phylo-node: A Molecular Phylogenetic Toolkit using Node.js

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ABSTRACT

Background: Node.js is an open-source and cross-platform environment that

provides a JavaScript codebase for back-end server-side applications. JavaScript

has been used to develop very fast and user-friendly front-end tools for bioinformatic

and phylogenetic analyses. However, no such toolkits are available using Node.js to

conduct comprehensive molecular phylogenetic analysis.

Results: To address this problem, I have developed, *phylo-node*, which was

developed using Node.js and provides a stable and scalable toolkit that allows the

user to perform diverse molecular and phylogenetic tasks. phylo-node can execute

the analysis and process the resulting outputs from a suite of software options that

provides tools for read processing and genome alignment, sequence retrieval,

multiple sequence alignment, primer design, evolutionary modeling, and phylogeny

reconstruction. Furthermore, phylo-node enables the user to deploy server

dependent applications, and also provides simple integration and interoperation with

other Node modules and languages using Node inheritance patterns, and a

customized piping module to support the production of diverse pipelines.

Conclusions: phylo-node is open-source and freely available to all users without

sign-up or login requirements. All source code and user guidelines are openly

available at the GitHub repository: https://github.com/dohalloran/phylo-node

Keywords: Node.js, JavaScript, phylogenetics

BACKGROUND

The cost of whole genome sequencing has plummeted over the last decade and as

a consequence, the demand for genome sequencing technology has risen

significantly [1]. This demand has meant that producing large complex datasets of

DNA and RNA sequence information is common in small research labs, and in terms

of human health this boom in sequence information and precipitous drop in

sequencing costs has had a direct impact in the area of personalized medicine [2-5].

However, once the sequence information becomes available, perhaps the greater

challenge is then processing, analyzing, and interpreting the data. To keep pace with

this challenge, the development of new, fast, and scalable software solutions is

required to visualize and interpret this information.

JavaScript is a lightweight programming language that uses a web browser as

its host environment. JavaScript is cross-platform and supported by all modern

browsers. Because JavaScript is client-side, it is very fast as it doesn't have to

communicate with a server and wait for a response in order to run some code. Web

browsers are ubiquitous and require no dependencies to deploy and operate, and so

JavaScript represents an obvious solution for visualizing sequence information.

Front-end developments using JavaScript have proven to be extremely efficient in

providing fast, easy-to-use, and embeddable solutions for data analysis [6-14]. A

very active community of developers at BioJS (http://www.biojs.io/) provides diverse

components for parsing sequence data types, data visualization, and bioinformatics

analysis in JavaScript [6,7,15-19].

Node.js provides server-side back-end JavaScript. Node.js is written in C,

C++, and JavaScript and uses the Google Chrome V8 engine to offer a very fast

cross-platform environment for developing server side Web applications. Node is a

single-threaded environment, which means that only one line of code will be

executed at any given time; however, Node employs non-blocking techniques for I/O

tasks to provide an asynchronous ability, by using *callback* functions to permit the

parallel running of code. Node holds much potential for the bioinformatic analysis of

molecular data. A community of Node developers provides modules for bioinformatic

sequence workflows (http://www.bionode.io/) which in time will likely parallel the

BioJS community for the number of modules versus components. However, as of

now there are no robust tools for phylogenetic analysis pipelines currently available

using the Node.js codebase. To fill this void I have developed, *phylo-node*, which

provides a Node.js toolkit that provides sequence retrieval, primer design, alignment,

phylogeny reconstruction and as well as much more, all from a single toolkit. *phylo-*

node is fast, easy to use, and offers simple customization and portability options

through various inheritance patterns. The Node package manager, *npm*

(https://www.npmjs.com/), provides a very easy and efficient way to manage

dependencies for any Node application. phylo-node is available at GitHub

(https://github.com/dohalloran/phylo-node), npm

(https://www.npmjs.com/package/phylo-node), and also BioJS

(http://www.biojs.io/d/phylo-node).

