The fitness of *Pseudomonas aeruginosa* quorum sensing signal cheats is influenced by the diffusivity of the environment

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Abstract

Experiments examining the social dynamics of bacterial quorum sensing (QS) have focused on mutants which do not respond to signals, and the role of QS-regulated exoproducts as public goods. The potential for QS signal molecules to themselves be social public goods has received much less attention. Here, we analyse how signal-deficient (*lasI*$^{-}$) mutants of the opportunistic pathogen *Pseudomonas aeruginosa* interact with wild-type cells in an environment where QS is required for growth. We show that when growth requires a 'private' intracellular metabolic mechanism activated by the presence of QS signal, *lasI*$^{-}$ mutants act as social cheats and outcompete signal-producing wild-type bacteria in mixed cultures, because they can use the signals produced by wild type cells. However, reducing the ability of signal molecules to diffuse through the growth medium, results in signal molecules becoming less accessible to mutants, leading to reduced cheating. Our results indicate that QS signal molecules can be considered as social public goods in a way that has been previously described for other exoproducts, but that spatial structuring of populations reduces exploitation by non-cooperative signal cheats.

Importance

Bacteria communicate via signaling molecules to regulate the expression of a whole range of genes. This process, termed quorum sensing (QS), moderates bacterial metabolism in many environmental conditions, from soil and water (where QS-regulated genes influence nutrient cycling) to animal hosts (where QS-regulated genes determine pathogen virulence). Understanding the ecology of QS could therefore yield vital clues as to how we might modify bacterial behaviour for environmental or clinical gains. Here, we demonstrate that QS signals act as shareable public goods. This means that their evolution, and therefore population-level responses to interference with QS, will be constrained by population structure. Further, we show that environmental structure (constraints on signal diffusion) alters the accessibility of QS signals and demonstrates that we need to consider population and environmental structure to help us further our understanding of QS signaling systems.

Introduction

Bacterial quorum sensing (QS) is a cell-to-cell signalling mechanism that coordinates a range of behaviours at the population level (1, 2). QS facilitates density-dependent production of extracellular molecules including nutrient-scavenging enzymes and virulence factors. These molecules have been termed 'public goods' because their benefits can be shared by all cells in the local population (3-6). Because these QS-regulated exoproducts are metabolically costly for cells to produce, QS can also be exploited by non-cooperating "cheats": cells that do not respond to QS signals and so pay no costs, but which exploit wild type populations because they benefit from the public goods produced by wild type neighbouring cells (4-7). Experiments into the social dynamics of QS have traditionally focused on these "signal blind" mutants, and a number of studies have shown that such mutants can arise in laboratory cultures and during infections (8-14). In various laboratory conditions and *in vivo* infection models, they have been shown to act as social cheats (6, 15-17).

However, little attention has been paid to whether QS signals themselves can act as exploitable public goods, despite there being a metabolic cost associated with the production of QS signals (18, 19). Here, we analyse how signal-negative (*lasI*-) mutants of the opportunistic pathogen *Pseudomonas aeruginosa* socially interact with wild-type cells in an environment where growth requires the cells to have a functional QS system, but where the fitness benefits of QS are 'private' to individual cells. *P. aeruginosa* regulates the production of many virulence factors through two *N*-acyl homoserine lactone (AHL) based QS systems. These systems, termed the *las* and *rhl* systems, produce and respond to the signals *N*-(3-oxododecanoyl)-L-homoserine lactone (3O-C12-HSL) and *N*-butanoyl-L-homoserine lactone (C4-HSL) respectively (1, 20). We conducted our experiments in a growth medium containing adenosine as a carbon source. Adenosine is deaminated to form inosine, which is degraded inside the cell by a nucleoside hydrolase (Nuh) to hypoxanthine plus ribose; hypoxanthine is then metabolised to produce glyoxylate plus urea (21). QS is crucial for growth in this medium because the *las* system (through the regulator LasR), positively regulates Nuh. Because Nuh acts intracellularly, any loss of fitness due to mutation of the signal gene *lasI* will be directly due to the lack of signal – not to any downstream effect on the production of extracellular enzymes. We demonstrate that, when provided with adenosine as a carbon source, *lasI*$^-$ mutants act as cheats: they grow poorly in monoculture but have a higher relative fitness than the signal-producing wild type in mixed cultures. In contrast, *lasR* mutants, which cannot regulate Nuh in the presence of signal, do not

80 gain any fitness benefits in mixed culture with wild type cells.

81

82 In contrast to experiments performed in well-mixed liquid medium in test tubes, interactions
83 between bacterial cells in natural environments (including infections) are affected by spatial
84 assortment and structuring (22-24). This affects how behaviours evolve (25-29). We tested how
85 simple spatial structuring, through the addition of agar to the growth medium, alters QS signal
86 diffusion and the social dynamics of wild-type and *lasI*$^-$ cells. Consistent with work on other
87 bacterial public goods (27), we demonstrate that the ability of lasI- mutants to cheat is
88 significantly reduced in structured populations. These results have implications for understanding
89 how and why bacterial signaling evolves, and the likely evolutionary fate of different types of
90 QS mutant in varied environmental conditions (30).

91

92

93 **Materials and Methods**

94

95 **Bacterial strains.** The strains used were the wild type *Pseudomonas aeruginos*a laboratory
96 strain PAO1 and isogenic mutants created via insertion of a gentamicin resistance gene in the QS
97 genes *lasI* (PAO1 *lasI*::Gm; referred to as *lasI*$^-$ (31)) or *lasR* (PAO1 *lasR*::Gm; referred to as
98 *lasR*$^-$ (16)). To test 3O-C12-HSL diffusion in different media, we used a reporter strain of the
99 *lasI*$^-$ mutant. This contains a chromosomal *luxABCDE* fusion to the promoter of the *lasB* gene,
100 which encodes the QS-dependent protease LasB (PAO1 p*lasB::lux* (31)).

101

102 **Growth conditions.** Quorum sensing medium (QSM) was modified from two previous studies
103 (5, 32). QSM consisted of M9 Minimal Salts supplemented with 6.8 g/L Na_2HPO_4, 3 g/L
104 KH_2PO_4, 0.5 g/L NaCl. 10μM NH_4Cl, 0.1 μM $CaCl_2$ and 1 μM $MgSO_4$. QSM was supplemented
105 with 0.1 % w/v of carbon sources as a mix of Casamino acids (CAA) and adenosine and the
106 medium was filter sterilized. The exact ratio of CAA and adenosine was varied as detailed in the
107 Results section. Liquid culture experiments were conducted in 24-well plates with a volume of 2
108 mL of media. Cultures were incubated overnight in LB medium at 37°C on an orbital shaker and
109 standardised to an OD_{600} of 0.8-0.9; 2 μL of pure or mixed inoculum was added to each
110 experimental culture. The starting frequency of the mutant was determined by diluting and
111 plating the starter cultures to determine the number of colony-forming units (CFU) of each

112 genotype. Experimental cultures in QSM were incubated at 37°C with orbital shaking for 24h or
113 48 h. After that time, cultures were diluted and replica plated on LB and LB + gentamycin (25
114 µg/ml) agar to enumerate the CFU of PAO1 and *lasI* or *lasR* mutants in mixed culture.
115 Experiments in solid media were conducted in QSM + 2% w/v agar in 1 ml volumes in 48-well
116 plates. Inoculation and culture conditions were otherwise identical to experiments in liquid
117 medium. To break up agar prior to dilution and plating, the solid 1 ml agar cubes were retrieved
118 from the plate and divided into thirds with a sterile metal spatula; each third was placed in a
119 screw-cap tube containing 500 µl phosphate-buffered saline and 6 metal beads (Cambio) and
120 homogenised using a FastPrep-24 5G bead beater (MP Biomedicals).

121

122 **Measures of signal concentration.** To measure the concentration of QS signal (*N*-3-oxo-
123 dodecanoyl-L-homoserine lactone; 3O-C12-HSL) present in 48 h cultures, 100 µl of each culture
124 supernatant was mixed with 100 µl of a log phase culture of a luminescent *Escherichia coli*
125 bioreporter (pSB1075; (33)) in the wells of a 96-well plate. This mixture was incubated for 4 h in
126 a Tecan multimode plate reader and luminescence and OD_{600} recorded at 15-min intervals. To
127 estimate 3O-C12-HSL concentration, the luminescence of experimental samples was compared
128 against a calibration curve constructed using QSM supplemented with known concentrations of
129 purified 3O-C12-HSL.

130

131 **Assaying the effect of agar on QS signal diffusion**. Agar has been successfully used to retard
132 the diffusion of other bacterial exoproducts (27). To verify that agar affects 3O-C12-HSL
133 diffusion in QSM, and to determine the optimal agar concentration to use in further experiments,
134 we devised a "sandwich experiment" in which a population of bacteria that switch on a
135 luminescent reporter gene in response to QS signal, but which cannot themselves produce signal,
136 were separated from a reservoir of purified signal by a layer of agar-supplemented medium. By
137 measuring the time to expression of the luminescent reported, we can assess the extent to which
138 the agar barrier delays diffusion of the signal from the reservoir to the reporter population. 0.1
139 ml LB supplemented with 0.5% w/v agar and containing 0.5 µM purified 3O-C12-HSL was
140 added to the wells of a 48-well plate and allowed to solidify. A second layer of 0.8 ml LB
141 supplemented with 1, 2, 3 or 4% w/v agar was then added on top of the signal-containing layer.

