Drosophila Mutants that Are Motile but Respond Poorly to All Stimuli Tested: An Intermediate Called "The Boss"

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ABSTRACT

Adult *Drosophila melanogaster* fruit flies were placed into one end of a tube near to repellents (benzaldehyde and heat) and away from the other end containing attractants (light and a favored temperature). They escaped from the repellents and went to the attractants. Five motile mutants that failed to do that were isolated. They did not respond to any external attractants tested or external repellents tested. In addition, they did not respond well to internal sensory stimuli like hunger, thirst, and sleep. The mutants, although motile, failed to respond to stimuli at both 34°C and at room temperature. Some of the mutants have been mapped. It is proposed that the information from the different sensory receptors comes together at an intermediate called "The Boss", that brings about a behavioral response.

I. INTRODUCTION

Organisms are constantly exposed to a variety of external attractants and external repellents as well as to a variety of internal sensory stimuli. How organisms respond to these to bring about behavior is a basic question of life.

One approach for discovering how this works is the isolation and study of mutants that fail here. In this report we show that *Drosophila* flies can be mutated in such a way that, although still motile, they no longer respond well to any sensory stimulus tested. This includes various external attractants and various external repellents as well as internal sensory functions like hunger, thirst, and sleep. An account of some of this work has appeared (Lar Vang and Julius Adler, 2016). A preliminary report of some of the results has been presented (Adler, 2011; Vang, Alexei Medvedev, and Adler, 2012).

An unusual feature here is that only a very few mutations cause a great many different defects in behavior. This makes it likely that the cause is a substance with

broad properties. It is now proposed that these mutants may be defective in "The Boss". The idea of The Boss was introduced by Adler in "My Life with Nature", p. 60 of Annual Review of Biochemistry, 2011: "The Boss is the thing inside every organism – humans, other animals, plants, microorganisms – that is in charge of the organism." All things that an organism does are controlled by The Boss: The Boss controls behavior, metabolism, development, immunological response, and reproduction (Figure 10 of Adler, 2016). While so far The Boss has been just an idea, this idea is now supported by these mutants.   Here the mutant organism would not respond to sensory stimuli due to lacking the behavior part of The Boss. Thus a single defect in The Boss could cause multiple defects in behavior. Possibly between The Boss and the mutants described here lies another component called "Inbetween" before (Vang and Adler, March 21, 2016).

Figure 1 summarizes current understanding of the mechanism of behavior including the role of The Boss:

Figure 1. The mechanism of behavior. This applies to all organisms: microorganisms, plants, animals including humans. Decision making occurs when several sensory stimuli are encountered together, which is usually the case. Central processing includes behavior, metabolism, development, immunological response, and reproduction. See The Boss in this text.

Work by others has shown that behavior in insects includes vision (Peter Weir and Michael Dickinson, 2015), smell and taste (Leslie Voshall and Reinhard Stocker,

2007), audition (Jan Clemens, Cyrille Girardin, Philip Coen and coworkers, 2015), and courtship (Haria Pavlou and Stephen Goodwin, 2013). Central processing includes the central complex, which is a system of neuropils consisting of the protocerebral bridge, the fan-shaped body, the ellipsoid body, and noduli (Ulrike Hanesch, Karl-Friedrich Fischbach, and Martin Heisenberg, 1989; J.M. Young and Douglas Armstrong, 2010; Tanya Wolf, Nirmala Iyer, and Gerald Rubin, 2015).

II. RESULTS

A. RESPONSES TO EXTERNAL STIMULI

1. RESPONSE TO STIMULI USED TOGETHER

In a 34°C dark room flies were started near two repellents (0.1M benzaldehyde and 37°C) at one end of a tube, away from two attractants (light at 1000 lux and 27°C) at the other end (Figure 2). The parent responded by going away from the repellents and to

Figure 2. Apparatus for isolating and testing mutants in a 34°C room. At the left end were repulsive 0.1M benzaldehyde and repulsive 37°C (due to a hot plate at 150°C). At the right end were attractive light (1000 lux) and attractive 27°C (due to ice water). The middle was close to 34°C.

the attractants (Figure 3A). Mutants that were not motile were rejected, only the motile mutants were studied. This consisted of five mutants, named 1 to 5. Such a mutant failed to respond when the four stimuli were present (Figure 3B). Each of the five mutants failed to respond when the four stimuli were present (Vang and Adler, 2016).

