1 TITLE:

5

13

- 2 Inferring the role of habitat dynamics in driving diversification: evidence for a species pump
- 3 in Lake Tanganyika cichlids
- 4 AUTHORS: Thijs Janzen<sup>1,2,3\*</sup>, Rampal S. Etienne<sup>2</sup>
- 6 AUTHOR AFFILIATION
- 7 1. Max Planck Institute for Evolutionary Biology, Department of Evolutionary Theory,
- 8 August-Thienemann-Straße 2, 24306, Plön, Germany.
- 9 2. University of Groningen, Groningen Institute for Evolutionary Life Sciences, Box
- 10 11103, 9700 CC Groningen, The Netherlands
- 3. Carl von Ossietzky University of Oldenburg, Institute for Biology, Department of
- Molecular Ecology, Carl-von-Ossietzky Straße 9, 26111, Oldenburg, Germany
- 4. \*Corresponding author: Thijs Janzen, Carl von Ossietzky University of Oldenburg,
- Institute for Biology, Department of Molecular Ecology, Carl-von-Ossietzky Straße 9,
- 16 26111, Oldenburg, Germany
- e-mail: thijs.janzen@uni-oldenburg.de

## ABSTRACT

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

Geographic isolation that drives speciation is often assumed to slowly increase over time, for instance through the formation of rivers, the formation of mountains or the movement of tectonic plates. Cyclic changes in connectivity between areas may occur with the advancement and retraction of glaciers, with water level fluctuations in seas between islands or in lakes that have an uneven bathymetry. These habitat dynamics may act as a driver of allopatric speciation and propel local diversity. Here we present a parsimonious model of the interaction between cyclical (but not necessarily periodic) changes in the environment and speciation, and provide an ABC-SMC method to infer the rates of allopatric and sympatric speciation from a phylogenetic tree. We apply our approach to the posterior sample of an updated phylogeny of the *Lamprologini*, a tribe of cichlid fish from Lake Tanganyika where such cyclic changes in water level have occurred. We find that water level changes play a crucial role in driving diversity in Lake Tanganvika. We note that if we apply our analysis to the Most Credible Consensus (MCC) tree, we do not find evidence for water level changes influencing diversity in the Lamprologini, suggesting that the MCC tree is a misleading representation of the true species tree. Furthermore, we note that the signature of habitat dynamics is found in the posterior sample despite the fact that this sample was constructed using a species tree prior that ignores habitat dynamics. However, in other cases this species tree prior might erase this signature. Hence we argue that in order to improve inference of the effect of habitat dynamics on biodiversity, phylogenetic reconstruction methods should include tree priors that explicitly take into account such dynamics.

## **INTRODUCTION**

Environmental changes such as the formation of mountain ridges, the formation of rivers and the movement of tectonic plates have long been known to be important drivers of speciation (Coyne and Orr 2004). Repeated environmental changes may thus lead to diversification patterns. Cyclic changes in the environment can cause populations to continuously switch between an allopatric and sympatric stage, providing a continuously renewed potential for speciation. And these cyclic changes can in turn drive diversity towards levels unexpected given the current geography, sometimes referred to as a "species pump" (Heaney 1985; Rossiter 1995). Examples of species pumps include environmental fluctuations fragmenting habitats on the slopes of mountains (Weir 2006; Sedano and Burns 2010; Hutter et al. 2013), glaciations and postglacial secondary contacts (Barnosky 2005), sea level changes causing the fusion and fragmentation of islands (Glor et al. 2004; Thorpe et al. 2008, but see Papadopoulou and Knowles 2015), and fluctuations in water level causing fragmentation and fusion of lakes with uneven bathymetry, as in the African Rift Lakes (Cohen et al. 1997b; Alin et al. 1999; McGlue et al. 2008; Ivory et al. 2016).

The African Rift Lakes provide a good starting point in studying the interplay between cyclic habitat dynamics and speciation, because they have been subject to frequent water level changes (Cohen et al. 1997b; Alin and Cohen 2003; Ivory et al. 2016), and are well known for their tremendous biodiversity (Seehausen 2000, 2006; Turner et al. 2001; Wagner et al. 2012, 2014; Brawand et al. 2014). An estimated number of 2000 cichlid fish species (Turner et al. 2001) have evolved in the African Rift Lakes over the past 10 million years (Genner et al. 2007; Meyer et al. 2016), and comprise one of the most spectacular adaptive radiations

67

68

69

70

71

72

73

74

75

76

77

78

79

80

81

82

83

84

85

86

87

88

89

90

(Seehausen 2006). The most prominent water level changes took place in Lake Tanganyika, where the water level has dropped substantially on multiple occasions over the past million years, sometimes splitting the lake into multiple smaller lakes (Lezzar et al. 1996; Cohen et al. 1997a, 2007). Being the oldest lake of the three large rift lakes (Cohen et al. 1993), Lake Tanganyika contains the highest behavioral diversity (Konings 2007) and is the only lake with a highly resolved phylogeny for cichlid fish. Evidence for the influence of changing water levels comes from analysis of mitochondrial DNA, which shows that for Tropheus species, some populations have experienced secondary contact upon changes in water level, potentially increasing genetic diversity and driving speciation (Sturmbauer et al. 2001; Koblmüller et al. 2011; Sefc et al. 2017). Similar patterns were found for Variabilichromis moorii and Ophthalmotilapia nasuta (Sturmbauer et al. 2001), Telmatochromis temporalis (Winkelmann et al. 2016), and Altolamprologus (Koblmüller et al. 2016). Comparison of mitochondrial DNA between populations from deep and shallow areas emphasizes that the deep areas are habitats that are more persistent over time, with lower genetic variation (Nevado et al. 2013). Furthermore, Eretmodus lineages identified using mitochondrial DNA are strongly associated with the bathymetric basins of Lake Tanganyika (Verheyen et al. 1996), suggesting that they have independently diversified at low water level.

Aguilée et al. (2013) developed a model for the African Rift Lakes in which populations at different locations diverge from each other depending on the local habitat, and at the same time allowed for sympatric speciation by implementing assortative mating that allows for a single branching point in trait values. Over time the different locations become separated or are reconnected, and this may drive the formation of new species. The authors conclude that stable numbers of diversity are best obtained by a fragmented habitat with recurrent merged states and rapid fluctuations. However, Aguilée et al. (2013) do not compare their results to

empirical data. By contrast, Pigot et al (2010) used a spatially explicit model of landscape fragmentation, where consecutive splitting of species' geographic ranges drives speciation, and compared phylogenies generated with their model, with known bird phylogenies. They found that including this geographical context of speciation explains a large part of the features exhibited by the reconstructed avian trees. Hence, including a geographical context of speciation seems a promising research avenue.

Here, we provide a method to infer whether or how cyclic changes in the environment influence both the generation and the maintenance of biodiversity. We use an extension of the standard constant-rates birth-death model. Because deriving an expression for the likelihood of this model for a given set of phylogenetic branching times is difficult, but simulation of phylogenies under the model is easy, we used approximate Bayesian computation (ABC) based on sequential Monte Carlo sampling (SMC) to estimate parameters from phylogenies. We applied our approach to an updated phylogeny of the *Lamprologini*, a tribe of cichlid fish from Lake Tanganyika in order to assess the importance of these habitat dynamics in shaping the current biodiversity of cichlids in Lake Tanganyika.

#### **METHODS**

Model

110

111

112

113

114

115

116

117

118

119

120

121

122

123

124

125

126

127

128

129

130

131

132

133

To model the interaction between environmental change and speciation, we envisage a lake that consists of a single pocket at high water level, but that splits into two pockets when the water level drops. When the water level drops, we assume that all species distribute themselves equally over the two pockets; similarly, when the water level rises, all species previously contained in the two pockets are combined into the single pocket. Allopatric speciation can only occur when the water level is low. We assume a constant probability rate for allopatric speciation, and hence the waiting time until the next speciation event is exponentially distributed. After this waiting time, one of the two incipient species in either pocket can speciate into a new species. If this allopatric speciation does not occur before the water level rises again, i.e. reflecting that there has not been enough genetic divergence, the two incipient species in the two pockets merge back into one species. This is conceptually similar to the idea of protracted speciation (Etienne and Rosindell 2012): the water level drop initiates the speciation process whereas the allopatric speciation event is the completion of speciation under the protracted speciation model. Sympatric speciation can always occur in our model, either at high water level in the lake, or in both pockets when the water level is low. Extinction is considered to be a background process that occurs locally, i.e. within a pocket. If the water level is high, this causes extinction of a species, if the water level is low, this causes local extinction in one of the pockets.

We implemented our model using a Gillespie algorithm, where the time steps are chosen depending on the rate of possible events. In the model there are five possible events:

- 1) A water level change event, inducing incipient species or merger of incipient species.
- 2) Sympatric speciation event at high water level, with rate  $\lambda_s^h$

- 3) Sympatric speciation event at low water level, with rate  $\lambda_s^l$
- 4) Allopatric speciation(-completion) event, at low water level, with rate  $\lambda_a^l$
- 5) Extinction event, with rate  $\mu$

When the water level drops, all species distribute themselves over both pockets. Thus, immediately after a water level drop, the number of incipient species is equal to twice the number of species. When the water level rises, all incipient species that belong to the same species merge into a single species. During a sympatric speciation event, a single species splits into two new species, and the original (incipient) species is consumed in the process. Here we assume that local disruptive selection causes divergence, similar to the implementation of speciation by Aguilee (Aguilée et al. 2011, 2013). If sympatric speciation occurs when the water level is low, the species in the other pocket is retained, and thus three new lineages arise: the first branching point occurs at the water level drop while the second occurs at the sympatric speciation event (Figure 1).