142 Each agar concentration was replicated in 6 wells. This layer was allowed to solidify and a final
143 layer of LB containing 0.5% w/v agar and the reporter PAO1 *lasI*⁻ p*lasB*::*lux* (overnight culture
144 at OD_{600} of 0.2) added. The plate was incubated in a Tecan multimode reader for 8 h and
145 luminescence read at 10 min intervals. As shown in Figure S1, increasing agar concentration
146 progressively delayed and reduced expression of luminescence. In order to check if higher
147 luminescence was due to increased bacterial numbers, bacteria were retrieved and CFU were
148 counted by plating. Median CFU was similar when 1 % or 2% agar was used (approx. 1.410^7),
149 but decreased by 30% when more agar was added (to approx. 110^7). It was difficult to determine
150 whether this was due to agar retarding growth at high concentrations, or simply due to the
151 increased difficulty of thoroughly homogenising media rich in agar. 1% agar was therefore
152 chosen for use in further experiments.

153

154 **Statistical analysis**. Relative fitness of mutants, v, was calculated as $x_2(1-x_1) / x_1(1-x_2)$, where x_1
155 is the starting frequency of the mutant and x_2 is the end frequency. It follows from the definition
156 that a relative fitness <1 signifies a decrease in mutant frequency, while a relative fitness >1
157 signifies an increase in mutant frequency. Statistical analysis of the results was conducted in R
158 3.2.3 (34) using generalized linear models assuming an underlying gamma distribution, with
159 adenosine treated as a continuous variable and block and treatment (liquid/solid medium) fitted
160 as factors. Raw data for all analyses reported is supplied as Supplemental Material (Data S1).

161

162 **Results**

163

164 ***las* mutants grow poorly in an environment where QS is required for growth**. Previous work
165 has shown that *las* mutants grow poorly in media where QS is required for growth (5, 7, 35). We
166 first confirmed that both *lasI*⁻ and *lasR*⁻ mutants were reduced in fitness in the specific medium
167 we chose for our experiments. We grew PAO1 and each mutant in a minimal medium base
168 containing 0.1% w/v carbon source. The carbon source was composed of casamino acids (CAA,
169 available for use by all cells, regardless of genotype) and adenosine (which can only be
170 metabolised when QS is functional in cells), in varying ratios. As the relative amount of

171 adenosine increased and cells were increasingly dependent on QS, the total cell density after 48 h
172 was reduced, and this effect was more pronounced in *lasI* monocultures than in wild-type
173 monocultures (Figure 1, CFU pure adenosine of CFU pure CAA: 4.6% for wild type, 0.1-0.2%
174 for *las* mutants). When all the available carbon was supplied as adenosine, *lasI*$^-$ monocultures
175 grew to approximately 7% of the density of wild-type monocultures.

176

177 **Signal-negative *lasI*$^-$ mutants act as social cheats in adenosine-based growth medium, but**
178 **signal-blind *lasR*$^-$ mutant do not.** We next tested whether adding purified 3O-C12-HSL, or co-
179 culturing with signal-producing wild-type bacteria, could restore the growth of *lasI*$^-$ mutants. We
180 calculated the fitness of *lasI*$^-$ mutants relative to the wild type (i) in pure culture with or without
181 exogenous 3O-C12-HSL, and (ii) in 1:1 co-culture with wild-type PAO1. Experiments were
182 conducted in quorum-sensing media with varying ratios of CAA and adenosine as above. A
183 relative fitness of 1 signifies similar growth of mutant and wild type bacteria, while values <1
184 reflect relatively poorer growth of the mutant and values >1 reflect better growth of the mutant.
185 Figure 2 shows fitted models describing how the relative fitness of *lasI*$^-$ (a,b) and *lasR*$^-$ (c,d)
186 mutants are affected by culture conditions. The raw data to which the models were fitted are
187 plotted in Figure S2.

188

189 Pure *lasI*$^-$ cultures became progressively less fit than the wild type as access to carbon depended
190 more on quorum sensing (negative correlation between relative fitness and % adenosine:
191 coefficient -21.6, $p < 0.001$). However, when 10 µM exogenous 3O-C12-HSL was supplied,
192 *lasI*$^-$ mutants surpassed the wild type in growth (positive correlation between relative fitness and
193 % adenosine: coefficient 8,74, $p = 0.003$; Figure 2a, S2a) This result is consistent with previous
194 work demonstrating a cost to 3O-C12 production (18). As predicted, this ability of *lasI*$^-$ mutants
195 to use exogenous signal, combined with the cost of signal production to the wild type, means that
196 *lasI*$^-$ mutants grown in co-culture with wild type bacteria act as social cheats: the average relative
197 fitness was consistently > 1 and did not decline as % adenosine increased (coefficient 1.8, $p =$
198 0.88 Figure 2b, S2a). There was, however, a slight drop in relative fitness when all carbon was
199 supplied as adenosine (Figure S2a. This is most likely attributable to the wild-type bacteria
200 growing more slowly and taking longer to fully switch on QS responses. Both wild-type growth
201 (Figure 1) and the pool of available signal (Figure S3) are reduced in this condition, leaving less
202 opportunity for exploitation by cheats.

203

204 Post-hoc comparisons confirmed that *lasI*$^-$ relative fitness was significantly increased in mixed
205 populations vs. pure cultures in all media containing adenosine (t-tests, $p < 0.01$). When all
206 carbon was supplied as CAA and signal is not required, there was no significant effect of pure vs.
207 mixed culture on fitness ($p > 0.95$) Taken together with the fact that mixed cultures grew to a
208 lower density than the wild-type cultures (Figure S5), these results indicate that *lasI* mutants
209 have an increased fitness when grown in the presence of wild-type bacteria under conditions
210 requiring social interaction, while in turn decreasing wild-type fitness.

211

212 To ensure that the effect described above was due to the social dynamics of signal production,
213 and not to the well-documented social dynamics of downstream exoproducts, this experiment
214 was repeated using a *lasR*$^-$ mutant. *lasR*$^-$ mutants are unable to respond to 3O-C12-HSL and
215 should therefore not be able to derive fitness benefits from exogenous signal in our setup. We
216 found the same negative correlations between % adenosine and CFU (Figure 1) and % adenosine
217 and fitness relative to the wild type (Figure 2c, S2b, coefficient -12.8, $p = 0.001$) as with the *lasI*$^-$
218 mutant for *lasR*$^-$ monocultures. Crucially, *lasR*$^-$ relative fitness was not rescued by adding 3O-
219 C12-HSL or by co-culturing with the wild type (coefficient -10.7, $p < 0.001$; Figure 2c,d and
220 S2b): i.e. these mutants could not exploit wild type bacteria.

221

222 **Slowing signal diffusion reduces *lasI*$^-$ mutant cheating.** As a last step, we tested whether
223 impeding the diffusion of signal molecules would make the *lasI*$^-$ mutant less effective as a cheat.
224 Reduced diffusion was achieved by adding agar to solidify the growth medium(Figure S1). *lasI*$^-$
225 monocultures showed comparable declines in fitness in liquid and solid medium (Figure 3a, S4a.
226 ANOVA: liquid/solid $F_{(1,124)} = 6.13$, $p = 0.01$; adenosine $F_{(1,123)} = 117.58$, $p < 0.001$; interaction
227 $F_{(1,122)} = 0.1658$, $p = 0.68$). In mixed cultures, the relative fitness of the *lasI*$^-$ mutant was
228 positively correlated with the percentage of carbon available as adenosine in liquid cultures, as
229 expected under cheating: but when the medium was solidified, *lasI*$^-$ relative fitness actually
230 showed a modest negative correlation with percentage adenosine (Figure 3b, S4b. ANOVA:
231 liquid/solid $F_{(1,123)} = 11.1324$, $p = 0.001$; adenosine $F_{(1,124)} = 0.0236$, $p = 0.88$; interaction
232 $F_{(1,122)} = 0.9760$, $p = 0.33$). This demonstrates that there is less opportunity for cheating in an
233 environment where signal molecules cannot diffuse freely.

234

235 **Discussion**

While there has been some discussion of QS signals as public goods (e.g. (36)), most published work on the social evolution of QS focuses on signal-blind mutants and the benefits of cheating on the production of QS-regulated exoproducts(5, 7, 14, 32). Here we provide the first direct evidence that, in addition to regulating the production of public goods, QS signal molecules are themselves capable of acting as public goods. Social cheating in the context of QS can therefore take multiple forms, depending on the environmental circumstances in which bacteria find themselves. Previous research had shown that (a) signal-blind mutants can be cheats when growth depends on the production of QS-dependent extracellular enzymes; and (b) cheating by signal-blind mutants can be constrained when some 'private' QS-controlled processes contribute to growth (35). Following recent confirmation that QS signals are costly to make (18), we now add two new perspectives to the social evolution of QS: (c) that signal-negative mutants can be cheats when growth depends entirely on private goods, regardless of any downstream effects on exoenzyme production; and (d) that this signal cheating can only occur when the environment permits sufficient diffusion of signal molecules.