Figure 3. Response to stimuli used together. Repellents (0.1M benzaldehyde and high temperature (37°C) were at the left end, attractants (light, 1000 lux, and a favored temperature (27°C) at the right end. *A*, Parental response (n=7). *B*, Mutant 2 (n=8). Flies were tested in a 34°C room with 10 to 20 flies used per trial. Data are mean±SEM.

2. RESPONSE TO INDIVIDUAL STIMULI

A single stimulus was presented to flies that were derived from ones that had already experienced the four stimuli. For example, the parent went to light only (Figure 4A) while a mutant did not (Figure 4B). Each of the five mutants failed to respond to light only (Vang and Adler, 2016).

Figure 4. Response to light alone. Light (1000 lux) was placed at the right end as in Fig. 2. *A*, Parental response (n=4). *B*, Mutant 1 response (n=5). Flies were tested at 34°C with 10 to 20 flies used per trial. Data are mean±SEM.

For heat alone, the parent was repelled (Figure 5A) but the mutant was not repelled (Figure 5B). That was the case for Mutants 1 and 2 (Vang and Adler, 2016). (The other mutants were not tested for this.)

Figure 5. Response to heat gradient alone. The heat source was placed at the left end as in Figure 2. *A*, Parental response (n=4). *B*, Mutant 1 response (n=5). Flies were tested at 34°C with 10 to 20 flies per trial. The warm side measured 37°C and the cool side 27°C. Data are mean±SEM.

A similar result was found for benzaldehyde alone: the parent was repelled by benzaldehyde while Mutants 1 and 2 were not repelled. See Vang and Adler, 2016, for the figures. (The other mutants were not tested for this.)

Thus the mutants were defective not only for the four stimuli used together but also for each stimulus used alone.

3. RESPONSE TO OTHER EXTERNAL STIMULI

These mutants were in addition tested with stimuli that were not among those four used to obtain the mutants:

The mutants were tested for response to the attractant sucrose after starvation (Robert Edgecomb, Cara Harth, and Anne Schneiderman, 1994) for 17 to 20 hours. Compared to the wild-type, both Mutants 1 and 2 consumed less sucrose, about 20% as much as the wild-type. See Vang and Adler, 2016, for the figures. (The other mutants were not tested for this.)

In the case of the repellent quinine, flies were started in a 0.1M quinine half and then they had the opportunity to go into a non-quinine half (see Vang, Medvedev, and Adler, 2012, for details of the method). The parent went into the non-quinine half but Mutant 1 and Mutant 2 did not. See Vang and Adler, 2016, for the figures. (The other mutants were not tested for this.)

To test response to gravity, these flies were placed into a vertical tube and pounded down, then at every minute the flies in each third of the tube were counted (see Vang, Medvedev, and Adler, 2012, for details of the method). The parent responded by climbing up while Mutants 1 and 2 climbed up 10% as well after subtraction of movement without any added stimuli. See Vang and Adler, 2016, for the figures. (The other mutants were not tested for this.)

Thus these mutants, isolated by use of the four stimuli, were defective even for stimuli that were not present during their isolation.

4. MOVEMENT WITHOUT ANY ADDED STIMULI

In the absence of any stimulus added by the experimenters, the parent (Figure 6A) and the mutant (Figure 6B) moved similarly, indicating that motility alone is about the

Figure 6. Response without added stimuli. **A**, Parental response (n=4). **B**, Mutant 1 response (n=6). Flies were tested at 34°C with 10 to 20 flies used per trial. Data are mean±SEM.

same in parent and mutant. This was found also for Mutants 2, 3, 4 and 5 (Vang and Adler, 2016). Aside from our seeing the flies, these results tell that the mutants are motile.

