

Quantifying the fitness benefit and cost of cefixime resistance in *Neisseria gonorrhoeae* to inform prescription policy

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Abstract

Gonorrhoea is one of the most common bacterial sexually transmitted infections in England. Over 41,000 cases were recorded in 2015, more than half of which occurred in men who have sex with men (MSM). As the bacterium has developed resistance to each first-line antibiotic in turn, we need an improved understanding of fitness benefits and costs of antibiotic resistance to inform control policy and planning. Cefixime was recommended as a single dose treatment for gonorrhoea from 2005 to 2010, during which time resistance increased and subsequently declined. We developed a stochastic compartmental model representing the natural history and transmission of cefixime sensitive and resistant strains of *Neisseria gonorrhoeae* in MSM in England, which was applied to data on diagnoses and prescriptions between 2008 and 2015. We estimated that asymptomatic carriers play a crucial role in overall transmission dynamics, with about 40% of infections remaining asymptomatic and untreated, accounting for 96% of onward transmission. The fitness cost of cefixime resistance in the absence of cefixime usage was estimated to be such that the number of secondary infections caused by resistant strains is only about half as much as for the susceptible strains, which is insufficient to maintain persistence. However, we estimated that treatment of cefixime-resistant strains with cefixime was unsuccessful in 84% of cases, representing a fitness benefit of resistance. This benefit was large enough to counterbalance the fitness cost when 31% of cases are treated with cefixime, and when more than 51% of cases were treated with cefixime the resistant strain had a net fitness advantage over the susceptible strain. Our findings have important implications for antibiotic stewardship and public health policies, and in particular suggest that cefixime could be used to treat a minority of gonorrhoea cases without raising resistance levels.

Introduction

Gonorrhoea, caused by the bacterial pathogen *Neisseria gonorrhoeae*, is one of the most common sexually transmitted infections in England. Incidence has increased year on year since 2008, culminating in over 41,000 cases in 2015 [1]. Around 22,000 of these cases were found in men who have sex with men (MSM), constituting a 20% annual increase. The greatest cause for concern, however, is the rapid growth in antimicrobial resistance. The bacterium has quickly developed resistance to each first-line antibiotic in turn, from penicillin through to cephalosporins, such as cefixime and ceftriaxone [2]. Treatment with ceftriaxone is the last remaining single-drug option in most settings worldwide; however susceptibility is diminishing rapidly [3]. As a result, England and many other countries now recommend treatment of gonorrhoea with a dual therapy of ceftriaxone and azithromycin [4]. Ceftriaxone resistance has been detected only sporadically in England; however azithromycin resistance is easily selected for and was prevalent in a recent outbreak [5]. Resistance to azithromycin effectively reduces the current treatment to a monotherapy, making resistance trends increasingly important to monitor against the threat of potentially untreatable gonorrhoea.

Public Health England (PHE) runs the Gonococcal Resistance to Antimicrobials Surveillance Programme (GRASP) [68], which has produced a report annually since 2000 [6–21]. GRASP monitors trends in resistance and susceptibility to a panel of antibiotics used to treat gonorrhoea in England and Wales, and thus informs national treatment guidelines and strategy. In 2004, GRASP began testing for cefixime resistance, defined as having a Minimum Inhibitory Concentration (MIC) of ≥ 0.125 mg/l [10]. In 2005, following worrying increases in resistance to the previous therapy, ciprofloxacin, a new recommendation was introduced that uncomplicated gonorrhoea should be treated with a single dose of cefixime [22].

