1 The theoretical molecular weight of

2 NaYF₄:RE upconversion nanoparticles

3 Lewis E. Mackenzie, ^[a] Jack A. Goode, ^[a] Alexandre Vakurov, ^[a] Padmaja P. Nampi, ^[b] Sikha Saha, ^[c] Gin

4 Jose,^[b] Paul A. Millner.^[a]

5 [a] School of Biomedical Sciences, Faculty of Biological Sciences, University of Leeds, United Kingdom, LS2 9JT.

6 [b] School of Chemical and Process Engineering, Faculty of Engineering, University of Leeds, United Kingdom, LS2 9JT.

7 [c] Leeds Institute of Cardiovascular and Metabolic Medicine (LICAMM), Faculty of Medicine and Health, University of

8 Leeds, United Kingdom, LS2 9JT.

9 Corresponding author: Lewis MacKenzie L.MacKenzie1@Leeds.ac.uk

10 Abstract

Upconversion nanoparticles (UCNPs) are utilized extensively for biomedical imaging, sensing, and 11 12 therapeutic applications, yet the molecular weight of UCNPs has not previously been reported. We present a theory based upon the crystal structure of UCNPs to estimate the molecular weight of 13 14 UCNPs: enabling insight into UCNP molecular weight for the first time. We estimate the theoretical 15 molecular weight of various UCNPs reported in the literature, predicting that spherical NaYF4 UCNPs ~ 10 nm in diameter will be ~1 MDa (i.e. 10^6 g/mol), whereas UCNPs ~ 45 nm in diameter 16 will be \sim 100 MDa (i.e. 10⁸ g/mol). We also predict that hexagonal crystal phase UCNPs will be of 17 18 greater molecular weight than cubic crystal phase UCNPs. Additionally we find that a Gaussian 19 UCNP diameter distribution will correspond to a lognormal UCNP molecular weight distribution. 20 Our approach could potentially be generalised to predict the molecular weight of other arbitrary 21 crystalline nanoparticles: as such, we provide standalone graphic user interfaces to calculate the 22 molecular weight both UCNPs and arbitrary crystalline nanoparticles. We expect knowledge of 23 UCNP molecular weight to be of wide utility in biomedical applications where reporting UCNP 24 quantity in absolute numbers or molarity will be beneficial for inter-study comparison and 25 repeatability.

26 Introduction

27 Photonic upconversion nanoparticles (UCNPs) have garnered widespread scientific interest due 28 to their unique near infra-red (NIR) excitation and visible luminescence properties; a process 29 known as photonic upconversion. UCNPs are inorganic crystalline nanostructures (typically NaYF₄) co-doped with rare-earth (RE) ions, (e.g. Yb^{3+} , Er^{3+} , Gd^{3+}); hereby referred to in general terms as 30 NaYF₄:RE UCNPs. The RE ions act as sensitizers and emitters for photonic upconversion of multiple 31 infra-red photons, resulting in visible luminescence emission. UCNP emission is highly stable,¹ with 32 no photo bleaching, and a relatively long luminescence emission lifetime ranging from hundreds of 33 microseconds to a few milliseconds.^{2,3} NIR excitation via upconversion is highly advantageous for 34 35 biomedical applications, where ultraviolet or visible excitation of fluorophores (e.g. dyes, proteins, 36 or quantum dots) is normally required, with the associated challenges of photo-bleaching and 37 photo-toxicity. Interactions between nearby molecules and the UCNPs crystal structure enables 38 molecular biosensing via luminescence resonance energy transfer (LRET) between UCNPs and molecules in proximity to them.⁴⁻¹⁰ As such, UCNPs have found wide utility in biomedical 39 applications, including as imaging contrast labels in cellulo, in vivo, and ex vivo^{5,11-19}; as biosensors 40 for detection of antibiotics²⁰ and toxins in food²¹⁻²³; as biosensors to measure biomarkers in 41 biological fluids (e.g. whole blood, serum, urine),^{6-8,24-26} and as therapeutic agents, against targets 42 such as cancer cells.^{27,28} Additionally UCNPs have been applied to nanoscale thermometry^{29,30} and 43 photovoltaic applications.^{31,32} However, to date, the molecular weight of UCNPs has not been 44 45 reported: as such, both the molarity of UCNPs in solution, and the absolute number of UCNPs in a 46 sample has been unknown.

The lack of molecular weight information for UCNPs is a considerable shortcoming in biomedical applications of UCNPs, where precise quantification of UCNP concentration would be highly beneficial for informing of dosage of UCNPs studies, as well as aiding inter-study comparison. Additionally, quantification of UCNP molarity and absolute number of UCNPs would be highly beneficial when constructing biosensors where the ratio of UCNPs compared to other molecules, e.g. antibodies^{6–8,25} or oligonucleotides,³³ is important for informing biosensor design.

The lack of information on UCNP molecular weight is likely due to lack of experimental techniques capable of measuring the molecular weight of large macromolecules such as UCNPs. Using the theory we present in this paper, we predict that the molecular weight of NaYF₄: RE UCNPs will range from a few mega Daltons (MDa) (i.e. 10⁶ g/mol) for exceptionally small UCNPs (~10 nm in diameter), to > 100 MDa (for UCNPs with a more typical diameter of ~45 nm). This large molecular weight range is well beyond the measurement limits both mass spectrometry and sedimentation velocity analytical ultracentrifugation (svUAC), which are limited to < 40 kDa and < 5 MDa respectively.³⁴ Despite this limitation, we attempted to employ svAUC to estimate the molecular weight of UCNPs ~30 nm in diameter (corresponding to a molecular weight of ~40 MDa), but reliable results were not obtained (see the Discussion section and supplementary material).

63 In this study, we present a theoretical method, based upon the extensively studied and 64 empirically proven theory of crystallography and UCNP structure, to calculate the molecular weight UCNPs, accounting for UCNP composition and morphology. In brief, the crystalline structure of 65 66 UCNPs is quantified by transmission electron microscopy (TEM), and x-ray diffraction (XRD) 67 experiments. From this information, the total atomic weight within a single NaYF₄:RE unit cell, and 68 the total number of unit cells within a UCNP can be calculated. Thus, the theoretical molecular 69 weight of UCNPs can be calculated by summing up the total molecular weight contained within all 70 unit cells in a UCNP.

We anticipate that this theoretical framework could be extended to crystalline nanoparticles of arbitrary morphology and composition, provided that the crystalline structure of such nanoparticles are known. As such, we also provide two stand-alone graphical user interfaces (GUIs) for simple calculation of the molecular weight of both NaYF₄:RE UCNPs and arbitrary crystalline nanoparticles. Knowledge of UCNP molecular weight will likely be highly beneficial for quantification of UCNP concentration in biomedical applications.

