

Extending R^2 and intra-class correlation coefficient from generalized linear mixed-effects models: capturing and characterizing biological variation

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1 **Abstract**

2 The coefficient of determination R^2 quantifies the proportion of variance explained by a statistical
3 model and is an important summary statistic of biological interest. However, estimating R^2 for
4 (generalized) linear mixed models (GLMMs) remains challenging. We have previously introduced
5 a version of R^2 that we called R^2_{GLMM} for Poisson and binomial GLMMs using biological examples,
6 but not for other distributional families. Similarly, we earlier discussed how to estimate intra-class
7 correlation coefficients ICC using only Poisson and binomial GLMMs. In this article, we expand
8 our methods to all the other non-Gaussian distributions such as negative binomial and gamma
9 distributions, which are common in biological data. While expanding our approach, we highlight
10 two useful concepts for biologists, Jensen's inequality and the delta method, both of which help us
11 in understanding the properties of GLMMs. Jensen's inequality has important implications for
12 biologically more meaningful interpretation of GLMMs, while the delta method allows a general
13 derivation of distribution-specific variances. We also discuss some special considerations for
14 binomial GLMMs with binary or proportion data. We illustrate the implementation of our extension
15 by worked examples from the field of ecology and evolution in the R environment although our
16 method can be used regardless of statistical environments.

17 **Key words:** *repeatability, regression, heritability, goodness of fit, information criteria,*
18 *variance explained, model fit, variance decomposition, reliability analysis.*

19

20 **1. Introduction**

21 One of the main purposes of linear modelling is to understand the sources of variation in biological
22 data. In this context, it is not surprising that the coefficient of determination R^2 is a commonly
23 reported statistic because it represents the proportion of variance explained by a linear model. The
24 intra-class correlation coefficient ICC is a related statistic that quantifies the proportion of variance
25 explained by a grouping (random) factor in multilevel/hierarchical data. In the field of ecology and
26 evolution, a type of ICC is often referred to as repeatability R , where the grouping factor is often
27 individuals that have been phenotyped repeatedly [1, 2]. We have reviewed methods for estimating
28 R^2 and ICC in the past, with a particular focus on non-Gaussian response variables in the context of
29 biological data [2, 3]. These previous articles featured generalized linear mixed-effects models
30 (GLMMs) as the most versatile engine for estimating R^2 and ICC (specifically R^2_{GLMM} and
31 ICC_{GLMM}). Our descriptions were limited to random-intercept GLMMs, but Johnson [4] has
32 recently extended the methods to random-slope GLMMs, widening the applicability of these
33 statistics (see also, [5, 6]).

34 However, at least one important issue seems to remain. Currently these two statistics are only
35 described for binomial and Poisson GLMMs. Although these two types of GLMMs are arguably the
36 most popular [7], there are other common families of distributions in biology, such as negative
37 binomial and gamma distributions [8, 9]. In this article, we revisit and extend R^2_{GLMM} and ICC_{GLMM}
38 to more distributional families with a particular focus on negative binomial and gamma
39 distributions. In this context, we discuss Jensen's inequality and two variants of the delta method,
40 which are rarely known among biologists. However, these concepts are useful not only for
41 generalizing our previous methods, but also for interpreting the results of GLMMs for biologists.
42 Furthermore, we refer to some special considerations when obtaining R^2_{GLMM} and ICC_{GLMM} from
43 binomially GLMMs for binary and proportion data, which we did not discuss in the past [2, 3]. We
44 provide worked examples inspired from the field of ecology and evolution, focusing on

45 implementation in the R environment [10] and finish by referring to two alternative approaches for
46 obtaining R^2 and ICC from GLMMs along with a cautionary note.

47 **2. Definitions of R^2_{GLMM} , ICC_{GLMM} and overdispersion**

48 To start with, we present R^2_{GLMM} and ICC_{GLMM} for a simple case of Gaussian error distributions
49 based on a linear mixed-effects model (LMM, hence also referred to as R^2_{LMM} and ICC_{LMM}).

50 Imagine a two-level dataset where the first level corresponds to observations and the second level to
51 some grouping factor (e.g. individuals) with k fixed effect covariates. The model can be written as
52 (model 1):

$$53 \quad y_{ij} = \beta_0 + \sum_{h=1}^k \beta_h x_{hij} + \alpha_i + \varepsilon_{ij}, \quad (2.1)$$

$$54 \quad \alpha_i \sim \text{Gaussian}(0, \sigma_\alpha^2), \quad (2.2)$$

$$55 \quad \varepsilon_{ij} \sim \text{Gaussian}(0, \sigma_\varepsilon^2), \quad (2.3)$$

56 where y_{ij} is the j th observation of the i th individual, x_{hij} is the j th value of the i th individual for the
57 h th of k fixed effects predictors, β_0 is the (grand) intercept, β_h is the regression coefficient for the
58 h th predictor, α_i is an individual-specific effect, assumed to be normally distributed in the
59 population with the mean and variance of 0 and σ_α^2 , ε_{ij} is an observation-specific residual, assumed
60 to be normally distributed in the population with mean and variance of 0 and σ_ε^2 , respectively. For
61 this model, we can define two types of R^2 as:

$$62 \quad R^2_{\text{LMM}(m)} = \frac{\sigma_f^2}{\sigma_f^2 + \sigma_\alpha^2 + \sigma_\varepsilon^2}, \quad (2.4)$$

$$63 \quad R^2_{\text{LMM}(c)} = \frac{\sigma_f^2 + \sigma_\alpha^2}{\sigma_f^2 + \sigma_\alpha^2 + \sigma_\varepsilon^2}, \quad (2.5)$$

$$64 \quad \sigma_f^2 = \text{var}\left(\sum_h^k \beta_h x_{hij}\right), \quad (2.6)$$

65 where $R_{LMM(m)}^2$ represents the marginal R^2 , which is the variance accounted for by the fixed effects,
66 $R_{LMM(c)}^2$ represents the conditional R^2 , which is the variance explained by both fixed and random
67 effects, and σ_f^2 is the variance explained by fixed effects [11, 12]. Since marginal and conditional
68 R^2 differ only in whether the random effect variance is included in the numerator, we avoid
69 redundancy and present equations only for marginal R^2 in the following.

70 Similarly, there are two types of ICC:

$$71 \quad ICC_{LMM(adj)} = \frac{\sigma_\alpha^2}{\sigma_\alpha^2 + \sigma_\varepsilon^2} \quad (2.7)$$

$$72 \quad ICC_{LMM} = \frac{\sigma_\alpha^2}{\sigma_\alpha^2 + \sigma_f^2 + \sigma_\varepsilon^2} \quad (2.8)$$

73 If no fixed effects are included, the two versions are identical and represent unadjusted ICC, but if
74 fixed effects are fitted, $ICC_{LMM(adj)}$ represents adjusted ICC, while ICC_{LMM} represented unadjusted
75 ICC (*sensu* [2]). Since the two versions of ICC differ only in whether the fixed effect variance,
76 calculated as in equation (2.6), is included in the denominator, we avoid redundancy and present
77 equations only for adjusted ICC in the following.

