

1 Content type: Article

2 **THE SYSTEMATIC CONSERVATION PLANNING FOR INTRASPECIFIC**
3 **GENETIC DIVERSITY.**

4 Ivan PAZ-VINAS^{1,2,3*}, Géraldine LOOT^{1,4}, Virgilio HERMOSO⁶, Charlotte
5 VEYSSIERE^{1,4}, Nicolas POULET⁷, Gaël GRENOUILLET^{1,4} and Simon
6 BLANCHET^{1,5}

7

8 ¹ Centre National de la Recherche Scientifique (CNRS), Université Paul Sabatier (UPS),
9 École Nationale de Formation Agronomique (ENFA); UMR 5174 EDB (Laboratoire
10 Évolution & Diversité Biologique), 118 route de Narbonne, 31062 Toulouse cedex 4,
11 France.

12 ² Aix-Marseille Université, CNRS, IRD, Avignon Université; UMR 7263 IMBE, Équipe
13 EGE, Centre Saint-Charles, Case 36, 3 place Victor Hugo, 13331 Marseille cedex 3,
14 France.

15 ³ Université de Lyon, CNRS, ENTPE; UMR 5023 LEHNA, Laboratoire d'Écologie des
16 Hydrosystèmes Naturels et Anthropisés, 6 rue Raphaël Dubois, 69622 Villeurbanne,
17 France.

18 ⁴ Université de Toulouse, UPS; UMR 5174 EDB, 118 route de Narbonne, 31062
19 Toulouse cedex 4, France.

20 ⁵ CNRS, Station d'Écologie Théorique et Expérimentale; UMR 5321, 09200 Moulis,
21 France.

22 ⁶ Centre Tecnologic Forestal de Catalunya, Crta. Sant Llorenç de Monunys, Km 2,
23 25280, Solsona. Lleida, Spain.

24 ⁷ Pôle Écohydraulique ONEMA-IMFT, Office National de l'Eau et des Milieux
25 Aquatiques, France.

26

27 **Short title:** Conservation planning of genetic diversity

28

29 **Keywords:** biodiversity, dendritic ecological networks, fish, freshwater, genetic
30 diversity, geostatistical tools, microsatellites, river basin, spatial patterns, systematic
31 conservation planning

32

33 ***Corresponding author:**

34

35 Ivan PAZ-VINAS

36 Université Lyon 1

37 CNRS, ENTPE, UMR 5023 LEHNA

38 Laboratoire d'Écologie des Hydrosystèmes Naturels et Anthropisés

39 3, rue Raphaël Dubois - Bât. Darwin C

40 69622 Villeurbanne Cedex France

41 Phone: (+33) 04 72 43 26 03

42 E-mail: ivanpaz23@gmail.com

43

44 **Co-author's e-mails:**

45

46 Simon Blanchet: simon.blanchet@sete.cnrs.fr

47 Virgilio Hermoso: virgilio.hermoso@gmail.com

48 Charlotte Veyssière: veyssiere.charlotte@gmail.com

49 Nicolas Poulet: nicolas.poulet@onema.fr

50 Gaël Grenouillet: gael.grenouillet@univ-tlse3.fr

51 Géraldine Loot: geraldine.loot@univ-tlse3.fr

52

53 **Abstract**

54

55 Intraspecific diversity is fundamental for species' adaptation to environmental
56 changes, and should hence be a main target for biodiversity conservation. However,
57 attempts to identify priority conservation areas for intraspecific diversity remain scarce.
58 Here, we used molecular data on six freshwater fish species sampled at a large spatial
59 scale, to determine hot- and cold-spots of genetic diversity, and to identify priority
60 conservation areas using a systematic conservation planning approach. We demonstrate
61 that the systematic conservation planning is an efficient and relevant approach for
62 preserving intraspecific diversity, although we identify weak congruencies and
63 surrogacies among conservation solutions found for each species. These weak
64 congruencies are due to among-species variation in the spatial distribution of hot-spots
65 of genetic diversity. We finally provide operational guidelines to efficiently use
66 systematic conservation planning methods with intraspecific genetic diversity data, and
67 to identify priority conservation areas for intraspecific diversity.

68

69

70 Biodiversity conservation is a major challenge that is often addressed by
71 identifying protected areas with high biodiversity and/or landscape values¹.
72 Conservation areas are generally identified as areas with high proportions of endemic,
73 rare or iconic species². Alternatively, conservation planning can be based on the concept
74 of complementarity between conservation areas³, and on cost-effectiveness analyses
75 such as systematic conservation planning methods (hereafter SCP⁴). SCP aims at
76 identifying a number of sites (i.e. *irreplaceable sites* that should be managed for
77 conservation in priority) best representing a predefined proportion of the biodiversity
78 observed in a region, at a minimum cost.

79 There have been attempts to include information on the phylogenetic history of
80 species assemblages into SCP approaches to preserve both species identities and their
81 macro-evolutionary history^{5,6}. However, genetic diversity observed at the *population*

82 *level* (i.e. within species) has rarely been considered in SCP. *Intraspecific genetic*
83 *diversity* is a fundamental facet of biodiversity, as it is the fuel for species to adapt to
84 global and environmental changes^{7–10}. Conservation geneticists have classically
85 considered this facet of biodiversity in conservation plans, for instance by identifying
86 “Evolutionary Significant Units” or unique genetic lineages¹¹. However, conservation
87 geneticists have almost ignored the possibility to combine genetic data (e.g. allele
88 identities) with dedicated planning tools such as SPC¹².

89 The relative lack of genetic datasets at *large* spatial scales may partly explain
90 why SCP has yet rarely been applied to intraspecific genetic diversity^{13,14}. Particularly,
91 conservation geneticists have been generally restricted by the amount and spatial range
92 of datasets. However, our capacity to compile genetic datasets at large spatial, temporal
93 and taxonomic scales has greatly increased in the last decades^{15,16}, so that it is now
94 possible to identify priority areas for the conservation of genetic diversity using
95 dedicated conservation planning tools.

96 Here, we tested the potential of SCP analyses to identify priority conservation
97 areas accounting for intraspecific genetic diversity measured at a large spatial scale. We
98 first considered a set of four common and representative freshwater fish species
99 (*Squalius cephalus*, *Gobio occitaniae*, *Barbatula barbatula* and *Phoxinus phoxinus*) to
100 test the influence of conservation targets (proportion of the total amount of genetic
101 diversity to be covered by irreplaceable sites) and analytical strategies (analysing each
102 species independently or all species pooled) on final conservation solutions (number
103 and identity of irreplaceable sites). We then included two rare species of particular
104 conservation interest (*Leuciscus burdigalensis* and *Parachondrostoma toxostoma*) to
105 test the relevance of the SCP approach in a “real conservation-oriented study”. For these
106 two species (and the four common species), we ran SCP analyses considering a typical

107 conservation target^{17,18} to (i) explore the spatial distribution of irreplaceable sites in a
108 riverscape, and (ii) test for congruency and surrogacy in irreplaceable sites among
109 species, and more particularly between rare and common species. We finally tested
110 whether –or not– irreplaceable sites were correctly predicted by classical indices of
111 genetic diversity (e.g. allelic richness). We demonstrate that preserving the genetic
112 diversity of a species assemblage is a feasible –yet complex– task necessitating
113 appropriate analyses to assist the decision-making process.

114

115 **Results**

116

117 *Descriptive statistics.* We assessed for the six species within-sites intraspecific genetic
118 diversity (i.e. α -IGD) by calculating both allelic richness (AR) and richness in private
119 alleles (PA). Overall, *P. toxostoma* (one of the two rare species) showed the lowest α -
120 IGD. Mean AR ranged from 2.114 for *P. toxostoma* to 5.821 for *P. phoxinus*, and mean
121 PA ranged from 0.036 for *P. toxostoma* to 0.162 for *L. burdigalensis* (Table S1). We
122 also assessed among-sites intraspecific genetic diversity (i.e. β -IGD) by quantifying (for
123 each species) how much a site is genetically unique compared to all others (using the
124 D_{est} index¹⁹, see the Methods section). *Parachondrostoma toxostoma* also showed the
125 lowest mean D_{est} value (0.069), while the highest mean value was found for *B.*
126 *barbatula* (0.383; Table S1).