77 Theory

78 Crystalline structure and photonic upconversion properties of UCNPs

79 The key to understanding both the optical properties of UCNPs and their molecular weight lies in the crystalline structure of UCNPs. UCNPs are a crystal lattice made up of repeating crystal unit 80 cells of NaYF₄, with a fraction of Y^{3+} ions selectively replaced by RE dopants (see Figure 1). In 81 82 UCNPs, photonic upconversion is enabled by the absorption of two or more near-infrared photons, 83 which, via excitation of several long-lived metastable electron states, and subsequent non-radiative 84 multi-phonon and radiative relaxation, produces luminescence emission at visible wavelengths (see 85 Figure 2). Efficient upconversion requires a crystalline host lattice, which is doped with multiple 86 different lanthanide ions (typically Yb^{3+} and Er^{3+}), where one lanthanide ion acts as a photosensitizer (typically Yb³⁺) and acts as a photonic emitter (typically Er^{3+}).³⁵ Although many different 87 combinations of lattice and RE dopants have been explored, ³⁶ the combination of Yb³⁺ and Er³⁺ ions 88 in a NaYF₄ host lattice has been found to provide high upconversion efficiency, and as such is 89 commonly used for UCNPs.^{37,38} Figure 2 shows an exemplar upconversion emission spectrum of 90

91 NaYF₄:Yb,Er cubic UCNPs (20% Yb³⁺, 2% Er^{3+}) and the corresponding Jablonski diagram for 92 upconversion.³⁹

93

94 NaYF₄:RE unit cells are either a cubic or a hexagonal crystal lattice arrangement (see Figure 1). In the face-centred cubic lattice arrangement (Na₂Y₂F₈), high-symmetry cation sites are formed, and 95 are randomly occupied by either Na⁺ or RE³⁺ ions (see Figure 1a), and Y³⁺ ions are substituted for 96 other RE³⁺ ions, enabling photonic upconversion. In hexagonal unit cells (Na₁₅Y₁₅F₆), there are two 97 relatively low-symmetry cation sites, which contain either Na⁺ or RE³⁺ ions (see Figure 1b).⁴⁰ 98 Characterisation of UCNP unit cells is typically conducted by XRD measurements. Several studies 99 100 have reported the crystal lattice parameters associated with cubic and hexagonal NaYF₄:RE UCNPs: these are summarised in Table 1. Wang et al., $(2010)^{40}$ report unit cell parameters for cubic (α 101 102 phase) and hexagonal (β phase) unit NaYF₄:RE unit cell configurations (see Figure 1). The 103 arrangement of ions within unit cells influences the crystal lattice parameters, consequently changing photonic properties, such as upconversion quantum efficiency.⁴⁰ 104

105

106 Synthesis of NaYF₄:RE UCNPs typically creates pseudo-spherical UCNPs with a range of 107 diameters. For example, Sikora et al., (2013) report a Gaussian diameter distribution of UCNPs, 108 ranging between 15 – 70 nm.¹⁴ and Haro-González et al. (2013)⁴¹ report a Gaussian diameter 109 distribution of UCNPs UCNPs ranging from ~10 – 50 nm in diameter (see Table 1).

- 110
- 111

Table 1. Crystal lattice parameters of NaYF₄:RE UCNPs reported in the literature.

Study	NaYF₄ RE dopant composition (%)	UCNP lattice structure	a (Å)	c (Å)	Mean UCNP diameter (nm)	UCNP diameter range (nm)
Sikora et al., (2013). ¹⁴	30% Yb ³⁺ , 2% Er ³⁺	Cubic	5.51	-	~ 30	15 -70
Cao et al., 2010. ¹⁵	20% Yb ³⁺ , 2% Er ³⁺	Hexagonal	5.960	3.510	33 ± 1 nm	32 - 34 nm
Wang et al., 2010. ⁴⁰	18% Yb ³⁺ , 2% Er ³⁺	Hexagonal	5.96	3.53	Not reported	Not reported
Wang et al., 2010. ⁴⁰	18% Yb ³⁺ , 2% Er ³⁺ , 60% Gd ³⁺	Hexagonal	6.02	3.60	Not reported	Not reported

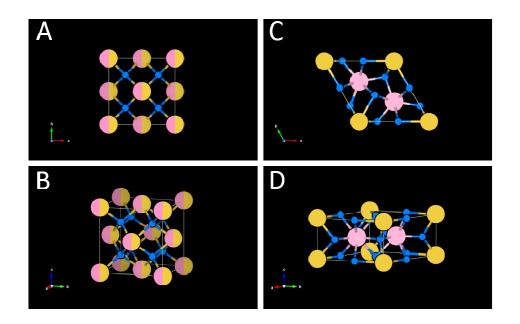
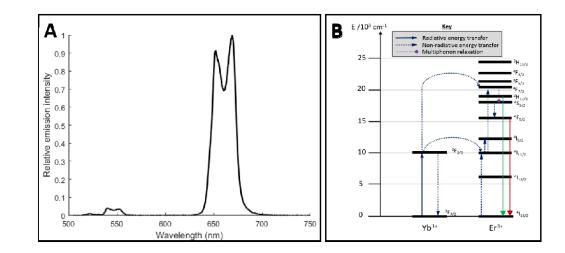


Figure 1. NaYF4: RE UCNPs unit cell structures. Colour key: Na⁺ ions are yellow; Y³⁺ and 114 RE³⁺ dopant ions are pink; F⁻ ions are smaller and blue. (A, B) Cubic lattice unit cell 115 116 structure. Sites that are randomly occupied by both Na⁺ and RE³⁺ are depicted as both pink and yellow. (C, D) Hexagonal lattice unit cell structure. This Figure is based upon 117 data from Kramer et al., (2004)⁴² and Wang et al., (2010).⁴⁰ Diagrams created with the 118 open-source software package VESTA.43 119





124

121 122 123

Figure 2. The upconversion emission of UCNPs. (A) Emission of a 1 mg/mL of NaYF₄:Yb,Er UCNPs (20% Yb and 2% Er) suspended in ultra-pure water (see supplementary material for UCNP synthesis details). (B) Corresponding Jablonski diagram depicting the upconversion process (based upon Heer et al., (2004)).³⁹

125 Estimating the number of unit cells in a UCNP

126 For the purposes of this study, we assume UCNPs to be spherical, with volume (V_{UCNP})

127 described by:

$$V_{UCNP} = \frac{4}{3} \pi r^3,$$
 (1)

where r is the radius of the UCNP. Note that non-spherical UCNP morphologies can be incorporated by modifying Equation 1 appropriately. If the UCNP consists of cubic unit cells, then the volume of an individual cubic unit cell (uV_{cubic}) is given by:

$$uV_{cubic} = a_c^3. \tag{2}$$

131 If the UCNP consists of hexagonal unit cells, volume of a hexagonal unit cell $(uV_{hexagonal})$ is given 132 by:

$$uV_{hexagonal} = \frac{2\sqrt{3}}{4} a_h^2 c_h. \tag{3}$$

133 Where a_h and c_h are lattice parameters describing hexagonal unit cells. Thus, the number of unit 134 cells in a UCNP (i.e. uN_{cubic} or $uN_{hexagonal}$) can be estimated by:

$$uN_{cubic} = V_{UCNP}/uV_{cubic},\tag{4}$$

$$uN_{hexagonal} = V_{UCNP} / uV_{hexagonal}.$$
(5)

135 This calculation assumes the effects of crystal dislocations and rounding error in the total number

of unit cells to be negligible. Further, we assume that UCNPs are composed of 100% cubic or

137 hexagonal unit cells because, to the best of our knowledge, hybrid crystal phase UCNPs have not

138 been reported.

