a gene (Supplementary
220 Material S2). Of these 53 coding SNPs, 32 of are in non-annotated genes, including one (on
221 scaffold02003 in position 98755) that introduced a stop codon into a gene which was annotated
222 as "hypothetical protein" in the *D. pulex* draft genome. 23 SNPs were synonymous and 30 were
223 non-synonymous mutations. Only one significantly associated SNP induced a non-synonymous
224 mutation in a candidate gene: the SNP at position 4433 on scf03156, inducing a change from
225 Valine to Leucine in the lysine specific histone demethylase. However, RAD-sequencing covers
226 only a small fraction of the genome (here 154745 loci were mapped, with reads of 95bp,
227 representing an estimated 6.12% of the genome). Hence, additional non-synonymous SNPs may

228 be present in parts of the NMP region not covered by RAD-sequencing reads. The same applies
229 for potential regulatory SNPs.

230 Discussion

231 Our results suggest that the NMP phenotype is determined by a single, large genomic
232 region located on LG3. The few significantly linked SNPs on other linkage groups may be in
233 linkage disequilibrium with the major region on LG3, maintained by pleiotropic effects.
234 Alternatively, they may be explained by errors in the genetic map or in the genome assembly, and
235 might, in fact, be variants within the NMP-region. The NMP phenotype mapped to the same LG3
236 region in all three crosses, involving populations with divergent mitochondrial lineages (Galimov
237 et al. 2011). This indicates either a single evolutionary origin of NMP in *D. magna* or parallel
238 evolution repeatedly involving the same genomic region. Given the divergent mitochondrial
239 sequences and the co-inheritance of mtDNA with the female-determining allele, a single
240 evolutionary origin of NMP in *D. magna* would imply that the female-determining mutation is
241 old. However, parallel (convergent) evolution remains possible, particularly as the NMP region
242 contains more than one gene potentially involved in sex determination and sex differentiation.
243 These genes could represent different mutational targets for NMP-inducing mutations, and hence
244 the mutation may not be the same in all populations, despite occurring in the same genomic
245 region. Finally, rare paternal transmission of mitochondria or transmission of the female-
246 determining mutation through rare males cannot be ruled out, but are unlikely (Galimov et al.
247 2011; Svendsen et al 2015). Interestingly, a dominant NMP phenotype has also been described in
248 *D. pulex* (Innes and Dunbrack 1993) and in *D. pulicaria* (Tessier and Caceres 2004). Considering
249 that an estimated 150 Myr separate *D. magna* and *D. pulex* (Kotov and Taylor 2011), parallel

250 evolution of the NMP phenotype seems more likely for these two species, rather than a
251 femaleness allele having been maintained for such a long evolutionary time.

252 According to the classical model of sex chromosome evolution discussed in the
253 introduction, the establishment of a female-determining mutation is followed by additional
254 mutations with sex antagonistic (SA) effects occurring at nearby loci. Recombination between
255 these loci and the female sex-determining locus is deleterious, and hence SA selection is thought
256 to favour a reduction of recombination between the incipient Z and W (or X and Y). Such
257 recombination reduction in the heterogametic sex (compared to recombination in the
258 homogametic one) is considered to be a hallmark of early sex chromosome evolution.
259 Alternatively, the sex-determining mutation may occur in a region with already low
260 recombination. Such regions occur on autosomes, for instance due to inversions or vicinity to the
261 centromere (Hoffman and Rieseberg 2008; Ironside 2010; Joron et al. 2011). If such a region
262 contains, from the outset, multiple linked potential target loci for SA mutations, then a further
263 reduction of recombination after the occurrence of the sex-determining mutation might not be
264 favoured by selection, at least not initially. In our study, the sex-determining mutation mapped to
265 the peri-centromeric region of LG3, which, as expected, has a low recombination rate not only in
266 MPxNMP crosses, but also in MPxMP crosses. Interestingly, the sex-determining mutations of
267 *Papaya* and *Populus* are also found in peri-centromeric locations (Yu et al. 2007, Kersten et al.
268 2014). In addition, we did not find strong evidence for a further reduction of recombination
269 between the incipient Z and the W chromosome, suggesting that SA selection did not act strongly
270 (if at all) to further extend the low-recombination region containing the female-determining
271 mutation. Yet, SA selection may still play a major role in determining the NMP phenotype.
272 Under the hypothesis that SA selection occurs, multiple loci should contribute to the NMP
273 phenotype, not only for the determination femaleness, but also for the expression or fitness of the

274 female phenotype, or for enhancing maleness of ZZ individuals. Here, we found highly
275 associated SNPs distributed on a total of 3.6 Mb, but separated by regions of low LD in which
276 there is strong evidence for historical recombination or gene conversion. This suggests a role for
277 selection to be acting on several genes in the NMP region. This selection is likely sex-
278 antagonistic, as it appears to maintain different W-linked vs. Z-linked alleles in several sub-
279 regions independently (and thereby maintain the high LD between associated SNPs despite
280 historical recombination). We identified 14 genes in the NMP region that are known to be
281 involved in sex determination/sex differentiation in other taxa. These genes might be potential
282 target for SA mutations. Hence, the NMP region shows all characteristics of a supergene (Joron
283 et al. 2011), with different alleles appearing to be selectively maintained between Z and W in
284 several genes throughout the region. Such a "sex-super gene" is nothing else than the first step
285 towards the evolution of a sex chromosome, even though low recombination in this region is not
286 due to secondary suppression of recombination, but due to a chance event (occurrence of the sex-
287 determining mutation in a large region with pre-existing low levels of recombination). However,
288 such a chance event may, in fact, be one of the possible mechanisms leading to initial suppression
289 of recombination between incipient sex chromosomes.

290 The transition from ESD to GSD is thought to happen rapidly in nature. However,
291 intermediate stages (such as gynodioecy, or partial GSD) can be evolutionary stable, depending
292 on their origin and genetic control (Charlesworth and Charlesworth 1978). The NMP mutation
293 might first have been favoured by selection because it leads to obligate outcrossing (Innes and
294 Dunbrack 1993). This is supported by the fact that inbreeding depression in *Daphnia* is strong
295 (e.g., Lohr and Haag 2015) and that within-clone mating occurs at an appreciable frequency (e.g.,
296 5 % of MP individuals in our association study were offspring of within-clone mating). Once it
297 attained an intermediate frequency, it was probably also affected by negative frequency-

298 dependent sex-ratio selection, which may have set the stage for additional mutations to gradually
299 improve maleness of ZZ individuals (Innes and Dunbrack 1993; Galimov et al. 2011). The
300 coexistence of ESD and GSD individuals in some populations of *D. magna* might be evolutionary
301 stable. This stability could be due to their reproductive cycle, called cyclical parthenogenesis,
302 which might prevent further evolution towards full GSD. Indeed, only females participate in
303 parthenogenetic reproduction. Hence, in order to profit from parthenogenetic multiplication, all
304 genotypes have to be females at some point during the seasonal cycle. ZZ females might increase
305 their male function during sexual reproduction by prolonging their investment in parthenogenetic
306 clutches (late season parthenogenetic clutches are usually male, Galimov et al. 2011). It is also
307 possible that the maleness of ZZ individuals may have been quantitatively improved by linked
308 genes with SA effects (see above). Nonetheless, at the very end of the season, it may still be more
309 profitable for these females to engage in sex rather than to abandon reproduction or to
310 parthenogenetically produce additional males which might not have the time to develop into
311 adults. If however, *Daphnia* was to complete the transition from ESD to pure GSD (as envisaged
312 by the theory outset in the introduction), the incipient Z homolog of the NMP region would need
313 to acquire a mutation determining the male sex (i.e. a recessive female-sterility mutation), and the
314 two chromosomes would then be called proto-sex chromosomes. This has not happened in *D.*
315 *magna*, as the ZZ individuals still have ESD and still contribute to the diapause stage production
316 also through female function. However, while hatchlings from diapause stages always seem to be
317 female in *Daphnia*, male hatchlings do occur in the related *Daphniopsis ephemeralis*, living in
318 short-lived environments (Schwartz and Hebert 1987). Hence, it is not inconceivable that some
319 *Daphnia* populations or species might evolve full GSD, perhaps especially under conditions that
320 reduce the importance of the pathenogenetic phase.