127

128 Testing the suitability of SCP analyses for intraspecific genetic diversity

129

130 In this first step we focused on the four common species for which genetic data
131 were available at a large spatial scale, allowing a thorough exploration of the suitability
132 of SCP for intraspecific genetic diversity.

133

134 *Spatial patterns of genetic diversity.* Using Generalized Linear Models for Stream
135 Networks (GLMSSN)^{20,21}, we showed that spatial patterns of genetic diversity largely
136 varied and were actually poorly congruent among the four common fish species (Figure
137 1). As extreme examples, hotspots of *AR* for *S. cephalus* were mainly found in the
138 Western part of the network and on the core streams, whereas these same areas were
139 identified as coldspots of *AR* for *B. barbatula* (Figure 1A1-1A3). Similarly, hotspots of
140 *PA* were inversely related between *G. occitaniae* and *P. phoxinus* (Figure 1B2-1B4).
141 Similar conclusions were reached for *D_{est}* (Figure 1C). For instance, hotspots of *D_{est}*
142 were observed in opposite areas of the river basin for the species pair *B. barbatula/P.*
143 *phoxinus* (Figure 1C3-1C4). As a consequence, the sign, slope and significance of
144 GLMSSNs explanatory variables strongly varied among species (Figure 1). This
145 incongruence in spatial patterns of genetic diversity among species was also reflected by
146 the low to moderate correlation coefficients measured among all possible pairs of
147 species and for each index of genetic diversity (i.e. *AR*, *PA* and *D_{est}*, Table S2). Indeed,
148 Pearson's correlation coefficients were lower than 0.6 for all comparisons but two (i.e.
149 between *B. barbatula/G. occitaniae* and between *P. phoxinus/B. barbatula* for *AR*, see
150 Table S2).

151

152 *The influence of conservation targets and analytical strategies to identify irreplaceable*
153 *sites for genetic conservation.* We used alleles' presence/absence data combined with
154 SCP procedures to test the influence of conservation targets and analytical strategies on

155 the number and identity of irreplaceable sites (i.e. sites that were selected in all the
156 solutions in SCP analyses; see the Methods section for more details).

157 When species were analysed independently, we found that the number of
158 irreplaceable sites increased as the conservation target (i.e. the percentage of alleles to
159 be covered by irreplaceable sites) increased, with a steep increase for conservation
160 targets higher than 75% of the total number of alleles present at the river basin scale
161 (Figure 2). However, the percentage of irreplaceable sites strongly varied among
162 species. For instance, for a 90% conservation target –which is a common target in
163 conservation genetics¹⁷–, the proportion of irreplaceable sites ranged from 3.61% of the
164 total number of sampled sites for *G. occitaniae* to 28.57% for *P. phoxinus* (Table S3;
165 Figure 2). For extreme conservation targets (100% of alleles to be covered), the
166 proportion of irreplaceable sites varied from 25.30% for *G. occitaniae* to 68.26% for *P.*
167 *phoxinus* (Table S3; Figure 2).

168 When alleles from the four common species were analysed in a single pooled
169 analysis, we similarly found that the proportion of irreplaceable sites increase as the
170 conservation target increases (Figure 2). Interestingly, we did not identify irreplaceable
171 sites for the 30% conservation goal, and only 3 irreplaceable sites were found for the
172 50% conservation goal (Figure 2). The proportion of irreplaceable sites increased
173 moderately to 17.39% for the 75% target, and then steeply increased for higher
174 conservation targets to reach 55.70% for the 90% target and 76.08% for the 100% target
175 (Figure 2). This later result suggests that almost all the river basin should be protected
176 to reach high conservation targets when adopting a pooled strategy.

177

178 A real conservation-oriented study using SCP approaches

179

180 We here focused on two rare species (in addition to the four common species) to
181 explore the usefulness of SCPs in a real case study.

182

183 *Identification of irreplaceable sites for genetic conservation.* We first visually explored
184 the spatial distribution of irreplaceable sites. Overall, the localization of irreplaceable
185 sites in the riverscape strongly varied among species, being spread all over the river
186 basin (Figure 3). We failed to identify areas (e.g. upstream or downstream locations)
187 clustering irreplaceable sites for any species (Figure 3A-F). This apparent lack of
188 clustering was statistically confirmed by our generalized linear models (Table S4).
189 Indeed, the two positional indices we used as explanatory variables (i.e. distance from
190 the outlet of sampling sites and the betweenness centrality of each sampling site, see the
191 Methods section) were not significant predictors of the irreplaceability of sites for all
192 species, except for distance from the outlet for *P. phoxinus* (Table S4). This indicates
193 that neither the position of sites in the riverscape, nor the positional importance of these
194 sites, determine the irreplaceability of sites.

195 Second, we tested whether conservation solutions found for each species were
196 spatially congruent among species. Over the six fish species, we identified forty-two
197 sites (out of the ninety-two sites, i.e. 45.65%) as irreplaceable at the 90% conservation
198 target for at least one species (Figure 4). Thirty-two of these forty-two sites were
199 irreplaceable for at least one of the four common species (Figure 4), and fourteen of the
200 forty-two sites were irreplaceable for at least one of the two rare species (Figure 4).
201 Among the six species, only eight out of these forty-two sites were irreplaceable for at
202 least two species, and only one of these sites was irreplaceable for three species (Figure
203 4).

204

205 *Surrogacy in irreplaceable sites among species.* We tested whether solutions found for
206 one species can be used as a surrogate for other species by calculating the percentage of
207 the total number of alleles observed for a given species that is covered by irreplaceable
208 sites identified for another species. Levels of surrogacy were generally low to moderate,
209 and strongly varied among species pairs (Table 1). For instance, irreplaceable sites
210 identified for *G. occitaniae* failed to cover the genetic diversity of the two rare species,
211 and covered only 17.66 to 42.60% of the total number of alleles of the other common
212 species. This indicates that irreplaceable sites identified for *G. occitaniae* (i.e. the most
213 widespread species) are poor surrogates for preserving the intraspecific genetic diversity
214 of other species (18.38% of surrogacy; Table 1). Conversely, irreplaceable sites found
215 for *P. phoxinus* and *L. burdigalensis* are better surrogates for *G. occitaniae*, as 79.58%
216 and 70.27% of the total number of alleles of *G. occitaniae* was covered by irreplaceable
217 sites identified for *P. phoxinus* and *L. burdigalensis* respectively (Table 1). Overall,
218 irreplaceable sites best covering genetic diversity of other species were those identified
219 for *B. barbatula*, which covered in average 68.28% of the total number of alleles of
220 other species (Table 1).

221 Interestingly, the thirty-two irreplaceable sites identified for the four common
222 species covered 79.87% and 90% of the total number of alleles of *L. burdigalensis* and
223 *P. toxostoma* respectively, suggesting that irreplaceable sites identified for a set of
224 common species can be good surrogates for intraspecific genetic diversity of rare
225 species. Conversely, the fourteen irreplaceable sites identified for the two rare species
226 covered a total number of alleles ranging from 65.18% for *S. cephalus* to 79.40% for *B.*
227 *barbatula*.

228

229 *Relationships between genetic indices and irreplaceable sites.* For all species but *G.*
230 *occitaniae* and *P. toxostoma* (for which none of the variables were significant
231 predictors), *PA* was identified as the only variable that significantly predicted the
232 probability for a site to be identified as an irreplaceable site (Table S5). This probability
233 increased with the number of *PA* in a site.

234

235 **Discussion**

236

237 Intraspecific diversity constitutes the fuel for species and populations to cope
238 with environmental changes^{7–9}. This biodiversity facet is hence the first that should
239 respond to global change, allowing populations and species to respond adaptively to
240 these changes¹⁰. However, this biodiversity facet has so far been poorly integrated in
241 dedicated optimization planning tools. We here fill this gap by demonstrating that the
242 systematic conservation planning of intraspecific genetic diversity is a feasible –yet
243 complex– task necessitating careful considerations.

244

245 *From idiosyncratic distributions of genetic diversity...*

246 Our results strongly suggest that the genetic diversity of the targeted species did
247 not follow a common spatial pattern, but rather species-specific (idiosyncratic) spatial
248 distributions. This conclusion holds true for all genetic diversity indices, and it
249 corroborates the few previous studies investigating simultaneously the spatial
250 distribution of genetic diversity at large spatial scales and for sympatric species
251 (e.g.^{22,23}). This was however unexpected given that recent meta-analyses on freshwater
252 organisms demonstrated that α -IGD is generally higher in downstream than in upstream
253 areas²⁴, and that β -IGD tends to be higher in upstream than in downstream sections²⁵.

