cAMP signaling mediates behavioral flexibility and consolidation of social status

in *Drosophila* aggression

Running title: cAMP and behavioral plasticity in aggression

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1   **SUMMARY STATEMENT**

2   Phasic recruitment of different cAMP signaling modalities in specific neuronal groups

3   lead to the formation of temporally distinct components of learning and memory in fly

4   aggression.

ABSTRACT

Social rituals, like male-male aggression in *Drosophila*, are often stereotyped and the

component behavioral patterns modular. The likelihood of transition from one behavioral

pattern to another is malleable by experience and confers flexibility to the behavioral

repertoire. Experience-dependent modification of innate aggressive behavior in flies

alters fighting strategies during fights and establishes dominant-subordinate

relationships. Dominance hierarchies resulting from agonistic encounters are

consolidated to longer lasting social status-dependent behavioral modifications resulting

in a robust loser effect.

We show that cyclic adenosine monophosphate (cAMP) dynamics regulated by the

calcium/calmodulin-dependent adenylyl cyclase, Rut and the cAMP phosphodiesterase,

Dnc but not the Amn gene product, in specific neuronal groups of the mushroom body

and central complex, mediate behavioral plasticity necessary to establish dominant-

subordinate relationships. *rut* and *dnc* mutant flies are unable to alter fighting strategies

and establish dominance relationships during agonistic interactions. This real-time

flexibility during a fight is independent of changes in aggression levels. Longer-term

consolidation of social status in the form of a loser effect, however, requires additional

Amn-dependent inputs to cAMP signaling and involves a circuit-level association

between the α/β and γ neurons of the mushroom body.

Our findings implicate cAMP signaling in mediating plasticity of behavioral patterns in

aggressive behavior and in the generation of a temporally stable memory trace that

manifests as a loser effect.

1 INTRODUCTION

Aggression is a social behavior involving competitive interactions over resources to

ensure reproductive success and survival. Conspecific agonistic interactions may result

in dominance hierarchies, where those at higher levels have better access to resources.

The social ritual of *Drosophila* aggression is modular and comprises of stereotyped

behavioral patterns analogous to a sequence of fixed action patterns. These sequences

are present in full complexity in socially naive animals and appear to be pre-wired in the

nervous system (Chen et al., 2002; Hoopfer, 2016; Lim et al., 2014). However, during

aggressive encounters, experience can alter the likelihood of transitions between

specific patterns and lead to experience-dependent plasticity (Trannoy et al., 2016;

Yurkovic et al., 2006). Initial agonistic interaction between naïve flies constitutes a

conditioning phase where male flies employ a combination of offensive and defensive

fighting strategies and display real time, experience-dependent plasticity to establish

dominant or subordinate status (Chen et al., 2002; Yurkovic et al., 2006). In flies, the

learned subordinate social status is consolidated into a long lasting loser effect,

resulting in an increased probability of losing agonistic encounters against familiar as

well as unfamiliar opponents (Trannoy and Kravitz, 2016; Trannoy et al., 2016; Yurkovic

et al., 2006).

Aggression is a complex social behavior influenced by a combinatorial interplay

between genetic factors, environmental cues and experience. Population level selection

for elevated aggression has implicated several genes, which show significant changes

in their expression (Dierick and Greenspan, 2006; Edwards et al., 2009; Wang et al.,

2008). However, a single social defeat can lead to the development of a robust loser

effect in such hyperaggressive lines, underscoring the modulatory role of experience in

relation to intrinsic abilities (Penn et al., 2010). While regulatory activities of

pheromones, neuromodulatory agents and neurotransmitters associated with

aggression levels are well documented (Alekseyenko et al., 2014; Alekseyenko et al.,

2013; Andrews et al., 2014; Certel et al., 2010; Dierick and Greenspan, 2007; Hoopfer,

2016; Hoyer et al., 2008; Kohl et al., 2015; Luo et al., 2014; Wang and Anderson, 2010;

Wang et al., 2011; Yuan et al., 2014), little is known about the neurogenetic

underpinnings of the relevant learning and memory components.

Social experience significantly alters the activity of flies and involves both classical and

operant conditioning components (Kamyshev et al., 2002). Experience-dependent

behavioral plasticity has also been observed in courtship conditioning assays where

males, after an unsuccessful mating experience, modulate their courtship behavior

(Siegel and Hall, 1979). Learning and memory in a social environment are likely to be

influenced by several factors, including the combinatorial inputs from multiple sensory

modalities. Consequently, central integration and interpretation of complex input

modalities are likely to involve coordination between multiple neurogenetic circuits.

Previous studies have established the centrality of the cyclic adenosine monophosphate

(cAMP) pathway in the formation of operant and classical conditioned memories and in

the integration of sensory inputs (Bragina and Kamyshev, 2003; Brembs, 2003; Busto et

al., 2010; Gailey et al., 1984). Several mutations associated with defective learning and

memory have been mapped to the genetic loci of cAMP pathway components (Dudai et

al., 1976; Feany and Quinn, 1995; Levin et al., 1992; Livingstone et al., 1984). In this

study, we implicate the cAMP second messenger pathway in behavioral plasticity during

aggressive encounters and in the development of the loser effect. *rutabaga* (calcium-

calmodulin-dependent adenylyl cyclase; Rut) and *dunce* (cAMP phosphodiesterase;

Dnc) mutants show compromised behavioral flexibility during agonistic encounters and

are unable to establish dominance hierarchies. *amnesiac* (predicted to code for three

putative neuropeptides; Amn) mutants, although competent in modifying behavioral

patterns during fights and establishing hierarchies, show no loser effect. These studies

demonstrate that *Rut* in specific neural circuits mediates behavioral flexibility leading to

the establishment of dominance relationships and in the long-term consolidation of

social status.

MATERIALS AND METHODS

Fly stocks and maintenance

Canton S (CS), rut^{2080} and amn^{c651} lines were obtained from Dr R. Strauss, University of

Mainz, Germany. dnc^1, rut^{2080};UAS-Rut and all Gal4 driver lines were obtained from the

stock center at Bloomington, USA. All fly lines were backcrossed for at least nine

generations. The autosomes of the X-linked cAMP pathway mutant alleles used were

equilibrated to that of the control strain. Stocks were maintained at 25^0C, 60% humidity

and a 14h:10h::light:dark cycle on standard food.

Analysis of aggressive behavior

Freshly eclosed flies were isolated and kept in social isolation for a period of 4-5 days

before testing. Acrylic paint marks on the upper thoracic region were used to identify

individuals. Male – male aggression assays were conducted as described earlier (Chen

et al., 2002) with the modification that they were conducted in a six-well chamber. All

experiments were conducted between Zeitgeber Time (ZT) 0 and ZT 6 unless otherwise

specified. A food cup with yeast paste and a headless female was placed inside a six-

well plate chamber. A pair of marked, un-anesthetized, age- and size- matched male

flies was introduced into the chamber through gentle aspiration. All fights were

conducted at 25^0C and 60% humidity and recorded using a Sony DCR-SR47E/S video

camera at 50 Hz. Fights between socially naïve flies involved three phases. In the 'fight

phase' a pair of naïve flies was allowed to fight for 60 min, this was followed by a 'rest

phase' of 60 min where flies were returned to their original food vials and finally a 'test

phase' where previously matched flies fought against unfamiliar, naïve opponents for 60

min. SONY PMB software on Windows OS was used for video playback and the fights

were manually curated. In all cases the analyzer was blind to the genotype of the fly.

Fights were analyzed on basis of encounters that involved physical interactions and

lasted at least 3 seconds. Encounters were considered separate if the time interval

between them exceeded 2 seconds. Winners and losers were designated based on an

ethologically characterized three lunge-three retreat rule (developed in (Yurkovic et al.,

2006)). Lunge is defined as a maneuver where a fly rears up on its hind legs and

collapses on the opponent (Chen et al., 2002; Zwarts et al., 2012). A retreat occurs

when a fly, in response to an offensive action, runs/flies away from the opponent or the

food cup (territory) (Chen et al., 2002; Zwarts et al., 2012). Within a trial, a fly was

assigned a status of 'winner' if it used three continuous lunges (without any interim

retreats) and its opponent executed three corresponding retreats (with no intervening

lunges) (Yurkovic et al., 2006). The latter was designated as a 'loser'.

We analyzed various parameters to investigate aggression in flies across genotypes.

They include: 1. Encounter frequency (encounters per minute measured as the total

number of encounters divided by total time of fighting); 2. Aggression vigor index (the

fraction of time spent fighting in the first 10 min following the commencement of the first

encounter). 3. Latency to engage in an encounter (time in seconds from the start of the

fight to initiate the first agonistic interaction that lasts for at least 3 seconds). In the

second fights, the 'Draw' outcomes, where 3 Lunge- 3 Retreat rule was not satisfied,

were further divided into three categories based on Penn *et al.* (2010). This

categorization was based on usage of lunges and retreats by experienced loser flies

against a naïve opponent. 'High intensity' draws consisted of experienced loser flies

predominantly using lunges while 'low intensity' draws included usage of retreats. Fights

in which the flies did not engage in agonistic interactions were considered as 'no

intensity' and categorized as draws. An experienced loser fly may not readily engage in

agonistic interactions against naïve opponents. This may preclude escalation during

fights and result in higher number of draws. Thus 'high/low intensity' categorization

facilitates better assessment of the status-dependent behavioral changes manifested as

the loser effect.