78 One of the main difficulties in extending R^2 from LMMs to GLMMs is defining the residual
79 variance σ_ε^2 . For binomial and Poisson GLMMs with an additive dispersion terms, we have
80 previously stated that σ_ε^2 is equivalent to $\sigma_e^2 + \sigma_d^2$ where σ_e^2 is the variance for the additive
81 overdispersion term, and σ_d^2 is the distribution-specific variance [2, 3]. Here overdispersion
82 represents the excess variation relative to what is expected from a certain distribution and can be
83 estimated by fitting an observation-level random effect (OLRE; see, [13, 14]). Alternatively,
84 overdispersion in GLMMs can be implemented using a multiplicative overdispersion term [15]. In
85 such an implementation, we stated that σ_ε^2 is equivalent to $\omega \cdot \sigma_d^2$ where ω is a multiplicative
86 dispersion parameter estimated from the model [2]. However, obtaining σ_d^2 for specific
87 distributions is not always possible, because in many families of GLMMs the parameters are less

88 clearly separated into a parameter for the expectation of the mean and a parameter for the
89 (over)dispersion. It turns out that binomial and Poisson distributions are special cases where σ_d^2 can
90 be usefully calculated, because either all overdispersion is modelled by an OLRE (additive
91 overdispersion) or by a single multiplicative overdispersion parameter (multiplicative
92 overdispersion). However, as we will show below, we can always obtain the GLMM version of σ_ϵ^2
93 (on the latent scale) directly. We refer to this generalised version of σ_ϵ^2 as ‘the observation-level
94 variance’ here rather than the residual variance (but we keep the notation σ_ϵ^2).

95 **3. Extension of R^2_{GLMM} and ICC_{GLMM}**

96 We now define R^2_{GLMM} and ICC_{GLMM} for an overdispersed Poisson (also known as quasi-Poisson)
97 GLMM, because the overdispersed Poisson distribution is similar to the negative binomial
98 distribution at least in their uses[9, 16]. Imagine count data repeatedly measured from a number of
99 individuals with associated data on k covariates. We fit an overdispersed Poisson (OP) GLMM with
100 the log link function (model 2):

$$101 \quad y_{ij} \sim \text{OP}(\lambda_{ij}, \omega), \quad (3.1)$$

$$102 \quad \ln(\lambda_{ij}) = \beta_0 + \sum_{h=1}^k \beta_h x_{hij} + \alpha_i, \quad (3.2)$$

$$103 \quad \alpha_i \sim \text{Gaussian}(0, \sigma_\alpha^2), \quad (3.3)$$

104 where y_{ij} is the j th observation of the i th individual and y_{ij} follows an overdispersed Poisson
105 distribution with two parameters, λ_{ij} and ω , $\ln(\lambda_{ij})$ is the latent value for the j th observation of the i th
106 individual, ω is the overdispersion parameter (when the multiplicative dispersion parameter ω is 1,
107 the model becomes a standard Poisson GLMM), α_i is an individual-specific effect, assumed to be
108 normally distributed in the population with the mean and variance of 0 and σ_α^2 , respectively (as in
109 model 1), and the other symbols are the same as above. For such a model, we can define $R^2_{\text{GLMM}(m)}$
110 and (adjusted) ICC_{GLMM} as:

$$111 \quad R_{\text{OP-In}(m)}^2 = \frac{\sigma_f^2}{\sigma_f^2 + \sigma_\alpha^2 + \ln(1 + \omega / \lambda)}, \quad (3.4)$$

$$112 \quad \text{ICC}_{\text{OP-In}} = \frac{\sigma_\alpha^2}{\sigma_\alpha^2 + \ln(1 + \omega / \lambda)}, \quad (3.5)$$

113 where the subscript of R^2 and ICC denote the distributional family, here OP-In for overdispersed
 114 Poisson distribution with log link, the term $\ln(1 + \omega / \lambda)$ corresponds to the observation-level
 115 variance σ_ε^2 (Table 1, for derivation see Appendix S1), ω is the overdispersion parameter, and λ is
 116 the mean value of λ_{ij} . We discuss how to obtain λ below.

117 The calculation is very similar for a negative binomial (NB) GLMM with the log link (model 3):

$$118 \quad y_{ij} \sim \text{NB}(\lambda_{ij}, \theta), \quad (3.6)$$

$$119 \quad \ln(\lambda_{ij}) = \beta_0 + \sum_{h=1}^k \beta_h x_{hij} + \alpha_i, \quad (3.7)$$

$$120 \quad \alpha_i \sim \text{Gaussian}(0, \sigma_\alpha^2), \quad (3.8)$$

121 where y_{ij} is the j th observation of the i th individual and y_{ij} follows a negative binomial distribution
 122 with two parameters, λ_{ij} and θ , where θ is the shape parameter of the negative binomial distribution
 123 (given by the software often as the dispersion parameter), and the other symbols are the same as
 124 above. $R_{\text{GLMM}(m)}^2$ and (adjusted) ICC_{GLMM} for this model can be calculated as:

$$125 \quad R_{\text{NB-In}(m)}^2 = \frac{\sigma_f^2}{\sigma_f^2 + \sigma_\alpha^2 + \ln(1 + 1 / \lambda + 1 / \theta)}, \quad (3.9)$$

$$126 \quad \text{ICC}_{\text{NB-In}} = \frac{\sigma_\alpha^2}{\sigma_\alpha^2 + \ln(1 + 1 / \lambda + 1 / \theta)}, \quad (3.10)$$

127 Finally, for a gamma GLMM with the log link (model 4):

$$128 \quad y_{ij} \sim \text{gamma}(\lambda_{ij}, \nu), \quad (3.11)$$

$$129 \quad \ln(\lambda_{ij}) = \beta_0 + \sum_{h=1}^k \beta_h x_{hij} + \alpha_i, \quad (3.12)$$

$$130 \quad \alpha_i \sim \text{Gaussian}(0, \sigma_\alpha^2), \quad (3.13)$$

131 where y_{ij} is the j th observation of the i th individual and y_{ij} follows a gamma distribution with two
132 parameters, λ_{ij} and ν , where ν is the shape parameter of the gamma distribution (sometimes
133 statistical programs report $1/\nu$ instead of ν ; also note that the gamma distribution can be
134 parameterized in alternative ways, Table 1), $R^2_{\text{GLMM}(m)}$ and (adjusted) ICC_{GLMM} can be calculated
135 as:

$$136 \quad R^2_{\text{gamma-ln}(m)} = \frac{\sigma_f^2}{\sigma_f^2 + \sigma_\alpha^2 + \ln(1 + 1/\nu)}, \quad (3.15)$$

$$137 \quad \text{ICC}_{\text{gamma-ln}} = \frac{\sigma_\alpha^2}{\sigma_\alpha^2 + \ln(1 + 1/\nu)}, \quad (3.16)$$

138 **4. Obtaining the observation-level variance by the ‘first’ delta method**

139 For overdispersed Poisson, negative binomial and gamma GLMMs with log link, the observation-
140 level variance σ_e^2 can be obtained via the variance of the log-normal distribution, as described
141 above (see Appendix S1). There are two more alternative methods to obtain the same target: the
142 delta method and the trigamma function. The two alternatives have different advantages and will be
143 discussed in some detail below.