254 Overall, these very general spatial patterns of intraspecific genetic diversity were
255 verified in our datasets. For instance, a negative relationship between allelic richness
256 and distance from the outlet is expected in freshwater organisms²⁴, and was actually
257 observed for three out of the six species considered in this study (Figure S1). However,
258 when using a precise and novel approach to map genetic diversity across the network,
259 we demonstrated that the distribution of cold- and hot-spots of α - and β -IGD was subtler
260 and idiosyncratic. This probably reflects interactions between colonization histories,
261 life-history-traits of species and the network structure, which are expected to drive
262 patterns of genetic diversity in rivers^{24,26,27}.

263

264 *...to the systematic conservation planning of intraspecific genetic diversity.*

265 The spatial mismatch in intraspecific genetic diversity among species probably
266 explains why we found that the level of congruency and surrogacy of irreplaceable sites
267 identified for each species was extremely low. For instance, there was an extremely low
268 proportion of irreplaceable sites that were common to two or three species (and never
269 more than three species; Figure 4). In the same way, we detected no clear patterns in the
270 spatial distribution of irreplaceable sites. In riverscapes, it is generally assumed that
271 small upstream areas are the “source” of genetic uniqueness, and hence the primary
272 areas to protect (i.e. the “small but mighty” paradigm²⁵). Our results did not confirm this
273 paradigm since irreplaceable sites (for any of the six species) were not particularly
274 situated in upstream areas and/or in areas of high connectivity (i.e. areas displaying high
275 centrality values^{28,29}), and rather suggests that priority areas for the conservation of
276 intraspecific genetic diversity should cover the whole distribution range of species.
277 Finally, the level of surrogacy among irreplaceable sites was low to moderate, and never
278 attained the 90% threshold we assumed when we considered all species

279 independently^{17,18}. Combined with our finding that the number of irreplaceable sites can
280 be relatively high for reasonable conservation targets (up to 46% sites are identified as
281 irreplaceable sites for at least one species at the conservation target of 90%), we
282 concluded that our ability to identify priority areas for intraspecific genetic diversity is
283 highly species-specific and depends on the capacity to tackle the trade-off between the
284 amount of genetic diversity to protect, and the extent of priority areas we can
285 realistically protect.

286 However, when surrogacy between all irreplaceable sites identified for the entire
287 set of common species and those identified for the rare species was tested, we reached
288 reasonable proportions of the total number of alleles to protect (~80-90%) for the two
289 rare species (i.e. *P. toxostoma* and *L. burdigalensis*). This result suggests that, in some
290 cases, genetic data obtained for a set of widely-distributed, “easier-to-sample” common
291 species displaying varying life-history traits can be used for identifying protection areas
292 for the intraspecific genetic diversity of other sympatric rare species that can be more
293 problematic to sample.

294 Overall, our results suggest that two different analytical strategies can be
295 employed in real-case SCP studies aiming at preserving intraspecific genetic diversity
296 (i) identification of conservation areas for each rare species independently or (ii)
297 identification of conservation areas for a set of representative common species. Both
298 strategies have their own advantages and inconveniences. The first strategy optimally
299 preserves genetic diversity of rare species at competitive costs (e.g. 14 irreplaceable
300 sites to protect in our demonstrative study), but this at the expense of the genetic
301 diversity of other sympatric species. Conversely, the second strategy will optimally
302 preserve genetic diversity of a set of common species while maintaining high levels of
303 genetic diversity for rare species, but this at a higher cost (e.g. 32 irreplaceable sites to

304 protect in our study). Whether to choose one of these two strategies will therefore
305 depend on many factors such as how difficult is the sampling of rare species compared
306 to common species, or the extent of the resources available for setting new protected
307 areas. We recommend however to adopt the second strategy when possible, since it
308 allows to simultaneously maintain genetic diversity from rare and common species.
309 Indeed, genetic diversity of common species is vital for ensuring ecosystem stability, as
310 it ultimately influence species interactions, population dynamics and ecosystem
311 functions³⁰, and we argue that it should be considered in conservation plans.

312

313 **Conclusions**

314

315 Our study provides novel, insightful and promising knowledge on the setting of
316 priority conservation areas for intraspecific diversity. It shows that systematic
317 conservation planning methods are useful objective tools for conservation geneticists
318 whose conservation solutions will strikingly depend on the species to be preserved and
319 the quantity of genetic information that managers aim at preserving in a landscape.
320 Given our results, we suggest that two strategies could be employed in real-case
321 conservation programs: (i) identification of priority conservation areas for each rare
322 species independently or (ii) identification of priority conservation areas on the basis of
323 the analysis of a set of representative common species that may serve as “umbrellas” for
324 rare sympatric species.

325 Our study also raises many additional questions that should be considered in the
326 near future. Among others, we believe that the next steps will be to formally identify
327 sound conservation targets for intraspecific diversity, to test whether neutral
328 intraspecific diversity appropriately mirrors quantitative and adaptive diversity⁹, and to

329 quantify the influence of intraspecific diversity on ecosystem functioning and services,
330 so as to better evaluate the added value of preserving such a facet of biodiversity^{9,31}.

331

332 **Methods**

333

334 Data collection

335

336 *Biological models.* We focused on an assemblage of six Cyprinid freshwater fish
337 species. Three of them are widespread in Europe (i.e. *Squalius cephalus*, *Phoxinus*
338 *phoxinus* and *Barbatula barbatula*) whereas three of them (*Gobio occitaniae*, *Leuciscus*
339 *burdigalensis* and *Parachondrostoma toxostoma*) are endemic to Southern France³².

340 This set of species covers a large functional trait space that is representative of many
341 freshwater fish communities. For instance, *S. cephalus* is a large-bodied fish species
342 with long lifespan (i.e. it can be 60 cm long and live up to 15 years³²) whereas at the
343 extreme *P. phoxinus* is a small-bodied species with shorter lifespan (i.e. it is less than 12
344 cm long and usually lives up to 4-5 years³²). From an ecological perspective, *G.*
345 *occitaniae*, *P. toxostoma* and *B. barbatula* are bottom feeders, whereas *S. cephalus* and
346 *P. phoxinus* are water column feeders and *L. burdigalensis* is more opportunistic.
347 Further, *B. barbatula* is mainly active during night, while the other species are
348 particularly active during the day. Four of these species are relatively abundant (i.e. *S.*
349 *cephalus*, *G. occitaniae*, *P. phoxinus* and *B. barbatula*), although they greatly vary in
350 their ecological niche and hence their spatial occupancy in the river network (see Figure
351 S2 for maps representing the spatial distribution of sampling sites for each species,
352 which roughly corresponds to their spatial distribution in the Garonne-Dordogne river
353 basin). We will hereafter refer to this set of species as the “common” species. The two

354 other species are rare (*L. burdigalensis*) to very rare (*P. toxostoma*; see Figure S2) in the
355 Garonne-Dordogne river basin, and are of particular interest for conservation. *Leuciscus*
356 *burdigalensis* is a recently described species that is locally experiencing both
357 demographic and genetic bottlenecks in many populations^{33,34}. *Parachondrostoma*
358 *toxostoma* is a vulnerable species³⁵ listed in the IUCN red list, in the Annex II of the
359 European Union Habitats Directive and in Appendix III of the Bern Convention³⁵.

360

361 *Sampling design.* During Spring/Summer 2010-2011, we used electric-fishing to sample
362 ninety-two sites distributed across thirty-five rivers from a large river basin, the
363 Garonne-Dordogne River basin (>100,000 km², South-Western France; Figure S2;
364 Table S6). Sampling sites were chosen to cover the whole distribution range of each
365 species at the riverscape scale, and to allow characterising spatial patterns of genetic
366 diversity for all these species. Up to 25 individuals *per species per site* were sampled
367 when possible. Not all species were present at all sampling sites (Figure S2; Table S6),
368 and some species were at a density that did not allow reaching the 25 individuals
369 threshold. In these cases, we captured as many individuals as possible. We anesthetized
370 each individual and then we collected and stored in 90% ethanol a fragment of their
371 pelvic fin. All individuals were released alive at their sampling location.