We devised the loser index in order to assess the ability of flies to demonstrate the

experience dependent loser effect. The loser index was calculated as a difference

between the numbers of encounters lost to the encounters won divided by the total

number of encounters in the second fight. Within an encounter, if a fly uses aggressive

actions like lunging, tussling (tugging and pulling the body of the opponent), boxing

(rearing up on hind legs and striking the opponent with forelegs), holding (rearing up on

hind legs and holding the other fly's abdomen), chasing (running after the opponent) or

fencing (extending its leg forward and pushing the other fly) (Chen et al., 2002; Zwarts

et al., 2012) and the other fly responds with a retreat then the former fly is a 'winner' and

the latter a 'loser' in that encounter.

Analysis of locomotor behavior

Locomotor behavior was analyzed using a negative geotaxis assay as described before

(Ali et al., 2011). A group of ten flies, age and size matched, were introduced into a food

vial one day prior to testing. The flies were placed in two head to head joined empty

food vials with a distance of 8 cm marked on the lower vial. Following a gentle tap to get

all the flies to the base of the vial, the number of flies able to climb above the 8 cm mark

in 10 seconds was scored. The experiment was repeated 10 times for each group and

the average pass rate calculated. Three experimental replicates were carried out for

each genotype.

Statistical analysis

Videotapes were analyzed and each encounter was scored for all fighting strategies and

documented on spreadsheets. All statistical analysis was performed using Statistica 8.0

and GraphPad Prism 6.0 statistical software. Various statistical tests were employed to

facilitate our analysis including Chi-square test, one-factor ANOVA and two-factor

repeated measures ANOVA followed by posthoc Tukey's or Dunnett's multiple

comparisons tests. Specific information on statistics used in each experiment is

included in the figure legends.

RESULTS

The cAMP pathway mediates establishment of hierarchies within a fight

Pairs of male flies demonstrate stereotyped aggressive behavior towards each other

while competing for resources like food or females. These dyadic interactions begin with

low aggression maneuvers like wing threat and fencing, which then escalate to high

aggression strategies like lunging and tussling. This escalation is a consequence of

experience-dependent behavioral transitions, with one fly demonstrating offensive

strategies with increasing frequency against an opponent fly employing more and more

defensive strategies. These real time behavioral modifications on shorter time scales

are stable within a fight and facilitate formation of social dominance hierarchies. We

have assigned dominance relationships using a previously developed three lunge –

three retreat rule (see Materials and Methods and (Yurkovic et al., 2006)). In wild-type

Canton S (CS) flies, 85% of the fights produced dominance relationships (Figure 1A).

Prior investigations have implicated the cAMP pathway in the development of learning

and memory in multiple conditioning paradigms. cAMP synthesis is regulated by the

enzymatic activity of the *Rut* encoded adenylyl cyclase that catalyzes the conversion of

ATP to cAMP (Levin et al., 1992; Livingstone et al., 1984). *Rut* has been implicated as a

biochemical coincidence detector necessary for the formation of short term memory

(STM) in olfactory conditioning assays (Dudai et al., 1988; McGuire et al., 2005). In

contrast to CS flies, rut^{2080} mutant flies were unable to form dominance relationships

with all their fights ending in draws (Figure 1A; $P < 0.001$). cAMP phosphodiesterase

activity encoded by *Dnc* negatively regulates cAMP levels (Davis and Kiger, 1981).

Consistent with the deregulation of cAMP dynamics, dnc^1 flies display attenuated

formation of dominance relationships with only 67% of the fights resulting in wins/losses

(Figure 1A; $P < 0.01$). The *Amn* gene product is thought to encode for neuropeptide/s

and is known to modulate cAMP signaling (Feany and Quinn, 1995; Moore et al., 1998)

However, *amn^{C651}* mutant flies displayed wild-type levels of dominance hierarchies

(Figure 1A; 82% of the fights, $P > 0.05$).

For further analysis of experience dependent behavioral modifications during a fight, we

focused on lunges and retreats. These strategies are commonly used offensive (lunges)

and defensive (retreats) maneuvers and previous work have indicated status dependent

modifications in their usage by winner/loser flies (Yurkovic et al., 2006). Behavioral

modulation was assessed by scoring numbers of lunges/retreats in first 25 encounters

in a fight. In line with previous observations, agonistic interactions resulted in an

escalation of aggression (Trannoy et al., 2016; Yurkovic et al., 2006). In aggressive

encounters between CS flies, the ultimate winners (as established by the 3 lunge-3

retreat rule; see Materials and Methods) progressively increased the deployment of

lunges while the losers increasingly adapted to the retreating behavior (Figure 1B, C). In

contrast, *rut^{2080}* mutant flies were unable to demonstrate any behavioral transitions

pertaining to lunges and retreats (Figure 1B, C; $P < 0.001$). *dnc^{1}* flies were not only

inefficient in establishing hierarchies but also displayed compromised behavioral

plasticity with significantly reduced ability to modify the frequency of lunges and retreats

compared to CS animals (Figure 1B, C; $P < 0.001$). *amn^{C651}* flies, which are competent

in establishing hierarchies, were able to modify the usage of offensive and defensive

strategies as well as CS flies (Figure 1B, C; $P > 0.05$). These results suggest a

correlation between experience dependent behavioral plasticity and formation of

dominance hierarchies following aggressive encounters.

No significant differences in locomotor activity between wild type and mutant flies were

found using a negative geotaxis assay (see Materials and Methods), thus ruling out the

possibility of overt motor deficits in these lines (Figure 2A; $P > 0.05$). Multiple

parameters were evaluated to assess the aggression levels of the wild type and mutant

lines. Encounter frequency was significantly lower in rut^{2080} mutants compared to CS

flies but dnc^1 and amn^{C651} mutants were comparable to wild-type flies (Figure 2B; $P_{rut} <$

0.05, P_{dnc} and $P_{amn} > 0.05$). Aggression vigor index was compromised in all the mutant

lines compared to CS but was not significantly different between rut^{2080}, dnc^1 and

amn^{C651} flies (Figure 2C; $P_{rut} < 0.001$, P_{dnc} and $P_{amn} < 0.05$).

While our studies implicate lack of behavioral plasticity in rut^{2080} mutants in establishing

dominance hierarchies in rut^{2080} versus rut^{2080} fights, it is also possible that the rut^{2080}

flies are unable to execute high intensity maneuvers like lunges. In $rut – rut$ fights,

neither opponent can adjust their fighting patterns depending upon experience resulting

in a lack of escalation beyond low intensity interactions precluding assessment of

inherent behavioral changes in rut^{2080} flies. CS flies have intact experience dependent

behavioral transitions compared to rut^{2080} mutants. Therefore, pairing CS vs rut^{2080} may

better reveal behavioral escalation during fights. CS vs rut^{2080} fights results in significant

dominance hierarchies with 37.5% of fights ending in a win or a loss, though it remains

attenuated compared to CS vs CS fights (Figure 3A; $P < 0.001$). Interestingly, rut^{2080}

flies demonstrate increased lunging and retreating behavior compared to their

performance in $rut-rut$ fights and also win 9% of the fights against wild-type opponents

(Figure 3B, C). rut^{2080} flies, when in a rut – CS fight, are capable of executing and

modifying the frequency of use of both lunges and retreats, although this ability remains

significantly compromised compared to that in CS flies (in CS – CS fights) (Figure 3B,

C; $P < 0.001$ for lunges and $P < 0.05$ for retreats). The usage of offensive and defensive

strategies for rut^{2080} flies against a wild type opponent is significantly better than rut-rut

fights (Figure 3B, C; $P < 0.001$ for lunges and $P < 0.01$ for retreats). These results

suggest that rut^{2080} flies are capable of executing high intensity maneuvers but are

compromised in modifying its frequency as well as CS flies during a fight.

Furthermore, dnc^{1} mutants are unable to modify fighting strategies as well as CS flies

but they are less compromised compared to the rut^{2080} flies in rut-rut fights (Figure 1A, B,

C). Consistent with the association of ability to modify fighting patterns with the

establishment of dominance relationships, dnc mutants do establish hierarchies though

less efficiently than wild-type CS flies (Figure 1A). Similarly, rut^{2080} flies against wild type

opponents demonstrate compromised plasticity and attenuated dominance structures

(Figure 3A, B, C). In fact, usage of lunges/retreats by rut^{2080} flies in rut – CS fights is

comparable to those seen in dnc^{1} (Figure 3B, C; $P > 0.05$ for lunges and retreats).