139

140 Estimating the total atomic weight within a single unit cell

Assuming no RE dopants, the atomic weight of a single cubic NaYF₄ (uAW_{cubic}) or hexagonal

142 NaYF₄ unit cell (uW_{hex}) is described by:

$$uAW_{cubic} = (2 \times AW_{Na}) + (2 \times AW_{Y}) + (8 \times AW_{F});$$
(6)

$$uAW_{hex} = (1.5 \times AW_{Na}) + (1.5 \times AW_{Y}) + (6 \times AW_{F});$$
⁽⁷⁾

where AW_{Na} , AW_{Y} , and AW_{F} are the atomic weight (Da or g/mol) of Sodium, Yttrium, and Fluorine respectively (see Table S1). We assume any mass difference due loss of electrons due to ionisation to be negligible. If RE dopant ions are added during UCNP synthesis, then a fraction of Y³⁺ ions are substituted for RE³⁺ dopant ions, altering the average atomic weight of unit cells within UCNPs. This RE doping can be accounted for by defining a total additive factor (*af*):

$$af = fRE_{d1} + fRE_{d2} \dots + fRE_{dn},\tag{8}$$

- 149 where RE_{d1} , fRE_{d2} , ... fRE_{dn} is the fractional percentage of an arbitrary number of RE dopants.
- 150 The total additive factor is a numeric value ranging between 0 and 1, representing the theoretical
- extremes of 0% and 100% Y substitution respectively. Thus, total the atomic mass contained within
- a single cubic or hexagonal unit cell with RE dopants is be calculated by:
- 153

$$uAW_{cubic} RE_{Doped} = (2 \times AW_{Na}) + (8 \times AW_{F}) + (2(1 - af) \times AW_{Y}) +$$

$$(2 \times fRE_{d1} \times AW_{RE_{d1}}) + (2 \times fRE_{d2} \times AW_{RE_{d2}}) + \dots + (2 \times fRE_{dn} \times AW_{RE_{dn}});$$
(9)

$$uAW_{hexagonal}RE_{Doped} = (1.5 \times AW_{Na}) + (6 \times AW_F) + (1.5(1 - af) \times (10))$$
$$W) + (1.5 \times fRE_{d1} \times AW_{RE_{d1}}) + (1.5 \times fRE_{d2} \times AW_{RE_{d2}}) + \dots + (1.5 \times fRE_{dn} \times AW_{RE_{dn}});$$

where $uAW_{cubic} RE_{Doped}$ and $uAW_{hexagonal}RE_{Doped}$ are the average atomic weight of RE doped cubic and hexagonal unit cells, respectively.

156

157 Estimating the theoretical molecular weight of a UCNP

Once the total number of unit cells within a UCNP (uN) and the total atomic weight (uAW)within each individual unit cell are estimated, the theoretical molecular weight of a cubic lattice UCNP (MWcubic) can be estimated by summing the atomic weight contributions from all unit cells:

$$MW_{cubic} = uAW_{cubic} RE_{Doped} \times uN_{cubic}, \tag{11}$$

$$MW_{hexagonal} = uAW_{hexagonal} RE_{Doped} \times uN_{hexagonal} .$$
(12)

From Equations 4, 5, 11, and 12, it can be seen that the molecular weight of UCNPs scales proportionally to volume, thus spherical UCNPs molecular weight will scale proportionally to the cube of UCNP radius.

165

167 Methods

168 Molecular weight predictions for cubic and hexagonal NaYF₄:RE UCNPs

- 169 Using the theory presented in Sections 2.4 2.6, the theoretical molecular weight of hexagonal
- and cubic lattice NaYF₄ UCNPs were calculated, assuming the following typical unit cell lattice
- 171 parameters: cubic: a = 5.51 Å; hexagonal: a = 5.91 Å, c = 3.53 Å; (see Table 1).
- 172

173 The effect of RE doping on theoretical molecular weight

- The effect of RE doping was investigated by using the theory presented in Sections 2.4 2.6 to calculate the theoretical molecular weight of NaYF₄:RE UCNPs incorporating various concentrations of Yb³⁺ and Er^{3+} dopant ions. We assume that UCNP lattice parameters will remain constant, neglecting the unit cell contraction effect demonstrated by Wang et al. (2010),⁴⁰ where UCNP unit cell lattice parameters are altered when the concentration of RE dopants is increased.⁴⁰
- 179

180 The theoretical molecular weight of UCNPs reported in the literature

- 181 The theoretical molecular weight of various $NaYF_4$: RE UCNPs reported in the literature was 182 calculated by incorporating various lattice parameters and mol% of RE dopants from the literature 183 into the theory presented in Sections 2.4 – 2.6.
- 184

185 UCNP diameter distribution vs. theoretical molecular weight distribution

UCNP synthesis typically produces a Gaussian distribution of UCNPs diameters. To investigate how such a distribution of UCNP diameters affects the distribution of theoretical UCNP molecular weights, the Gaussian diameter distribution data for a single batch of NaYF₄:Yb,Er UCNPs was reproduced from data presented in Sikora et al (2013).¹⁴ The theoretical molecular weight for each UCNP diameter in this distribution was calculated by the theory presented in Sections 2.4 – 2.6. Gaussian fits to the data were calculated by using non-linear least squares fitting in MATLAB (MATLAB 2016a, MathWorks).

