# Evolution of the sex ratio and effective number under gynodioecy and androdioecy

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Abstract

We analyze dynamic models of the evolution of androdioecy and gynodioecy under autosomal modifiers of weak effect. In our zygote control models, the sex expressed by a zygote depends on its own genotype, while in our maternal control models, the sex expressed by a zygote depends on the genotype of its maternal parent. Our study addresses the Li-Price equation, which for all its heuristic appeal, describes evolutionary change over a single generation. Our analysis unifies full multi-dimensional local stability analysis with the Li-Price equation by identifying a point in the neighborhood of a fixation state from which a single-generation step indicates the asymptotic behavior of a rare, introduced allele initiated at an arbitrary location near the fixation state. We incorporate our theoretical analysis into our previously-developed Bayesian inference framework to develop a new method for inferring the viability of gonochores (males or females) relative to hermaphrodites. Applying this approach to microsatellite data derived from natural populations of the gynodioecious plant *Schiedea salicaria* and the the androdioecious killifish *Kryptolebias marmoratus*, we find that while female and hermaphrodite *S. salicaria* appear to have similar viabilities, male *K. marmoratus* appear to survive to reproductive age at less than half the rate of hermaphrodites.

Keywords:
sex ratio evolution, gynodioecy, androdioecy, effective population size, Li-Price equation

# 1 Introduction

Changes in the breeding system and effective number induce genome-wide transformations of the context in which evolution operates. Here, we address the evolution of androdioecy and gynodioecy under nuclear control and explore the implications of that process for effective number.

Among the central questions regarding the evolution of breeding systems is the nature of Darwinian fitness. Reproductive success of an individual may depend not only on its own sex expression but on the sex of other members of the present or descendant population. Numerous authors have explored definitions of Darwinian fitness under androdioecy and gynodioecy (Ross and Weir 1975; Lloyd 1975; Charlesworth and Charlesworth 1978). An alternative approach, and the one we have adopted here, entails modeling the genetic dynamics without appeal to an external definition of fitness (Ross and Weir 1975, 1976; Wolf and Takebayashi 2004).

The evolution of gynodioecy under the joint control of cytoplasmic and nuclear factors has been well-studied (reviewed by Bailey and Delph 2007; McCauley and Bailey 2009). Clearly, the mechanism of sex expression in systems in which heritability of sex appears to be purely nuclear may comprise a cytoplasmic component as well. For example, upon the fixation in a population of a cytoplasm that induces cytoplasmic male sterility (CMS), variation among individuals in sex expression would no longer depend on cytoplasmic variation ("cryptic CMS," Schultz 1994; Fishman and Willis 2006). Similarly, the genetic basis of sex expression may shift from a single major locus to many loci of minor effect upon fixation at the major locus.

We present models for the evolutionary modification of sex expression by autosomal modifiers of weak effect and arbitrary dominance. Our models address determination of sex of a zygote by either its own genotype (zygote control) or the genotype of its maternal parent (maternal control). Previous workers have studied the evolution of sex expression under zygote control, with simplifying restrictions on dominance (Ross and Weir 1975, 1976; Charlesworth and Charlesworth 1978; Wolf and Takebayashi 2004). The evolution of androdioecy and gynodioecy under maternal control has not to our knowledge been explored previously.

In addition, we address the heuristically-appealing Li-Price framework (Li 1967; Price 1970) in the context of full multi-dimensional local stability analyses. We use our results to explore the definition of heritability of sex expression and the evolution effective number.

Incorporation of our theoretical analysis into a previously-developed Bayesian inference framework (Redelings *et al.* 2015) yields a new method for using multilocus data as a basis for inferring the viability of gonochores (males or females) relative to hermaphrodites.

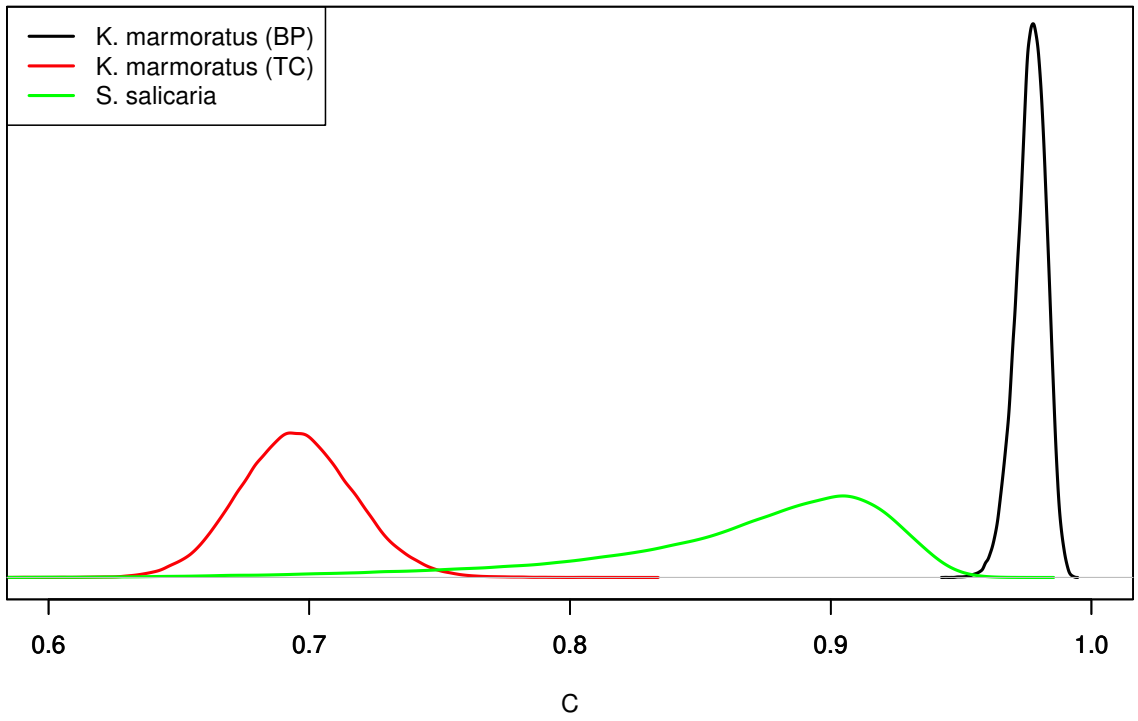


Figure 1: Posterior distributions of the collective contribution of hermaphrodites to the population gene pool ($C$) for two populations of androdioecious *Kryptolebias marmoratus* and for gynodioecious *Schiedea salicaria*.

## 1.1 Collective contribution of a sex

Of fundamental importance to breeding system evolution is the collective contribution of a sex to the population gene pool. It is a key determinant of both reproductive value (Fisher 1930) and effective number (Wright 1931).

**Empirical observations:** Figure 1 presents posterior distributions of $C$, the collective contribution of hermaphrodites, inferred from the three microsatellite data sets analyzed by Redelings *et al.* (2015). In the androdioecious killifish *Kryptolebias marmoratus*, hermaphrodites collectively contribute a substantially greater proportion of the population gene pool in the more highly inbred BP population than in the TC population. In *Schiedea salicaria*, the collective contribution of females (male-steriles) lies close to the population proportion of females of 12% reported by Campbell *et al.* (2010).

**Effective number:** Wright (1931) introduced the notion of effective population size in the context of generalizing fundamental aspects of evolutionary change to populations structured by sex, fluctuations through time in numbers of individuals, or other factors. In their analysis of the concept of effective population size, Ewens (1982) and Crow and Denniston (1988) noted departures among the various definitions in specific models.

We refer to the probability that a pair of genes randomly sampled from distinct reproduc-

tives in the present (offspring) generation derive from the same reproductive in the preceding (parental) generation as the rate of parent-sharing ($1/N_P$):

$$\frac{1}{N_P} = \frac{C^2}{N_H} + \frac{(1-C)^2}{N_G}. \tag{1}$$

for $N_H$ and $N_G$ representing the effective number of hermaphrodites and gonochores (males or females) in the parental generation, respectively, and $C$ the probability that an autosomal gene randomly sampled from a reproductive in the offspring generation derives from a hermaphroditic parent. Here, $C^2$ corresponds to the probability that two genes, each randomly sampled from a reproductive, both derive from hermaphrodites, with $1/N_H$ the probability that the same hermaphrodite contributed both genes. Crow and Denniston (1988) denoted the inverse of the rate of parent-sharing ($N_P$) as "inbreeding effective size."

We define relative effective number ($R$) as the ratio of inbreeding effective size and the total effective number of reproductives ($N = N_G + N_H$):

$$R = \frac{N_P}{N}. \tag{2}$$

From (1), we obtain

$$\begin{aligned} R &= \frac{N_P}{N_H + N_G} = \frac{1}{\frac{C^2}{h} + \frac{(1-C)^2}{1-h}} \\ &= \frac{h(1-h)}{h(1-h) + (h-C)^2}, \end{aligned} \tag{3}$$

for $h$ the proportion of hermaphrodites among reproductives:

$$h = \frac{N_H}{N_G + N_H}. \tag{4}$$

Clearly, relative effective number cannot exceed unity ($R \leq 1$), attaining unity only for

$$h = C, \tag{5}$$

at which the proportion of hermaphrodites among reproductives ($h$) is identical to $C$, the probability that a random gene sampled from reproductives derives from a hermaphrodite in the parental generation. Both (1) and (2) differ conceptually and quantitatively from indices proposed by Laporte *et al.* (2000), who explored effective number in gynodioecious populations. That distinct concepts of effective number exist is not unexpected under even the most basic forms of population structure, including sex (Ewens 1982; Crow and Denniston 1988).

Figure 2 presents posterior distributions of relative effective number $R$ (2) for the three data sets studied by Redelings *et al.* (2015), including those derived from two populations of the androdioecious killifish *Kryptolebias marmoratus* (Mackiewicz *et al.* 2006; Tatarenkov *et al.* 2012). While the gynodioecious *S. salicaria* population (green) shows relative effective number $R$ near its maximum of unity, the killifish populations appear to have lower $R$, especially the Belize population TC, in which higher male proportions (ca. 17%, Mackiewicz *et al.* 2006) have been observed than in the Florida Keys population BP (ca. 1%, Turner *et al.* 1992; Tatarenkov *et al.* 2012).

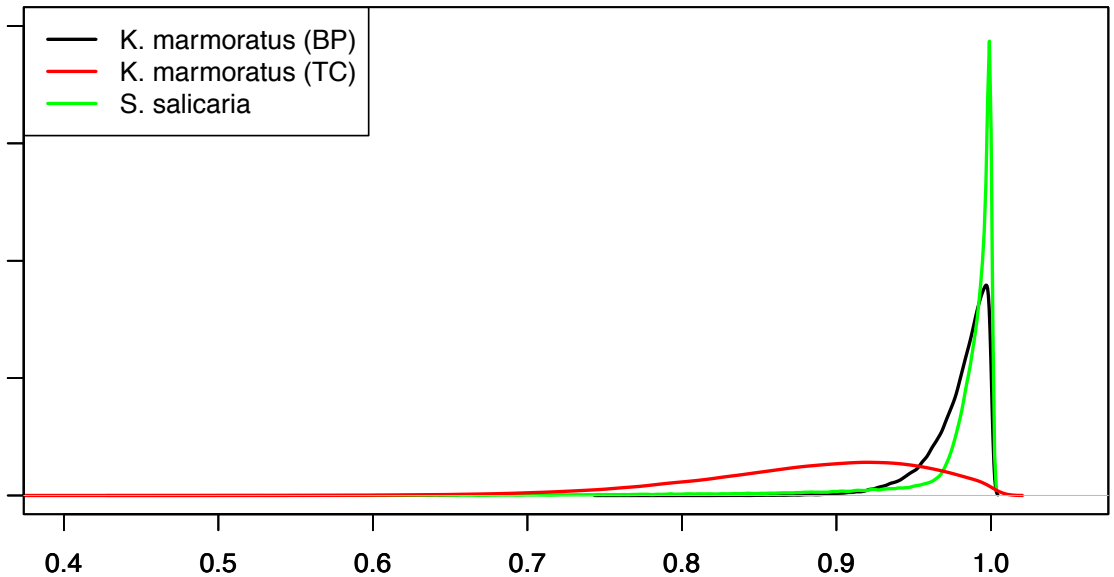


Figure 2: Posterior distributions of relative effective number $R$ (2).

## 1.2 Reproductive value

Fisher (1930) explored the evolutionary modification of the sex ratio under gonochorism, with $N_f$ females and $N_m$ males participating in reproduction. Under the assumption that reproduction is limited by the number of females, the total number of zygotes is proportional to $N_f$ and the *reproductive value* of a male relative to a female corresponds to $N_f/N_m$. Because males and females make equal collective contributions at each autosomal locus, autosomal modifiers evolve toward equal investment in male and female offspring (Fisher 1930). Edwards (2000) provides an account of the origins of this argument.

The evolution of the sex ratio has also been addressed in the context of the marginal value of parental investment in offspring of each sex (*e.g.*, Shaw and Mohler 1953; Lloyd 1975; Charnov *et al.* 1976). Increased investment in the sex with the highest marginal value affords increased transmission to the grandoffspring generation. Equal marginal value among mating types implies that no transmission advantage accrues to any change in investment strategy. For sexual forms corresponding to hermaphrodites and gonochores, the per capita contribution of hermaphroditic offspring to the grandoffspring generation corresponds to $C/N_H$, reflecting the partitioning among $N_H$ reproductive hermaphrodites of the collective contribution to the gene pool by hermaphrodites ($C$). The marginal value of investing in hermaphroditic offspring exceeds the marginal value of investing in gonochorous offspring only if

$$\frac{C}{N_H} > \frac{Z(1-C)}{N_G}, \tag{6}$$

for $Z$ the expected number of gonochores of reproductive age that can be produced with the investment required to produce a single hermaphrodite of reproductive age. In this context, the reproductive value of a sex is proportional to a ratio of marginal values.

An evolutionarily stable strategy (ESS, Maynard Smith and Price 1973) corresponds to an investment allocation against which no other allocation can increase when rare. Candidate ESS hermaphrodite proportions ($h^*$) correspond to points of equality between the marginal values of hermaphrodites and gonochores:

$$\frac{h^*}{1-h^*} = \frac{C}{Z(1-C)}. \tag{7a}$$

If the departure of $Z$ from unity derives entirely from differential viability of gonochores relative to hermaphrodites, this candidate ESS corresponds to a sex ratio immediately before the sex-specific viability phase (rather than at reproduction) of

$$\frac{\hat{h}}{1-\hat{h}} = \frac{C}{1-C}. \tag{7b}$$

A candidate hermaphrodite proportion would in fact correspond to an ESS only if any rare modifier of the sex ratio fails to increase at a geometric rate in a monomorphic population exhibiting the candidate sex ratio. Further, $\hat{h}$ would correspond to an ESS that is locally attracting in parameter space if rare autosomal enhancers of hermaphrodite production invade a population with hermaphrodite proportion $h_c$ only if $h_c < \hat{h}$ and suppressors invade only if $h_c > \hat{h}$. Such an investment allocation has been described as a continuously stable strategy (CSS, Eshel and Motro 1981) or as showing $m$-stability (Taylor 1989) or convergence stability (Christiansen 1991).