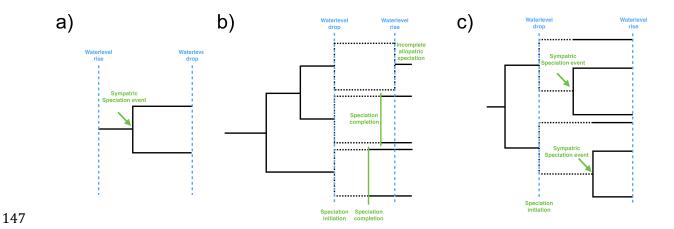


Figure 1. Schematic representation of the consequences of the three different types of speciation. Time proceeds from left to right. The dotted blue lines indicate water level changes. (a) During a sympatric speciation at high water level event, diversification is not aligned with any associated water level change. (b) During allopatric speciation at low water level, speciation initiation (incipient species are indicated with a dotted line) coincides with the water level drop, causing branching events (if speciation-completion occurs before water level rise) to line up in time. Branching events are conditionally independent of the time of speciation completion, hence, even when the actual speciation completion events occur at different time points, branching events in the species tree are identical. (c) During a sympatric

speciation event at low water, the speciation event is independent of the water level changes.

Because the original species is consumed in the process, a new branching event is also added

at the water level change event. Hence, both speciation-completion (b) and sympatric

speciation at low water level (c) cause branching times to line up at the time of water level

drop. Please note that (a) and (c) represent the reconstructed species tree, but (b) does not; the

reconstructed species tree for (b) would not show the branching event in the uppermost part

of the tree.

157

158

159

161

164

165

166

167

168

169

170

171

172

173

174

175

176

177

178

179

180

181

182

During an extinction event, one (possibly incipient) species is removed from the simulation.

If the water level is low, this need not lead to the extinction of a species, because the sister

incipient species might remain in the other pocket, ensuring survival of the species.

## Maximum Likelihood

Without water level changes, our model reduces to the constant rates birth-death model (Nee

et al. 1994). As a reference therefore, we estimated parameters of the standard birth-death

model using Maximum Likelihood. The likelihood of the birth-death model was calculated

using the function "bd ML" from the R package DDD. (Etienne et al. 2012).

## Fitting the model to empirical data

We performed two different fitting procedures: firstly, we performed a model selection procedure, where three different water level scenarios were fitted simultaneously to the data (more information about the chosen scenarios can be found in the next section). The model selection procedure simultaneously estimates parameter estimates and assesses the fit of the models. However, because the model selection procedure primarily samples the best fitting model (by design), it does not allow for the comparison of parameter estimates across different models. Therefore, we also fitted the three different water level scenarios independently to the empirical data, and obtained posterior distributions for the parameters relevant to these scenarios.

We fitted our model to 100 trees randomly sampled from the MCMC chain obtained from the

\*BEAST analysis (see below), and to the Most Credible Consensus (MCC) tree.

#### Water level scenarios

183

184

185

186

187

188

189

190

191

192

193

194

195

196

197

198

199

200

201

202

203

204

205

206

The main focus of our approach is to assess the impact of water level changes on the diversification rate. Lake Tanganyika experienced low water level stands 35 - 40 k years ago (kya) (-160 meter), 169 - 193 kya (-250 meter), 262 - 295 kya (-350 meter), 363 - 393 kya (-350 meter) and 550 - 1100 kya (-650 – 700 meter) (Lezzar et al. 1996; Cohen et al. 1997a). The southern and northern basin of Lake Tanganyika are separated from each other by a ridge at a depth of 500 meter below present level. Although some of these water level changes may not have split up the lake completely, we assume here that these water level changes still caused sufficient disruption of migration between the northern and southern basin, to be equivalent to physical separation. Consequently, high water levels occurred between 0-35kya, 40 - 169 kya, 193 - 262 kya, 295 - 363 kya and 393 - 550 kya. Unfortunately the geological record does not reveal whether any low water level stands occurred beyond 1.1 million years ago (Ma). This leaves us with two alternative scenarios: either no low water level stands occurred beyond 1.1 Ma, or these low water level stands have not been preserved accurately in the geological record. In order to capture these two scenarios we performed inference using two alternative water level implementations. Firstly we used the exact literature values, assuming a high water level stand until 1.1 Ma. We refer to this scenario as LW (Literature Waterlevels). Secondly we assumed that before 1.1. Ma, water level changes occurred at the same average rate of water level change in the most recent 1.1 million years. In the recent 1.1 million years, the lake experienced 5 high water level stands, and 5 low water level stands, which amounts to 10 water level changes in total. To extrapolate water level changes to more than 1.1 Ma, we

drew waiting times until the next water level change from an exponential distribution with rate 10. We refer to this scenario as EW (Extrapolated Water levels). Thirdly we also tested the null expectation: no effect of water level changes on speciation, we refer to this scenario as NW (No Water levels). Without water level changes, the model reduces to the constant-rates birth death model.

## Parameter estimation

207

208

209

210

211

212

213

214

215

216

217

218

219

220

221

222

223

224

225

226

227

228

229

230

To fit the model to empirical data we used Approximate Bayesian Computation, in combination with a Sequential Monte Carlo scheme (ABC-SMC) (Toni et al. 2009). As summary statistics for the ABC analysis we chose the normalized Lineages Through Time statistic (Janzen et al. 2015), tree size, Phylogenetic Diversity (AvPD, Schweiger et al. 2008) and the  $\gamma$  statistic (Pybus and Harvey 2000). On all parameters  $(\lambda_s^h, \lambda_s^l, \lambda_a^l, \mu)$  we chose uniform priors U(-3, 2), on a <sup>10</sup>log scale, such that the eventual prior distribution spans (10<sup>-3</sup>, 10<sup>2</sup>). A <sup>10</sup>log scale was chosen to explore parameter space uniformly, and put extra emphasis on low values. The standard deviation of the normal distribution used to perturb the parameters was chosen to have a mean of 0, and a standard deviation of 0.05 (on the <sup>10</sup>log transformed parameter), and we updated one parameter each time (e.g. jumps were only made in one dimension, to avoid extremely low acceptance rates). The number of particles used per SMC step was 10,000, where a particle is a data structure containing the model choice and the parameter estimates. To assess the fit of the model to the data we calculated the Euclidian distance between the summary statistic of the simulated data and the empirical data. To ensure that the differences in summary statistics were on the same scale, we normalized the differences. Differences were normalized by dividing each difference by the standard deviation of that summary statistic of 1,000,000 trees simulated using parameter values sampled from the prior.

## Model selection

232

233

234

235

236

237

238

239

240

241

242

243

244

245

246

247

248

249

250

251

252

253

254

255

To identify which model best explains the data, we performed ABC model selection, as described in Toni et al. (2009; 2010). The main difference between standard ABC-SMC and ABC-SMC including model selection is that the latter adds one parameter, which keeps track of the model. As jumping kernel between models we assumed a 50% probability of staying at the same model, and a 25% probability of jumping to either other model. We assumed a uniform prior across all three models; this translates to a probability of 1/3 for each model in the first iteration of the ABC-SMC procedure, and hence an expected number of 3333 particles assigned to each model in the first iteration. This reversible jump ABC-SMC model selection procedure results in a posterior distribution over the three models, where the model with most support is the model selected most across all particles. We can calculate the Bayes factor by taking the ratio of the number of particles assigned to the respective models (Toni et al. 2009). For example, the Bayes factor of LW/EW is the number of particles assigned to the model with literature water level changes divided by the number of particles assigned to the model with extrapolated water level changes. Because a model can receive zero particles, we set the Bayes factor for each model compared to the model with zero particles to the maximum support possible, which is the total number of particles: 10,000. To calculate the posterior support for a model, we calculate 2 ln (Bayes factor), following Kass and Raftery (1995). A transformed Bayes factor over a value of 2 then corresponds to substantial support for the considered model (Kass and Raftery 1995).

## Model selection validation

To assess whether our ABC-SMC method can accurately infer the correct model, we simulated 100 datasets for each model (NW, LW & EW), with parameter values drawn from the prior. We report the median Bayes factor across the 100 replicates. If our method can

accurately infer the correct model, we expect the median Bayes Factor (after 2 ln transformation) to be above 2 when comparing posterior support for the model with which the data was simulated to the other two models.

## Measurement uncertainty

256

257

258

259

260

261

262

263

264

265

266

267

268

269

270

271

272

273

274

275

276

277

A phylogeny generated with a high rate of allopatric speciation and a high rate of water level changes tends to have multiple speciation events that are aligned in time (Figure 1, b). This is due to the fact that the onset of speciation is given by the time of water level change. Phylogenetic reconstruction methods such as BEAST (Bouckaert et al. 2014) currently do not allow for simultaneous branching events. Hence, when fitting the model, trees are generated that are by definition dissimilar from the empirical tree constructed using BEAST, even if underlying events are close to the original events. To circumvent this we perturbed the branching time of each node in the trees simulated using our model. In this way speciation events that were previously aligned in time now occur on slightly different time points, as in a tree from a BEAST analysis. We perturbed branching times by adding a random number drawn from a truncated normal distribution with mean 0, standard deviation  $\sigma$ , truncated by the minimum distance to either the daughter or the parent species. If there were no daughter lineages present, and the node gave rise to an extant species, the normal distribution was truncated to the minimum distance to the parent or the present time. Nodes were perturbed from past to present (leaving the crown in place, to ensure a phylogenetic tree with the same age as the empirical tree). The standard deviation of the perturbation kernel was included as an extra parameter to be inferred, with a uniform prior on  $(10^{-3}, 10^{0})$ .