144 The delta method for variance approximation uses a first order Taylor series expansion, which is
145 often employed to approximate the standard error (error variance) for transformations (or functions)
146 of a variable x when the (error) variance of x itself is known (see [17]; for an accessible reference
147 for biologists, [18]). A simple case of the delta method for variance approximation can be written
148 as:

$$149 \quad \text{var}[f(x)] \approx \text{var}[x] \left(\frac{d}{dx} f(x) \right)^2, \quad (4.1)$$

150 where x is a random variable (typically represented by observations), f represents a function (e.g.
151 log or square-root), var denotes variance, and d/dx is a (first) derivative with respect to variable x .

152 Taking derivatives of any function can be easily done using the R environment (examples can be
153 found in the Appendices). It is the delta method that Foulley and colleagues [19] used to derive the
154 distribution specific variance σ_d^2 for Poisson GLMMs as $1/\lambda$: Given that $\text{var}[\lambda_{ij}] = \lambda$ in Poisson
155 distributions and $d \ln(\lambda) / dx = 1/\lambda$, it follows that $\text{var}[\ln(\lambda_{ij})] \approx \lambda(1/\lambda)^2$ (note that for Poisson
156 distributions without overdispersion, σ_d^2 is equal to σ_ε^2 because $\sigma_\varepsilon^2 = 0$). One clear advantage of
157 the delta method is its flexibility, and we can easily obtain the observation-level variance σ_ε^2 for all
158 kinds of distributions/link functions. For example, by using the delta method, it is straightforward to
159 obtain σ_ε^2 for the Tweedie (compound Poisson-gamma) distribution, which has been used to model
160 non-negative real numbers in ecology (e.g., [20, 21]). For the Tweedie distribution, the variance on
161 the observed scale has the relationship $\text{var}[y] = \phi\mu^p$ where μ is the mean on the observed scale and
162 ϕ is the dispersion parameter, comparable to λ and ω in equation (3.1), and p is a positive constant
163 called an index parameter. Therefore, when used with the log-link function, an approximated σ_ε^2
164 value can be obtained by $\phi\mu^{(p-2)}$ according to equation (4.1). The log-normal approximation
165 $\ln(1 + \phi\mu^{(p-2)})$ is also possible (see Appendix S1; cf. Table 1).

166 The use of the trigamma function ψ_1 is limited to distributions with log link, but it should provide
167 the most accurate estimate of the observation level variance σ_ε^2 . This is because the variance of a
168 gamma-distributed variable on the log scale is equal to $\psi_1(\nu)$ where ν is the shape parameter of the
169 gamma distribution [22] and hence σ_ε^2 is $\psi_1(\nu)$. At the level of the statistical parameters (Table 1;
170 on the ‘expected data’ scale; *sensu* [23]; see their Figure 1), Poisson and negative binomial
171 distributions can be both seen special cases of gamma distributions, and σ_ε^2 can be obtained using
172 the trigamma function (Table 1). For example, σ_ε^2 for the Poisson distribution is $\psi_1(\lambda)$ with the
173 speciality that in the case of Poisson distributions $\sigma_\varepsilon^2 = \sigma_d^2$. As we show in Appendix S2, $\ln(1+1/\lambda)$
174 (log-normal approximation), $1/\lambda$ (delta method approximation) and $\psi_1(\lambda)$ (trigamma function) are

175 similar if λ is greater than 2. Nonetheless, our recommendation is to use the trigamma function for
 176 obtaining σ_ε^2 whenever this is possible.

177 We note that in calculations of heritability (which can be seen as a type of ICC although in a strict
 178 sense, it is not; see [23]) using negative binomial GLMMs, the trigamma function has been
 179 previously used to obtain observation-level variance ([22, 24]; cf. [23]). Table 1 summarises
 180 observation-level variance σ_ε^2 for overdispersed Poisson, negative binomial and gamma
 181 distributions for commonly used link functions.

182 **5. How to estimate λ from data**

183 Imagine a Poisson GLMM with log link and additive overdispersion fitted as an observation-level
 184 random effect (model 5):

$$185 \quad y_{ij} \sim \text{Poisson}(\lambda_{ij}), \quad (5.1)$$

$$186 \quad \ln(\lambda_{ij}) = \beta_0 + \sum_{h=1}^p \beta_h x_{hij} + \alpha_i + e_{ij}, \quad (5.2)$$

$$187 \quad \alpha_i \sim \text{Gaussian}(0, \sigma_\alpha^2), \quad (5.3)$$

$$188 \quad e_{ij} \sim \text{Gaussian}(0, \sigma_e^2), \quad (5.4)$$

189 where y_{ij} is the j th observation of the i th individual, and follows a Poisson distribution with the
 190 parameter λ_{ij} , e_{ij} is an additive overdispersion term for j th observation of the i th individual, and the
 191 other symbols are the same as above. Using the log-normal approximation $R^2_{\text{GLMM}(m)}$ and (adjusted)
 192 ICC_{GLMM} can be calculated as:

$$193 \quad R^2_{\text{P-ln}(m)} = \frac{\sigma_f^2}{\sigma_f^2 + \sigma_\alpha^2 + \sigma_e^2 + \ln(1 + 1/\lambda)}, \quad (5.5)$$

$$194 \quad \text{ICC}_{\text{P-ln}} = \frac{\sigma_\alpha^2}{\sigma_\alpha^2 + \sigma_e^2 + \ln(1 + 1/\lambda)}, \quad (5.6)$$

195 where, as mentioned above, the term $\ln(1+1/\lambda)$ is σ_ϵ^2 (or σ_d^2) for Poisson distributions with the log
196 link (Table 1).