321 The molecular mechanisms underlying male differentiation in *D. magna* and its
322 relationship with ESD are the object of much research (Kato et al. 2008, 2010, 2011; LeBlanc
323 and Medlock 2015). To date, we know that a homolog of the *Doublesex* gene (*dsx*) is found in *D.*
324 *magna (dapmadsx)* and that it is a major effector regulating the male phenotype (Kato et al. 2011;
325 Salz 2011; Beukeboom and Perrin 2013). The regulation of its expression appears, however, to be
326 different in *Daphnia* compared to other arthropods, as no evidence for sex-specific splicing was
327 identified for the *transformer* gene in male versus female embryos (Kato et al. 2010, 2011;
328 Verlhulst et al. 2010). Instead, the expression of *dapmadsx* might be controlled by an "on/off"
329 mechanism whose activation may involve elements responsive to juvenile hormone (Kato et al.
330 2011). Our results show that the NMP region does not contain *dapmadsx* nor *transformer*, and
331 hence neither of these genes is the likely location of the female-determining mutation. However,
332 the NMP mutation does contain multiple genes that are potentially members of the same
333 pathways. These genes include *transformer2 (tra2)* on scf02569, a splicing regulator known to
334 interact with *transformer* to control the female sex-specific splicing of *dsx* in insects. It will be
335 interesting to investigate if and how *tra2* interacts with *dapmadsx.* Moreover, we found four
336 genes involved in hormonal pathways (*zip9*, *zip11*, *Broad* complex, aldo-keto reductase), which
337 might be relevant as *dsx* expression might be controlled by hormone-responsive elements.
338 Furthermore, scf02569 contains a gene resembling transcription factor *sox9*, which inactivates the
339 female differentiation pathway and promotes spermatogenesis in males mammals. Overall, the
340 presence of these 14 genes in the NMP region is congruent with a major role of this region in sex
341 determination/differentiation. In addition, scf02569 is a strong candidate for carrying the original
342 sex-determining mutation. Not only does it contain 25% of the significantly associated SNPs and
343 eight of the 14 candidate genes, but one of these candidate genes (*tra2*) appears to be one of the

344 most promising ones, based on its function in other organisms, and on what is currently known
345 about the mechanisms underlying male differentiation in *D. magna*.

346 In the present study, we determined that the dominant female sex-determining locus
347 observed in multiple population of *D. magna* maps within a single genomic region in the peri-
348 centromeric region of LG3. With a pre-existing low recombination rate, the region contains
349 multiple genomic regions in high LD spanning a total of 3.6Mb, separated by regions of low LD
350 due historical recombination or gene conversion likely occurred. The strong LD is likely
351 maintained by sex antagonistic selection, since multiple genes involved in both males and
352 females sex differentiation were found throughout the region. We conclude that *D. magna*'s LG3
353 carries a sex-related supergene, and is an incipient W chromosome, the youngest stage that might
354 be possible to empirically observe in sex chromosome evolution. *D. magna* is thus a very
355 promising model for studying the evolutionary transitions from ESD to GSD and early stages of
356 sex chromosome evolution.

357

358 Material and Methods

359 Genetic linkage mapping of the NMP-determining region, and assessment of
360 recombination rates

361 In order to map the genomic region responsible for the NMP phenotype, we performed
362 experimental crosses between known NMP and MP genotypes. Because the NMP locus is
363 believed to be heterozygous dominant in females, the goal was to maximize heterozygosity of the
364 mother NMP clone, while ensuring the MP father was homozygous or had different alleles. All
365 three crosses involved outcrossing between two populations to ensure that a sufficient number of

366 markers had different genotypes between fathers and mothers. One of the crosses also involved a
367 NMP mother that was already a hybrid between two populations, again in order to maximize
368 heterozygosity. The first cross called "NMPxMP_1" involved a NMP female from Volgograd,
369 Russia (48.53 N, 44.486944 E) and a male from Orog-Nur, Mongolia (45.032708 N, 100.718133
370 E) as well as 66 of their F1 offspring. The second cross ("NMPxMP_2") used a hybrid NMP
371 female, which was produced by crossing a NMP female from Moscow, Russia, (55.763514 N,
372 37.581667 E), with a male from Orog-Nur. The cross then involved this hybrid female and a male
373 from Vääränmaanruskia, Finland (60.271617 N, 21.896317 E), as well as 54 of their offspring.
374 The third cross ("NMPxMP_3") involved a NMP female from Yakutsk, Russia (61.964047 N,
375 129.630956 E) and a male from Rybnoye, Russia (56.425003 N, 37.602672 E), as well as 22 of
376 their offspring. The three NMP females had divergent mitochondrial haplotypes (based on a 534
377 bp alignment of the COI sequences, the average number of substitutions per site was 0.037
378 between Yakutsk and Moscow, 0.041 between Yakutsk and Volgograd, and 0.010 between
379 Moscow and Volgograd; Galimov et al. 2011; Genbank accession number JF750770.1 for
380 Moscow, AY803073.1 for Volvograd, and [added upon acceptance] for Yakutsk). Microsatellite
381 markers distributed across the genome were used to investigate the parental lines. Markers that
382 were heterozygous in the NMP mother and for which father and mother had different genotypes
383 were selected and genotyped in the offspring. For all these markers, it could unambiguously be
384 determined which of two maternal alleles was transmitted to a given offspring. Hence, linkage of
385 markers to the NMP-determining region could be assessed, by assaying co-transmission of
386 maternal alleles with the phenotype (i.e. with the W vs. Z chromosome). Before genotyping, the
387 offspring were thus phenotyped using the juvenile hormone Methyl Farnesoate, which triggers
388 the production of males in MP strains but not in NMP strains of *Daphnia* (Galimov et al. 2011).

389 Microsatellite loci were amplified using the M13-protocol (Schuelke 2000): For each
390 locus, unlabeled forward and reverse primers were used together with fluorescently labelled,
391 universal M13 primer. The forward primer consisted of a locus-specific part as well as an
392 overhang complementary to M13. PCR reactions were carried out using the Type-it
393 Microsatellite PCR Kit (Qiagen) according to the manufacturer's protocol with an annealing
394 temperature of 60°C. After 22 cycles, the annealing temperature was lowered to 53°C for another
395 20 cycles in order to allow for proper M13 annealing. The resulting PCR products were diluted
396 four times and mixed with a LIZ5000 size ladder (Applied Biosystems). Samples were genotyped
397 using ABI 3730 capillary sequencer and GENEMAPPER software v. 3.0 (Applied Biosystems).
398 A total of 81 microsatellite loci (see Supplementary Material S1) were tested in the parents. Of
399 these, 60 were polymorphic in one or both parents and thus genotyped in the offspring (47 in
400 NMPxMP_1 and 21 in NMPxMP_2, partially overlapping). Linkage to the NMP phenotype was
401 assessed with a Fisher's Exact tests (two-tailed). Some of the markers were specifically designed
402 in regions for which linkage to the NMP phenotype was suspected based on information from an
403 earlier version of the genetic map (Routtu et al. 2010; Routtu et al. 2014) and the initial finding of
404 weak but significant linkage of one marker (dm_scf00243_208642) in the NMPxMP_1 cross.
405 Therefore, the markers do not represent a random sample throughout the genome. The
406 NMPxMP_3 cross, which included the NMP female with the most divergent mitochondrial
407 haplotype, was done at a later stage. It was only used to test whether NMP maps to the same
408 genomic region as in the two other crosses. Hence, only three loci closely linked to NMP in the
409 first two crosses were also genotyped in the offspring of this cross.