Interestingly, the aggression vigor index is similar for rut - CS and rut - rut fights (Figure

3D; $P > 0.05$).

This series of experiments establish the role of the cAMP signaling in behavioral

plasticity underlying modification of fighting strategies during agonistic interactions.

Behavioral flexibility, rather than changes in levels of aggression, is correlated with the

generation of dominance hierarchies.

Specific neural circuits are recruited in cAMP-mediated behavioral plasticity

We next asked which components of the *Drosophila* brain were involved in the status-

dependent modifications of aggressive strategies and formation of dominance

relationships. As multiple sensory modalities are likely to be involved in aggressive

encounters, a complex pattern of recruitment of neuronal circuits may underlie the

integration of these sensory inputs (Asahina et al., 2014; Hoopfer, 2016; Hoopfer et al.,

2015; Lim et al., 2014; Ramin et al., 2014; Wang and Anderson, 2010; Wang et al.,

2011; Yoon et al., 2013). To test this, we used the UAS-Gal4 binary system to express

the Rut gene product in restricted neuronal populations in a rut^{2080} mutant background.

We chose well-characterized Gal4 drivers that have restricted expression in defined

neuronal populations of the mushroom body and central complex of the fly brain as

these regions are strongly implicated as central integrators in multiple behavioral

paradigms (Table 1) (Aso et al., 2009; Blum et al., 2009; Joiner and Griffith, 1999;

Neuser et al., 2008; Pan et al., 2009; Torroja et al., 1999; Zars et al., 2000a; Zars et al.,

2000b). All experiments were conducted at the same time of the day. The genetic

rescue and the control experiments were done in parallel. Pan-neuronal expression of

Rut using the *Appl*-Gal4 driver completely rescued the inability to form dominance

relationships seen in rut^{2080} mutants (Figure 4A; $P < 0.001$). Similar restoration of

dominance hierarchies to wild-type levels was seen with expression limited to the α/β

and γ lobe neurons of the mushroom body using the c309 driver (Figure 4A; $P < 0.001$).

Next, we tested a panel of Gal4 drivers expressing *Rut* in different subpopulations of

neurons in the MB and the central complex of rut^{2080} flies (Table 1). UAS-Rutabaga

driven by Gal4 drivers c739, 201Y, c305a and c205, but not c819, partially rescued the

inability of rut^{2080} to form hierarchical relationships (Figure 4A; $P < 0.001$). These

experiments implicate the independent involvement of the α/β, γ and α'/β' lobes of the

MB together with the F5 neurons of the fan-shaped body (FB) in neuronal processing

leading to the formation of dominance. The ellipsoid body (EB) of the central complex

does not appear to be involved in this function (Figure 4A; $P > 0.05$).

Analysis of behavioral patterns revealed that distinct neuronal circuits mediate *rut*-

dependent behavioral plasticity within a fight (Figure 4 and Table 1). Pan-neuronal

expression of *Rut* using the *Appl*-Gal4 driver rescued the inability to alter the frequency

of use of lunges and retreats seen in rut^{2080} mutants (Figure 4B, C; $P < 0.01$). We also

observed a complete rescue of behavioral transitions pertaining lunges and retreats with

Rut expression limited to the α/β and γ lobe neurons of the mushroom body using the

c309 driver (Figure 4D, E; $P < 0.001$). Furthermore, expression of *Rut*, independently in

the α/β, α'/β' or γ neurons of the MB or in the FB also rescued the deficit in the

progressive increase of lunges and retreats seen in rut^{2080} flies (Figure 4D, E, F, G; $P <$

0.01). The ellipsoid body (EB) of the central complex does not appear to be involved in

this function as no rescue was observed with the EB only driver (Figure 4F, G; $P > 0.05$).

Restricted expression of *Rut* in specific brain regions rescued the behavioral plasticity of

lunges/retreats and the ability to establish dominance hierarchies within a fight.

To confirm that experience-dependent behavioral flexibility was a major substrate of the

rescue and not an indirect effect of changes in other general features of aggression, we

evaluated multiple aggression related parameters. As the rescue was strongest

(comparable to wild-type) with the *Appl*-Gal4 (pan neuronal) and c309 (α/β and γ

neurons of the MB) drivers, these were used to evaluate changes in aggressiveness.

However, no change in the frequency of encounters, aggression vigor and latencies to

engage was found (Figure 5A – E; $P > 0.05$). These results suggest that though pan

neuronal and α/β and γ drivers fully rescue the ability to alter the intensities of specific

fighting patterns during a fight and the establishment of dominance hierarchies, they do

not affect the levels of aggression in these animals.

Our analysis indicates that in accordance with the multimodal inputs associated with

dyadic interactions, multiple neuronal groups in the MB (α/β, α'/β', γ) and CC (FB) are

involved in the processing of these inputs that lead to the progressive changes in

offensive and defensive strategies within a fight. The rescue experiments also highlight

the role of cAMP signaling in behavioral plasticity in aggression to be independent of

general changes in aggression levels.

cAMP signaling is necessary for the development of the loser effect

To test if aggression-associated hierarchies developed in the first fights influenced the

outcomes of subsequent fights, losers from first fights were paired with unfamiliar,

socially naïve opponents, after a 60-minute rest period. Dominance relationships were

assigned using the three lunge- three retreat rule as in the first fights. Fights that fell

under the 'draw' category were further subdivided into three groups. 'High intensity'

draws consisted of experienced flies predominantly using lunges in their second fight

without satisfying 3 lunge-3 retreat criteria for dominance relationships. Similarly, 'low

intensity' draws consisted of experienced flies predominantly using retreats. Fights in

which the flies did not engage in agonistic interactions were considered as 'no intensity'

and categorized as draws. For statistical analysis, high and low intensity draws were

grouped with wins and losses, respectively. Consistent with previous studies (Trannoy

et al., 2016; Yurkovic et al., 2006), wild-type CS loser flies always lost to naïve

opponents in their second fights (Figure 6A). This indicates that an experience of social

loss results in a strong loser effect in subsequent fights.

As rut^{2080} flies did not generate winners or losers in the first fight, both the individuals

were considered as experienced and used in second fights. However, as in their first

fights, rut^{2080} mutants were unable to form any dominance relationships in their second

fights (Figure 6A; $P < 0.001$). dnc^{1} also displayed no significant loser effect with 18%

losses, 18% wins and 64% draws (Figure 6A*; P* < 0.001).

Surprisingly, amn^{C651} flies did not show any loser effect. 50% of analyzed second fights

resulted in losses and the remaining in wins for losers (Figure 6A; $P < 0.001$). Unlike rut

and dnc, amn^{C651} flies develop dominant-subordinate relations as efficiently as CS in

their first fights (Figure 1C) but are still unable to consolidate this experience into a loser

effect. This suggests a special requirement of Amn gene product in stabilizing the

hierarchical structures in flies in the form of a loser effect.

A loser index, calculated as the difference between the number of encounters lost and

the number of encounters won divided by the total number of encounters in the second

fight (see Materials and Methods), was used to represent the loser effect as a

consequence of past experience. This allowed direct evaluation of experience-

dependent alterations in fighting strategy in the losers across genotypes in subsequent

fights. CS flies showed a high loser index consistent with their robust loser effect (Figure

6B). In contrast, the amn^{C651} and dnc^{1} losers, and the experienced rut^{2080} flies displayed

significantly lower loser indices (Figure 6B; $P < 0.001$).

These results show that dnc^{1} and rut^{2080} flies, which display compromised development

of dominance hierarchies in their first fights, did not display a loser effect. In contrast,

amn^{C651}, which could form dominance hierarchies in their first fights, also lacked the

loser effect. *Amn* appears to have an independent function from *Rut* and *Dnc* and is

necessary for the stabilization of the social status from the first fight in the form of a

loser effect.

α/β and γ lobes of the MB cooperate to mediate cAMP-dependent loser effect

Previous experience of loss results in a robust loser effect in wild-type flies. We

investigated the neurogenetic circuitry involved in the development of this effect.

In the second fights, both *Appl* and c309 Gal4 driver lines were able to restore the loser

effect in *rut^{2080}* flies to levels statistically indistinguishable from wild-type CS flies (Figure

7A, B and Table 1). Pan neuronal (Appl) or combined α/β and γ lobe–specific (c309)

expression resulted in most losers losing and a small proportion of non-aggressive

draws (Figure 7A; $P < 0.001$). Loser index in these flies was significantly greater than

controls (Figure 7B; $P < 0.001$) and comparable to wild-type flies (Figure 7B; $P > 0.05$).

Interestingly, expression of UAS-*Rut* limited to individual substructures of the MB and

CC was not sufficient to rescue impaired loser effect observed in *rut* flies (Figure 7B;

Table 1). Expression in individual neuronal subpopulations using the c739, 201Y, c305a

or c205 Gal4 drivers (in the *rut^{2080}* background), resulted in a higher proportion of non-

draw outcomes in losers paired with naïve opponents but failed to show a robust loser

effect (Figure 7A; $P < 0.001$). As expected, there was no significant change in the loser

index for these flies (Figure 7B; $P > 0.05$). Consistent with compromised learning in the

first fights, expression limited to the EB using the c819 Gal4 driver did not rescue the

loser effect (Figure 7; $P > 0.05$).