193

194 Stand-alone GUIs for calculation of nanoparticle theoretical molecular weight

Two stand-alone executable graphic user interfaces (GUIs) were created in MATLAB to enable rapid calculation of UCNP theoretical molecular weight. Each GUI incorporates different features and assumptions. The first, more simple, GUI was developed to enable other researchers to calculate the theoretical molecular weight of spherical NaYF₄:RE UCNPs for a user-defined nanoparticle size range. The second, more powerful, GUI was designed to enable users to estimate the theoretical molecular weight of crystalline nanoparticles with arbitrary nanoparticle geometry;

arbitrary lattice parameters; and arbitrary elemental composition, across a user-defined range of characteristic nanoparticle sizes. Additional technical information for both GUIs is provided in the supplementary material section. The stand-alone GUIs developed are shown in supplemental Figures S1 and S2. These GUIs are freely available from the University of Leeds Research Data Depository and are attributed with their own citable DOI (<u>https://doi.org/10.5518/173</u>).⁴⁴

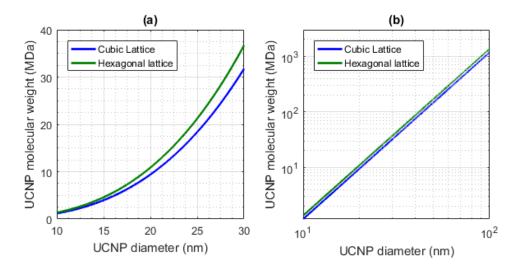
206

207 **Results**

208 Theoretical molecular weight of cubic and hexagonal NaYF4: RE UCNPs

Hexagonal lattice UCNPs have a greater theoretical molecular weight than cubic lattice UCNPs (see Figure 3); this is due to the lower volume of hexagonal unit cells, and correspondingly higher density of hexagonal lattice UCNPs. Additionally, because molecular weight scales to UCNP volume, relatively small changes in UCNP diameter increased molecular weight considerably: e.g. a 20 nm cubic UCNP has a molecular weight of ~10 MDa, whereas a 30 nm UCNP has a molecular weight of > 30 MDa (an increase of 20 MDa for a 5 nm change in UCNP diameter).





216 217

218

219 220

Figure 3. Diameter versus theoretical molecular weight for hexagonal and cubic NaYF₄ UCNPs (green and blue respectively). (a) UCNP diameter vs. molecular weight on a standard x-axis. (b) The same data plotted with a logarithmic scale. Lattice parameters were assumed to be: a = 5.51 Å for cubic UCNPs; a, c = 5.91 Å and 3.53 Å for hexagonal UCNPs.

223

224 The effect of RE doping on UCNP molecular weight

Increasing Yb³⁺ or Er³⁺ dopant % increased the theoretical molecular weight of UCNPs (see Figure 4) because Yb³⁺ and Er³⁺ have a greater atomic mass than Y³⁺. However, the difference in theoretical molecular weight between UCNPs doped with Yb³⁺ and Er³⁺ was relatively small to the small difference between the atomic weight of Yb³⁺ and Er³⁺ (173.054 and 167.259 g/mol respectively, see Table S1). Hexagonal lattice UCNPs show a slightly higher increase in theoretical molecular weight for a given dopant concentration than cubic lattice UCNPs because hexagonal lattice UCNPs have a greater unit cell density compared to their cubic counterparts.

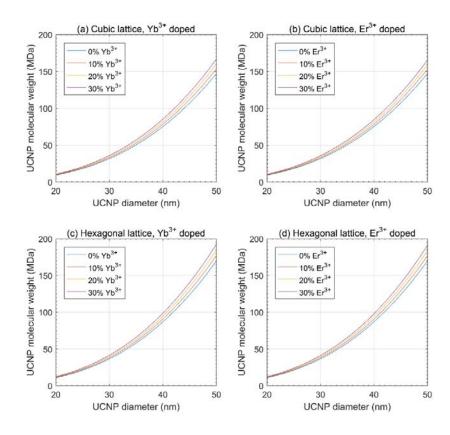




Figure 4. The effect of RE doping on theoretical UCNP molecular weight. (a, b)

234theoretical molecular weight vs. RE dopant mol% for cubic lattice UCNPs. (c, d)235theoretical molecular weight vs. RE dopant mol% for cubic lattice UCNPs.236Calculations assume that lattice parameters are a = 5.51 Å for cubic lattice UCNPs,237a = 5.91 Å; c = 3.53 Å for hexagonal lattice UCNPs, and that lattice parameters are238independent of dopant mol%.

239

241 The theoretical molecular weight of NaYF₄:RE UCNPs reported in the literature

242 The theoretical molecular weight of various NaYF₄:RE UCNPs reported in the literature are

shown in Figure 5.

244

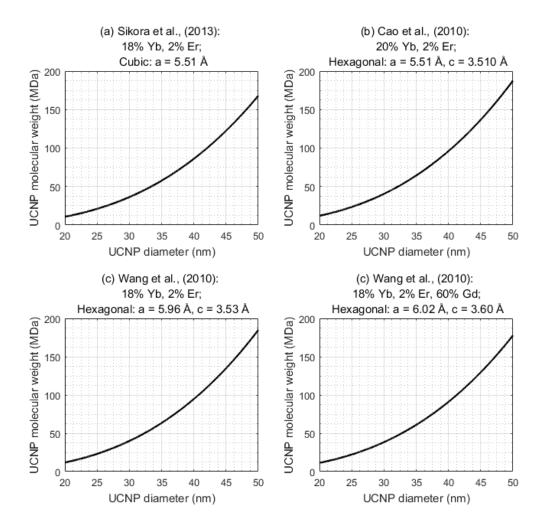


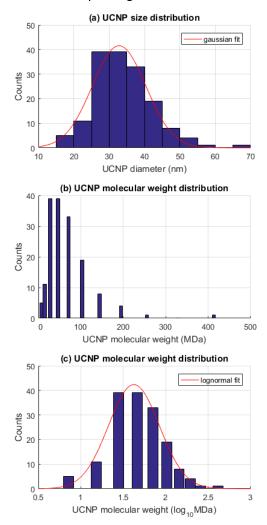


Figure 5. Theoretical molecular weight of various UCNPs reported in the literature. (a) Sikora et al., (2013).¹⁴ (b) Cao et al., (2010).¹⁵ (c, d) Wang et al., (2010).⁴⁰

250 UCNP diameter distribution vs. theoretical molecular weight distribution

251 The UCNP diameter distribution data from Sikora et al., (2013)¹⁴ was well-fitted by a Gaussian

- distribution ($R^2 = 0.96$) (see Figure 6a). The corresponding theoretical molecular weight distribution
- 253 (shown in Figure 6b), demonstrates the exponential relation between UCNP diameter and UCNP
- 254 molecular weight distribution. Plotted on a logarithmic x-axis scale (Figure 6c), the resulting
- molecular weight distribution was well fitted by a Gaussian distribution ($R^2 = 0.98$), indicating that
- the molecular weight distribution corresponding to a Gaussian diameter distribution is lognormal.



257

Figure 6. Gaussian UCNP diameter distributions give arise to lognormal distribution of theoretical molecular weights. (a) A Gaussian diameter distribution of UCNPs is well described by a normal distribution ($R^2 = 0.96$). (b) The corresponding theoretical molecular weight distribution of UCNPs on a linear molecular weight scale. (c) The molecular weight distribution on a logarithmic x-axis is well fitted by a lognormal distribution ($R^2 = 0.98$).