IMPLEMENTATION

phylo-node was developed using the Node.js codebase. The phylo-node core

contains a base wrapper object that is used to prepare the arguments and directory

prior to program execution. The base wrapper module is contained within the

Wrapper_core directory (Figure 1). An individual software tool can be easily

accessed and executed by importing the module for that tool so as to get access to

the method properties on that object. These method properties are available to the

user by using the *module.exports* reference object. Inside a driver script file, the user

can import the main module object properties and variables by using the *require*

keyword which is used to import a module in Node.js. The *require* keyword is

actually a global variable, and a script has access to its context because it is

wrapped prior to execution inside the *runInThisContext* function (for more details,

refer to the Node.js source code: https://github.com/nodejs). Once imported, the

return value is assigned to a variable which is used to access the various method

properties on that object. For example: a method property on the *phyml* object is

phyml.getphyml(), which invokes the *getphyml* method on the *phyml* object to

download and decompress the PhyML executable. For a complete list of all

methods, refer to the *README.md* file at the GitHub repository

(https://github.com/dohalloran/phylo-node/blob/master/README.md). In order to

correctly wrap and run each executable, new shells must be spawned so as to

execute specific command formats for each executable. This was achieved by using

child.process.exec, which will launch an external shell and execute the command

inside that shell while buffering any output by the process. Binary files and

executables were downloaded and executed in this manner and the appropriate file

and syntax selected by determining the user's operating system. phylo-node was

validated on Microsoft Windows 7 Enterprise ver.6.1, MacOSX El Capitan

ver.10.11.5, and Linux Ubuntu 64-bit ver.14.04 LTS.

RESULTS AND DISCUSSION

phylo-node is a toolkit to interface with key applications necessary in building a

phylogenetic pipeline (Table 1). Firstly, phylo-node allows the user to remotely

download sequences by building a unique URL and passing this string to the NCBI

e-utilities API (http://www.ncbi.nlm.nih.gov/books/NBK25501/). Any number of genes

can be supplied as command-line arguments to phylo-node by accessing the

fetch_seqs.fasta method on the *fetch_seqs* object in order to retrieve sequence

information in FASTA format. The module for remote sequence retrieval is contained

within the *Sequence* directory. phylo-node also provides methods on specific objects

to download various executable files using the *download* module (Figure 1). Any

binary can be downloaded using the base module *get_executable* contained within

the *Download* directory, however objects pertaining to specific tools such as PhyML

also contain methods for downloading and unpacking binaries (see README.md file

for details). phylo-node also enables the user to create a web server to deploy

embeddable applications such as JBrowse [11] which provides genome visualization

from within a web browser. To facilitate interoperation between various applications

and components, the phylo-node package also contains a module called *phylo-*

node_pipes inside the *Pipes* directory. The *phylo-node_pipes* module allows the user

to easily pipe data between different applications by requiring the *child_process*

module which provides the ability to spawn child processes. Through *phylo-*

node_pipes, the user can chain commands together that will be executed in

sequence to build consistent, and extensive pipelines. The *Pipes* directory contains

sample driver scripts for using the *phylo-node_pipes* module.

phylo-node is highly scalable and new modules for diverse applications can

easily be plugged in. The modules required to wrap and execute applications are all

contained within the *Run* directory. The following tools can be implemented using

phylo-node from within the *./Tool/Run* directory: Trimmomatic [20] to process reads

prior to read alignment; Bowtie2 [21] for read alignment to a reference sequence;

Primer3 [22-24] to facilitate primer design; Clustal Omega [25], K-align [26], and

MUSCLE [27,28] for multiple sequence alignments; Codeml [29], PAL2NAL [30], and

Slr [31] for selection analysis; jModelTest2 [32] and ProtTest3 [33] to determine the

best-fit model of evolution; and PhyML [34,35] for phylogeny reconstruction. The

PhyML executable is also employed by jModelTest2 and ProtTest3. These specific

tools were selected because they are some of the most popular choices and

applications in many bioinformatics pipelines: for example, Primer3 is the most

popular software (over 15,000 citations) for primer design [36]; Clustal Omega, K-

align, and MUSCLE are very fast and accurate multiple sequence alignment tools

that are commonly used to build robust DNA, RNA, or protein alignments [37];

Codeml is part of the PAML suite [29], and alongside PAL2NAL [30] and Slr [31] are

commonly used to determine rates of selection [38,39]. ProtTest3 and jModelTest2

are widely used to determine best-fit models of amino-acid replacements and

nucleotide substitution [40-42]; PhyML is also a popular program (over 12,000

citations) for building phylogenies using maximum likelihood [34]; for next generation

sequencing data, Trimmomatic and Bowtie2 are commonly implemented in read

processing and mapping pipelines [43]. Sample input files for all applications

deployed by phylo-node can be found in the *Input_examples* directory and sub-

directories. Taken together, phylo-node provides a diverse toolkit that allows the user

to develop robust pipelines and instances using Node.