410 Genetic linkage mapping of the NMP region was carried out in R/qtl (Broman et al.
411 2003). It soon became evident that NMP mapped to a region of LG3 of the *D. magna* reference
412 genetic map (Dukič et al. in press), of which a first version called v4.0.1 was published in

413 Svendsen et al. (2015). Hence, map construction was done using markers that either showed
414 significant linkage with the NMP phenotype ($P < 0.01$ in pairwise Fisher's exact tests) or were
415 found on scaffolds of the *D. magna* genome v2.4 (bioproject reference PRJNA298946, on the
416 NCBI repository: https://www.ncbi.nlm.nih.gov/bioproject/?term=PRJNA298946) that had been
417 mapped to LG3. Markers that had more than one third of missing genotypes (amplification
418 failures, etc.) were discarded.

419 The mapping procedure consisted in first positioning and ordering the markers according
420 to previously available data (Dukič et al. in press), if possible, and secondly by estimating genetic
421 distances among the ordered markers according to standard procedures. Specifically, we ordered
422 our markers by using the cM position of the nearest mapped SNP from the same scaffold in the
423 reference genetic map v4.0.1. Microsatellite markers on scaffolds that were not mapped in the
424 reference genetic map were positioned according to the estimated recombination fraction in our
425 crosses between these markers and already mapped markers. The only exception to this
426 procedure was done for microsatellite marker scf02066_483524, which is located on a mis-
427 assembled part of scf02066, closely linked to the end of scf00494 (Dukič et al. in press), and thus
428 was ordered according to this position. Once ordered, Kosambi-corrected genetic map distances
429 among all markers were recalculated from the offspring genotypes of our crosses using R/qtl
430 (with the option sliding-window = 8 markers).

431 To test for a reduction of recombination around the NMP region in the MPxNMP crosses
432 versus the reference MPxMP cross of the genetic map v.4.0.1, we compared the genetic distances
433 for intervals of adjacent markers between the crosses. Specifically, for each interval, we assessed
434 the number of recombinant vs. non-recombinant individuals in each of the NMPxMP crosses
435 versus the reference cross, and tested for significant differences using Fisher's Exact tests (two-
436 tailed) implemented in R core package stats (R Development Core Team, 2008).

437

438 Association mapping of SNPs in the NMP region

439 To further characterize the NMP region, we used single-nucleotide polymorphism (SNP)
440 data obtained by RAD-sequencing of a random sample of 72 individuals (17 NMP and 55 MP)
441 obtained by hatching resting stages from the Moscow population. The details of the RAD-
442 sequencing protocols, alignment to the *D. magna* reference genome, and SNP calling are given in
443 Supplementary Material S4. We obtained 7376 SNPs to be analyzed (Supplementary Material S2
444 for an excel document containing all SNPs). We first performed a genome-wide association
445 study, using the expectation that any bi-allelic SNP functionally related to NMP or tightly linked
446 to it should be heterozygous in NMP individuals and homozygous for the more frequent of the
447 two alleles in MP individuals, corresponding to the ZW and ZZ genotypes, respectively. Only
448 sites with a minor allele frequency larger than 0.1 and less than one third of the individuals with
449 missing genotypes were used in the analysis. For each retained bi-allelic site, we grouped
450 individuals into four categories: Heterozygous NMP individuals, homozygous NMP individuals,
451 homozygous MP individuals for the major allele, and MP individuals non-homozygous for the
452 major allele (either heterozygous or homozygous for the minor allele). For each site, we counted
453 the number of individuals in each of the four categories and calculated the expected number of
454 individuals (under the null-hypothesis of no association) using standard Hardy-Weinberg
455 proportions with allele frequencies estimated across all individuals. We then used Pearson's Chi-
456 square tests with two degrees of freedom to evaluate the genotype-phenotype association at each
457 site. However, in order to only test the hypothesis specified above, any excess that went in the
458 direction opposite the hypothesis (for instance an excess of non-heterozygous individuals among
459 NMP) was discarded (i.e., was not taken into account for the overall Chi-square value). The R

460 script of this association analysis is given in Supplementary Material S5. The significance of the
461 association was assessed by correcting the *P*-value of the Chi-square test according to an overall
462 false discovery rate (FDR) of 10^{-5} using the p.adjust function of the R core stats package.
463

464 Differentiation, heterozygosity, linkage disequilibrium, and historical recombination in
465 the NMP region

466 To confirm that the NMP associated region is highly differentiated between NMP and the
467 MP individuals, we estimated F_{ST} for each SNP with the R package PopGenome (Pfeiffer et al.
468 2014). We also performed a linkage disequilibrium (LD) analysis of the NMP region and
469 investigated relative levels of heterozygosity for MP and NMP individuals. Note that all three
470 analyses test essentially for the same thing (Charlesworth et al 1997): If there are many strongly
471 associated SNPs, we would expect F_{ST} to be high, heterozygosity to be high in NMP individuals,
472 and LD to be high, at least between associated SNPs. All three analyses were run to check for
473 consistency, and in the case of heterozygosity also as a comparison with other genomic regions.
474 The subsequent analyses were restricted to cM positions between 85 and 95 of the genetic map,
475 which includes the NMP region as well as some flanking regions on either side. However, due to
476 the dearth of recombination in this region, the relative position and orientation of many of the
477 scaffolds is unknown (several entire scaffolds having the exact same cM position). Hence, we
478 first inferred the likely relative position and orientation of these scaffolds by LD mapping in MP
479 individuals, assuming no structural rearrangement of those scaffold between MP and NMP
480 individuals (see Supplementary Material S6 for details of the LD mapping procedures). Once the
481 inferred physical map was established, we used it to plot the genotype-phenotype association in
482 the region as well as F_{ST}, and heterozygosity.

483 Linkage disequilibrium for all individuals (i.e., including NMP) was then estimated via
484 pairwise r^2 for each pair of SNPs (see Supplementary Material S7), using the program MCLD
485 (Zaykin et al. 2008). Significance was tested using 9999 permutations, and the extent of LD was
486 visualized using a heatmap constructed in R using the LDheatmap package (Shin et al. 2006). To
487 test for signatures of historical recombination between W and Z haplotypes, we used a filtered
488 dataset composed of 140 polymorphic sites in the region, retaining just one site per read (a
489 maximum of two polymorphic sites on the same read were present in the whole data set, but
490 SNPs on the same read were always in full linkage). We then phased these data using the
491 GERBIL program implemented in the package GEVALT V2.0 (Davidovich et al. 2007). We did
492 not allow the program to infer missing genotypes, since the algorithm favors the use of the more
493 common allele, here the Z allele (only 17 out of 144 haplotypes are W haplotypes). The phasing
494 resulted in two Z haplotypes for each MP individual and in one W and one Z haplotype for each
495 NMP individual (Z haplotypes were identified according to the allelic state of the two most
496 strongly associated SNPs named scf2723_2194 and scf2723_13482). Because we could not
497 exclude genotyping nor phenotyping error (the latter only in the direction of falsely identifying an
498 individual as NMP), we used conservative criteria for the test: we first removed the NMP
499 individual (RM1-01) which had a high genotypic resemblance to MP and thus resulted the
500 highest evidence for recombination (this individual was most likely mis-phenotyped during the
501 hormonal exposure). Furthermore, before carrying out the test we corrected singleton variants
502 within each haplotype group: if a variant was present in only one haplotype in the group, we
503 reverted its state to the majority allele in this group. Overall, 27 loci and 6 loci out of 140 loci
504 were reverted in W and Z, respectively. This conservatively assumes that all these singleton
505 variants were due to genotyping error (note that loci with a minor allele frequency of < 0.1 across
506 both groups had already been excluded during the initial filtering, see Supplementary Material S8

