Cysteine proteases of hookworm *Necator Americanus* as virulence factors and implications for future drug design: A bioinformatics-based study.

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Abbreviations:

NA:	Necator Americanus
CPs:	Cysteine proteases.
SmSP1:	Schistosoma mansoni Serine Protease1
HMWK	High Molecular Weight Kininogen
T.Cruzi	Trypanosoma Cruzi
S.mansoni:	Schistosoma mansoni
P.falciparum	Plasmodium falciparum
P.Westermani	Paragonimus westermani
C. elegans	Caenorhabditis elegans
L. major	Leishmania major
A. Caninum	Ancylostoma caninum
S.japonicum	Schistosoma japonicum
H. Contortus	Haemonchus contortus
O. ostertagi	Ostertagia ostertagi
A. Suum	Ascaris suum

Abstract:

Human hookworm *Necator Americanus* causes iron deficiency anemia, as the parasite ingests blood from the gastrointestinal tract of its human host. The virulence factors of this blood feeding nematode have not been researched extensively. This bioinformatics based study focuses on eight of the cathepsin B like cysteine proteases (CPs) of the worm, which could have immense pathogenic potential. The role of the individual CPs remain vaguely determined except for CP3 which has been shown to act as globinase in the hemoglobin degradation pathway. In this study, the cysteine proteases were subjected to predictive molecular characterizations viz: probability of extracellular secretion to the interface between pathogen and host, ability for hemoglobin degradation, and/or interaction with host plasma proteins. CP1 - CP6, which harbored the active site cysteine and were observed to have N terminal signal peptide for extracellular localization, were relevantly predicted to be secretory. Amongst these, CP2 and CP3 showed the presence of hemoglobinase motif derived in this study that could be a prerequisite for globin or hemoglobin degradation. Similar S2 subsite feature of CP1 and CP6 that is shared with cruzain and cathepsin B is suggestive of involvement of the hookworm CPs in binding HMWK as substrate
CP1, CP2, CP3, CP5 and CP6 were predicted to bind heparin, which is the glycosaminoglycan molecule that has been demonstrated to aid the substrate-cleaving functionality of other cysteine proteases like human cathepsin B and cruzain. Heparin docked onto the *NA* CPs at the C terminal domain, away from the active site, similar to what has been shown for heparin binding to cathepsin B and cruzain. These observations therefore lead us to the hypothesize that the functions of the hookworm CPs, which could include preventing blood clot formation, might be assisted by heparin. This study underscores the potential of synthetic heparin analogs as molecular treatment for hookworm infection, which could have implications for future drug design.

Introduction:

Hookworm infection in humans is a neglected tropical disease that affects over 700 million people worldwide, mostly in the developing countries of the tropical and subtropical regions. (Hotez *et al*, 2004; Hotez *et al*, 2010) *Necator Americanus (NA)* species of hookworm constitutes the majority of these infections (~85%) (Loukas *et al*, 2011).

The clinical manifestation of the disease includes anemia, malnutrition in pregnant women, and cognitive and/or physical development impairment in children (Diemert *et al*, 2008). These helminth blood feeders on reaching maturity can feed up to 9ml of blood per day in an infected individual by attaching themselves to the intestinal mucosa of the host, through cutting plates as in *NA* (Pearson *et al* 2012). Iron deficiency anemia is the direct effect of the hookworm's blood feeding (Kassebaum *et al*, 2014), resulting in other subsidiary consequences of the hookworm disease (Sakti *et al* 1999; Hotez *et al*, 2008).

The infective larval stage (L3) worm penetrates into the host skin from soil (Vetter *et al*, 1977; Vetter *et al*, 1977) and then invades the circulatory system to reach heart and lungs, wherefrom it migrates to alveoli and then to trachea. The parasite eventually reaches gastrointestinal tract as fourth stage larvae (L4) to develop into blood feeding adult stage hookworms (Hotez *et al*, 2004).

An array of diverse enzymes and molecules in *NA*'s biomolecule repertoire facilitate the pathogen's survival in the host for up to seven years or longer during the different stages of its lifecycle (Pearson *et al* 2012). The most important therapeutic targets are the enzymes involved in interaction with host and in nutrient acquisition. These are often found in the excretory-secretory (ES) products of the worm. The ES proteins have been shown to engage in crucial functions like tissue degradation for host invasion (Pearson *et al* 2012, Brooker *et al*, 2004), fibrinogen degradation (Brown *et al*, 1995) for preventing blood clots, hemoglobin degradation (Brown *et al*, 1995) for nutrient acquisition, and evasion of the host immune system (Bungiro *et al*, 2011). The enzymes in the ES potpourri of *NA* are not yet completely characterized. However, Brown *et al*, had reported cysteine and serine proteases from ES products to degrade hemoglobin and fibrinogen, and detected the presence of at least two cysteine proteases operating at different pH optima (Brown *et al*, 1995).

The cysteine proteases (CPs) in *NA* form the ninth most gut-expressed abundant gene family (Ranjit *et al* 2006) These are most similar to *H.Contortus* (a blood feeding ruminant) CPs that dominate (~16%) intestinal transcriptome of the barber pole worm (Jasmer *et al*, 2001), highlighting the importance of pathogenic CPs in host blood degradation. The CPs are synthesized as precursor molecules, where in their folded proenzyme form, a self-inhibitory peptide blocks their catalytic domain. The removal of the inhibitory peptide upon proteolytic action of other peptidases releases the mature cysteine proteases, which are

active. The enzymatic clefts of the NA-CPs contain the catalytic triad residues: Cysteine, Histidine and Asparagine. Classification of cysteine proteases relies on the sequence homology spanning the catalytic residues (Sajid *et al*, 2002). CPs of parasitic organisms are divided into clans CA and CD. The clans are further classified into families in which Cathepsin B-like proteases belong to C1. The *NA*-CPs which are cathepsin B-like, therefore belong to the clan CA C1 family of cysteine proteases, which have a mostly α-helical amino-terminal known as the L domain, and the antiparallel β strands dominated carboxy-terminal is known as the R domain. The active site for proteolytic degradation is at the interface of these L and R domains. The entire cascade of hemoglobin degradation in hookworm has not been elucidated. However, certain key enzymes have been shown to participate in the degradation pathway. Aspartic protease *NA*-APR1 acts on hemoglobin, whereas metalloprotease MEP1 and cysteine protease CP3 degrade globin fragments (Ranjit *et al* 2009). While ingestion and digestion of blood, anticoagulant proteins are secreted to prevent clot formation (Stanssens *et al*, 1996; Harrison *et al*, 2002; Furmidge *et al*, 1995). Cysteine proteases, from *NA* (Brown *et al*, 1995) and from phylogenetically close *H. contortus* (Cox *et al*, 1990) have been implicated to have anticoagulation properties (Brown *et al*, 1995). *NA* adopts a number of complementary strategies to evade host procoagulation system (Furmidge *et al*, 1995), few of which have been elucidated so far.

