Environmental cholera (*Vibrio cholerae*) dynamics in an estuarine system in southern

coastal Ecuador

RH: Dynamics of Vibrio cholerae in southern Ecuador

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Abstract

Cholera emergence is strongly linked to local environmental and ecological context. The 1991-

2004 pandemic emerged in Perú and spread north into Ecuador's El Oro province, making this a

key site for potential re-emergence. Machala, El Oro, is a port city of 250,000, near the Peruvian

border. Many livelihoods depend on the estuarine system, from fishing for subsistence and trade,

to domestic water use. In 2014, we conducted biweekly sampling for 10 months in five estuarine

locations, across a gradient of human use, and ranging from inland to ocean. We measured

water-specific environmental variables implicated in cholera growth and persistence: pH,

temperature, salinity, and algal concentration, and evaluated samples in 5 months for pathogenic

and non-pathogenic *Vibrio cholerae*, by polymerase chain reaction (PCR). We found

environmental persistence of pandemic strains O1 and O139, but no evidence for toxigenic

strains. Cholera presence was coupled to algal and salinity concentration, and sites exhibited

considerable seasonal and spatial heterogeneity. This study indicates that environmental

conditions in Machala are optimal for cholera re-emergence, with risk peaking during

September, and higher risk near urban periphery low-income communities. This highlights a

need for surveillance of this coupled cholera– estuarine system to anticipate potential future

cholera outbreaks.

Introduction

Cholera remains a severe global threat to public health and development efforts (WHO 2013). According to the World Health Organization, the burden of cholera is at least 100 times greater than current estimates (Zuckerman et al. 2007; WHO 2013), with 120,000 deaths and 3-5 million cases each year worldwide. Previous studies suggest that outbreaks of cholera can be explained by oceanographic variables (e.g., sea surface temperature, pH, salinity) and phytoplankton blooms, indicating the potential to predict disease outbreaks (Jutla et al. 2010; Jutla et al. 2013). A recent analysis of global cholera pandemics indicates that cholera outbreaks originate in coastal regions, often during flood events, before spreading inland (Jutla et al. 2010). Our own previous work suggests that both current and future coastal hotspots of cholera transmission are far larger than current surveillance efforts can capture, with considerably higher potential exposure than previously estimated (Escobar et al. 2015). Estuarine systems are a natural intersection of coastal oceanographic conditions and human use; as productive systems for fisheries, port locations for transport, and rich riparian soils, they are a highly exposed interface for humans. Particularly because coastal estuarine systems often represent subsistence-level dependence on the interface, in terms of artisanal fisheries, a higher likelihood of direct water use, and simply greater physical exposure by proximity, it is also the most vulnerable of populations that are most likely to be exposed to pathogens and the most flooding prone areas in the world (Nicholls 1995; Dixon et al. 2006; De Sherbinin et al. 2007; Hanson et al. 2011; Hallegatte et al. 2013; Cai et al. 2014).

The causative agent of human cholera, *Vibrio cholerae*, is thought to originate from estuarine waters, based on phylogenetic information and its physiological requirements for growth and persistence (Colwell and Huq 1994; Colwell 2004). *Vibrio cholerae* is endemic in

the Bay of Bengal (Bangladesh and India) and along coastal areas in Latin America (Lipp et al.

2002; Mutreja et al. 2011), and it persists environmentally in riverine, estuarine, and coastal

waters around the world (Lipp et al. 2002), Cholera epidemics have been found to follow

coastlines (Colwell 1996), and *V. cholerae* can be transmitted to humans via a wide range of

marine organisms, including zooplankton, aquatic plants, shellfish, and fish (Vezzulli et al.

2010).

78   Ecuador is a critical location to understand cholera and other climate and water-sensitive

diseases due to its (1) high potential for cholera outbreaks and the high incidence of other

climate-sensitive infectious diseases (e.g., leptospirosis, dengue), and (2) the strong influence of

oceanographic conditions on local climate and flooding during El Niño events (Rossel et al.