phylo-node is highly scalable and customizable, and was inspired by projects

such as BioPerl [44] which provides very diverse tools that include Perl modules for

many bioinformatic tasks and also parsers and wrappers for diverse sequence

formats and applications. BioPerl's open source structure and architecture allows

users to plug new modules into BioPerl pipelines to design new applications. Node.js

implements prototypal inheritance as per JavaScript but also provides access to the

module.exports object which permits easy portability between the phylo-node toolkit

and any other modules, and also interoperation between different languages by

using the *child.process.exec* process. Therefore, *phylo-node* can be integrated with

existing Node.js bioinformatics tools [45,46] or software written in other languages.

For example, jModelTest2, ProtTest3 and Trimmomatic require a Java runtime

environment (http://www.oracle.com/technetwork/java/javase/downloads/jre8-

downloads-2133155.html), and by using *require* to import each module, the user can

execute the analysis of these tools.

CONCLUSIONS

In conclusion, phylo-node is a novel package that leverages the speed of Node.js to

provide a robust and efficient toolkit for researchers conducting molecular

phylogenetics. phylo-node can be easily employed to develop complex but

consistent workflows, and integrated with existing bioinformatics tools using the

Node.js codebase.

AVAILABILITY AND REQUIREMENTS

- Project name: phylo-node
- Project home page: https://github.com/dohalloran/phylo-node
- Operating system(s): Platform independent
- Programming language: Node.js
- Other requirements: none
- License: MIT
- Any restrictions to use by non-academics: no restrictions or login requirements

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AUTHOR CONTRIBUTIONS

D.O'H. conceived the idea for *phylo-node*, wrote and tested the code, and wrote the manuscript.

COMPETING INTERESTS

The author declares no competing interests.

226   References

1. Shaffer C. Next-generation sequencing outpaces expectations. Nat Biotechnol.
2007;25: 149. doi: nbt0207-149 [pii].

2. Wade N. The quest for the $1,000 human genome: DNA sequencing in the
doctor's office? At birth? It may be coming closer. N Y Times Web. 2006: F1, F3.

3. Mardis ER. Anticipating the 1,000 dollar genome. Genome Biol. 2006;7: 112. doi:
gb-2006-7-7-112 [pii].

4. Service RF. Gene sequencing. The race for the $1000 genome. Science.
2006;311: 1544-1546. doi: 311/5767/1544 [pii].

5. Hayden EC. The $1,000 genome. Nature. 2014;507: 294-295.

6. Yachdav G, Goldberg T, Wilzbach S, Dao D, Shih I, Choudhary S, et al. Anatomy
of BioJS, an open source community for the life sciences. Elife. 2015;4:
10.7554/eLife.07009. doi: 10.7554/eLife.07009 [doi].

7. Gomez J, Garcia LJ, Salazar GA, Villaveces J, Gore S, Garcia A, et al. BioJS: an
open source JavaScript framework for biological data visualization. Bioinformatics.
2013;29: 1103-1104. doi: 10.1093/bioinformatics/btt100 [doi].

8. Salazar GA, Meintjes A, Mulder N. PPI layouts: BioJS components for the display
of Protein-Protein Interactions. F1000Res. 2014;3: 50-50.v1. eCollection 2014. doi:
10.12688/f1000research.3-50.v1 [doi].

9. Gomez J, Jimenez R. Sequence, a BioJS component for visualising sequences.
F1000Res. 2014;3: 52-52.v1. eCollection 2014. doi: 10.12688/f1000research.3-52.v1
[doi].

10. Cui Y, Chen X, Luo H, Fan Z, Luo J, He S, et al. BioCircos.js: an interactive
Circos JavaScript library for biological data visualization on web applications.
Bioinformatics. 2016;32: 1740-1742. doi: 10.1093/bioinformatics/btw041 [doi].

11. Buels R, Yao E, Diesh CM, Hayes RD, Munoz-Torres M, Helt G, et al. JBrowse:
a dynamic web platform for genome visualization and analysis. Genome Biol.
2016;17: 66-016-0924-1. doi: 10.1186/s13059-016-0924-1 [doi].