5. EFFECT OF INCUBATION TEMPERATURE

All the work reported above was carried out in a 34°C room in order to allow, if necessary, isolation and study of conditional mutants, i.e. mutants defective at 34°C but not defective at room temperature. We measured response to light (1000 lux) at room temperature (21 to 23°C). The parent responded to light but all five of the mutants failed to respond to light or responded only 10% as well as the parent, just as they did at 34°C (Vang and Adler, 2016). Thus the mutations are not conditional.

Presumably these mutants are defective to all stimuli at room temperature, not just to light. Figures below show defects at room temperature for hunger, thirst, and sleep. Then how could the mutants survive and grow at room temperature? It must be that the mechanism studied here is not an essential one: flies live and reproduce without it.

B. RESPONSES TO INTERNAL STIMULI

1. HUNGER

Here we focus on hunger (Robert Edgecomb, C.E.Harth, and A. M. Schneiderman, 1994; Christoph Melche, Ruediger Barder, and Michael Pankratz, 2007; Katzuyo Fujikawa, Aya Takahashi, Asuza Nishimura and coauthors., 2009; Shelli Farhadian, Mayte Suarez-Farinas, Leslie Voshall and coauthors, 2012; Seung-Hyun Hong, Kyu-Sun Lee, Su-Jin Kwak and coauthors, 2012; Pavel Itskov and Carlos Ribeiro, 2012). To measure hunger we used an apparatus (Fig. 7), inspired and modied

Figure 7. Apparatus for measuring hunger and for measuring thirst. For details see (Vang and Adler, 2016). Tube 1 is called "origin". Flies were tested at room temperature (21-23°C) for up to 40 hours.

from an earlier design (L. Barton-Brown and D.R. Evans, 1960), that we described (Vang and Adler, 2016).

Briefly, in a dark room at 21-23°C male flies – parent or mutants - were transferred into one end (tube 1) of a 5 x 140 cm apparatus containing throughout its length a 5 cm wide strip of wet paper to satisfy thirst but containing no food. Starvation for food began once the flies were put in. Every 10 hours the location of the flies was measured with light on for a few seconds.

At 20 hours the parent had largely left the origin (tube 1) and had begun to accumulate at the end (tube 4) (Figure 8A, solid bars), while the mutant had moved

Figure 8. Movement of flies at 20 hours in search for food. Solid: water but no food (no sucrose). Open: water and food (0.1M sucrose). *A*, Parental response with water only (n=5) and with water + sucrose (n=9). *B*, Mutant 2 response with water only (n=5) and with water + sucrose (n=4). Data are mean±SEM. See (Vang and Adler, 2016) for Mutant 1; the other mutants were not tested for this. Flies were tested at room temperature (21-23°C) with 40 to 60 flies used per trial.

towards the end very little (Figure 8B, solid bars). This is interpreted to mean that the parent is searching for food while the mutant is defective in searching for food.

When food (0.1M sucrose) was added throughout the tube along with the wet strip of paper, the parent moved less far (rather than accumulating at the end) (Figure 8A open bars), while the mutant remained mostly where placed (Figure 8B, open bars). Since sucrose inhibited the movement of the parent, it is supposed that movement without

sucrose is due largely to hunger. From these results we conclude that the mutants are defective in hunger.

2. THIRST

To study thirst, flies were deprived of water. The procedure is the same as for hunger except that water was omitted and solid sucrose was layered throughout (Vang and Adler, 2016). Mutants 1 and 2 were tested, the other mutants not (Vang and Adler, 2016). By 30 hours the parent had moved out, presumably to search for water since addition of water inhibited this (Vang and Adler, 2016). The mutant moved out less well than the parent (Vang and Adler, 2016), so we conclude that the mutants are defective in thirst.

3. SLEEP-WAKE

The parent and mutants isolated here were studied for sleep and wake according to the procedure of Cory Pfeiffenberger *et al*., 2010. The parent was different from the mutants (Figure 9). The parent showed greatest activity at the start and end of the day but

Figure 9. Circadian response. Individual flies are placed into a tube (5 x 20 mm) with an infrared light beam intersecting at the middle of the tube. Mutant 1 (n=24), Mutant 2 (n=24), and parental response (n=24) are recorded over a 24 hour period at 22° C. Data are mean±SEM. There are smaller differences between the parent and the three other mutants (Vang and Adler, 2016).

not in the middle of the day. Mutant 2 showed high activity throughout the day. Mutant 1 was less active than the parent at the start of the day.