Fig 1 shows the trends in cefixime prescription and resistance in England. Very little resistance was detected until 2007, however by 2009 the total level of resistance had passed the 5% threshold at which the WHO recommends that first-line treatment guidelines should be changed [13,23]. At this time almost 60% of gonorrhoea diagnoses were being treated with cefixime [15]. The majority of the resistance was concentrated in the MSM population, where it reached a peak of 33% in 2010 [16]. This evidence, combined with increasingly common reports of cefixime treatment failure, formed the basis for the decision in May 2011 for another update to the treatment guidelines for uncomplicated gonorrhoea [24,25]. Cefixime was no longer recommended as a first-line treatment, and was replaced with a combination of 500mg ceftriaxone and 1g azithromycin [4]. Since 2011, cefixime prescribing has fallen drastically, in

line with the updated guidelines. Over the same period the proportion of cefixime-resistant isolates has declined steadily in MSM, falling to less than 1% in 2014 [20].

We hypothesize that the resistance trend observed can be explained by a net fitness benefit to cefixime resistance when cefixime is widely prescribed but a net fitness cost when cefixime prescriptions decline. Understanding the relationship between antibiotic use and the emergence of resistance in gonorrhea has been identified as a key research agenda [26]. Here our main aim is to further the understanding of the evolutionary dynamics of cefixime resistance, and to use this newfound knowledge to inform public health practice. There is still much we do not understand about the natural history of gonorrhea, especially since unobserved asymptomatic infections have long been thought to be an important reservoir of infection. The proportion of incident cases that are asymptomatic at each bodily site of infection is known to vary, but has not been definitively measured [27–29]. Furthermore, the expected duration of carriage of asymptomatic gonococcal infection is not well studied. Estimates have been traditionally in the region of six months, however recent work using genomic data on pairs of known sexual contacts has suggested that a longer duration of carriage can occasionally happen [30]. We therefore developed and applied a Bayesian statistical approach to account for these uncertainties in the epidemiology of gonorrhea.

Materials and methods

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Epidemiological Data

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The total number of diagnoses of gonorrhoea in MSM in England between 2008 and 2015 were extracted from the Genitourinary Medicine Clinic Activity Dataset (GUMCAD) [60]. This mandatory reporting system provides data on diagnoses of sexually transmitted infections from sexual health services in England, and the GUMCAD data is published annually by PHE on their website at <http://www.gov.uk/government/statistics/sexually-transmitted-infections-stis-annual-data-tables>. This yearly number of gonorrhoea diagnoses is denoted $Y(t)$.

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The number of cases of gonorrhoea in MSM that were cefixime-resistant and reported by GRASP between 2008 and 2015 were extracted from the corresponding GRASP reports [14–21] and denoted $Y^{\text{res}}(t)$. The coverage of GRASP was calculated for every year between 2008 and 2015 by taking the ratio between the number of cases included in GRASP (irrespective of resistance) and the number of GUMCAD diagnoses in the same year. This GRASP coverage proportion is denoted $q(t)$. The proportion of gonorrhoea cases that were treated with cefixime, as opposed to other antibiotics, was also extracted from the GRASP reports between 2008 and 2015. This time-dependent proportion is denoted $\pi(t)$ and illustrated in Fig 1.

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The MSM population is estimated at $N = 1.5$ million, based on a sexually active male population of 20 million [33]. In the third National Survey of Sexual Attitudes and Lifestyles (Natsal) 8.4% of men reported same-sex experience at least once, with 2.6% of men having had a same-sex partner in last five years, putting a plausible range for the MSM population at 0.5 and 1.7 million [34]. Natsal is known to under-represent MSM so an estimate towards the top of the range was adopted [35]. Given the low prevalence of gonococcal infection in the population, the total population size is not expected to excessively affect the results.