Our results implicate combined processing by the α/β and γ neurons of the MBs in the

development of the loser effect.

DISCUSSION

Two distinct experience-dependent behavioral modifications are observed in *Drosophila*

aggression. The first results in dominance relationships correlated with real time,

progressive changes in fighting strategies. The second is the consolidation of the

experience of a previous loss to a loser effect in subsequent fights. Our data indicate

that these behavioral changes involve specific cAMP-mediated signaling and are

processed by overlapping neuronal circuits involving two temporally separated memory

traces.

Rut and Dnc enzymes have been implicated in learning and/or STM in multiple

paradigms, including classical conditioning and courtship-associated learning and

memory (Bragina and Kamyshev, 2003; Gailey et al., 1984; Livingstone et al., 1984;

McGuire et al., 2005). *rut* and *dnc* flies do not develop dominance hierarchies as well as

CS flies, with all *rut – rut* fights resulting in draws. Compared to CS, both *rut* and *dnc*

are unable to display experience-dependent modification of fighting strategies and are

deficient in generating losers and winners.

rut and *dnc* mutant flies are deficient in memory formation in various behavioral assays

(Dudai et al., 1976; Gailey et al., 1984; Livingstone et al., 1984; McGuire et al., 2005). In

olfactory conditioning, both *rut* and *dnc* mutants demonstrate rapid memory degradation

within an hour of training but have no impact on later memory (Chen et al., 1986; Levin

et al., 1992; Tamura et al., 2003). Also, *rut* mutant flies were unable to demonstrate

robust 30 min-memory after an hour of training in a courtship conditioning paradigm

(Ishimoto et al., 2013). Therefore, *rut* and *dnc* mutants' inability to alter behavioral states

might be due to poor memory acquisition pertaining winner-loser status, which results in

lack of hierarchical relationships in a fight.

The Amn gene product encodes for three putative neuropeptides (Feany and Quinn,

1995 and Moore et al., 1998). Genetic and biochemical studies have implicated the

Amn locus in cAMP dynamics (Feany and Quinn, 1995; Moore et al., 1998;

Bhattacharya et al., 2004). It is assumed, from analogous data in other organisms, that

the Amn gene product/s activate adenylyl cyclase activity via GPCR signaling (Abel and

Kandel, 1995; McGuire et al., 2005). However, there is no direct evidence of the Amn

gene products being secreted and acting as a classical neuropeptide via GPCR

activation. In our experiments, *amn* flies are as competent in modifying fighting

strategies and establishing hierarchal relationships as CS flies but are unable to retain

the loser status in subsequent fights. These results suggest that the plasticity in

modifying aggressive strategies during a fight is mediated by cAMP dynamics under the

control of Rut and Dnc gene products (but not Amn) and is necessary to establish

dominance structures.

All three cAMP pathway mutants displayed a reduction in aggression vigor as compared

to CS. This includes *amn*, which displays normal behavioral flexibility and dominance

hierarchies. These results suggest that the level of aggression in flies is independent of

their ability to modify behavior in an experience-dependent manner. The latency to

engage in fights in cAMP mutants was also comparable to CS. The latter suggests no

overt changes in motivational states and support the previous conclusion.

Brain region specific expression of the Rut gene product in a *rut* mutant background

implicates specific neuronal circuits in status-dependent behavioral changes during

agonistic encounters. All the drivers that rescue the establishment of dominance also

restore the ability to alter the usage of lunges and retreats during a fight.

Rescue by Appl or c309 did not alter the encounter frequency, aggression vigor or the

latency to engage displayed by *rut* mutants. In line with our analysis of cAMP pathway

mutants, these results demonstrate that the rescue of dominance is attributable to a

restoration of experience-dependent plasticity mediated by *Rut* in specific circuits and

not a consequence of modified aggressiveness.

Our study suggests a functional role for MBs and Central Complex (CC) in learning and

memory associated with aggression, consistent with those described in courtship

conditioning (Joiner and Griffith, 1999; McBride et al., 1999; Sitnik et al., 2003).

Mushroom bodies, which are central to olfactory learning, have been previously

correlated with changes in agonistic behavior. *Neuralized* mutants with altered MB

organization have been reported to increase aggressiveness when food is limiting

(Rollmann et al., 2008). Inhibition of synaptic output from the MB has also been shown

to reduce levels of aggression (Liu et al., 2011).

Interestingly, not only neuronal groups of the MB (the α/β, γ and α'/β' neurons) were

implicated, the F5 neurons of the FB but not the EB of the CC was also found to be

functionally involved. The FB has been previously reported to mediate visual learning of

specific pattern features (Liu et al., 2006; Pan et al., 2009). A recent study has

underscored the importance of the CC in aggression by demonstrating the modulation

of aggressive behavior by dopaminergic PPM3 neurons that synapse onto the FB

(Alekseyenko et al., 2013). While there is no evidence of direct anatomical connectivity

between the MB and the F5 neurons of the FB (Li et al., 2009; Young and Armstrong,

2010), it is possible that they are functionally linked by neuromodulatory dopaminergic

signaling.

In the courtship suppression paradigm, multiple biochemically and anatomically distinct

neural circuits have been implicated (Joiner and Griffith, 2000). Our observations in

aggression-associated behaviors are analogous and possibly reflect the integration of

multiple sensory inputs contributing to the establishment of dominance hierarchies.

The reduced aggression seen in cAMP pathway mutants could be due to alteration in

the production of aggression-associated signal/s and/or their perception. While this

possibility awaits systematic investigation, the rescue of behavioral flexibility without

altering aggressiveness seen in our experiments suggests that the two may be

separable. It is also possible that the observed behavioral deficits originate from

neurodevelopmental functions of these genes. Our study does not explicitly rule this out

though adult specific expression of Rut is sufficient to rescue learning and memory

defects in other behavioral paradigms (Mao et al., 2004; McGuire et al., 2003; Putz and

Heisenberg, 2002; Tan et al., 2010). Future studies will need to address this question

specifically in the context of aggression.

The robust loser effect seen in wild-type flies is absent in *rut*, *dnc* and *amn* mutant lines.

As in their first fights, all *rut-rut* second fights end in draws. No loser effect was also

seen in *dnc* flies. Both *rut* and *dnc* have compromised development of hierarchies

suggesting that learning within a fight and establishing dominance relationships are

necessary for the development of the loser effect. Interestingly, *amn* flies also lack the

loser effect, though they can adjust fighting strategies and develop dominance

relationships as well as wild-type flies. The Amn gene product appears to mediate the

consolidation of the social status acquired during the first fight into the loser effect.

Alternatively, Amn may be necessary for a distinct memory phase resulting in the loser

effect. A similar function for Amn has been reported in olfactory conditioning

experiments where *amn* mutants fail to develop intermediate-term memory or are

unable to consolidate STM into this relatively longer lasting phase (Feany and Quinn,

1995; Yu et al., 2006). Amn has also been reported to mediate memory stability in

courtship conditioning (Siegel and Hall, 1979). Our findings support this model as *amn*

flies do establish winner-loser relationships but fail to consolidate these dominance

hierarchies. If the Amn gene product functions as a secreted neuropeptide, future

rescue experiments involving the cognate receptor/s will be necessary to determine the

neural circuits subserving Amn-dependent establishment of the loser effect. A less

robust, short duration winner effect has been recently reported in flies (Trannoy and

Kravitz, 2016; Trannoy et al., 2016). It remains to be seen if cAMP signaling and circuit

features analogous to our observations mediate this phenomenon.

Rescue of *rut* simultaneously in the α/β and γ lobe neurons of the MB, not only fully

rescues its ability to generate dominance relationships but also the loser effect.

However, *rut* expression independently in the α/β or the γ neurons were unable to

restore the loser effect, though social hierarchy was rescued. We hypothesize that two

distinct Rut dependent memory traces facilitate the formation of the temporally distinct

memory phases. A short-lived engram in distinct MB and CC substructures mediates

behavioral plasticity within a fight leading to the formation of dominance relationships.

Combinatorial processing between α/β and γ lobe neurons of the MB enable formation

of a second, longer lasting memory trace that is necessary for the establishment of the

loser effect.

Olfactory conditioning studies have suggested that the α/β neurons are indispensable

for memory consolidation and retrieval, while odor-shock coincidence detection maps to

the γ neurons (Dubnau and Chiang, 2013; Dubnau et al., 2001; Qin et al., 2012). These

observations demonstrate recruitment of specific subsets of MB neurons for initial

associations and others for memory consolidation suggesting circuit level coordination

between MB substructures. Our experiments provide evidence towards such systems

level memory consolidation in aggression where an association between α/β and γ

lobes of the MB is critical for the establishment of the loser effect. However, the

existence of two parallel, independent traces with differing kinetics of formation and

decay cannot be formally ruled out.