372

373 *Genotyping.* Genomic DNA was extracted using a salt-extraction protocol³⁶. We used
374 multiplexed Polymerase Chain Reactions (PCRs) to co-amplify 8 to 15 microsatellite
375 loci depending on the species (8 for *G. occitaniae*, 9 for *B. barbatula*, 10 for *S. cephalus*
376 and *P. phoxinus*, 14 for *L. burdigalensis* and 15 for *P. toxostoma*). We used 5-20 ng of
377 genomic DNA and QIAGEN® Multiplex PCR Kits (Qiagen, Valencia, CA, USA) to
378 perform PCR amplifications. Details on loci, primer concentrations, PCR conditions and

379 multiplex sets can be found in Table S7. The genotyping was conducted on an ABI
380 PRISM™ 3730 Automated Capillary Sequencer (Applied Biosystems, Foster City, CA,
381 USA). The scoring of allele sizes was done using GENEMAPPER® v.4.0 (Applied
382 Biosystems).

383

384 *Genetic diversity assessment.* Given the good spatial resolution of the sampling obtained
385 for the four common species (i.e. *S. cephalus*, *G. occitaniae*, *P. phoxinus* and *B.*
386 *barbatula*), descriptive genetic analyses were conducted for sampling sites displaying a
387 minimum sample size of N=10 individuals for these species, so as to maximize
388 consistency on subsequent allelic frequency-based genetic analyses (see Figure S2 and
389 Table S6 for details on sample sizes). For the two rare species, for which the sampling
390 was more restricted (i.e. *L. burdigalensis* and *P. toxostoma*), genetic analyses were
391 conducted for sampling sites displaying a minimum sample size of N=6 individuals, so
392 as to maximize the number of sampling sites included in the SCP procedures. We then
393 determined for each of the six species the occurrence of null alleles and potential
394 scoring errors with the program MICROCHECKER 2.3³⁷. We tested for departures
395 from Hardy-Weinberg (HW) equilibrium with the ‘adegenet’ R package v1.6-2³⁸. The
396 program GENEPOP v4.0³⁹ was used to assess linkage disequilibrium among loci within
397 sites. We found significant deviations from HW for a few locus/population pairs for
398 each six species (see Appendix A1 and Supplementary File 1 for details and raw tables),
399 and significant linkage disequilibrium and homozygote excesses for only the four
400 common species (Appendix A1; Supplementary File 1). However, no clear patterns
401 were observed for any species across loci and populations for these deviations. Given
402 the small extent of these deviations and given the large spatial extent of the databases,

403 we assumed that they weakly affected our main findings (Appendix A1; Supplementary
404 File 1).

405 To assess within-sites intraspecific genetic diversity (i.e. α -IGD), we applied
406 rarefaction procedures implemented in ADZE v1.0⁴⁰ to calculate both allelic richness
407 (AR^{41}) and private allelic richness (PA^{42}) at the sampling site level (based on a
408 minimum of N=10 individuals for common species or N=6 individuals for rare species).
409 To assess the among-sites component of intraspecific genetic diversity (i.e. β -IGD), we
410 used the R package ‘mmod’⁴³ to calculate –for each species– a pairwise genetic
411 differentiation index (i.e. D_{est}^{19}). For each site (and species), we then derived the
412 averaged value of all pairwise D_{est} values estimated between one given site and all the
413 remaining sites, so as to obtain a single value *per* site.

414

415 Testing the suitability of SCP analyses for intraspecific genetic diversity

416

417 In the first step, we tested the influence of conservation targets and analytical
418 strategies on final conservation solutions. In this step, we focused specifically on data
419 from the four common species, as their large coverage of the sampling area is more
420 suited for the demonstrative exercise done in this step.

421

422 *Spatial patterns of genetic diversity.* We first used geostatistical modelling tools to
423 explore spatial patterns of α and β genetic diversity for the four common species at the
424 riverscape scale by predicting the distributions of AR , PA and D_{est} from the observed
425 empirical values using Generalized Linear Models for Spatial Stream Networks
426 (GLMSSN^{20,21}). This was done using the ‘STARS’ toolset of ARCGIS v10.2 and the R
427 package ‘SSN’^{20,21}. We conducted a model selection procedure based on a comparison

428 of Akaike Information Criteria (AIC) estimated for several competing GLMSSNs.
429 These models were built by (i) assuming three geographic descriptors (i.e. topological
430 distance from the outlet, longitude and latitude) as explanatory variables, and (ii)
431 choosing a tail-down covariance structure model among the following ones:
432 exponential, Matérn, spherical, linear-with-sill and Epanechnikov. As the number of
433 explanatory variables differed among the GLMSSNs we built, we used the maximum
434 likelihood estimation method for each GLMSSN, so as to allow AIC-based model
435 comparisons. For each common species and genetic index, the best model had the
436 lowest AIC score (see Supplementary File 1 for raw results). This model was used to
437 estimate the slope and the significance of the relationships between explanatory
438 variables and each genetic index, so as to test whether or not spatial patterns of
439 intraspecific genetic diversity can be detected. We finally used predictions from the best
440 models to produce krigged maps for each common species and each genetic index, so as
441 to visually represent the spatial distribution of intraspecific genetic diversity across the
442 whole river drainage, and to visually highlight hot- and cold-spots of intraspecific
443 genetic diversity. We also calculated Pearson's correlation coefficients between values
444 calculated at the site level for each genetic index (i.e. AR , PA and D_{est}) for each pair of
445 common species, so as to test for spatial congruency in patterns of genetic diversity
446 among common species.

447

448 *Identification of irreplaceable sites.* We then tested whether conservation targets (i.e.
449 the percentage of total number of conservation units to be present in the final
450 conservation solution) and analytical approaches (i.e. species-specific or species-pooled
451 analyses) influence the identification of irreplaceable sites. SCP methods traditionally
452 use species presence/absence data as input data to identify irreplaceable sites for the

453 conservation of taxonomic diversity at the community level (i.e. sites that cannot be
454 excluded from an optimal selection of sites for conservation)⁴⁴. Here, we replaced
455 species presence/absence data by alleles' presence-absence data to identify irreplaceable
456 sites for the conservation of intraspecific genetic diversity of each species in the river
457 drainage. We used the program Marxan v2.1⁴⁴ and genetic data from the common
458 species to identify, for each common species independently, an optimal set of sites that
459 best represent at least 50, 75, 90 or 100% of the total number of alleles present in the
460 whole riverscape at a minimum "cost", which corresponds to four conservation targets.
461 Given the lack of ground estimates for conservation cost, we used a constant cost *per*
462 site, so our objective translated into identifying the minimum number of sites that
463 represent a given proportion of intraspecific genetic diversity (i.e. 50, 75, 90 or 100% of
464 the total pool of alleles represented at least once⁴⁵). We arbitrarily choose the 50, 75 and
465 100% conservation targets to explore how the proportion of alleles to protect affects the
466 selection of irreplaceable sites. We additionally tested the 90% conservation target, as it
467 corresponds to a threshold target being typically assumed in *ex-situ* conservation
468 plans^{17,18}. In order to estimate the relevance of each site to preserve a given proportion
469 of the allelic diversity in the river basin, we used two different methods. In case of
470 100% of allelic diversity, we used the traditional irreplaceability measure reported by
471 Marxan, which ranges between 100% (highly irreplaceable) and 0% (not irreplaceable).
472 This measure is estimated by running the optimization algorithm a number of times
473 (N=100 runs in our case) and then computing the frequency of selection of each site
474 within the solutions obtained. Sites with unique allelic composition will be selected
475 across all runs and reported as highly irreplaceable, whereas sites with more common
476 alleles, replaceable by other sites with the same alleles, will appear poorly irreplaceable.
477 For the other conservation targets, we selected a random pull of 50, 75 and 90% of the

478 total number of alleles existing at the basin level and for each species. We then ran the
479 optimization algorithm to identify the minimum set of sites representing this particular
480 selection of alleles. Given that the analyses were run only on a subset of alleles, we
481 replicated this process 100 times to minimize the effect of the arbitrary selection of
482 alleles. For each subset of alleles, we ran the Marxan procedure as explained above (e.g.
483 constant cost and 100 runs for each) and retained the best solution for subsequent
484 analyses (solution with the lowest value for the objective function). We then calculated
485 the irreplaceability as the frequency of selection of each site within the 100 random pull
486 of alleles. For a given species and a given conservation target, we considered a site as
487 irreplaceable for genetic diversity conservation when it displayed an irreplaceability
488 value of 100%. We selected such a high threshold so as to be conservative. We tested
489 and compared visually how the proportions of irreplaceable sites vary among species
490 and conservation targets.