507 for the list of uncorrected and corrected haplotypes). Finally, to test only for recombination
508 between Z and W haplotypes (as opposed to recombination within Z), we performed the
509 Hudson's four-gamete test, which stipulates that the presence of the four possible gametes in a
510 pair of segregating bi-allelic sites is a sign of historical recombination or gene conversion. We
511 inspected all instances where recombination was detected by the test and retained only those
512 where the inferred recombination had occurred between the Z and the W haplotypes.
513

514 SNP effect and identification of candidate genes involved in sex determination

515 To assess whether the NMP region contains candidate genes with known functions related
516 to sex differentiation or sex determination, we extracted all 1306 protein sequences
517 corresponding to transcript sequences located on the scaffolds in the NMP region, and reduced
518 isoform redundancy using BlastClust (http://toolkit.tuebingen.mpg.de/blastclust) with the
519 following parameters: minimum length coverage of 60%, minimum identity of 90%, minimum
520 transcript size of 100 amino acids. This resulted in a set of 361 protein sequences, which we
521 trimmed by hand to remove further the redundancy (we only kept one transcript for each gene).
522 The retained 283 protein sequences (see Supplementary Material S9 for the complete list) were
523 blasted against the NCBI nr database with Blast2GO (Conesa and Götz 2008), using blastp and a
524 maximum e-value of 10^{-10}. Annotated genes were then compared with a list of 601 genes
525 obtained from the NCBI gene data base using the keywords "sex determination" and "sex
526 differentiation". We also used the GFF file (also available on NCBI, bioproject reference
527 PRJNA298946), which contains gene features of *D. magna*, to classify each SNP in the NMP
528 region according to whether it induces a synonymous or a non-synonymous change. This analysis
529 was done using the software tool IGV (Robinson et al. 2011).

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Figures

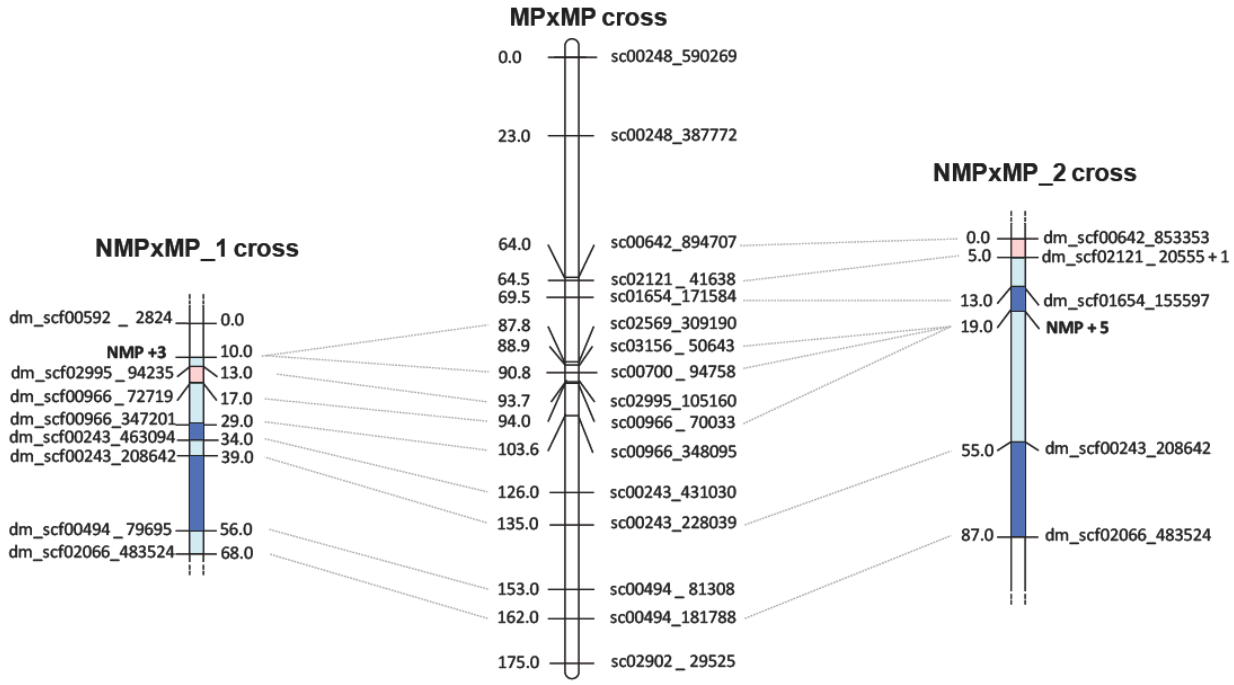

Figure 1. Genetic map of the two NMPxMP crosses (microsatellite markers) and of the MPxMP cross used to create the *Daphnia magna* reference genetic map (SNP markers, Dukič et al. in revision). Only Linkage Group 3 (LG3) is shown. Map distances are in centiMorgans, calculated with the Kosambi mapping function in R/qtl. Areas in light blue / light red show a non-significant reduction / expansion of recombination by comparison to the MP cross. while areas in bright blue indicates a significant reduction of recombination. For the two NMPxMP crosses, one marker per position is represented. In NMPxMP_1, NMP+3 indicates that three markers were in full linkage with the NMP locus. In NMPxMP_2, dm_scf00532_1398 was fully linked with dm_scf02121_20555; also, NMP+5 indicates that five markers were fully linked with the NMP locus.