## Empirical data

279

280

281

282

283

284

285

286

287

288

289

290

291

292

293

294

295

296

297

298

299

300

301

302

303

We fitted our model to the phylogenetic tree of the tribe of *Lamprologini*, the most diverse tribe within Lake Tanganyika, containing 79 species of cichlids in Lake Tanganyika (Day et al. 2007; Koblmüller et al. 2007; Sturmbauer et al. 2010). The Lamprologini are endemic to Lake Tanganyika and its surrounding rivers and all species are substrate brooders with shared paternal and maternal care. In contrast to the mouthbrooding species from the Haplochromini, the Lamprologini show little sexual dimorphism and dichromatism, which are well-known indicators for sexual selection (Kraaijeveld et al. 2011). We therefore expect that the *Lamprologini* is a good candidate for picking up signals from water level changes. We reconstructed a new Lamprologini tree following the workflow of the most complete Lamprologini tree to date, which is a consensus tree based on the mitochondrial ND2 gene (Sturmbauer et al. 2010), but we added three newly described species (Lepidiolamprologus mimicus (Schelly et al. 2007), Neolamprologus timidus (Kullander et al. 2014b) and Chalinochromis cyanophleps (Kullander et al. 2014a)). Using phyloGenerator (Pearse and Purvis 2013), we downloaded sequences from GenBank for nine genes (GenBank access numbers can be found in the Supplementary Information). Genes were selected on the basis of species coverage (at least 25% of the 79 Lamprologini species for which molecular data is available), and whether or not the gene was crucial for inclusion of a species (e.g. for a number of species, the only available gene was ND2). After selection, our full dataset consisted of three mitochondrial genes: the NADH dehydrogenase subunit 2 (ND2 gene, sequences from Kocher et al. 1995; Klett and Meyer 2002; Clabaut et al. 2005; Duftner et al. 2005; Schelly et al. 2006; Day et al. 2007; Koblmüller et al. 2007, 2016; Schwarzer et al. 2009; Wagner et al. 2009; Sturmbauer et al. 2010; O'Quin et al. 2010; Kullander et al. 2014b; Weiss et al. 2015). The cytochrome b (cytb) gene (sequences from Salzburger et al. 2002; Nevado et al. 2009; Wagner et al. 2009; O'Quin et al. 2010; Matschiner et al. 2011, 2016;

305

306

307

308

309

310

311

312

313

314

315

316

317

318

319

320

321

322

323

324

325

326

327

328

Kullander et al. 2014b; Shirai et al. 2014) and the cytochrome c oxidase subunit I (COI gene, sequences from Sparks and Smith 2004; Nevado et al. 2013; Kullander et al. 2014a, 2014b; Breman et al. 2016; Matschiner et al. 2016) and six nuclear genes: the nuclear locus 38A (38A, sequences from Muschick et al. 2012; Meyer et al. 2016), the 18S ribosomal RNA internal-transcribed spacer 1-2 with 5.8S and 28S ribosomal RNA partial sequences (18S, sequences from Nevado et al. 2009; Koblmüller et al. 2016), the recombinase activating protein 1 (rag1, sequences from Clabaut et al. 2005; Nevado et al. 2009; Kullander et al. 2014b; Shirai et al. 2014; Koblmüller et al. 2016; Meyer et al. 2016), the endothelin receptor B1 gene (ednrb1, sequences from Muschick et al. 2012; Santos et al. 2014), the ribosomal protein S7 (rps7, sequences from Schelly et al. 2006; Meyer et al. 2016)) gene and the rod opsin gene (RH1, sequences from Sugawara et al. 2002; Spady et al. 2005; Nagai et al. 2011; Meyer et al. 2015). GenBank access numbers for the used sequences can be found in the supplementary material. Sequences were aligned using MAFFT (setting: --auto) (Katoh and Standley 2013), and subsequently, sequences were cleaned using trimAI (sites with >80% data missing were removed, e.g. setting -gt 0.2) (Capella-Gutiérrez et al. 2009). Rather than concatenating the alignments, we partitioned the data into subsets with independent sequence evolution models. which is more suitable for a dataset which is expected to show incomplete lineage sorting or hybridization (Meyer et al. 2016). To prepare alignments for use with partitionFinder, alignments were combined using SequenceMatrix 1.8 (Vaidya et al. 2011). The best partitioning found by partitionFinder 2.1.1 (Lanfear et al. 2012, 2016), partitioned the data into 5 subsets (unlinked branches, AICc selection criterion), with all nuclear genes into one subset (Rps7, ednrb1, 38A, 18S, RAG1 and RH1), with substitution model HKY+I+Γ. The remaining three mitochondrial genes (ND2, COI and cytb) were placed in separate subsets, each with a GTR+I+  $\Gamma$  substitution model.

Using \*BEAST (Heled and Drummond 2010) within the BEAST 2 package (Bouckaert et al. 2014), we inferred the time-calibrated species tree. We used an uncorrelated log-normal relaxed clock and applied two calibration points. Firstly, we calibrated the crown of the Lamprologini to be 4 million years old (log-normal prior, mean of 4 Myr 95% conf interval: [3, 5]), based on the results from Meyer et al. (2016). Secondly, we included two riverine Lamprologini species (L. congoensis and L. teugelsi), and calibrated the onset of their branching event at 1.7 Ma (offset 1.1, log normal distribution with mean 1.7, 95% conf interval [1.15, 3.47], "use originate = true"), following Koblmuller (2010). We applied 1/Xpriors on the clock rates, and log-normal priors on the substitution rates. All other priors were left at their default setting. As tree model we used the birth death model. The used BEAST configuration file (the Beauti xml) can be found in the supplementary material. We ran 10 independent STARBEAST MCMC chains, of 750M trees each. Each chain was verified to have ESS values of at least 100 for all parameters. The first 10M trees were pruned from these chains as burn-in and then they were combined (we used the species tree, rather than the individual gene trees) into one large chain (of 7400M trees). Chains were thinned by taking only each 5,000<sup>th</sup> tree. Using TreeAnnotator (from the BEAST 2 suite) we constructed a Maximum Clade Credibility tree (using all 1.48M trees after thinning), storing the mean heights. We then pruned the tree from riverine species to obtain the pure *Lamprologini* tree on which we fitted our model. Instead of performing one ABC-SMC inference on the obtained MCC tree using a huge number of particles, which would be more accurate but computationally extremely demanding, we performed 100 parallel inferences using 10,000 particles each. We report the mean Bayes factor across these replicates.

329

330

331

332

333

334

335

336

337

338

339

340

341

342

343

344

345

346

347

348

349

350

# Branching time uncertainty in the empirical tree

To account for uncertainty in the estimates of branching times in the *Lamprologini* tree we sampled 100 trees from the posterior distribution obtained by \*BEAST. Sampling was performed at random, irrespective of the likelihood of the trees. In the Supplementary material we show that the distribution of summary statistics of the subset of 100 trees is similar to the distribution of the thinned chain. The 100 sampled trees were, like the Maximum Clade Credibility tree, also pruned to remove the riverine taxa and stored separately. For all 100 trees we performed both the ABC-SMC model selection algorithm and the ABC-SMC parameter estimation algorithm, to determine the impact of different branching times on the inferred water level model and associated parameters, and to determine whether the MCC tree is a good representation of the underlying variability.

## RESULTS

364

365

366

367

368

369

370

371

372

373

374

375

376

377

378

379

380

381

382

383

384

385

386

387

388

# Lamprologini phylogeny

The onset of diversification within the *Lamprologini* is estimated to be around 3.96 Ma (95%) Highest Posterior Density interval (HPD): [3.09, 4.91]), which is very close to the prior we put on the node age, based on previous estimates (Meyer et al. 2016). Furthermore, we estimate the branching off of the Congo species (N. congoensis and N. teugelsi) from the Lamprologini in Lake Tanganyika to have occurred around 1.35 Ma (HPD: [1.12, 1.68]). which is a bit younger than previously obtained estimates (1.70 Ma, (Sturmbauer et al. 2010)). The topology of the Maximum Clade Credibility tree is largely consistent with previous findings (Sturmbauer et al. 2010) (Figure 1). Placement of Neolamprologus fasciatus as a close relative to N. wauthioni seems to re-iterate previously published evidence for introgressive hybridization (Koblmüller et al. 2007). For the three species not previously included in the Lamprologini phylogeny, Lepidiolamprologus mimicus was placed as a close relative to the other species within the genus Lepidiolamprologus, Chalinochromis cyanophleps was placed as a sister species to Chalinochromis brichardi, within the group of Chalinochromis and Julidochromis species, and in agreement with previous analysis (Kullander et al. 2014b). In contrast to previous findings (Kullander et al. 2014b), Neolamprologus timidus is not placed as a sister species to Neolamprologus furcifer, but rather associates with N. mondabu and N. falcicula. Again, in contrast to other previous findings (Gante et al. 2016), we place N. olivaceous outside the brichardi complex, which includes the model system species N. brichardi and N. pulcher. (but see the DensiTree representation, which shows that this is not true for all trees). Interestingly, we also do not infer N. savoryi to be phylogenetically clustered within the brichardi complex (the 'Princess cichlids' (Gante et al. 2016)), in contrast to Gante et al. (2016). We should take into account

390

391

392

393

394

395

396

397

398

399

400

401

402

403

404

405

406

407

408

bit different.

however that the analysis by Gante et al. is based on on full genome sequences from only a small group of species, in contrast to the limited number of markers from a large number of species that we used. As a reference we inferred speciation and extinction using the constant-rates birth-death model (Nee et al. 1994). Using Maximum Likelihood (the function bd ML in the DDD package (Etienne et al. 2012)), we obtained an estimate of 1.871 myr<sup>-1</sup> for the speciation rate, and an estimate of 0.993 myr<sup>-1</sup> for the extinction rate, for the MCC tree. We find an estimate of 0.87 mvr<sup>-1</sup> for the diversification rate (speciation - extinction) and an estimate of 0.531 myr<sup>-1</sup> for the turnover rate (extinction / speciation). For the 100 trees sampled from the full chain, we obtain estimates of 3.02 myr<sup>-1</sup> (95% HPD: [1.608, 4.947]) for the speciation rate and 2.409 myr<sup>-1</sup> (95% HPD: [0.884, 4.530]) for the extinction rate. This translates into estimates of 0.61 myr<sup>-1</sup> (95% HPD: [0.313, 0.985]) for the diversification rate, and 0.765 myr<sup>-1</sup> <sup>1</sup> (95% HPD: [0.518, 0.930]) for the turnover rate. Estimates for the birth-death model obtained during reconstruction of the tree using BEAST indicate an estimate of 0.864 myr<sup>-1</sup> (95% HPD: [0.287, 1.459]) for the speciation rate and an estimate of 0.613 myr<sup>-1</sup> (95% HPD: [0.181, 0.953]) for the relative death rate, which translates into an estimate for the extinction rate of 0.52 myr<sup>-1</sup> (95% HPD: [0.156, 0.823]) per million years. This yields estimates of 0.334 myr<sup>-1</sup> and 0.613 myr<sup>-1</sup> for the diversification and turnover rate respectively. The BEAST inferences include the riverine species, so speciation and extinction rates are expected to be a

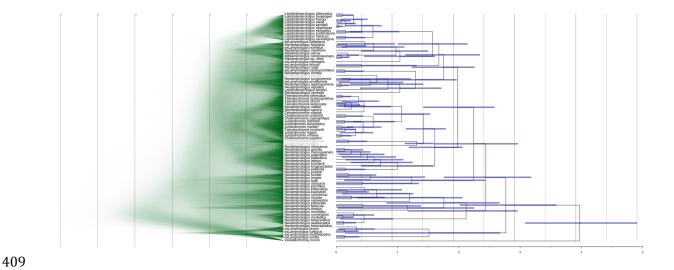


Figure 2. Phylogenetic hypothesis for the *Lamprologini* and outgroups, based on 3 mitochondrial and 6 nuclear genes, and two calibrations: 4 million years for the root of the *Lamprologini* clade, and 1.1 - 3.5 million years for the Congo *Lamprologini* species. Left panel: DensiTree (Bouckaert and Heled 2014) representation of the MCMC chain obtained using \*BEAST. Shown are trees from a thinned posterior chain, after selecting every 100,000<sup>th</sup> tree. Riverine species are indicated in grey. Right panel: Maximum Clade Credibility tree. Bars around the node span the 95% HPD for each node. Please note that for the dual display of both the densitree representation and the MCC phylogeny, some tips of the MCC phylogeny might appear slightly misaligned. A high resolution version of both the Densitree representation and the MCC phylogeny can be found in the supplementary information.