A *lasI*$^{-}$ mutant grew poorly in monoculture, but growth could be rescued by adding either purified 3O-C12-HSL signal or by co-culturing with wild-type, signal-producing bacteria. In co-cultures, the fitness of *lasI*$^{-}$ mutants relative to the wild type increased as we forced the bacteria to rely more on adenosine for carbon. As the population became more reliant on QS, *lasI* mutants gained a greater fitness pay-off from exploiting costly, diffusible signal produced by the wild-type. *lasR*$^{-}$ mutants did not gain a similar advantage from co-culture with the wild type in adenosine medium. These signal-blind mutants can take up signals, but cannot respond to them and so cannot switch on expression of the *nuh* hydrolase required for growth on adenosine (21, 35). The contrasting results for the two different QS mutants confirm that, in this environment, the QS signal itself acts as a public good (Figure 4)

Adding agar to the growth medium lowered both the diffusion of signal molecules and the relative fitness of cheats in mixed culture. Thus adding simple spatial structuring into our system had a significant impact on the ability of signal-negative mutants to cheat on the wild type. There was no effect of structuring on relative fitness in pure cultures: even though agar enhanced the overall growth rate of the bacteria, the basic costs and benefits of signalling remained the same in liquid and solid media. This final observation is consistent with work on other bacterial public

goods (27). We thus predict that the evolution of QS signalling strategies will be influenced by population genetic and spatial structure, and that signal-negative cheats might only rise to appreciable frequencies in environments where signals diffuse freely. For example, the thick, adhesive mucus and bacterial biofilm polymers that block the airways of cystic fibrosis patients with chronic lung infection may partially protect producers from cheating by signal-negative mutants (24). Spatial dynamics play a huge role in the real-life ecology of environmentally and clinically important microbial ecosystems, and are therefore of considerable interest to microbiologists investigating the roles of bacteria in processes as diverse as geochemical cycling, soil health, fouling and infection (29, 37, 38).

Our work opens up new avenues for exploring how, when and why bacterial signalling evolves in different environments and why we find a variety of QS mutant genotypes and phenotypes in natural environments (8-14, 39, 40). Given what we are now learning about the evolution of traits such as QS and how spatial structure changes the evolutionary dynamics, we suggest that there is a need to carefully consider the experimental design of *in vitro* experiments to increase their relevance to actual infections (41). To be forewarned is to be forearmed: a more accurate understanding of microbial ecology and evolution, gained from more realistic lab experiments, will be a vital weapon in the fight against antibiotic resistant infection.

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417

418 **Figure legends**

419

420 **Figure 1.** Population density (colony-forming units: CFU) after 48h of growth in quorum
421 sensing medium with varying ratios of casamino acids:adenosine supplied to a total
422 concentration of 0.1% w/v carbon source. Black dots: indvididual wild-type cultures; grey dots:
423 indicate *lasI*$^-$ cultures*;* white dots: *lasR* cultures.

424

425 **Figure 2.** Results of fitting generalized linear models to relative fitness data from experiments
426 with *lasI*$^-$ (a,b) and *lasR*$^-$ (c,d) mutants. Lines show fitted models, shaded areas denote standard
427 deviation. Relative fitness is compared between pure cultures of each mutant with added 3O-
428 C12-HSL or with a solvent-only control (a,c), and between pure cultures and mixed culture with
429 wild-type bacteria (b,d). Raw data for these experiments are shown in Figure S2.

430

431 **Figure 3.** Comparison of *lasI*$^-$ relative fitness in pure culture (a) or mixed culture with the wild
432 type (b). Results of fitting generalized linear models to relative fitness data from experiments in
433 liquid *versus* agar-supplemented medium. Lines show fitted models, shaded areas denote
434 standard deviation. Raw data for these experiments are shown in Figure S4.

435

436 **Figure 4.** Schematic of the effects of LasIR quorum sensing on the production of intracellular
437 and extracellular enzymes by *Pseudomonas aeruginosa*.

438

439 **Figure S1**. Supplementing growth medium with agar retards diffusion of 3O-C12-HSL. (a)
440 Expression of a 3O-C12-HSL-dependent luminescent reporter construct is delayed and reduced
441 when signal-negative reporter bacteria are separated from a reservoir or purified signal by an
442 agar barrier. Lines and shading show means and s.d. of 15 replicates. (b) The experimental set-
443 up, as described in the Materials and Methods, is easily visualised by replicating the experiment
444 on a larger scale and adding red food colouring in place of bacterial signal.

445

446 **Figure S2**. Relative fitness of (a) *lasI*$^-$ mutants and (b) *lasR*$^-$ mutants when in pure culture (black
447 circles), grown in 1:1 mixture with wild type (grey circles) and in pure culture supplemented
448 with 10 μM 3O-C12-HSL (white circles).

449

450 **Figure S3.** Measurements of 3O-C12-HSL concentration in wild-type supernatant after 48h. (a)
451 The total concentration of 3O-C12-HSL declines with increasing adenosine. (b) After dividing
452 the signal concentration by the CFU of the producing culture, one observes the reverse trend –
453 concentration of 3O-C12-HSL per CFU increases with adenosine level and reaches its maximum
454 in pure adenosine.

455

456 **Figure S4.** Relative fitness of *lasI*$^-$ mutants in liquid medium (black circles) *versus* agar-
457 supplemented medium (grey circles), (a) in pure culture and (b) in mixed culture with the wild
458 type.

459

460 **Figure S5.** Population density (colony-forming units: CFU) after 48h of growth in quorum
461 sensing medium, Blue dots: indvididual wild-type cultures; yellow dots: indicate *lasI* and wild-

462 type mixed cultures; red dots: *lasI* cultures.

463

464 **Data S1.** Data for all analyses reported in the text will be made available on with the published
465 version of this article.