Attainment of the presumptive ESS sex ratio (7) implies maximization of relative effective number (2) only if gonochores and hermaphrodites have equal viability ($Z = 1$). Otherwise ($Z \neq 1$), the departure from unity of relative effective number at the candidate sex ratio ESS provides a basis for inference of $Z$. Under the assumption that the natural populations under study (Fig. 1) have in fact evolved to the ESS, we use the Bayesian sampler of Redelings *et al.* (2015) to obtain posterior densities for the relative viability of gonochores ($Z$) in *Kryptolebias marmoratus* and *Schiedea salicaria*. While we find little support for differential viability of the alternative sexual forms in *S. salicaria*, *K. marmoratus* males appear to have significantly lower viability than hermaphrodites.

# 2 Methods

## 2.1 Candidate ESS sex expression levels

We derive candidate ESS values under zygote and maternal control of sex expression in populations comprising $N_H$ hermaphrodites and $N_G$ gonochores (males or females). These candidate ESS levels extend those proposed by Lloyd (1975). Our full local stability analysis (Section 3) demonstrates that these candidates do in fact correspond to continuously stable strategies.

**Life cycle:** Figure 3 depicts the major phases of the life cycle. A proportion $\tilde{s}$ of egg cells produced by hermaphrodites are self-fertilized (uniparental). In the gynodioecy models, females produce offspring (all biparental) at rate $\tilde{\sigma}$ relative to hermaphrodites ($\tilde{\sigma}$ corresponds to $\sigma$ in Redelings *et al.* 2015). Inbreeding depression occurs immediately after zygote formation, with uniparental offspring surviving to the juvenile stage at rate $\tau$ relative to biparental offspring. Under a rescaling at the juvenile stage, a female has an average of $\sigma$ surviving offspring relative to a hermaphrodite, for which uniparentals constitute a proportion $s$ of its surviving offspring. Gonochorous offspring survive to reproductive age at rate $Z$ relative to hermaphroditic offspring, irrespective of whether they are uniparental or biparental.

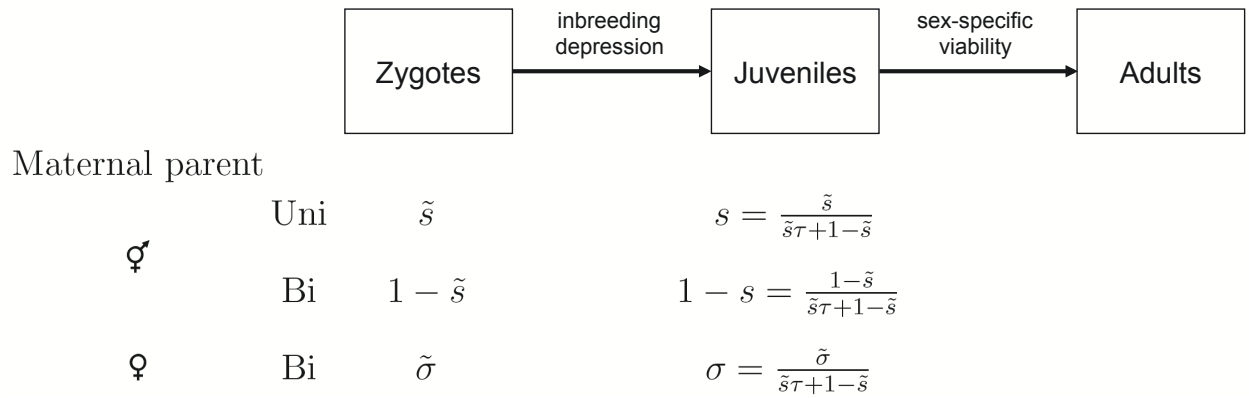


Figure 3: Life cycle. Hermaphrodites (⚥) self-fertilize a proportion $\tilde{s}$ of their egg cells, with the complement randomly outcrossed. Under gynodioecy, females (♀) generate $\tilde{\sigma}$ egg cells relative to hermaphrodites ($\tilde{\sigma}$ corresponds to $\sigma$ in Redelings *et al.* 2015). Inbreeding depression occurs immediately after zygote formation, with uniparental zygotes ("Uni") surviving at rate $\tau$ relative to biparental zygotes ("Bi"). At the juvenile stage, a proportion $s$ of the remaining offspring of hermaphrodites are uniparental, with $\sigma$ the relative number of juvenile offspring produced per female parent. Sex-specific viability selection occurs between the juvenile and adult phases, with gonochores (males or females) surviving to reproductive age at rate $Z$ relative to hermaphrodites. In the absence of heritable variation in sex expression (*e.g.*, fixation of a modifier allele that induces the ESS), the sex of an offspring is independent of parental sex and of whether it is uniparental or biparental; in such cases, $s$ and $\sigma$ are identical at the juvenile and adult stages.

Our full dynamical models depict evolving autosomal modifiers of sex expression. In contrast, our ESS derivation assumes the absence of heritable variation in sex expression: for example, upon the fixation of a modifier allele that induces the ESS sex ratio. Under this assumption, offspring sex (gonochore or hermaphrodite) is independent of parental sex and independent of the level of inbreeding. Accordingly, the relative proportions of uniparental and biparental offspring ($s$ and $\sigma$ in Fig. 3) are identical at the juvenile and adult stages and the sex ratio among zygotes is identical to the sex ratio among juveniles.

**Androdioecy:** Under androdioecy ($N_G$ males and $N_H$ hermaphrodites), outcrossing entails fertilization of egg cells from the pollen cloud, to which a female-sterile (male) individual

contributes at rate $\omega$ relative to hermaphrodites. In accordance with the laboratory experiments of Furness *et al.* (2015) on *Kryptolebias marmoratus*, our Kryptolebias model imposes the additional assumption that all biparental individuals have a male parent ($\omega = \infty$).

Hermaphrodites alone produce egg cells, of which are $\tilde{s}$ are fertilized by self-pollen. The uniparental proportion among juveniles,

$$s_A = \frac{\tilde{s}\tau}{\tilde{s}\tau + 1 - \tilde{s}}, \tag{8}$$

is independent of the population sex ratio.

The probability that an autosomal gene randomly sampled from juvenile offspring (Fig. 3) derives from a hermaphrodite in the parental generation corresponds to

$$C_A = 1 - (1 - s_A)G_A/2,$$

in which $G_A$ reflects the relative contribution of males of the parental generation to the pollen pool:

$$G_A = \frac{\omega N_G}{\omega N_G + N_H} = \frac{\omega(1-h)}{\omega(1-h)+h}, \tag{9}$$

for $(1-h)$ the frequency of males among reproductives in the parental generation (4). In the Kryptolebias model, in which all biparental offspring have a male parent ($G_A = 1, \omega = \infty$), the collective contribution of hermaphrodites reduces to

$$C_A = 1 - (1 - s_A)/2 = (1 + s_A)/2.$$

As indicated in our exposition of the life cycle (Fig. 3), the absence of heritable genetic variation for sex expression (*e.g.*, at a genetically monomorphic ESS state) implies that the uniparental proportion $s_A$ is identical at the juvenile and adult stages. At such an ESS state, $C_A$ corresponds to the probability that a random autosomal gene sampled at either the juvenile or the adult stage derives from a hermaphrodite in the preceding generation.

Candidate ESS sex ratios at reproductive age (7) satisfy

$$\frac{h^*}{1-h^*} = \frac{1 - (1 - s_A)/2\left[\frac{\omega(1-h^*)}{\omega(1-h^*)+h^*}\right]}{Z(1 - s_A)/2\left[\frac{\omega(1-h^*)}{\omega(1-h^*)+h^*}\right]}.$$

Solving, we obtain candidates for the unbeatable sex ratio at reproduction under androdioecy:

$$\frac{h_A^*}{1-h_A^*} = \begin{cases} \frac{\omega(1+s_A)/2}{Z\omega(1-s_A)/2-1} & \text{if valid} \\ \infty & \text{otherwise.} \end{cases} \tag{10a}$$

Maintenance of androdioecy ($1 > h_A^* > 0$) requires that the expected contribution of a juvenile male to the subsequent generation exceed that of a juvenile hermaphrodite by at least twofold:

$$Z\omega(1 - s_A) > 2. \tag{10b}$$

This condition becomes more stringent as the rate of outcrossing $(1 - s_A)$ or the relative viability of males $(Z)$ decline. If (10b) fails, the sole candidate ESS corresponds to pure hermaphroditism $(h_A = 1)$.

At the juvenile (rather than adult) stage, the candidate ESS (10a) corresponds to a sex ratio of

$$\frac{\hat{h}_A}{1 - \hat{h}_A} = \begin{cases} \frac{Z\omega(1+s_A)/2}{Z\omega(1-s_A)/2-1} & \text{if valid} \\ \infty & \text{otherwise.} \end{cases} \tag{10c}$$

indicating that the the composite parameter $Z\omega$ represents the net effects on the ESS of differential viability and pollen success of males.

In his treatment of androdioecy, Lloyd (1975, his equation (7)) proposed an unbeatable proportion of males of

$$q = \frac{t - 2lv[t + i(1-t)]}{2iv(1-t)(1-l) + t(1 + v - 2lv)}, \tag{11}$$

for $q$ the proportion of males at reproductive age, $t$ the proportion of seeds set by non-self pollen, $l$ the pollen production of a hermaphrodite (described as "female") relative to a male, $v$ the rate of survival to reproduction of a hermaphrodite relative to a male, and $i$ the viability of uniparental offspring relative to biparental offspring. Substitution of

$$\begin{aligned} t &= 1 - \tilde{s} \\ 1/l &= \omega \\ 1/v &= Z \\ i &= \tau. \end{aligned}$$

into (11) corresponds to our non-zero ESS candidate (10).

**Gynodioecy:** Under gynodioecy ($N_G$ females and $N_H$ hermaphrodites), females set seeds at rate $\tilde{\sigma}$ relative to hermaphrodites (Fig. 3). An autosomal gene randomly sampled from a juvenile offspring derives from a hermaphrodite parent with probability

$$\begin{aligned} C_G &= \frac{N_G\tilde{\sigma}/2 + N_H(\tau\tilde{s} + 1 - \tilde{s})}{N_G\tilde{\sigma} + N_H(\tau\tilde{s} + 1 - \tilde{s})} \\ &= \frac{(1-h)\sigma/2 + h}{(1-h)\sigma + h} \end{aligned} \tag{12}$$

for $h$ the proportion of hermaphrodites among parents in the preceding generation (4) and $\sigma$ the scaled seed fertility of females (Fig. 3). This expression also corresponds to

$$C_G = 1 - (1 - s_G)G_G/2,$$

for the uniparental proportion among juveniles given by

$$s_G = \frac{N_H\tau\tilde{s}}{N_G\tilde{\sigma} + N_H(\tau\tilde{s} + 1 - \tilde{s})} = \frac{hs}{(1-h)\sigma + h}, \tag{13}$$

and the proportion of biparental offspring that have a female parent by

$$G_G = \frac{N_G\tilde{\sigma}}{N_G\tilde{\sigma} + N_H(1-\tilde{s})} = \frac{(1-h)\sigma}{(1-h)\sigma + h(1-s)}. \tag{14}$$

In contrast with androdioecy (8), the uniparental fraction $s_G$ (13) depends on the population sex ratio. Once again, at a monomorphic ESS (absence of heritable genetic variation for sex expression), the uniparental proportion among offspring of reproductive age is identical to $s_G$ among juvenile offspring.

From (7) and (12), the candidate ESS at reproductive age corresponds to

$$h = \frac{(1-h)\sigma/2 + h}{(Z+1)(1-h)\sigma/2 + h}.$$

Solving, we obtain candidates for the unbeatable sex ratio under gynodioecy:

$$\frac{h_G^*}{1-h_G^*} = \begin{cases} \frac{\sigma}{Z\sigma-2} & \text{if valid} \\ \infty & \text{otherwise.} \end{cases} \tag{15}$$

Maintenance of gynodioecy ($1 > h_G^* > 0$) requires that the expected number of offspring produced by a juvenile female exceed that of a juvenile hermaphrodite by at least twofold:

$$Z\sigma > 2. \tag{16}$$

More intense inbreeding depression (smaller $\tau$) and higher female viability or fertility (larger $Z$ or $\tilde{\sigma}$) tend to promote gynodioecy. For cases in which (16) fails, (15) indicates that the sole candidate ESS corresponds to pure hermaphroditism ($h_G^* = 1$).

At the juvenile stage (Fig. 3), the candidate ESS (15) corresponds to

$$\frac{\hat{h}_G}{1-\hat{h}_G} = \begin{cases} \frac{Z\sigma}{Z\sigma-2} & \text{if valid} \\ \infty & \text{otherwise,} \end{cases} \tag{17}$$

with composite parameter $Z\sigma$ comprising the net effects on the ESS of differential viability and seed set of females.

Equation (2) of Lloyd (1975) provides the unbeatable sex ratio under gynodioecy:

$$\frac{p}{1-p} = \frac{b-2SX}{bS}, \tag{18}$$

for $p$ the proportion of females at reproductive age, $b$ the seed set of females, $S$ the viability of a hermaphrodite (described as "male") relative to a female, and $X$ the number of zygotes surviving to reproduction produced by a hermaphrodite relative to a female. In Lloyd's notation,

$$X = i[a + be(1-a)(1-w) + (1-be)(1-a)r] + be(1-a)w,$$

in which $i$ corresponds to the relative viability of uniparental offspring (our $\tau$), the first bracket the proportion of seeds of hermaphrodites set by self-pollen (our $\tilde{s}$), and $be(1-a)w$

the proportion of seeds of hermaphrodites set by pollen from the pollen cloud (our $(1-\tilde{s})$). Substitution of

$$\begin{aligned} i &= \tau \\ X &= (\tau\tilde{s} + 1 - \tilde{s}) \\ b &= \tilde{\sigma} \\ 1/S &= Z. \end{aligned}$$

into (18) corresponds to our non-zero ESS candidate (15).

## 2.2 Li-Price equation

Li (1967) and Price (1970) expressed the one-generation change in the frequency of an allele as a covariance between fitness and the frequency of the allele across genotypes. Here, we extend this framework to the evolution of effective number and sex ratio in inbred populations.

Table 1 presents measures associated with genotypes at a biallelic autosomal locus. In the population, genotypes $AA$, $Aa$, and $aa$ occur in frequencies to $u_0$, $u_1$, and $u_2$ ($\sum_i u_i = 1$). The locus may influence the expression of a trait, with genotype $i$ associated with trait

**Table 1**
Phenotypic and genetic values

| | Genotypes | | |
|---|---|---|---|
| | $AA$ | $Aa$ | $aa$ |
| Frequency | $u_0$ | $u_1$ | $u_2$ |
| Trait deviation | $P_0 - \bar{P}$ | $P_1 - \bar{P}$ | $P_2 - \bar{P}$ |
| Additive genotypic value | $2\alpha_0$ | $\alpha_0 + \alpha_1$ | $2\alpha_1$ |
| Fitness deviation | $T(u_0' - u_0)/u_0$ | $T(u_1' - u_1)/u_1$ | $T(u_2' - u_2)/u_2$ |

deviation $(P_i - \bar{P})$, in which the average value of the trait corresponds to

$$\bar{P} = \sum_i u_i P_i.$$

Price (1970) defined the fitness of genotype $i$ as proportional to the number of gametes transmitted to the offspring generation. In panmictic populations, in which genotypic frequencies at the point of zygote formation conform to Hardy-Weinberg proportions, this definition of fitness corresponds to the expected rate of survival to reproduction, as assumed by Li (1967). Because fitness in the present context may include various components, we here define the fitness of genotype $i$ as the ratio of numbers of individuals of genotype $i$ at

the same point in the life cycle in consecutive generations:

$$W_i = \frac{Tu_i'}{u_i}, \tag{19}$$

for the prime representing the next generation forward in time. Denniston (1978) observed that this measure departs from more conventional notions of fitness: high genotypic fitness reflects high production *of* the genotype rather than *by* the genotype. Under this definition, fitness is virtually always frequency-dependent: even for the most basic model of constant viability selection, (19) ceases to change only at equilibria ($u_i' = u_i$).