197 In our earlier papers, we proposed to use the exponential of the intercept (from the intercept-only
198 model or models with centred fixed factors) $\exp(\beta_0)$ as an estimator of λ [2, 3]. We also suggested
199 that it is possible to use the mean of observed values y_{ij} . Unfortunately, these two recommendations
200 are often inconsistent with each other. This is because, given the model 5 (and all the models in the
201 previous section), the following relationships hold:

$$202 \quad \exp(\beta_0) \leq E[y_{ij}], \quad (5.7)$$

$$203 \quad E[\lambda_{ij}] = \exp(\beta_0 + 0.5\sigma_\tau^2), \quad (5.8)$$

$$204 \quad E[y_{ij}] = E[\lambda_{ij}], \quad (5.9)$$

205 where E represents the expected value (i.e., mean) on the observed scale, β_0 is the mean value on
206 the latent scale (i.e. β_0 from the intercept-only model), σ_τ^2 is the total variance on the latent scale
207 (e.g., $\sigma_\alpha^2 + \sigma_\epsilon^2$ in the models 1 and 5, and σ_α^2 in models 2-4[2]; see also [25]). In fact, $\exp(\beta_0)$ gives
208 the median value of y_{ij} rather than the mean of y_{ij} , assuming a Poisson distribution. Thus, the use of
209 $\exp(\beta_0)$ will often overestimate σ_d^2 , providing conservative (smaller) estimates of R^2 and ICC,
210 compared to when using averaged y_{ij} , which is a better estimate of $E[y_{ij}]$. Quantitative differences
211 between the two approaches may often be negligible, but when λ is small, the difference can be
212 substantial so the choice of the method needs to be reported for reproducibility (Appendix S2). Our
213 new recommendation is to obtain λ via equation (5.8). When sampling is balanced (i.e. observations
214 are equally distributed across individuals and covariates), equation (5.8) and the mean of the
215 observed values will give similar values, but when unbalanced, method equation (5.8) is preferable.
216 This recommendation for obtaining λ also applies to negative binomial GLMMs (see Table 1).

217 **6. Jensen's inequality and the 'second' delta method**

218 A general form of equation (5.7) is known as Jensen's inequality, $g(\bar{x}) \leq \overline{g(x)}$ where g is a convex
219 function. Hence, the transformation of the mean value is equal to or larger than the mean of
220 transformed values (the opposite is true for a concave function; that is, $g(\bar{x}) \geq \overline{g(x)}$; [26]). In fact,
221 whenever the function is not strictly linear, simple application of the inverse link function (or back-
222 transformation) cannot be used to translate the mean on the latent scale into the mean value on the
223 observed scale. This inequality has important implications for the interpretation of results from
224 GLMMs, and also generalized linear models GLMs and linear models with transformed response
225 variables.

226 Although log-link GLMMs (e.g., model 5) have an analytical formula, equation (5.8), this is not
227 usually the case. Therefore, converting the latent scale values into observation-scale values requires
228 simulation using the inverse link function. However, the delta method for bias correction can be
229 used as a general approximation to account for Jensen's inequality when using link functions or
230 transformations. This application of the delta method uses a second order Taylor series expansion
231 [17, 27]. A simple case of the delta method for bias correction can be written as:

$$232 \quad E[f(x)] \approx f(x) + 0.5\sigma_x^2 \frac{d^2}{dx^2} f(x), \quad (6.1)$$

233 where d^2/dx^2 is a second derivative with respect to the variable x and the other symbols are as in
234 equations (4.1) and (5.8). By employing this bias correction delta method (with
235 $d^2 \exp(x) / dx^2 = \exp(x)$), we can approximate equation (5.8) using the same symbols as in
236 equations (5.7)-(5.9):

$$237 \quad E[\lambda_{ij}] = E[\exp(\beta_0)] \approx \exp(\beta_0) + 0.5\sigma_\tau^2 \exp(\beta_0) \quad (6.2)$$

238 The comparison between equation (5.8) (exact) and equation (6.2) (approximate) is shown in
 239 Appendix S3. The approximation is most useful when the exact formula is not available as in the
 240 case of a binomial GLMM with logit link (model 6):

$$241 \quad y_{ij} \sim \text{binomial}(p_{ij}, n_{ij}), \quad (6.3)$$

$$242 \quad \text{logit}(p_{ij}) = \beta_0 + \sum_{h=1}^k \beta_h x_{hij} + \alpha_i + e_{ij}, \quad (6.4)$$

$$243 \quad \alpha_i \sim \text{Gaussian}(0, \sigma_\alpha^2), \quad (6.5)$$

$$244 \quad e_{ij} \sim \text{Gaussian}(0, \sigma_e^2), \quad (6.6)$$

245 where y_{ij} is the number of ‘success’ in n_{ij} trials by the i th individual at the j th occasion (for binary
 246 data, n_{ij} is always 1), p_{ij} is the underlying probability of success, and the other symbols are the same
 247 as above.

248 To obtain corresponding values between the latent scale and data (observation) scale, we need to
 249 account for Jensen’s inequality (note the logit function combines of concave and convex sections).

250 For example, the overall intercept, β_0 on the latent scale could be transformed not just with the
 251 inverse (anti) logit function ($\text{logit}^{-1}(x) = \exp(x)/(1 + \exp(x))$) but also the bias corrected
 252 approximation. For the case of the binomial GLMM, we can use this approximation below given

253 that $d^2 \text{logit}^{-1}(x) / dx^2 = \exp(x)(1 - \exp(x)) / (1 + \exp(x))^3$:

$$254 \quad E[y_{ij}] = E[\text{logit}^{-1}(\beta_0)] \approx \frac{\exp(\beta_0)}{1 + \exp(\beta_0)} + 0.5\sigma_\tau^2 \frac{\exp(\beta_0)(1 - \exp(\beta_0))}{(1 + \exp(\beta_0))^3}. \quad (6.7)$$

255 We can replace β_0 with any value obtained from the fixed part of the model (i.e. $\beta_0 + \sum \beta_h x_{hij}$).

256 Another approximation proposed by Zeger and colleagues [28] produces similar (but slightly better)
 257 estimates than equation (6.7). Using our notation, this approximation can be written as:

$$258 \quad E[p_{ij}] \approx \text{logit}^{-1} \left(\beta_0 \sqrt{1 + \left(\frac{16\sqrt{3}}{15\pi} \right)^2 \sigma_\tau^2} \right). \quad (6.8)$$

259 A comparison between equations (6.7) and (6.8) is also shown in Appendix S3. This approximation
260 uses the exact solution for the inverse probit function, which can be written for a model like model
261 6 but using the probit link: i.e., $\text{probit}(p_{ij}) = \beta_0 + \sum_{h=1}^k \beta_h x_{hij} + \alpha_i + e_{ij}$ in place of equation (6.4):

$$262 \quad E[p_{ij}] = \text{probit}^{-1} \left(\beta_0 \sqrt{1 + \sigma_\tau^2} \right). \quad (6.9)$$

263 Simulation will give the most accurate conversions when no exact solutions are available. The use
264 of the delta method for bias correction accounting for Jensen's inequity is a very general and
265 versatile approach that is applicable for any distribution with any link function (see Appendix S3)
266 and can save computation time. We note that the accuracy of the delta method (both variance
267 approximation and bias correction) depends on the form of the function f , the conditions for and
268 limitation of the delta method are described in the article by Oehlert [27].