Experience-dependent modification of innate behaviors involves multiple components

and previous studies have demonstrated the central importance of pheromonal,

aminergic and other modulatory activities in aggressive behavior (Alekseyenko et al.,

2014; Alekseyenko et al., 2013; Andrews et al., 2014; Certel et al., 2010; Chan and

Kravitz, 2007; Dankert et al., 2009; Dierick and Greenspan, 2007; Hoopfer, 2016; Hoyer

et al., 2008; Kohl et al., 2015; Liu et al., 2011; Luo et al., 2014; Wang and Anderson,

2010; Yuan et al., 2014; Zwarts et al., 2012). However, these studies focus on

aggression levels and do not directly assess behavioral plasticity and memory

components in aggression. A recent study indicated that rut^{2080} and amn^1 are unable to

demonstrate dominance hierarchies due to compromised aggression (Trannoy et al.,

2016). However, this study did not explore the lack of behavioral flexibility in these

mutants. In our study, rut^{2080} has low proclivity to engage in the first 10 min of fights, fail

to form dominance relations but are capable of executing high intensity maneuvers. In

contrast, amn^{C651} (a functionally strong allele of *amn* (Rosay et al., 2001)), showed

reduced aggression vigor but was still able to establish dominance hierarchies. Further,

the rescue of *Rut* restores dominance patterns but not aggressiveness. Therefore, low

aggression is not correlated with the inability to establish winner-loser statuses in a

fight.

cAMP signaling appears to have a conserved role in establishing dominance

relationships across phyla. As observed previously in the olfactory conditioning and

courtship paradigms, *rut* and *dnc* mutants display similar behavioral deficits in

aggression-associated behavioral flexibility despite having opposing biochemical

functions. Amn activity is also thought to increase cAMP levels (perhaps with different

kinetics than Rut). Thus, cAMP levels and fine-tuning of cAMP levels may encode signal

integration by the cAMP second messenger system. In crayfish, accumulation of cAMP

is necessary to form the loser effect while a reduction in cAMP levels is associated with

the winner effect (Momohara et al., 2016). Biogenic amines have been associated with

aggression and winner/loser effects in animals including flies (Alekseyenko et al., 2014;

Alekseyenko et al., 2013; Andrews et al., 2014; Hoyer et al., 2008; Zhou and Rao,

2008), crayfish (Momohara et al., 2016; Panksepp, 2003), crickets (Rillich and

Stevenson, 2014; Rillich and Stevenson, 2015; Stevenson et al., 2005) and mammals

(de Almeida et al., 2005). Aminergic signaling commonly impinges on cAMP/Protein

Kinase A (PKA) pathways implicating the cAMP signaling as a central effector of

neuromodulation by biogenic amines (Neckameyer and Leal, 2017). In mammals, the

cAMP pathway has been shown to influence aggressive behavior (Breuillaud et al., 2012). cAMP signaling in the basolateral amygdala has also been correlated with memory associated with conditioned defeat, a paradigm analogous to the loser effect (Jasnow et al., 2005; Markham et al., 2010). At the circuit level, recent work in zebrafish has implicated antagonistic regulation by two sub-regions of the dorsal habenula to establish winner-loser status (Chou et al., 2016).

Our results implicate sequential recruitment of cAMP signaling components with Rut and Dnc activities required for behavioral flexibility within a fight and, consequently, to establish dominance hierarchies. Behavioral plasticity, rather than aggressiveness, is correlated with social status. Establishment of social status facilitates the formation of a more stable and longer lasting memory phase that requires the additional involvement of the Amn gene product. Neuronal circuits subserving aggression associated learning and memory show phasic recruitment. While a short-lived Rut-dependent trace in multiple MB and CC substructures mediate learning during agonistic encounters, combinatorial processing by both α/β and γ lobe neurons of the MB, is necessary for the development of the longer lasting loser effect.

This study provides mechanistic insight into circuit level associations specific to different phases of behavioral plasticity and memory in *Drosophila* aggression and the integration of biochemical signaling at the single neuron level with systems level consolidation.

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REFERENCES

Alekseyenko, O. V., Chan, Y. B., Fernandez, M. P., Bulow, T., Pankratz, M. J. and Kravitz, E. A. (2014). Single serotonergic neurons that modulate aggression in *Drosophila*. *Curr Biol* **24**, 2700-7.

Alekseyenko, O. V., Chan, Y. B., Li, R. and Kravitz, E. A. (2013). Single dopaminergic neurons that modulate aggression in *Drosophila*. *Proc Natl Acad Sci USA* **110**, 6151-6.

Ali, Y., Escala, W., Ruan, K. and Zhai, R. (2011). Assaying locomotor, learning, and memory deficits in *Drosophila* models of neurodegeneration. *J Vis Exp*, e2504.

Andrews, J. C., Fernandez, M. P., Yu, Q., Leary, G. P., Leung, A. K., Kavanaugh, M. P., Kravitz, E. A. and Certel, S. J. (2014). Octopamine neuromodulation regulates Gr32a-linked aggression and courtship pathways in *Drosophila* males. *PLoS Genet* **10**, e1004356.

Asahina, K., Watanabe, K., Duistermars, B. J., Hoopfer, E., Gonzalez, C. R., Eyjolfsdottir, E. A., Perona, P. and Anderson, D. J. (2014). Tachykinin-expressing neurons control male-specific aggressive arousal in *Drosophila*. *Cell* **156**, 221-35.

Aso, Y., Grubel, K., Busch, S., Friedrich, A. B., Siwanowicz, I. and Tanimoto, H. (2009). The mushroom body of adult *Drosophila* characterized by GAL4 drivers. *J Neurogenet* **23**, 156-72.

Blum, A. L., Li, W., Cressy, M. and Dubnau, J. (2009). Short- and long-term memory in *Drosophila* require cAMP signaling in distinct neuron types. *Curr Biol* **19**, 1341-50.

Bragina, Y. V. and Kamyshev, N. G. (2003). Comparative studies of four *Drosophila* P-insertion mutants with memory defects. *Neurosci Behav Physiol* **33**, 73-9.

Brembs, B. (2003). Operant conditioning in invertebrates. *Curr Opin Neurobiol* **13**, 710-7.

Breuillaud, L., Rossetti, C., Meylan, E. M., Merinat, C., Halfon, O., Magistretti, P. J. and Cardinaux, J. R. (2012). Deletion of CREB-regulated transcription coactivator 1 induces pathological aggression, depression-related behaviors, and neuroplasticity genes dysregulation in mice. *Biol Psychiatry* **72**, 528-36.

Busto, G., Cervantes-Sandoval, I. and Davis, R. (2010). Olfactory learning in *Drosophila*. *Physiology* **25**, 338-346.

Certel, S. J., Leung, A., Lin, C. Y., Perez, P., Chiang, A. S. and Kravitz, E. A.
(2010). Octopamine neuromodulatory effects on a social behavior decision-making
network in *Drosophila* males. *PLoS One* **5**, e13248.

Chan, Y.-B. and Kravitz, E. (2007). Specific subgroups of FruM neurons control
sexually dimorphic patterns of aggression in *Drosophila melanogaster*. *Proc Natl Acad*
Sci USA **104**, 19577-19582.

Chen, C. N., Denome, S. and Davis, R. L. (1986). Molecular analysis of cDNA
clones and the corresponding genomic coding sequences of the *Drosophila* dunce+
gene, the structural gene for cAMP phosphodiesterase. *Proc Natl Acad Sci U S A* **83**,
9313-7.

Chen, S., Lee, A. Y., Bowens, N. M., Huber, R. and Kravitz, E. A. (2002).
Fighting fruit flies: a model system for the study of aggression. *Proc Natl Acad Sci USA*
99, 5664-8.

Chou, M. Y., Amo, R., Kinoshita, M., Cherng, B. W., Shimazaki, H.,
Agetsuma, M., Shiraki, T., Aoki, T., Takahoko, M., Yamazaki, M. et al. (2016). Social
conflict resolution regulated by two dorsal habenular subregions in zebrafish. *Science*
352, 87-90.

Dankert, H., Wang, L., Hoopfer, E., Anderson, D. and Perona, P. (2009).
Automated monitoring and analysis of social behavior in *Drosophila*. *Nat Meth* **6**, 297-
303.

Davis, R. L. and Kiger, J. A. (1981). Dunce mutants of *Drosophila*
melanogaster: mutants defective in the cyclic AMP phosphodiesterase enzyme system.
J Cell Biol **90**, 101-7.

de Almeida, R. M., Ferrari, P. F., Parmigiani, S. and Miczek, K. A. (2005).
Escalated aggressive behavior: dopamine, serotonin and GABA. *Eur J Pharmacol* **526**,
51-64.

Dierick, H. A. and Greenspan, R. J. (2006). Molecular analysis of flies selected
for aggressive behavior. *Nat Genet* **38**, 1023-31.