264 **Discussion**

We have provided a theory to estimate the molecular weight of UCNPs. Our theory is required because, to the best of our knowledge, there are no experimental techniques measuring the molecular weight of UCNPs, which we predict will be > 5 MDa for UCNPs ~15 nm in diameter, and 100 MDa for UCNPs ~45 nm in diameter. Mass spectrometry is limited to molecules < 40 kDa, and svAUC is limited to measurements of macromolecules < 5 MDa.³⁴ Our theory predicts that UCNPs with a molecular weight of < 5 MDa would be < 15 nm in diameter. To the best of our knowledge monodisperse synthesis of such small UCNPs has not been reported in the literature.

272

273 Despite the aforementioned challenges of experimental verification, we attempted svAUC 274 measurements of UCNPs, because successful svAUC studies of other types of nanoparticles (e.g. SiO₂ nanoparticles) with unknown molecular weight have been reported.^{45,46} If accurate svAUC 275 276 measurements of UCNPs could be made, then UCNP molecular weight could potentially be 277 calculated by the theory described by Carney et al., (2011),³⁴ which is based upon accurate 278 quantification of sedimentation and diffusion coefficients from svAUC measurements, and which 279 has been verified for gold nanoparticles ~2 MDa in molecular weight. The full details of the method 280 of our svAUC experiment are provided in the supplementary information. However, our svAUC 281 experiment studying UCNPs was not successful. In brief, our avAUC results showed that the UCNPs 282 (diameter = 32 ± 5 nm, average theoretical molecular weight of ~ 43 MDa) sedimented very rapidly, 283 even at low centrifuge rotor speeds (3,000 rpm), limiting the amount of useable data. At higher 284 rotor speeds UCNPs sedimented too rapidly for data collection. When the recovered sedimentation 285 coefficient was extrapolated to zero sample concentration, a negative sedimentation coefficient 286 was returned. Additionally, UCNPs were observed to diffuse considerably, further complicating AUC 287 experiments. This unusual behaviour is not typical of the nanoclusters and gold nanoparticles used to demonstrated the molecular weight estimation technique described by Carney et al., (2011),³⁴ 288 289 and as such UCNP molecular weight could not be estimated by svAUC. The challenges associated 290 with svAUC measurement of UCNPs serve to further highlight the need for a method to estimate 291 the molecular weight of UCNPs theoretically.

292

Although it has not been possible for us to experimentally validate our estimates of UCNP molecular weight, it may be possible in future to verify some limited predictions of our theory. For example, it may be possible to measure the difference in bulk densities of cubic and hexagonal UCNPs and compare this with predictions from our theory. However, we could not attempt this

297 measurement because we did not have access to the high temperature crucible equipment
 298 required for hexagonal UCNP synthesis.⁴⁰

299

Despite this lack of current and direct experimental verification, we can be reasonably confident in the accuracy of our theory because it stems directly from the theory of crystallography, which has been a subject of intense study in the past century,⁴⁷ combined with empirical measurements of UCNP crystal structure.

304

305 Our method to calculate the theoretical molecular weight of NaYF₄:RE UCNPs relies on two basic 306 assumptions: 1. that UCNPs are crystals of homogenous elemental composition and unit cell phase, 307 and 2. that the lattice parameters and diameter data utilized is accurate. These assumptions can be 308 verified by TEM and XRD measurements of UCNP crystal structure. Ensuring accurate lattice 309 parameters is particularly important when estimating the molecular weight of UCNPs with arbitrarily large dopant concentrations. For example, Wang et al., (2010)⁴⁰ experimentally 310 demonstrated that by doping a hexagonal phase NaYF₄:Yb,Er UCNP (18% Yb, 2% Er) with increasing 311 concentrations of Gd³⁺ increases the lattice parameters of the UCNP significantly, resulting in an 312 313 increased unit cell volume. Thus, because of this dependence of lattice parameter on RE dopant 314 percentage, our estimations of UCNP molecular weight in Figure 3 may be an over-estimation on 315 true values if lattice parameters are not independently verified for each RE dopant concentration 316 of interest. UCNP volume/morphology also influences theoretical UCNP molecular weight. We 317 recommend using TEM to directly quantify UCNP morphology with limited assumptions. Other 318 techniques such as such as dynamic light scattering (DLS) and nanoparticle tracking analysis can be 319 used to estimate the equivalent hydrodynamic radius of nanoparticles but incorporate various 320 assumptions into calculations.^{46,48} As such, direct TEM imaging of UCNPs is preferable to ensure 321 theoretical molecular weight is as accurate as possible. In this study we assumed UCNPs are 322 perfectly spherical, but our method could be trivially adapted for arbitrary nanoparticle 323 geometries; e.g. rods, 40,49 triangular, 50 or prism-shaped 51 nanoparticles, and for nanoparticles of 324 varying crystalline composition. The extension of our technique to arbitrary geometries, arbitrary 325 crystal lattice parameters, and arbitrary elemental composition is demonstrated by the development and application of an advanced GUI incorporating all of these variables (see Figure 326 327 S2). Our theory does not account for any dislocations in the regular UCNP crystal structure. Instead 328 we assume the influence of any such dislocations to be negligible compared to the molecular 329 weight of whole UCNPs. Our theory also does not account for any surface functionalisation with 330 amorphous layers or other molecules. Thus the molecular weight of UCNPs modified by addition of

a silica^{8,35,52} or calcium fluoride⁵³ shell coating will be greater than the theoretical molecular weight
 estimated by our technique.

333

334 It should be noted that a simple theory for estimation of the molecular weight of a single 335 homogenous gold nanoparticle based upon bulk density of materials was proposed by Lewis et al. (2006).⁵⁴ However, this simple theory did not account for crystalline unit cell parameters or 336 337 elemental doping. Further, their theory was not extended to describe the molecular weight 338 distributions of a population of nanoparticles. Our results demonstrate that a Gaussian distribution 339 of UCNP diameters corresponds to a lognormal distribution in molecular weight (as shown in Figure 340 6). Mathematically, it is reasonable to expect similar logarithmic relations between UCNP diameter 341 and molecular weight for arbitrary diameter distributions. Such molecular weight distributions may 342 of consequence when studying behaviour of UCNP populations, because minor outliers in UCNP 343 diameter will be extreme outliers in terms of molecular weight.