1996; Rossel and Cadier 2009; Hanson et al. 2011; Hallegatte et al. 2013; Cai et al. 2014).

Indeed, in January 1991, cholera re-emerged in Latin America after more than a century without

cases (Lacey 1995). In Ecuador, the 1991 cholera epidemic emerged in the south of the country

from a small fishing village in El Oro Province, and it is suspected that a fisherman introduced

the index case was traveling north from Perú (Dixon et al. 2006). From 1991 to 2004 over 90,000

cases of cholera were reported in Ecuador, with most cases from coastal provinces. El Oro and

Guayas provinces, located in southern coastal Ecuador, encompassed one of two disease

epicenters in the country. Recent studies suggest there is a high risk of a second epidemic in

Ecuador due to the presence of important risk factors including the growth of vulnerable urban

populations, decreased investment in cholera surveillance and prevention programs, increased

flood risk associated with climate change, and a street food culture that includes eating raw

shellfish (ceviche) (Malavade et al. 2011). In addition, Guayaquil (Guayas Province), the largest

city in Ecuador, has been identified as the third most vulnerable city in the world to future flood

risk (Hallegatte et al. 2013). Furthermore, it has been found that in populations with a high

prevalence of blood group O, such as in Latin America, illness from cholera is more severe, and

the requirements for rehydration and hospitalization of infected individuals are considerably

higher (Swerdlow et al. 1994; Nelson et al. 2009). Given these conditions, there is compelling

evidence that people in southern coast of Ecuador are a high-risk population and there is a

critical need for active cholera surveillance in this region.

To address this, we evaluated local variability in the presence of cholera in the estuarine

environment surrounding the city of Machala, El Oro province, a site identified as a current and

future coastal cholera hotspot (Escobar et al. 2015). We selected five sampling sites associated

with estuarine water access in Machala, Ecuador, representing a range of economic and human

activity conditions, in addition to different proximity to the ocean. Using water sampling

methodology, coupled with laboratory identification of *Vibrio cholerae* bacteria, we assessed the

local environmental and pathogenic conditions over a period of ten months. Strengthening

climate and water-sensitive infectious disease surveillance systems (WHO 2003; Zuckerman et

al. 2007) and further understanding of the role of environmental factors in disease outbreak and

transmission over time and space (Sedas 2007; Akanda et al. 2013) are urgently needed to target

cholera and other climate and water sensitive diseases.

Methodology

Study Site

Machala is a port city of approximately 250,000 inhabitants, with major economic activities

stemming from agriculture (bananas), aquaculture (shrimp farming), and fishing/shellfish

collection, both small-scale and semi-industrial scale. Five sampling sites were established

within the Machala estuarine system (Figure 1), selected for maximum heterogeneity, to include

highly built urban areas, ports, mangrove, and coastal sampling areas. The five sites were: Isla

Jambelí, Boca del Macho, Puerto Bolívar Boca, Puerto Bolívar Adentro, and Héroes de Jambelí.

Isla Jambelí site is on the outer edge of the coastal draining estuary, and the entrance to Jambelí

is interspersed with mangroves and shrimp farms. Boca del Macho is the open edge of the inner

estuary, in open water on shallow sand, with mangroves. Puerto Bolívar Boca is near the mouth

of the open harbor, characterized by heavy boat traffic, commercial fishing, and residences lining

the waterway, with mangroves and shrimp farming on the far side of the waterway. Puerto

Bolívar Adentro is further into the city, along the estuary, characterized by residential low-

income housing, with shrimp farms and mangroves across the Héroes waterway. Heroes de

Jambelí is the most inland site, characterized by low income and poor quality housing built along

mangroves at the edge of the city; outflow from the houses is visible directly into the water

(Figure 1). The port city of Machala is an important sentinel surveillance site, due to its location

along the Pan American highway, approximately 80km north of the Peruvian border, facilitating

significant movement of people and potential pathogens by land and sea.