Dierick, H. A. and Greenspan, R. J. (2007). Serotonin and neuropeptide F have
opposite modulatory effects on fly aggression. *Nat Genet* **39**, 678-82.

Dubnau, J. and Chiang, A. S. (2013). Systems memory consolidation in
Drosophila. *Curr Opin Neurobiol* **23**, 84-91.

Dubnau, J., Grady, L., Kitamoto, T. and Tully, T. (2001). Disruption of
neurotransmission in *Drosophila* mushroom body blocks retrieval but not acquisition of
memory. *Nature* **411**, 476-80.

Dudai, Y., Corfas, G. and Hazvi, S. (1988). What is the possible contribution of
Ca^{2+}-stimulated adenylate cyclase to acquisition, consolidation and retention of an
associative olfactory memory in *Drosophila*. *J Comp Physiol A* **162**, 101-9.

Dudai, Y., Jan, Y. N., Byers, D., Quinn, W. G. and Benzer, S. (1976). dunce, a
mutant of *Drosophila* deficient in learning. *Proc Natl Acad Sci USA* **73**, 1684-8.

Edwards, A., Zwarts, L., Yamamoto, A., Callaerts, P. and Mackay, T. (2009).
Mutations in many genes affect aggressive behavior in *Drosophila melanogaster*. *BMC*
biology **7**, 29.

Feany, M. B. and Quinn, W. G. (1995). A neuropeptide gene defined by the
Drosophila memory mutant amnesiac. *Science* **268**, 869-73.

Gailey, D., Jackson, F. and Siegel, R. (1984). Conditioning mutations in
Drosophila melanogaster affect an experience dependent behavioral modification in
courting males. *Genetics* **106**, 613-623.

Hoopfer, E. D. (2016). Neural control of aggression in *Drosophila*. *Curr Opin*
Neurobiol **38**, 109-18.

Hoopfer, E. D., Jung, Y., Inagaki, H. K., Rubin, G. M. and Anderson, D. J.
(2015). P1 interneurons promote a persistent internal state that enhances inter-male
aggression in *Drosophila*. *Elife* **4**.

Hoyer, S. C., Eckart, A., Herrel, A., Zars, T., Fischer, S. A., Hardie, S. L. and
Heisenberg, M. (2008). Octopamine in male aggression of *Drosophila*. *Curr Biol* **18**,
159-67.

Ishimoto, H., Wang, Z., Rao, Y., Wu, C. F. and Kitamoto, T. (2013). A novel
role for ecdysone in *Drosophila* conditioned behavior: linking GPCR-mediated non-
canonical steroid action to cAMP signaling in the adult brain. *PLoS Genet* **9**, e1003843.

Jasnow, A. M., Shi, C., Israel, J. E., Davis, M. and Huhman, K. L. (2005).
Memory of social defeat is facilitated by cAMP response element-binding protein
overexpression in the amygdala. *Behav Neurosci* **119**, 1125-30.

Joiner, M. and Griffith, L. (1999). Mapping of the anatomical circuit of CaM
kinase-dependent courtship conditioning in *Drosophila*. *Learn Mem* **6**, 177-192.

Joiner, M. A. and Griffith, L. C. (2000). Visual input regulates circuit configuration in courtship conditioning of Drosophila melanogaster. *Learn Mem* **7**, 32-42.

Kamyshev, N. G., Smirnova, G. P., Kamysheva, E. A., Nikiforov, O. N., Parafenyuk, I. V. and Ponomarenko, V. V. (2002). Plasticity of social behavior in *Drosophila*. *Neurosci Behav Physiol* **32**, 401-8.

Kohl, J., Huoviala, P. and Jefferis, G. S. (2015). Pheromone processing in *Drosophila*. *Curr Opin Neurobiol* **34**, 149-57.

Levin, L. R., Han, P. L., Hwang, P. M., Feinstein, P. G., Davis, R. L. and Reed, R. R. (1992). The *Drosophila* learning and memory gene rutabaga encodes a Ca^{2+}/Calmodulin-responsive adenylyl cyclase. *Cell* **68**, 479-89.

Lim, R. S., Eyjolfsdottir, E., Shin, E., Perona, P. and Anderson, D. J. (2014). How food controls aggression in *Drosophila*. *PLoS One* **9**, e105626.

Liu, G., Seiler, H., Wen, A., Zars, T., Ito, K., Wolf, R., Heisenberg, M. and Liu, L. (2006). Distinct memory traces for two visual features in the *Drosophila* brain. *Nature* **439**, 551-6.

Liu, W., Liang, X., Gong, J., Yang, Z., Zhang, Y. H., Zhang, J. X. and Rao, Y. (2011). Social regulation of aggression by pheromonal activation of Or65a olfactory neurons in *Drosophila*. *Nat Neurosci* **14**, 896-902.

Livingstone, M. S., Sziber, P. P. and Quinn, W. G. (1984). Loss of calcium/calmodulin responsiveness in adenylate cyclase of rutabaga, a *Drosophila* learning mutant. *Cell* **37**, 205-15.

Luo, J., Lushchak, O. V., Goergen, P., Williams, M. J. and Nassel, D. R. (2014). *Drosophila* insulin-producing cells are differentially modulated by serotonin and octopamine receptors and affect social behavior. *PLoS One* **9**, e99732.

Mao, Z., Roman, G., Zong, L. and Davis, R. L. (2004). Pharmacogenetic rescue in time and space of the *rutabaga* memory impairment by using Gene-Switch. *Proc Natl Acad Sci U S A* **101**, 198-203.

Markham, C. M., Taylor, S. L. and Huhman, K. L. (2010). Role of amygdala and hippocampus in the neural circuit subserving conditioned defeat in Syrian hamsters. *Learn Mem* **17**, 109-16.

McBride, S., Giuliani, G., Choi, C., Krause, P., Correale, D., Watson, K., Baker, G. and Siwicki, K. (1999). Mushroom body ablation impairs short-term memory

and long-term memory of courtship conditioning in *Drosophila melanogaster*. *Neuron* **24**, 967-977.

McGuire, S. E., Deshazer, M. and Davis, R. L. (2005). Thirty years of olfactory learning and memory research in *Drosophila melanogaster*. *Prog Neurobiol* **76**, 328-47.

McGuire, S. E., Le, P. T., Osborn, A. J., Matsumoto, K. and Davis, R. L. (2003). Spatiotemporal rescue of memory dysfunction in *Drosophila*. *Science (New York, N.Y.)* **302**, 1765-1768.

Momohara, Y., Minami, H., Kanai, A. and Nagayama, T. (2016). Role of cAMP signalling in winner and loser effects in crayfish agonistic encounters. *Eur J Neurosci* **44**, 1886-95.

Moore, M. S., DeZazzo, J., Luk, A. Y., Tully, T., Singh, C. M. and Heberlein, U. (1998). Ethanol intoxication in *Drosophila*: Genetic and pharmacological evidence for regulation by the cAMP signaling pathway. *Cell* **93**, 997-1007.

Neckameyer, W. and Leal, S. (2017). Diverse Functions of Insect Biogenic Amines as Neurotransmitters, Neuromodulators, and Neurohormones. In *Hormones, Brain and Behavior*, pp. 367-401: Elsevier

Neuser, K., Triphan, T., Mronz, M., Poeck, B. and Strauss, R. (2008). Analysis of a spatial orientation memory in *Drosophila*. *Nature* **453**, 1244-1247.

Pan, Y., Zhou, Y., Guo, C., Gong, H., Gong, Z. and Liu, L. (2009). Differential roles of the fan-shaped body and the ellipsoid body in *Drosophila* visual pattern memory. *Learn Mem* **16**, 289-95.

Panksepp, J. (2003). Neuroscience. Feeling the pain of social loss. *Science* **302**, 237-9.

Penn, J. K., Zito, M. F. and Kravitz, E. A. (2010). A single social defeat reduces aggression in a highly aggressive strain of *Drosophila*. *Proc Natl Acad Sci USA* **107**, 12682-6.

Putz, G. and Heisenberg, M. (2002). Memories in *drosophila* heat-box learning. *Learn Mem* **9**, 349-359.

Qin, H., Cressy, M., Li, W., Coravos, J. S., Izzi, S. A. and Dubnau, J. (2012). Gamma neurons mediate dopaminergic input during aversive olfactory memory formation in *Drosophila*. *Curr Biol* **22**, 608-14.

Ramin, M., Domocos, C., Slawaska-Eng, D. and Rao, Y. (2014). Aggression
and social experience: genetic analysis of visual circuit activity in the control of
aggressiveness in *Drosophila*. *Mol Brain* **7**, 55.

Rillich, J. and Stevenson, P. A. (2014). A fighter's comeback: dopamine is
necessary for recovery of aggression after social defeat in crickets. *Hormones and*
behavior **66**, 696-704.

Rillich, J. and Stevenson, P. A. (2015). Releasing stimuli and aggression in
crickets: octopamine promotes escalation and maintenance but not initiation. *Front*
Behav Neurosci **9**.