466

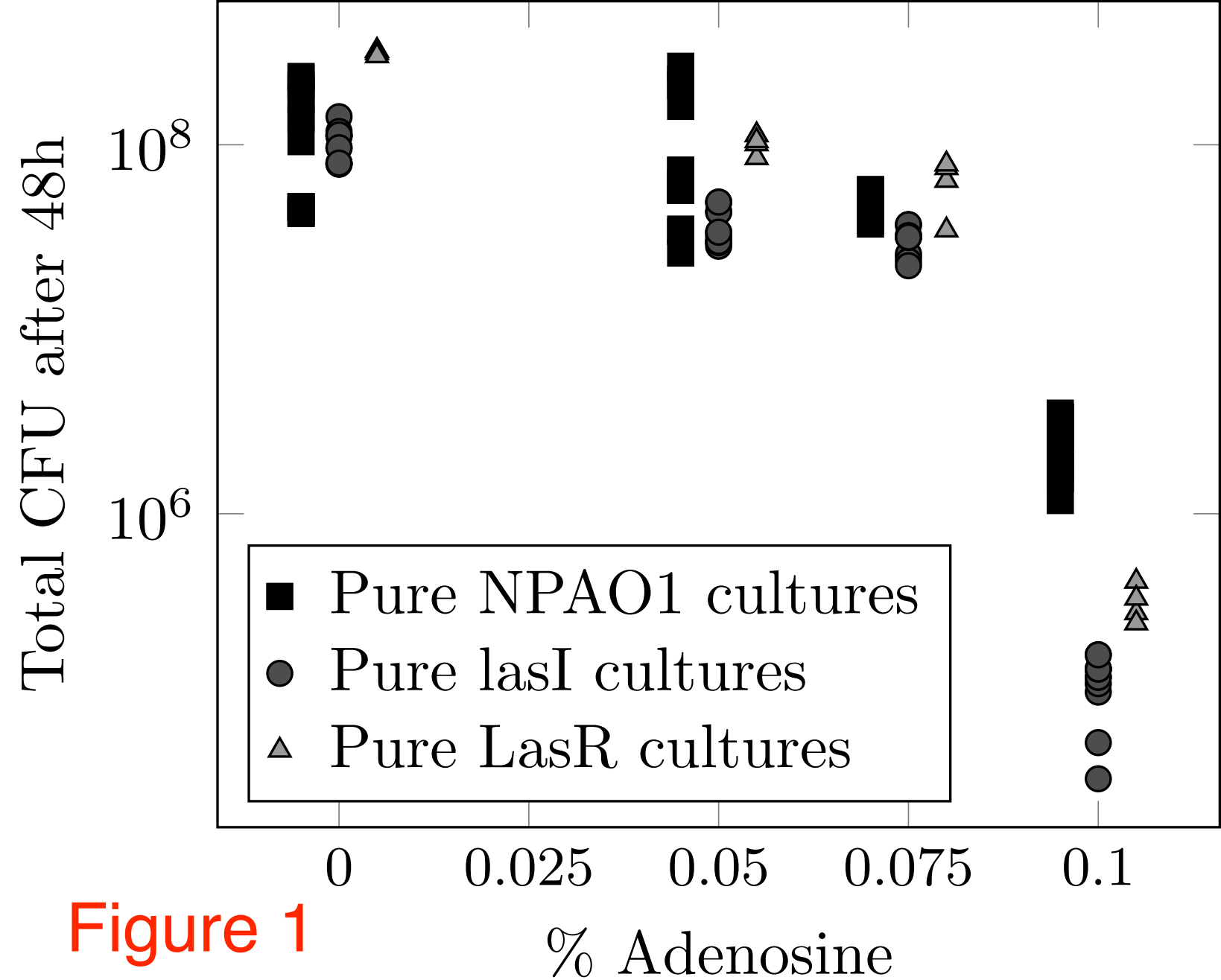
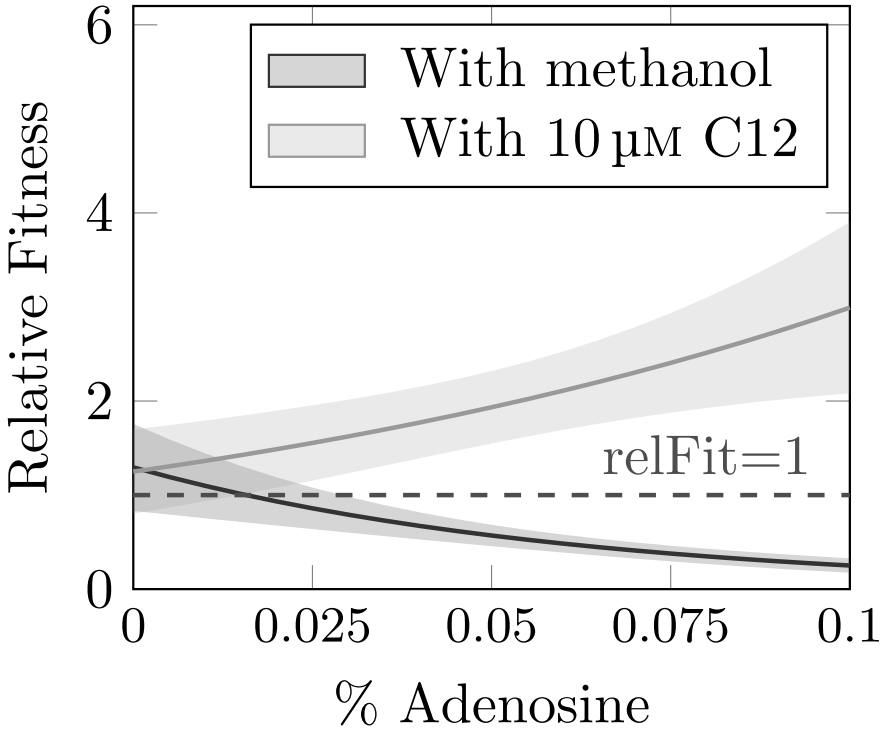
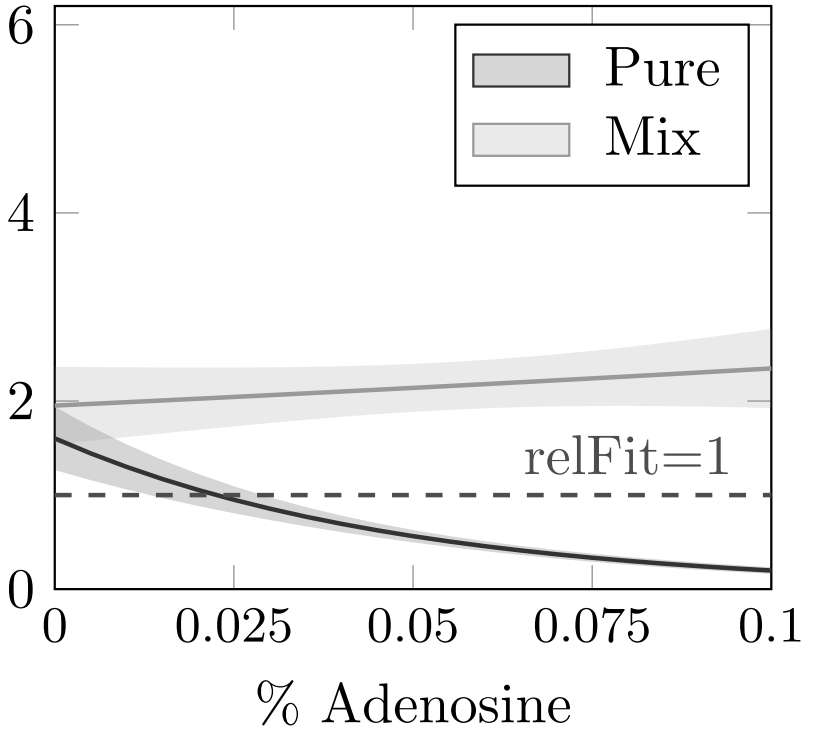

Total CFU after 48h
10^8
10^6
Pure NPAO1 cultures
Pure lasI cultures
Pure LasR cultures
0
0.025
0.05
0.075
0.1
% Adenosine