Taken together, cysteine proteases from different pathogenic organisms perform diverse functions pertaining to blood feeding. This study focuses on eight cysteine proteases viz: CP1, CP2, CP3, CP4, CP4b, CP5, CP6, CP7 encoded by *NA* genome; of which CP2, CP3, CP4 and CP5 genes are reportedly expressed in abundance in the gut tissue of the adult worm (Ranjit *et al* 2008). Only CP3 amongst these has been characterized as a globinase (Ranjit *et al* 2009). The expression of the other CP genes in the *NA* gut is suggestive of their involvement in digestive or other assistive functions. Despite *NA*-CPs' importance in the parasite's physiology, they are under-researched and not much is known about the individual proteases and which of these constitute the ES products of *NA*. This bioinformatics based study on the molecular characterization of the CPs probes into those aspects of the proteases, which could have pathogenic potential such as hemoglobin degradation and blood clot prevention. Several bioinformatics methodologies have been applied ranging from sequence-based predictive methods, homology modeling, docking, motif derivation from sequence patterns, and mapping of molecular interactions to elucidate the role of the CPs as possible virulence factors and hence target for therapeutics. The approaches to some of the methods adopted here are in the context of other relevant cysteine proteases. Implications for the usage of heparin-analogs as worm-inhibitors have been outlined based on docking of heparin in the *NA*-CPs.

Materials and Methods:

1. Sequence alignments and pattern detection:

The following *NA* cysteine protease sequences were retrieved for analyses from Uniprot protein sequence database (The UniProt Consortium, 2015)(Uniprot ID in parentheses): Necpain or CP1 (Q9U938), CP2 (A1YUM4), CP3 (A1YUM5), CP4 (A1YUM6), CP4b (W2TRZ7), CP5 (A1YUM7), CP6 (W2T0C4) and CP7 (W2SQD9) (the organism code part of the ID is omitted for brevity). The alignments were done in ICM (Abagyan *et al*, 1994) by BLOSUM62 scoring matrix, with gap opening penalty of 2.40 and gap extension penalty of 0.15. Patterns of relevance and predicted or deciphered sites of functional importance were mapped onto the aligned CP sequences to denote their positions in the protein sequence.

1.1 N-terminal:

The N-terminal pre-sequences of *NA* CP1-6 were used to derive PRATT (Jonassen *et al*, 1995) patterns within them. The lack of these patterns was looked for in CP7 by ScanProsite (Sigrist *et al*, 2002) to determine CP1-6 specific N-terminal signature, as such signals often holds clue to protein sorting (Blobel, 1980).

1.2 Signal peptide cleavage:

SignalP (Petersen *et al*, 2011) was used to predict cleavage sites in the proteases, where the signal peptide would be cleaved off to generate the proenzymes.

1.3. Subcellular localization:

The *NA*-CP sequences were submitted for subcellular location prediction to TargetP (Emanuelsson *et al*, 2007), iPSORT (Bannai *et al*, 2002), TMHMM (Krogh *et al*, 2001), LocSigDB (King *et al*, 2007), Bacello (Pierleoni *et al*, 2006), Protein Prowler (Bodén *et al*, 2005), Cello (Yu *et al*, 2006) and PrediSi (Hiller *et al*, 2004) webservers to determine which of the proteases would be prone to secretion. While the algorithms for most of these programs take N-terminal signals into account, Bacello predicts localization on the basis of the information contained in the folded protein and LocSigDB is a signature pattern database derived from proteins whose localization has been confirmed by experiments.

1.4 Hemoglobinase motif:

The incompletely elucidated hemoglobin degradation pathway in *NA* describes the role of only CP3 as a globinase, amongst other CPs in the cysteine protease repertoire. In an effort to investigate the involvement of the rest of the *NA* CPs in hemoglobin degradation, cysteine protease sequences from other organisms - known to degrade hemoglobin - were taken along with *NA*-CP3 to derive conserved patterns unique to these proteins. Those organisms, the proteins, and their genbank (Benson *et al*, 2013) accession numbers (in parenthesis) are: *Necator Americanus* CP3 (ABL85237.1), *A. Caninum* CP1 (Q11006), *A. Caninum* CP2 (AIG62903.1), *S.mansoni* CB1 (3QSD_A), *P.falciparum* falcipain2 (AAK06665.1), *P.falciparum* falcipain 3(KOB61544.1), *S.japonicum* Cathepsin B (P43157.1), *H. Contortus* AC3 (Q25032), *H.Contortus* AC4 (Q25031), *P.Westermani* CP1 (AAF21461.1) and *O. ostertagi* CP1 (P25802.3) and *A. Suum* CP (AAB40605.1) Conserved patterns from the aforementioned proteins were derived in PRATT (Jonassen *et al*, 1995) and the motifs were scanned against some other non-hemoglobin degrading proteins in ScanProsite (Sigrist *et al*, 2002) to pinpoint patterns specific to the hemoglobin degrading enzymes. The set of non-hemoglobin degrading organisms and their relevant proteins were: *C. elegans*_CPR3 (AAA98789.1), *C.elegans*_CPR4 (AAA98785.1) and *L. major* cathepsin B (AAB48119.1). Such derived patterns specific to the hemoglobin degrading enzymes (when found) were scanned in the rest of the *NA* CP sequences.

1.5 Kallikrein-like activity:

Kallikrein-like activity of cleaving high molecular weight kininogen (HMWK) has been reported for cruzain, which is a cysteine protease of *T.Cruzi* – a parasite that traverses blood capillary vessels as trypomastigotes (Del Nery *et al*, 1997; Lima *et al,* 2002). Lys-bradykinin, a potent vasodilator, is one of the cleavage products generated by the Kininogen-cleaving kallikrein-like activity of cruzain (Maurer et *al*, 2011). As *NA* larvae too traces migratory route through blood capillaries of human hosts, the cysteine proteases of the human hookworm were scrutinized for molecular features for possible kallikrein-like activity, by aligning the cathepsin B-like *NA* CPs with cathepsin B (PDB: 1HUC) and cruzain (PDB: 2OZ2) – both of which cleave HMWK (Barros *et al*, 2004, Del Nery *et al*, 1997) The sequence alignment was done in ICM with the same parameters as mentioned before. The distances between the C-alpha atoms of crucial residues within the enzymatic cleft of relevant protease structures were measured in Chimera (Pettersen *et al*, 2004) for estimating the functional role of the residues.

2. Heparin binding:

Heparin was included in the study to explore the feasibility of its interaction with the *NA*-CPs, as glycosylation is an important post-translational modification in cysteine proteases of many pathogenic organisms (Sajid *et al*, 2002). Also, heparin-like glycosaminoglycan (GAGs) displayed on host proteoglycans (Bartlett

et al, 2010*)* are most probably encountered for recognition by the worm ES products at the host-pathogen interface.

2.1 Heparin binding domain prediction:

The sequences were subjected to query by ProDom (Bru *et al*, 2005), a protein domain family database derived from Uniprot knowledgebase. The search was carried out for the purpose of finding any domains from other organisms, with known heparin binding functionality.