Rollmann, S. M., Zwarts, L., Edwards, A. C., Yamamoto, A., Callaerts, P.,
Norga, K., Mackay, T. F. and Anholt, R. R. (2008). Pleiotropic effects of *Drosophila*
neuralized on complex behaviors and brain structure. *Genetics* **179**, 1327-36.

Rosay, P., Armstrong, J. D., Wang, Z. and Kaiser, K. (2001). Synchronized
neural activity in the *Drosophila* memory centers and its modulation by amnesiac.
Neuron **30**, 759-70.

Siegel, R. W. and Hall, J. C. (1979). Conditioned responses in courtship
behavior of normal and mutant *Drosophila*. *Proc Natl Acad Sci USA* **76**, 3430-4.

Sitnik, N., Tokmacheva, E. and Savvateeva-Popova, E. (2003). The ability of
Drosophila mutants with defects in the central complex and mushroom bodies to learn
and form memories. *Neurosci Behav Physiol* **33**, 67-71.

Stevenson, P. A., Dyakonova, V., Rillich, J. and Schildberger, K. (2005).
Octopamine and experience-dependent modulation of aggression in crickets. *J.*
Neurosci. **25**, 1431-1441.

Tamura, T., Chiang, A. S., Ito, N., Liu, H. P., Horiuchi, J., Tully, T. and Saitoe,
M. (2003). Aging specifically impairs amnesiac-dependent memory in *Drosophila*.
Neuron **40**, 1003-11.

Tan, Y., Yu, D., Pletting, J. and Davis, R. L. (2010). Gilgamesh Is Required for
rutabaga-Independent Olfactory Learning in *Drosophila*. *Neuron* **67**, 810-820.

Torroja, L., Chu, H., Kotovsky, I. and White, K. (1999). Neuronal
overexpression of APPL, the *Drosophila* homologue of the amyloid precursor protein
(APP), disrupts axonal transport. *Curr Biol* **9**, 489-92.

Trannoy, S. and Kravitz, E. A. (2016). Strategy changes in subsequent fights as consequences of winning and losing in fruit fly fights. *Fly*, 0.

Trannoy, S., Penn, J., Lucey, K., Popovic, D. and Kravitz, E. A. (2016). Short and long-lasting behavioral consequences of agonistic encounters between male *Drosophila melanogaster*. *Proc Natl Acad Sci USA* **113**, 4818-23.

Wang, L. and Anderson, D. J. (2010). Identification of an aggression-promoting pheromone and its receptor neurons in *Drosophila*. *Nature* **463**, 227-31.

Wang, L., Dankert, H., Perona, P. and Anderson, D. (2008). A common genetic target for environmental and heritable influences on aggressiveness in *Drosophila*. *Proc Natl Acad Sci USA* **105**, 5657-5663.

Wang, L., Han, X., Mehren, J., Hiroi, M., Billeter, J. C., Miyamoto, T., Amrein, H., Levine, J. D. and Anderson, D. J. (2011). Hierarchical chemosensory regulation of male-male social interactions in *Drosophila*. *Nat Neurosci* **14**, 757-62.

Yoon, J., Matsuo, E., Yamada, D., Mizuno, H., Morimoto, T., Miyakawa, H., Kinoshita, S., Ishimoto, H. and Kamikouchi, A. (2013). Selectivity and plasticity in a sound-evoked male-male interaction in *Drosophila*. *PLoS One* **8**, e74289.

Yu, D., Akalal, D. B. and Davis, R. L. (2006). *Drosophila* alpha/beta mushroom body neurons form a branch-specific, long-term cellular memory trace after spaced olfactory conditioning. *Neuron* **52**, 845-55.

Yuan, Q., Song, Y., Yang, C. H., Jan, L. Y. and Jan, Y. N. (2014). Female contact modulates male aggression via a sexually dimorphic GABAergic circuit in *Drosophila*. *Nat Neurosci* **17**, 81-8.

Yurkovic, A., Wang, O., Basu, A. C. and Kravitz, E. A. (2006). Learning and memory associated with aggression in *Drosophila melanogaster*. *Proc Natl Acad Sci USA* **103**, 17519-24.

Zars, T., Fischer, M., Schulz, R. and Heisenberg, M. (2000a). Localization of a short-term memory in *Drosophila*. *Science* **288**, 672-5.

Zars, T., Wolf, R., Davis, R. and Heisenberg, M. (2000b). Tissue-specific expression of a type I adenylyl cyclase rescues the rutabaga mutant memory defect: in search of the engram. *Learn Mem* **7**, 18-31.

Zhou, C. and Rao, Y. (2008). A subset of octopaminergic neurons are important for *Drosophila* aggression. *Nat Neurosci* **11**, 1059-67.

1   **Zwarts, L., Versteven, M. and Callaerts, P.** (2012). Genetics and neurobiology
2   of aggression in *Drosophila*. *Fly* **6**, 35-48.
3

FIGURE LEGENDS

Figure 1. cAMP signaling is required for establishing dominance relationships.

(*A*) CS and *amn*^{C651} flies form stable dominance relationships (win/loss) in 85% and

82% in their first fights, respectively. *rut*^{2080} displayed no dominance structures while

dnc^{1} showed dominance in 67% of their fights. (Two-tailed Chi-square test; ** $P < 0.01$,

*** $P < 0.001$, n=30). *, indicates comparison with CS. (*B*) CS winner flies progressively

increase lunging. A similar trend was seen with *amn*^{C651} flies ($P > 0.05$). In contrast,

rut^{2080} ($P < 0.001$) and *dnc*^{1} ($P < 0.01$) flies demonstrated significantly deficient lunging

trends. (Two factor repeated measures ANOVA followed by post hoc Tukey's multiple

comparison test, *'P'* is the interaction term, mean ± SEM, n=30). (*C*) Both CS and

amn^{C651} loser flies ($P > 0.05$) showed progressive increase in retreats while this was

significantly compromised in *rut*^{2080} ($P < 0.001$) and *dnc*^{1} ($P < 0.01$).

dnc^{1} mutants, although compromised in modifying fighting strategies compared to CS

flies, demonstrate better trends compared to *rut*^{2080} mutants ($P < 0.001$ for lunges; $P <$

0.01 for retreats) (Two factor repeated measures ANOVA followed by post hoc Tukey's

multiple comparison test, *'P'* is the interaction term, mean ± SEM, n=30).

Total number of lunges/retreats in every three successive encounters is analyzed and

their mean is reported. SEM: Standard error of the mean.

Figure 2. Aggression levels in cAMP pathway mutant flies.

(*A*) Locomotor activity in flies was analyzed using negative geotaxis assay. No

significant differences were observed between CS and cAMP pathway mutants in their

average pass rate (One factor ANOVA followed by post hoc Tukey's multiple

comparison test, mean ± SEM, n=30). (*B*) Encounters per minute is lower in rut^{2080} flies

(*P* < 0.05) but is comparable in amn^{c651} and dnc^{1} flies compared to CS flies (*P* > 0.05)

(One factor ANOVA followed by post hoc Tukey's multiple comparison test, mean ±

SEM n=30). *, indicates comparison with CS. (*C*) Aggression vigor index, defined as the

fraction of time spent fighting in first 10 minutes, is significantly lower in rut^{2080} (*P* <

0.001), amn^{c651} (*P* < 0.05) and dnc^{1} (*P* < 0.05) flies in comparison to CS flies (One

factor ANOVA followed by post hoc Tukey's multiple comparison test, mean ± SEM

n=30). *, indicates comparison with CS. (*D*) Latency to engage in an encounter is not

significantly different in rut^{2080} (*P* > 0.05), dnc^{1} (*P* > 0.05) and amn^{c651} (*P* > 0.05) flies in

comparison to CS flies. (One factor ANOVA followed by post hoc Tukey's multiple

comparison test, mean ± SEM ,n=30). SEM: Standard error of the mean.

Figure 3. rut^{2080} flies display improved lunging/retreating behavior against wild type

opponents.

(*A*) The rut^{2080} flies show attenuated dominance structures against a wild type opponent

compared to CS v CS fights (*P* < 0.001) but improved hierarchical relationships

compared to rut^{2080} v rut^{2080} fights (*P* < 0.01) (Two-tailed Chi-square test; *** *P* < 0.001,

++ *P* < 0.01, n=25). *, indicates comparison with CS and +, with rut^{2080}. (*B*) and (*C*) The

rut^{2080} flies against a wild type opponent demonstrate significantly better status-

dependent modification of lunges/retreats compared to rut^{2080} v rut^{2080} fights (*P* < 0.001

for lunges and *P* < 0.01 for retreats) and is comparable to dnc^{1} flies (*P* > 0.05). The

lunge/retreat trends for rut^{2080} flies in rut^{2080} v CS fights are compromised compared to

CS flies ($P < 0.001$ for lunges; $P < 0.05$ for retreats). (Two factor repeated measures

ANOVA followed by post hoc Tukey's multiple comparison test, 'P' is the interaction

term, mean ± SEM, n=20). (*D*) Aggression vigor index is significantly lower in flies

engaged in rut^{2080} v rut^{2080} ($P < 0.001$) and CS v rut^{2080} ($P < 0.001$) fights in comparison

to flies in CS v CS fights (One factor ANOVA followed by post hoc Tukey's multiple

comparison test, mean ± SEM n=30). *, indicates comparison with flies in CS v CS

fights.