491 To test how pooling data from several species affect the identification of
492 irreplaceable sites, we further performed a “pooled” analysis, in which all alleles found
493 for each common species at a site were pooled together in a single input dataset. We
494 then selected a random pull of 30, 50, 75, 90 and 100% of the total number of alleles
495 existing at the basin level (all common species confounded), and performed 100 Marxan
496 runs *per* conservation targets to identify the minimum set of sites representing these
497 particular selections of alleles.

498

499 A real conservation-oriented study using SCP approaches

500

501 In this second step, we (i) explored the spatial distribution of irreplaceable sites,
502 (ii) tested whether conservation solutions found for each species are congruent among

503 species, (iii) tested whether solutions found for one species can be used as a surrogate
504 for other species, and (iv) tested whether –or not– irreplaceable sites are correctly
505 predicted by classical indices of genetic diversity. In this step, we included the two rare
506 species, since congruency and surrogacy are particularly important to measure for rare
507 species for which data are more difficult to collect. We therefore focus more specifically
508 on the comparisons implying common vs. rare species.

509

510 *Identification of irreplaceable sites for genetic conservation.* We focused only on
511 irreplaceable sites identified for the 90% target and used the program Marxan as
512 described above to identify these sites for *L. burdigalensis* and *P. toxostoma*
513 independently, in addition of the four common species.

514 We mapped these irreplaceable sites (for the six species pooled or independently)
515 on the river network, so as to test (i) whether or not specific areas harboured more
516 irreplaceable sites (e.g. upstream areas that are generally thought to be of high
517 conservation priority²⁵) and (ii) if irreplaceable sites are spatially congruent among
518 species and, most notably, among common and rare species. In addition, we ran GLMs
519 (assuming a binomial error terms distribution) including whether or not a site has been
520 designated as an irreplaceable site at the 90% target as a binomial dependent variable,
521 and distance to the outlet of sampling sites and betweenness centrality values^{46,47} for
522 each sites explanatory variables. Betweenness centrality is an index quantifying the
523 positional importance of each sampling site within the river basin^{28,29}. The significance
524 of each term was tested at the $\alpha=0.05$ threshold.

525

526 *Surrogacy in irreplaceable sites among species.* We then estimated the levels of
527 surrogacy among species by calculating the percentage of the total number of alleles

528 observed for a given species that is covered by irreplaceable sites identified for another
529 species. Although surrogacy was calculated for all species pairs, we specifically focused
530 on rare species by calculating (i) the percentage of the total number of alleles observed
531 for rare species covered by all the irreplaceable sites identified for all the common
532 species, and (ii) the percentage of the total number of alleles observed for common
533 species covered by all the irreplaceable sites identified for the rare species.

534

535 *Relationships between irreplaceable sites and indices of genetic diversity.* To test the
536 ability of classical genetic indices to predict the propensity for a site to be irreplaceable
537 from a conservation viewpoint, we ran for each species Generalized Linear Models
538 (GLMs, with a binomial error terms distribution) including whether or not a site has
539 been designated as an irreplaceable site at the 90% target as a binomial dependent
540 variable, and AR , PA and D_{est} as explanatory variables. We tested the significance of
541 each term at the $\alpha=0.05$ threshold. Explanatory variables were centred and scaled, in
542 order to compare the relative strength of the predictors among species.

543

544 **Acknowledgements**

545

546 We are very grateful to all the ONEMA staff that contributed to the field
547 sampling, in particular Laurence Blanc, Leslie Faggiano, Charlotte Evangelista,
548 Christine Lauzeral, Loïc Tudesque, Roselyne Étienne, Sébastien Villéger, Julien
549 Cucherousset, Vincent Dubut, Alain Blanchet and South-Western France ONEMA staff
550 are thanked for their help in the field. Roselyne Étienne and Vincent Dubut are also
551 thanked for their help with molecular analyses. Lisa Fourtune is acknowledged for
552 having calculated the betweenness centrality values of our river network. We also thank

553 Marie-Hélène Lizée for her help with GIS tools and Camille Pagès for her comments.
554 Further, the authors thank the “Génopole Toulouse” for help with genotyping and Keoni
555 Saint-Pée for correcting the English. IPV was financially supported by a MESR
556 (“Ministère de l'Enseignement Supérieur et de la Recherche”) PhD scholarship during
557 this study. VH was funded by the National Environmental Research Program Northern
558 Australia Hub and a “Ramon y Cajal” contract (RYC-2013-13979) funded by the
559 Spanish government. This study is part of the European project “IMPACT”. This
560 project has been carried out with financial support from the Commission of the
561 European Communities, specific RTD programme “IWRMNET”. This work has been
562 done in two research units (EDB & SETE) that are part of the “Laboratoire
563 d'Excellence” (LABEX) entitled TULIP (ANR-10-LABX-41).

564

565 **References**

566

- 567 1. Watson, J. E. M., Dudley, N., Segan, D. B. & Hockings, M. The performance and
568 potential of protected areas. *Nature* **515**, 67–73 (2014).
- 569 2. Filipe, A. F. *et al.* Selection of priority areas for fish conservation in Guadiana River
570 Basin, Iberian Peninsula. *Conserv. Biol.* **18**, 189–200 (2004).
- 571 3. Kirkpatrick, J. B. An iterative method for establishing priorities for the selection of
572 nature reserves: An example from Tasmania. *Biol. Conserv.* **25**, 127–134 (1983).
- 573 4. Margules, C. R. & Pressey, R. L. Systematic conservation planning. *Nature* **405**,
574 243–253 (2000).
- 575 5. Asmyhr, M. G., Linke, S., Hose, G. & Nipperess, D. A. Systematic conservation
576 planning for groundwater ecosystems using phylogenetic diversity. *PLoS ONE* **9**,
577 e115132 (2014).

- 578 6. Buerki, S. *et al.* Incorporating evolutionary history into conservation planning in
579 biodiversity hotspots. *Philos. Trans. R. Soc. B Biol. Sci.* **370**, 20140014–20140014
580 (2015).
- 581 7. Hoffmann, A. A. & Sgrò, C. M. Climate change and evolutionary adaptation.
582 *Nature* **470**, 479–485 (2011).
- 583 8. Carroll, S. P. *et al.* Applying evolutionary biology to address global challenges.
584 *Science* **346**, 1245993–1245993 (2014).
- 585 9. Mittell, E. A., Nakagawa, S. & Hadfield, J. D. Are molecular markers useful
586 predictors of adaptive potential? *Ecol. Lett.* **18**, 772–778 (2015).
- 587 10. Rey, O., Danchin, E., Mirouze, M., Loot, C. & Blanchet, S. Adaptation to Global
588 Change: A Transposable Element–Epigenetics Perspective. *Trends Ecol. Evol.* **31**,
589 514–526 (2016).
- 590 11. Pritchard, J. K., Stephens, M. & Donnelly, P. Inference of population structure
591 using multilocus genotype data. *Genetics* **155**, 945–959 (2000).
- 592 12. Diniz-Filho, J. A. F. *et al.* Planning for optimal conservation of geographical
593 genetic variability within species. *Conserv. Genet.* **13**, 1085–1093 (2012).
- 594 13. Grantham, H. S., Pressey, R. L., Wells, J. A. & Beattie, A. J. Effectiveness of
595 Biodiversity Surrogates for Conservation Planning: Different Measures of
596 Effectiveness Generate a Kaleidoscope of Variation. *PLoS ONE* **5**, e11430 (2010).
- 597 14. Hermoso, V., Januchowski-Hartley, S. R. & Pressey, R. L. When the suit does not
598 fit biodiversity: Loose surrogates compromise the achievement of conservation
599 goals. *Biol. Conserv.* **159**, 197–205 (2013).
- 600 15. Andrew, R. L. *et al.* A road map for molecular ecology. *Mol. Ecol.* **22**, 2605–2626
601 (2013).