Water sampling

At each of the five study sites (Figure 1), water sampling was conducted at high tide, twice

monthly along a transect with 3 sub-sites spaced 250 m apart, and 3 replicates per subsite. Three

1L surface water samples per subsite were collected in sterile polypropylene bottles, and placed

in coolers with ice for transport for the laboratory. For environmental sampling, using a YSI

water probe* (600 XLM V2 Sonde), we recorded Surface Temperature (ºC), Conductivity, pH,

Salinity, and Optic-T BGA PE (Phycoerythrin) [blue-green algae] (cells/ml, which we converted

to cells/µl for ease of visualization) at each end of the transect.

Laboratory analyses

Water samples were transferred to the laboratory in coolers for *Vibrio cholerae* testing and were

processed within 24 hours of collection. For laboratory analysis, a 1L water sample was filtered

through a Whatman membrane No. 1 and 0.22 µm membrane (Millipore) by vacuum. Then,

10mL of Phosphate Buffered Saline (PBS) (pH 7.4) was pipetted onto the retained contents on

the membrane and gently washed by pipette 15x. The PBS was left on the membrane to incubate

at room temperature for 15 minutes prior to collection in 50mL conical tube.

DNA isolation and PCR

Genomic DNA was extracted from bacterial pellet of the previous step with a QIAamp DNA

mini kit (Qiagen), following manufacture instructions. Diagnosis of cholera serogroups and the

detection of toxigenic genes were performed each by duplex PCR. Table 1 describes primers sets

used to amplify the *rfb* region of O1 and O139 serogroups and the toxin subunit A (ctxA) and

toxin coregulated pilus (tcpA) genes. For both duplex PCRs, master mix was as follows: 0.05

U/ul of JumpStart REDTaq DNA Polymerase (Sigma), 1X buffer, 0.2 mM dNTPs, 0.2 mM of

each primer set, 1 µl of template and ultrapure water to a final volume of 25 µl. The

amplification program for diagnosis of serogroups was adapted from Hoshino et al.(Hoshino et

al. 1998), using the following conditions: 5 minutes at 94°C, 35 cycles of 94°C for 1 minute,

55°C for 1 minute and 72°C for 1 minute and final extension of 72°C for 7 minutes. Positive

samples for either or both serogroups were subjected to toxigenic genes duplex PCR. The

amplification program was according to conditions described in Kumar et al.(Kumar et al. 2010):

3 minutes at 94°C, 30 cycles of 94°C for 30 seconds, 59°C for 30 seconds and 72°C for 1.2

minutes, and final extension of 72°C for 10 minutes. PCR products were resolved in a 2%

agarose gel and sequenced to verify gene amplification.

Statistical analyses

As the data were not normally distributed, we conducted non-parametric tests throughout. We

characterized the sampling sites for each environmental variable: Temperature, pH, Salinity, and

BGA, conducting Kruskal-Wallis rank sum tests on site means, and on monthly means. We then

examined whether *V. cholerae* prevalence at sites, and strain (i.e., O1, O139) prevalence

separately, was associated with environmental variables using a series of non-parametric

Kendall's tau correlations.

Results

Environmental characteristics

The probe recorded a range of 9-104 readings at each sub-site biweekly for 10 months. We

pooled all readings by month for analyses. Our sample sites differed significantly in

environmental characteristics (Figure 2), as shown by a series of Kruskal-Wallis rank sum tests

(Temperature: χ^2= 206.19, df = 4, p < 0.0001; Salinity: χ^2= 2257.5, df = 4, p < 0.0001; pH: χ^2=

1347.3, df = 4, p < 0.0001; BGA χ^2= 1824.8, df = 4, p < 0.0001). We found that Héroes de

Jambelí, the most inland site, had the highest BGA, and that Isla de Jambelí, the most coastal

site, had the highest salinity; while there were statistical differences between all sites in all the

environmental variables, there were no clear outliers in the other variables. Our sample sites

exhibited significant change in environmental characteristics across months (Figure 3), as shown

by a series of Kruskal-Wallis rank sum tests (Table 2). Temperature was lowest in August for all

sites – likely reflecting pacific upwelling, which cools the water, regardless of air temperature.