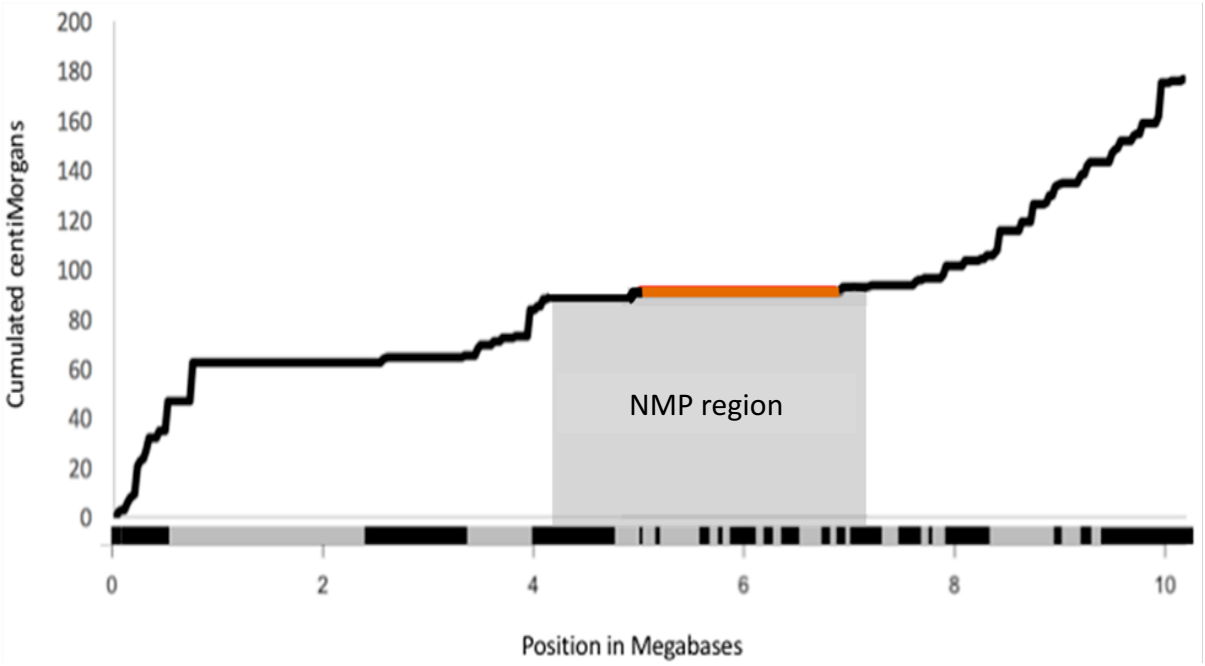

Figure 2. Marey map of LG3 in the MPxMP cross, showing the evolution of the recombination rate along the physical position on LG3. The x axis shows the position in Megabases. Alternated grey and black colors on the x axis represent the length of the different scaffolds that compose LG3. The centromeric region (90.8cM) is highlighted in orange. The shaded area corresponds to the genomic region containing fully linked markers with the NMP phenotype.

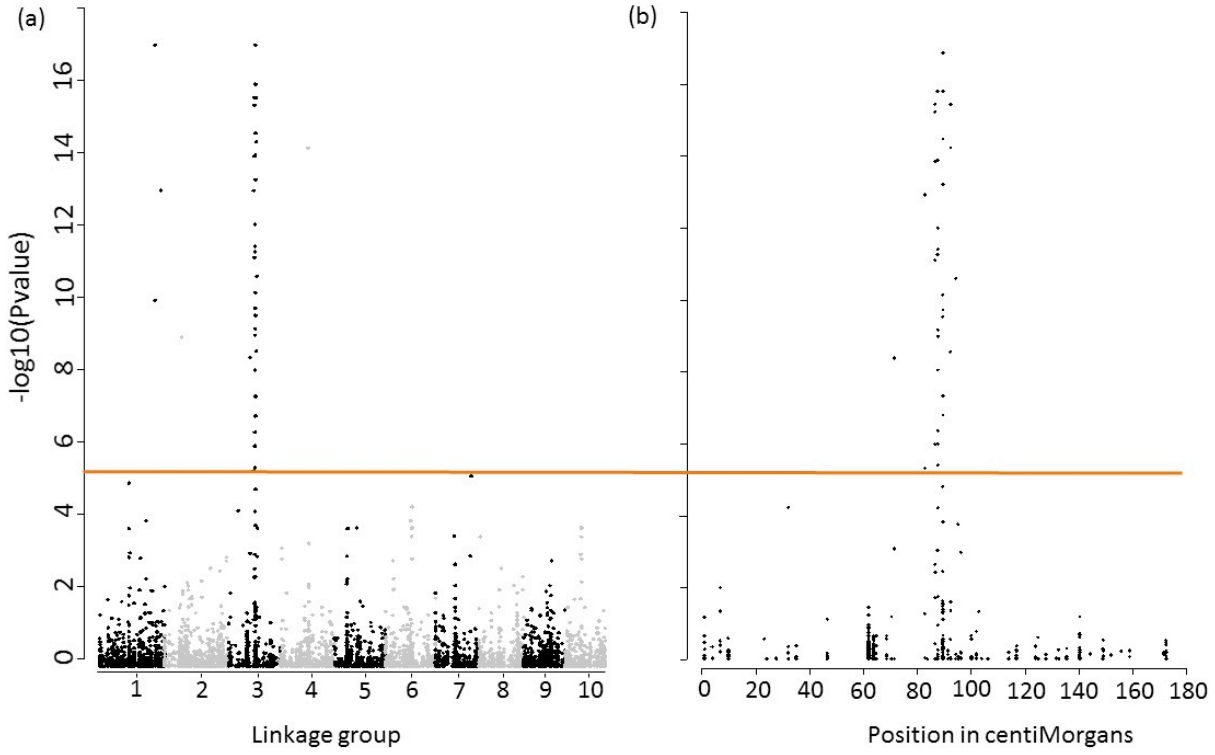

Figure 3. Genome-wide association results. Associations of SNP loci with the NMP polymorphism in a sample of 53 MP and 17 NMP individuals (a) across the entire genome and (b) on LG3. On LG3, markers between 72.3cM and 95.7cM show significant associations with the NMP phenotype (the orange line shows significance, with FDR-corrected P-values < 10^{-3}).