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Transmission model

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In order to investigate the fitness cost and benefit of cefixime resistance in gonorrhoea, we created a stochastic compartmental model, illustrated in Fig 2 with notation summarised in Table 1. High rates of reinfection with gonorrhoea have been observed, suggesting low levels of acquired immunity [31], and experimental urethral infection in male volunteers found no protection was conferred on repeat infection with an identical strain six months apart [32]. It was therefore assumed that no immunity was conferred upon recovery from infection. The analysis was restricted to MSM, the population in which the cefixime-resistant outbreak of gonorrhoea was concentrated. A closed population of size N was assumed due to the short time period under consideration. Individuals are initially susceptible (S). They become infected with strain $s \in \{\text{sus}, \text{res}\}$, denoting cefixime-susceptible and resistant strains respectively. The model assumes that strains do not vary in transmissibility, and that the rate of infection from an infectious individual to a susceptible individual is θ/N . Infected individuals initially pass through an incubation period (U_s) which they leave at rate σ . A proportion ψ of those infected then go on to develop symptoms (E_s), whereas the remainder enters an asymptomatic stage (A_s). Recovery from asymptomatic infection happens (either naturally or following unrelated antibiotic treatment) at rate ν for the susceptible strain and at rate $\alpha\nu$ for the resistant strain. The parameter α therefore represents the fitness cost of cefixime resistance. The infected population for each strain s is denoted $I_s = U_s + E_s + A_s$, and the total infected population denoted $I = I_{\text{sus}} + I_{\text{res}}$. All infected individuals are assumed to be infectious. The symptomatic individuals (E_s) seek treatment at rate μ . A time-varying proportion $\pi(t)$ are treated with cefixime ($T_{s;\text{cef}}$) whereas the remaining $1 - \pi(t)$ are treated with other antibiotics ($T_{s;\text{oth}}$). The treated individuals recover from the infection and become susceptible again at rate ρ , with the exception of a proportion ϕ of the individuals infected with a cefixime-resistant strain and who have been treated with cefixime ($T_{\text{res};\text{cef}}$) for whom treatment failure happens and who become asymptotically infected (A_{res}) [51, 69].

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Calculation of the basic reproduction number

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The basic reproduction number, R_0 , is a measure of the reproductive capacity of an infectious agent and is defined as the average number of secondary cases of gonorrhoea arising from the introduction of a typical infected individual in a completely susceptible population. Where there is direct competition between strains, as in the situation we are modelling, the strain with the highest R_0 will outcompete the others.

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To calculate R_0 we must consider the generation-time, defined as the expected time from an individual becoming infected to infecting another individual [37]. By considering the expected time spent in each compartment of the model corresponding to infection with the susceptible strain (ie. states U_{sus} , E_{sus} and A_{sus} in Fig 2), we derive an analytical expression of the basic reproduction number R_0^{sus} for the susceptible strains:

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$$R_0^{\text{sus}} = \theta \left(\frac{1}{\sigma} + \frac{1 - \psi}{\nu} + \frac{\psi}{\mu} \right) \quad (1)$$

Similarly, the basic reproduction number R_0^{res} for the resistant strains is given by:

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$$R_0^{\text{res}} = \theta \left(\frac{1}{\sigma} + \frac{1 - \psi + \phi\pi\psi}{\alpha\nu} + \frac{\psi}{\mu} \right) \quad (2)$$

Bayesian inference

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We considered the data as a Partially Observed Markov Process, with the number of GUMCAD recorded cases, $Y(t)$, and GRASP reported resistant cases, $Y^{\text{res}}(t)$, being the observed realisations of the underlying unobserved processes: the total incidence of gonorrhoea infections, $Z(t)$, and incidence of cefixime-resistant infections, $Z^{\text{res}}(t)$. The reporting process for the total number of cases recorded by GUMCAD was set to allow for a 10% under-reporting rate on average, with a 10% margin of error:

$$Y(t) \sim \text{Normal}(0.9Z(t), 0.05Z(t))$$

The probability of a cefixime-resistant case of gonorrhoea being sampled by the GRASP study was assumed to be Poisson distributed with a sampling probability denoted $q(t)$ derived from the coverage of the GRASP study over 2008 to 2014:

$$Y^{\text{res}}(t) \sim \text{Poisson}(q(t)Z^{\text{res}}(t))$$

Based on these observations we aimed to infer the values of the ten parameters: $A_{\text{sus}}(0)$, $A_{\text{res}}(0)$, θ , ψ , σ , ν , α , μ , ρ and ϕ .