2.2 Heparin glycosylation motif prediction:

The sequences were scanned in Scanprosite (Sigrist *et al*, 2002) for searching putative glycosylation motifs, which could be the attachment sites for pathogenic glycosaminoglycan like heparin.

2.3. Heparin binding docking simulation:

Lack of experimental three-dimensional structure of the cysteine proteases prompted BLAST search for homology model templates, against Protein Data Bank (PDB) (Berman *et al,* 2000). 3QSD - mature CathepsinB1 enzyme from *Schistosoma Mansoni* - was chosen from the search results for building models as it aligned well at the active site and had a resolution of 1.3 Å. Also, this structure had co-ordinates for the two occluding loop residues near the active site, which have been designated crucial for the exopeptidase activity for this class of cathepsin B like proteases (Illy *et al,* 1997). Homology models were built within the internal co-ordinates mechanics protocol of ICM software (Abagyan *et al,* 1994). The sequence alignment between the template and the model sequence was generated by using BLOSUM62 matrix, with gap opening penalty of 2.40 and gap extension penalty of 0.15. Further, for generating reliable models, the alignment around the active site was edited wherever necessary, according to conservation propensity of residues, and for modeling the occluding loop residues. Loops were sampled for the alignment gaps where the template did not have co-ordinates for the model sequence. The loop refinement parameters were used according to the default settings of the procedure. Acceptance ratio during the simulation was 1.25. The *NA*-CP model structures were then built within the full refinement module of the software. The quality of the homology models were checked using PROCHECK (Laskowski *et al*, 1993) which showed 100% of the residues from most of the CPs to lie within the allowed regions of the Ramachandran plot. CP2 and CP5 were the exceptions, which had 99.5% of residues in the allowed regions.

The co-ordinates for heparin were taken from its complex deposited in PDB (ID: 5D65) and saved as SDF formatted ligand. The CP homology models were converted to ICM formatted receptors for docking the heparin molecule. The sequence stretch of the CPs encompassing the predicted fibronectin domain, which can putatively bind heparin like molecules (Pankov *et al,* 2002) was selected for docking the heparin tetrasachharide which had alternating units of N,

O6-disulfo-glucosamine (PDB ID: SGN) and 2-O-sulfo-alpha-L-idopyranuronic acid (PDB ID: IDS). The receptor maps were generated with grid step of 0.50. The dockings were performed with a thoroughness level of 3, with the generation of three initial ligand conformations for each simulation.

The heparin-bound *NA* CP models were rendered in electrostatic surface representation by ICM (Abagyan *et al,* 1994), where the potential scale was set to 5.0 along with the assignment of simple charges, for the purpose of viewing the electrostatics of the protein sites occupied by the negatively charged heparin.

Results:

1. Sequence alignments and pattern detection:

1.1 N-terminal:
The *NA*-CP alignment showed that the CP7 lacked the N-terminal signal pre-sequence present in the other proteases (**Figure1**). The PRATT derived motif unique to CP1-6's pre-sequence was M-x(4,5)-L. CP7's N-terminal however had a lysosomal targeting pattern [DE]x{3}L[LI], according to LocSigDB. The specific residues in the pre-sequences are summarized in **Table1**, along with the lysosome targeting peptide in CP7.

1.2 Signal peptide cleavage:
SignalP derived cleavage sites predicted the length and the peptide sequence for the signals contained in the pre-sequences of CP1-6. **Figure 1** shows the positioning of the cleavage sites where the signal peptide would be cleaved off to release the proenzymes. The lengths of these signal peptides across the *NA* CPs were approximately the same.

1.3 Subcellular localization:
The consensus from the subcellular localization prediction methods deemed CP1-6 to be secretory proteins, with the aforementioned presence of pre-sequences, which signal for the proteases' extracellular localization. CP7 was predicted to be a lysosome directed protease (**Table 1**).

1.4 Hemoglobinase motif:
The hemoglobinase motif Y-[WY]-[IL]-[IV]-x-N-S-W-x-[DEGNQST]-[DGQ]-W-G-E-x(1,2)-G-x-[FI]-[NR]-[FILM]-x(2)-[DG]-x-[DGNS] was derived from the hemoglobin degrading cysteine proteases of the following blood feeders viz: *NA* (CP3), *A. Caninum*, *S.mansoni*, *S.japonicum*, *H. Contortus*, *O. ostertagi* and *A. Suum*. The pattern detected here is longer than the previously reported Y-W-[IL]-[IV]–A-N-SW-X–X–D-W-G-E motif by Baig *et al*, (Baig *et al*, 2002). See comparison in **Figure 1**. The training data set for deriving this new motif additionally included falcipain2 and falcipain3 of *P.falciparum* and CP1 of *P.westermani* and the derived motif was absent in the cysteine proteases of the non-blood feeders viz:

C. elegans and *L. major*. The observations from this study are similar to the previous study in the context of the presence of the motif in blood feeding proteases and its absence in the non-blood feeding proteases. The derived motif when searched in the *NA*-CPs (excluding CP3 of the training dataset) was detected only in CP2.

1.5 Kallikrein-like activity:

NA CP1 and CP6 were observed to have an aspartic acid and a glutamic acid respectively at the bottom of their enzymatic pocket similar to human cathepsin B's and cruzain's (both of which cleave HMWK) critical Glu at their S2 pocket, **(Figure 2)** which forms an important specificity determinant for substrates having Arg or Phe (Gillmor *et al*, 1997). *NA* CP1 and CP6, due to their similarity in enzymatic-pocket molecular feature with cruzain and human cathepsin B, are likely to bind human HMWK as their substrate in the region Leu^{373}-Gly-Met-Ile-Ser-Leu-Met-Lys-Arg-Pro-Pro-Gly-Phe-Ser-Pro-Phe-Arg-Ser-Ser-Arg-Ile^{393} of the kininogen; the same region that cruzain cleaves between Met^{379}-Lys^{380} and Arg^{389}-Ser^{390} to generate the vasodilator Lys - bradykinin (Del Nery *et al*, 1997). With the catalytic cysteine of the cysteine protease enzymes located between S1' and S1 subsites, where the peptide bond hydrolysis occurs, the cleavage sites of HMWK indicate that Arg^{381} and Phe^{388} of the kininogen are likely to get accommodated at the S2 subsite of cruzain. The side chain of the Glu208 at the S2 site could form salt-bridge interaction with the charged P2 Arg^{381}. When P2 is hydrophobic Phe^{388}, the glutamate could turn away to face the solvent. Therefore, the proteolytic action on HMWK by *NA*-CP1 and CP6 - which possess acidic Asp246 and Glu245 near their S2 pockets - could possibly release vasoactive peptides.