## **Parameter estimation**

We estimated parameter values for the three models for all 100 trees sampled from the posterior. We report the parameter values across the combined posterior across all 100 trees. Note that variation in the parameter estimates results from two sources of variation: branching time variation across the 100 trees, and secondly variation in the parameter estimate within each ABC-SMC inference.

The model without water level changes is identical to the constant-rates birth-death model, and we find that our ABC-SMC estimates for sympatric speciation at high water level ( $\lambda_s^h$ ) are slightly lower than the Maximum Likelihood estimate of the birth rate under the constant rates birth-death model (2.644 Myr<sup>-1</sup>(95% HPD: [1.208, 4.633], see also Table 1) versus 3.02,

433

434

435

436

437

438

439

440

441

442

443

444

445

446

447

448

449

450

451

452

453

454

455

see also Table 1). Similarly, we infer the extinction rate ( $\mu$ ) to also be slightly lower (1.950) Myr<sup>-1</sup> (95% HPD: [0.188, 4.101]) versus 2.409, see also Table 1). We obtain estimates of 0.694 and 0.738 for diversification and turnover respectively, which are close to the estimates obtained using Maximum Likelihood (a diversification rate of 0.610 and a turnover rate of 0.765 respectively). Taking into account the 95% confidence intervals on the obtained parameter estimates and the fact that the ABC-SMC estimates are potentially affected by the prior while the ML estimates are not, we are confident that estimates obtained using our ABC-SMC method for the model without water level changes are consistent with the maximum likelihood estimates under the constant-rates birth-death model. Using the LW model, which implements water level changes following the literature (e.g. high water level until ~1.1 Ma, after which a series of water level changes took place), we infer a lower rate of sympatric speciation at high water level (0.871 Myr<sup>-1</sup> (95% HPD: [0.227, 3.642])), which is compensated with a high rate of allopatric speciation (6.412 Myr<sup>-1</sup> (95%) HPD: [0.001, 14.195])) but not with a high rate of sympatric speciation at low water level (0.028 Myr<sup>-1</sup> (95% HPD: [0.001, 0.651])), suggesting that water level dynamics are important drivers of biodiversity, but only through allopatric speciation. Extinction is inferred to be low (0.037 Myr<sup>-1</sup> (95% HPD: [0.001, 2.133])). Because of the non-trivial relationship between speciation at high and low water level, we can no longer calculate diversification and turnover rates. Using the EW model, where water level changes are extrapolated beyond 1.1 Ma, we observe that the rate of sympatric speciation at high water level is inferred to be similar to without water level changes (2.753 Myr<sup>-1</sup> (95% HPD: [1.347, 4.383])). Extinction, however, is lower (0.111 Myr<sup>-1</sup> (95% HPD: [0.001, 1.627])), and allopatric speciation and sympatric speciation at low water level are both inferred to be much lower than for the literature water scenario

(0.022 Myr<sup>-1</sup> (95% HPD: [0.001, 0.466]) and 0.033 Myr<sup>-1</sup> (95% HPD: [0.001, 0.504]) 456 respectively). 457 Across the three water level models we observe that the distribution of the post-hoc 458 perturbations  $\sigma$  does not differ substantially from the prior for the NW and EW water models, 459 with low estimates (0.036 (95% HPD: [0.001, 0.569]) and 0.030 (95% HPD: [0.001, 0.484]) 460 for the NW and EW model respectively, Table 1). We notice a much higher value of  $\sigma$ 461 462 associated with LW (0.174, (95% HPD: [0.001, 0.680])), which also has a much higher estimate for allopatric speciation at low water level. Allopatric speciation at low water level 463 464 potentially causes temporal alignment of branching times and we introduced the parameter  $\sigma$ to correct simulated phylogenies for this, to allow comparison with phylogenies generated by 465 \*BEAST, which does not allow for temporally aligned branching times. Hence, the higher 466 inferred value of  $\sigma$  for the LW model confirms the validity of the application of our post-hoc 467 perturbation. 468

469

Table 1. Median posterior density estimate, for sympatric speciation at high water  $(\lambda_s^h)$ , extinction  $(\mu)$ , perturbation  $(\sigma)$ , sympatric speciation at low water  $(\lambda_s^l)$  and allopatric speciation  $(\lambda_a^l)$ . Shown are results for the model with no water level changes (NW), literature values for water level changes (LW) and water level changes extrapolated beyond the literature range (EW). The 95% credibility interval is shown between square brackets. All values are rates per million years.

	$\lambda_s^h$	μ	σ	$\lambda_a^l$	$\lambda_s^l$
NW	2.644 [1.208, 4.633]	1.950 [0.188, 4.101]	0.036 [0.001, 0.569]		
LW	0.871 [0.227, 3.642]	0.037 [0.001, 2.133]	0.174 [0.001, 0.68]	6.412 [0.001, 14.195]	0.028 [0.001, 0.651]
EW	2.753 [1.347 4.383]	0 111 [0 001 1 627]	0 030 [0 001 0 484]	0 022 [0 001 0 466]	0.033 [0.001_0.504]

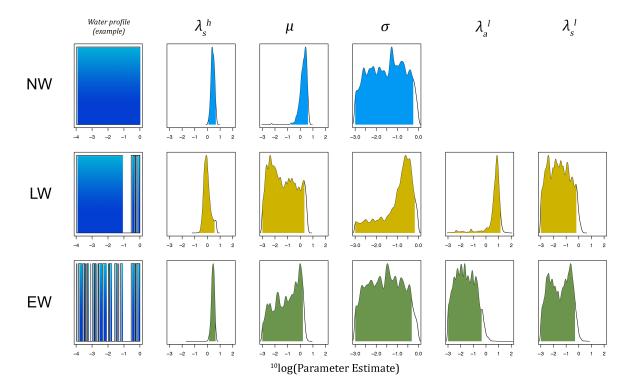
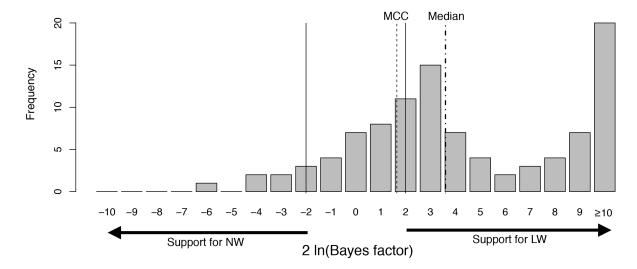


Figure 3. Posterior densities of the pooled posterior distribution across 100 randomly drawn trees from the posterior MCMC chain. Shown are estimates for the three water level scenarios (no water level changes (NW), literature values for water level changes (LW) and extrapolated values for water level changes (EW)). Shown are the posterior density (black line) and the 95% credibility interval (shaded area, blue for NW, gold for LW and green for EW. X-axes are on a  $^{10}$ log scale. The first column shows a sample water level profile, with the water level on the y-axis, and the time before present (in million years) on the x-axis. Note that for the EW model, for each simulation a new profile was generated, and that the shown profile is only one example of such a profile. Because without water level changes,  $\lambda_a^l$  and  $\lambda_s^l$  have no meaning, their posterior distribution is not shown for the NW scenario.

## Model fitting



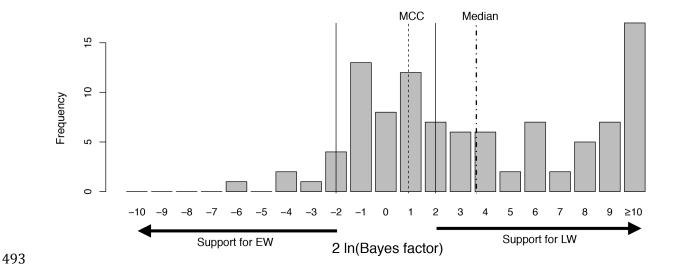


Figure 4. Model selection results on 100 trees randomly drawn from the \*BEAST posterior of the Lamprologini tree. The top row shows 2 ln (Bayes factors) comparing posterior support of the LW (literature water changes) model with the NW (no water level changes) model, the bottom row shows 2 ln (Bayes factors) of the comparison between the posterior support for the LW model with the EW (extrapolated water level changes) model. A 2ln(Bayes factor) higher than 2 is generally considered to provide substantial evidence in favor of the respective model (Kass and Raftery 1995), which is indicated by the solid lines. The thin dotted line indicates the median 2 ln (Bayes factor) obtained for the MCC tree, for which we do not find substantial support for any of the three models. The thick dotted line indicates the median 2 ln (Bayes factor) for the trees drawn from the \*BEAST posterior (e.g. the median of the distribution shown), which is in both cases above 2, indicating substantial support for the LW model compared to the other two models. 2 ln(Bayes factors) higher than 10 are grouped together into one category.