To genotypes $AA$, $Aa$, and $aa$, we associate additive genotypic values $2\alpha_0$, $\alpha_0 + \alpha_1$, and $2\alpha_1$. Much previous work, designed for panmictic populations, has defined additive genotypic value as the frequency of allele $A$ in a genotype (Li 1967; Price 1970). Here, we use the definition of Fisher (1941), under which the additive effects $\alpha_i$ are obtained by minimizing the mean squared deviation (MSD) of the phenotype from the additive genotypic value across genotypes:

$$\text{MSD} = u_0[P_0 - \bar{P} - 2\alpha_0]^2 + u_1[P_1 - \bar{P} - (\alpha_0 + \alpha_1)]^2 + u_2[P_2 - \bar{P} - 2\alpha_1]^2. \tag{20}$$

For general systems of mating, the *average effect* of substitution (Fisher 1941), the expected effect on the trait of substituting allele $A$ for allele $a$, corresponds to

$$\begin{aligned} \alpha_0 - \alpha_1 &= \frac{2u_0(P_0 - \bar{P}) + u_1(P_1 - \bar{P})}{4p(1-p) - u_1} \\ &= \frac{F(P_0 - P_2) + (1-F)[p(P_0 - P_1) + (1-p)(P_1 - P_2)]}{1+F}, \end{aligned} \tag{21}$$

for $p$ representing the frequency of allele $A$ ($p = u_0 + u_1/2$) and $F$ the fixation index (Wright 1933). In the additive case, in which

$$(P_0 - P_2) = 2(P_1 - P_2),$$

the average effect reduces to

$$\alpha_0 - \alpha_1 = (P_1 - P_2),$$

irrespective of the magnitude of $F$ or intensity of any selection.

Using the definitions summarized in Table 1, we obtain the covariance across genotypes between fitness $W$ (19) and additive genotypic value $G_\alpha$ with respect to the trait:

$$\begin{aligned} \text{Cov}(WG_\alpha) &= u_0\frac{T(u_0' - u_0)}{u_0}2\alpha_0 + u_1\frac{T(u_1' - u_1)}{u_1}(\alpha_0 + \alpha_1) + u_2\frac{T(u_2' - u_2)}{u_2}2\alpha_2 \\ &= 2(\alpha_0 - \alpha_1)T\Delta p, \end{aligned} \tag{22}$$

in which $\Delta p$ represents the change in frequency of allele $A$ over a single generation. This expression indicates that the frequency of the $A$ allele increases ($\Delta p > 0$) if either (1) its average effect of substitution on the trait is positive ($(\alpha_0 - \alpha_1) > 0$) and the trait is positively

correlated with fitness ($\mathrm{Cov}(WG_\alpha) > 0$) or (2) its average effect of substitution on the trait is negative ($(\alpha_0 - \alpha_1) < 0$) and the trait is negatively correlated with fitness ($\mathrm{Cov}(WG_\alpha) < 0$). To address Fisher's (1958) fundamental theorem of natural selection, Li (1967) and Price (1970, 1971) considered the trait of fitness itself, in which case the covariance $\mathrm{Cov}(WG_\alpha)$ reduces to the additive variance in fitness.

For all its heuristic appeal, expression (22) provides a one-dimensional description of evolutionary change across a single generation. In the present context, the trait of interest corresponds to the long-term evolution of sex expression in a multi-dimensional state space. Unless sex expression is uncorrelated with fitness ($\mathrm{Cov}(WG_\alpha) = 0$) or the focal modifier locus has no additive variance with respect to this trait ($(\alpha_0 - \alpha_1) = 0$), natural selection will induce genetic change. Because both the average effect of substitution (21) and the covariance $\mathrm{Cov}(WG_\alpha)$ depend on the genotypic frequencies, the relationship between the one-generation description provided by (22) and the outcome of the evolution process needs clarification.

Key to the application of the Li-Price framework to the evolution of sex expression is the elucidation of the component of the population to which the genotypic frequencies ($u_i$) in Table 1 correspond. In the present context, populations may comprise both gonochores and hermaphrodites, and sex expression in a zygote depends on either its own genotype or the genotype of its maternal parent. As the average effect of substitution (21) is determined with respect to this genotypic distribution ($u_i$), it defines heritability for the evolutionary process under study.

## 2.3 Dynamic models of sex ratio evolution

We address two genetic mechanisms for the determination of sex expression. In the zygote control models, zygotes of genotypes $AA$, $Aa$, and $aa$ respectively develop into hermaphrodites at rates $h_0$, $h_1$, and $h_2$ ($0 \leq h_i \leq 1, i = 0, 1, 2$). In the maternal control models, it is the genotype of the maternal parent of the zygotes that determines sex expression rates.

Hermaphrodites set a proportion $\tilde{s}$ of seeds by self-fertilization. Uniparental offspring survive to reproduction at rate $\tau$ relative to biparental offspring, with this differential survival occurring immediately upon zygote formation, even before sex expression.

### 2.3.1 Zygotic control of sex expression

Following differential survival of uniparental and biparental offspring (but before sex expression and reproduction by the offspring), genotypes $AA$, $Aa$, and $aa$ occur in proportions $z_0$, $z_1$, and $z_2$ ($z_0 + z_1 + z_2 = 1$).

**Androdioecy:** In the next generation forward in time, genotypic frequencies correspond

to

$$
\begin{aligned}
z_0' &\propto \tilde{s}\tau(z_0h_0 + z_1h_1/4) + (1-\tilde{s})(z_0h_0 + z_1h_1/2)q \\
z_1' &\propto \tilde{s}\tau z_1h_1/2 + (1-\tilde{s})[(z_0h_0 + z_1h_1/2)(1-q) + (z_1h_1/2 + z_2h_2)q] \\
z_2' &\propto \tilde{s}\tau(z_1h_1/4 + z_2h_2) + (1-\tilde{s})(z_1h_1/2 + z_2h_2)(1-q),
\end{aligned}
$$

for $q$ denoting the frequency of the $A$ allele in the pollen pool:

$$
q = \frac{h_0z_0 + h_1z_1/2 + \omega Z[(1-h_0)z_0 + (1-h_1)z_1/2]}{h_0z_0 + h_1z_1 + h_2z_2 + \omega Z[(1-h_0)z_0 + (1-h_1)z_1 + (1-h_2)z_2]}. \tag{23a}
$$

These expressions imply

$$
\begin{aligned}
Tz_0' =& s_A(z_0h_0 + z_1h_1/4) + (1-s_A)(z_0h_0 + z_1h_1/2)q \\
Tz_1' =& s_Az_1h_1/2 + (1-s_A)[(z_0h_0 + z_1h_1/2)(1-q) + (z_1h_1/2 + z_2h_2)q] \\
Tz_2' =& s_A(z_1h_1/4 + z_2h_2) + (1-s_A)(z_1h_1/2 + z_2h_2)(1-q),
\end{aligned} \tag{23b}
$$

for $s_A$ given in (8) and the normalizer by

$$
T = h_0z_0 + h_1z_1 + h_2z_2. \tag{23c}
$$

In the absence of selection on the modifier locus ($h_0 = h_1 = h_2$), recursion system (23) indicates that allele frequency in seeds and pollen ($z_0 + z_1/2 = q$) remains at its initial value, with asymptotic convergence at rate $s_A/2$ of the frequency of heterozygotes ($z_1$) to

$$
2q(1-q)(1-F_{neut}),
$$

for $F_{neut}$ the fixation index (Wright 1933):

$$
F_{neut} = \frac{s}{2-s}, \tag{24}
$$

with $s_A$ substituted for $s$.

**Gynodioecy:** Genotypic frequencies in the next generation forward in time correspond to

$$
\begin{aligned}
z_0' &\propto \tilde{s}\tau(z_0h_0 + z_1h_1/4) \\
&\quad + \{(1-\tilde{s})(z_0h_0 + z_1h_1/2) + \tilde{\sigma}Z[z_0(1-h_0) + z_1(1-h_1)/2]\}q \\
z_1' &\propto \tilde{s}\tau z_1h_1/2 \\
&\quad + \{(1-\tilde{s})(z_0h_0 + z_1h_1/2) + \tilde{\sigma}Z[z_0(1-h_0) + z_1(1-h_1)/2]\}(1-q) \\
&\quad + \{(1-\tilde{s})(z_1h_1/2 + z_2h_2) + \tilde{\sigma}Z[z_1(1-h_1)/2 + z_2(1-h_2)]\}q \\
z_2' &\propto \tilde{s}\tau(z_1h_1/4 + z_2h_2) \\
&\quad + \{(1-\tilde{s})(z_1h_1/2 + z_2h_2) + \tilde{\sigma}Z[z_1(1-h_1)/2 + z_2(1-h_2)]\}(1-q),
\end{aligned}
$$

in which $q$ represents the frequency of the $A$ allele in the pollen pool (which derives entirely from hermaphrodites),

$$
q = \frac{h_0z_0 + h_1z_1/2}{h_0z_0 + h_1z_1 + h_2z_2}. \tag{25a}
$$

After division by $(\tilde{s}\tau + 1 - \tilde{s})$, we obtain

$$\begin{aligned}
Tz_0' &= s(z_0h_0 + z_1h_1/4) \\
&\quad + \{(1-s)(z_0h_0 + z_1h_1/2) + \sigma Z[z_0(1-h_0) + z_1(1-h_1)/2]\}q \\
Tz_1' &= sz_1h_1/2 \\
&\quad + \{(1-s)(z_0h_0 + z_1h_1/2) + \sigma Z[z_0(1-h_0) + z_1(1-h_1)/2]\}(1-q) \\
&\quad + \{(1-s)(z_1h_1/2 + z_2h_2) + \sigma Z[z_1(1-h_1)/2 + z_2(1-h_2)]\}q \\
Tz_2' &= s(z_1h_1/4 + z_2h_2) \\
&\quad + \{(1-s)(z_1h_1/2 + z_2h_2) + \sigma Z[z_1(1-h_1)/2 + z_2(1-h_2)]\}(1-q),
\end{aligned} \tag{25b}$$

for the normalizer corresponding to

$$T = \sum_{i=0}^{2} z_i[h_i + \sigma Z(1-h_i)]. \tag{25c}$$

In the absence of selection on the modifier locus ($h_0 = h_1 = h_2 = h$), allele frequency in seeds and pollen ($z_0 + z_1/2 = q$) remains at its initial value. Unlike the uniparental fraction $s_A$ (8) under androdioecy, $s_G$ (13) depends on the population sex ratio. The frequency of heterozygotes ($z_1$) converges asymptotically at rate $s_G/2$ (13) to

$$2q(1-q)(1-F_{neut}),$$

for $F_{neut}$ given in (24) but with $s_G$ (13) substituted for $s$. Selective neutrality at the modifier locus entails that the transformation (25) has an eigenvalue of unity (reflecting no changes in allele frequency) and an eigenvalue of $s_G/2$ (reflecting convergence of $z_1$ under inbreeding).

### 2.3.2 Maternal control of sex expression

Under the maternal control model, the genotype of the maternal parent determines sex expression in zygotes. We describe recursions in genotypic frequencies at the point of reproduction (rather than zygote formation), with genotypes $AA$, $Aa$, and $aa$ occurring in proportions $x_0$, $x_1$, and $x_2$ in hermaphrodites and $y_0$, $y_1$, and $y_2$ in gonochores ($x_0+x_1+x_2+y_0+y_1+y_2 = 1$).

**Androdioecy:** At the point of reproduction, genotypic frequencies among hermaphrodites correspond to

$$\begin{aligned}
Tx_0' &= s_A(x_0h_0 + x_1h_1/4) + (1-s_A)(x_0h_0 + x_1h_1/2)q \\
Tx_1' &= s_Ax_1h_1/2 + (1-s_A)[(x_0h_0 + x_1h_1/2)(1-q) + (x_1h_1/2 + x_2h_2)q] \\
Tx_2' &= s_A(x_1h_1/4 + x_2h_2) + (1-s_A)(x_1h_1/2 + x_2h_2)(1-q),
\end{aligned} \tag{26a}$$

for $q$ the frequency of the $A$ allele in the pollen cloud,

$$q = \frac{x_0 + x_1/2 + \omega(y_0 + y_1/2)}{x_0 + x_1 + x_2 + \omega(y_0 + y_1 + y_2)}, \tag{26b}$$

and $s_A$ given in (8). Substitution of $Z(1-h_i)$ for $h_i$ in the hermaphrodite recursion $Tx'_i$ produces the male recursion $Ty'_i$, which implies the normalizer

$$T = \sum_{i=0}^{2} x_i[h_i + Z(1-h_i)]. \tag{26c}$$

Because male genotypic frequencies ($y_i$) affect transmission only through the pollen cloud (26b), description of the transformation requires a smaller set of variables, including $x_0$, $x_1$, $x_2$, $(y_0 + y_1/2)$, and $(y_1/2 + y_2)$.

In the absence of selection on the modifier locus ($h_0 = h_1 = h_2 = h$), the population ratio of hermaphrodites to males converges in a single generation to

$$\frac{\sum_i x'_i}{\sum_i y'_i} = \frac{h}{h + Z(1-h)} \tag{27a}$$

and the genotypic frequencies in hermaphrodites and males are proportional:

$$\frac{x'_i}{y'_i} = \frac{h}{Z(1-h)}. \tag{27b}$$

Accordingly, the frequencies of allele $A$ among hermaphrodites ($x_0+x_1/2$), males ($y_0+y_1/2$), and pollen ($q$) converge to equality in a single generation,

$$q' = \frac{x'_0 + x'_1/2}{\sum_i x'_i} = \frac{y'_0 + y'_1/2}{\sum_i y'_i},$$

and attain their common equilibrium value in two generations,

$$p = \frac{x_0 + x_1/2}{\sum_i x_i}(1+s_A)/2 + q(1-s_A)/2, \tag{28}$$

in which the uniparental proportion $s_A$ is given in (8) and $x_i$ and $q$ represent the initial values of those variables. The frequency of heterozygotes converges asymptotically, at rate $s_A/2$, to

$$x_1 + y_1 = 2p(1-p)(1-F_{neut}),$$

for $p$ given in (28) and $F_{neut}$ in (24), with $s_A$ (8) substituted for $s$.

Near the state of fixation of the $a$ allele, the neutral transformation has a single eigenvalue of unity (corresponding to allele frequency), a single eigenvalue of $s_A/2$ (governing convergence of the frequency of heterozygotes to the value dictated by $F_{neut}$ and allele frequency), and two eigenvalues of zero (representing the near-instantaneous convergence to equality of allele frequencies in hermaphrodites, males, and pollen).