C. MAPPING OF THE MUTANTS

We found that Mutant 1 maps in a small gap between 12E3 and 12E5 on the X-chromosome (Vang and Adler, 2016) which we call "*theboss A*". (Earlier we called it *inbetween A*). We found that Mutant 2 maps in the *CG1791* gene, a part of the fibrinogen gene of the X-chromosome, or next to it (Vang and Adler, 2016), which we call "*theboss B*". (Earlier we called it *inbetween B*.)

III. DISCUSSION

Here we describe the isolation and some properties of *Drosophila* mutants that are motile but yet they each fail in response to all external attractants and all external repellents tested (Figures 3-5) and also they are deficient in response to internal stimuli tested (Figures 8 and 9). Thus, although the mutants are motile, they have:

decreased responsiveness to light
decreased responsiveness to heat and to favorable temperature
decreased responsiveness to repulsive chemicals (like benzaldehyde)
decreased responsiveness to sweet tastants (like sucrose)
decreased responsiveness to bitter tastants (like quinine)
decreased responsiveness to gravity
decreased responsiveness to hunger
decreased responsiveness to thirst
abnormality in some sleep

Because all of these different behaviors are defective in the mutants, it seems reasonable to say that there is a single place that is responsible, rather than a defect in each of the many different sensory receptors. Here is a place that when mutated leads to a defect in many places. What is this place? It could be various activities but, as offered in the Introduction, this newly discovered place is considered to be the behavior part of The Boss. This part of The Boss would be defective in the *theboss* mutants found here; these have a defect in the mechanism that dictates a behavioral response. It is proposed that The Boss controls responses to all sensory stimuli.

The path from sensory stimuli to behavioral responses in vertebrates and flies has been compared by Nicholas Strausfeld and Frank Hirth (2013); they reported that the basal ganglia of mammals (the frontal cortex) and the central complex of insects (the superior medial protocerebrum) have multiple similarities and they suggest deep homology between these cerebral cortices, see Figure 10. Origin of the cerebral cortex

After Strausfeld and Hirth, Science 340:157 (2013)

Figure 10. Comparison of the basal ganglia of vertebrates and the central complex of insects according to modification of Fig. 2 of Strausfeld and Hirth (2013). The green arrows lead to the response.

has been studied by use of an ancient ancestor by Raju Tomer, Dedlev Arendt and coauthors (2010) and Heather Marlow and Arendt (2014).

Starting in the 1870's it became apparent to some psychologists that there is a part of the mammalian brain, the frontal cortex, that is master of the whole brain; see reviews up to 1970 by the neurophysiologist Aleksandr Luria, who himself modernized this concept and studied syndromes resulting from deficiencies of the frontal cortex (Luria, 1973, 1980). This part of the brain became known as the "central executive" through the research of the psychologist Alan Baddeley (1966) or as the "executive brain" through the research of the neuropsychologist Elkhonon Goldberg (2001), a student of Luria's. It is now known as "executive control" or "executive function" as well as "prefrontal cortex". For reviews see Goldberg and Dmitri Bougakov (2007), Joaquin Fuster (2008), Sam Gilbert and Paul Burgess (2008), Alfredo Ardila (2008), and Hyun Chung, Lisa Weyandt, and Anthony Swentosky (2014).