Model selection

When we apply the model selection algorithm to the MCC tree, we find median Bayes factors (we report here not the raw numbers, but 2 ln(Bayes factor), but for brevity refer to them as Bayes factors) of 1.64 and 0.9 when comparing the LW model with the NW and EW model respectively. We thus find no convincing evidence for any of the three models, when fitting our model to the MCC tree. Alternatively, when we fit to 100 trees randomly sampled from the \*BEAST posterior, we find Bayes factors of 3.60 and 3.65 when comparing LW model with the NW and EW model respectively. Furthermore, in 77 out of the 100 trees we select the LW model as the most likely model (based on the Bayes factor), in 17 out of 100 trees we select the EW model, and only in 6 out of 100 trees we select the model without any water level changes.

## Validation of the model selection procedure

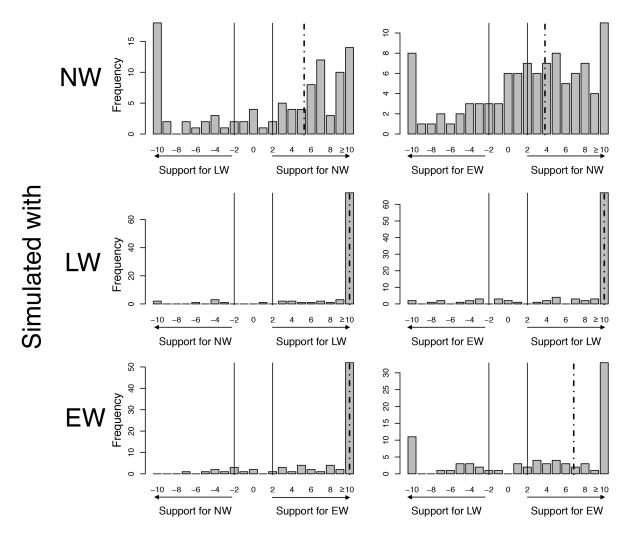


Figure 5. Validation of the ability of our ABC-SMC algorithm to infer the correct model. 100 replicate datasets were generated for each water level model (no water level changes NW, water level changes from the literature LW, or water level changes extrapolated beyond the literature range, EW). The plots show the distribution of the 2 ln(Bayes factor) across all 100 replicate inferences. The dotted line indicates the median 2 ln(Bayes factor). A 2ln(Bayes factor) higher than 2 is generally considered to provide substantial evidence in favor of the respective model (Kass and Raftery 1995). 2 ln(Bayes factors) higher than 10 are grouped together into one category.

Model validation shows that when we simulated data using the NW model, the NW model was selected using our model validation algorithm more than the other two models (59 out of 100 replicates). Median Bayes factors are both higher than 2, with a median of 5.28 and 3.83 versus the LW and EW model respectively, supporting considerable support for the NW

model over the other two models. When data was simulated with the LW model, we selected the correct model in the majority of 100 replicates (84 out of 100 replicates). The Bayes factors reflect this, with medians of 18.4 (this is the maximum score) versus both the NW and EW model. Lastly, when we simulated data using the EW model, we selected the correct model more than the other two models, in 65 out of 100 replicates. This was reflected by the Bayes factors as well, as the median Bayes factor versus the NW model was 18.4, and the median Bayes factor versus the LW model was 7.47. More interesting is the correct detection rate of a model, which is given by the number of trees simulated by the model that is selected for that tree. This is equal to asking whether, given posterior support for a respective model, we also find that the tree for which we find this support was simulated with the respective model. If our model selection procedure can not detect models accurately, we expect a detection rate of around 50%, as support is always divided between two (not three) models. Detection rates larger than 50% support the conclusion that our model selection procedure can adequately infer the correct model. We find that across the 300 simulated trees, 120 trees received considerable support for the LW model over the NW model (e.g. 2 ln (BF LW/NW) > 2), of these 120 trees, 92 trees were simulated with the LW model, which leads to a correct detection rate of 77% (See Figure 6). Furthermore, out of 107 trees that received considerable support for the LW model over the EW model, we find that 83 trees were simulated using the LW model, which translates to a detection rate of 78%. We find similar detection rates for the NW model: 90% against the LW model (62 out of 69 detected trees) and 83% against the EW model (54 out of 65 detected trees). Lastly, detection rates for the EW model mirror these findings: a detection rate of 79% against the NW model (79 out of 100 detected trees), and of 86% against the LW model (68 out of 79 detected trees).

538

539

540

541

542

543

544

545

546

547

548

549

550

551

552

553

554

555

556

557

558

559

560

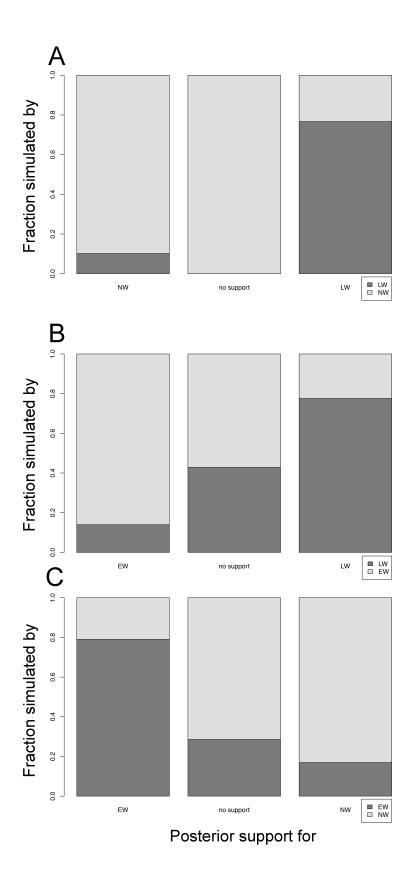


Figure 6. Accuracy of assignment of models depending on their posterior support. A: The relative fraction of trees simulated with either LW (dark) or NW (light), receiving support for LW  $(2\ln(\text{Bayes Factor LW/NW}) > 2)$ , support for NW  $(2\ln(\text{Bayes Factor LW/NW}) < -2)$ , or receiving no support for either model. **B**: The relative fraction of trees simulated with either

LW (dark) or EW (light), receiving support for LW (2ln(Bayes Factor LW/EW) > 2), support for EW (2ln(Bayes Factor LW/EW) < -2), or receiving no support for either model. C: The relative fraction of trees simulated with either NW (dark) or EW (light), receiving support for NW (2ln(Bayes Factor NW/EW) > 2), support for EW (2ln(Bayes Factor NW/EW) < -2), or receiving no support for either model.

## DISCUSSION

574

575

576

577

578

579

580

581

582

583

584

585

586

587

588

589

590

591

592

593

594

595

596

597

We have presented a model that infers past speciation and extinction rates, and their interactions with changes in the environment, from a given phylogeny. We have shown that our model is able to accurately select between different scenarios, including or excluding environmental change. We applied our model to an updated phylogeny of the cichlid fish tribe of Lamprologini and found evidence that past water level changes have shaped current cichlid diversity in Lake Tanganyika, when we applied our model to a sample from the posterior distribution of trees of the Lamprologini, as inferred by \*BEAST. We asked the model to select the best fitting of three scenarios: a scenario without any water level changes, a scenario using the values found in the literature, and a scenario using the mean rate of water level change found in the literature to extrapolate water level changes beyond the range of literature values available. We found that the model following literature water levels received most support, which suggests that water level changes have been an important driver of diversity in the Lamprologini. We note that a model without effect of water level changes on diversification (NW) can sometimes generate patterns that resemble the predictions of the preferred model (LW). Yet, we find when fitting our model to trees drawn from the \*BEAST posterior that the distribution of Bayes Factors is skewed towards the model following literature water levels (LW) and we find support for the model without an effect of water level changes on diversification (NW) only for a small number of trees, suggesting that this effect is relatively small.

When we applied our model selection algorithm on the Most Credible Consensus (MCC) tree, we found contrasting results. Support for both models including water level changes diminished, and posterior support for the model without any water level changes increased.

Nevertheless, no single model could yield enough support to convincingly reject the other two. Moreover, results using the MCC tree are markedly different from those using trees sampled from the posterior. We conclude therefore that the MCC tree, at least for the *Lamprologini*, but most likely more generally, provides a poor summary of the true species tree and of the underlying variation in branching patterns. Hence, we suggest to avoid reporting MCC trees, and instead to provide the reader with the full posterior distribution, for instance through a DensiTree plot (Bouckaert and Heled 2014). Posterior inference, for instance of speciation and extinction rates should preferentially also be performed on multiple independent samples from the posterior, rather than on the MCC tree, as the underlying variation might lead to very different results, as we have shown here.

Discrepancies between the MCC tree and the posterior distribution of trees could also potentially clarify previously recovered inconsistencies when studying diversification, for example in shrews in the Philippines. The Philippines have been subject to strong sea level fluctuations, causing the fission and fusion of several islands, primarily during the Pleistocene (Brown et al. 2013). Population genetic evidence has convincingly shown that the location of such fused islands correlates strongly with genetic divergence between populations in many different species (Evans et al. 2003; Linkem et al. 2010; Siler et al. 2010; Oaks et al. 2013). Phylogenetic analysis however, has failed to show any evidence of diversification associated with Pleistocene water level changes (Esselstyn and Brown 2009). The basis for this phylogenetic analysis however, was an MCC tree. Repeating the analysis on the posterior distribution underlying the MCC tree could mitigate these problems, and could clarify the impact of Pleistocene water level changes on diversification in the Philippines archipelago.

623

624

625

626

627

628

629

630

631

632

633

634

635

636

637

638

639

640

641

642

643

644

645

When allopatric speciation rates are high, the resulting phylogenetic trees have internal nodes that have synchronized branching times, e.g. branching times that align with episodes of water level change. Although Phylogenetic reconstruction software is able to infer simultaneous branching events, it typically uses only two parameters (birth and death) to infer all branching events of the tree. Therefore, if it can accommodate the simultaneous events, it is unlikely to fit well to the non-simultaneous events, and vice-versa. Our finding of evidence for a substantial role of habitat dynamics in diversification can therefore be regarded as conservative. To improve the fit of trees generated by our model with trees generated by \*BEAST we included an a posteriori perturbation parameter in our model. This parameter determines the standard deviation of a Gaussian perturbation kernel that is applied to each node after the simulation has completed. By perturbing each node, we minimized the probability that branching times align in time. We found that standard deviation increased in size with an increase in allopatric speciation, as expected. A less ad hoc solution to deal with the alignment of branching times in the tree would be to incorporate the model presented here as a tree prior in phylogenetic reconstruction software. Although this need not introduce any significant differences in the tree topology, the distribution of branching times could be substantially influenced, and any subsequent inference focusing on such patterns could be very different. Including such models in tree reconstruction software may require incorporation of ABC methods, and will be extremely computationally demanding, but our results justify such an endeavor.