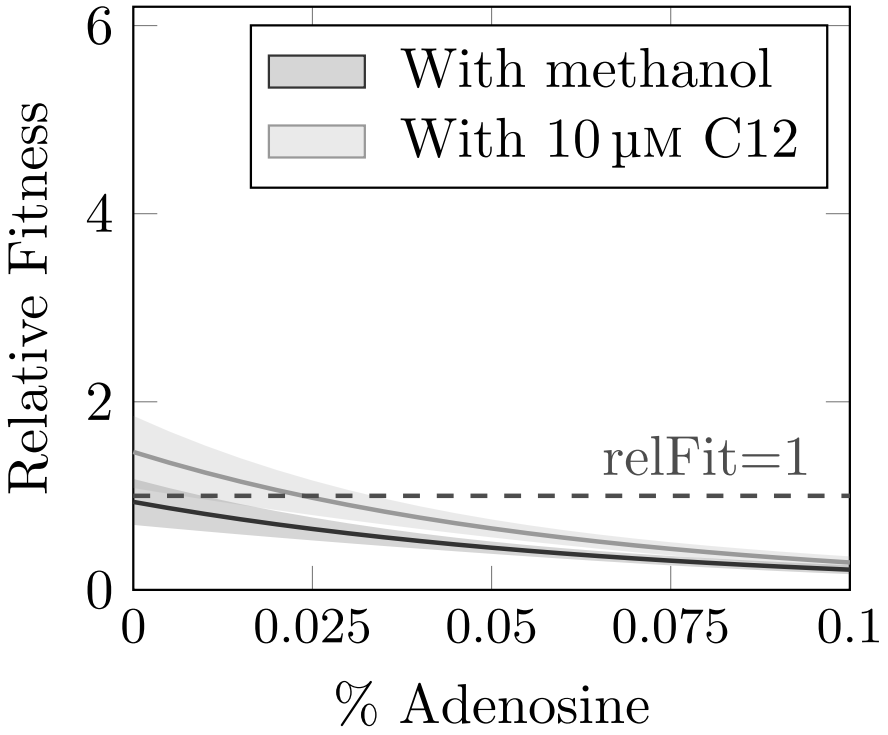
Figure 1



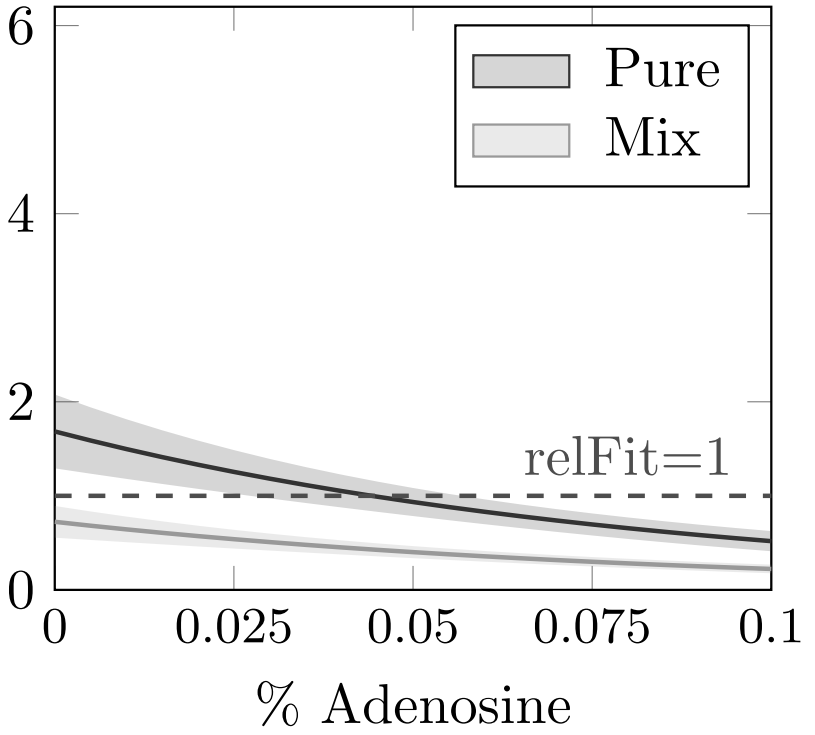
(a) lasI fitness with 10 µM C12



(b) lasI fitness in mixed/pure culture



(c) lasR fitness with 10 µM C12



(d) lasR fitness in mixed/pure culture

Figure 2

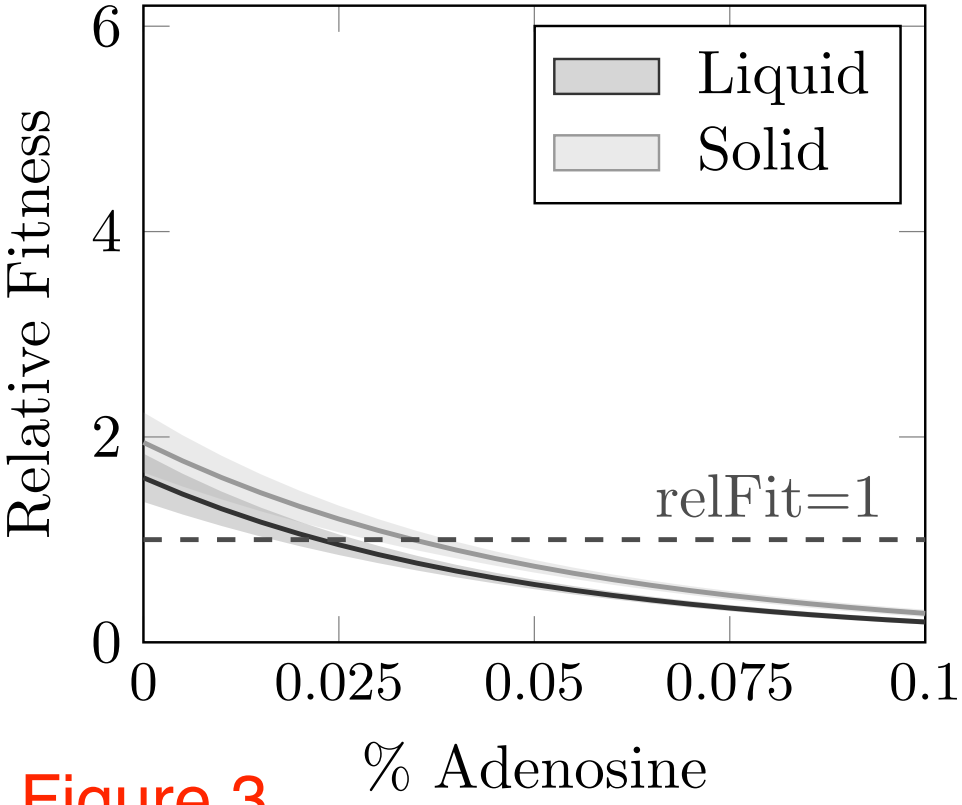
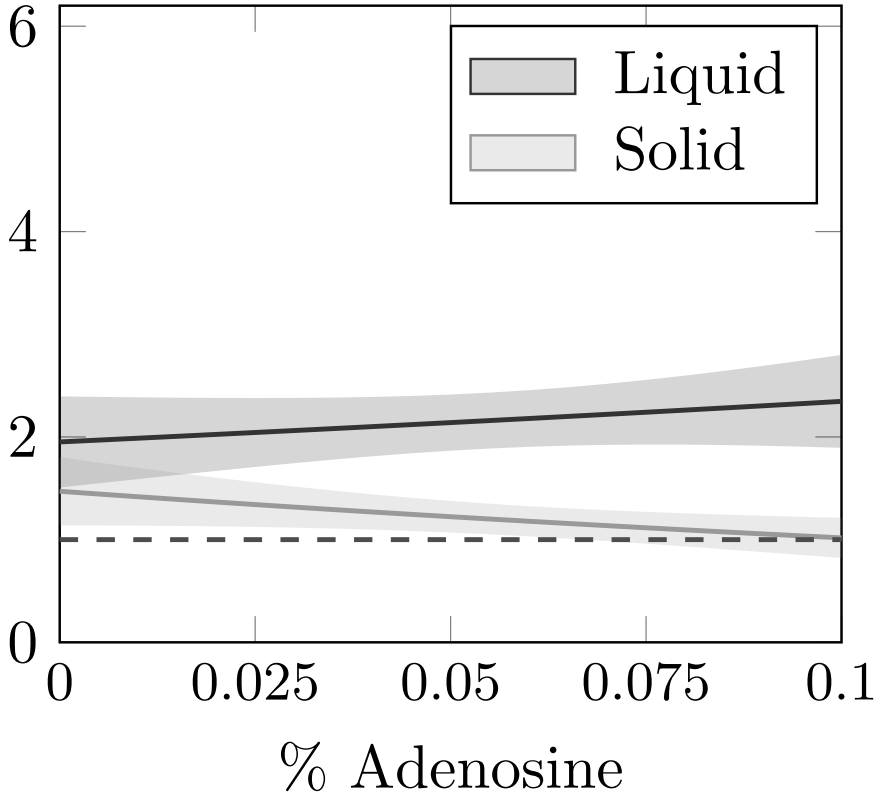