12. Salavert F, Garcia-Alonso L, Sanchez R, Alonso R, Bleda M, Medina I, et al.
Web-based network analysis and visualization using CellMaps. Bioinformatics. 2016.
doi: btw332 [pii].

13. Franz M, Lopes CT, Huck G, Dong Y, Sumer O, Bader GD. Cytoscape.js: a
graph theory library for visualisation and analysis. Bioinformatics. 2016;32: 309-311.
doi: 10.1093/bioinformatics/btv557 [doi].

14. Vanderkam D, Aksoy BA, Hodes I, Perrone J, Hammerbacher J. pileup.js: a
JavaScript library for interactive and in-browser visualization of genomic data.
Bioinformatics. 2016. doi: btw167 [pii].

15. Garcia L, Yachdav G, Martin MJ. FeatureViewer, a BioJS component for
visualization of position-based annotations in protein sequences. F1000Res. 2014;3:
47-47.v2. eCollection 2014. doi: 10.12688/f1000research.3-47.v2 [doi].

16. Kalderimis A, Stepan R, Sullivan J, Lyne R, Lyne M, Micklem G. BioJS
DAGViewer: A reusable JavaScript component for displaying directed graphs.
F1000Res. 2014;3: 51-51.v1. eCollection 2014. doi: 10.12688/f1000research.3-51.v1
[doi].

17. Villaveces JM, Jimenez RC, Habermann BH. KEGGViewer, a BioJS component
to visualize KEGG Pathways. F1000Res. 2014;3: 43-43.v1. eCollection 2014. doi:
10.12688/f1000research.3-43.v1 [doi].

18. Villaveces JM, Jimenez RC, Habermann BH. PsicquicGraph, a BioJS component
to visualize molecular interactions from PSICQUIC servers. F1000Res. 2014;3: 44-
44.v1. eCollection 2014. doi: 10.12688/f1000research.3-44.v1 [doi].

19. Yachdav G, Hecht M, Pasmanik-Chor M, Yeheskel A, Rost B. HeatMapViewer:
interactive display of 2D data in biology. F1000Res. 2014;3: 48-48.v1. eCollection
2014. doi: 10.12688/f1000research.3-48.v1 [doi].

20. Bolger AM, Lohse M, Usadel B. Trimmomatic: a flexible trimmer for Illumina
sequence data. Bioinformatics. 2014;30: 2114-2120. doi:
10.1093/bioinformatics/btu170 [doi].

21. Langmead B, Salzberg SL. Fast gapped-read alignment with Bowtie 2. Nat
Methods. 2012;9: 357-359. doi: 10.1038/nmeth.1923 [doi].

22. You FM, Huo N, Gu YQ, Luo MC, Ma Y, Hane D, et al. BatchPrimer3: a high
throughput web application for PCR and sequencing primer design. BMC
Bioinformatics. 2008;9: 253-2105-9-253. doi: 10.1186/1471-2105-9-253 [doi].

23. Untergasser A, Cutcutache I, Koressaar T, Ye J, Faircloth BC, Remm M, et al.
Primer3--new capabilities and interfaces. Nucleic Acids Res. 2012;40: e115. doi:
gks596 [pii].

24. Untergasser A, Nijveen H, Rao X, Bisseling T, Geurts R, Leunissen JA.
Primer3Plus, an enhanced web interface to Primer3. Nucleic Acids Res. 2007;35:
W71-4. doi: gkm306 [pii].

25. Sievers F, Wilm A, Dineen D, Gibson TJ, Karplus K, Li W, et al. Fast, scalable
generation of high-quality protein multiple sequence alignments using Clustal
Omega. Mol Syst Biol. 2011;7: 539. doi: 10.1038/msb.2011.75 [doi].

26. Lassmann T, Sonnhammer EL. Kalign--an accurate and fast multiple sequence
alignment algorithm. BMC Bioinformatics. 2005;6: 298. doi: 1471-2105-6-298 [pii].

27. Edgar RC. MUSCLE: multiple sequence alignment with high accuracy and high
throughput. Nucleic Acids Res. 2004;32: 1792-1797. doi: 10.1093/nar/gkh340.