Given that water level changes are only prevalent during the last million years before present, we cannot exclude the possibility that increased diversification due to reasons other than

647

648

649

650

651

652

653

654

655

656

657

658

659

660

661

662

663

664

665

666

667

668

669

670

changing water levels has driven diversification during this period. On average, the LW model could be represented by a simple birth-death model with a rate shift around one million years ago. We expect however that although such a model could accommodate the increased average diversification, it cannot replicate temporal alignment in branching events due to water level changes. To examine this in more detail, we fitted a simple birth-death model with a rate shift around one million years ago to the trees obtained from the posterior (see Supplementary Information).. In the absence of a likelihood for the LW model, we compared the nLTT statistic for the rate-shift model with that of the LW model, as the nLTT statistic should be sensitive to detecting temporal alignment of branching events, We find that our model is much closer to the empirical data than the rate shift model. We attempted to improve the fit of the rate-shift model by allowing the speciation rate in the model to shift up and down in line with the literature values of the water level changes. The two rates inferred by the model then represent speciation at low, and at high water level respectively. Although we do find an increase in the rate of speciation at low water level, the fit of this rate-shift model is still worse than that of the LW model. This supports our conclusion that water level changes influence the phylogeny not only through an increased speciation rate, but also through temporal alignment of branching times.

Although we refer in our model to the different implementations of speciation as sympatric and allopatric, care should be taken in interpreting these forms of speciation. We consider here allopatric speciation only on a large scale, where populations become allopatric over stretches of hundreds of kilometers (Sturmbauer et al. 2001). Large-scale isolation might not be necessary for cichlids, as some species can already be limited in gene flow by a sand stretch of 50 meters separating populations (Rico and Turner 2002). Such micro-allopatric speciation events are not captured by the allopatric speciation rate in our model. Rather, these

local scale events are captured in our model by sympatric speciation. Hence, sympatric speciation in our model covers all degrees of speciation ranging from full sympatry to allopatry, providing that geographical isolation is smaller than that imposed by a water level change. Allopatric speciation in our model then solely refers to speciation events caused by geographical isolation over a large distance, driven by changes in water level, and inducing simultaneous branching events.

In our model we have assumed that when the water level drops, species distribute themselves equally over the two pockets of water that survive the water level drop. A more realistic model would allow for a skew towards one of the pockets, either dependent on the respective sizes of the pockets, the distribution of the species over the lake at high water level, or both. We have here refrained from including a parameter that regulates the distribution of species over the two pockets in order to avoid over fitting. Another possible extension of our model would lie into extending the approach towards three or more pockets, possibly combined with a parameter governing the distribution of species across these three pockets during a water level drop. Bathymetric maps of Lake Tanganyika suggest that for some water level changes it might split into three lakes (Coulter 1991). How a split of a species into three populations, and associated allopatric divergence and speciation, affects phylogenetic structure and affects temporal alignment in branching times remains currently unexplored and would be an interesting avenue for future work.

Our results are strongly in line with population genomic analyses in a number of cichlid species including *Eretmodus cyanostictus* (Verheyen et al. 1996), *Tropheus moorii* (Koblmüller et al. 2011; Nevado et al. 2013; Sefc et al. 2017), *Variabilichromis moorii* 

(Nevado et al. 2013), *Altolamprologus* (Koblmüller et al. 2016) and *Telmatochromis* temporalis (Winkelmann et al. 2016), and resonate with population genomic findings across the three African Rift Lakes (Sturmbauer et al. 2001). Furthermore, population genetic studies have shown that water level fluctuations in Lake Malawi have been associated with population expansion in cichlid species (Arnegard et al. 1999; Sturmbauer et al. 2001; Genner et al. 2010), suggesting a potential role for water level changes in Lake Malawi as well. Phylogenetic reconstruction for Malawi cichlid species is problematic however, partially due to the young age of the species. However, considering that the geological record of Lake Malawi spans a much larger part of the total lifespan of the lake (Delvaux 1995; Lyons et al. 2015; Ivory et al. 2016) and thus provides a much better record about water level fluctuations since the colonization of the lake by cichlids, we expect that modern genetic developments will soon allow for a thorough understanding of the impact of water level changes on cichlids in Lake Malawi as well.

# Conclusion

Our model integrates standard constant-rate birth-death mechanics with environmental change and with speciation induced by geographical isolation. We analyzed the phylogeny of the tribe of *Lamprologini* to see whether past water level changes in Lake Tanganyika have contributed to the current diversity of cichlid fish in Lake Tanganyika. We find an important role for environmental changes in driving diversity, and find evidence that past water level changes have shaped current standing diversity in the tribe of *Lamprologini*. However, we found that inference of past environmental changes from a single phylogeny, and more specifically, from the MCC tree, tends to lead to unreliable results. We therefore advocate caution when using the MCC tree as a basis for further analysis. Furthermore, we argue for

the inclusion of more detailed branching models in phylogenetic reconstruction software, which allow for the inclusion of an interaction between the environment, and speciation rates.

Acknowledgements

We thank Lucas Molleman for useful discussions. We thank the Netherlands Organisation for Scientific Research (NWO) for financial support through VIDI and VICI grants awarded to RSE. We thank the Donald Smits Center for information Technology of the University of Groningen for their support and providing access to the Millipede and Peregrine high-performance computing cluster. We thank the Max Planck Institute for Evolutionary Theory for their support and providing access to their high-performance computing cluster. We thank the Carl von Ossietzky Universität Oldenburg for their support and providing access to the CARL computing cluster.

#### REFERENCES

- Aguilée R., Claessen D., Lambert A. 2013. Adaptive radiation driven by the interplay of ecoevolutionary and landscape dynamics. Evolution (N. Y). 67:1291–1306.
- Aguilée R., Lambert A., Claessen D. 2011. Ecological speciation in dynamic landscapes. J.
   Evol. Biol. 24:2663–77.
- Alin S., Cohen A., Bills R. 1999. Effects of landscape disturbance on animal communities in Lake Tanganyika, East Africa. Conservation. 13:1017–1033.
- Alin S., Cohen A.S. 2003. Lake-level history of Lake Tanganyika, East Africa, for the past
   2500 years based on ostracode-inferred water-depth reconstruction. Palaeogeogr.
- Palaeoclimatol. Palaeoecol. 199:31–49.
- Arnegard M.E., Markert J.A., Danley P.D., Stauffer J.R., Ambali A.J., Kocher T.D. 1999.
- Population structure and colour variation of the cichlid fishes Labeotropheus fuelleborni
- Ahl along a recently formed archipelago of rocky habitat patches in southern Lake
- 745 Malawi. Proc. R. Soc. Lond. B. 266:119–130.
- Barnosky A. 2005. Effects of Quaternary climatic change on speciation in mammals. J.
   Mamm. Evol. 12:247–264.
- Bouckaert R., Heled J. 2014. DensiTree 2: Seeing trees through the forest. bioRxiv.:1–11.
- Bouckaert R., Heled J., Kühnert D., Vaughan T., Wu C.H., Xie D., Suchard M.A., Rambaut
- A., Drummond A.J. 2014. BEAST 2: A Software Platform for Bayesian Evolutionary Analysis. PLoS Comput. Biol. 10:1–6.
- Brawand D., Wagner C.E., Li Y.I., Malinsky M., Keller I., Fan S., Simakov O., Ng A.Y.,
- Lim Z.W., Bezault E., Turner-Maier, J. Johnson J., Alcazar R., Noh H.J., Russell P.,
- Aken B., Alföldi J., Amemiya C., Azzouzi N., Baroiller J.-F., Barloy-Hubler F., Berlin
- A., Bloomquist R., Carleton K.L., Conte M.A., D'Cotta H., Eshel O., Gaffney L.,
- Galibert F., Gante H.F., Gnerre S., Greuter L., Guyon R., Haddad N.S., Haerty W.,
- Harris R.M., Hofmann H. a., Hourlier T., Hulata G., Jaffe D.B., Lara M., A.P. L.,
- MacCallum I., Mwaiko S., Nikaido M., Nishihara H., Ozouf-Costaz C., Penman D.J.,
- Przybylski D., Rakotomanga M., Renn S.C.P., Ribeiro F.J., Ron M., Salzburger W.,
- Sanchez-Pulido L., Santos M.E., Searle S., Sharpe T., Swofford R., Tan F.J., Williams
- L., Young S., Yin S., Okada N., Kocher T.D., Miska E. a., Lander E.S., Venkatesh B.,
- Fernald R.D., Meyer A., Ponting C.P., Streelman J.T., Lindblad-Toh K., Seehausen O.,
- Di Palma F. 2014. The genomic substrate for adaptive radiation in African cichlid fish.
- 764 Nature. 513:375–381.
- Breman F.C., Loix S., Jordaens K., Snoeks J., Van Steenberge M. 2016. Testing the potential
   of DNA barcoding in vertebrate radiations: the case of the littoral cichlids (Pisces,
- Perciformes, Cichlidae) from Lake Tanganyika. Mol. Ecol. Resour. 16:1455–1464.
- Brown R.M., Siler C.D., Oliveros C.H., Esselstyn J. a., Diesmos A.C., Hosner P. a., Linkem
- 769 C.W., Barley A.J., Oaks J.R., Sanguila M.B., Welton L.J., Blackburn D.C., Moyle R.G.,
- Townsend Peterson a., Alcala A.C. 2013. Evolutionary processes of diversification in a
- model island archipelago. Annu. Rev. Ecol. Evol. Syst. 44:411–435.