Cruzain's Ala138 and human cathepsin B's Ala171 at the S2 subsites of the enzymes, located near their critical Glu residue, corresponded with Ala175 of CP1 and Ala174 of CP6. The other *NA* CPs did not have this Ala residue at their S2 site. See alignment in **Figure 2.** The distances between the C-alpha atoms of the conserved Ala and the acidic Asp/Glu residues are listed in **Supplementary Table 1.** Such measurements were made in order to get an estimate of the distance of the critical Asp/Glu from the active site cleft, given the lack of ligand in the structures (except cruzain). The Ala and Asp/Glu around the S2 site of *NA* CP1 and CP6 were spaced further away compared to the distance between the relevant Ala and Glu in cruzain and cathepsin B. However, in the dynamic *NA* CP proteins, the side chains of the charged acidic residues could probably move closer (more so Glu245 of CP6, because of its longer side chain) to make the salt-bridge interaction with charged Arg^{381} of HMWK substrate.

2. Heparin binding:

2.1 Heparin binding domain prediction:

CP2 and CP5 sequences were predicted by ProDom to have fibronectin domain type III (entry: PDC9H7K4), which is known to harbor N-linked and O-linked glycosylation sites (Pankov *et al*, 2002) and has been shown to bind heparin (Takahashi *et al*, 2007). The predicted domain was 74 amino acids in length. CP3 having the longest sequence had 79 residues in the region. CP5 showed the RGD motif bordering the predicted fibronectin region (See alignment in **Figure 1**), which is a signature for type III_{10} repeat (Pankov *et al*, 2002). The region did not have any cysteine residues in any of the *NA* CPs for disulfide bond formation, which is another feature for fibronectin type III repeat (Pankov *et al*, 2002). With CP5 as reference, the fibronectin domain-like region in the rest of the secretory CPs showed sequence identity within 62.5% to 68.75% and the sequence similarity ranged from 56.45% to 71.43%. The high sequence identity and similarity implied that all the CPs would be predisposed to heparin binding, and so the entire stretch of the predicted fibronectin domain was considered for docking heparin.

2.2 Heparin glycosylation motif prediction:

N-glycosylation sites were predicted within the fibronectin domain region, which is another hallmark of type III repeats. 'NGTD' motifs were detected in *NA* CP1, CP3, CP4, CP4b and CP7, as per prosite entry PS00001. The glycosylation site region in the alignment (**Figure1**) showed CP2 to retain at least the Asn residue for N-glycosylation in its 'NGVK' stretch.

2.3. Heparin binding docking simulation:

Heparin on being docked at the putative *heparin-binding* fibronectin domain of the cathepsin B-like CPs showed the best scored conformation to bind surface loops (**Figure 3**), away from the enzymatic cleft (**Figure 4**), at a site similar to what has been reported in an earlier docking/MD simulation study on human cathepsin B - heparin interaction (*Costa et al, 2010*). The crucial interactions made there by the human cathepsin B's mentioned basic residues Lys156 and Arg233 for binding the negatively charged heparin, mapped close to this study's heparin-interacting Lys and Arg residues in most of the *NA*-CPs (derived from the alignment of human cathepsin B with *NA* CPs: **Figure 2**). This is the closest comparison (**Figure 1**), which could be drawn, with no structures of cathepsin B – heparin complex available in PDB. The binding site residues within 4Å of the ligand are underscored in the sequence alignment. Barring CP6, which showed the least sequence similarity with the reference sequence CP5 in the fibronectin domain region (**Supplementary Table 2**), heparin occupied similar sites in the CP homology models **(Figure 3)**. The positioning of heparin was slightly shifted in CP6, albeit away from the active site (**Figure 4**). The overall electrostatics (**Figure 4**) at the heparin-bound sites of the CPs varied from predominantly

neutral to positive (except CP2, CP4 and CP4b), depending on the site-residues and long-range electrostatic effects from residues beyond. The negative sulfate groups in the glycosaminoglycan molecule mostly interacted with basic/neutral residues at the binding site. Some of the heparin-contacts of the CPs showed the patterns: BBX, XBBX, BXB, and BXXBB, which were entirely or partially in conformity with previously reported heparin-binding motifs (Forster *et al,* 2006; Proudfoot *et al,* 2001*;* Mann *et al,* 1994*;* Fromm *et al,* 1997), where B is a basic residue and X is any amino acid. The highly negative binding-site electrostatics of CP4 and CP4b contributed to unfavorable docking scores for these proteases. The rest of the CPs showed scores for good binding of heparin. **Table 2** summarizes the scores, contact residues, sequence patterns, and H-bonding interactions of the highest scored conformations of heparin.

Discussion:

NA infection and survival in human hosts requires a repertoire of proteolytic enzymes. The parasite's physiology involves cysteine proteases for digestive purposes and evading potentially damaging host hemostatic events, only some of which have been characterized – that this study attempts to decode.

1. Sequence alignments and pattern detection:

1.1 N-terminal:

CP1-6 proteins' hydrophobic N-terminal pre-sequence are presumably signal peptides for extracellular localization. N-terminal signals are extremely degenerate across various proteins. The conserved hydrophobic M-x(4,5)-L sequence pattern in the *NA*-CPs therefore forms part of the unique signal peptide for these proteins.

1.2 Signal peptide cleavage:

CP7, which lacked the N-terminal pre-sequence, was not predicted to have any signal peptide cleavage site. The observation implies and re-emphasizes that this protease gets synthesized without the signal for extracellular localization, unlike the other CPs.

1.3 Subcellular localization:

CP1-6 are secreted out as per the subcellular localization predictions from this study, suggesting their presence in the ES products for host-pathogen interactions. CP3, amongst these, has been implicated to be present in the gut of an adult worm (Ranjit *et al*, 2008) and has been shown to be involved in the hemoglobin degradation pathway (Ranjit *et al* 2009). Therefore, CP3 being predicted to be secretory; CP1, CP2, CP4, CP4b, CP5 and CP6 also could be expected to be in the ES products of *NA* for similar or other supportive functions pertaining to blood feeding. Whereas, CP7 that lacks the active site cysteine

residue is predicted to reside in lysosome for unknown purposes.

1.4 Hemoglobinase motif:

The cysteine proteases from the *NA* ES products of the parasite had been demonstrated to cleave hemoglobin (Brown *et al*, 1995). However, only CP3 in the worm has been characterized to have such role. The molecular features pertaining to such function in *NA* CPs has not been researched. Hemoglobin degrading activity in Cathepsin B like proteases from blood feeding helminths were attributed to Y-W-[IL]-[IV]–A-N-SW-X–X–D-W-G-E sequence motif by Baig *et al*. In this study, a specific sequence pattern generic to hemoglobin degrading enzymes was sought, without emphasis on a particular family of cysteine proteases. This study therefore included hemoglobin-cleaving non-cathepsin B enzymes from *P.falciparum* and *P.Westermani*, which were not taken into account by Baig *et al*. Despite adopting a different methodology (mentioned in materials and methods) from the previous study for deriving the motif, a pattern unique to the blood degrading enzymes emerged. The hemoglobinase motif Y-[WY]-[IL]-[IV]-x-N-S-W-x-[DEGNQST]-[DGQ]-W-G-E-x(1,2)-G-x-[FI]-[NR]-[FILM]-x(2)-[DG]-x-[DGNS] which is being reported here is a longer pattern and working with such motif is advantageous in terms of avoiding false positives. The derived motif could be investigated in different enzymes across blood feeding pathogens. CP2 was the only protein in the repertoire of the *NA* cysteine proteases to have the motif, other than the already established globinase CP3. This observation is suggestive of CP2's involvement in hemoglobin degradation, along with CP3.