Figure 3

(a) Fitted model for relative fitness of pure cultures



(b) Fitted model for relative fitness of mixed cultures

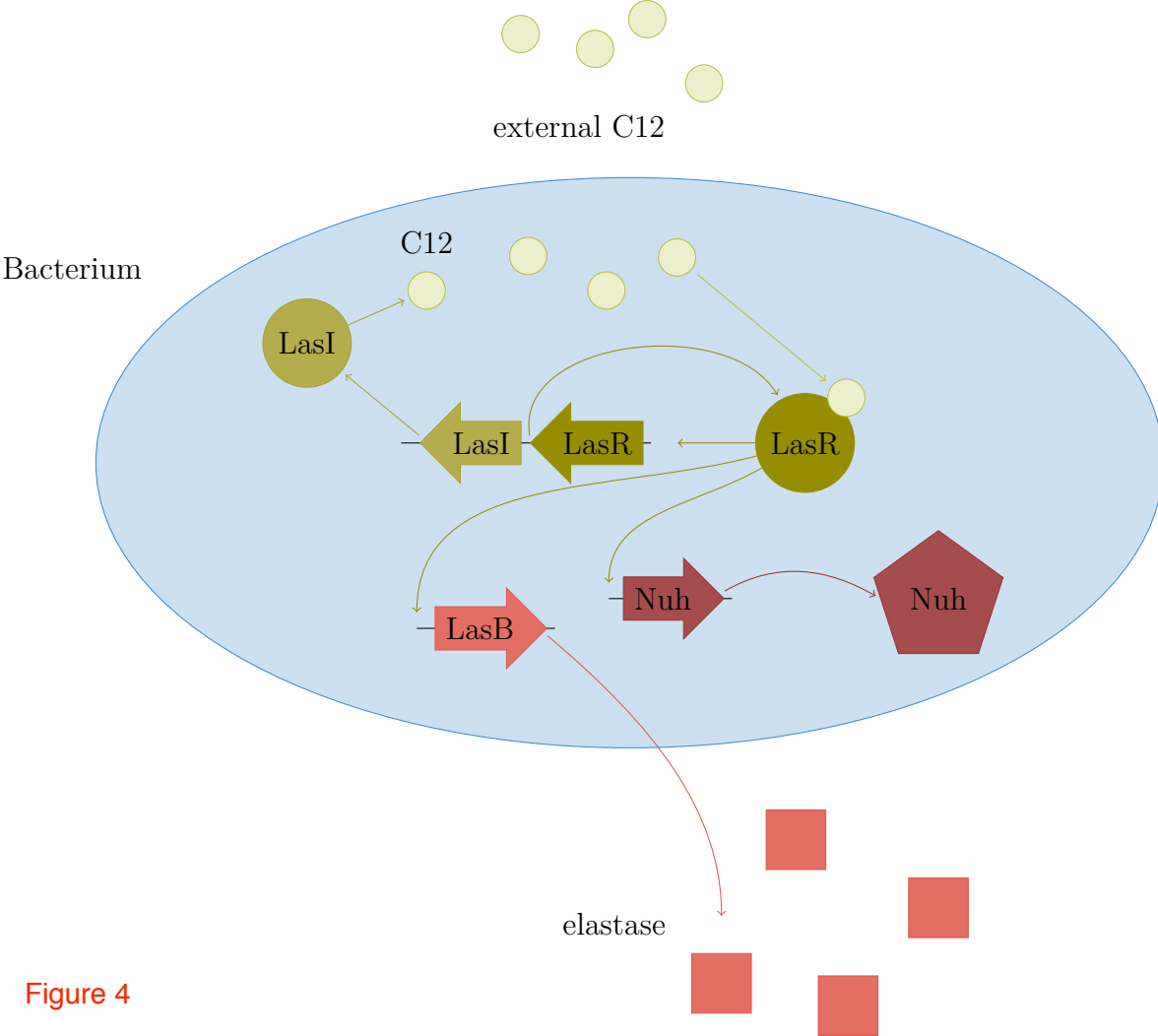

external C12
Bacterium
C12
LasI
LasI
LasR
LasR
Nuh
Nuh
LasB
elastase

Figure 4

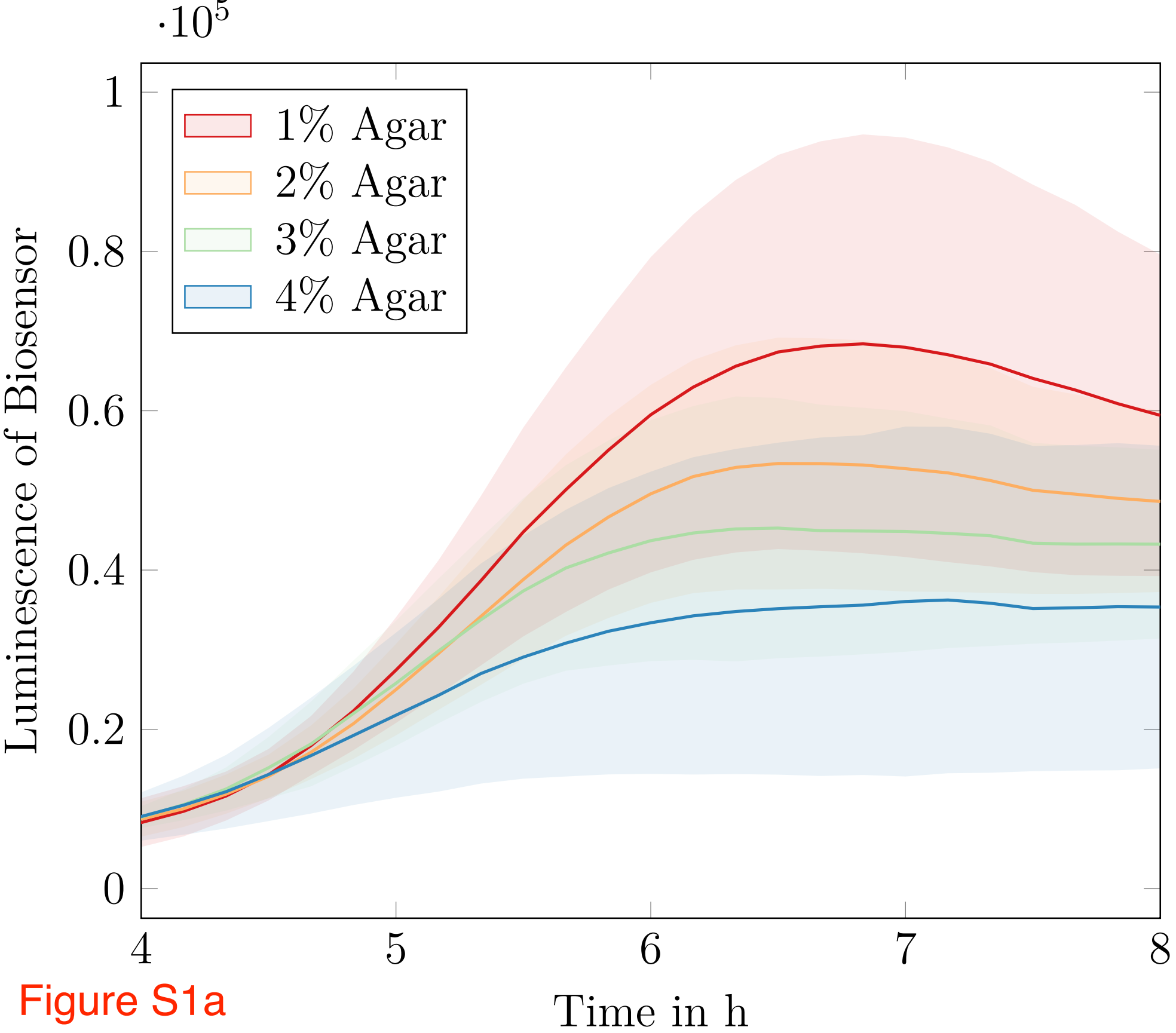

$\cdot 10^5$
1
0.8
0.6
0.4
0.2
0
Luminescence of Biosensor
1% Agar
2% Agar
3% Agar
4% Agar
4
5
6
7
8
Time in h