Salinity at the most inland site, Héroes de Jambelí, was consistently lowest, and showed the

smallest change across months, while the other sites had a decrease in salinity in May, then a rise

from July-December. Isla de Jambelí had the highest salinity, reflecting its location on the most

coastal site. BGA was highest at the most inland site, Héroes de Jambelí, peaking in May,

lagging temperature by a month. BGA shows the least temporal or spatial clustered pattern and

has no obvious seasonality across the year. Heroes de Jambeli, however, registered the highest

BGA values during the study (~25,000). pH appears to peak in December-January across all

sites, with a decrease in July-August; the coastal and inland sites showed low pH values across

seasons, while Boca del Macho registered consistently high pH values across months.

Laboratory analyses

Of a total of 405 individual water samples, collected between May – September, 382 were

diagnosed by PCR. We found 139 (36.4%) samples positive for *V. cholerae*, and 243 (64%)

negative. We found both O1 and O139 serogroups of *V. cholerae* present in the estuarine system

studied in Machala, Ecuador. Serogroup O139 was predominant; 118 (83.5%) samples were

O139, and 51 (35.3%) were O1 (30 samples contained both). We were able to detect *V.*

cholerae during each of the 5 months of sampling, nevertheless we found that prevalence

decreased drastically in July (Figure 4). By sequencing the samples, we confirmed that the PCR

protocol applied was proper for detection of *V. cholerae* serogroups O1 and O139 strains.

Vibrio cholerae **characteristics**

We pooled water samples within sites, to derive monthly *V. cholerae* prevalences across and

within sites (prevalence = positive/total samples tested). Overall monthly prevalence of *V.*

cholerae ranged from 0.3 (n=68) in July to 0.58 (n=45) in September, with site prevalence

ranging 0-1, with a mean monthly site prevalence of 0.35 (Figure 4A). Individual strain

prevalence was generally higher for O139 than O1, but we see that Puerto Bolívar Adentro and

Héroes de Jambelí were *V. cholerae* positive in every month and also had higher prevalences

than the other sites (Figure 4B and 4C). We found that the prevalence of *V. cholerae*, and O139

and O1 strains separately, were significantly associated with higher BGA (blue-green algae

densities), and that prevalence of *V. cholerae* the O1 strain were significantly associated with

lower salinity. We found no significant association between prevalence and temperature or pH

(Table 3).

Discussion and Recommendations

We found evidence of an environmental reservoir of *V. cholerae* in the estuarine waters of

Machala, Ecuador, in 2014. We confirmed the presence of *V. cholerae*, including pandemic

strains O1 and O139. We cannot rule out ongoing toxigenic presence, but we did not detect it in

our samples.

Our sites exhibited considerable seasonal and spatial heterogeneity in environmental

variables and *V. cholerae* prevalence, with clear peaks (and troughs) during specific months. For

example, we found peak *V. cholerae* prevalence in September, with highest values in two sites:

– Héroes de Jambelí and Puerto Bolívar Adentro (Figure 1), these sites are characterized by low

income housing on the edge of the city, while being inland sites, facing mangroves and shrimp

farms, and were found to have *V. cholerae* present in every month sampled. The lowest *V.*

cholerae prevalence was found in July, in which only the two most inland sites had *V. cholerae*

detection. Water temperature had the clearest temporal pattern, falling rapidly through July,

likely corresponding to Pacific upwelling, cooling the waters, and increasing nutrients in the

system (Strutton et al. 2001). We found that there was lowest salinity in the most inland site,

Héroes de Jambelí, and a higher concentration of BGA than in other sites. This is in contrast to

Isla Jambelí, a small island community furthest from the mainland and closest to the ocean, with

high salinity due to its coastal location; however, it did not have lower BGA than other sites.