344

345 Estimation of molecular weight of NaYF₄:RE UCNPs will likely be of utility in various applications, 346 particularly in biomedical imaging, biosensing, and therapeutics. Knowledge of UCNP molecular weight will likely be of great utility in studies where UCNP surfaces are functionalised with 347 additional molecules, e.g. antibodies ^{6-8,25} or oligonucleotides, ³³ because If the molecular weight of 348 349 UCNPs is known, then the molar concentrations of substances in the functionalisation processes 350 can be determined. When combined with estimation of UCNP surface area, this could inform the 351 UCNP functionalisation for biosensing applications. Knowledge of UCNP molecular weight would 352 also be beneficial in the processing of particles for downstream applications. In particular, steps 353 taken to functionalise the nanoparticles may require separation procedures to remove unreacted 354 moieties or unwanted reactants. If the molecular weight of UCNPs were known, then it may be 355 beneficial for the optimisation of conjugation stoichiometry, which can be concentration 356 dependant; the reaction rates of UCNPs will be heavily influenced by their molecular weight; thus a 357 greater understanding of their molecular weight may increase the knowledge of thermodynamic 358 properties of UCNP systems. This is particularly important when considering the use of bio-359 receptors with UCNPs where the mass of the particle may affect the binding kinetics of the UCNP-360 receptor construct.

The molecular weight of UCNPs will also be of interest in the study of cytotoxicity, biodistribution, cellular uptake, metabolism, and excretion of UCNPs in biological systems.^{12,14} Currently, it is extremely challenging to compare the results from various imaging and therapeutic studies because UCNP concentration is reported as weight of UCNPs per volume of aqueous media

(i.e. mg/mL or similar).¹² This is a crude measure which does not quantify number of UCNPs in a 365 given sample. For example, nanoparticles can induce membrane damage⁵⁵ and initiate apoptosis 366 (programmed cell-death).^{56,57} Reporting the molar concentration of UCNPs would help assessment 367 368 of UCNP cytotoxic effects. A standardised protocol based on molecular weight of UCNPs would help assessment of accumulation of UCNPs *in vivo* and their clearance time from organs¹³ or tumours.⁵³ 369 Reporting the molar concentration of UCNP composites may also help to develop highly-localised 370 371 targeted delivery of therapeutic drugs to the required sites in the body, leading to better controlled targeted photodynamic therapy,²⁷ and potential improvements in targeted drug delivery.¹⁶ 372

373 **Conclusions**

374 We have provided a method to estimate the theoretical molecular weight of UCNPs. This theory 375 is based upon UCNP crystal parameters which can be measured for batches of UCNPs by TEM and 376 XRD techniques. The theory presented here is generalizable to other crystalline nanoparticles 377 where the relevant crystalline lattice parameters are known, i.e. nanoparticle unit cell elemental 378 composition, unit cell size parameters, and nanoparticle morphology. To enhance dissemination of 379 our theory we provide two stand-alone GUIs for calculation of the molecular weight of both UCNPs 380 and arbitrary crystalline nanoparticles respectively. We could not, however, experimentally verify 381 our predictions of UCNP molecular weight with mass spectrometry or svAUC. We did attempt 382 svAUC experiments but could not recover reliable svAUC data because UCNPs were observed to 383 sediment and diffuse rapidly. Nevertheless, our theory provides some key predications about the 384 molecular weight of UCNPs. Firstly, that the theoretical molecular weight of UCNPs scales with 385 volume of the nanoparticle. As an example, we predict that a spherical UCNP ~10 nm diameter will have a molecular weight of ~1 MDa (10^6 g/mol), whereas a UCNP ~ 45 nm in diameter will be ~100 386 MDa (10^8 g/mol) . From this relation, we find that a Gaussian distribution of nanoparticle diameters 387 388 corresponds to a lognormal distribution of UCNPs molecular weights, and that a small change in 389 UCNP diameter distribution can potentially represent a large change in overall UCNP molecular 390 weight. We also report that Hexagonal crystal lattice phase UCNPs will be of greater molecular 391 weight than cubic lattice phase UCNPs, and that increasing RE dopant % will increase UCNP 392 molecular weight.

We expect that the knowledge of UCNP molecular weight will be of utility in a wide variety of biomedical applications, as UCNP concentrations can now be reported in terms of molarity or absolute number of UCNPs instead of the relatively crude measure of sample weight. This will likely aid inter-study comparison of both UCNP dosage and improve methods for creating UCNP biosensors.

398

399 **References**

400	1.	Zhou, J., Xu, S., Zhang, J. & Qiu, J. Upconversion luminescence behavior of single
401		nanoparticles. <i>Nanoscale</i> 7, (2015).
402	2.	Hyppänen, I., Höysniemi, N., Arppe, R., Schaeferling, M. & Soukka, T. Environmental Impact
403		on the Excitation Path of the Red Upconversion Emission of Nanocrystalline NaYF $_4$ D:Yb $^{3+}$,Er
404		³⁺ . J. Phys. Chem. C acs.jpcc.7b01019 (2017). doi:10.1021/acs.jpcc.7b01019
405	3.	Plohl, O. et al. Optically Detected Degradation of NaYF4: Yb, Tm Based Upconversion
406		Nanoparticles in Phosphate Buffered Saline Solution. Langmuir acs.langmuir.6b03907
407		(2016). doi:10.1021/acs.langmuir.6b03907
408	4.	Chen, F., Bu, W., Cai, W. & Shi, J. Functionalized upconversion nanoparticles: versatile
409		nanoplatforms for translational research. Curr. Mol. Med. 13, 1613–32 (2013).
410	5.	Wang, F., Banerjee, D., Liu, Y., Chen, X. & Liu, X. Upconversion nanoparticles in biological
411		labeling, imaging, and therapy. Analyst 135, 1839–1854 (2010).
412	6.	Tang, J., Lei, L., Feng, H., Zhang, H. & Han, Y. Preparation of K+-Doped Core-Shell NaYF4:Yb,
413		Er Upconversion Nanoparticles and its Application for Fluorescence
414		Immunochromatographic Assay of Human Procalcitonin. J. Fluoresc. 26, 2237–2246 (2016).
415	7.	Lei, L. et al. A rapid and user-friendly assay to detect the Neutrophil gelatinase-associated
416		lipocalin (NGAL) using up-converting nanoparticles. <i>Talanta</i> 162, 339–344 (2017).
417	8.	Jo, E. J., Mun, H. & Kim, M. G. Homogeneous Immunosensor Based on Luminescence
418		Resonance Energy Transfer for Glycated Hemoglobin Detection Using Upconversion
419		Nanoparticles. <i>Anal. Chem.</i> 88, 2742–2746 (2016).
420	9.	Doughan, S., Uddayasankar, U. & Krull, U. J. A paper-based resonance energy transfer
421		nucleic acid hybridization assay using upconversion nanoparticles as donors and quantum
422		dots as acceptors. Analytica Chimica Acta 878, (Elsevier B.V., 2015).
423	10.	Zhang, S. et al. Fluorescence resonance energy transfer between NaYF4:Yb,Tm upconversion
424		nanoparticles and gold nanorods: Near-infrared responsive biosensor for streptavidin. J.
425		Lumin. 147, 278–283 (2014).
426	11.	Mader, H. S., Kele, P., Saleh, S. M. & Wolfbeis, O. S. Upconverting luminescent nanoparticles
427		for use in bioconjugation and bioimaging. <i>Curr. Opin. Chem. Biol.</i> 14, 582–596 (2010).
428	12.	Gnach, A., Lipinski, T., Bednarkiewicz, A., Rybka, J. & Capobianco, J. a. Upconverting
429		nanoparticles: assessing the toxicity. <i>Chem. Soc. Rev. Chem. Soc. Rev</i> 44, 1561–1584 (2015).
430	13.	Zou, R. et al. Silica shell-assisted synthetic route for mono-disperse persistent
431		nanophosphors with enhanced in vivo recharged near-infrared persistent luminescence.