- 602 16. Pauls, S. U. *et al.* Integrating molecular tools into freshwater ecology: developments
603 and opportunities. *Freshw. Biol.* **59**, 1559–1576 (2014).
- 604 17. Frankham, R., Briscoe, D. A. & Ballou, J. D. *Introduction to conservation genetics*.
605 (Cambridge University Press, 2002).
- 606 18. Neel, M. C. & Cummings, M. P. Effectiveness of conservation targets in capturing
607 genetic diversity. *Conserv. Biol.* **17**, 219–229 (2003).
- 608 19. Jost, L. G(ST) and its relatives do not measure differentiation. *Mol. Ecol.* **17**, 4015–
609 4026 (2008).
- 610 20. Peterson, E. & Ver Hoef, J. M. V. STARS: STARS: An ArcGIS toolset used to
611 calculate the spatial information needed to fit spatial statistical models to stream
612 network data. *J. Stat. Softw.* **56**, 1–17 (2014).
- 613 21. Ver Hoef, J., Peterson, E., Clifford, D. & Shah, R. SSN: An R Package for Spatial
614 Statistical Modeling on Stream Networks. *J. Stat. Softw.* **56**, 1–43 (2014).
- 615 22. Fortuna, M. A., Albaladejo, R. G., Fernandez, L., Aparicio, A. & Bascompte, J.
616 Networks of spatial genetic variation across species. *Proc. Natl. Acad. Sci.* **106**,
617 19044–19049 (2009).
- 618 23. Taberlet, P. *et al.* Genetic diversity in widespread species is not congruent with
619 species richness in alpine plant communities. *Ecol. Lett.* **15**, 1439–1448 (2012).
- 620 24. Paz-Vinas, I., Loot, G., Stevens, V. M. & Blanchet, S. Evolutionary processes
621 driving spatial patterns of intraspecific genetic diversity in river ecosystems. *Mol.*
622 *Ecol.* **24**, 4586–4604 (2015).
- 623 25. Finn, D. S., Bonada, N., Múrria, C. & Hughes, J. M. Small but mighty: headwaters
624 are vital to stream network biodiversity at two levels of organization. *J. North Am.*
625 *Benthol. Soc.* **30**, 963–980 (2011).

- 626 26. Paz-Vinas, I. & Blanchet, S. Dendritic connectivity shapes spatial patterns of
627 genetic diversity: a simulation-based study. *J. Evol. Biol.* **28**, 986–994 (2015).
- 628 27. Thomaz, A. T., Christie, M. R. & Knowles, L. L. The architecture of river networks
629 can drive the evolutionary dynamics of aquatic populations. *Evolution* **70**, 731–739
630 (2016).
- 631 28. Altermatt, F. Diversity in riverine metacommunities: a network perspective. *Aquat.*
632 *Ecol.* **47**, 365–377 (2013).
- 633 29. Fourtune, L., Paz-Vinas, I., Loot, G., Prunier, J. G. & Blanchet, S. Lessons from the
634 fish: a multi-species analysis reveals common processes underlying similar species-
635 genetic diversity correlations. *Freshw. Biol.* (2016). doi:10.1111/fwb.12826
- 636 30. Mimura, M. *et al.* Understanding and monitoring the consequences of human
637 impacts on intraspecific variation. *Evol. Appl.* **10**, 121–139 (2017).
- 638 31. Siefert, A. *et al.* A global meta-analysis of the relative extent of intraspecific trait
639 variation in plant communities. *Ecol. Lett.* **18**, 1406–1419 (2015).
- 640 32. Kottelat, M. & Freyhof, J. *Handbook of European freshwater fishes*. (Publications
641 Kottelat, 2007).
- 642 33. Poulet, N., Beaulaton, L. & Dembski, S. Time trends in fish populations in
643 metropolitan France: insights from national monitoring data. *J. Fish Biol.* **79**, 1436–
644 1452 (2011).
- 645 34. Paz-Vinas, I., Quéméré, E., Chikhi, L., Loot, G. & Blanchet, S. The demographic
646 history of populations experiencing asymmetric gene flow: combining simulated
647 and empirical data. *Mol. Ecol.* **22**, 3279–3291 (2013).
- 648 35. Paz-Vinas, I. *et al.* Combining genetic and demographic data for prioritizing
649 conservation actions: insights from a threatened fish species. *Ecol. Evol.* **3**, 2696–
650 2710 (2013).

- 651 36. Aljanabi, S. M. & Martinez, I. Universal and rapid salt-extraction of high quality
652 genomic DNA for PCR-based techniques. *Nucleic Acids Res.* **25**, 4692–4693
653 (1997).
- 654 37. Van Oosterhout, C., Hutchinson, W. F., Wills, D. P. M. & Shipley, P. micro-
655 checker: software for identifying and correcting genotyping errors in microsatellite
656 data. *Mol. Ecol. Notes* **4**, 535–538 (2004).
- 657 38. Jombart, T. adegenet: a R package for the multivariate analysis of genetic markers.
658 *Bioinformatics* **24**, 1403–1405 (2008).
- 659 39. Rousset, F. genepop'007: a complete re-implementation of the genepop software for
660 Windows and Linux. *Mol. Ecol. Resour.* **8**, 103–106 (2008).
- 661 40. Szpiech, Z. A., Jakobsson, M. & Rosenberg, N. A. ADZE: a rarefaction approach
662 for counting alleles private to combinations of populations. *Bioinformatics* **24**,
663 2498–2504 (2008).
- 664 41. Petit, R. J., El Mousadik, A. & Pons, O. Identifying Populations for Conservation
665 on the Basis of Genetic Markers. *Conserv. Biol.* **12**, 844–855 (1998).
- 666 42. Kalinowski, S. T. Counting alleles with rarefaction: Private alleles and hierarchical
667 sampling designs. *Conserv. Genet.* **5**, 539–543 (2004).
- 668 43. Winter, D. J. mmod: an R library for the calculation of population differentiation
669 statistics. *Mol. Ecol. Resour.* **12**, 1158–1160 (2012).
- 670 44. Ball, I. R., Possingham, H. P. & Watts, M. in *Spatial conservation prioritisation:*
671 *Quantitative methods and computational tools* (eds. Moilanen, A., Wilson, K. A. &
672 Possingham, H. P.) **Chapter 14**, 185–195 (Oxford Iniversity Press, Oxford, UK,
673 2009).

674 45. Hermoso, V. & Kennard, M. J. Uncertainty in coarse conservation assessments
675 hinders the efficient achievement of conservation goals. *Biol. Conserv.* **147**, 52–59
676 (2012).

677 46. Freeman, L. C. A set of measures of centrality based on betweenness. *Sociometry*
678 **40**, 35 (1977).

679 47. Estrada, E. & Bodin, Ö. Using network centrality measures to manage landscape
680 connectivity. *Ecol. Appl.* **18**, 1810–1825 (2008).

681

682 **Author Contributions:** The study was designed by SB, GL and IPV. IPV and SB
683 wrote the manuscript with the help of VH, GG, NP and GL. IPV, SB, GL and CV
684 collected the samples. GL and CV produced the genetic data. IPV, VH and SB
685 conducted the population genetic and statistical analyses.

686

687 **Competing Financial Interests statement:** Authors declare not having any competing
688 financial interest.

689

690 **Figures legends**

691

692 **Figure 1**

693 Spatial distribution of observed (coloured circles) and interpolated (coloured lines)
694 values of AR , PA and D_{est} (A, B, C respectively in the figure) for *Squalius cephalus*,
695 *Gobio occitaniae*, *Barbatula barbatula*, and *Phoxinus phoxinus* (1, 2, 3, 4 respectively
696 in the figure) obtained with GLMSSNs. The width of coloured lines is inversely related
697 to the prediction standard error. The cursor on the vertical coloured scale indicates the
698 mean value of AR , PA and D_{est} . The slope (β) of each explanatory variable (i.e.
699 topological distance to the outlet, longitude and latitude) and its significance is also
700 reported. N.I. indicates that the explanatory variable has not been included in the model;
701 * p-value <0.05; ** p-value <0.01; *** p-value <0.001.

702

703 **Figure 2**

704 Percentage of irreplaceable sites identified by Marxan for conservation targets of 50, 75,
705 90 and 100% of the total number of alleles present in the river basin for each common
706 species and for a pooled analysis in which all alleles from all common species were
707 pooled together. N_{SITES} represents the number of sites included in the Marxan analyses.