269 **7. Special considerations for binomial GLMMs**

270 The observation-level variance σ_ϵ^2 can be thought of as being added to the latent scale on which
271 other variance components are also estimated in a GLMM (equations (3.2), (3.7), (3.12), (5.2) and
272 (6.4) for models 2-6). Since the proposed R^2_{GLMM} and ICC_{GLMM} are ratios between variance
273 components and their additive combinations, we can show using the delta method that R^2_{GLMM} and
274 ICC_{GLMM} calculated via σ_ϵ^2 approximate to those of R^2 and ICC on the observation (original) scale
275 (shown in Appendix S4). In some cases, there exist specific formulas for ICC on the observation
276 scale [2]. In the past, we distinguished between ICC on the latent scale and on the observation scale
277 [2]. Such a distinction turns out to be strictly appropriate only for binomial distributions but not for
278 Poisson distributions (and probably also not for other non-Gaussian distributions). This is because

279 the property of what we have called the distribution-specific variance σ_d^2 for binomial distributions
280 (e.g. $\pi^2/3$ for binomial error distribution with the logit link function) is quite different from what we
281 have discussed as the observation-level variance σ_e^2 although these two types of variance are
282 related conceptually (i.e., both represents variance due to non-Gaussian distributions with specific
283 link functions). Let us explain this further.

284 A binomial distribution with a mean of p (the proportion of successes) has a variance of $p(1-p)$ and
285 we find that the observation-level variance is $1/(p(1-p))$ using the delta method on the logit-link
286 function (see Table 2). This observation-level variance $1/(p(1-p))$ is clearly different from the
287 distribution-specific variance $\pi^2/3$. As with the observation-level variance for the log-Poisson model
288 (which is $1/\lambda$ and changes with λ ; note that we would have called $1/\lambda$ the distribution-specific
289 variance; [2, 3]), the observation-level variance of the binomial distribution changes as p changes
290 (see Appendix S5), suggesting these two observation-level variances ($1/\lambda$ and $1/(p(1-p))$) are
291 analogous while the distribution-specific variance $\pi^2/3$ is not. Further, the minimum value of
292 $1/(p(1-p))$ is 4, which is larger than $\pi^2/3 \approx 3.29$, meaning that the use of $1/(p(1-p))$ in R^2 and ICC will
293 always produce larger values than those using $\pi^2/3$. Consequently, Browne and colleagues [15]
294 showed that ICC values (or variance partition coefficients, VPCs) estimated using $\pi^2/3$ were higher
295 than corresponding ICC values on the observation (original) scale using logistic-binomial GLMMs
296 (see also [29]). Then, what is $\pi^2/3$?

297 Three common link functions in binomial GLMMs (logit, probit and complementary log-log) all
298 have corresponding distributions on the latent scale: the logistic distribution, standard normal
299 distribution and Gumbel distribution, respectively. Each of these distributions has a theoretical
300 variance, namely, $\pi^2/3$, 1 and $\pi^2/6$, respectively (Table 2). As far as we are aware, these theoretical
301 variances only exist for binomial distributions. It is important to notice that, for example, the
302 meaning of $1/(p(1-p))$, which is the variance on the latent scale that approximates to the variance
303 due to binomial distributions on the observation scale is distinct from the meaning of $\pi^2/3$, which is

304 the variance of the latent distribution (i.e., the logistic distribution) according to which the original
305 data are theoretically distributed on the logit scale. We need distinguishing these theoretical
306 (distribution-specific) variances from the observation-level variance. Put another way, R^2 and ICC
307 values using the theoretical distribution-specific variance can rightly be called the latent (link) scale
308 (*sensu* [2]) while, as mentioned above, R^2 and ICC values using the observation-level variance
309 estimate the counterparts on the observation (original) scale (cf. [23]). The use of the theoretical
310 distribution-specific variance will almost always provide different values of R^2_{GLMM} and ICC_{GLMM}
311 from those using the observation-level obtained via the delta method (see Appendix S5). In any
312 case, we should be aware that binomial GLMMs are special cases for obtaining R^2_{GLMM} and ICC_{GLMM}
313 GLMM from binomial GLMMs.

314 **8. Worked examples: revisiting the beetles**

315 In the following, we present a worked example by expanding the beetle dataset that was generated
316 for the previous work [3]. In brief, the dataset represents a hypothetical species of beetle that has the
317 following life cycle: larvae hatch and grow in the soil until they pupate, and then adult beetles feed
318 and mate on plants. Larvae are sampled from 12 different populations ('Population'; see Figure 1).
319 Within each population, larvae are collected at two different microhabitats ('Habitat'): dry and wet
320 areas as determined by soil moisture. Larvae are exposed to two different dietary treatments
321 ('Treatment'): nutrient rich and control. The species is sexually dimorphic and can be easily sexed
322 at the pupa stage ('Sex'). Male beetles have two different color morphs: one dark and the other
323 reddish brown ('Morph', labeled as A and B in Figure 1). Sexed pupae are housed in standard
324 containers until they mature ('Container'). Each container holds eight same-sex animals from a
325 single population, but with a mix of individuals from the two habitats ($N_{\text{container}} = 120$; $N_{\text{animal}} =$
326 960).
327 We have data on the five phenotypes, two of them sex-limited: (i) the number of eggs laid by each
328 female after random mating which we had generated previously using Poisson distributions (with

329 additive dispersion) and we revisit here for analysis with quasi-Poisson models (i.e. multiplicative
330 dispersion), (ii) the incidence of endo-parasitic infections that we generated as being negative
331 binomial distributed, (iii) body length of adult beetles which we had generated previously using
332 Gaussian distributions and that we revisit here for analysis with gamma distributions, (iv) time to
333 visit five predefined sectors of an arena (employed as a measure of exploratory tendencies) that we
334 generated as being gamma distributed, and (v) the two male morphs, which was again generated
335 with binomial distributions. We will use this simulated dataset to estimate R^2_{GLMM} and ICC_{GLMM} .

336 All data generation and analyses were conducted in R 3.3.1 [10]. We used functions to fit GLMMs
337 from the three R packages: 1) the *glmmadmb* function from *glmmADMB* [30], 2) the *glmmPQL*
338 function from MASS [31], and 3) the *glmer* and *glmer.nb* functions from *lme4* [32]. In Table 1, we
339 only report results from *glmmADMB* because this is the only function that can fit models with all
340 relevant distributional families. All scripts and results are provided as an electronic supplement
341 (Appendix S6). In addition, Appendix S6 includes an example of a model using the Tweedie
342 distribution, which was fitted by the *cpglmm* function from the *cplm* package [21]. Notably, our
343 approach for R^2_{GLMM} is kindly being implemented in the *rsquared* function in the R package,
344 *piecewiseSEM* [33]. Another important note is that we often find less congruence in GLMM results
345 from the different packages than those of linear mixed-effects models, LMM. Thus, it is
346 recommended to run GLMMs in more than one package to check robustness of the results although
347 this may not always be possible.