Figure S1a

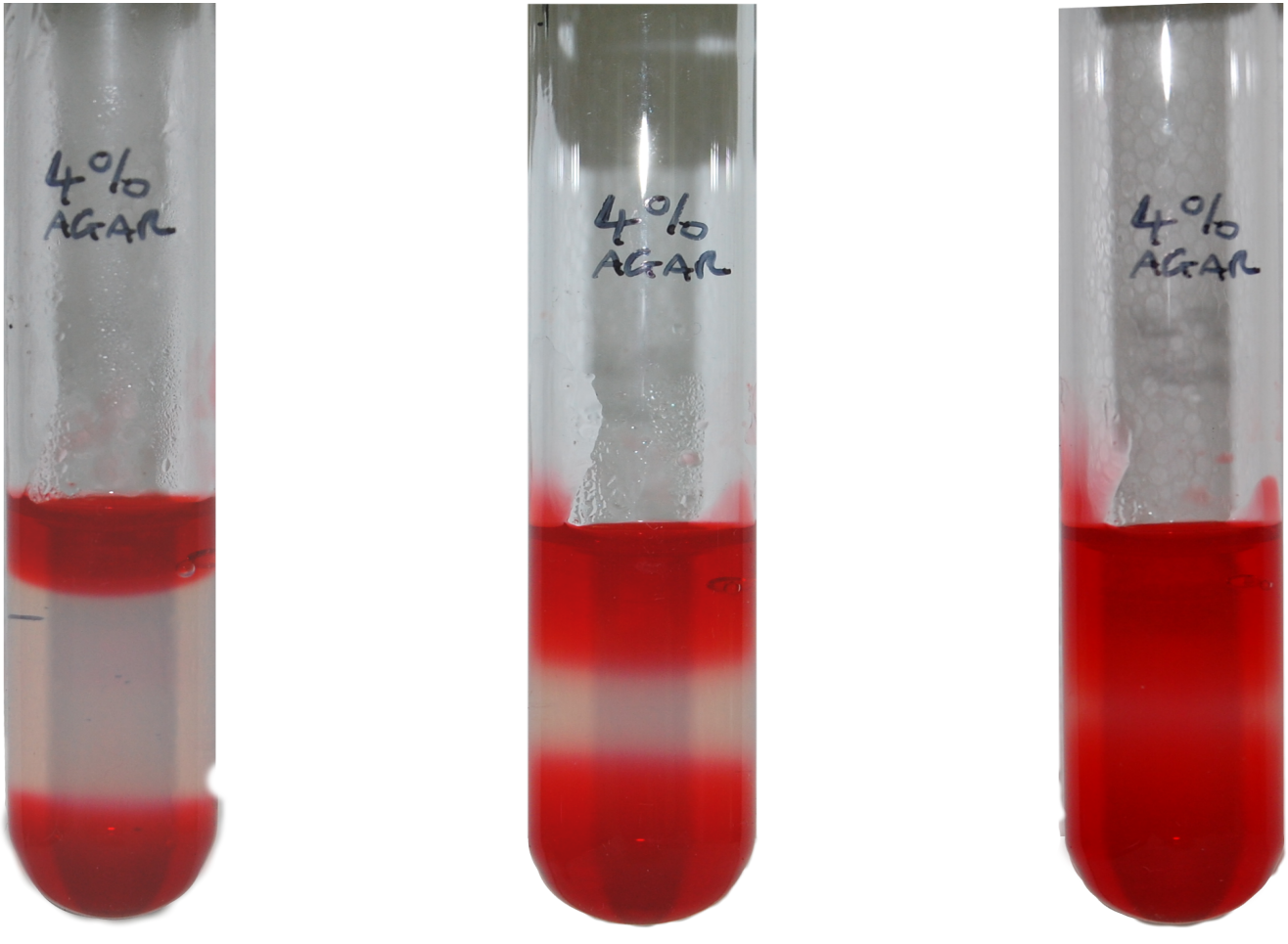


Figure S1b

time

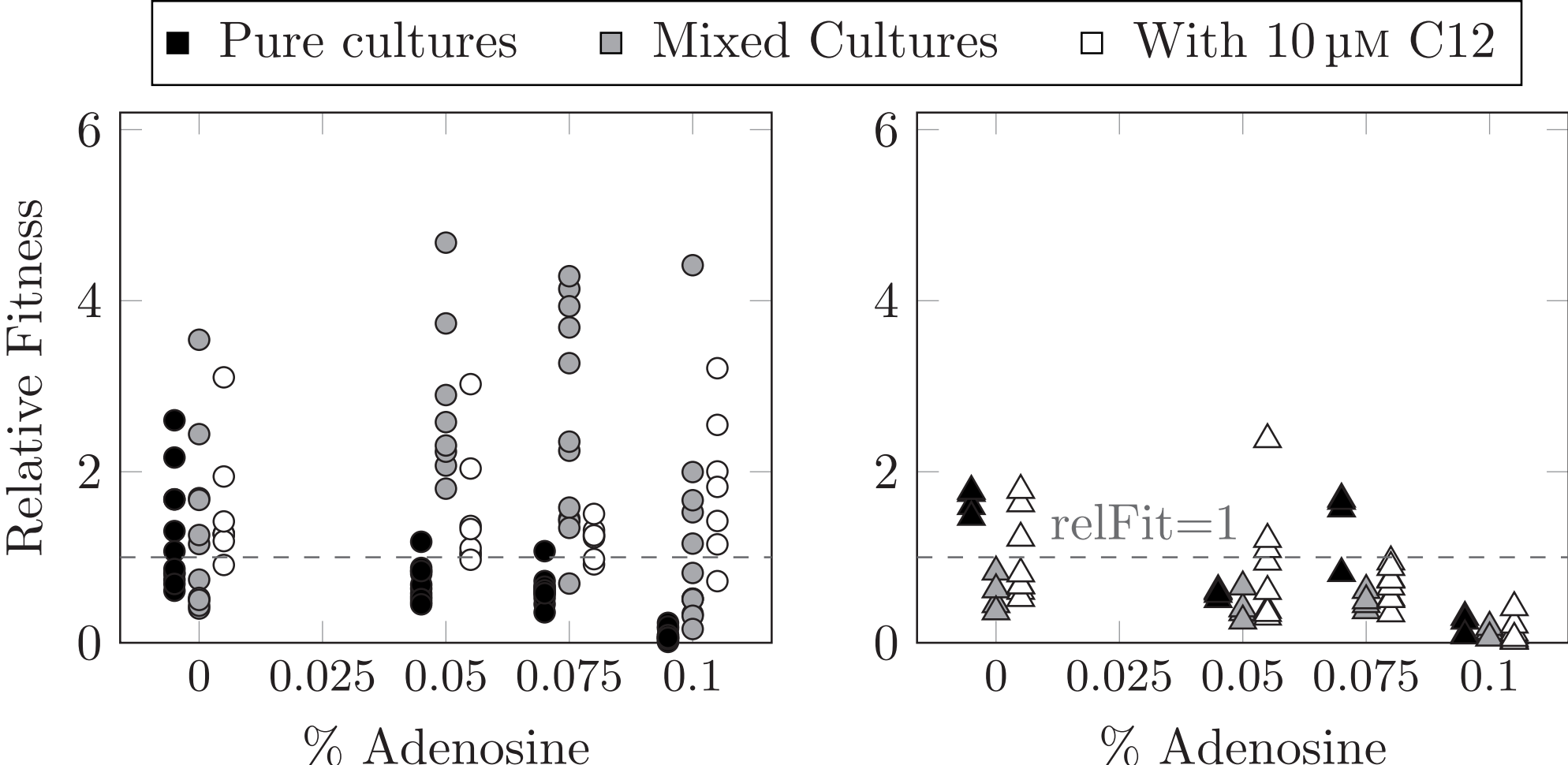


Figure S2

(a) Relative fitness of lasI⁻

(b) Relative fitness of lasR⁻

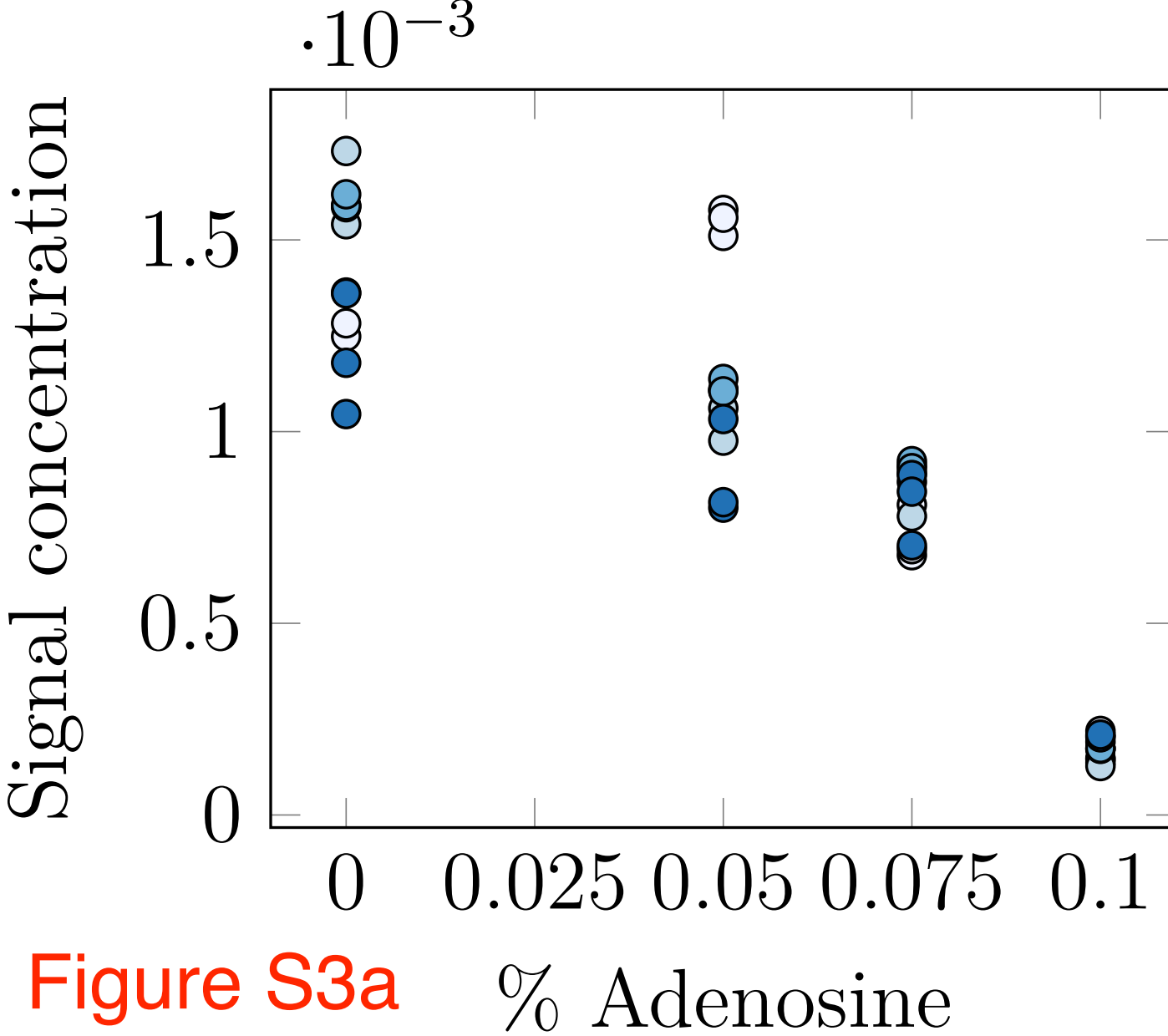

$\cdot 10^{-3}$
Signal concentration
1.5
1
0.5
0
0
0.025
0.05
0.075
0.1
% Adenosine

Figure S3a

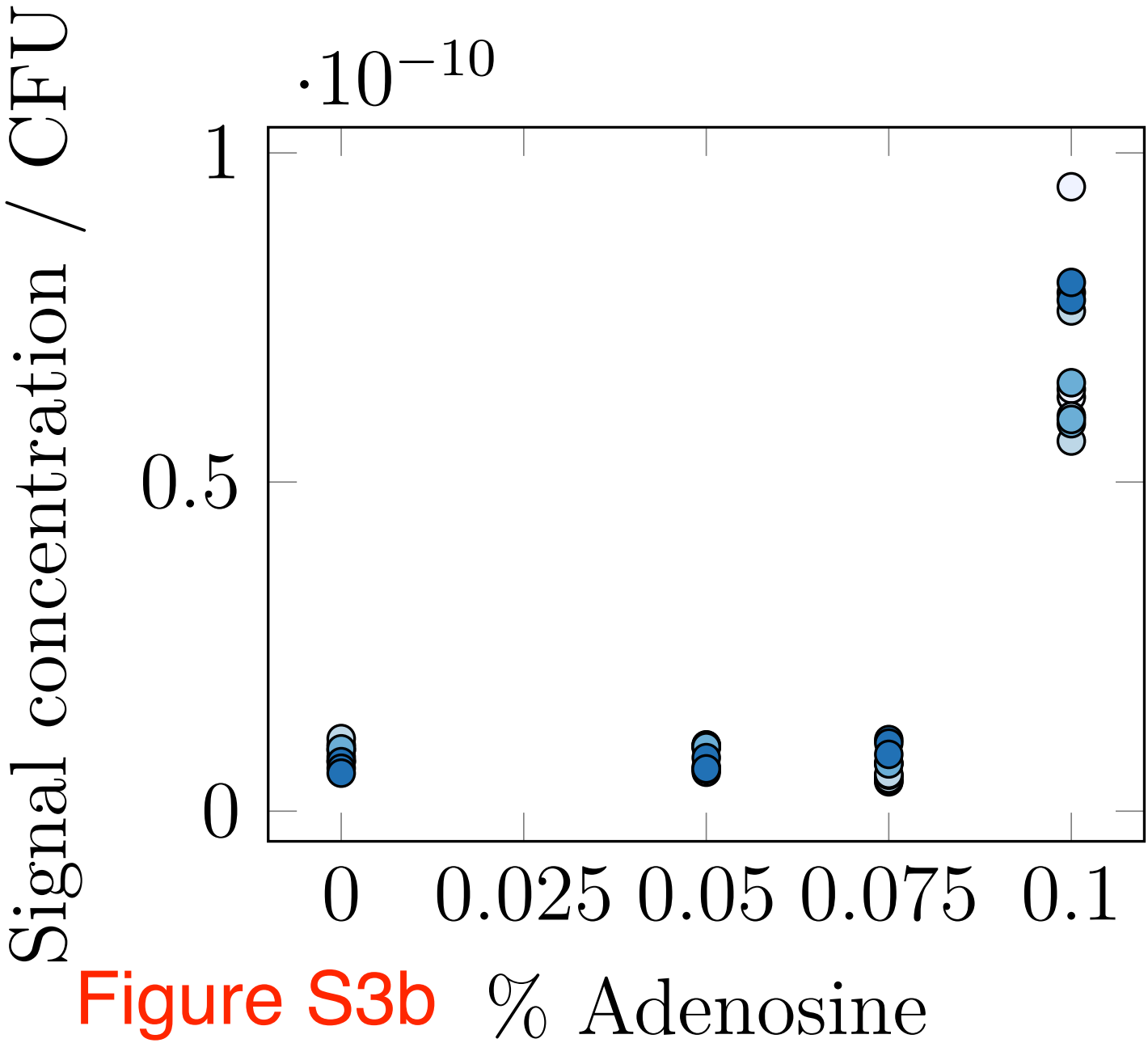
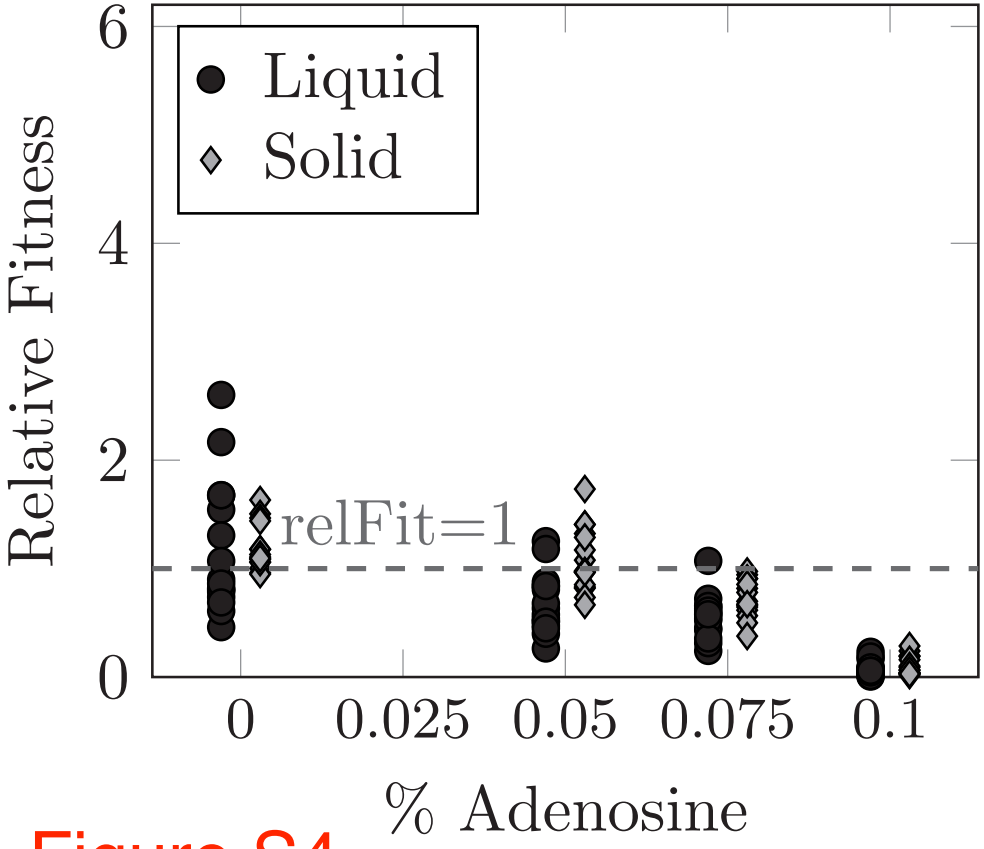