708

709 **Figure 3**

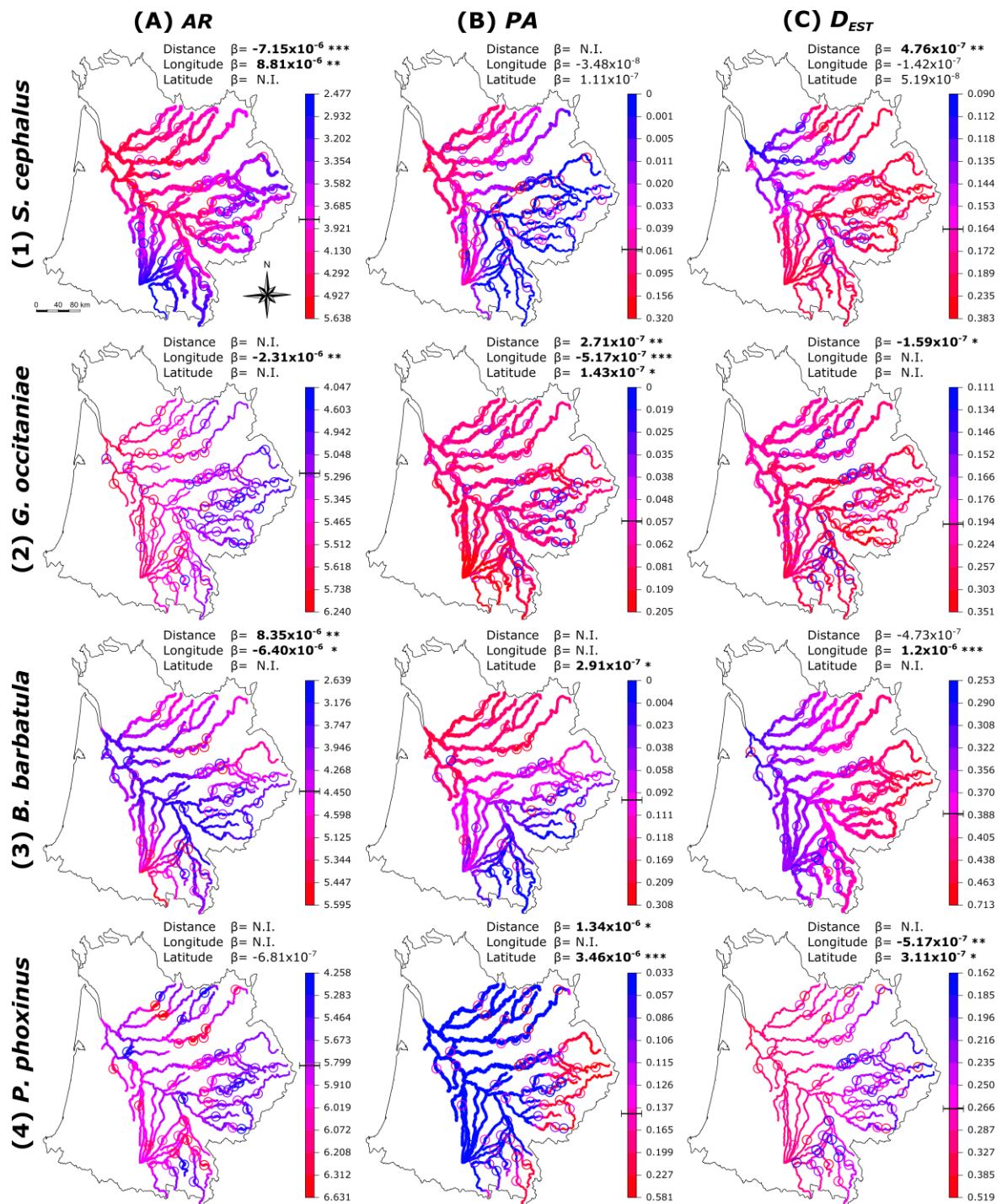
710 Irreplaceable sites that have been identified for each species by Marxan for preserving
711 90% of the total number of alleles present in the river basin (red-filled circles).

712

713 **Figure 4**

714 Irreplaceable sites that have been identified by Marxan for at least one (unicoloured-
715 filled points), two (black dotted circles surrounding bicoloured points) or three (black
716 bolded circle surrounding a tricoloured point) species, assuming a conservation target of
717 90% of the total number of alleles present in the river basin when considering the six
718 species.
719