We found that the timing of *V. cholerae* was coupled to the environmental variables we

measured. For example, water temperature, BGA, and pH decreased in most sites through

July/August, so did the overall prevalence of *V. cholerae*, but we only demonstrated significant

associations between prevalence and site and month specific levels of salinity and BGA. Average

ocean salinity is around 35 ppt, while freshwater rivers average around 0.5 ppt; clearly in this

estuarine system, we see a mixed or brackish system, ranging from the lower average of around

15 ppt at our most inland site, to a high approaching 34 ppt at our coastal site. We detected *V.*

cholereae at a range of salinities, finding a negative correlation with increasing salinity,

suggesting that the lower salinity may provide a more suitable environment for the growth of *V.*

cholereae, but that even the higher salinity approaching ocean concentrations do not prevent that

growth. This finding is consistent with previous work demonstrating the suitability of coastal

oceans for *V. cholereae*(Strutton et al. 2001), but reveals a finer scale relationship with salinity as

we move inland in an estuarine system, up the gradient to fresh water.

BGA (blue-green algae; a.k.a. cyanobacteria) are photosynthetic prokaryotes that can be

found in freshwater, marine, and terrestrial environments (Stanier and Bazine 1977). The

photosynthetic pigments of cyanobacteria include chlorophyll-*a* and the phycobiliprotein

phycoerythrin. Here we use BGA values to characterize water features and because BGA has

been associated with *V. cholereae* persistence (Epstein 1993). However, BGA itself also poses a

significant threat to humans through its production of cyanotoxins. BGA toxins include

neurotoxins, hepatotoxins, cytotoxins, irritants and gastrointestinal toxins (Codd et al. 2005).

Among these chemicals, microsystin, is a known liver carcinogen (Hunter 1998), and has been

detected in marine ecosystems (Miller et al. 2010). Exposure to these via skin contact, inhalation,

or ingestion, can result in a range of effects, from skin irritation and conjunctivitis, to kidney

damage and respiratory arrest (Codd et al. 1999). The World Health Organization (WHO)

recognizes BGA blooms as an emerging public health risk, and recommends the development of

early warning systems to detect scums (World Health Organization 1999; Falconer 2001;

Manganelli et al. 2012). We argue that BGA related diseases should be included in differential

diagnosis in Héroes de Jambelí, particularly during May, when a BGA increase was evident in

this study.

Temperature increase, coupled with high nutrient load, low flow, and thermal

stratification, generally results in increased growth rates of cyanobacteria, and its dominance in

the phytoplankton community (Davis et al. 2009; Elliott 2010; Huber et al. 2012). This could

explain the high BGA values early in the year (Figure 3). In addition, warm temperatures

promote increases in the number of days where BGA biomass exceeds thresholds established by

WHO (Davis et al. 2009; Elliott 2012). High temperature influences water column stability and

mixing depth, producing favorable conditions for BGA blooms (Robarts and Zohary 1987; Stal

et al. 2003). This association of temperature increase with BGA blooms is consistent across

coastal, estuarine, and inland waters (Paerl 1988). Thus, long term monitoring to measure BGA

biomass should be considered at least in Heroes de Jambeli and Puerto Bolivar -the sites report

the highest BGA values (Figure 2), particularly considering that a rise in water temperature is

associated with BGA emergence (Wasmund 1997; Kanoshina et al. 2003; Suikkanen et al. 2007),

increasing the risk of BGA diseases with imminent future climate warming.

In Ecuador, seawater eutrophication is a public health problem (World Health

Organization 1999).In fact, several species of BGA have been identified in aquatic environments

in Ecuador (Gunkel and Casallas 2002; Nedbalová and Sklenár 2008; Ramírez-Luna et al. 2008).