348 In all the models, estimated regression coefficients and variance components are very much in
349 agreement with what is expected from our parameter settings (Table 1 and Appendix S6). When
350 comparing the null and full models, which had ‘sex’ as a predictor, the magnitudes of the variance
351 component for the container effect always decrease in the full models. This is because the variance
352 due to sex is confounded with the container variance in the null model. As expected, (unadjusted)
353 ICC values from the null models are usually smaller than adjusted ICC values from the full models
354 because the observation-level variance (analogous to the residual variance) was smaller in the full

355 models, implying that the denominator of equation (3.2) shrinks. However, the numerator also
356 becomes smaller for ICC values for the container effect from the parasite, size and exploration
357 models so that adjusted ICC values are not necessarily larger than unadjusted ICC values.
358 Accordingly, adjusted $ICC_{\text{container}}$ is smaller in the parasite and size models but not in the
359 exploration model. The last thing to note is that for the morph models (binomial mixed models),
360 both R^2 and ICC_{values} are larger when using the distribution-specific variance rather than the
361 observation-level variance, as discussed above (Table 3; also see Appendix S4).

362 **9. Alternatives and a cautionary note**

363 Here we extended our simple methods for obtaining R^2_{GLMM} and ICC_{GLMM} for Poisson and
364 binomial GLMMs to other types of GLMMs such as negative binomial and gamma. We have
365 described three different ways of obtaining the observational-level variance and how to obtain the
366 key rate parameter λ for Poisson and negative binomial distributions. We discussed important
367 considerations which arise for estimating R^2_{GLMM} and ICC_{GLMM} with binomial GLMMs. As we
368 have shown, the merit of our approach is not only its ease of implementation but also that our
369 approach encourages researchers to pay more attention to variance components at different levels.
370 Research papers in the field of ecology and evolution often report only regression coefficients but
371 not variance components of GLMMs [3].

372 We would like to highlight two recent studies that provide alternatives to our approach. First, Jaeger
373 and colleagues [5] have proposed R^2 for fixed effects in GLMMs, which they referred to as $R^2_{\beta^*}$ (an
374 extension of an R^2 for fixed effects in linear mixed models or R^2_{β} by Edwards and colleagues [34]).
375 They show that $R^2_{\beta^*}$ is a general form of our marginal R^2_{GLMM} ; in theory, $R^2_{\beta^*}$ can be used for any
376 distribution (error structure) with any link function. Jaeger and colleagues highlight that in the
377 framework of $R^2_{\beta^*}$, they can easily obtain semi-partial R^2 , which quantifies the relative importance
378 of each predictor (fixed effect). As they demonstrate by simulation, their method potentially gives a
379 very reliable tool for model selection. One current issue for this approach is that implementation

380 does not seem as simple as our approach. We note that our R^2_{GLMM} framework could also provide
381 semi-partial R^2 via commonality analysis (see [35]; note that unique variance for each predictor in
382 commonality analysis corresponds to semi-partial R^2 ; [36]).

383 Second, de Villemereuil and colleagues [23] provided a framework with which one can estimate
384 exact heritability using GLMMs at different scales (e.g. data and latent scales). Their method can be
385 extended to obtain exact ICC values on the data (observation) scale, which is analogous to, but not
386 the same as, our ICC_{GLMM} using the observation-level variance, σ_{ϵ}^2 described above. Further, this
387 method can, in theory, be extended to estimate R^2_{GLMM} on the data (observation) scale. One
388 potential difficulty is that the method of de Villemereuil and colleagues. is exact but that a
389 numerical method is used to solve relevant equations so one will require a software package (e.g.,
390 the QGglmm package; [23]).

391 Finally, we finish by repeating what we said at the end of our original R^2 paper [3]. Both R^2 and
392 ICC are indices that are likely to reflect only one or a few aspects of a model fit to the data and
393 should not be used for gauging the quality of a model. We encourage biologists use R^2 and ICC in
394 conjunctions with other indices like information criteria (e.g. AIC, BIC and DIC), and more
395 importantly, with model diagnostics such as checking for model assumptions, heteroscedasticity
396 and sensitivity to outliers.

397 **Authors' contribuions**

398 SN conceived ideas and conducted analysis. Both developed the ideas further, and contributed to
399 writing and editing of the manuscript.

400 **Competing interests**

401 We have not competing interests

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409

410 References

- 411 [1] Lessells, C.M. & Boag, P.T. 1987 Unrepeatable repeatabilities - a common mistake. *Auk* **104**,
412 116-121.
- 413 [2] Nakagawa, S. & Schielzeth, H. 2010 Repeatability for Gaussian and non-Gaussian data: a
414 practical guide for biologists. *Biol Rev* **85**, 935-956. (doi:10.1111/j.1469-185X.2010.00141.x).
- 415 [3] Nakagawa, S. & Schielzeth, H. 2013 A general and simple method for obtaining R^2 from
416 generalized linear mixed-effects models. *Methods Ecol Evol* **4**, 133-142. (doi:10.1111/j.2041-
417 210x.2012.00261.x).
- 418 [4] Johnson, P.C.D. 2014 Extension of Nakagawa & Schielzeth's R^2 -GLMM to random slopes
419 models. *Methods Ecol Evol* **5**, 944-946. (doi:10.1111/2041-210x.12225).
- 420 [5] Jaeger, B.C., Edwards, L.J., Das, K. & Sen, P.K. 2016 An R^2 statistic for fixed effects in the
421 generalized linear mixed model. *Journal of Applied Statistics*,
422 10.1080/02664763.02662016.01193725. (doi:10.1080/02664763.2016.1193725).
- 423 [6] LaHuis, D.M., Hartman, M.J., Hakoyama, S. & Clark, P.C. 2014 Explained Variance Measures
424 for Multilevel Models. *Organ Res Methods* **17**, 433-451. (doi:10.1177/1094428114541701).
- 425 [7] Bolker, B.M., Brooks, M.E., Clark, C.J., Geange, S.W., Poulsen, J.R., Stevens, M.H.H. &
426 White, J.S.S. 2009 Generalized linear mixed models: a practical guide for ecology and evolution.
427 *Trends Ecol Evol* **24**, 127-135. (doi:Doi 10.1016/J.Tree.2008.10.008).
- 428 [8] Bolker, B.M. 2008 *Ecological models and data in R*. Princeton, NJ, Princeton University Press.
- 429 [9] Ver Hoef, J.M. & Boveng, P.L. 2007 Quasi-Poisson vs. negative binomial regression: how
430 should we model overdispersed count data? *Ecology* **88**, 2766-2772.
- 431 [10] R Development Core Team. 2016 R: A language and environment for statistical computing.
432 (version 2.15.0 ed. Vienna, Austria, R Foundation for Statistical Computing.
- 433 [11] Snijders, T. & Bosker, R. 1999 *Multilevel Analysis: an Introduction to basic and advanced*
434 *multilevel modeling*. London, Sage.