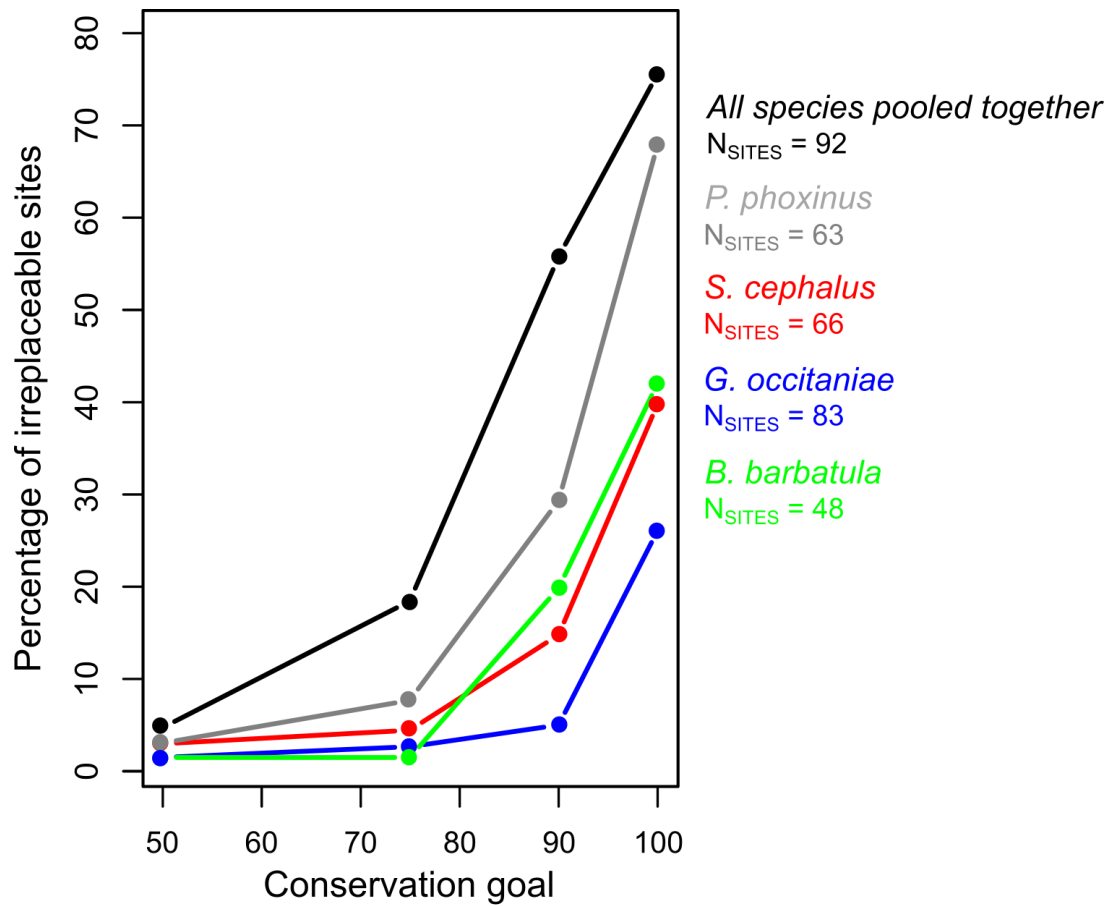
720 **FIGURE 1**



721

722

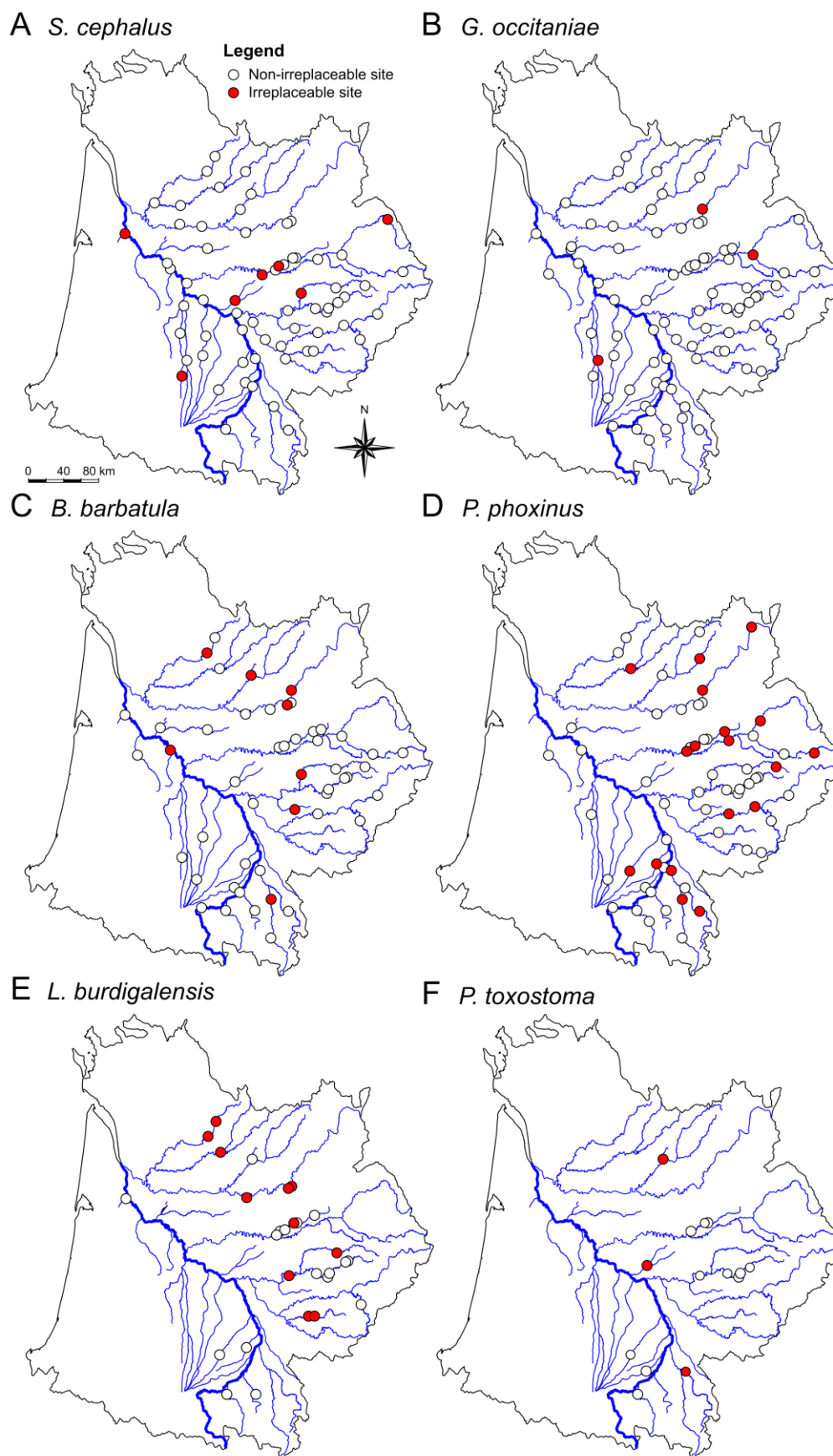
723 **FIGURE 2**

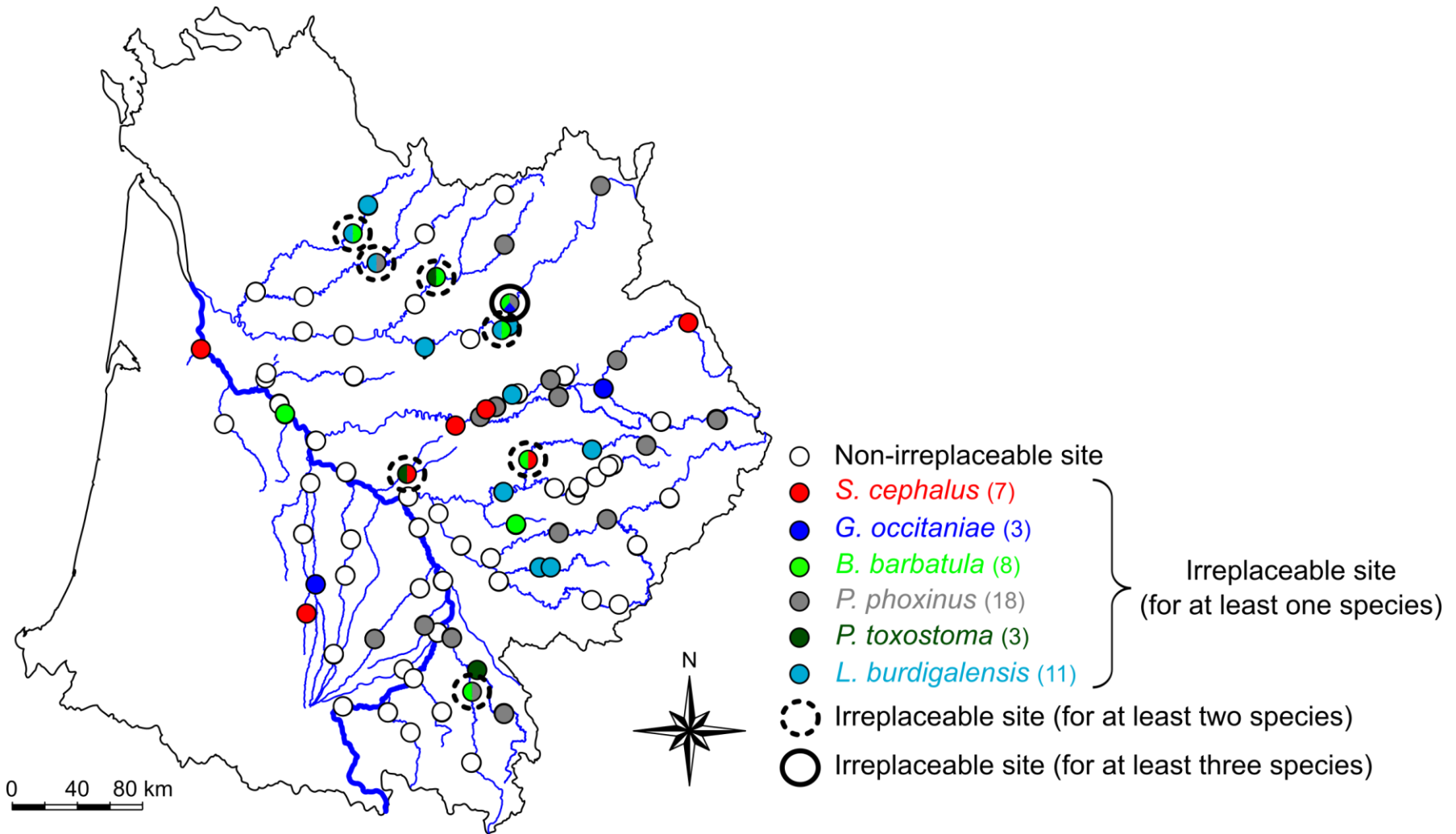


724 (Percentage of the total number of alleles to protect)

725

726 **FIGURE 3**





730 **TABLE 1:** Table reporting for each species the percentages of their total number of alleles that are covered by the sampling sites having
 731 been identified as irreplaceable sites for the other species (considering the 90% conservation target).

732

Proportion of alleles from (below) that fell within irreplaceable sites from (right):	<i>Squalius cephalus</i>	<i>Gobio occitaniae</i>	<i>Barbatula barbatula</i>	<i>Phoxinus phoxinus</i>	<i>Leuciscus burdigalensis</i>	<i>Parachondrostoma toxostoma</i>	Common species	Rare species
<i>Squalius cephalus</i>	90.00	17.66	61.52	56.52	61.60	39.73	—	65.18
<i>Gobio occitaniae</i>	65.60	90.00	66.76	79.58	70.27	50.97	—	73.40
<i>Barbatula barbatula</i>	49.79	31.65	90.00	79.57	76.78	38.95	—	79.40
<i>Phoxinus phoxinus</i>	48.25	42.60	68.43	90.00	63.49	44.61	—	68.88
<i>Leuciscus burdigalensis</i>	44.73	0	66.13	67.09	90.00	36.42	79.87	—
<i>Parachondrostoma toxostoma</i>	78.55	0	78.55	50.00	67.14	90.00	90.00	—
<i>Average over all other species</i>	55.58	18.38	68.28	66.55	67.86	42.14		

733