**Gynodioecy:** Genotypic frequencies in the next generation forward in time correspond to

$$\begin{aligned}
Tx'_0 &= s(h_0x_0 + h_1x_1/4) + q[(1-s)(h_0x_0 + h_1x_1/2) + \sigma(h_0y_0 + h_1y_1/2)] \\
Tx'_1 &= sh_1x_1/2 + (1-s)[(1-q)(h_0x_0 + h_1x_1/2) + q(h_1x_1/2 + h_2x_2)] \\
&\qquad + \sigma[(1-q)(h_0y_0 + h_1y_1/2) + q(h_1y_1/2 + h_2y_2)] \\
Tx'_2 &= s(h_1x_1/4 + h_2x_2) + (1-q)[(1-s)(h_1x_1/2 + h_2x_2) + \sigma(h_1y_1/2 + h_2y_2)]
\end{aligned} \tag{29a}$$

for $q$ the frequency of $A$ in the pollen cloud (to which hermaphrodites alone contribute):

$$q = \frac{x_0 + x_1/2}{x_0 + x_1 + x_2}. \tag{29b}$$

Similar to the androdioecy model, the $Ty_i'$ have the same form as $Tx_i'$, but with $h_i$ replaced by $Z(1-h_i)$, which implies

$$T = \sum_{i=0}^{2} (x_i + \sigma y_i)[h_i + Z(1-h_i)]. \tag{29c}$$

In the absence of selection on the modifier locus ($h_0 = h_1 = h_2 = h$), the population converges in a single generation to the state (27), with the $y_i$ now representing genotypic frequencies in females. Frequencies of allele $A$ among hermaphrodites, females, and pollen in the first generation correspond to

$$\begin{aligned} q' &= \frac{x_0' + x_1'/2}{\sum_i x_i'} = \frac{y_0' + y_1'/2}{\sum_i y_i'} \\ &= \frac{(x_0 + x_1/2) + (y_0 + y_1/2)\sigma + \sigma \sum_i y_i \left( \frac{x_0 + x_1/2}{\sum_i x_i} - \frac{y_0 + y_1/2}{\sum_i y_i} \right) /2}{\sum_i (x_i + y_i \sigma)}, \end{aligned}$$

for the $x_i$ and $y_i$ representing genotypic frequencies at initialization, and attain their common equilibrium value in two generations. The frequency of heterozygotes converges asymptotically, at rate $s_G/2$ (13), to

$$x_1 + y_1 = 2q(1-q)(1 - F_{neut}),$$

for $F_{neut}$ given by (24), with $s_G$ (13) substituted for $s$.

Near the state of fixation of the $a$ allele, the neutral transformation has a single eigenvalue of unity (corresponding to allele frequency), a single eigenvalue of $s_G/2$ (governing convergence of the frequency of heterozygotes to the value dictated by $F_{neut}$ and allele frequency), and two eigenvalues of zero (representing the convergence in two generations of allele frequencies in hermaphrodites and females to their common equilibrium value).

## 2.4 Weak selection

To explore the nature of selection on the sex ratio, we restrict most of the remaining analysis to weak selection on the modifier of the sex ratio, viewed as a perturbation from selective neutrality. Selective neutrality of the variation segregating at the focal locus entails that all genotypes induce identical hermaphrodite fractions:

$$h_0 = h_1 = h_2. \tag{30}$$

Weak selection implies that differences among genotypes,

$$\begin{aligned} d_0 &= h_0 - h_2 \\ d_1 &= h_1 - h_2, \end{aligned} \tag{31}$$

are sufficiently small to justify treating as negligible quantities of the second order in the $d_i$ or smaller. This assumption of weak selection at the modifier locus implies no restriction on the magnitude of differences viability or fertility between inbred and outbred offspring or between the sexes.

For each of the four models under study, we determine the conditions for local stability of the fixation of the $a$ allele against the introduction of the $A$ allele in small frequencies. In the preceding section, we have shown that in the absence of selection on the modifier locus (30), all systems show rapid convergence to a state in which associations between genes within genotypes reflect inbreeding and associations between allele frequency and sex are absent. For each model, we enumerated the eigenvalues of the neutral transformation: a single eigenvalue of unity (representing allele frequency) and a single eigenvalue of $s/2$ (reflecting asymptotic convergence of the frequency of heterozygotes), with any additional eigenvalues corresponding to zero. Because eigenvalues are continuous in complex space (*e.g.*, Serre 2010, Chapter 5), the eigenvalues of the perturbed (weak-selection) transformation depart continuously in the $d_i$ (31) from those of the neutral transformation. Accordingly, the dominant eigenvalue of the weak-selection transformation lies near unity, with the moduli of the other eigenvalues remaining strictly less than unity. Because the maternal control models have two eigenvalues of zero under neutrality, the perturbed transformation may have conjugate pairs of imaginary eigenvalues. Even so, any imaginary eigenvalues do not determine asymptotic local stability because the dominant eigenvalue of a non-negative matrix corresponds to a simple, real root of the characteristic polynomial (Gantmacher 1959). Accordingly, the dominant eigenvalue of the perturbed transformation lies near unity, with the moduli of the other eigenvalues remaining strictly less than unity. These properties of the weak-selection transformation imply that examination of the sign of the characteristic polynomial of the local stability matrix evaluated at unity is sufficient to determine local stability.

While a conventional local stability analysis provides a full determination of the fate of modifiers with weak effects on sex expression, we further undertake to elucidate the process of evolution by interpreting the results of our local stability analysis in terms of the Li-Price equation (Li 1967; Price 1970). Appendix A describes this method, which modifies an approach developed previously (Uyenoyama 1988, 1991).

# 3 Analysis

We perform local stability analyses for each of the four multidimensional models of the evolutionary modification of sex expression in androdioecious and gynodioecious populations (Section 2.3). We first determine the change of basis used in local stability analysis for our models of androdioecy and gynodioecy, under zygote and maternal control of sex expression.

We then show that the candidate ESS sex ratios (Section 2.1) do in fact correspond to continuously stable strategies.

## 3.1 Evolution of androdioecy

### 3.1.1 Zygote control of sex expression

Under zygote control of sex expression (23), genotype $i$ occurs with frequency $z_i$, of which a proportion $h_i$ develop into hermaphrodites and the complement into males.

**Local stability condition:** A necessary condition for the exclusion of allele $A$ introduced in low frequency into a population monomorphic for the $a$ allele, which induces hermaphroditism at rate $h_2$, is positivity of the characteristic polynomial of the local stability matrix evaluated at unity. Under zygote control (23), this condition corresponds to

$$(h_2 - \hat{h})[d_0 h_2 s/2 + d_1 h_2(1-s) - d_0 d_1 s/2] > 0, \tag{32}$$

in which the uniparental proportion $s$ corresponds to $s_A$ (8), $\hat{h}$ to the ESS candidate (10c), $h_2$ the proportion of the common $aa$ genotype that develop into hermaphrodites, and the $d_i$ (31) the phenotypic deviations of genotypes bearing the rare $A$ allele. Under weak selection (small $d_i$), this condition reduces to

$$(h_2 - \hat{h})[d_0 s/2 + d_1(1-s)] > 0, \tag{33}$$

and is also sufficient for local stability. For the Kryptolebias model, in which males alone fertilize outcrossed eggs ($\omega = \infty$), we show in Appendix B that the sole condition for local stability corresponds to

$$(\hat{h} - h_2)(h_2 - r_L) > 0, \tag{34}$$

in which $r_L$ denotes the larger root of the bracketed term in (32), viewed as a quadratic in $h_2$, under arbitrary dominance levels and intensities of selection on the modifier of sex expression ($d_i$).

**Average effect of substitution:** A fundamental notion of heritability of sex expression is that hermaphrodites and gonochores differ in the frequencies of alleles that modify sex expression. In any generation, the difference in frequency of the $A$ allele between hermaphrodites and gonochores corresponds to

$$\frac{z_0 h_0 + z_1 h_1/2}{\sum_i z_i h_i} - \frac{z_0(1-h_0) + z_1(1-h_1)/2}{\sum_i z_i(1-h_i)} = \frac{z_0(h_0 - \bar{h}) + z_1(h_1 - \bar{h})/2}{\bar{h}(1-\bar{h})}, \tag{35}$$

for

$$\bar{h} = \sum_i z_i h_i.$$

This expression corresponds to the average effect of substitution (21), with the genotypic frequencies at the point of sex expression ($z_i$) assuming the role of the $u_i$ in Table 1.

**New basis system:** In accordance with (35), we designate as the new basis vectors near the fixation of the $a$ allele (small $z_0$ and $z_1$)

$$\begin{aligned} t_0 &= z_0 + z_1/2 \\ t_1 &= z_0 - (z_0 + z_1/2)F_{neut}, \end{aligned} \tag{36a}$$

for $F_{neut}$ corresponding to (24) with the uniparental fraction $s_A$ (8) substituted for $s$. To the first order in the frequencies of rare genotypes, the genotypic frequencies correspond to

$$\begin{aligned} z_0 &= t_0 F_{sel} \\ z_1 &= 2t_0(1 - F_{sel}), \end{aligned} \tag{36b}$$

for $F_{sel}$ the fixation index under weak selection. From (36) we obtain

$$F_{sel} = F_{neut} + \frac{t_1}{t_0}. \tag{37}$$

Near the fixation of the $a$ allele, the average effect of substitution (21) corresponds to

$$\frac{z_0 d_0 + z_1 d_1/2}{4(z_0 + z_1/2) - z_1} = \frac{d_0 F_{sel} + d_1(1 - F_{sel})}{1 + F_{sel}}. \tag{38}$$

For $F_{sel}$ determined at the key vector (A.1) defined in Appendix A, this expression (38) for the average effect of substitution corresponds to the bracketed factor in (32).

Under weak selection (31), $t_1$ is $O(d_i)$ (Appendix A), implying that the departure between $F_{sel}$ and $F_{neut}$ is also $O(d_i)$. To the first order in the intensity of selection on the modifier locus ($O(d_i)$), the average effect of substitution (38) corresponds to

$$\frac{d_0 F_{neut} + d_1(1 - F_{neut})}{1 + F_{neut}} = d_0 s/2 + d_1(1 - s),$$

in agreement with (33).

### 3.1.2 Maternal control of sex expression

Under maternal control of sex expression (26), genotype $i$ occurs with frequency $x_i$ among maternal parents, all of which are hermaphrodites, and with frequency $y_i$ among reproductive males.

**Local stability condition:** The conditions for local stability under maternal control mirror those under zygote control. The characteristic polynomial evaluated at unity is positive (necessary for local stability) only if (32) holds. Under weak selection (31), (33) provides the necessary and sufficient condition for local stability.

**Average effect of substitution:** To address heritability, we again address differences between hermaphrodites and gonochores in the frequency of a modifier of sex expression.

In the next generation forward in time, the difference in frequency of the $A$ allele between hermaphrodites and gonochores corresponds to

$$\begin{aligned}
&\frac{x_0' + x_1'/2}{\sum_i x_i'} - \frac{y_0' + y_1'/2}{\sum_i y_i'} \\
&\qquad = (1+s_A)/2 \left[ \frac{x_0 h_0 + x_1 h_1/2}{\sum_i x_i h_i} - \frac{x_0(1-h_0) + x_1(1-h_1)/2}{\sum_i x_i (1-h_i)} \right] \\
&\qquad = (1+s_A)/2 \left[ \frac{x_0(h_0 - \bar{h}) + x_1(h_1 - \bar{h})/2}{\bar{h}(1-\bar{h})} \right],
\end{aligned} \tag{39}$$

for

$$\bar{h} = \sum_i x_i h_i.$$

This expression suggests that the average effect of substitution corresponds to (21) with the $u_i$ replaced by

$$\frac{x_i(1+s_A)/2}{\sum_i x_i(1+s_A)/2} = \frac{x_i[\tilde{s}\tau + (1-\tilde{s})/2]}{\sum_i x_i[\tilde{s}\tau + (1-\tilde{s})/2]}. \tag{40}$$

Under maternal control model of androdioecy, the maternal genotypic frequencies ($x_i$) are weighted by the production of uniparental offspring, at rate

$$\frac{\tilde{s}\tau}{\tilde{s}\tau + (1-\tilde{s})/2},$$

and of biparental offspring, at rate

$$\frac{(1-\tilde{s})/2}{\tilde{s}\tau + (1-\tilde{s})/2},$$

in which the 1/2 appears to represent the relatedness of biparental offspring to their maternal parent relative to the relatedness of uniparental offspring.

**New basis system:** We use (40) to specify the change in basis. Under androdioecy, males contribute to future generations only through pollen or sperm. In populations fixed for the $a$ allele, the ratio of hermaphrodites to males at reproductive age corresponds to

$$\frac{x_2}{y_2} = \frac{h_2}{Z(1-h_2)}. \tag{41}$$

Near this fixation state, we designate as the new basis vectors

$$\begin{aligned}
t_0 &= x_0 + x_1/2 \\
t_1 &= x_0 - (x_0 + x_1/2) F_{neut} \\
t_2 &= \frac{x_0 + x_1/2}{h_2} - \frac{y_0 + y_1/2}{Z(1-h_2)},
\end{aligned} \tag{42}$$

for $F_{neut}$ corresponding to (24) with the uniparental fraction $s_A$ (8) substituted for $s$.

At the key vector (A.1) defined in Appendix A, $t_2$ (42), representing the difference in allele frequency between hermaphrodites and males, is proportional to the average effect of substitution (38). Also at this key vector, the fixation index under selection $F_{sel}$ corresponds to (37) and the average effect of substitution (38) again corresponds to the bracketed factor in (32).

## 3.2 Evolution of gynodioecy

### 3.2.1 Zygote control of sex expression

For the zygote control model of gynodioecy (25), the condition for positivity of the characteristic polynomial of the local stability matrix evaluated at unity is identical to (32), with the uniparental proportion $s$ now corresponding to $s_G$ (13) and $\hat{h}$ to the ESS candidate (17). Under weak selection (31), (33) provides the necessary condition for local stability of the fixation state.

Also identical to the expressions under zygote control of androdioecy are the average effect of substitution (35) and the definition of the new basis system (36), but with $s_G$ (13) substituted for $s$ in $F_{neut}$ (24).

### 3.2.2 Maternal control of sex expression

**Local stability condition:** For the maternal control model (29), the condition for local stability under weak selection corresponds to

$$(h_2 - \hat{h})\{B[h_2 + (1-h_2)Z\sigma](1+s_G) - (d_0 - d_1)[d_0 s_G + d_1(1-s_G)]s_G Z\sigma/2\} > 0, \quad (43)$$

in which $s_G$ corresponds to the uniparental proportion (13), $\hat{h}$ to the ESS candidate (17), and $B$ the bracketed factor in (32):

$$B = d_0 h_2 s_G/2 + d_1 h_2(1-s_G) - d_0 d_1 s_G/2. \quad (44)$$

Under weak selection (31), (43) reduces to (33), which provides the necessary and sufficient condition for local stability of the fixation state.