1.5 Kallikrein-like activity:

Human plasma kallikrein (pKal) is a component of the anticoagulation pathway that cleaves HMWK - a protein involved in blood coagulation and fibrinolysis - to produce a potent vasodilator bradykinin (Maurer *et al*, 2011) that stimulates prostacyclin (PGI_2) release from endothelial cells. PGI_2 in turn is an inhibitor of platelet activation and degranulation pathway (Maurer *et al*, 2011). Emulation of plasma kallikrein activity has been reported for cruzain (Del Nery *et al*, 1997; Lima *et al, 2002*) – a cysteine protease from *T.Cruzi* that traverses blood capillary vessels. Such function has been suggested also for SmSP1 - serine protease of blood fluke *S.mansoni* (Mebius *et al*, 2013), as kallikrein-like activity had been reported for a serine protease from *S.mansoni* (Carvalho *et al*, 1998, Da'dara *et al*, 2011). The blood stream navigating physiology of the mentioned pathogens explains their need for cleaving HMWK to generate bradykinin for vasodilation and blood clot prevention.

Cruzain, although being cathepsin L-like, shares similar substrate specificity with cathepsin B. The S2 pocket of the enzyme is the major specificity determinant, which has a glutamic acid residue at its bottom, same as what is found in cathepsin B's S2 subsite. This particular feature is responsible for their similar substrate bindings, (Gillmor *et al*, 1997) which includes HMWK - the cleavage of which by these proteases generates vasoactive bradykinin (Del Nery *et al*, 1997,

Barros *et al*, 2004). Cathepsin B, which has been implicated to impair coagulation and fibrinolysis under pathological conditions, (Herszenyi *et al*, 1997) has been shown also to generate bradykinin from HMWK (Barros *et al*, 2004). Cathepsin B-like extracellular *NA* CPs therefore are likely to exhibit HMWK-cleaving kininogenase activity. Especially CP1 and CP6 of the *NA* hookworm could possibly generate bradykinin by the dint of their critically positioned Asp246 and Glu245. The hookworm might be exploiting this component of the anticoagulation pathway (amongst others) to evade host blood clots for easing migration and/or facilitating feeding.

2. Heparin binding:

2.1 Heparin binding domain prediction:

Fibronectin can non-covalently bind a number of biologically important ligands that includes glycosaminoglycan such as heparin or heparan sulfate, which are present on the surfaces of extracellular proteoglycans (Pankov *et al*, 2002). The molecule's type II domain forms the primary binding site for heparin. However, fibronectin fragments with type III_{7-10} and type III_{10-12} repeats have been shown to bind heparin (Takahashi *et al*, 2007). In the cathepsin B-like *NA*-CPs, the predicted fibronectin domain type III overlapped with the β sheet and loop dominated R domains; structurally resembling the R domains of human cathepsin B and cruzain (**Figure 3**), where heparin has been implicated to bind (Costa *et al*, 2010, Lima *et al,* 2002). The negatively charged heparin occupied neutral-positive electrostatic patches in most of the different *NA* CPs (**Figure 4**) at about the same location as the low-energy heparin-docked region of human cathepsin B (Costa *et al*, 2010). Similar results were obtained in this study despite adopting a different methodology of pattern-based prediction method to specify the region for docking heparin, as compared to the Costa *et al* study, where they relied on protein patches having positive electrostatic potential. Heparin bound to the *NA*-CPs, away from the active site, making the K and R contacts, as made by human cathepsin B's K156 and R233 (**Figure 1**).

2.2 Heparin glycosylation motif prediction:

The prediction of N-glycosylation motifs within the predicted fibronectin type III domain region of the *NA* CPs hint that these proteases can have GAG molecules covalently attached to these sites as post-translational modifications. Such modifications could include heparin, which are reported to occupy N-glycosylation sites, and even sites without the Asn attachment point (Fenaille *et al*, 2007). N-glycosylation sites harboring *NA,* CP1, CP2, CP3, CP4 and CP4b, could be transporting the hookworm's indigenous heparin (if present) to the host-pathogen interface. CP5 and CP6, which did not show any N-motif in the region, could also be glycosylated with hookworm heparin. Heparin when released at the interface could possibly assist in evading blood coagulation, as has been

implicated for indigenous worm heparin/heparan sulfate found on the tegumental surface of blood fluke *S.mansoni* (Mebius *et al*, 2013).

2. 3 Heparin binding docking simulation:

Heparin-like glycosaminoglycan present in animal plasma membrane and ECM, form components of the host-pathogen interface (Bartlett *et al,* 2010). The GAGs serve as recognition factors for molecular interactions, controlling activities like cell adhesion and parasitic infection (Bartlett *et al,* 2010*;* Lima *et al,* 2002*;* Judice *et al,* 2013*).* These negatively charged molecules are covalently linked to syndecans and glypicans proteoglycans of host cells, harboring important binding sites for parasitic cysteine proteases (Bartlett *et al,* 2010*).* The simultaneous binding of these proteases with GAG and their protein substrate results in the formation of ternary complexes (Lima *et al,* 2002*;* Judice *et al,* 2013*),* which facilitates the enzymatic action of the proteases. This type of non-covalent heparin binding modulates the activity of the cysteine proteases, specifically cathepsin B - which otherwise tends towards alkaline pH induced inactivation. (Almeida *et al* 2001*,* Costa *et al,* 2010). Soluble proteoglycans or GAG chains released upon proteolytic cleavage of ECM or cell surface components, like their immobilized counterpart, can perform relevant functions (Bartlett *et al,* 2010). The implications of heparin binding to *NA* CP1, CP2, CP3, CP5 and CP6, as indicated by the docking scores, (**Table 2**) are discussed in the light of heparin binding to other cysteine proteases as follows.

2.2.1 Modulation of catalytic activity:

Human Cathepsin B bound by heparin/heparin-sulfate does not undergo alkaline pH induced loss of catalytic activity. Inactivation of cathepsin B under alkaline conditions occurs due to disruption of electrostatic interactions like the thiolate-imidazolium ion pair of the active site, and breakage of crucial salt bridge interactions. Heparin binding prevents the loss of such interactions and helps stabilize the protease's structure at alkaline pH by maintaining the helical content of the active enzyme (Almeida *et al* 2001). This allosteric mechanism mediated by heparin's binding away from the active site has been corroborated by computational studies (Costa *et al,* 2010*). NA*-CPs being cathepsin B-like are likely to be affected by heparin binding in a similar way. The molecule docked on to the *NA*-CPs, at a region similar to what has been reported by Costa *et al* in their cathepsin B–heparin dockings (**Figure 1**). This is suggestive of possible allosteric control that could be exercised by heparin over the enzymatic action of the *NA*-CPs to make them functional even at alkaline pH. Such controls could be crucial for the survival of hookworm, which encounters different pH conditions within the host, and the cysteine proteases of which have varying degrees of overall surface electrostatics (**Figure 4**).