Unfortunately, while there is an increasing recognition of the negative health effects of BGA

blooms, monitoring in coastal marine waters is rare, and efforts are strongly biased to freshwater

systems (Nedbalová and Sklenár 2008), making our exploration in estuarine areas a crucial

update to the status of BGA in the country. This study was conducted in an average climate year,

providing a preliminary framework for monitoring coupled *V. cholerae* – estuarine dynamics for

potential emergence of cholera outbreaks in the region. This is particularly useful baseline

information for anticipating El Niño years, extreme climate events associated with warming

temperatures of surface ocean water and increased rainfall and flooding events. Climate change

projections indicate that the frequency of extreme El Niño events will increase in the future, (Cai

et al. 2014) increasing the risk of water-borne diseases endemic in the region, such as cholera,

typhoid, and leptospirosis, and provides valuable information to add *V. cholerae* in the public

health agenda to consider infectious diseases beyond the already important vector-borne

diseases, such as dengue fever, chikungunya, zika, and malaria.

Indeed, by May 2016, two years after the initiation of this study, the first case of cholera

was reported in Machala, after approximately 12 years with no case reports in Ecuador (2016).

An immuno-compromised individual was confirmed positive for *V. cholerae* serotype O1 non-

toxigenic, by the National Public Health Research Institute of the Ministry of Health. Our team

diagnosed the patient using the same PCR assay described here. Although the source of the

infection was not confirmed, this case report suggests a worrisome link to the results of our

epidemiological survey, and merits further examination.

This study highlights the urgency for active epidemiological surveys and the need for

public health interventions to reduce the risk of water-borne pathogens in vulnerable populations

from a holistic social-ecological perspective. The community Héroes de Jambelí is a low-income

peri-urban settlement with less than 50 families, established informally in 2002. The community

continues to lack adequate access to piped water, sewerage, and garbage collection due to their

status as an illegal settlement. Simple bamboo homes have built over the mangrove system, with

direct discharge of wastewater into the estuary. At the same time, this community's livelihood

depends on artisanal fisheries (e.g., crabs, mollusks) from these same estuaries. This vulnerable

coupled human-natural system results in high risk of emerging epidemics from water-borne

pathogens.

The results of this study, coupled with ongoing and previously published remote sensing

and GIS assessments, will allow us to identify geographic areas for future *V. cholerae*

surveillance across coastal Ecuador. In the future, we anticipate sampling additional sites where

we have identified geographic algal bloom hotspots, but which do not have historic reports of

cholera emergence – to serve as control sites for analyses, and to provide background levels of

Vibrio and other pathogens. This information will inform the development of predictive maps

and population attributable fractions to help translate surveillance and modeling data into

numbers that can inform policy development, identification of communities at increased risk of

cholera, and preventive interventions. Other future projects will include: continued development

of a training program in infectious disease surveillance, development of a web-based GIS

platform to integrate data sources and examine the role of environmental factors in *V. cholerae*

transmission over time and space, and development of an early warning system for climate-

sensitive diseases.

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Table 1. PCR primers set used in this study

Set	Primer	Sequence	Product	Reference
1	O1F2-1	GTT TCA CTG AAC AGA TGG G	192 bp	(Hoshino et al. 1998)
	O1R2-2	CGG TCA TCT GTA AGT ACA AC		Development and evaluation of a multiplex PCR assay for rapid
2	O139F2	AGC CTC TTT ATT ACG GGT GG	449bp	detection of toxigenic *Vibrio cholerae* O1 and O139
	O139R2	GTC AAA CCC GAT CGT AAA GG		
3	tcpA-F	ATG CAA TTA TTA AAA CAG CTT TTT AAG	675bp	(Kumar et al. 2010) Rapid Detection of Virulence-Associated Genes in Environmental
	tcpA-R	TTA GCT GTT ACC AAA TGC AAC AG		Strains of *Vibrio cholerae* by Multiplex PCR
4	ctxA-F	CGG GCA GAT TCT AGA CCT CCT G	564bp	(Singh et al. 2002) Development of a hexaplex PCR
	ctxA-R	CGA TGA TCT TGG AGC ATT CCC AC		assay for rapid detection of virulence and regulatory genes in *Vibrio cholerae* and *Vibrio mimicus*