An analytical expression for the likelihood of the observed data given our model is not available, so we obtained an unbiased estimate of the likelihood using a particle filter [38]. The estimated likelihood was then incorporated into a particle Monte Carlo Markov Chain (pMCMC) which was used to obtain a sample from the posterior distribution of the model parameters [39].

The model fitting was implemented using the R package pomp, which includes a pMCMC algorithm that can be used to perform Bayesian inference [40]. The algorithm was modified to enable parallel computation. The particle filter estimation of the likelihood was based on 1,000 particles, which was sufficiently robust to estimate the likelihood. 6 million iterations of the pMCMC were thinned by a factor of 100. Four separate chains were run with dispersed starting points, and compared using the R package coda [41]. The chains were found to have converged to the same posterior distribution based on the multivariate version of the Gelman-Rubin diagnostic, which was less than 1.1 for all inferred parameters [42, 43]. To ensure maximum robustness, the samples from the four chains were then combined, and found to have an effective sample size of more than 150 for all parameters.

Prior distributions of parameters

Bayesian inference requires setting plausible priors for the model parameters. We used highly uninformative Uniform(0,1) priors for the two proportion parameters ϕ and ψ and Uniform(0, ∞) priors for the three parameters $A_{\text{sus}}(0)$, $A_{\text{res}}(0)$ and α , which is an improper distribution but does not lead to an improper posterior distribution. For the five remaining parameters θ , ν , σ , μ and ρ we assigned informative Gamma priors based on literature review, as summarised in Table 1.

The transmission rate of infection, represented by the parameter θ , encompasses both the average number of sexual partners per year and the transmission probability per partnership. The Natsal-3 survey observed a mean number of sexual partners per year for MSM of 4.4 [34] and we would therefore expect θ to be slightly lower, to reflect the fact that not all contacts result in transmission. The prior distribution for θ was therefore set such that it was between 2.9 and 6.3 with 99% prior weight.

The expected duration of carriage for asymptomatic gonorrhoea is not well measured. A study of 18 asymptomatic infected men saw no resolution in urethral infection in the 165 days until they received treatment [51]. Estimates of the duration of carriage in modelling studies have been based on calculations that take into account observed prevalence and assumed proportion of unobserved infection, and are often in the region of 6 months [52–54]. However, in recent work using genomic data, the greatest observed time to most recent common ancestor for bacterial genomes from known contact pairs was 8 months, suggesting that this estimate needs to be increased [30]. Duration of carriage may depend on infection site, for pharyngeal gonorrhoea it has been estimated at 12 weeks, and for rectal infection has been estimated at one year [49, 55]. The parameter ν was therefore assigned a prior that corresponded to a mean duration of carriage between three months and one year with 99% prior weight.

The duration of the incubation, symptomatic and treatment stages of infection have been estimated to be short, in the region of days rather than weeks [45–47]. Gamma priors were accordingly assigned to each of the three parameters σ , μ and ρ .

Results

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Estimation of model parameters

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We fitted our model of gonorrhoea transmission to two different time series over the years 2008 to 2014: the total number of gonorrhoea diagnoses in MSM in England [44] and the incidence of cefixime-resistant gonorrhoea [14–20]. The posterior distribution of parameters shown in Fig 3 was obtained through Bayesian inference, implemented using a pMCMC method [39]. For each parameter we report the posterior mean estimate and 95% credibility interval shown in brackets (Table 1). The model suggests that at the end of 2007, the first year that cefixime-resistant cases were detected by GRASP [13], there were 630 cases (417, 888) of asymptomatic cefixime-susceptible gonorrhoea ($A_{\text{sus}}(0)$), and 49 cases (19, 94) of asymptomatic cefixime-resistant gonorrhoea ($A_{\text{res}}(0)$).