- 435 [12] Snijders, T. & Bosker, R. 2011 *Multilevel Analysis: an Introduction to basic and advanced*
436 *multilevel modeling*. 2nd ed. London, Sage.
- 437 [13] Harrison, X.A. 2014 Using observation-level random effects to model overdispersion in count
438 data in ecology and evolution. *Peerj* **2**, e616. (doi:ARTN e61610.7717/peerj.616).
- 439 [14] Harrison, X.A. 2015 A comparison of observation-level random effect and Beta-Binomial
440 models for modelling overdispersion in Binomial data in ecology & evolution. *Peerj* **3**, e1114.
441 (doi:ARTN e111410.7717/peerj.1114).
- 442 [15] Browne, W.J., Subramanian, S.V., Jones, K. & Goldstein, H. 2005 Variance partitioning in
443 multilevel logistic models that exhibit overdispersion. *J R Stat Soc a Stat* **168**, 599-613.
- 444 [16] Gelman, A. & Hill, J. 2006 *Data analysis using regression and multilevel/hierarchical models*
445 Cambridge, Cambridge University Press.
- 446 [17] Ver Hoef, J.M. 2012 Who Invented the Delta Method? *Am Stat* **66**, 124-127.
447 (doi:10.1080/00031305.2012.687494).
- 448 [18] Powell, L.A. 2007 Approximating variance of demographic parameters using the delta method:
449 A reference for avian biologists. *Condor* **109**, 949-954. (doi:Doi 10.1650/0010-
450 5422(2007)109[949:Avodpu]2.0.Co;2).
- 451 [19] Foulley, J.L., Gianola, D. & Im, S. 1987 Genetic Evaluation of Traits Distributed as Poisson-
452 Binomial with Reference to Reproductive Characters. *Theor. Appl. Genet.* **73**, 870-877. (doi:Doi
453 10.1007/Bf00289392).
- 454 [20] Foster, S.D. & Bravington, M.V. 2013 A Poisson-Gamma model for analysis of ecological
455 non-negative continuous data. *Environ Ecol Stat* **20**, 533-552. (doi:10.1007/s10651-012-0233-0).
- 456 [21] Zhang, Y.W. 2013 Likelihood-based and Bayesian methods for Tweedie compound Poisson
457 linear mixed models. *Stat Comput* **23**, 743-757. (doi:10.1007/s11222-012-9343-7).
- 458 [22] Tempelman, R.J. & Gianola, D. 1999 Genetic analysis of fertility in dairy cattle using negative
459 binomial mixed models. *J. Dairy Sci.* **82**, 1834-1847.

- 460 [23] de Villemereuil, P., Schielzeth, H., Nakagawa, S. & Morrissey, M. 2016 General Methods for
461 Evolutionary Quantitative Genetic Inference from Generalized Mixed Models. *Genetics* **204**, 1281-
462 1294. (doi:10.1534/genetics.115.186536).
- 463 [24] Matos, C.A.P., Thomas, D.L., Gianola, D., Tempelman, R.J. & Young, L.D. 1997 Genetic
464 analysis of discrete reproductive traits in sheep using linear and nonlinear models .1. Estimation of
465 genetic parameters. *J. Anim. Sci.* **75**, 76-87.
- 466 [25] Carrasco, J.L. 2010 A generalized concordance correlation coefficient based on the variance
467 components generalized linear mixed models for overdispersed count data. *Biometrics* **66**, 897-904.
468 (doi:Doi 10.1111/J.1541-0420.2009.01335.X).
- 469 [26] Rao, C.R. 2002 *Linear statistical inference and its applications*. 2nd ed. ed. New York, John
470 Wiley & Sons.
- 471 [27] Oehlert, G.W. 1992 A note on the delta method. *Am Stat* **46**, 27-29. (doi:Doi
472 10.2307/2684406).
- 473 [28] Zeger, S.L., Liang, K.Y. & Albert, P.S. 1988 Models for Longitudinal Data - a Generalized
474 Estimating Equation Approach. *Biometrics* **44**, 1049-1060. (doi:Doi 10.2307/2531734).
- 475 [29] Goldstein, H., Browne, W. & Rasbash, J. 2002 Partitioning variation in multilevel models.
476 *Understanding Statistics* **1**, 223-231.
- 477 [30] Fournier, D.A., Skaug, H.J., Ancheta, J., Ianelli, J., Magnusson, A., Maunder, M.N., Nielsen,
478 A. & Sibert, J. 2012 AD Model Builder: using automatic differentiation for statistical inference of
479 highly parameterized complex nonlinear models. *Optim Method Softw* **27**, 233-249.
480 (doi:10.1080/10556788.2011.597854).
- 481 [31] Venables, W.N. & Ripley, B.D. 2002 *Modern applied statistics with S*. 4 ed. New York,
482 Springer.
- 483 [32] Bates, D., Machler, M., Bolker, B.M. & Walker, S.C. 2015 Fitting Linear Mixed-Effects
484 Models Using lme4. *J Stat Softw* **67**, 1-48.

- 485 [33] Lefcheck, J.S. 2016 PIECEWISESEM: Piecewise structural equation modelling in R for
486 ecology, evolution, and systematics. *Methods Ecol Evol* **7**, 573-579. (doi:10.1111/2041-
487 210x.12512).
- 488 [34] Edwards, L.J., Muller, K.E., Wolfinger, R.D., Qaqish, B.F. & Schabenberger, O. 2008 An R^2
489 statistic for fixed effects in the linear mixed model. *Stat Med* **27**, 6137-6157. (doi:Doi
490 10.1002/Sim.3429).
- 491 [35] Ray-Mukherjee, J., Nimon, K., Mukherjee, S., Morris, D.W., Slotow, R. & Hamer, M. 2014
492 Using commonality analysis in multiple regressions: a tool to decompose regression effects in the
493 face of multicollinearity. *Methods Ecol Evol* **5**, 320-328. (doi:Doi 10.1111/2041-210x.12166).
- 494 [36] Nimon, K.F. & Oswald, F.L. 2013 Understanding the Results of Multiple Linear Regression:
495 Beyond Standardized Regression Coefficients. *Organ Res Methods* **16**, 650-674.
496 (doi:10.1177/1094428113493929).
- 497
- 498
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Table 1. The observation-level variance σ_ε^2 for the three distributional families: quasi-Poisson (overdispersed Poisson), negative binomial and gamma with the three different methods for deriving σ_ε^2 : the delta method, long-normal approximation and the trigamma function, ψ_1 .