Helen Barbas reported (2000, 2013), as described in Figure 11, that the orbitofrontal

Figure 11. The orbitofrontal cortex receives information from sensory cortices and also from the amygdala's recording of past events and emotions and also (not shown here) from the thalamus, as described in Fig. 3 of Barbas (2000). For more detail, see Figs. 41.14 and 41.31 of Barbas (2013).

cortex of humans and other primates receives information from the sensory cortices (which contain visual, auditory, somatosensory, gustatory, and olfactory data as it is received) and also from the amygdala (which contains data about memory and emotion) and also from the thalamus. Barbas suggested that "the orbitofrontal cortex is thus capable of sampling the entire external and internal environment and may act as an environmental integrator" (2000). The orbitofrontal cortex is a part of the prefrontal cortex (p. 3 of Barbas, 2013). After receiving all this information, the prefrontal cortex decides what action should be brought about by informing the premotor cortex (Barbas, 2013), whose function is to produce action. The prefrontal cortex has been reported also in rodents (Verity Brown and Eric Bowman, 2002; Harry Uylings, Henk Groenwegen, and Bryan Kolb, 2003). For a review of what constitutes the prefrontal cortex in mammals, rats, and mice see Marie Carlen (2017).

It is suggested by us that the excitatory pathways and the inhibitory pathways of the prefrontal cortex of vertebrates (Figure 41.14 of Barbas, 2013) are directed by what is here called the "The Boss". The Boss would be in charge of the prefrontal cortex, The Boss would control the prefrontal cortex. The Boss occurs inside the organism. It was proposed that The Boss is to be found throughout biology, in animals, plants, and microorganisms, and that The Boss directs each organism (Adler, 2011, "My life with nature", page 60; Adler, 2016, "A search for The Boss: The thing inside each organism that is in charge"). Thus every organism has something in it that is in control of the organism, namely The Boss.

The prefrontal cortex of vertebrates occurs in the frontal cortex (Barbas, 2013), which corresponds to the superior medial protocerebrum of insects: see Figure 10 above. The frontal cortex and the superior medial protocerebrum might well have analogous functions, so a system like that in the prefrontal cortex might occur in insects, too. The anatomy of the superior medial protocerebrum has been studied in flies, see James Phillips-Portillo and Strausfeld, 2012, and Cynthia Hsu and Vikas Bhandawat, 2016. The superior medial protocerebrum could respond to *theboss* genes.

The control of other genes by The Boss could be studied by finding conditional mutants in The Boss: these could be missing the response to all stimuli at some higher temperature but would be normal at room temperature in order to keep The Boss intact there. In the present report such mutants did not show up, the mutants described here failed at 34°C but they also failed at room temperature. However, only five mutants were studied here; possibly if a much larger number of mutants were isolated there would be some among them that are defective at 34°C but normally responsive at room temperature. In our work on decision making (Adler and Vang, 2016) mutants were isolated that fail at 34°C but do respond normally at room temperature, see next.

Decision making has been studied in animals, plants, and microorganisms (see Figure 1). Decision making occurs in primates and rodents, as described by Jacob Cloke, Derek Jacklin, and Boyer Winters (2015). For a review of decision making in mice, *Drosophila*, and *Caenorhabditis elegans* see Nilay Yapici, Manuel Zimmer, and Ana Domingos (2014); about *Drosophila* they say, "Nutrient sensing that guides decision-making has also been studied in *Drosophila* and found to be regulated at the central nervous system level…These neurons are located in the lateral protocerebrum". In *Drosophila* integration of conflicting sensory information (repellent heat of one color plus non-repellent room-temperature of another color) takes place in the mushroom body, according to Shiming Tang and Aike Guo (2001). In *Drosophila* integration of conflicting sensory information (repellent CO_2 plus attractant vinegar) requires the mushroom body, according to Laurence Lewis, Gerald Rubin, Ilona Grunwald Kadow, and coworkers (2015). *Drosophila* mutants were isolated by Adler and Vang (2016) that fail in making a decision (repellent methyl eugenol plus attractant light) at 34°C but the mutants respond normally in making a decision at room temperature, so it seems that The Boss could be in control of decision making and central processing. These *Drosophila* mutants (Adler and Vang, 2016) should be studied further to find out if some of them may act in the superior medial protocerebrum, see above, to bring about the central processing mentioned in Figure 1.

In conclusion for a major portion here: The Boss is in control of the *internal* parts of the organism – behavior, metabolism, development, immunologic response, and reproduction – and behavior is defective in The Boss mutants described here. In contrast to The Boss, the Leader is in control of the *external* parts of behavior, not described in this report – for example the relation of the organism to other organisms.