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Table 1. Parameter notations, prior and posterior distributions

Parameter	Description	Prior	Posterior mean (95% CI)
$A_{\text{sus}}(0)$	initial carriage of cefixime-susceptible infection	Uniform($0, \infty$)	630 (417, 888)
$A_{\text{res}}(0)$	initial carriage of cefixime-resistant infection	Uniform($0, \infty$)	49 (19, 94)
θ	rate of transmission	Gamma(44, 10)	5.2 (4.2, 6.5)
ψ	proportion of infections that become symptomatic	Uniform(0, 1)	62% (46%, 77%)
σ	rate of departing incubation period	Gamma(17, 0.22)	79 (46, 119)
ν	rate of recovery from asymptomatic infection	Gamma(8, 3.45)	1.9 (0.9, 3.1)
μ	rate of seeking treatment when symptomatic	Gamma(3, 0.02)	131 (33, 303)
ρ	rate of recovery following treatment	Gamma(101, 1.9)	53 (43, 63)
α	increased recovery from cefixime-resistant infection	Uniform($0, \infty$)	1.7 (1.4, 2.4)
ϕ	proportion of treatment failures for cefixime resistant infections treated with cefixime	Uniform(0, 1)	84% (55%, 100%)

The posterior distribution of the rate of transmission, θ , suggests a higher mean rate of infection than the prior expectation: 5.2 (4.2, 6.5), but the prior and posterior credible intervals overlap to a large extent, suggesting that the results are consistent with our prior knowledge. Our model predicts that the proportion ψ of infections that become symptomatic is 62% (46%, 77%). The three parameters σ , μ and ρ , corresponding respectively to the durations of the incubation period, symptomatic infection before seeking treatment, and the treatment phase, had posterior distributions similar to their prior distributions, indicating that the prior distributions were appropriate but that there is little additional information on these parameters in the data set. The posterior distribution of ν has a slightly lower mean than prior, implying a longer mean duration of carriage of 193 days (118, 397). The prior and posterior credible intervals still intersect to a large extent so there is not significant evidence of a departure from the prior based on the data.

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The last two parameters, α and ϕ , capture the difference between the susceptible and resistant strains in our model. The model predicts that in order to replicate observed incidence patterns recovery from asymptomatic cefixime-resistant gonorrhoea occurs $\alpha = 1.7$ (1.4, 2.4) times faster than asymptomatic cefixime-susceptible gonorrhoea, giving rise to a fitness cost. The model suggests a treatment failure proportion of $\phi = 84\%$ (55%, 100%) for resistant gonorrhoea treated with cefixime, so that resistance confers a fitness benefit in an environment in which cefixime is highly prescribed.

Beyond the marginal posterior distributions of the parameters described above, it is informative to study their posterior correlations. The pairwise posterior relationships between the ten parameters are depicted in Fig 3. Parameters σ , μ and ρ did not show a strong correlation with any parameters; as expected the short duration of the incubation, symptomatic and treatment stages of infection led to these parameters contributing relatively little to the dynamics of infection. The strongest correlation was found between ν and α , -0.87 (-0.89, -0.86), corresponding to the trade-off required to maintain the duration of carriage of resistant infection, which is equal to $1/(\alpha\nu)$. Parameters ν and ψ were also highly negatively correlated, -0.84 (-0.85, -0.82), which corresponds to the trade-off between duration of carriage, $1/\nu$, and the proportion of infections entering the carriage state, $(1 - \psi)$. The two negative correlations of both α and ψ with ν lead to a positive correlation between α and ψ .

Posterior predictive analysis

The total number of gonorrhoea cases in MSM observed by GUMCAD and the number of cefixime-resistant infections isolated in MSM by GRASP were compared with simulated datasets using parameters sampled from their posterior distributions. Fig 4 demonstrates that the simulated data closely emulate the real data. The real data are within the 95% predictive intervals at all time points, indicating a good fit of the model to the data [57].

The total number of cases of gonorrhoea observed by GUMCAD, and the number of cefixime-resistant cases observed by GRASP in 2015 [21] were not used in the model fitting process, and were used to provide an independent check of the model fit. Both data points are within the 95% probability intervals predicted by our model: 21,915 gonorrhoea diagnoses were recorded by GUMCAD, compared to 27,668 (18,144, 41,054) diagnoses predicted by the model; 10 cefixime-resistant cases were recorded by GRASP, compared to 4 (0, 14) cases predicted by the model. Our modelling suggests that in 2015 0.7% (0.4%, 1.1%) of MSM in England may be carriers of asymptomatic gonorrhoea.