28. Edgar RC. MUSCLE: a multiple sequence alignment method with reduced time
and space complexity. BMC Bioinformatics. 2004;5: 113. doi: 10.1186/1471-2105-5-
113 [doi].

29. Yang Z. PAML: a program package for phylogenetic analysis by maximum
likelihood. Comput Appl Biosci. 1997;13: 555-556.

30. Suyama M, Torrents D, Bork P. PAL2NAL: robust conversion of protein
sequence alignments into the corresponding codon alignments. Nucleic Acids Res.
2006;34: W609-12. doi: 34/suppl_2/W609 [pii].

31. Massingham T, Goldman N. Detecting amino acid sites under positive selection
and purifying selection. Genetics. 2005;169: 1753-1762. doi: genetics.104.032144
[pii].

32. Darriba D, Taboada GL, Doallo R, Posada D. jModelTest 2: more models, new
heuristics and parallel computing. Nat Methods. 2012;9: 772. doi:
10.1038/nmeth.2109 [doi].

33. Darriba D, Taboada GL, Doallo R, Posada D. ProtTest 3: fast selection of best-fit
models of protein evolution. Bioinformatics. 2011;27: 1164-1165. doi:
10.1093/bioinformatics/btr088; 10.1093/bioinformatics/btr088.

34. Guindon S, Gascuel O. A simple, fast, and accurate algorithm to estimate large
phylogenies by maximum likelihood. Syst Biol. 2003;52: 696-704.

35. Guindon S, Dufayard JF, Lefort V, Anisimova M, Hordijk W, Gascuel O. New
algorithms and methods to estimate maximum-likelihood phylogenies: assessing the
performance of PhyML 3.0. Syst Biol. 2010;59: 307-321. doi: 10.1093/sysbio/syq010
[doi].

36. Rozen S, Skaletsky H. Primer3 on the WWW for general users and for biologist
programmers. Methods Mol Biol. 2000;132: 365-386.

37. Thompson JD, Linard B, Lecompte O, Poch O. A comprehensive benchmark
study of multiple sequence alignment methods: current challenges and future
perspectives. PLoS One. 2011;6: e18093. doi: 10.1371/journal.pone.0018093 [doi].

38. Zhang C, Wang J, Long M, Fan C. gKaKs: the pipeline for genome-level Ka/Ks
calculation. Bioinformatics. 2013;29: 645-646. doi: 10.1093/bioinformatics/btt009
[doi].

39. Huerta-Cepas J, Serra F, Bork P. ETE 3: Reconstruction, Analysis, and
Visualization of Phylogenomic Data. Mol Biol Evol. 2016;33: 1635-1638. doi:
10.1093/molbev/msw046 [doi].

40. Guesmi-Mzoughi I, Archidona-Yuste A, Cantalapiedra-Navarrete C, Regaieg H,
Horrigue-Raouani N, Palomares-Rius JE, et al. First Report of the Spiral Nematode
Rotylenchus incultus (Nematoda: Hoplolaimidae) from Cultivated Olive in Tunisia,
with Additional Molecular Data on Rotylenchus eximius. J Nematol. 2016;48: 136-
138.

41. Mazza G, Menchetti M, Sluys R, Sola E, Riutort M, Tricarico E, et al. First report
of the land planarian Diversibipalium multilineatum (Makino & Shirasawa, 1983)
(Platyhelminthes, Tricladida, Continenticola) in Europe. Zootaxa. 2016;4067: 577-
580. doi: 10.11646/zootaxa.4067.5.4 [doi].

42. Matassi G. Horizontal gene transfer drives the evolution of Rh50 permeases in
prokaryotes. BMC Evol Biol. 2017;17: 2-016-0850-6. doi: 10.1186/s12862-016-0850-
6 [doi].

43. Conesa A, Madrigal P, Tarazona S, Gomez-Cabrero D, Cervera A, McPherson
A, et al. A survey of best practices for RNA-seq data analysis. Genome Biol.
2016;17: 13-016-0881-8. doi: 10.1186/s13059-016-0881-8 [doi].

44. Stajich JE, Block D, Boulez K, Brenner SE, Chervitz SA, Dagdigian C, et al. The
Bioperl toolkit: Perl modules for the life sciences. Genome Res. 2002;12: 1611-1618.
doi: 10.1101/gr.361602 [doi].