**Average effect of substitution:** To address heritability, we return to (39). From the full system of recursions for maternal control of sex expression (29), we obtain

$$\begin{aligned}
&\frac{x_0' + x_1'/2}{\sum_i x_i'} - \frac{y_0' + y_1'/2}{\sum_i y_i'} \\
&\qquad = \Gamma_1 \left( \frac{x_0 + x_1/2}{\sum_i x_i} - \frac{y_0 + y_1/2}{\sum_i y_i} \right) \\
&\qquad\quad + \Gamma_2 \{ (1-s)[x_0(h_0 - \bar{h}) + x_1(h_1 - \bar{h})/2] \\
&\qquad\qquad + \sigma[y_0(h_0 - \bar{h}) + y_1(h_1 - \bar{h})/2] \}
\end{aligned}$$

in which

$$\Gamma_1 = \frac{\sigma s \sum_i y_i \sum_i x_i (h_i - \bar{h})}{2 \sum_i (x_i + \sigma y_i) h_i \sum_i (x_i + y_i \sigma)(1 - h_i)}$$
$$\Gamma_2 = \frac{\sum_i (x_i + y_i \sigma)}{2 \sum_i (x_i + \sigma y_i) h_i \sum_i (x_i + y_i \sigma)(1 - h_i)}$$

and

$$\bar{h} = \frac{\sum_i \{x_i(1+s) + y_i \sigma\} h_i}{\sum_i \{x_i(1+s) + y_i \sigma\}}.$$

Under weak selection, for which terms of the form $(h_i - h_j)$ are small, the difference in allele frequency between the sexes are also small, with the difference converging rapidly to

$$\frac{x_0' + x_1'/2}{\sum_i x_i'} - \frac{y_0' + y_1'/2}{\sum_i y_i'} = \frac{E}{2h(1-h)\sum_i (x_i + y_i \sigma)} + o(d) \tag{45a}$$

in which

$$E = (1+s)[x_0(h_0 - \bar{h}) + x_1(h_1 - \bar{h})/2] + \sigma[y_0(h_0 - \bar{h}) + y_1(h_1 - \bar{h})/2], \tag{45b}$$

$h$ represents any of the $h_i$, and $o(d)$ comprises quantities smaller than terms of the form $(h_i - h_j)$.

Expression (45b) suggests that the average effect of substitution corresponds to (21) with the $u_i$ replaced by

$$\frac{x_i[\tilde{s}\tau + (1-\tilde{s})/2] + y_i \tilde{\sigma}/2}{\sum_i \{x_i[\tilde{s}\tau + (1-\tilde{s})/2] + y_i \tilde{\sigma}/2\}}. \tag{46}$$

A major feature that distinguishes this gynodioecy model from the corresponding androdioecy model (39) is that gonochores (females) as well as hermaphrodites may serve as maternal parents, the individuals that control sex expression. Comparison of (40) and (46) indicates that the weighting of the contributions to the offspring generation of hermaphroditic to female maternal parents corresponds to

$$\frac{\tilde{s}\tau + (1-\tilde{s})/2}{\tilde{\sigma}/2} = \frac{(1+s)}{\sigma}, \tag{47}$$

implying a twofold weighting of uniparental offspring relative to biparental offspring.

**New basis system:** In defining the new basis system, we adopt the weighted average of allele frequencies in hermaphrodites and females described in (47):

$$\begin{aligned}
t_0 &= \frac{(x_0 + x_1/2)(1+s) + (y_0 + y_1/2)\sigma}{h_2(1+s) + Z(1-h_2)\sigma} \\
t_1 &= \frac{[x_0 - (x_0 + x_1/2)F_{neut}](1+s) + [y_0 - (y_0 + y_1/2)F_{neut}]\sigma}{h_2(1+s) + Z(1-h_2)\sigma} \\
t_2 &= \frac{x_0 + x_1/2}{h_2} - \frac{y_0 + y_1/2}{Z(1-h_2)} \\
t_3 &= \frac{x_0}{h_2} - \frac{y_0}{Z(1-h_2)}
\end{aligned} \tag{48}$$

for $F_{neut}$ corresponding to (24) with the uniparental fraction $s_G$ (13) substituted for $s$. These expressions reflect that near the fixation state, the ratio of hermaphrodites to gonochores in the population ($x/y$) lies close to (41).

At the key vector (A.1) defined in Appendix A, both $t_2$ (48), representing the difference in allele frequency between hermaphrodites and males, and the factor of $(h_2 - \hat{h})$ in local stability condition (43) are proportional to the average effect of substitution (45b).

# 4 Data analysis

Redelings *et al.* (2015) developed a Bayesian method for the analysis of multilocus data sampled from populations reproducing through pure hermaphroditism, androdioecy, or gynodioecy. Using an explicitly coalescent-based framework, it generates posterior distributions for the uniparental fraction (probability that a random individual is uniparental), the analogue to estimates of selfing rates generated by earlier methods (*e.g.*, Ritland 2002; Enjalbert and David 2000; David *et al.* 2007).

In any empirical investigation, modifiers of the sex ratio may have not yet evolved to the ESS even if the model is appropriate. Subject to this caveat, we use our new theoretical results to infer the viability of gonochores (males or females) relative to hermaphrodites in the natural populations analyzed by Redelings *et al.* (2015).

Using microsatellite data derived from natural populations of the androdioecious killifish *Kryptolebias marmoratus* (Tatarenkov *et al.* 2012) and the gynodioecious Hawaiian endemic *Schiedea salicaria* (Wallace *et al.* 2011), Redelings *et al.* (2015) generated posterior distributions of the basic parameters of the models, including the population sex ratio among reproductives (7a). Those estimates imply posterior distributions of $C$, the collective contribution of hermaphrodites to the next generation (Fig. 1), from which we infer the sex ratio at the juvenile stage (7b). Under the assumption that the natural populations under study have converged on the attracting ESS sex ratio, we use the departure between the sex ratios at the two points in the life cycle ((7a) and (7b)) to obtain the posterior distribution of $Z$.

Figure 4 presents posterior distributions of $Z$ in the Schiedea and Kryptolebias populations. We find little evidence of a difference in viability between females and hermaphrodites in the gynodioecious *S. salicaria* (median=1.08, 95% BCI=(0.34, 1.78)), in which the Bayesian Credible Interval (BCI) denotes the interval comprising the highest posterior density. In contrast, male *K. marmoratus* appear to have substantially lower viability than hermaphrodites in both the BP population (median=0.45, 95% BCI=(0.20, 0.81)) and the TC population (median=0.48, 95% BCI=(0.25, 0.77)), even though the frequency of males is several-fold higher in the TC population (0.17 versus 0.01; Turner *et al.* 1992; Tatarenkov *et al.* 2012; Mackiewicz *et al.* 2006).

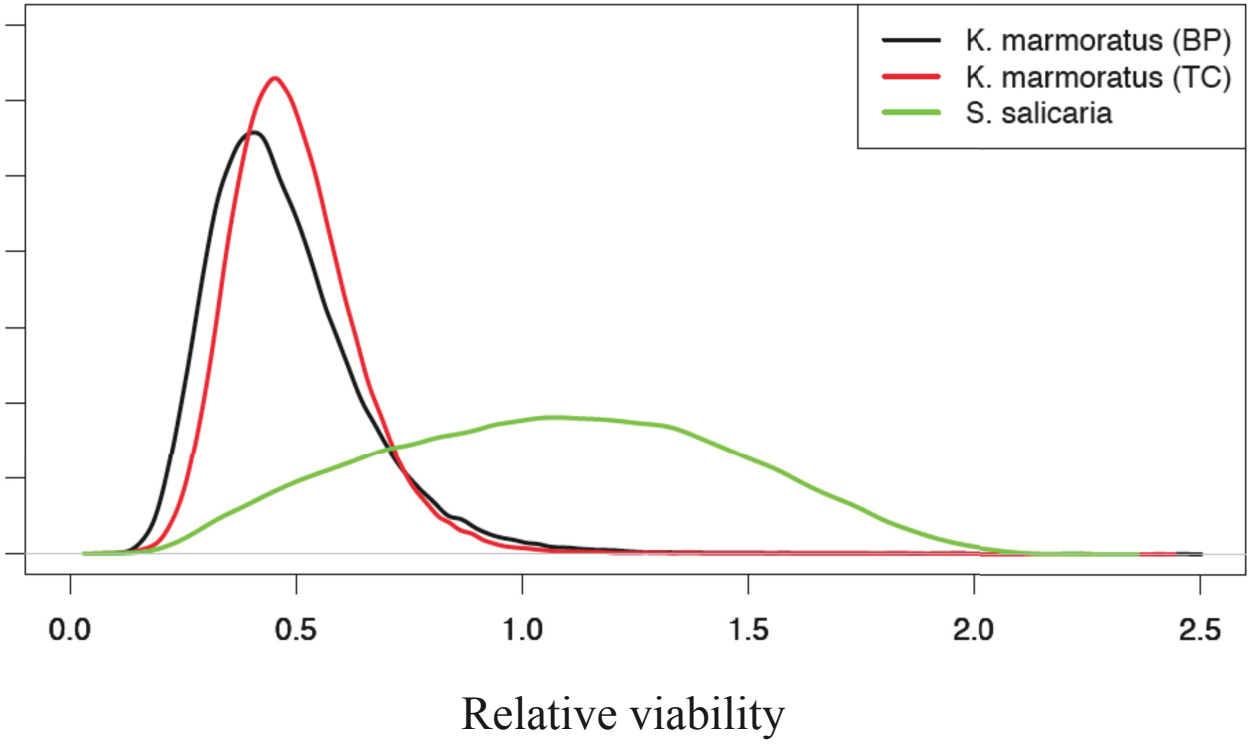


Figure 4: Posterior distributions of the viability of gonochores relative to hermaphrodites ($Z$).

# 5 Discussion

We here explore the evolution of androdioecy and gynodioecy under the influence of autosomal modifiers of weak effect. Our study unifies full multi-dimensional local stability analysis with the heuristically-appealing Li-Price equation (Li 1967; Price 1970) by identifying a point in the neighborhood of a fixation state from which a single-generation step indicates the asymptotic behavior of a rare, introduced allele initiated at an arbitrary location near the fixation state. In addition, we use our theoretical analysis to infer the viability of gonochores (males or females) relative to hermaphrodites in the gynodioecious plant *Schiedea salicaria* and the the androdioecious killifish *Kryptolebias marmoratus*. We find that female and hermaphrodite *Schiedea salicaria* appear to have similar viabilities, but that male *Kryptolebias marmoratus* survive to reproductive age at rates significantly lower than do hermaphrodites.

## 5.1 Relative viability of gonochores

We have explored the evolution of modifiers of the expression of zygote sex under the control of the zygotes themselves or of their maternal parents in androdioecious and gynodioecious systems. Our analysis shows that the central determinant of the evolutionary modification of the sex ratio corresponds to $C$ (1), the collective contribution of hermaphroditic parents to autosomal genes transmitted to offspring (Fisher 1930). Natural selection on modifiers of weak effect promotes convergence to the evolutionarily stable sex ratio among juveniles of

$$\frac{\hat{h}}{1-\hat{h}} = \frac{C}{1-C}$$

(7b), for $\hat{h}$ the ESS proportion of hermaphrodites. In the absence of sex-specific differences in rate of survival to reproductive age ($Z = 1$), this sex ratio holds at reproductive age at well, implying maximization ($R = 1$) of relative effective number (2). Here, we interpret a departure of relative effective number $R$ from unity as an indication of a change in the population sex ratio from the juvenile stage to reproductive age, at which point the ESS corresponds to

$$\frac{h^*}{1-h^*} = \frac{C}{Z(1-C)}$$

(7a), for $h^*$ the proportion of hermaphrodites among offspring of reproductive age and $Z$ the relative rate of survival of gonochores from the juvenile to the adult stages. As the Bayesian MCMC method of Redelings *et al.* (2015) permits inference of $h^*$ and $C$, it also yields posterior distributions for $Z$, the relative viability of gonochores (Fig. 4).

Inference of near-maximal relative effective number (Fig. 2) for a natural population of the gynodioecious *Schiedea salicaria* (Wallace *et al.* 2011) suggests close convergence to the ESS (7b). In agreement, the posterior distribution of $Z$ (Fig. 4) provides little evidence of differential viability between the sexes. In contrast, Figure 4 indicates that males of the androdioecious killifish *Kryptolebias marmoratus* (Kelley *et al.* 2016) appear to have about

twofold lower viability than hermaphrodites. Our analysis suggests similar male viabilities in the highly inbred BP population, in which reproductively mature males are very rare (posterior median = 1%), and the more outbred TC population, in which they are more abundant (posterior median = 17%, Redelings *et al.* 2015).

Turner *et al.* (2006) conducted common garden experiments to address male development in the killifish *K. marmoratus*, an emerging model system for environmental sex determination (Kelley *et al.* 2016). Lines derived from the progeny of individual hermaphrodites derived from natural populations in Belize that showed marked differences in the proportion of adult males were reared in the laboratory under identical conditions. While fewer males appeared in broods derived from the rare-male population, both sets of broods showed substantially higher frequencies of males than observed in the natural populations from which they were derived. Turner *et al.* (2006) proposed the novel hypothesis that outcrossing or high heterozygosity may directly induce male development. Our inference that the rate of survival to reproductive age of males is less than half the rate of hermaphrodites (Fig. 4) is consistent with the discrepancies in sex ratio between the laboratory and the wild. Turner *et al.* (2006) described the orange-hued mature males as "highly conspicuous." The considerable body of work on guppies indicates that predation can generate intense selection, with various indices of crypsis responding rapidly to predator abundance under both laboratory and field conditions (Endler 1980; Reznick *et al.* 1996).

Turner *et al.* (2006) suggested that under current theoretical models of androdioecy, maintenance of males in highly inbred populations of *K. marmoratus*, would require "implausibly large" male fertility. Low viability of males would further increase the stringency of the condition (10b). However, our analysis indicates that the existence of any viable biparental offspring ($1 > s_A$) is sufficient to favor the maintenance of males if males alone fertilize eggs that are not self-fertilized ($\omega = \infty$, Furness *et al.* 2015).

## 5.2 Evolution by means of major and minor genes

Previous studies have shown that complete dominance of a major gene inducing gonochorous development in a zygote implies the direct convergence of the sex ratio to the ESS (7a) under both androdioecy (Ross and Weir 1976; Wolf and Takebayashi 2004) and gynodioecy (Ross and Weir 1975). The proportion of offspring that have a gonochorous parent corresponds to $2(1-C)$, for $(1-C)$ the probability that an autosomal gene randomly sampled from the offspring that survive inbreeding depression derive from a gonochorous parent (1). We now restrict consideration to complete dominance of the gonochore allele ($h_0 = 0$) and determination of zygote sex by its own genotype. Because gonochorous parents transmit the dominant allele to half their offspring, the sex ratio among offspring of reproductive age corresponds to

$$\frac{h'}{1-h'} = \frac{C}{Z2(1-C)/2},$$

for $h'$ the proportion of hermaphrodites among offspring. Accordingly, the ESS (7a) represents the equilibrium population sex ratio among reproductives. This property holds neither

under other dominance schemes (*e.g.*, recessive gynodioecy, Ross and Weir 1975) nor under maternal determination of zygote sex, even with complete dominance.

Upon the direct evolution of a major nuclear gene to the ESS, any selective pressure on minor genes for sex expression to modify the sex ratio toward the ESS ceases. In all other cases, natural selection favors modifier genes across with genome that reduce the disparity between the population sex ratio and the ESS, even in the face of opposing selection against such pleiotropic effects on the sex ratio.