Comparative analysis of basic reproduction numbers

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A key threshold in epidemic theory associates the persistence of disease in a population with a basic reproduction number greater than one [58]. Using Eq 1 we obtain a posterior estimate for the basic reproduction number for cefixime-susceptible infection of $R_0^{\text{sus}} = 1.18$ (1.09, 1.35), which suggests that the cefixime-susceptible strain of gonorrhea is expected to persist in the population without further intervention (Fig 5A). Under our hypothesis the basic reproduction number for cefixime-resistant gonorrhea depends on the frequency of cefixime prescription (Eq 2) and so can be considered as a function $R_0^{\text{res}}(\pi)$ where π is the proportion of gonorrhea diagnoses being treated by cefixime. In the two extreme cases when no cefixime is prescribed ($\pi = 0$, meaning that treatment is always effective) and only cefixime is prescribed ($\pi = 1$, meaning that only a proportion $1 - \phi$ of treatment is effective) we estimate a basic reproduction number for resistant gonorrhea of $R_0^{\text{res}}(0) = 0.73$ (0.62, 0.83) and $R_0^{\text{res}}(1) = 1.60$ (1.38, 1.90), respectively. At its height in 2008 the frequency of cefixime prescriptions was 70%, we estimate that at this time the basic reproduction number for resistant gonorrhea was $R_0^{\text{res}}(0.7) = 1.34$ (1.22, 1.51). The former estimate is ≤ 1 whereas the latter two are > 1 , which is consistent with the fact that between 2005 and 2010, when cefixime was often used to treat gonorrhea, resistance to cefixime increased whereas with the discontinuation of cefixime usage from 2011 resistance has decreased.

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We can estimate the frequency of cefixime prescriptions above which we expect the resistant strain to persist, corresponding to when the fitness benefit of cefixime resistance is greater than its fitness cost, by setting $R_0^{\text{res}}(\pi) = 1$ and solving for π in Eq 2. We denote this threshold $\pi^{\{R_0^{\text{res}}=1\}}$, and thus obtain a posterior estimate of $\pi^{\{R_0^{\text{res}}=1\}} = 0.31$ (0.26, 0.36) (Fig 5B). This result suggests that up to a quarter of gonorrhea treatments could be with cefixime monotherapy without causing a cefixime-resistant epidemic. Another important threshold is the level of cefixime prescriptions above which the resistant strain of gonorrhea is fitter than the susceptible strain. We denote this threshold $\pi^{\{R_0^{\text{sus}}=R_0^{\text{res}}\}}$. By setting $R_0^{\text{res}} = R_0^{\text{res}}$ and equating Eqs 1 and 2 we obtain a posterior estimate of $\pi^{\{R_0^{\text{sus}}=R_0^{\text{res}}\}} = 0.51$ (0.43, 0.62) (Fig 5C).

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Impact of cefixime usage on simulated resistance trends

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The basic reproduction numbers derived above are informative, but do not capture completely the complex dynamics of infection transmission that occur when accounting for stochasticity, competition between susceptible and resistant strains, and non-negligible fractions of the population becoming infected. To further study the impact of cefixime prescribing on the cefixime-resistant and susceptible epidemics, we performed stochastic model simulations over eight years from 2008 to 2015 using parameters drawn from their posterior distributions and examining scenarios with a constant frequency of cefixime prescriptions ranging from no use of cefixime ($\pi = 0$) to all gonorrhea cases being treated by cefixime ($\pi = 1$). Fig 6A shows that, when more than 35% of gonorrhea cases were treated with cefixime, there was a 95% probability that the resistant outbreak persisted in 2015. This is comparable to our estimate of $\pi^{\{R_0^{\text{res}}=1\}} = 0.31$ (0.26, 0.36) (Fig 5B), the level of prescriptions above which the fitness benefit of cefixime resistance is greater than the fitness cost.