- 772 Capella-Gutiérrez S., Silla-Martínez J.M., Gabaldón T. 2009. trimAl: A tool for automated
- alignment trimming in large-scale phylogenetic analyses. Bioinformatics. 25:1972–
- 774 1973.
- Clabaut C., Salzburger W., Meyer A. 2005. Comparative phylogenetic analyses of the
- adaptive radiation of Lake Tanganyika cichlid fish: nuclear sequences are less
- homoplasious but also less informative than mitochondrial DNA. J. Mol. Evol. 61:666–
- 778 81.
- 779 Cohen A.S., Lezzar K.E., Tiercelin J.J., Soreghan M. 1997a. New palaeogeographic and lake-
- level reconstructions of Lake Tanganyika: implications for tectonic, climatic and
- biological evolution in a rift lake. Basin Res. 9:107–132.
- Cohen A.S., Soreghan M., Scholz C.A. 1993. Estimating the age of formation of lakes: an example from Lake Tanganyika, East African Rift system. Geology. 21:511.
- Cohen A.S., Stone J.R., Beuning K.R.M., Park L.E., Reinthal P.N., Dettman D., Scholz C.A.,
- Johnson T.C., King J.W., Talbot M.R., Brown E.T., Ivory S.J. 2007. Ecological
- consequences of early Late Pleistocene megadroughts in tropical Africa. Proc. Natl.
- 787 Acad. Sci. 104:16422–7.
- Cohen A.S., Talbot M.R., Awramik S.M., Dettman D.L., Abell P. 1997b. Lake level and
- paleoenvironmental history of Lake Tanganyika, Africa, as inferred from late Holocene
- and modern stromatolites. Geol. Soc. Am. Bull. 109:444–460.
- 791 Coulter G. 1991. Lake Tanganyika and its life. Oxford University Press.
- 792 Coyne J., Orr H. 2004. Speciation. Sunderland, Massachusets U.S.A.: Sinauer Associates.
- 793 Day J.J., Santini S., Garcia-Moreno J. 2007. Phylogenetic relationships of the Lake
- Tanganyika cichlid tribe Lamprologini: the story from mitochondrial DNA. Mol.
- 795 Phylogenet. Evol. 45:629–42.
- 796 Delvaux D. 1995. Age of Lake Malawi (Nyasa) and water level fluctuations. Mus. roy. Afrr.
- 797 centr., Tervuren (Belg.), Dept. Geol. Min., Rapp. ann. 1993 1994. 108:99–108.
- 798 Duftner N., Koblmüller S., Sturmbauer C. 2005. Evolutionary relationships of the
- Limnochromini, a tribe of benthic deepwater cichlid fish endemic to Lake Tanganyika.
- 800 East Africa. J. Mol. Evol. 60:277–289.
- 801 Esselstyn J.A., Brown R.M. 2009. The role of repeated sea-level fluctuations in the
- generation of shrew (Soricidae: Crocidura) diversity in the Philippine Archipelago. Mol.
- 803 Phylogenet. Evol. 53:171–181.
- 804 Etienne R.S., Haegeman B., Stadler T., Aze T., Pearson P.N., Purvis A., Phillimore A.B.
- 2012. Diversity-dependence brings molecular phylogenies closer to agreement with the
- 806 fossil record. Proc. R. Soc. B Biol. Sci. 279:1300–1309.
- Etienne R.S., Rosindell J. 2012. Prolonging the past counteracts the pull of the present:
- protracted speciation can explain observed slowdowns in diversification. Syst. Biol.
- 809 61:204–13.
- 810 Evans B.J., Brown R.M., McGuire J.A., Supriatna J., Andayani N., Diesmos A., Iskandar D.,
- Melnick D.J., Cannatella D.C. 2003. Phylogenetics of fanged frogs: testing

- biogeographical hypotheses at the interface of the asian and Australian faunal zones.
- 813 Syst. Biol. 52:794–819.
- Gante H.F., Matschiner M., Malmstrøm M., Jakobsen K.S., Jentoft S., Salzburger W. 2016.
- Genomics of speciation and introgression in Princess cichlid fishes from Lake
- 816 Tanganyika. Mol. Ecol.
- Genner M.J., Knight M.E., Haesler M.P., Turner G.F. 2010. Establishment and expansion of
- Lake Malawi rock fish populations after a dramatic Late Pleistocene lake level rise. Mol.
- 819 Ecol. 19:170–82.
- Genner M.J., Seehausen O., Lunt D.H., Joyce D.A., Shaw P.W., Carvalho G.R., Turner G.F.
- 821 2007. Age of cichlids: new dates for ancient lake fish radiations. Mol. Biol. Evol.
- 822 24:1269–82.
- Glor R.E., Gifford M.E., Larson A., Losos J.B., Schettino L.R., Chamizo Lara A.R., Jackman
- T.R. 2004. Partial island submergence and speciation in an adaptive radiation: a
- multilocus analysis of the Cuban green anoles. Proc. R. Soc. B Biol. Sci. 271:2257–65.
- Heaney L.R. 1985. Zoogeographic Evidence for Middle and Late Pleistocene Land Bridges to the Philippine Islands. Mod. Quat. Res. Southeast Asia. 9:127–143.
- Heled J., Drummond A.J. 2010. Bayesian Inference of Species Trees from Multilocus Data.
- 829 Mol. Biol. Evol. 27:570–580.
- Hutter C.R., Guayasamin J.M., Wiens J.J. 2013. Explaining Andean megadiversity: The
- evolutionary and ecological causes of glassfrog elevational richness patterns. Ecol. Lett.
- 832 16:1135–1144.
- 833 Ivory S.J., Blome M.W., King J.W., McGlue M.M., Cole J.E., Cohen A.S. 2016.
- 834 Environmental change explains cichlid adaptive radiation at Lake Malawi over the past
- 835 1.2 million years. Proc. Natl. Acad. Sci.:201611028.
- Janzen T., Höhna S., Etienne R.S.R.S. 2015. Approximate Bayesian Computation of
- diversification rates from molecular phylogenies: introducing a new efficient summary
- statistic, the nLTT. Methods Ecol. Evol. 6:566–575.
- Kass R., Raftery A. 1995. Bayes Factors. J. Amer. Stat. Assoc. 90:773–795.
- Katoh K., Standley D.M. 2013. MAFFT multiple sequence alignment software version 7:
- Improvements in performance and usability. Mol. Biol. Evol. 30:772–780.
- Klett V., Meyer A. 2002. What, if anything, is a Tilapia?-mitochondrial ND2 phylogeny of
- tilapiines and the evolution of parental care systems in the African cichlid fishes. Mol.
- Biol. Evol. 19:865–883.
- Koblmüller S., Duftner N., Sefc K.M., Aibara M., Stipacek M., Blanc M., Egger B.,
- Sturmbauer C. 2007. Reticulate phylogeny of gastropod-shell-breeding cichlids from
- Lake Tanganyika--the result of repeated introgressive hybridization. BMC Evol. Biol.
- 848 7:7.
- Koblmüller S., Nevado B., Makasa L., van Steenberge M., Vanhove M.P.M., Verheyen E.,
- Sturmbauer C., Sefc K.M. 2016. Phylogeny and phylogeography of Altolamprologus:
- ancient introgression and recent divergence in a rock-dwelling Lake Tanganyika cichlid

- genus. Hydrobiologia.:1–16.
- Koblmüller S., Salzburger W., Obermüller B., Eigner E., Sturmbauer C., Sefc K.M. 2011.
- Separated by sand, fused by dropping water: habitat barriers and fluctuating water levels
- steer the evolution of rock-dwelling cichlid populations in Lake Tanganyika. Mol. Ecol.
- 856 20:2272–90.
- Kocher T.D., Conroy J. a, McKaye K.R., Stauffer J.R., Lockwood S.F. 1995. Evolution of
- NADH dehydrogenase subunit 2 in east African cichlid fish. Mol. Phylogenet. Evol.
- 859 4:420–432.
- Kraaijeveld K., Kraaijeveld-Smit F.J.L., Maan M.E. 2011. Sexual selection and speciation:
- the comparative evidence revisited. Biol. Rev. Camb. Philos. Soc. 86:367–77.
- Kullander S.O., Karlsson M., Norén M. 2014a. Chalinochromis cyanophleps, a new species
- of cichlid fish (Teleostei: Cichlidae) from Lake Tanganyika. Zootaxa. 3790:425–438.
- Kullander S.O., Norén M., Karlsson M. 2014b. Description of Neolamprologus
- timidus, new species, and review of N. furcifer from Lake Tanganyika (Teleostei:
- Cichlidae). Ichthyol.Explor.Freshwaters. 24:301–328.
- Lanfear R., Calcott B., Ho S.Y.W., Guindon S. 2012. PartitionFinder: Combined selection of
- partitioning schemes and substitution models for phylogenetic analyses. Mol. Biol. Evol.
- 869 29:1695–1701.
- 870 Lanfear R., Frandsen P.B., Wright A.M., Senfeld T., Calcott B. 2016. PartitionFinder 2: New
- Methods for Selecting Partitioned Models of Evolution for Molecular and
- Morphological Phylogenetic Analyses. Mol. Biol. Evol.:msw260.
- 873 Lezzar K.E., Tiercelin J.J., Batist M., Cohen A.S., Bandora T., Rensbergen P., Turdu C.,
- Mifundu W., Klerkx J. 1996. New seismic stratigraphy and Late Tertiary history of the
- North Tanganyika Basin, East African Rift system, deduced from multichannel and
- high-resolution reflection seismic data and piston core evidence. Basin Res. 8:1–28.
- 877 Linkem C.W., Hesed K.M., Diesmos A.C., Brown R.M. 2010. Species boundaries and
- cryptic lineage diversity in a Philippine forest skink complex (Reptilia; Squamata;
- 879 Scincidae: Lygosominae). Mol. Phylogenet. Evol. 56:572–585.
- Lyons R.P., Scholz C.A., Cohen A.S., King J.W., Brown E.T., Ivory S.J., Johnson T.C.,
- Deino A.L., Reinthal P.N., Mcglue M.M., Blome M.W. 2015. Continuous 1.3-million-
- year record of East African hydroclimate, and implications for patterns of evolution and
- biodiversity. PNAS.:2–7.
- Matschiner M., Hanel R., Salzburger W. 2011. On the origin and trigger of the notothenioid
- adaptive radiation. PLoS One. 6.
- Matschiner M., Musilová Z., Barth J.M.I., Starostova Z., Salzburger W., Steel M., Bouckaert
- R. 2016. Bayesian Phylogenetic Estimation of Clade Ages Supports Trans-Atlantic
- Dispersal of Cichlid Fishes. Syst. Biol.
- McGlue M.M., Lezzar K.E., Cohen A.S., Russell J.M., Tiercelin J.-J., Felton A. a., Mbede E.,
- Nkotagu H.H. 2008. Seismic records of late Pleistocene aridity in Lake Tanganyika,
- tropical East Africa. J. Paleolimnol. 40:635–653.