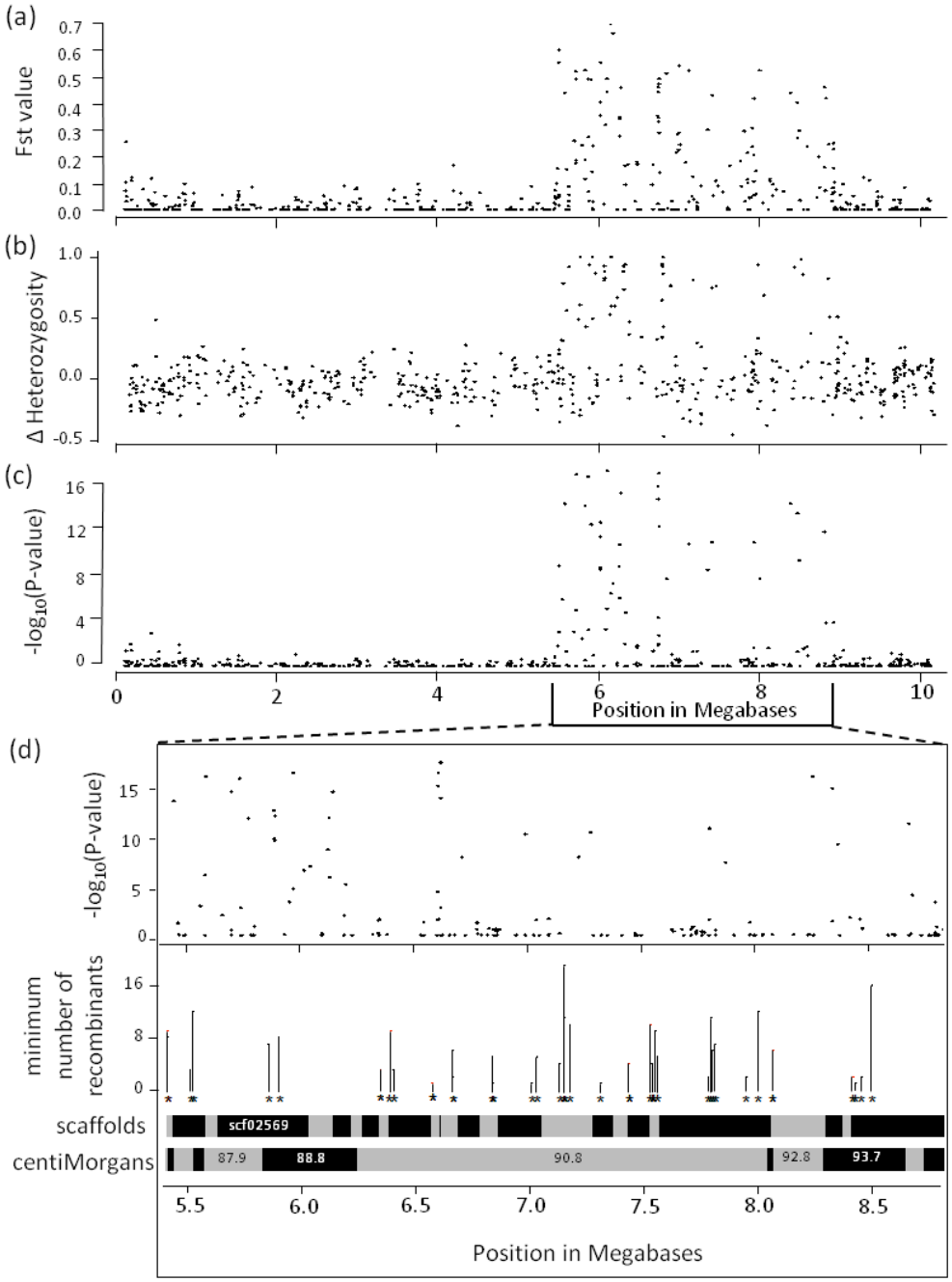

Figure 4. Subdivision between NMP and MP individuals (F_{ST} values, in panel a), the difference in heterozygote frequencies (panel b), and log transformed P values for associations on LG3 (panel c). Panel (d), for the NMP and flanking regions, shows the associations and the minimum numbers of recombinant haplotypes at different positions represented as both the genetic map locations in centiMorgans, and the physical positions of scaffolds.

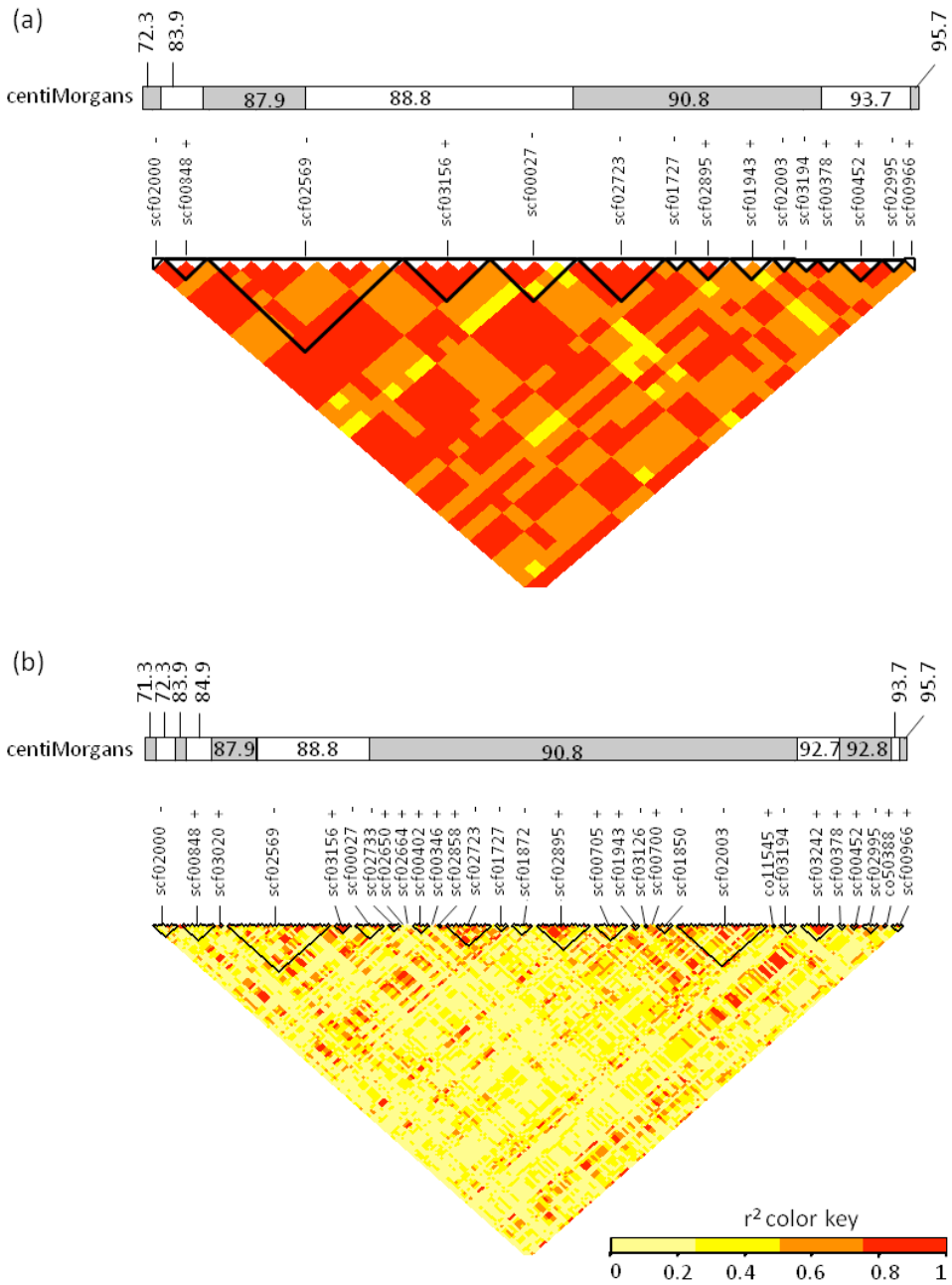

Figure 5. Linkage Disequilibrium Heatmap (r^2 coefficients) in the NMP region. Results are shown for (a) the NMP associated SNPs, (b) all SNPs mapping to the NMP and flanking region corresponding to the NMP region using the 72 individuals (55 MP, 17 NMP). Black triangles represent scaffolds, and the bi-colored band represents the genetic map positions of each SNP, in centiMorgans. + and – represent the orientations of the scaffolds (forward and reverse respectively).

Tables

Table 1. Comparison between crosses with one parent heterozygous (NMP x MP) and both parents homozygous for the MP factor. The table shows Fisher's Exact Test *P*-values for all pairs of markers studied in (a) the NMPxMP_1 cross and (b) the NMPxMP_2 cross. Table also gives the proportion of recombinants for each cross, and for the reference cross (MPxMP, Dukič et al. in revision).

(a)	**NMPxMP_1 versus MPxMP**	***P*-value**	**% recombinant NMPXMP_1**	**% recombinant MPxMP**
	nmp - dm_scf02995_94235	1.000	1.515	1.905
	dm_scf02995_94235 - dm_scf00966_72719	0.300	4.545	0.952
	dm_scf00966_72719 - dm_scf00966_347201	0.655	12.121	15.238
	dm_scf00966_347201 - dm_scf00243_463094	>0.001	4.545	30.476
	dm_scf00243_463094 - dm_scf00243_208642	0.070	4.545	13.333
	dm_scf00243_208642 - dm_scf00494_79695	0.016	13.636	30.476
	dm_scf00494_7969 - dm_scf02066_483524	1.000	12.121	13.333
(b)	**NMPxMP_2 versus MPxMP**	**Pvalue**	**% recombinant NMPXMP_2**	**% recombinant MPxMP**
	dm_scf00642_853353 - dm_scf02121_20555	0.115	3.704	0
	dm_scf02121_20555 - dm_scf01654_155597	1.000	7.404	7.692
	dm_scf01654_155597 - nmp	0.015	7.404	23.077
	nmp - dm_scf00243_208642	0.474	27.778	34.615
	dm_scf00243_208642 - dm_scf02066_483524	0.016	25.926	46.154