$\cdot 10^{-10}$
1
0.5
0
Signal concentration / CFU
0
0.025
0.05
0.075
0.1
% Adenosine

Figure S3b



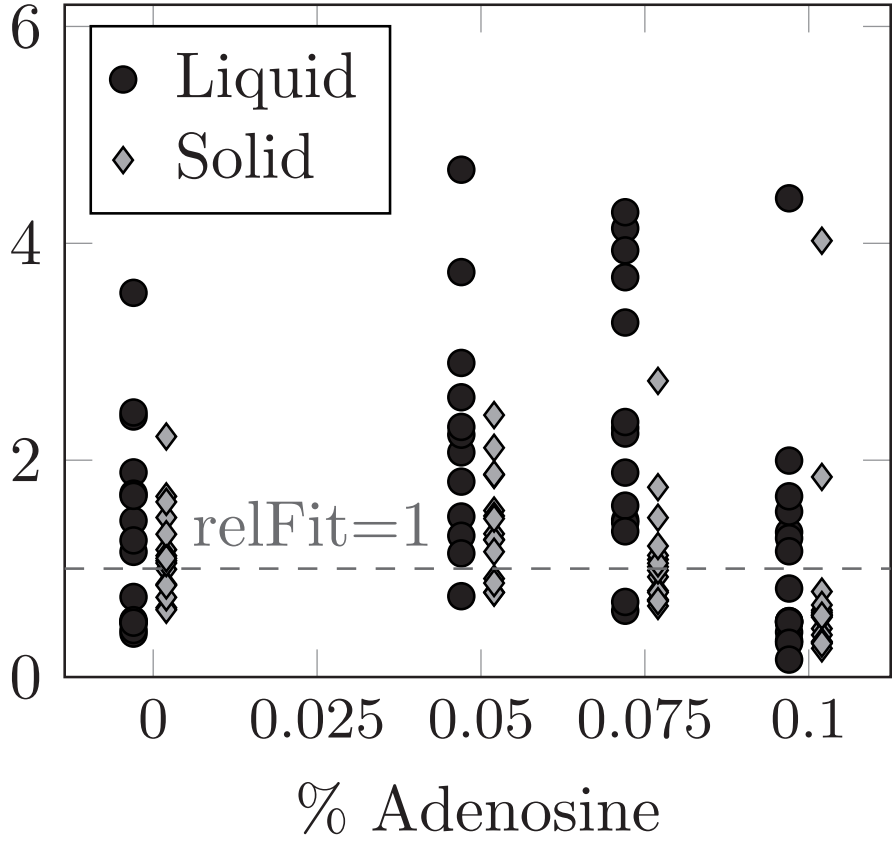


Figure S4

(a) Pure lasI⁻ cultures

(b) Mixed cultures

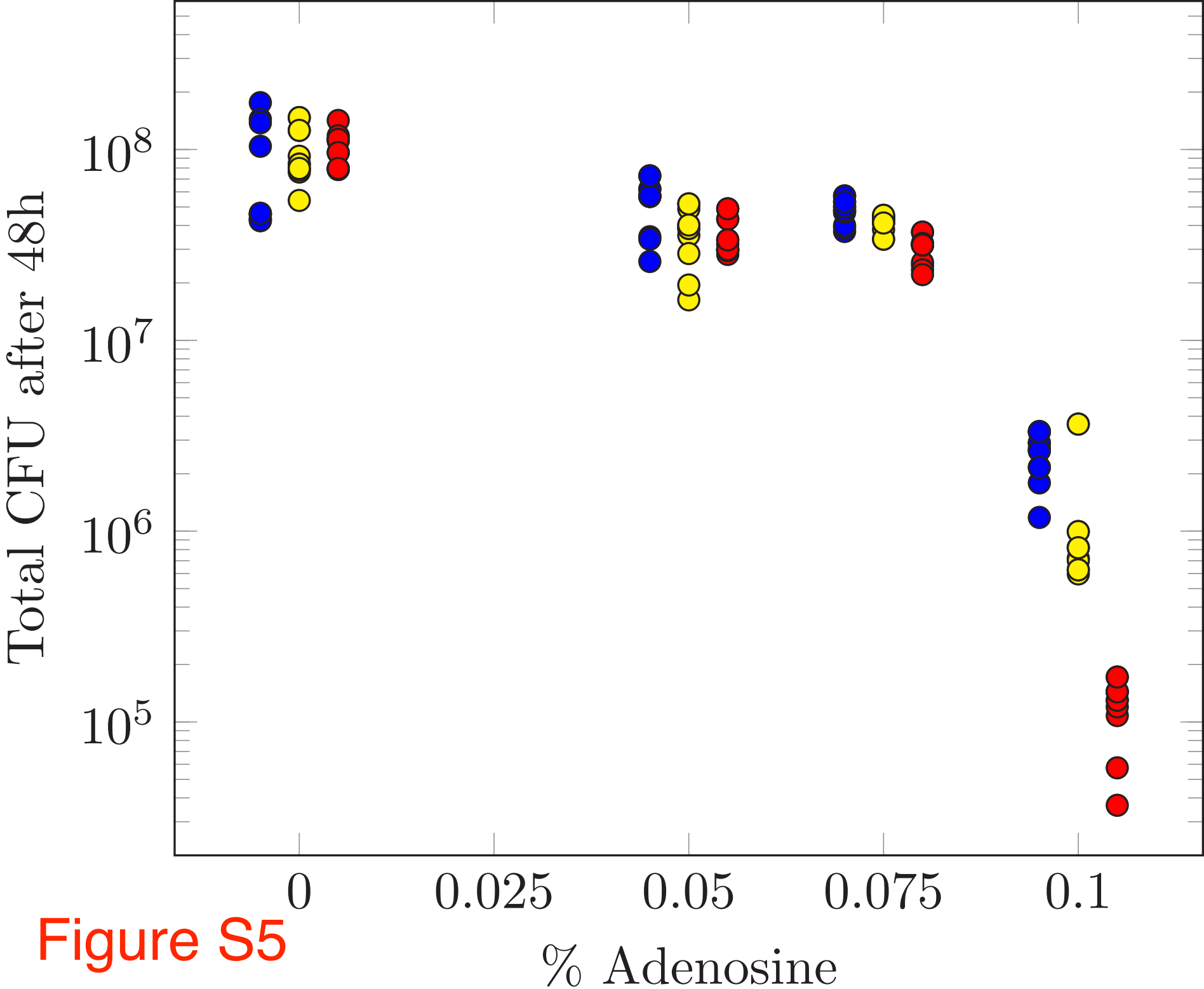

Total CFU after 48h
10^8
10^7
10^6
10^5
0
0.025
0.05
0.075
0.1
% Adenosine

Figure S5