In their analysis of sex within broods generated by controlled crosses between females and hermaphrodites of the gynodioecious *Schiedea salicaria*, Weller and Sakai (1991) recognized two major groups of hermaphroditic pollen donors: those that generated hermaphroditic offspring almost exclusively and those that generated the two sexes in approximately equal proportions. Weller and Sakai (1991) proposed that male sterility derives from a recessive allele at a single locus. The analysis of Ross and Weir (1975) shows that recessivity of a major allele for male sterility implies an equilibrium population sex ratio that departs from the ESS (7b). Even so, Weller and Sakai (2005) found approximate agreement between the population sex ratio and the predicted ESS, and our inference of near-maximal values of relative effective number ($R$ near unity in Fig. 2) suggests that the ESS (7b) has in fact been attained, provided that females and hermaphrodites have equal viability ($Z = 1$). Reconciliation between the proposed genetic basis of male sterility in *S. salicaria* and the apparent attainment of the ESS (7b) suggests that modifier loci distinct from the major gene may have induced the convergence of the population sex ratio to the evolutionarily stable strategy.

## 5.3 Evolutionarily stable strategies

Our analysis shows that the sex ratios proposed by Lloyd (1975) correspond to attracting evolutionarily stable strategies (ESSs) under arbitrary schemes for dominance of rare alleles introduced at a monomorphic locus that modifies sex expression. Fully-specified models of sex expression with inbreeding are inherently multi-dimensional, reflecting multiple genotypes in both gonochores and hermaphrodites in our studies. Imposition of restrictions on dominance can reduce the dimensionality of the system, permitting specification of equilibrium states on the basis of allele frequency alone (Ross and Weir 1975, 1976; Charlesworth and Charlesworth 1978; Wolf and Takebayashi 2004). We regard the removal of restrictions on dominance as essential to the spirit of an ESS analysis, under which the attainment of the ESS reflects the filtering of newly-arisen modifier alleles throughout the genome.

The conceptual origins of the Li-Price equation (Li 1967; Price 1970) lie in Robertson's (1966) exploration of the effects of culling, on the basis of informal criteria, on the genetic variance of a desired trait (high milk yield in dairy cattle). With respect to the evolution of mating systems, sex may influence various components of transmission of genes to future generations, possibly including relatedness (Lloyd 1975). Here, the focal trait corresponds to the propensity of a zygote to develop into a gonochore or a hermaphrodite under the control

of the genotype at a modifier locus of its maternal parent (maternal control models) or its own genotype (zygote control models).

The heuristically-appealing Li-Price equation (22) provides a condition for the change over a single generation in the frequency of an allele at a locus modifying the focal trait. Our analysis relates this one-dimensional, one-generational description of the evolutionary process to the asymptotic conditions for invasion of a rare modifier allele in multi-dimensional models of sex expression with inbreeding under the assumption of weak selection. Weak selection entails that alleles segregating at modifier loci have small effects on the prevailing rate of sex expression. This assumption permits selection of any intensity on relative viability or fertility of uniparentals or gonochores relative to hermaphrodites. We define a key initial state (A.1) in the full multi-dimensional space such that the change in frequency of the rare allele over a single generation starting from this state indicates its asymptotic fate (invasion or extinction) starting from arbitrary states in a sufficiently small neighborhood of the fixation state (Appendix A). We show that the change in frequency of the rare allele is proportional to the value of the characteristic polynomial of the full transformation evaluated at unity. While this criterion provides necessary and sufficient conditions for local stability, strong selection (introduction of genes with major effects on sex expression) may cause the sign of the characteristic polynomial evaluated at unity to become insufficient as an indicator of local stability, with the key initial state (A.1) invalid or undefined (Fig. 5).

Central to the Li-Price equation (22), which relates the focal trait to fitness, is the average effect of substitution (Fisher 1941), an index of the influence of segregating variation on the trait. In our models, comprising two sex forms and either zygote or maternal control of sex expression, the average effect may be determined with respect to the genotypic distribution ($u_i$ in Table 1) in more than one component of the population. An additional question concerns the relevance of relatedness of the controlling genotype to the two sex forms. Our approach entails permitting the answers to these questions emerge naturally from the models themselves. We adopt a notion of heritability that reflects associations between sex and allele frequency. This approach indicates that under zygote control of androdioecy (23) and gynodioecy (25), the average effect is defined with respect to genotypic frequencies among zygotes at the point of sex expression (35). In this case, the controlling entities (zygotes) are equally related to themselves regardless of sex.

In contrast, relatedness plays a role under maternal control of sex expression in offspring. For the androdioecy model (26), hermaphrodites alone determine offspring number, with gonochores (males) serving only as pollen or sperm donors. Under our notion of heritability, the average effect is defined with respect to genotypic frequencies among maternal parents (hermaphrodites) at reproductive age (39). Uniparental offspring bear twofold higher relatedness to their maternal parents than do biparental offspring, irrespective of the sex of the offspring (39).

Among the unique features of maternal control of gynodioecy (29) is that gonochores (females) as well as hermaphrodites contribute to offspring number. Accordingly, the average effect of substitution depends on both sexes, with the offspring of females weighted by a factor of 1/2, reflecting their biparental derivation, and the biparental and uniparental offspring of

hermaphrodites weighted by 1/2 and 1, respectively (47). Our analysis confirms that under this definition, the asymptotic local stability conditions agree with the Li-Price equation (22).

### 5.4 Heritability

In populations in which modifiers of sex expression segregate, the average effect of substitution (21) at a modifier locus is proportional to the difference in allele frequency between gonochores and hermaphrodites. This property holds in all models studied: both zygote and maternal control of androdioecy and gynodioecy ((35), (39), (45)). Because only hermaphrodites can self-fertilize in our models, heritability of sex may induce heritability of level of inbreeding. For example, hermaphroditic offspring may have a higher chance both of having descended from hermaphroditic parents and of being uniparental. Consistent with this expectation is the observation that gonochorous adults are more outbred than hermaphroditic adults (Wolff *et al.* 1988; Collin and Shykoff 2003) in some gynodioecious species, including *Schiedea salicaria* (Weller and Sakai 2005).

The uniparental proportion under androdioecy (8) or gynodioecy (13) corresponds to the fraction of juveniles that are uniparental (Fig. 3). Among the key parameters of the coalescent process that underlies the determination of the likelihood in the Bayesian sampler of Redelings *et al.* (2015) is the uniparental proportion among adults. In the absence of sex-specific viability selection ($Z = 1$), the uniparental proportion is identical between the juvenile and adult stages. Alternatively ($Z \neq 1$), the uniparental proportions at the juvenile and adult stages may differ if sex expression is heritable. As a consequence, our estimates of the relative viability of gonochores ($Z$) are subject to the assumption that the population has attained the ESS level of sex expression as a genetically monomorphic state: the outcome of filtering of newly-arisen mutations of minor effect, for example. In particular, the segregation of modifiers of sex expression in *S. salicaria* (Weller and Sakai 2005) may violate this assumption.

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## References

Bailey, M. F. and Delph, L. F., 2007. A field guide to models of sex-ratio evolution in gynodioecious species. *Oikos* **116**, 1609–1617.

Campbell, D. R., Weller, S. G., Sakai, A. K., Culley, T. M., Dang, P. N., and Dunbar-Wallis,

A. K., 2010. Genetic variation and covariation in floral allocation of two species of *Schiedea* with contrasting levels of sexual dimorphism. *Evolution* **65**, 757–770.

Charlesworth, B. and Charlesworth, D., 1978. A model for the evolution of dioecy and gynodioecy. *Am. Nat.* **112**, 975–997.

Charnov, E. L., Maynard Smith, J., and Bull, J. J., 1976. Why be a hermaphrodite. *Nature* **263**, 125–126.

Christiansen, F. B., 1991. On conditions for evolutionary stability for a continuously varying character. *Am. Nat.* **138**, 37–50.

Cockerham, C. C. and Weir, B. S., 1968. Sib mating with two linked loci. *Genetics* **60**, 629–640.

Collin, C. L. and Shykoff, J. A., 2003. Outcrossing rates in the gynomonoecious-gynodioecious species *Dianthus silvestris* (Caryophyllaceae). *Amer. J. Bot.* **90**, 579–585.

Crow, J. F. and Denniston, C., 1988. Inbreeding and variance effective population numbers. *Evolution* **42**, 482–495.

David, P., Pujol, B., Viard, F., Castella, V., and Goudet, J., 2007. Reliable selfing rate estimates from imperfect population genetic data. *Mol Ecol* **16**, 2474–2487.

Denniston, C., 1978. An incorrect definition of fitness revisited. *Ann. Hum. Genet.* **42**, 77–85.

Edwards, A. W. F., 2000. Carl Düsing (1884) on *The Regulation of the Sex-Ratio*. *Theor. Pop. Biol.* **58**, 255–257.

Endler, J. A., 1980. Natural selection on color patterns in *Poecilia reticulate*. *evol* **34**, 76–91.

Enjalbert, J. and David, J. L., 2000. Inferring recent outcrossing rates using multilocus individual heterozygosity: application to evolving wheat populations. *Genetics* **156**, 1973–1982.

Eshel, I. and Motro, U., 1981. Kin selection and strong evolutionary stability of mutual help. *Theor. Pop. Biol.* **19**, 420–433.

Ewens, W. J., 1982. On the concept of effective population size. *Theor. Pop. Biol.* **21**, 373–378.

Fisher, R. A., 1930. The genetical theory of natural selection. Oxford Univ. Press, Oxford, first edition.

Fisher, R. A., 1941. Average excess and average effect of a gene substitution. *Ann. Eugen.* **11**, 53–63.

Fisher, R. A., 1958. The Genetical Theory of Natural Selection. Dover, New York, second edition.

Fishman, L. and Willis, J. H., 2006. A cytonuclear incompatibility causes anther sterility in *Mimulus* hybrids. *Evolution* **60**, 1372–1381.

Furness, A. I., Tatarenkov, A., and Avise, J. C., 2015. A genetic test for whether pairs of hermaphrodites can cross-fertilize in a selfing killifish. *J. Hered.* **106**, 749–752.

Gantmacher, F. R., 1959. The Theory of Matrices, volume II. Chelsea Publishing Co., New York.

Kelley, J. L., Yee, M.-C., Brown, A. P., Richardson, R. R., Tatarenkov, A., Lee, C. C., Harkins, T. T., Bustamante, C. D., and Earley, R. L., 2016. The genome of the self-fertilizing mangrove rivulus fish, *Kryptolebias marmoratus*: a model for studying phenotypic plasticity and adaptations to extreme environments. *Genome Biol. Evol.* **8**, 2145–2154.

Laporte, V., Cuguen, J., and Couvet, D., 2000. Effective population sizes for cytoplasmic and nuclear genes in a gynodioecious species: the role of the sex determination system. *Genetics* **154**, 447–458.

Li, C. C., 1967. Fundamental theorem of natural selection. *Nature* **214**, 505–506.

Lloyd, D. G., 1975. The maintenance of gynodioecy and androdioecy in angiosperms. *Genetica* **45**, 325–339.

Mackiewicz, M., Tatarenkov, A., Taylor, D. S., Turner, B. J., and Avise, J. C., 2006. Extensive outcrossing and androdioecy in a vertebrate species that otherwise reproduces as a self-fertilizing hermaphrodite. *Proc. Natl. Acad. Sci. (USA)* **103**, 9924–9928.

Maynard Smith, J. and Price, G. R., 1973. The logic of animal conflict. *Nature* **246**, 15–18.

McCauley, D. E. and Bailey, M. F., 2009. Recent advances in the study of gynodioecy: the interface between theory and empiricism. *Ann. Bot. (Lond)* **104**, 611–620.

Price, G. R., 1970. Selection and covariance. *Nature* **227**, 520–521.

Price, G. R., 1971. Fisher's "fundamental theorem" made clear. *Ann. Hum. Genet.* **36**, 126–140.

Redelings, B. D., Kumagai, S., Tatarenkov, A., Wang, L., Sakai, A. K., Weller, S. G., Culley, T. M., Avise, J. C., and Uyenoyama, M. K., 2015. A Bayesian approach to inferring rates of selfing and locus-specific mutation. *Genetics* **201**, 1171–1188.

Reznick, D. N., Butler IV, M. J., Rodd, F. H., and Ross, P., 1996. Life-history evolution in guppies (*Poecilia reticulata*) 6. differential mortality s a mechanism for natural selection. *evol* **50**, 1651–1660.

Ritland, K., 2002. Extensions of models for the estimation of mating systems using $n$ independent loci. *Heredity* **88**, 221–228.

Robertson, A., 1966. A mathematical model of the culling process in dairy cattle. *Animal Production* **8**, 95–108.

Ross, M. D. and Weir, B. S., 1975. Maintenance of male sterility in plant populations. III. Mixed selfing and random mating. *Heredity* **35**, 21–29.

Ross, M. D. and Weir, B. S., 1976. Maintenance of males and females in hermaphrodite populations and the evolution of dioecy. *Evolution* **30**, 425–441.

Schultz, S. T., 1994. Nucle-cytoplasmic male sterility and alternative routes to dioecy. *Evolution* **48**, 1933–1945.

Serre, D., 2010. Matrices: Theory and Applications. Springer, New York.

Shaw, R. D. and Mohler, J. D., 1953. The selective significance of the sex ratio. *Am. Nat.* **87**, 337–342.

Tatarenkov, A., Earley, R. L., Taylor, D. S., and Avise, J. C., 2012. Microevolutionary distribution of isogenicity in a self-fertilizing fish (*Kryptolebias marmoratus*) in the Florida Keys. *Integr. Comp. Biol.* **52**, 743–752.

Taylor, P. D., 1989. Evolutionary stability in one-parameter models under weak selection. *Theor. Pop. Biol.* **36**, 125–143.

Turner, B. J., Davis, W. P., and Taylor, D. S., 1992. Abundant males in populations of a selfing hermaphrodite fish, *Rivulus marmoratus*, from some Belize cays. *J. Fish Biol.* **40**, 307–310.

Turner, B. J., Fisher, M. T., Taylor, D. S., Davis, W. P., and Jarrett, B. L., 2006. Evolution of 'maleness' and outcrossing in a population of the self-fertilizing killifish, *Kryptolebias marmoratus*. *Evolutionary Ecology Research* **8**, 1475–1486.

Uyenoyama, M. K., 1988. On the evolution of genetic incompatibility systems. IV. Modification of response to an existing antigen polymorphism under partial selfing. *Theor. Pop. Biol.* **34**, 347–377.

Uyenoyama, M. K., 1991. On the evolution of genetic incompatibility systems. VI. A three-locus modifier model for the origin of gametophytic self-incompatibility. *Genetics* **128**, 453–459.

Wallace, L. E., Culley, T. M., Weller, S. G., Sakai, A. K., Kuenzi, A., Roy, T., Wagner, W. L., and Nepokroeff, M., 2011. Asymmetrical gene flow in a hybrid zone of Hawaiian Schiedea (Caryophyllaceae) species with contrasting mating systems. *PLoS ONE* **6**, e24845. doi:10.1371/journal.pone.0024845.

Weller, S. G. and Sakai, A. K., 1991. The genetic basis of male sterility in *Schiedea* (Caryophyllaceae), an endemic Hawaiian species. *Heredity* **67**, 265–273.

Weller, S. G. and Sakai, A. K., 2005. Inbreeding and resource allocation in *Schiedea salicaria* (Caryophyllaceae), a gynodioecious species. *J. Evol. Biol.* **18**, 301–308.