432		Nano Res. (2017). doi:10.1007/s12274-016-1396-z
433	14.	Sikora, B. <i>et al.</i> Transport of NaYF4:Er3+, Yb3+ up-converting nanoparticles into HeLa cells.
434		Nanotechnology 24, 235702 (2013).
435	15.	Cao, T. <i>et al.</i> Water-soluble NaYF4:Yb/Er upconversion nanophosphors: Synthesis,
436		characteristics and application in bioimaging. Inorg. Chem. Commun. 13, 392–394 (2010).
437	16.	Ma, Y. et al. Labeling and long-term tracking of bone marrow mesenchymal stem cells in
438		vitro using NaYF4:Yb3+,Er3+ upconversion nanoparticles. Acta Biomater. 42, 199–208
439		(2016).
440	17.	Kostiv, U. <i>et al.</i> RGDS- and TAT-Conjugated Upconversion of NaYF4:Yb3+/Er3+&SiO2
441		Nanoparticles: In Vitro Human Epithelioid Cervix Carcinoma Cellular Uptake, Imaging, and
442		Targeting. ACS Appl. Mater. Interfaces 8, 20422–20431 (2016).
443	18.	Shi, Y. et al. Stable Upconversion Nanohybrid Particles for Specific Prostate Cancer Cell
444		Immunodetection. <i>Nat. Publ. Gr.</i> 1–11 (2016). doi:10.1038/srep37533
445	19.	Rao, L. et al. Erythrocyte Membrane-Coated Upconversion Nanoparticles with Minimal
446		Protein Adsorption for Enhanced Tumor Imaging. Appl. Mater. Interfaces 2159–2168 (2017).
447		doi:10.1021/acsami.6b14450
448	20.	Hu, G. et al. Upconversion Nanoparticles and Monodispersed Magnetic Polystyrene
449		Microsphere Based Fluorescence Immunoassay for the Detection of Sulfaquinoxaline in
450		Animal-Derived Foods. J. Agric. Food Chem. 64, 3908–3915 (2016).
451	21.	Dai, S., Wu, S., Duan, N. & Wang, Z. A luminescence resonance energy transfer based
452		aptasensor for the mycotoxin Ochratoxin A using upconversion nanoparticles and gold
453		nanorods. <i>Microchim. Acta</i> 183, 1909–1916 (2016).
454	22.	Guo, X., Wu, S., Duan, N. & Wang, Z. Mn 2 + -doped NaYF 4🛛: Yb / Er upconversion
455		nanoparticle-based electrochemiluminescent aptasensor for bisphenol A. Anal. Bioanal.
456		Chem. 408, 3823–3831 (2016).
457	23.	Chen, Q., Hu, W., Sun, C., Li, H. & Ouyang, Q. Synthesis of improved upconversion
458		nanoparticles as ultrasensitive fluorescence probe for mycotoxins. Anal. Chim. Acta 938,
459		137–145 (2016).
460	24.	Fu, X., Chen, L. & Choo, J. Optical Nanoprobes for Ultrasensitive Immunoassay. Anal. Chem.
461		1, 124–137 (2016).
462	25.	Gao, N., Ling, B., Gao, Z., Wang, L. & Chen, H. Near-infrared-emitting NaYF4:Yb,Tm/Mn
463		upconverting nanoparticle/gold nanorod electrochemiluminescence resonance energy
464		transfer system for sensitive prostate-specific antigen detection. Anal. Bioanal. Chem.
465		(2017). doi:10.1007/s00216-017-0212-2

466	26.	Juntunen, E. <i>et al.</i> Effects of blood sample anticoagulants on lateral flow assays using
467		luminescent photon-upconverting and Eu(III) nanoparticle reporters. Anal. Biochem. 492,
468		13–20 (2016).
469	27.	Liang, L. et al. Facile Assembly of Functional Upconversion Nanoparticles for Targeted
470		Cancer Imaging and Photodynamic Therapy. ACS Appl. Mater. Interfaces 8, acsami.6b00713
471		(2016).
472	28.	Yang, X. et al. Synthesis of a core/satellite-like multifunctional nanocarrier for pH- and NIR-
473		triggered intracellular chemothermal therapy and tumor imaging. RSC Adv. 7, 7742–7752
474		(2017).
475	29.	Geitenbeek, R. G. <i>et al.</i> NaYF $_4$: Er $^{3+}$,Yb $^{3+}$ /SiO $_2$ Core/Shell Upconverting Nanocrystals for
476		Luminescence Thermometry up to 900 K. J. Phys. Chem. C acs.jpcc.6b10279 (2017).
477		doi:10.1021/acs.jpcc.6b10279
478	30.	Zheng, K., Zhao, D., Zhang, D., Liu, N. & Qin, W. Temperature-dependent six-photon
479		upconversion fluorescence of Er 3+. J. Fluor. Chem. 132, 5-8 (2011).
480	31.	Shao, W. et al. A core-multiple shell nanostructure enabling concurrent upconversion and
481		quantum cutting for photon management. <i>Nanoscale</i> 11, 11081–11095 (2017).
482	32.	Li, FC. & Kitamoto, Y. Fabrication of UCNPs/TiO2 aerogel photocatalyst to improve
483		photocatalytic performance. 20013, 20013 (2017).
484	33.	Park, Y. II et al. Facile Coating Strategy to Functionalize Inorganic Nanoparticles for
485		Biosensing. Bioconjug. Chem. (2016). doi:10.1021/acs.bioconjchem.6b00524
486	34.	Carney, R. P. et al. Determination of nanoparticle size distribution together with density or
487		molecular weight by 2D analytical ultracentrifugation. Nat. Commun. 2, 335 (2011).
488	35.	Arppe, R. et al. Quenching of the upconversion luminescence of NaYF 42:Yb $^{3+}$,Er $^{3+}$ and NaYF
489		$_4$ ${ m I}$:Yb $^{3+}$,Tm $^{3+}$ nanophosphors by water: the role of the sensitizer Yb $^{3+}$ in non-radiative
490		relaxation. <i>Nanoscale</i> 7, 11746–11757 (2015).
491	36.	Cong, T. <i>et al.</i> Upconversion luminescence enhancement in NaYF4: Yb3+, Er3+ nanoparticles
492		induced by Cd2+ tridoping. <i>Mater. Res. Bull.</i> (2017). doi:10.1016/j.materresbull.2017.02.032
493	37.	Haase, M. & Schäfer, H. Upconverting nanoparticles. <i>Angew. Chemie - Int. Ed.</i> 50, 5808–
494		5829 (2011).
495	38.	Menyuk, N., Dwight, K. & Pierce, J. W. NaYF4: Yb,Er - An efficient upconversion phosphor.
496		Appl. Phys. Lett. 21, 159–161 (1972).
497	39.	Heer, S., Kömpe, K., Güdel, H. U. & Haase, M. Highly efficient multicolour upconversion
498		emission in transparent colloids of lanthanide-doped NaYF4 nanocrystals. Adv. Mater. 16,
499		2102–2105 (2004).