45. Kim J, Levy E, Ferbrache A, Stepanowsky P, Farcas C, Wang S, et al. MAGI: a
Node.js web service for fast microRNA-Seq analysis in a GPU infrastructure.
Bioinformatics. 2014;30: 2826-2827. doi: 10.1093/bioinformatics/btu377 [doi].

46. Page M, MacLean D, Schudoma C. blastjs: a BLAST+ wrapper for Node.js. BMC
Res Notes. 2016;9: 130-016-1938-1. doi: 10.1186/s13104-016-1938-1 [doi].

47. Skinner ME, Uzilov AV, Stein LD, Mungall CJ, Holmes IH. JBrowse: a next-
generation genome browser. Genome Res. 2009;19: 1630-1638. doi:
10.1101/gr.094607.109 [doi].

48. Yang Z. PAML 4: phylogenetic analysis by maximum likelihood. Mol Biol Evol.
2007;24: 1586-1591. doi: msm088 [pii].

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FIGURE LEGENDS

Figure 1. Workflow for *phylo-node*.

phylo-node is organized into a workflow of connected modules and application

scripts. In order to interface with a software tool, the base wrapper module is invoked

to process command-line requests that are then passed into the software specific

module. The input for the specific software can be passed into the base wrapper

from a folder specified by the user or by using the sequence retrieval module which

is contained within the *Sequence* directory. The *Pipes* directory contains a module

for easy piping of data between applications while binaries and executables can be

downloaded using the *get_executable* module from within the *Download* folder to

deploy software specific modules within the *Run* directory or to provide applications

to a web server from within the *Server* directory.

392 **Table 1**. Summary of *phylo-node* applications.

Module	Description	Source	Application	Citation
fetch_seqs	Remotely retrieve sequence data	https://www.ncbi.nlm.nih.gov/books/NBK25501/	ASN.1 and FASTA sequences	-
get_executable	Download binaries and executables	https://www.npmjs.com/package/download	batch, exe, jar etc..	-
http_server	Creates a web server	https://nodejs.org/api/http.html and http://jbrowse.org/jbrowse-1-12-1/	local version of JBrowse	Skinner et al. 2009 [47]
bowtie2	Sequence aligner	http://bowtie-bio.sourceforge.net/bowtie2/index.shtml	align fastq reads onto reference genome	Langmead and Salzberg (2012) [21]
trimmomatic	Read Trimming	http://www.usadellab.org/cms/?page=trimmomatic	preprocessing for reference alignment	Bolger et al. (2014) [20]
phyml	Maximum-Likelihood Phylogenies	http://www.atgc-montpellier.fr/phyml/binaries.php	model testing (ProtTest3 and jModelTest2) and tree building	Guindon et al. (2010) [35]
primer3	Primer design	https://sourceforge.net/projects/primer3/	PCR and sequencing	Untergasser et al. (2012) [23]
muscle	MSA	http://www.drive5.com/muscle/downloads.htm	sequence alignment	Edgar (2004a); Edgar (2004b) [27,28]
clustal_Omega	MSA	http://www.clustal.org/omega/#Download	sequence alignment	Sievers et al. (2011) [25]
kalign	MSA	http://msa.sbc.su.se/cgi-bin/msa.cgi	sequence alignment	Lassmann and Sonnhammer (2005) [26]
pal2nal	Generate codon alignments	http://www.bork.embl.de/pal2nal/	processing of alignments for selection analysis	Suyama et al. (2006) [30]
Slr	Selection analysis	http://www.ebi.ac.uk/goldman-srv/SLR/#download	detect rates of selection in coding DNA	Massingham and Goldman (2005) [31]
codeml	Selection analysis	http://abacus.gene.ucl.ac.uk/software/paml.html	ML analysis of coding DNA using codon substitution models	Yang (1997); Yang (2007) [29,48]
prottest	Model selection	https://github.com/ddarriba/prottest3	best-fit model determination of amino-acid replacement	Darriba et al. (2011) [33]
jmodeltest2	Model selection	https://github.com/ddarriba/jmodeltest2	best-fit model determination of nucleotide substitutions	Darriba et al. (2012) [32]
base_wrap	Base module for program execution	https://nodejs.org/api/child_process.html	handles arguments and spawns child processes	-
phylo-node_pipes	Module for chaining commands	-	used to pipe data between applications	-