Wolf, D. E. and Takebayashi, N., 2004. Pollen limitation and the evolution of androdioecy from dioecy. *Am. Nat.* **163**, 122–137.

Wolff, K., Friso, B., and van Damme, J. M. M., 1988. Outcrossing rates and male sterility in natural populations of *Plantago coronopus*. *Theor. Appl. Genet.* **76**, 190–196.

Wright, S., 1931. Evolution in Mendelian populations. *Genetics* **16**, 97–159.

Wright, S., 1933. Inbreeding and homozygosis. *Proc. Natl. Acad. Sci. (USA)* **19**, 411–420.

# Appendix A Change of basis

Here, we describe the relationship between the one-generational, one-dimensional description of evolution given by the Li-Price equation (22) and a full asymptotic, multi-dimensional local stability analysis. We describe a state of the population from which the change in allele frequency over a single generation does in fact correctly reflect the asymptotic condition for initial increase in the full multi-dimensional system under weak selection.

## A.1 Weak selection

Under selective neutrality of variation at the modifier locus, the genotypic frequencies initiated at any state comprising both alleles rapidly converge to a configuration characterized by equality between sex forms of genotypic frequencies ($x_i = y_i$) and fixation index (Wright 1933) given by (24). In the absence of differences among genotypes in sex expression (30), the multi-dimensional transformations we address (Section (2.3)) have a dominant eigenvalue of unity, reflecting preservation of allele frequency, with all remaining eigenvalues, corresponding to classical measures of disequilibrium, having moduli strictly less than unity. Weak-selection systems (31) represent perturbations in parameter space of such neutral transformations. For cases, including the maternal control model of gynodioecy (29), in which the neutral transformation has repeated eigenvalues, the perturbed transformation may have conjugate pairs of imaginary eigenvalues. Even so, any imaginary eigenvalues do not determine asymptotic local stability because the dominant eigenvalue of a non-negative matrix corresponds to a simple, real root of the characteristic polynomial (Gantmacher 1959). Because eigenvalues are continuous in complex space (*e.g.*, Serre 2010, Chapter 5), the eigenvalues of the weak-selection transformation depart continuously in the $d_i = (h_i - h_2)$ (31) from those of the neutral transformation. In particular, the dominant eigenvalue of the local stability matrix under weak selection lies near unity, with the moduli of the other eigenvalues remaining strictly less than unity. As a consequence, examination of the value of the characteristic polynomial of the local stability matrix under weak selection is sufficient to establish local stability.

## A.2 Elucidating the Li-Price equation

To relate the Li-Price equation (22) to the full multi-dimensional local stability analysis, we introduce a change of basis from the genotypic frequencies of the rare genotypes ($AA$ and $Aa$) in hermaphrodites and gonochores to allele frequency and disequilibrium measures. Here, measures of disequilibrium reflect any departures of variables other than allele frequency from their equilibrium values under the mating system in the absence of selection on the modifier locus ($h_0 = h_1 = h_2$). In particular, disequilibrium corresponds to the departure of the frequency of heterozygotes ($Aa$) from the frequency associated with $F_{neut}$ (24) and not, in particular, from Hardy-Weinberg proportions ($F = 0$).

**Change of basis:** We determine a key vector such that the direction of change in allele frequency over a single generation starting from this vector reflects the asymptotic behavior of the system starting from an arbitrary position in the neighborhood of the fixation state. Let $\boldsymbol{M}$ denote the local stability matrix under the original basis system. Because $\boldsymbol{M}$ is a non-negative matrix, its dominant eigenvalue is non-negative and corresponds to a simple root of its characteristic polynomial (Gantmacher 1959). Under the new basis, the local stability matrix corresponds to

$$\boldsymbol{N} = \boldsymbol{AMB},$$

for $\boldsymbol{A}$ translating from the old basis to the new basis and $\boldsymbol{B}$ translating from the new basis to the old basis ($\boldsymbol{AB} = \boldsymbol{I}$). For $\boldsymbol{z}$ an arbitrary vector in the neighborhood of the fixation state,

$$(\boldsymbol{I} - \boldsymbol{N})\boldsymbol{z}$$

describes change over a single generation. We define *key vector* $\tilde{\boldsymbol{z}}$ such that change may occur only in the first dimension (allele frequency), irrespective of the magnitude of disequilibria in other dimensions:

$$(\boldsymbol{I} - \boldsymbol{N})\tilde{\boldsymbol{z}} = \begin{pmatrix} -\Delta z \\ 0 \\ \vdots \\ 0 \end{pmatrix}, \tag{A.1}$$

in which $\Delta z$ denotes the change in allele frequency over a single generation. For $\boldsymbol{M}$ and $\boldsymbol{N}$ $n$-dimensional matrices, $\tilde{\boldsymbol{z}}$ is determined by the last $(n-1)$ rows of $(\boldsymbol{I} - \boldsymbol{N})\tilde{\boldsymbol{z}}$.

**Asymptotic behavior:** Here, we show that under weak selection (31), a one-generation step from key vector $\tilde{\boldsymbol{z}}$ (A.1) indicates the asymptotic behavior of the system initiated from an arbitrary location in the neighborhood of the fixation state.

Let $\boldsymbol{X}$ represent the matrix obtained by replacing the first column of an $n$-dimensional identity matrix by $\tilde{\boldsymbol{z}}$. Multiplication of $(\boldsymbol{I} - \boldsymbol{N})$ by $\boldsymbol{X}$ on the right produces

$$(\boldsymbol{I} - \boldsymbol{N})\boldsymbol{X} = \begin{pmatrix} -\Delta z & \boldsymbol{R_2} \\ \boldsymbol{R_1} & (\boldsymbol{I} - \boldsymbol{N})_{n-1} \end{pmatrix}, \tag{A.2}$$

in which $\boldsymbol{R_1}$ is an $(n-1)$-dimensional column vector of zeros, $\boldsymbol{R_2}$ is an $(n-1)$-dimensional row vector with elements equal to the corresponding elements of the first row of $(\boldsymbol{I} - \boldsymbol{N})$,

and $(\boldsymbol{I}-\boldsymbol{N})_{n-1}$ is the matrix obtained by removing the first row and column from $(\boldsymbol{I}-\boldsymbol{N})$. Taking the determinant of both sides of (A.2) produces

$$\mathrm{Det}[\boldsymbol{I}-\boldsymbol{N}]\mathrm{Det}[\boldsymbol{X}] = -\Delta z \mathrm{Det}[(\boldsymbol{I}-\boldsymbol{N})_{n-1}], \tag{A.3a}$$

for $\mathrm{Det}[(\boldsymbol{I}-\boldsymbol{N})_{n-1}]$ the principal minor obtained by deleting the first row and column of $(\boldsymbol{I}-\boldsymbol{N})$.

To achieve our objective of relating the Li-Price equation (22) to a full multi-dimensional local stability analysis, we demonstrate that

$$\mathrm{Det}[\boldsymbol{I}-\boldsymbol{N}] \propto -\Delta z$$

under weak selection ($d_i$ near zero). This expression implies that the direction of change over a single generation of the system initiated at $\tilde{\boldsymbol{z}}$ (A.1) corresponds to the sign of the characteristic polynomial of the multi-dimensional stability matrix evaluated at unity. Weak selection (31) entails small differences among genotypes in sex expression (small $d_i = h_i - h_2$). Because $\Delta z$ is $O(d_i)$, (A.3a) implies

$$\mathrm{Det}[\boldsymbol{I}-\boldsymbol{N}]\mathrm{Det}[\boldsymbol{X}] = -\Delta z \mathrm{Det}[(\boldsymbol{I}-\boldsymbol{N}^*)_{n-1}] + o(d_i), \tag{A.3b}$$

for $\boldsymbol{N}^*$ the linearized transition matrix under neutrality ($d_i = 0$). To show that

$$\mathrm{Det}[(\boldsymbol{I}-\boldsymbol{N}^*)_{n-1}] > 0, \tag{A.4}$$

we note that under neutrality, the absence of all disequilibria implies invariant gene frequency in all models studied here (Section 2.3). Accordingly,

$$(\boldsymbol{I}-\boldsymbol{N}^*)\boldsymbol{e} = \boldsymbol{0},$$

in which $\boldsymbol{N}^*$ denotes the linearized transition matrix under neutrality and $\boldsymbol{e}$ the unit vector with first element equal to 1 and zeros elsewhere. This expression implies that the element in the first column and row of $\boldsymbol{N}^*$ corresponds to unity. Further, that the neutral system converges to the state in which all disequilibria are absent implies that all elements in the first column of $\boldsymbol{N}^*$ other than the first are zero. As a result, $\boldsymbol{N}^*$ has the form

$$\boldsymbol{N}^* = \begin{pmatrix} 1 & \cdots \\ \boldsymbol{R_1} & (\boldsymbol{N}^*)_{n-1} \end{pmatrix},$$

in which $(\boldsymbol{N}^*)_{n-1}$ denotes the submatrix obtained by removing the first row and column from $\boldsymbol{N}^*$ and $\boldsymbol{R_1}$ is again an $(n-1)$-dimensional column vector of zeros. The characteristic polynomial of $\boldsymbol{N}^*$,

$$\mathrm{Det}[\lambda\boldsymbol{I}-\boldsymbol{N}^*] = (\lambda-1)\mathrm{Det}[(\lambda\boldsymbol{I}-\boldsymbol{N}^*)_{n-1}] = 0,$$

has a unit eigenvalue (corresponding to allele frequency), with the remaining eigenvalues (corresponding to disequilibria) given by the roots of $\mathrm{Det}[(\lambda\boldsymbol{I}-\boldsymbol{N}^*)_{n-1}]$. That all eigenvalues associated with disequilibria are strictly less than unity in absolute value implies (A.4).

# Appendix B Local stability analysis of Kryptolebias model under zygote control of sex

We address the local stability of recursion system (23) near the state of fixation of the $a$ allele at the modifier locus. Under Kryptolebias model, sex expression is determined by the genotype of the zygote itself and only males fertilize outcrossed eggs ($\omega = \infty$ and $Z > 0$). In the absence of males prior to the introduction of genetic variation at the modifier locus ($h_2 = 1$), eggs that are not self-fertilized fail to become zygotes. As a consequence, any allele that induces the development of males ($h_0 > 0$ or $h_1 > 0$) derives an enormous selective advantage from the fertilization of the proportion $(1 - s_A)$ of all eggs produced. Accordingly, we restrict further consideration to cases in which the common genotype produces some males ($h_2 < 1$).

We demonstrate that the sole condition for local stability corresponds to (34):

$$(\hat{h} - h_2)(h_2 - r_L) > 0, \tag{B.1}$$

for $r_L$ the larger root of the bracketed term in (32), viewed as a quadratic in $h_2$:

$$q_{AZ}(h_2) = h_2[d_0 s_A/2 + d_1(1 - s_A)] - d_0 d_1 s_A/2, \tag{B.2}$$

in which

$$\begin{aligned} d_0 &= h_0 - h_2 \\ d_1 &= h_1 - h_2. \end{aligned}$$

These results imply that the proposed ESS (10c) corresponds to an attracting evolutionarily stable strategy under arbitrary dominance levels and intensities of selection on the modifier of sex expression.

## B.1 Linearized recursion system

At the fixation of the $a$ allele, the population comprises only $aa$ individuals ($z_2 = 1$), with normalizer

$$T = h_2.$$

Upon the introduction of the rare alternative allele $A$, genotypes $AA$ and $Aa$ arise in low frequencies ($\delta_0$ and $\delta_1$). Linearization of the full recursion system (23) by ignoring terms of the second order in the $\delta_i$ produces

$$\begin{aligned} \delta_0' &= \frac{s_A(\delta_0 h_0 + \delta_1 h_1/4)}{h_2} \\ \delta_1' &= \frac{[\delta_1 s_A h_1/2 + (1 - s_A)(\delta_0 h_0 + \delta_1 h_1/2)]}{h_2} \\ &\quad + \frac{(1 - s_A)[\delta_0(1 - h_0) + \delta_1(1 - h_1)/2]}{1 - h_2}, \end{aligned}$$

with local stability determined by the dominant eigenvalue of

$$\boldsymbol{M} = \begin{pmatrix} \frac{s_A h_0}{h_2} & \frac{s_A h_1}{4h_2} \\ (1-s_A)\left(\frac{h_0}{h_2} + \frac{1-h_0}{1-h_2}\right) & \left[s_A \frac{h_1}{h_2} + (1-s_A)\left(\frac{h_1}{h_2} + \frac{1-h_1}{1-h_2}\right)\right]/2 \end{pmatrix}. \tag{B.3}$$

Because this matrix is non-negative, its dominant eigenvalue is real and non-negative (Gantmacher 1959, Chapter XIII). Its characteristic polynomial is proportional to

$$C_{AZ}(\lambda) = [(\lambda-1)(1-h_2) - (h_2 - \hat{h})]\left[\lambda - \frac{s_A(h_0 - h_1/2)}{h_2}\right] + (h_2 - \hat{h})\frac{h_1}{h_2}\left[\lambda - \frac{s_A h_0}{2h_2}\right], \tag{B.4}$$

in which the proposed ESS proportion of hermaphrodites at birth corresponds to (10c), with all biparental offspring derived from male parents ($\omega = \infty$):

$$\hat{h} = (1 + s_A)/2. \tag{B.5}$$

Setting the resident hermaphrodite fraction to the proposed ESS ($h_2 = \hat{h}$), we find that $C_{AZ}(\lambda)$ (B.4) reduces to

$$(\lambda - 1)(1 - h_2)\left[\lambda - \frac{s_A(h_0 - h_1/2)}{h_2}\right] = 0,$$

confirming a dominant eigenvalue of unity near the fixation of an allele that induces the candidate ESS, as required for an ESS. Further, we show that ESS is evolutionarily attracting: in a population fixed for an allele that specifies a sex ratio different from the ESS ($h_2 \neq \hat{h}$), only alleles that locally bring the sex ratio closer to the ESS increase when rare (B.1).

A necessary condition for local stability is positivity of the characteristic polynomial $C_{AZ}(\lambda)$ (B.4) evaluated at unity. In addition, we determine the sign of $C_{AZ}(\lambda)$ at two values:

$$\begin{aligned} \lambda_0 &= 1 + \frac{(\hat{h} - h_2)(h_0 - h_2)}{h_2(1-h_2)} \geq 0 \\ \lambda_1 &= 1 + \frac{(\hat{h} - h_2)(h_1 - h_2)}{h_2(1-h_2)} \geq 0. \end{aligned} \tag{B.6a}$$

We find that $C_{AZ}(\lambda)$ changes sign between these values:

$$\begin{aligned} C_{AZ}(\lambda_0) &\propto (\hat{h} - h_2)(h_0 - h_1) \\ C_{AZ}(\lambda_1) &\propto (\hat{h} - h_2)(h_1 - h_0). \end{aligned} \tag{B.6b}$$

## B.2 Special cases

Under random mating ($s_A = 0$), the ESS $\hat{h}$ (B.5) reduces to 1/2 and the dominant eigenvalue of local stability matrix $\boldsymbol{M}$ (B.3) corresponds to

$$\frac{1}{2}\left(\frac{h_1}{h_2} + \frac{1-h_1}{1-h_2}\right).$$

This condition implies that the fixation state is locally stable only if

$$(1/2 - h_2)(h_2 - h_1) > 0,$$

confirming both (B.1) and the classical results of Fisher (1930, Chapter VI): an equal sex ratio at birth corresponds to an attracting ESS under random mating.