500	40.	Wang, F. et al. Simultaneous phase and size control of upconversion nanocrystals through
501		lanthanide doping. <i>Nature</i> 463, 1061–1065 (2010).
502	41.	Haro-González, P. <i>et al</i> . Optical trapping of NaYF4:Er3+,Yb3+ upconverting fluorescent
503		nanoparticles. <i>Nanoscale</i> 5, 12192–9 (2013).
504	42.	Krämer, K. W. et al. Hexagonal Sodium Yttrium Fluoride Based Green and Blue Emitting
505		Upconversion Phosphors. Chem. Mater. 16, 1244–1251 (2004).
506	43.	Momma, K. & Izumi, F. VESTA 3 for three-dimensional visualization of crystal, volumetric
507		and morphology data. J. Appl. Crystallogr. 44, 1272–1276 (2011).
508	44.	MacKenzie, L. E. Graphic User Interfaces for the calculation of nanoparticle molecular
509		weight. (2017). doi:https://doi.org/10.5518/173
510	45.	Mittal, V., Völkel, A. & Cölfen, H. Analytical ultracentrifugation of model nanoparticles:
511		Comparison of different analysis methods. <i>Macromol. Biosci.</i> 10, 754–762 (2010).
512	46.	Wohlleben, W. Validity range of centrifuges for the regulation of nanomaterials: From
513		classification to as-tested coronas. J. Nanoparticle Res. 14, (2012).
514	47.	Bragg, W. H. & Bragg, W. L. The Reflections of X-rays by Crystals. Proc. R. Soc. A1 88, 428–
515		438 (913).
516	48.	Domingos, R. F. et al. Characterizing manufactured nanoparticles in the environment:
517		Multimethod determination of particle sizes. Environ. Sci. Technol. 43, 7277–7284 (2009).
518	49.	Na, H., Woo, K., Lim, K. & Jang, H. S. Rational morphology control of β -NaYF4:Yb,Er/Tm
519		upconversion nanophosphors using a ligand, an additive, and lanthanide doping. Nanoscale
520		5, 4242–51 (2013).
521	50.	Jia, H., Xu, W., An, J., Li, D. & Zhao, B. A simple method to synthesize triangular silver
522		nanoparticles by light irradiation. 64, 956–960 (2006).
523	51.	Shan, J., Uddi, M., Wei, R., Yao, N. & Ju, Y. The Hidden Effects of Particle Shape and Criteria
524		for Evaluating the Upconversion Luminescence of the Lanthanides Doped Nanophosphors. J.
525		Phys. Chem. C 114, 2452–2461 (2010).
526	52.	Lü, Q., Guo, F., Sun, L., Li, A. & Zhao, L. Silica-/titania-coated Y2O3: Tm3+, Yb 3+
527		nanoparticles with improvement in upconversion luminescence induced by different
528		thickness shells. J. Appl. Phys. 103, (2008).
529	53.	Li, H., Hao, S., Yang, C. & Chen, G. Synthesis of Multicolor Core/Shell
530		NaLuF4:Yb3+/Ln3+@CaF2 Upconversion Nanocrystals. Nanomaterials 7, 34 (2017).
531	54.	Lewis, D. J., Day, T. M., MacPherson, J. V. & Pikramenou, Z. Luminescent nanobeads:
532		attachment of surface reactive Eu(III) complexes to gold nanoparticles. Chem. Commun.
533		1433–1435 (2006). doi:10.1039/B518091K

534 55. Nel, A. E. *et al.* Understanding biophysicochemical interactions at the nano-bio interface.

535 Nat. Mater. 8, 543–557 (2009).

- 536 56. Bexiga, M. G. *et al.* Cationic nanoparticles induce caspase 3-, 7- and 9-mediated cytotoxicity 537 in a human astrocytoma cell line. *Nanotoxicology* **5**, 557–567 (2011).
- 538 57. Hou, Z. et al. UV-Emitting Upconversion-Based TiO ₂ Photosensitizing Nanoplatform: Near-
- 539 Infrared Light Mediated *in Vivo* Photodynamic Therapy *via* Mitochondria-Involved Apoptosis
 540 Pathway. *ACS Nano* 9, 2584–2599 (2015).
- 541

542 Acknowledgements

The authors would like to extend a special thanks to Amy Barker (School of Molecular and Cellular Biology, University of Leeds) for her technical assistance and expertise in conducting and analysing AUC experiments. We are also grateful to Professor Peter Stockley (School of Molecular and Cellular Biology, University of Leeds) for granting access to the AUC facilities.

547

548 **Funding Acknowledgements**

- L.E. MacKenzie was supported by a grant from the Biotechnology and Biological Sciences
 Research Council Tools and Development Resources Fund (BBSRC TDRF) (BB/N021398/1).
- J.A. Good is supported by a grant from the Medical Research Council (MRC) (MR/N029976/1).
- A. Vakurov was supported by a grant from the Natural Environment Research Council (NERC).
 (NE/N007581/1).
- Padmaja P. Nampi is supported by a European Commission Marie Skłodowska-Curie Individual
 Fellowship for Experienced Researchers (H2020-MSCA-IF-2015).
- 556 Competing financial interest statement
- 557 The authors declare no competing financial interests

558 Author contributions statement

- L.E.M and J.A.G. conceived the research concept and wrote the manuscript.
- L.E.M. performed all calculations, provided Figures 2, 3, 4, 5, 6, and created the stand-alone
- 561 GUIs.
- 562 J.A.G. provided Figure 1.
- A.V, P.P.N, S.S, G.J., and P.M, contributed to and reviewed the manuscript.