V. METHODS

Details of methods used here are found in in the previous paper (Vang and Adler, 2016): A. Isolation of mutants. B. How to study response to external stimuli. C. How to study response to internal stimuli.

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BIBLIOGRAPHY

Adler J (2011) My life with nature. Annu Rev Biochem. 80:42–70.

Adler J (2016) A search for The Boss: The thing inside each organism that is in charge. Anatomy Physiol Biochem Int J. Vol. 1, July 2016.

Adler J, Vang LL (2016) Decision making By *Drosophila* flies. bioRxiv, doi: https://doi.org/10.1101/045666.

Ardila A. (2008) On the evolutionary origins of executive functions. Brain Cogn 68:92-99.

Baddeley A (1966) Exploring the central executive. Quar J Expl Psychol 49A(1):5-28.

Barbas H (2000) Connections underlying the synthesis of cognition, memory, and emotion in primate prefrontal cortices. Brain Res Bull 52:319-330.

Barbas H (2013) Frontal cortex. Chapter 41, pp. 1289-1334. *Neuroscience in the 21st Century*, ed. Donald W. Pfaff, SpringerLink.

Barton-Brown L, Evans DR (1960) Locomotor activity of the blowfly as a function of feeding and starvation. J Insect Physiol 4:27-37.

Brown VJ, Bowman EM (2002) Rodent models of prefrontal cortical function. Trends Neurosci 25:340-343.

Carlen M (2017) What constitutes the prefrontal cortex? Science 358:478-482.

Chung HJ, Weyandt LL, Swentosky A (2014) The physiology of executive functioning. pp. 13-27, in Goldstein S, Naglieri JA, *Handbook of Executive Functioning*, SpringerLink, Springer Science.

Clemens J, Giradin CC, Coen P, Guan X-J, Dickson BJ, Murthy M (2015) Connecting neural codes with behavior in the auditory system of *Drosophila*. Neuron 87:1332-1343.

Cloke JM, Jacklin DL, Winters BD (2015) The neural bases of crossmodal object recognition in non-human primates and rodents: a review. Behav Brain Res 15:118-130.

Edgecomb RS, Harth CE, Schneiderman AM (1994) Regulation of feeding behavior in adult *Drosophila melanogaster* varies with feeding regime and nutritional state. J Exp Biol 197:215-35.

Farhadian SF, Suárez-Fariñas M, Cho CE, Pellegrino M, Vosshall LB (2012) Post-fasting olfactory, transcriptional, and feeding responses in *Drosophila*. Physiol Behav 105:544–53.

Fujikawa K, Takahashi A, Nishimura A, Itoh M, Takano-Shimizu T, Ozaki M (2009) Characteristics of genes up-regulated and down-regulated after 24 h starvation in the head of *Drosophila*. Gene 446:11–17.

Fuster JM. IV. Executive function. In: *The Prefrontal Cortex*, Fourth Edition. London: Academic Press, 2008. pp. 178-193.

Gilbert SJ, Burgess PW (2008) Executive function. Curr Biol 18(3):R110-R114.

Goldberg E (2001) *The Executive Brain, Frontal Lobes and the Civilized Mind*. Oxford University Press, USA.

Goldberg EM, Bougakov DH (2007) Goals, executive control, and action. Chapter 12 In: Baars BJ and Gage NM (Eds) *Cognition, Brain, and Consciousness: Introduction to Cognitive Neuroscience*, Elsevier, pp. 343-368.

Hanesch U, Fischbach K-F, Heisenberg M (1989) Neuronal architecture of the central complex in *Drosophila melanogaster*. Cell Tissue Res 257:343-366.

Hong S-H, Lee K-S, Kwak S-J, Kim A-K, Bai H, Jung M-S, Kwon OY, Song W-J, Tatar M, and Yu K (2012) Minibrain/Dyrk1a regulates food intake through the Sir2-FOXO-sNPF/NPY pathway in *Drosophila* and mammals. PLoS Genet 8:e1002857. doi: 10.1371/annotation/8c2c8644-1beb-4410-8bef-c388b7256738.