2.2.2 Activation of anticoagulation pathway:

2.2.2.1 Kallikrein-like activity:

The kininogenase activity in cruzain is induced by heparin by forming a complex with kininogen and cruzipain, for assisting the proteolytic cleavage of kininogen to release Lys-bradykinin (Lima *et al,* 2002). The docking results from this study suggest high likelihood of heparin binding to most of the cathepsin B-like *NA* CPs, which (especially CP1 and CP6) in turn can bind HMWK as their substrate to form ternary complex. The implications of such interactions, if validated, could underscore hookworm CPs' role in possible kininogen-cleaving activity for generating vasodilators to aid hookworm larvae's migration through small blood vessels.

2.2.2.2 Antithrombin activity:

The disruption in flow (stasis) of the ingested blood at the adult hookworm's gut would tend to trigger coagulation (Lowe 2003*;* Bagot *et al,* 2008). Inhibiting such coagulation would be a prerequisite for blood degradation. Heparin is known to serve as co-factors to antithrombin - an inhibitor of thrombin and other coagulation factors in the blood plasma. Heparin increases the thrombin binding efficiency of antithrombin by 2,000 to 10,000 fold (Beck *et al,* 1985) and thereby becomes extremely effective in preventing blood clots. Soluble free heparin-like GAG chains could be possibly released for such functions, by the proteolytic action of the *NA* CPs on host proteoglycans, or by deglycosylation of the *NA* CPs. Either way, heparin thus released could bind plasma antithrombin to enhance anticoagulation activity of hookworm.

2.2.5 Therapy:

NA-CPs' suggestive role in hemoglobin degradation, anticoagulation and cell - aided by heparin-like molecules as mechanistically described before, probably requires abolishment of GAG binding to the *NA*-CPs for preventing the proteases' pathogenic activities. Blocking the putative GAG binding site on the parasite's CPs, (see residues in **Table 2**) could help thwart hookworm infection. Soluble heparin/heparin sulfate usage as competitive inhibitors have the chances of causing adverse physiological effects due to their anticoagulant/immunogenic properties (Bartlett *et al,* 2010). However, synthetic GAG mimetics with limited biological activity (minimum active structure) or polysaccharides (Vann *et al*, 1981*;* Copeland *et al,* 2008) targeted at GAG binding site, could be used for inhibiting hookworm infection in conjunction with active-site inhibitors.

This study was initiated to explore the role of *NA* CPs in human hookworm pathogenesis*,* as the proteases remain uncharacterized despite their demonstrated pathogenic potential (Brown *et al*, 1995). Bioinformatics based analyses of the *NA*-CPs is indicative of the presence of CP1, CP2, CP3, CP4,

CP4b, CP5 and CP6 in the ES content of the hookworm for host-pathogen interactions. The hemoglobinase motif derived here is harbored by CP2 and CP3, which suggests hemoglobinase activity for these two proteins, of which only CP3 has been confirmed to degrade globin (Ranjit *et al*, 2009). Possible HMWK – cleaving activity by *NA* CP1 and CP6 hint towards the proteases' role in evading the host hemostatic system for preventing blood clots, in order to facilitate feeding and survival. The heparin docking results from this study is speculative of *NA* CPs' heparin-assisted HMWK-cleaving activity. Such function, already verified for the proteases from other parasites like *T.Cruzi* (Del Nery *et al*, 1997; Lima *et al,* 2002) and *S.mansoni* (Carvalho *et al*, 1998) that navigate blood capillaries, leads to the hypothesis of similar survival strategy adoption by larvae-stage *NA* for migrating through blood capillaries. The predicted extracellular localizations of CP1-CP6 and their predicted pathogenic roles render these CPs to be possibly multi-targeted for heparin-analog binding, which could inhibit hookworm infection. This study attempts to provide molecular level information based on computational predictions, decoding previously unreported possible functions of the *NA* cysteine proteases, which could be subjected to further experimental validation.

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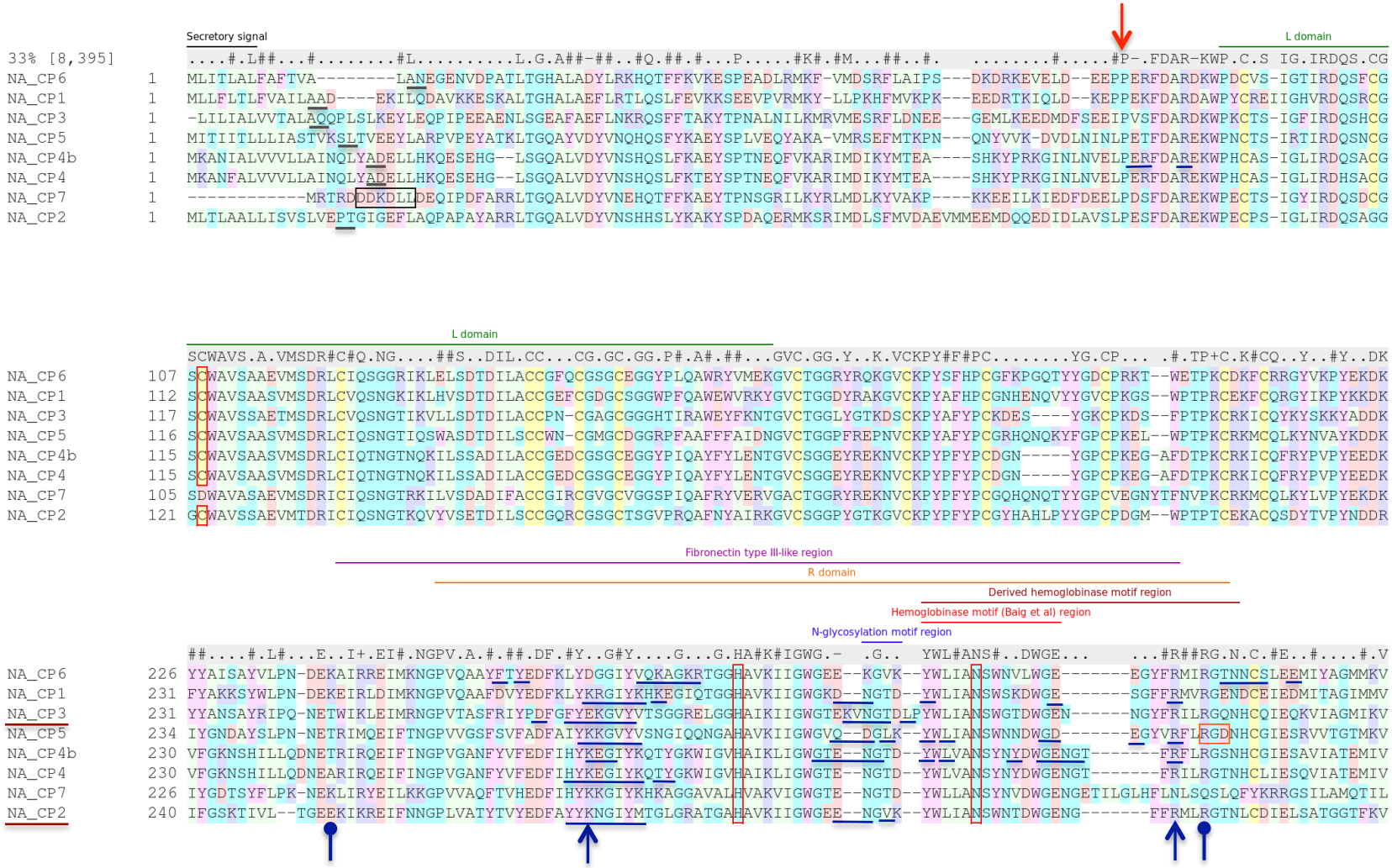
NA Cysteine Protease	Localization	Signal motif	Motif specific to protea
CP1	Secretory	M-x(4,5)-L	MLLFLTL
CP2	Secretory	M-x(4,5)-L	MLTLAAL
CP3	Secretory	M-x(4,5)-L	LILIAL
CP4	Secretory	M-x(4,5)-L	MKANFAL
CP4b	Secretory	M-x(4,5)-L	MKANIAL
CP5	Secretory	M-x(4,5)-L	MITIITL
CP6	Secretory	M-x(4,5)-L	MLITLAL
CP7	Non-secretory (Lysosomal)	[DE]x{3}L[LI]	DDKDLL