The data set for control CS flies, which were trained at the same time of day as

experimental lines, is the same as shown in Fig 1 and 2. Total number of

lunges/retreats in every three successive encounters is analyzed and their mean is

reported. SEM: Standard error of the mean

Figure 4. Distinct neuronal circuits mediate behavioral plasticity during agnostic

encounters.

Restricted expression of *Rut* in specific sub-population of neurons in rut^{2080} flies yields

differential rescue of learning impairment in first fights. (*A*) Lack of development of

hierarchies in rut^{2080} flies is rescued upon expression of UAS-Rut (in a rut^{2080}

background) pan-neuronally (Appl), in the α/β and γ lobes of the MB (c309), α/β lobes of

the MB (c739), γ lobes of the MB (201Y), α'/β' lobes of the MB (c305a) and in the F5

neurons of the FB (c205). No rescue was seen upon reintroduction of *Rut* to the EB

(c819). (Two-tailed Chi-square test; ** $P < 0.01$, *** $P < 0.001$, +++ $P < 0.001$, n≥15). *,

indicates comparison with CS and **+**, with *rut²⁰⁸⁰*; UAS-Rut/+. (*B*) - (*G*) Ability to

progressively increase lunges/retreats was rescued (relative to *rut²⁰⁸⁰* flies) by

expressing *Rut* either pan-neuronally (Appl; $P < 0.001$ for lunges and retreats) or in α/β

and γ (c309; $P < 0.001$ for lunges and retreats), α/β alone (c739; $P < 0.001$ for lunges, P

< 0.01 for retreats), α'/β' (c305a; $P < 0.01$ for lunges, $P < 0.05$ for retreats) alone, γ

(201y; $P < 0.001$ for lunges, $P < 0.05$ for retreats) alone and in the FB (c205; $P < 0.001$

for lunges, $P < 0.01$ for retreats). EB (c819; $P > 0.05$ for lunges and retreats) expression

did not show any rescue. (*B*) and (*C*) are comparisons of *Appl*-Gal4 with CS and no

Gal4 control, (*D*) and (*E*) are comparisons of MB restricted Gal4 drivers (c309, c739,

201y and c305a) with no Gal4 control and (*F*) and (*G*) comparisons are of central

complex restricted Gal4 drivers (c205 and c819) with no Gal4 control (Two factor

repeated measures ANOVA followed by post hoc Tukey's multiple comparison test, *'P'*

is the interaction term, mean ± SEM, n≥15).

The data set for control CS flies, which were trained at the same time of day as

experimental lines, is the same as shown in Fig 1. Total number of lunges/retreats in

every three successive encounters is analyzed and their mean is reported. SEM:

Standard error of the mean

Figure 5. Aggression levels are unchanged upon rescue of aggression-associated

learning.

(*A*) In comparison to *rut²⁰⁸⁰*;UAS-Rut/+ control flies, *Rut* expression in α/β and γ lobes of

the MB (c309; $P > 0.05$) or pan-neuronally (Appl; $P > 0.05$) resulted in a comparable

number of encounter frequencies (One factor ANOVA followed by post hoc Dunnett's

multiple comparison test, mean ± SEM, n=20). (*B*) There is no significant improvement,

as compared to *rut²⁰⁸⁰*;UAS-Rut/+ control flies, in aggression vigor index in response to

Rut expression either pan-neuronally or in the α/β and γ lobes (*P* > 0.05; One factor

ANOVA followed by post hoc Dunnett's multiple comparison test, mean ± SEM, n=20).

(*C*) Latency to engage in an encounter (*P* > 0.05) remains comparable upon *Rut*

expression using either Appl or c309 Gal4 driver lines in comparison to *rut²⁰⁸⁰*;UAS-

Rut/+ control flies (One factor ANOVA followed by post hoc Dunnett's multiple

comparison test, mean ± SEM, n=20).

SEM: Standard error of the mean.

Figure 6. cAMP signaling is necessary for developing loser mentality.

(*A*) CS losers lost all their second fights against naïve opponents while *amn^C651* flies lost

and won equal number of fights. *dnc¹* flies show weak dominance relationships and

their interactions largely ended in draws, again suggestive of memory defects. All

previously experienced *rut²⁰⁸⁰* flies engaged in draws. (Two-tailed Chi-square test; *** *P*

< 0.001, n≥20). *, indicates comparison with CS. (*B*) Loser index assesses the

experience dependent modulation of fighting strategies of experienced flies in their

second fights against a naïve opponent. *rut²⁰⁸⁰*, *dnc¹* and *amn^C651* flies, all showed

severely compromised loser indices compared to CS. (One factor ANOVA followed by

post hoc Dunnett's multiple comparison test, *** *P* < 0.001, mean ± SEM, n≥20). *,

indicates comparison with CS.

Figure 7. Recruitment of specific neuronal circuits for the development of the loser

effect.

(*A*) Rescue experiments expressing *Rut* (in a *rut^{2080}* background) either pan-neuronally

(Appl) or in the α/β and γ lobes of the MB (c309) showed a strong recovery of the loser

effect. Expression in the α/β lobes (c739), γ lobes (201Y), α'/β' lobes (c305a) and in the

FB (c205) of *rut* mutants resulted in partial recovery of dominance relationships. No

rescue was seen upon the reintroduction of *Rut* in the EB (c819) of *rut^{2080}*. (Two-tailed

Chi-square test; *** $P < 0.001$, $^{+++}$ $P < 0.001$ n≥15). *, indicates comparison with CS and

+, with *rut^{2080}*; UAS-Rut/+. (*B*) Loser index analysis revealed that pan-neuronal and α/β

and γ lobe combined expression is able to rescue the loser effect. All other

combinations did not show any rescue. (One factor ANOVA followed by post hoc

Tukey's multiple comparison test, *** $P < 0.001$, $^{+++}$ $P < 0.001$, mean ± SEM, n≥15).

*, indicates comparison with CS and **+**, with *rut^{2080}*; UAS-Rut/+. SEM: Standard error of

the mean. The data set for control CS flies, which were trained at the same time of day

as experimental lines, is the same as shown in Fig 6.

Table I: Expression patterns of Gal4 drivers used and rescue results

Gal4 drivers	Expression pattern	Rescue of learning (1st fight)	Rescue of loser effect (2nd fight)
Appl	Pan-neuronal (Neuser et al., 2008; Torroja et al., 1999)	Winners: Y* Losers: Y*	Y*
c309	αβ and γ lobes of the MB (Aso et al., 2009; Blum et al., 2009; Joiner and Griffith, 1999)	Winners: Y* Losers: Y*	Y*
c739	αβ lobes of the MB (Aso et al., 2009; Blum et al., 2009; Joiner and Griffith, 1999; Zars et al., 2000b)	Winners: Y* Losers: Y*	N
201Y	γ lobes and αβ lobes (sparse) of the MB (Aso et al., 2009; Blum et al., 2009; Joiner and Griffith, 1999; Zars et al., 2000a)	Winners: $Y^\dagger$ Losers: Y*	N
c305a	50% of α'β' lobes of the MB; weak expression in AL and EB (Aso et al., 2009; Blum et al., 2009)	Winners: $Y^\dagger$ Losers: Y*	N
c205	F5 neurons of the FB (Liu et al., 2006; Pan et al., 2009)	Winners: Y* Losers: Y*	N
c819	R2/R4m neurons of the EB (Liu et al., 2006; Pan et al., 2009)	Winners: N Losers: N	N

Y: yes; N: no; MB: Mushroom bodies; EB: Ellipsoid body; FB: Fan shaped body; AL: Antennal lobes
(*, $P<0.001$; †, $P<0.01$; Indices were compared between rut^{2080}/Y; UAS-rut/+ and rut^{2080}/Y; UAS-rut/Gal 4).

A

B

C

Figure 1

Figure 2

Figure 3

Figure 4

Figure 5

Figure 6

A

			+++	+++	+++	+++	+++	+++	ns
	***	***	ns	ns	***	***	***	***	***

Rut	+	-	-	-	-	-	-	-	-	-
UAS-Rut	-	+	-	+	+	+	+	+	+	+
Gal4	-	-	Appl	Appl	c309	c739	201y	c305a	c205	c819

B

			+++	+++	ns	ns	ns	ns	ns
***	***	ns	ns	***	***	***	***	***

Rut	+	-	-	-	-	-	-	-	-	-
UAS-Rut	-	+	-	+	+	+	+	+	+	+
Gal4	-	-	Appl	Appl	c309	c739	201y	c305a	c205	c819

Figure 7