Hsu CT, Bhandawat V (2016) Organization of descending neurons in *Drosophila melanogaster*. Scientif Reports 6:20259:1-41.

Itskov PM, Ribeiro C (2013) The dilemmas of the gourmet fly: the molecular and neuronal mechanisms of feeding and nutrient decision making in *Drosophila.* Front Neurosci 7:1–7. doi: 10.3389/fnins.2013.00012.

Lewis LPC, Siju KP, Aso Y, Friedrich AB, Bulteel AJB, Rubin GM, Grunwald Kadow IC (2015) A higher brain circuit for immediate integration of conflicting sensory information in *Drosophila*. Curr Biol 25:2202-2214.

Luria AR (1973) *The Working Brain, an Introduction to Neuropsychology*, translated by Basil Haigh, Penguin Books Ltd., pp. 79-99, 187-225.

Luria AR (1980) *Higher Cortical Functions in Man*, second American edition, translated by Basil Haigh, pp. 246-365, Basic Books Inc.

Marlow H, Arent D (2014) Evolution: Ctenophore genomes and the origin of neurons. Curr Biol 24:R757-R761.

Melche C, Bader R, Pankratz MJ (2007) Amino acids, taste cirquits, and feeding behavior in *Drosophila*: towards understanding the psychology of feeding in flies and man. J Endocrinol 192:467-462.

Parker MO, Brock AS, Walton RT, Brennan CH (2013) The role of zebrafish (*Danio verio*) in dissecting the genetics and neural circuits of executive function. Front Neural Circuits 7:63-81.

Pavlou HJ, Goodwin SF (2013) Courtship behavior in *Drosophila melanogaster*: towards a 'courtship connectome'. Curr Opin Neorobiol 23:76-83.

Pfeiffenberger C, Lear BC, Keagan KP, Allada R (2010) Processing circadian data collected from the *Drosophila* activity monitoring (DAM) system. Cold Spring Harb Protoc 2010:1242–1250.

Phillips-Portillo J, Strausfeld NJ (2012) Representation of the brain's superior protocerebrum of the flesh fly, *Neobellieria bullata*, in the central body. J Comp Neurol 520:3070-3087.

Strausfeld NJ, Hirth F (2013) Deep homology of arthropod central complex and vertebrate basal ganglia. Science 340:157-161.

Tang S, Guo A (2001) Choice behavior of *Drosophila* facing contradictory visual cues. Science 294:1543-1547.

Tomer R, Denes AS, Tessmar-Raible,K, Arendt D (2010) Profiling by image registration reveals common origin of annelid mushroom bodies and vertebrate pallium. Cell 142:800-809.

Uylings HBM, Groenewegen HJ, Kolb B (2003) Do rats have a prefrontal cortex? Behav Brain Res 146:3-17.

Vang LL, Medvedev AV, Adler J (2012) Simple ways to measure behavioral responses of *Drosophila* to stimuli and use of these methods to characterize a novel mutant. PLoS ONE 7:e37495. doi: http://dx.doi.org/10.1371/journal.pone.0037495.

Vang LL, Adler J (2016) *Drosophila* mutants that are motile but respond poorly to all stimuli tested: These may have a defect in interaction with stimuli or in executive function. bioRxiv March 21, 2016.

Vosshall LB, Stocker RF (2007) Molecular architecture of smell and taste in *Drosophila*. Annu Rev Neurosci 30:505-533.

Weir PT, Dickinson MH (2015) Functional divisions for visual processing in the central brain of flying *Drosophila*. Proc Natl Acad Sci USA 112:E5523-E5532.

Wolff T, Iyer NA, Rubin GM (2015) Neuroarchitecture and anatomy of the *Drosophila* central complex: A GAL4-based dissection of protocerebral bridge neurons and circuits. J Comp Neurol 523:997-1037.

Yapici N, Zimmer M, Domingos AI (2014) Cellular and molecular basis of decision-making. EMBO Rep 15:1023-1035.

Young JM, Armstrong JD (2010) Structure of the adult central complex in *Drosophila*: Organization of distinct neuronal subsets. J Comp Neurol 518:1500-1524.