- Meyer B.S., Matschiner M., Salzburger W. 2015. A tribal level phylogeny of Lake
- Tanganyika cichlid fishes based on a genomic multi-marker approach. Mol. Phylogenet.
- 894 Evol. 83:56–71.
- Meyer B.S., Matschiner M., Salzburger W. 2016. Disentangling incomplete lineage sorting
- and introgression to refine species-tree estimates for Lake Tanganyika cichlid fishes.
- 897 Syst. Biol. syw069:1–20.
- Muschick M., Indermaur A., Salzburger W. 2012. Convergent Evolution within an Adaptive
- Radiation of Cichlid Fishes. Curr. Biol.:1–7.
- 900 Nagai H., Terai Y., Sugawara T., Imai H., Nishihara H., Hori M., Okada N. 2011. Reverse
- evolution in RH1 for adaptation of cichlids to water depth in Lake Tanganyika. Mol.
- 902 Biol. Evol. 28:1769–1776.
- Nee S., May R.M., Harvey P.H. 1994. The reconstructed evolutionary process. Philos. Trans.
- 904 R. Soc. Lond. B. Biol. Sci. 344:305–11.
- Nevado B., KoblmÜller S., Sturmbauer C., Snoeks J., Usano-Alemany J., Verheyen E. 2009.
- Complete mitochondrial DNA replacement in a Lake Tanganyika cichlid fish. Mol.
- 907 Ecol. 18:4240–4255.
- Nevado B., Mautner S., Sturmbauer C., Verheyen E. 2013. Water-level fluctuations and
- metapopulation dynamics as drivers of genetic diversity in populations of three
- 910 Tanganyikan cichlid fish species. Mol. Ecol. 22:3933–48.
- 911 O'Quin K.E., Hofmann C.M., Hofmann H.A., Carleton K.L. 2010. Parallel Evolution of
- opsin gene expression in African cichlid fishes. Mol. Biol. Evol. 27:2839–2854.
- Oaks J.R., Sukumaran J., Esselstyn J. a, Linkem C.W., Siler C.D., Holder M.T., Brown R.M.
- 2013. Evidence for climate-driven diversification? A caution for interpreting ABC
- inferences of simultaneous historical events. Evolution (N. Y). 67:991–1010.
- Papadopoulou A., Knowles L.L. 2015. Genomic tests of the species-pump hypothesis: Recent
- 917 island connectivity cycles drive population divergence but not speciation in Caribbean
- 918 crickets across the Virgin Islands. Evolution (N. Y). 69:1501–1517.
- Pearse W.D., Purvis A. 2013. phyloGenerator: An automated phylogeny generation tool for
- ecologists. Methods Ecol. Evol. 4:692–698.
- 921 Pybus O., Harvey P. 2000. Testing macro–evolutionary models using incomplete molecular
- 922 phylogenies. Proc. R. Soc. B Biol. Sci. 267:2267–72.
- 923 Rico C., Turner G.F. 2002. Extreme microallopatric divergence in a cichlid species from
- 924 Lake Malawi. Mol. Ecol. 11:1585–90.
- Rossiter A. 1995. The cichlid fish assemblages of Lake Tanganyika: ecology, behaviour and
- evolution of its species flocks. Adv. Ecol. Res.
- 927 Salzburger W., Baric S., Sturmbauer C. 2002. Speciation via introgressive hybridization in
- 928 East African cichlids? Mol. Ecol. 11:619–25.
- 929 Santos M.E., Braasch I., Boileau N., Meyer B.S., Sauteur L., Böhne A., Belting H.-G.,
- Affolter M., Salzburger W. 2014. The evolution of cichlid fish egg-spots is linked with a

- 931 cis-regulatory change. Nat. Commun. 5:5149.
- 932 Schelly R., Salzburger W., Koblmüller S., Duftner N., Sturmbauer C. 2006. Phylogenetic
- relationships of the lamprologine cichlid genus Lepidiolamprologus (Teleostei:
- Perciformes) based on mitochondrial and nuclear sequences, suggesting introgressive
- hybridization. Mol. Phylogenet. Evol. 38:426–438.
- 936 Schelly R., Takahashi T., Bills R., Hori M. 2007. The first case of aggressive mimicry among
- lamprologines in a new species of Lepidiolamprologus (Perciformes: Cichlidae) from
- 938 Lake Tanganyika. Zootaxa. 49:39–49.
- 939 Schwarzer J., Misof B., Tautz D., Schliewen U.K. 2009. The root of the East African cichlid 940 radiations. BMC Evol. Biol. 9:186.
- 941 Schweiger O., Klotz S., Durka W., Kühn I. 2008. A comparative test of phylogenetic diversity indices. Oecologia. 157:485–95.
- 943 Sedano R.E., Burns K.J. 2010. Are the Northern Andes a species pump for Neotropical birds?
- Phylogenetics and biogeography of a clade of Neotropical tanagers (Aves: Thraupini). J.
- 945 Biogeogr. 37:325–343.
- Seehausen O. 2000. Explosive speciation rates and unusual species richness in
- haplochromine cichlid fishes: effects of sexual selection. Adv. Ecol. Res. 31:237–274.
- 948 Seehausen O. 2006. African cichlid fish: a model system in adaptive radiation research. Proc.
- 949 R. Soc. B Biol. Sci. 273:1987–98.
- 950 Sefc K.M., Mattersdorfer K., Ziegelbecker A., Neuhüttler N., Steiner O., Goessler W.,
- Koblmüller S. 2017. Shifting barriers and phenotypic diversification by hybridisation.
- 952 Ecol. Lett.
- 953 Shirai K., Inomata N., Mizoiri S., Aibara M., Terai Y., Okada N., Tachida H. 2014. High
- prevalence of non-synonymous substitutions in mtDNA of cichlid fishes from Lake
- 955 Victoria. Gene. 552:239–245.
- 956 Siler C.D., Oaks J.R., Esselstyn J.A., Diesmos A.C., Brown R.M. 2010. Phylogeny and
- biogeography of Philippine bent-toed geckos (Gekkonidae: Cyrtodactylus) contradict a
- prevailing model of Pleistocene diversification. Mol. Phylogenet. Evol. 55:699–710.
- 959 Spady T.C., Seehausen O., Loew E.R., Jordan R.C., Kocher T.D., Carleton K.L. 2005.
- Adaptive molecular evolution in the opsin genes of rapidly speciating cichlid species.
- 961 Mol. Biol. Evol. 22:1412–1422.
- Sparks J.S., Smith W.L.W.L. 2004. Phylogeny and biogeography of cichlid fishes (Teleostei:
- Perciformes: Cichlidae). Cladistics. 20:501–517.
- 964 Sturmbauer C., Baric S., Salzburger W., Rüber L., Verheyen E. 2001. Lake level fluctuations
- synchronize genetic divergences of cichlid fishes in African lakes. Mol. Biol. Evol.
- 966 18:144–54.
- 967 Sturmbauer C., Salzburger W., Duftner N., Schelly R., Koblmüller S. 2010. Evolutionary
- history of the Lake Tanganyika cichlid tribe Lamprologini (Teleostei: Perciformes)
- derived from mitochondrial and nuclear DNA data. Mol. Phylogenet. Evol. 57:266–84.

- 970 Sugawara T., Terai Y., Okada N. 2002. Natural Selection of the Rhodopsin Gene During the
- 971 Adaptive Radiation of East African Great Lakes Cichlid Fishes. Mol. Biol. Evol.
- 972 19:1807–1811.
- 973 Thorpe R.S., Surget-Groba Y., Johansson H. 2008. The relative importance of ecology and
- geographic isolation for speciation in anoles. Philos. Trans. R. Soc. Lond. B. Biol. Sci.
- 975 363:3071–81.
- Toni T., Stumpf M.P.H. 2010. Simulation-based model selection for dynamical systems in systems and population biology. Bioinformatics. 26:104–110.
- 978 Toni T., Welch D., Strelkowa N., Ipsen A., Stumpf M.P.H. 2009. Approximate Bayesian
- computation scheme for parameter inference and model selection in dynamical systems.
- 980 J. R. Soc. Interface. 6:187–202.
- Turner G.F., Seehausen O., Knight M.E., Allender C.J., Robinson R. 2001. How many
- species of cichlid fishes are there in African lakes? Mol. Ecol. 10:793–806.
- Vaidya G., Lohman D.J., Meier R. 2011. SequenceMatrix: Concatenation software for the
- fast assembly of multi-gene datasets with character set and codon information.
- 985 Cladistics. 27:171–180.

- Verheyen E., Rüber L., Snoeks J., Meyer A. 1996. Mitochondrial phylogeography of rock-
- dwelling cichlid fishes reveals evolutionary influence of historical lake level fluctuations
- of Lake Tanganyika, Africa. Philos. Trans. R. Soc. Lond. B. Biol. Sci. 351:797–805.
- Wagner C.E., Harmon L.J., Seehausen O. 2012. Ecological opportunity and sexual selection together predict adaptive radiation. Nature. 487:366–9.
- Wagner C.E., Harmon L.J., Seehausen O. 2014. Cichlid species-area relationships are shaped by adaptive radiations that scale with area. Ecol. Lett. 17:583–592.
- Wagner C.E., McIntyre P.B., Buels K.S., Gilbert D.M., Michel E. 2009. Diet predicts
   intestine length in Lake Tanganyika's cichlid fishes. Funct. Ecol. 23:1122–1131.
- Weir J.T. 2006. Divergent Timing and Patterns of Species Accumulation in Lowland and
   Highland Neotropical Birds. Evolution (N. Y). 60:842–855.
- 997 Weiss J.D., Cotterill F.P.D., Schliewen U.K. 2015. Lake Tanganyika A 'Melting Pot' of 998 Ancient and Young Cichlid Lineages (Teleostei: Cichlidae)?:1–29.
- Winkelmann K., Rüber L., Genner M.J. 2016. Lake level fluctuations and divergence of cichlid fish ecomorphs in Lake Tanganyika. Hydrobiologia::1–14.