Family	Distributional parameters	Mean (E[y]) Variance (var[y])	Link function	Delta method	log-normal approximation	trigamma function
Quasi-Poisson (OP: overdispersed Poisson)	OP(λ, ω)	E[y] = λ	log	$\frac{\omega}{\lambda}$	$\ln\left(1 + \frac{\omega}{\lambda}\right)$	$\psi_1\left(\frac{\lambda}{\omega}\right)$
Poisson (when $\omega = 1$)	$\lambda > 0$ $\omega > 0$	var[y] = $\lambda\omega$	square-root	0.25ω	-	
Negative binomial (NB)	NB(λ, θ)	E[y] = λ	log	$\frac{1}{\lambda} + \frac{1}{\theta}$	$\ln\left(1 + \frac{1}{\lambda} + \frac{1}{\theta}\right)$	$\psi_1\left(\left[\frac{1}{\lambda} + \frac{1}{\theta}\right]^{-1}\right)$

		$\lambda > 0$	$\text{var}[y] = \lambda + \frac{\lambda^2}{\theta}$	square-root	$0.25 \left(1 + \frac{\lambda}{\theta} \right)$	-	
		$\theta > 0$					
Gamma	$\text{gmma}(\lambda, \nu)$	$E[y] = \lambda$		log	$\frac{1}{\nu}$	$\ln \left(1 + \frac{1}{\nu} \right)$	$\psi_1(\nu)$
		$\lambda > 0$	$\text{var}[y] = \frac{\lambda^2}{\nu}$	inverse	$\frac{1}{\nu \lambda^2}$	-	
		$\nu > 0$		(reciprocal)			
Gamma (alternative parameterization)	$\text{gamma}(\nu, \kappa)$	$E[y] = \frac{\nu}{\kappa}$		log	$\frac{1}{\nu}$	$\ln \left(1 + \frac{1}{\nu} \right)$	$\psi_1(\nu)$
		$\nu > 0$	$\text{var}[y] = \frac{\nu}{\kappa^2}$	inverse	$\frac{\kappa^2}{\nu^3}$	-	
		$\kappa > 0$		(reciprocal)			

$\text{var}[\ln(x)] = \psi_1(\nu) = \sum_{n=1}^{\infty} 1/(\nu+n)$ when x follows gamma distribution. In the R environment, the function, *trigamma* can be used to obtain $\psi_1(\nu)$.

Table 2. The distribution-specific variance σ_d^2 and observation-level variance σ_ϵ^2 for binomial (and Bernoulli) distributions; note that only one of them should be used for obtaining R^2 and ICC.

Family	Distributional parameters, mean & variance	Link name	Link function	Distribution-specific variance	Observation-level variance using the delta method (min. values and corresponding p)
Binomial (Bernoulli; $n = 1$)	binomial(p, n) $0 < p < 1$ $n > = 1$ (integers)	logit	$\ln\left(\frac{p}{1-p}\right)$	$\frac{\pi^2}{3} \sim 3.29$ (logistic distribution)	$\frac{1}{p(1-p)}$ (min = 4; $p = 0.5$)
	$E[y] = np$ $\text{var}[y] = np(1-p)$	probit ($\Phi(p)$)	$\sqrt{2}\text{erf}^{-1}(2p-1)$	1 (standard normal distribution)	$2\pi p(1-p)\left(\exp\left[\text{erf}^{-1}(2p-1)\right]^2\right)^2$ (min ~ 1.57 ; $p = 0.5$)

cloglog		$\frac{\pi^2}{6} \sim 1.65$	$\frac{p}{(\ln(1-p))^2 (1-p)}$
(complimentary log-log)	$\ln(-\ln(1-p))$	(Gumbel distribution)	(min ~ 1.54 ; $p \sim 0.8$; ~ 2.08 ; $p = 0.5$)

'erf⁻¹' is the inverse of the Gauss error function, which is often denoted as 'erf'.

Table 3. Mixed-effects model analysis of a simulated dataset estimating variance components and regression slopes for nutrient manipulations on fecundity, endoparasite loads, body length, exploration levels and male morph types; $N_{[population]}=12$, $N_{[container]}=120$ and $N_{[animal]}=960$.

Model name	Fecundity models (log-link)		Parasite models (log-link)		Size models (log-link)		Exploration models (log-link)		Morph models (logit-link)	
	Quasi-Poisson mixed models		Negative binomial mixed models		Gamma mixed models		Gamma mixed models		Binomial (binary) mixed models	
	Null Model	Full Model	Null Model	Full Model	Null Model	Full Model	Null Model	Full Model	Null Model	Full Model
Fixed effects	<i>b</i>	<i>b</i>	<i>b</i>	<i>b</i>	<i>b</i>	<i>b</i>	<i>b</i>	<i>b</i>	<i>b</i>	<i>b</i>
	[95% CI]	[95% CI]	[95% CI]	[95% CI]	[95% CI]	[95% CI]	[95% CI]	[95% CI]	[95% CI]	[95% CI]
Intercept	1.630	1.261	0.766	1.752	2.682	2.737	4.752	4.056	-0.108	-0.740
	[1.379, 1.882]	[0.989, 1.532]	[0.330, 1.202]	[1.282, 2.223]	[2.616, 2.689]	[2.699, 2.775]	[4.555, 4.949]	[3.842, 4.269]	[-0.718, 0.501]	[-1.450, -0.030]
Treatment (experiment)	-	0.491	-	-0.768	-	0.033	-	2.007	-	0.840
		[0.391, 0.591]		[-0.870, -0.667]		[0.023, 0.044]		[1.965, 2.050]		[0.422, 1.258]
Habitat (wet)	-	0.152	-	0.700	-	0.009	-	-0.560	-	0.414
		[0.055, 0.249]		[0.599, 0.801]		[-0.001, 0.019]		[-0.603, -0.518]		[0.002, 0.826]
Sex (male)	-	-	-	-2.198	-	-0.213	-	-1.105	-	-

				[-2.511, -1.884]		[-0.230, -0.196]		[-1.256, -0.955]		-
Random effects	σ^2	σ^2	σ^2	σ^2	σ^2	σ^2	σ^2	σ^2	σ^2	σ^2
Population	0.178	0.187	0.375	0.541	0.0026	0.0039	0.071	0.104	1.002	1.111
Container	0.042	0.059	1.976	0.613	0.0140	0.0014	0.364	0.163	0.136	0.186
Observation-level (Distribution-specific)	0.477	0.349	0.873	0.397	0.0069	0.0064	1.664	0.118	4.010 (3.290)	4.010 (3.290)
Fixed factors	-	0.066	-	1.479	-	0.0116	-	1.393	-	0.220
$R^2_{GLMM(m)}$	-	10.01%	-	48.83%	-	49.54%	-	78.34%	-	3.98% (4.57%)
$R^2_{GLMM(c)}$	-	47.19%	-	86.91%	-	72.52%	-	93.34%	3.98% (4.57%)	27.46% (31.55%)
ICC _[Population]	25.50%	31.47%	11.62%	34.89%	11.38%	33.17%	3.40%	26.94%	19.49% (22.63%;)	20.96% (24.23%)

ICC _[Container]	5.98%	9.84%	61.30%	39.53%	59.57%	12.37%	17.34%	42.34%	2.67% (3.07%;)	3.50% (4.05%)
AIC	2498.8	2412.3	4342.6	3920.5	3379.9	3139.5	11223.8	9004.3	605.5	589.6

95 % CI (confidence intervals) were calculated by the *confint* function in lme4. The observation-level variance was obtained by using the trigamma function. In the Morph models, both the observation-level variance and distribution-specific variance were used; note that ones in brackets use the distribution-specific variance for R^2 and ICC. ICC_[Container] is not a typical ‘repeatability’ but the proportion of variance due to the container effect beyond the population variance.

Figure legends

Figure 1. A schematic of how hypothetical datasets are obtained (see the main text for details).