Table1: The subcellular localization of the *NA* cysteine proteases, and the motifs pertaining to the signals for the localization

NA-CP	Score	Contact Residues	Motif pattern	H-bonding residues
CP1	-21.75	**K186, R187, G188**, I189, Y190, K191, K193, D211, N212, T214, Y216, E229, R234	BBX	R187, K191, N212, R234
CP2	-24.22	Y182, Y183, K184, N185, G186, I187, Y188, M189, E209, N210, V212, R232.		Y182, K184, N185, M189, E209, N210, R232
CP3	-20.33	D176, F179, Y180, E181, K182, G183, V184, Y185, K207, V208, N209, G210, T211, L213		E181, K182, V184, N209, G210
CP4	-3.809	**H180, Y181, K182**, E183, G184, I185, Y186, K187, T189, Y190.	BXB	K182, I185, K187, T189
CP4b	-8.224	E2, R3, R7, K182, E183, G184, G205, T206, E207, N208, G209, Y212, L214, Y221, G224, E225, N226, G227, T228, R230		R3, E183, T206, E207, N208, G209, Y212, E225, N226
CP5	-15.4	**Y183, K184, K185, G186,** V187, Y188, V189, Q209, D210, L212, Y214, L216, G226, D227, E228, R232	XBBX	K184, K185, Q209
CP6	-27.69	F175, Y177, V189, Q190, **K191, A192, G193, K194, R195**, T237, N238, N239, C240, S241, E244	BXXBB	Q190, G193, R195, N238, N239, S241

Table 2: Docking scores, contact residues, sequence motif patterns, and H-bonding interactions for the highest scored conformations of heparin molecule docked onto the *NA* mature cysteine proteases.



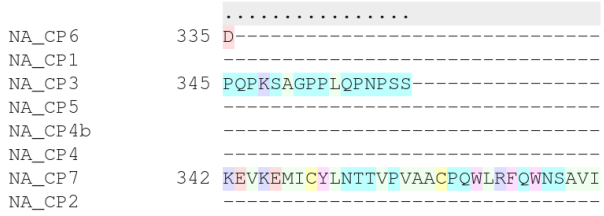


Figure 1 : Sequence alignment of the *NA* CPs with annotated functional regions. The begin position of the mature proteases are indicated by red arrow and the catalytic triad residues are boxed in red. The cleavage sites between the extracellular signal peptide and zymogen-sequence are underscored in black. The lysosome-targeting peptide of *NA* CP7 is boxed in black. *NA* CP2 and CP3 which show the presence of the derived hemoglobinase motif are underscored in dark red. The RGD motif signature in *NA* CP5 for fibronectin type III is boxed in orange. The residues in contact with the docked heparin molecule are underscored in blue. The K and R residues contacting heparin in most of the docked complexes are shown by blue arrows to denote their positions with respect to the regions corresponding to human cathepsin B's K156 and R233 residues, (indicated by spherical arrowheads) shown to be crucial for binding heparin (Costa *et al*, 2010).

Figure 2 : Sequence alignment of the *NA* CPs with HMWK-cleaving and heparin-binding cruzain and human cathepsin B. The Glu of the S2 subsite of cruzain and cathepsin B, shown to be crucial for substrate binding, are within red oval. The position corresponding to CP1's Asp and CP6's and Glu placed near their S2 pockets, is indicated by red arrow. The position of Ala of the S2 pocket of HMWK-cleaving cruzain and cathepsin B, also conserved between CP1 and CP6, is indicated by green arrow. The putative glycosylation motifs within the region are boxed in blue. The K156 and R233 residues hypothesized to be crucial for heparin-binding in human cathepsin B (Costa *et al*, 2010) are boxed in blue.