455 **Table 2.** Kruskal-Wallis rank sum test results for each site and environmental variable

456 differences by month.

Environmental Variable	Site	X^2	DF	p-value
Temperature	Boca de Macho	832.65	9	0.0001
	Héroes de Jambelí	643.85	8	0.0001
	Isla de Jambelí	622.85	9	0.0001
	Puerto Bolívar Adentro	445.99	9	0.0001
	Puerto Bolívar Boca	625.44	9	0.0001
Salinity				
	Boca de Macho	837.16	9	0.0001
	Héroes de Jambelí	230.17	8	0.0001
	Isla de Jambelí	671.41	9	0.0001
	Puerto Bolívar Adentro	464.85	9	0.0001
	Puerto Bolívar Boca	619.21	9	0.0001
pH				
	Boca de Macho	534.3	9	0.0001
	Héroes de Jambelí	431.66	8	0.0001
	Isla de Jambelí	245.91	9	0.0001
	Puerto Bolívar Adentro	378.53	9	0.0001
	Puerto Bolívar Boca	416.76	9	0.0001
BGA				
	Boca de Macho	650.84	9	0.0001
	Héroes de Jambelí	309.2	8	0.0001
	Isla de Jambelí	469.75	9	0.0001
	Puerto Bolívar Adentro	219.78	9	0.0001
	Puerto Bolívar Boca	519.81	9	0.0001

459 **Table 3:** Kendall tau tests for correlation between prevalence of cholera, and each strain

460 separately, and environmental variables at sites, pooled monthly.

Environmental Variable	Prevalence	Kendall's τ	z	*p*-value
Temperature	*V. cholerae*	0.08	0.56	0.57
	O1	0.12	0.80	0.42
	O139	0.02	0.12	0.91
Salinity	*V. cholerae*	-0.34	-2.35	**0.02**
	O1	-0.35	-2.29	**0.02**
	O139	-0.28	-1.92	0.06
pH	*V. cholerae*	-0.10	-0.68	0.49
	O1	-0.14	-0.92	0.36
	O139	-0.10	-0.64	0.52
BGA	*V. cholerae*	0.55	3.76	**0.00**
	O1	0.48	3.09	**0.00**
	O139	0.49	3.34	**0.00**

Figures

Figure 1: Location of sampling sites. **A.** Ecuador (in yellow) in South America, indicating the location of Machala (red point); **B.** Location of Machala on the southern coast of Ecuador (red point); **C.** Location of the five sampling sites: Isla Jambelí, Boca del Macho, Puerto Bolívar Boca, Puerto Bolívar Adentro, and Héroes de Jambeli (red points), in and around Machala (green).

Figure 2: Environmental features in the study area. Water characteristics by site (means and standard errors Temperature (TEMP, °C), Salinity (SAL), pH, and measured total concentration of blue-green algae (BGA, cells/µL).

Figure 3: Environmental features in the study period. Water characteristics by month (means and standard errors) and sites: **A.** Temperature (TEMP, °C), **B.** Salinity (SAL), **C.** measured total concentration of blue-green algae (BGA, cells/µL), and **D.** pH.

Figure 4: *Vibrio cholerae* detection. Monthly site prevalence of **A.** cholera as given by positive PCR test, **B.** O1 strain, **C.** O139 strain.

 Figure 1

486 **Figure 2**

490   **Figure 3**

493   **Figure 4**

494