Under complete selfing ($s_A = 1$), the ESS $\hat{h}$ (B.5) is equal to unity. Matrix $\boldsymbol{M}$(B.3) is triangular, with the fixation state locally stable to the introduction of the $A$ allele only if

$$(1 - h_2)[h_2 - \max(h_0, h_1/2)] > 0,$$

again confirming (B.1).

Under complete dominance of the rare allele ($h_0 = h_1$), characteristic polynomial (B.4) reduces to

$$[(\lambda - 1)(1 - h_2) - (h_2 - \hat{h})(h_2 - h_0)/h_2][\lambda - s_A h_0/(2h_2)],$$

and the larger ($r_L$) and smaller ($r_S$) roots of (B.2) correspond to

$$\begin{aligned} r_L &= h_0 \\ r_S &= h_0 s_A/2. \end{aligned}$$

Local stability requires that both

$$\begin{aligned} (\hat{h} - h_2)(h_2 - r_L) &> 0 \\ h_2 &> r_S. \end{aligned} \tag{B.7}$$

Because

$$\hat{h} > r_S,$$

the necessary and sufficient condition for local stability under complete dominance corresponds to the first inequality in (B.7), in accordance with (B.1).

## B.3 General dominance and selection intensity

In the remainder of this section, we assume partial inbreeding ($1 > s_A > 0$) and $h_0 \neq h_1$. We first demonstrate that (B.1) implies positivity of the characteristic polynomial (B.4) evaluated at unity for all $h_0$ and $h_1$. We then show that this necessary condition for local stability is in fact sufficient: the (non-negative) dominant eigenvalue of $\boldsymbol{M}$ (B.3) is less than unity under (B.1).

Substitution of $\lambda = 1$ into the characteristic polynomial (B.4) indicates

$$C_{AZ}(1) \propto -(\hat{h} - h_2) q_{AZ}(h_2), \tag{B.8}$$

for $q_{AZ}(h_2)$ given in (B.2). In accordance with our earlier exposition of the full recursion system (23), neutrality ($d_0 = d_1 = 0$) implies that the eigenvalue associated with allele

frequency corresponds to unity, with the frequency of $Aa$ heterozygotes converging to the state corresponding to $F_{neut}$ (24) at rate $s_A/2$.

We now assume that $d_0$ or $d_1$ is non-zero ($h_0 \neq h_2$ or $h_1 \neq h_2$). Because only hermaphrodites produce egg cells, the existence of the population monomorphic for the $a$ allele implies $h_2 > 0$. If the rare allele determines complete male development ($h_0 = 0$ or $h_1 = 0$), then smaller root $r_S = 0$ and

$$C_{AZ}(1) \propto \begin{cases} (\hat{h} - h_2)[h_2 - h_1(1 - s_A/2)] & \text{if } h_0 = 0 \\ (\hat{h} - h_2)(h_2 - h_0 s_A) & \text{if } h_1 = 0. \end{cases} \tag{B.9}$$

If $h_0 = 0$ and $C_{AZ}(1) > 0$, then (B.6) indicates that

$$\begin{aligned} h_2 < \hat{h} &\Rightarrow \lambda_0 < 1 \text{ and } C_{AZ}(\lambda_0) < 0 \\ \hat{h} < h_2 &\Rightarrow \lambda_1 < 1 \text{ and } C_{AZ}(\lambda_1) < 0, \end{aligned}$$

Similarly, under $h_1 = 0$ and $C_{AZ}(1) > 0$,

$$\begin{aligned} h_2 < \hat{h} &\Rightarrow \lambda_1 < 1 \text{ and } C_{AZ}(\lambda_1) < 0 \\ \hat{h} < h_2 &\Rightarrow \lambda_0 < 1 \text{ and } C_{AZ}(\lambda_0) < 0. \end{aligned}$$

These relationships indicate the existence of a root of characteristic polynomial $C_{AZ}(\lambda)$ in $(0, 1)$, which confirms (34) and (B.1): $C_{AZ}(1) > 0$ is both necessary and sufficient for local stability under $h_0 = 0$ or $h_1 = 0$.

Restricting consideration to the remaining case ($h_0, h_1, h_2 > 0$), we find that $q_{AZ}(h_2)$ (B.2) corresponds to a quadratic in $h_2$ with a negative leading term with

$$q_{AZ}(0), q_{AZ}(\max(h_0, h_1)) < 0 < q_{AZ}(\min(h_0, h_1)),$$

which implies that the larger ($r_L$) and smaller ($r_S$) roots of this quadratic lie in

$$\begin{aligned} r_L &\in (\min(h_0, h_1), \max(h_0, h_1)) \\ r_S &\in (0, \min(h_0, h_1)). \end{aligned} \tag{B.10}$$

We first establish that

$$r_S < 1/2 \leq \hat{h} \tag{B.11}$$

for all $h_0$ and $h_1$ in $(0, 1]$. In cases satisfying

$$\min(h_0, h_1) < 1/2,$$

the smaller root $r_S$ (B.10) lies below $1/2$ and consequently $\hat{h}$. If

$$1/2 < \min(h_0, h_1),$$

both $d_0$ and $d_1$ are positive for $h_2 = 1/2$, which implies

$$q_{AZ}(1/2) = [d_1(1 - s_A) + d_0 s_A(1 - h_1)]/2 > 0$$

and confirms (B.11).

For small $h_2$, satisfying

$$h_2 < r_S < \hat{h}, \min(h_0, h_1), \tag{B.12}$$

$C_{AZ}(1) > 0$ (B.8) and both $\lambda_0$ and $\lambda_1$ exceed unity (B.6). That the quadratic characteristic polynomial (B.4) is negative at one of these values ($C_{AZ}(\lambda_1) < 0$ or $C_{AZ}(\lambda_0) < 0$) implies that an eigenvalue in excess of unity exists. We conclude that under (B.12), alleles that increase the proportion of hermaphrodites beyond the level specified by the resident homozygote ($h_2$) increase when rare, confirming (B.1).

We now consider higher hermaphroditic frequencies at the fixation,

$$r_S < h_2, \tag{B.13}$$

under which (B.8) indicates

$$C_{AZ}(1) \propto (\hat{h} - h_2)(h_2 - r_L),$$

the left side of (B.1). Accordingly, (B.1) ($C_{AZ}(1) > 0$) is a necessary condition for local stability. We now demonstrate that it is in fact sufficient for local stability under (B.13). For

$$r_S < h_2 < \hat{h},$$

$C_{AZ}(1) > 0$ implies

$$r_S < \min(h_0, h_1) < r_L < h_2 < \hat{h}.$$

Expressions (B.6) indicate that if $h_1 > h_0$, $\lambda_0 < 1$ and $C_{AZ}(\lambda_0) < 0$ If $h_0 > h_1$, $\lambda_1 < 1$ and $C_{AZ}(\lambda_1) < 0$. We conclude that quadratic characteristic polynomial (B.4) is negative at a value ($\lambda_0$ or $\lambda_1$) less than unity, which implies that that $C_{AZ}(1) > 0$ (B.1) is sufficient for local stability. We now restrict consideration to

$$\hat{h} < h_2,$$

which together with $C_{AZ}(1) > 0$ implies

$$\hat{h} < h_2 < r_L < \max(h_0, h_1).$$

Similar to the preceding case, $h_1 > h_0$ implies $\lambda_1 < 1$ and $C_{AZ}(\lambda_1) < 0$, while $h_0 > h_1$ implies $\lambda_0 < 1$ and $C_{AZ}(\lambda_0) < 0$ (B.6). We again conclude that the quadratic characteristic polynomial (B.4) has a root less than unity, which implies that (B.1) is indeed necessary and sufficient for local stability.

## B.4 Limits of the Li-Price equation

Here, we illustrate that the weak-selection assumption is essential to the heuristically-appealing Li-Price equation (22) and the change of basis that relates it to a full local stability analysis. We provide an example showing that under strong selection on the modifier locus,

the key vector $\tilde{\boldsymbol{z}}$ (A.1) can become invalid and the sign of the characteristic polynomial evaluated at unity insufficient to determine the asymptotic fate of a rare allele introduced into a population monomorphic at the modifier locus.

Local stability matrix $\boldsymbol{M}$ (B.3) represents the linearized transformation with respect to a basis comprising the frequencies of rare genotypes $AA$ ($\delta_0$) and $Aa$ ($\delta_1$). We adopt the new basis described in Appendix A, which comprises the frequency of the rare allele ($A$) and the departure of the heterozygote frequency from the proportion expected under neutrality ($h_0 = h_1 = h_2$):

$$\begin{aligned} t_0 &= \delta_0 + \delta_1/2 \\ t_1 &= \delta_0 - (\delta_0 + \delta_1/2)F_{neut}, \end{aligned} \tag{B.14}$$

for

$$F_{neut} = s_A/(2 - s_A),$$

the fixation index under uniparental fraction $s_A$ (8). Matrix $\boldsymbol{A}$,

$$\boldsymbol{A} = \begin{pmatrix} 1 & 1/2 \\ 1 - F_{neut} & -F_{neut}/2 \end{pmatrix}, \tag{B.15}$$

translates points from the original to the new coordinate system. In the original coordinate system, the key vector (A.1) $\tilde{z}$ corresponds to

$$\boldsymbol{B}\tilde{z} = \begin{pmatrix} \tilde{\delta}_0 \\ \tilde{\delta}_1 \end{pmatrix}, \tag{B.16}$$

for $\boldsymbol{B} = \boldsymbol{A}^{-1}$.

For illustrative purposes, we assume additivity in sex expression,

$$h_1 = (h_0 + h_2)/2, \tag{B.17a}$$

and set

$$\begin{aligned} h_2 &= 1/4 \\ s_A &= 3/4. \end{aligned} \tag{B.17b}$$

Under these assignments, the ESS $\hat{h}$ corresponds to 7/8 and the characteristic polynomial evaluated at unity (B.8) reduces to

$$C_{AZ}(1) \propto (\hat{h} - h_2)(h_2 - h_0)[h_2(2 + s_A) - h_0 s_A].$$

The sole condition for local stability (B.1), which reduces to

$$(\hat{h} - h_2)(h_2 - h_0) > 0,$$

indicates that the fixation of the $a$ allele resists the invasion of the rare $A$ allele only for

$$h_0 < h_2 = 1/4. \tag{B.18}$$

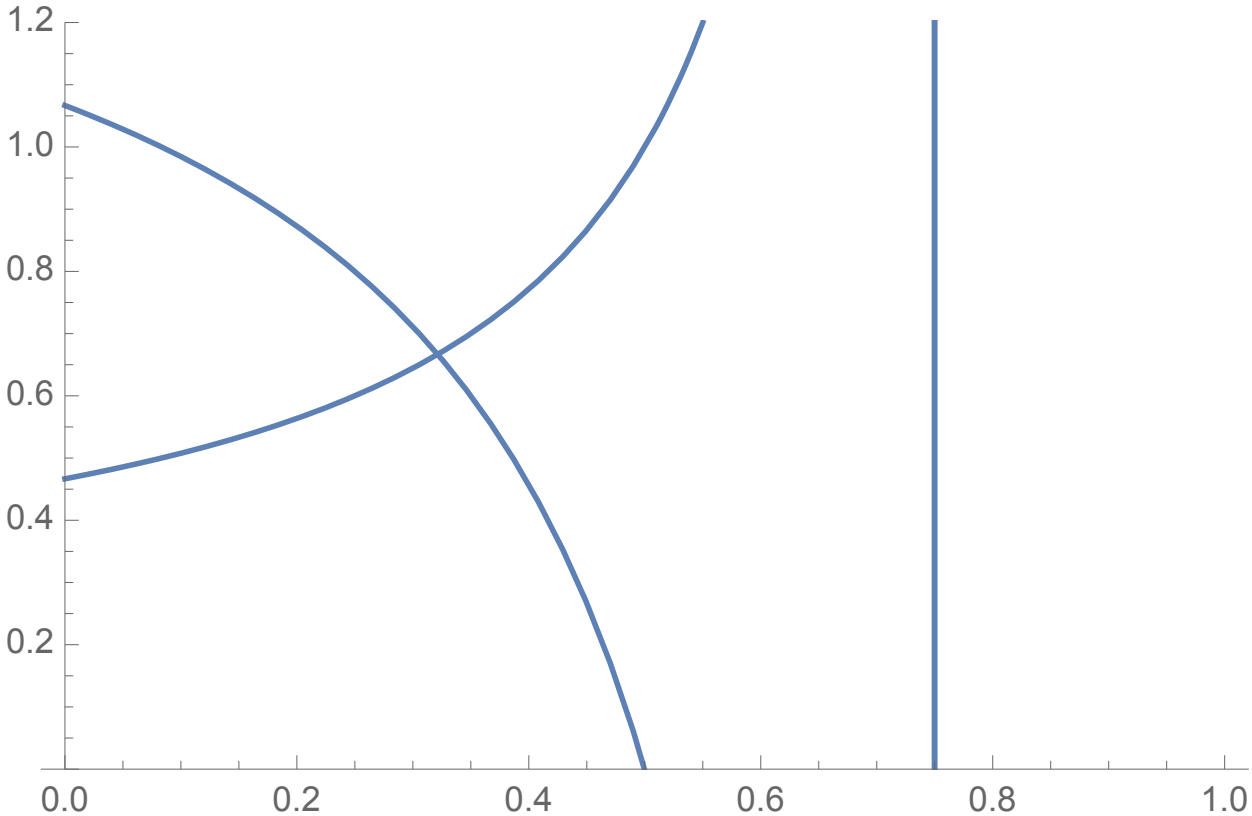


Figure 5: Relative magnitudes of the frequencies of rare homozygotes ($\tilde{\delta}_0$, increasing curve) and heterozygotes ($\tilde{\delta}_1$, declining curve) at the key vector (B.16) as a function of $h_0$, the sex expression parameter associated with the rare homozygote. At the vertical bar ($h_0 = 3/4$), both elements have a discontinuity, which corresponds to the passage through zero of $\mathrm{Det}[(\boldsymbol{I} - \boldsymbol{N})_{n-1}]$ in (A.3).

Indeed, the characteristic polynomial evaluated at unity $C_{AZ}(1)$ is positive in this range and changes sign at $h_0 = 1/4$. However, under intense selection,

$$h_0 > h_2(2 + s_A)/s_A = 11/12, \tag{B.19}$$

$C_{AZ}(1)$ is positive in spite of the local instability of the fixation state.

Key vector (A.1), which connects the local stability criterion to the Li-Price equation (22), remains valid only in the range

$$h_0 < 1/2.$$

Figure 5 plots $\tilde{\delta}_0$ and $\tilde{\delta}_1$, elements of key vector (B.16), as a function of the value of $h_0$. The relative frequency of heterozygotes ($\tilde{\delta}_1$) becomes non-positive for $h_0 \geq 1/2$. In addition, at $h_0 = 3/4$, the principal minor $\mathrm{Det}[(\boldsymbol{I} - \boldsymbol{N})_{n-1}]$ in (A.3) passes through zero, inducing a discontinuity in the key vector (vertical line in Fig. 5).

This simple example illustrates that the connection between the Li-Price equation (22)

and the full local stability analysis holds only for weak selection, which corresponds under (B.17) to the range $0 < h_0 < 1/2$ under additivity of sex expression.