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843
844 Table 2. List of scaffolds containing SNPs significantly associated to NMP (Chi square test; $P < 10^{-5}$). LG: linkage group; Size: total
845 size of the scaffold (in basepair); nb. SNPs: number of associated SNPs on the scaffold; *P*-values: resulting Chi square P value.
846

LG	scaffold	Size (bp)	Scaffold position (cM)	nb SNPs	Position of SNPs on scaffold	*P*-values
1	scf00512	3718170	188,7	2	1898250; 1898257	2.04E-17; 1.48E-10
1	scf00205	87516	210,8	3	7803; 7820; 7827	1.63E-13;1.63E-13; 1.63E-13
2	scf02190	2111488	58,2	1	49642	1.46E-09
3	scf02000	31222	72,3	1	4181	5.10E-09
3	scf00848	142519	83,9	2	34647; 68264	5.62E-06; 1.67E-13
3	scf02569	397658	87,9 - 88,8	9	384651; 381715; 268729; 232532; 193880; 80640; 80555; 77438; 77432	1.25E-06; 5.24E-16; 1.99E-14; 8.42E-16; 1.04E-11; 1.35E-12; 8.62E-10; 1.29E-09; 5.30E-12
3	scf03156	106208	88,8	4	4433; 50649; 50655; 79120	2.30E-16; 1.20E-06; 1.20E-06; 5.14E-07
3	scf00027	84679	88,8	4	34526; 29953; 26878; 12813	1.11E-08; 7.27E-12; 4.61E-06; 1.84E-14
3	scf02723	46460	90,8	5	2032; 2194; 13416; 13482	8.44E-14; 2.04E-17;2.30E-16; 4.78E-15
3	scf01727	97879	90,8	1	6345	5.62E-08
3	scf02895	190495	90,8	2	190140; 190147	3.76E-10; 3.76E-10
3	scf01943	93640	90,8	2	13984; 67370	5.79E-08; 2.42E-10
3	scf02003	324342	90,8	1	27223	9.30E-11
3	scf03194	167565	90,8	1	124008	1.91E-07
3	scf00378	73438	93,7	1	22714	5.24E-16
3	scf00452	41028	93,7	2	34895; 34916	8.23E-15; 8.23E-15
3	scf02995	126150	93,7	1	108378	3,47E-09
3	scf00966	460511	95,7	1	174527	3,33E-11
4	scf00311	941766	95,0	1	669922	1,18E-14

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851
852 Table 3. Results of the Blast run showing the 14 candidate genes and their location on the *D. magna* genome. The table reports the start
853 and end position of the gene on the scaffold it maps to, the size of the expected protein (number of amino-acid), the NCBI attributed
854 gene name, the taxon with the best blast hit and its classification, the corresponding e-value, and the percentage of sequence similarity.
855

scaffold	start position	end position	size (aa)	NCBI name	Species	Classification	e-value	similarity
scf00027	2877	6078	316	Serine arginine-rich splicing factor 7	*Harpegnatos saltator*	Insecta	4.0E-50	82.8%
scf00848	96321	97283	136	Aldo-keto reductase family 1, member C4	*Riptortus pedestris*	Insecta	4.1E-59	72.5%
scf02003	35289	35935	136	Poly-U-binding splicing factor Half Pint	*Acyrthosiphon pisum*	Insecta	5.9E-67	95.1%
scf02003	213333	214454	115	Cytochrome P450 314 family	*Daphnia magna*	Crustacea	3.8E-46	88.3%
scf02569	3227	4315	108	Zinc transporter zip11	*Tribolium castaneum*	Insecta	3.9E-29	80.2%
scf02569	9179	10907	300	Zinc transporter zip9	*Poecilia formosa*	Vertebrata	2.1E-75	74.8%
scf02569	35151	44725	606	SOX-9-like transcription factor	*Acromyrmex echinatior*	Insecta	5.0E-48	89.8%
scf02569	218892	220701	292	DnaJ homolog dnaj-5	*Acromyrmex echinatior*	Insecta	4.4E-109	72.1%
scf02569	334258	337000	462	Broad-complex	*Oncopeltus fasciatus*	Insecta	8.3E-50	85.2%
scf02569	340469	342584	281	Transformer2	*Daphnia pulex*	Crustacea	2.9E-119	88.7%
scf02569	76814	79370	558	Protein SPT2 homolog	*Acyrthosiphon pisum*	Insecta	2,00E-33	63.9%
scf02569	228772	229714	158	Histone deacetylase complex subunit sap18	*Metaseiulus occidentalis*	Arachnida	1.1E-55	82.1%
scf02723	1124	6033	287	Epidermal growth factor receptor kinase	*Zootermopsis nevadensis*	Insecta	1.7E-30	71.2%
scf03156	4200	8559	794	Lysine-specific histone demethylase 1A	*Stegodyphus mimosarum*	Arachnida	0.0	85.3%

856 ## Supplementary material

857

858 S1: Table: microsatellites.xls

859 Excel file giving the list of the 81 microsatellite markers tested in this study.

860

861 S2 : Table: snp_data.xls

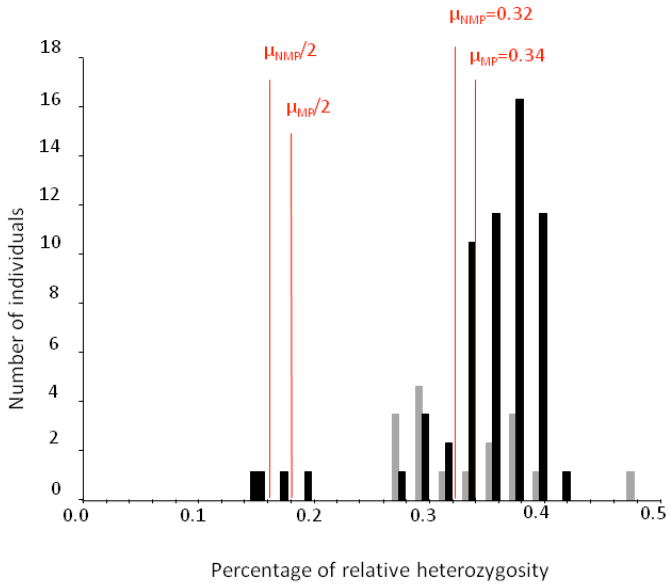
862 Excel file containing the list of SNPs obtained from the RAD-sequencing panel for all
863 individuals. Information listed: Linkage Group; order of SNP on the genetic map; CentiMorgan
864 position on the genetic map; basepair position on the physical map; MegaBase position; scaffold
865 where the SNP maps to; position of the SNP on the scaffold (in bp); major allele; minor allele.
866 Additional information: Chi square value; associated *P*-value; mutation location and type
867 (intergenomic, intron, 5' / 3' UTR; amino acid substitution); synonymous or non synonymous
868 mutation; gene impacted.

869

870 S3 : Figure.

871 Caption: Distribution of the percentage of relative heterozygosity in MP (black) and NMP (grey)
872 individuals. Calculations were performed without LG3, as this chromosome shows a higher
873 heterozygosity in NMP individuals.