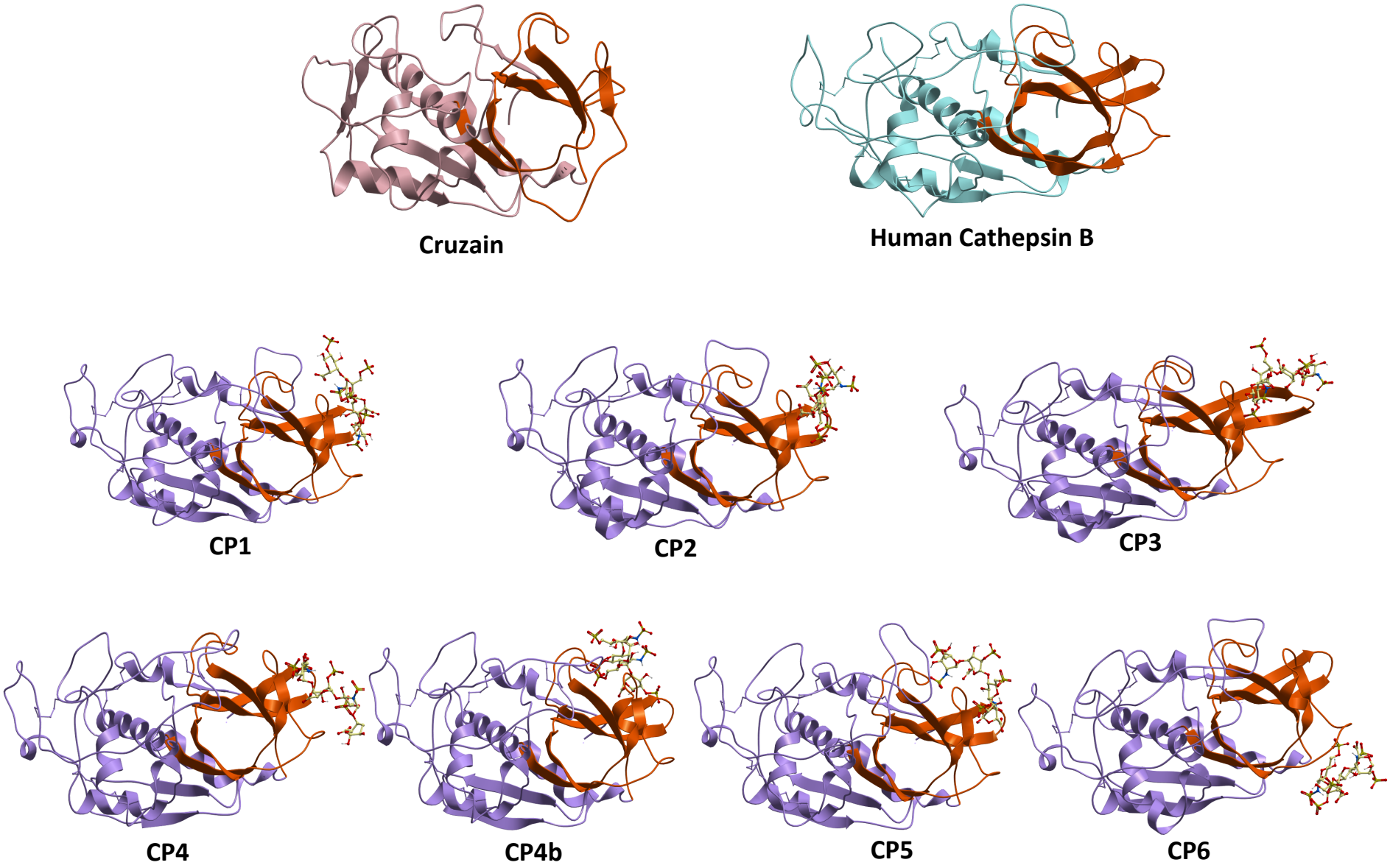


Figure 3: The R domains (orange) of Cruzain (PDB ID: 2OZ2) and human Cathepsin B (PDB ID: 1HUC) implicated to bind heparin, compared to structurally similar R domains (orange) of the *NA*-CP homology models. The antiparallel beta strand and loop dominated domain overlaps with the fibronectin type III-like region at the C-terminus of the *NA* CPs. The docked heparin molecule (represented in stick) mostly makes contact with the loops of the highlighted domain.

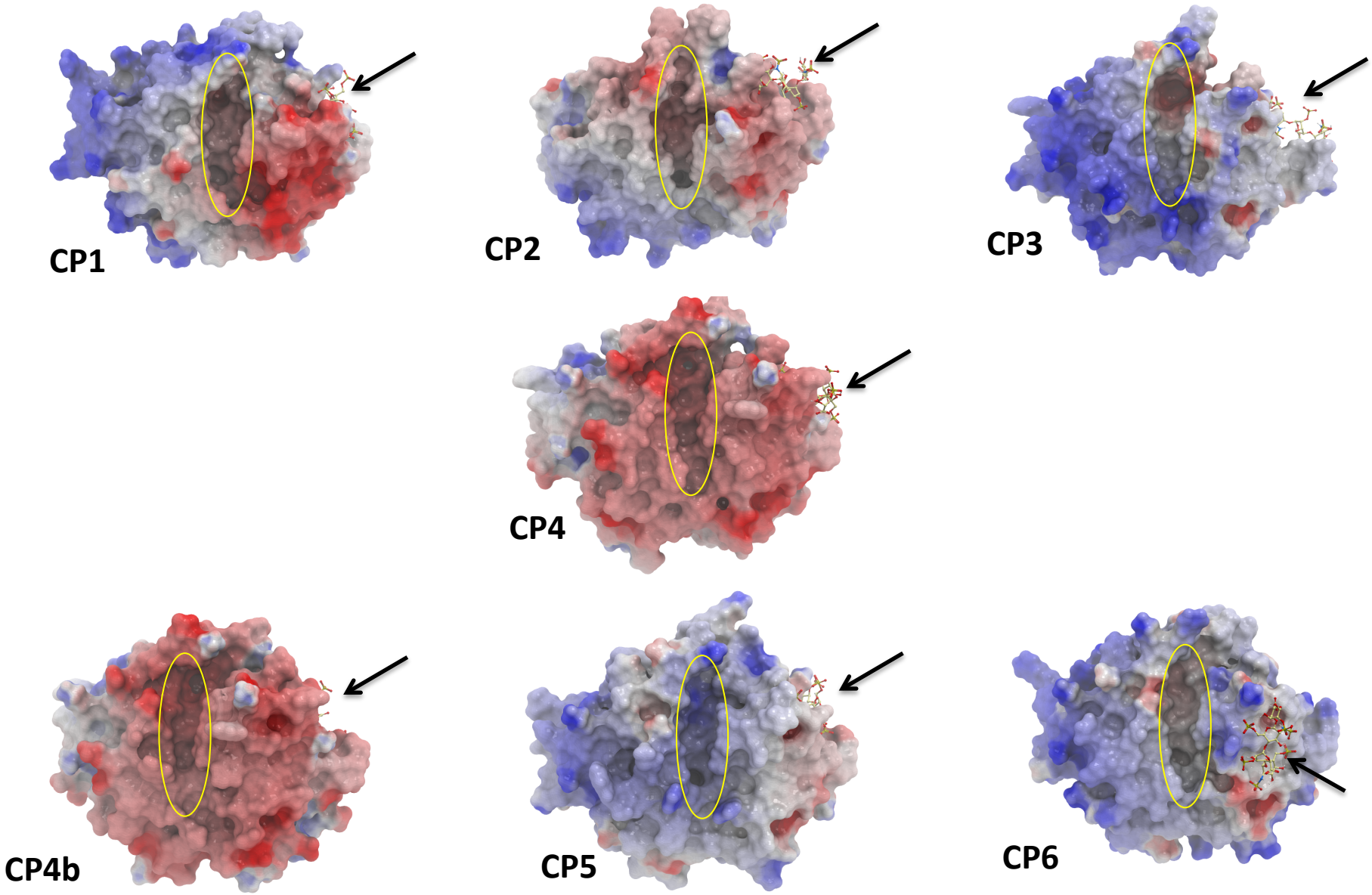


Figure 4: Heparin-docked homology models of the *NA*-CPs in electrostatic surface represention. The active sites of the cysteine proteases, appearing as clefts, are shown within yellow ovals. The location of the docked heparin (represented in stick) which bind away from the active site are indicated by arrows.