874



875

876 S4 : Text : RAD-sequencing and SNP calling protocol

877 We used the RAD-sequencing protocol developed by Etter et al. (2011) with a few
878 modifications. The 72 individuals were divided in 2 libraries. Prior to DNA extraction,
879 individuals were treated for 72 hours with three antibiotics (Streptomycin, Tetracyclin,
880 Ampicilin) at a concentration of 50 mg/L of each antibiotic and fed with microscopic glass beads
881 (Sephadex "Small" by Sigma Aldrich: 50 µm diameter) at a concentration to 0.5g/100 mL. The
882 aim of this treatment was to minimize contaminant DNA (i.e., bacterial DNA or algal DNA) in in
883 the gut and on the surface of the carapace. Genomic DNA was extracted using the Qiagen Blood
884 and Tissue kit following manufacturer's instructions and digested with PstI (New England
885 Biolabs). Digested DNA was barcoded with individual-specific P1 adapters and pooled to create
886 a library containing 2100ng DNA. The pooled library was sheared on a Bioruptor using 2 times 3
887 cycles (1 cycle 30 seconds ON, 1 minute OFF), and fragments between 300 and 500bp were
888 selected through agarose gel electrophoresis. DNA fragments were blunted and a P2 adapter was
889 ligated. The library was amplified through PCR (30 seconds at 98°C, followed by 18 cycles of 10

890 sec. at 98°C, 30 sec. at 65°C and 30 sec. at 72°C; a final elongation step was performed at 72°C
891 for 5 min.). A final electrophoresis was performed to select and purify fragments between 350
892 and 600bp. Each library was sequenced on a single lane of an Illumina HiSeq 2000, using single-
893 end 100 cycle sequencing by the Quantitative Genomics Facility service of the Department of
894 Biosystem Science and Engineering (D-BSSE, ETH), Basel, Switzerland.

895 The quality of the raw sequencing reads (library-wide and per-base) was assessed with
896 FastQC (http://www.bioinformatics.babraham.ac.uk/projects/fastqc/), and reads were checked for
897 barcode integrity, absence of adapter sequences within the reads, and integrity of the PstI cut site.
898 The reads were sorted individually by barcode and filtered to remove reads with uncalled bases
899 and an overall base quality score of less than 24. Reads were subsequently aligned to the *Daphnia*
900 *magna* genome (V2.4; *Daphnia* Genomic Consortium, bioproject reference PRJNA298946, on
901 the NCBI repository: https://www.ncbi.nlm.nih.gov/bioproject/?term=PRJNA298946) using
902 BWA v.0.7.10 (Li and Durbin 2009). Reads that did not map to the reference genome or that
903 mapped to more than one place were discarded. The successfully mapped reads were filtered
904 according to mapping quality (end-to-end mapping with a mapping quality score of at least 25, no
905 more than eight high quality substitutions).

906 Assignment of reads to RAD loci (defined by unique 95 bp locations on the reference
907 genome) and genotype calling was performed in Stacks V1.19 with a bounded SNP model in
908 pstacks (--bound_high of 0.04, according to the base call error rate provided by the sequencing
909 facility) and allowing a maximum of two high frequency haplotypes (i.e. alleles) per locus per
910 individual. Loci with more than two high frequency alleles were excluded because of a too high
911 risk of falsely mapping paralogous reads to a single locus. Cstacks and sstacks were operated
912 with default settings and with the -g option to use genomic location as method to group reads.
913 The distribution of the minor allele frequency indicated that heterozygous loci usually had a

914 minor allele frequency ranging between 0.2 and 0.5 within an individual. We thus fixed the
915 max_het_seq parameter to 0.2 in the program genotypes. As such, potentially heterozygous
916 genotypes with a minor allele frequency of between 0.05 (default homozygote cut-off) and 0.2
917 were considered ambiguous and were scored as missing in the results. Loci were also filtered
918 according to sequencing depth: Loci with less than 20 reads were discarded (to reduce
919 uncertainty in genotype calls) as were reads with a more than five times higher depth than the
920 average depth across individuals (to reduce the risk of including repetitive elements).

921 After final genotype calling, loci were mapped to the Daphnia magna genetic map v.3.0
922 (Dukič et al, submitted). This was done by extracting for each RAD locus the linkage group and
923 cM position of the nearest map-markers on the same scaffold and, if needed, by extrapolating the
924 cM position of the RAD locus by linear extrapolation between the two nearest map-markers.

925

926 *References*

927 Etter PD, Preston JL, Bassham S, Cresko W, Johnson EA. 2011. Local de novo assembly of RAD
928 paired-end contigs using short sequencing reads. PLoS ONE,6:755,e18561.

929

930 S5 : Script: association.R

931 Annotated R script providing the function we used to perform the association analysis at the
932 genome wide level. This script comes partly from the one used in Joron et al. 2011, referenced in
933 the article.

934

935 S6 : Text. Protocol for the physical ordering of scaffold in the NMP associated non-
936 recombining region of LG3.

937 The region controlling the NMP phenotype maps to a region with a low recombination
938 rate in the reference genetic map. This results in the fact that numerous scaffolds are mapped to
939 the exact same cM position, and their relative position and orientation amongst each other are not
940 resolved. Hence, no physical order of the scaffolds in the region can be obtained from the genetic
941 map. This is problematic for genome-wide association studies and fine mapping of the NMP
942 locus, especially for determining whether the NMP phenotype maps to one or multiple specific
943 sub-regions. In an attempt to physically order and orientate the scaffold in the NMP region, we
944 performed linkage disequilibrium (LD) mapping, which uses data on LD from a single population
945 and therefore can make use of historical recombination events present in the data. LD mapping
946 relies on the expectations that two physically close loci should show a high correlation in their
947 segregation patterns in a population (and thus high LD), since recombination events between the
948 two loci should be rare. It is a population based method, so that individual discrepancies with the
949 global population pattern cannot be tested.

950 Because the NMP region on the incipient W chromosome might carry phenotype specific
951 rearrangements, we based the physical ordering using LD mapping only on the 54 MP individuals
952 sampled from the MOS population. We performed LD mapping on a region of LG3 between
953 85cM and 95cM, in order to use SNPs just outside the NMP linked region as anchor. The MOS
954 dataset contains SNPs on 30 mapped scaffolds in this region. Among these, there are three groups
955 of scaffolds for which the relative position and orientation could not be resolved with the genetic
956 map: two scaffolds at position 88.8 cM, 17 scaffolds at 90.8 cM and 4 scaffolds at 93.7 cM. For
957 physical ordering of the scaffolds within each of these groups, we first calculated r^2 values for

958 each pair of SNPs with MCLD (Zaykin 2008), which is based on the correlation of segregation of
959 genotypes in natural populations, avoiding the need to phase the data, but removing any
960 individual particularities in segregation pattern. We then averaged the r^2 values of the three
961 terminal SNPs on each side of each scaffold and created a matrix of pairwise average r^2 values
962 between each pair of scaffold extremities for each of the cM groups separately. When more than
963 two scaffolds had to be ordered in a group, we perform a hierarchical clustering analysis to
964 identify "starting clusters" (highly linked scaffold extremities), using the hclust function of the R
965 core package stats. Scaffolds were then added one by one to the starting clusters following the
966 hierarchical order obtained from the hclust dendogram, and oriented in a way that maximized the
967 average r^2 values between adjacent scaffold extremities.
968

969 S7 : Table: LD_nmp_region.xls

970 Excel document containing calculated r^2 values for the SNPs present in the NMP non
971 recombining region, in order to order and orientate scaffolds in the region not recombining in the
972 reference MPxMP cross.
973

974 S8 : Table: phased_haplotypes.xls

975 Excel document containing the list of the raw and the corrected haplotypes phased in this study,
976 along with the SNP coordinates for each position.
977

978 S9 : Text: nmp_region_gene_content.fasta

979 FASTA formatted document listing the 283 genes used in the analysis.
980

981 Raw genomic data:

982 The FASTQ files of all the individuals used in this study will be available on the SRA database

983 upon acceptance of the manuscript.