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Fig 6B shows that, when more than 50% of gonorrhea cases were treated with cefixime, the simulated incidence of cefixime-susceptible infection began to fall, with the cefixime-resistant strain becoming more common. This supports our analytical estimate of $\pi^{\{R_0^{\text{sus}}=R_0^{\text{res}}\}} = 0.51$ (0.43, 0.62) (Fig 5C), the level of cefixime prescriptions above which the resistant strain becomes fitter than the susceptible strain. If cefixime were used to treat more than 60% of cases then the level of cefixime resistance would become greater than 50% at the end of the eight year simulation period and if cefixime were used to treat all cases resistance would be close to 100%.

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Discussion

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We have used mathematical modelling and Bayesian inference methods to uncover insights into the dynamics of cefixime resistance in gonorrhoea. We quantified both the fitness cost and fitness benefit of resistant strains, which allowed us to make predictions about the future prevalence of resistance as a function of how often cefixime is prescribed. Our results indicate that cefixime could be used to treat uncomplicated cases of gonorrhoea without incurring the risk of causing a resistant epidemic like the one that happened in 2007-2012, provided its frequency of use were controlled, enabling continued use of an 'abandoned' antibiotic. Our analysis suggests that cefixime could be used to treat up to 25% of cases, but this threshold should be used cautiously for reasons described below.

Our modelling approach requires making a number of assumptions, and it is important to consider their suitability. For example, it was assumed that all cefixime-susceptible infections were cured, regardless of which antibiotic was prescribed. The prescription data shows that between 2008 and 2015 the vast majority of non-cefixime prescriptions were for ceftriaxone, either alone or in combination with azithromycin, so the assumption of cure is reasonable given that ceftriaxone resistance reports remain sporadic in England. Our model implicitly assumes that there is no co-infection with both strains, and no evolution of resistance happening within-host. This simplification has been used in a number of other studies on the epidemiology of antimicrobial resistance [61, 62]. Ignoring within-host competition between resistant and susceptible strains following co-infection is justified here by the fact that both strains have low prevalence, making co-infection very unlikely. Within-host evolution of resistance was included in a recent gonorrhoea modelling study [67] but clearly this is a rare event which only increases by one the number of resistant infections, which we estimated to be already equal to 49 at the start of the epidemic simulation on the 1st January 2008.

Our model also makes assumptions concerning the cost and benefit of cefixime resistance. The fitness cost of the mutation conferring resistance is assumed to be constant over time, however compensatory mutations have been observed in other bacterial pathogens that reduce the initially high fitness cost of antibiotic resistance [59, 63]. It is clear from our analysis that there was a substantial fitness cost to cefixime resistance when the prescription protocol was changed in 2010, which is the reason why the resistance level subsequently fell. We cannot rule out that compensatory mutations took place after resistance initially emerged, but this would mean that the initial cost was even higher and in these conditions resistance would have been unlikely to emerge at all. Our formulation of the dynamics of the

fitness cost of resistance was via a reduction in the duration of asymptomatic carriage. In the absence 271
of evidence of the resistance mechanism the fitness cost could plausibly be modelled through reduced 272
transmissibility of the resistant strain [67], which would not affect our overall conclusions, in particular 273
regarding the basic reproduction number analysis and predictions of the impact of cefixime usage on 274
future resistance trends. 275

The ceftriaxone-azithromycin dual therapy is currently effective, but it represents a last resort, so that we 276
urgently need a strategy for what would be done if it stopped working. It is likely to be just a matter of 277
time before this happens, with the first reported failure of the dual therapy having occurred in 2015 [65]. 278
Resistance to azithromycin was detected in a recent outbreak that started in the North of England [5], 279
and is now reported in almost 10% of tested isolates [21]. Resistance to ceftriaxone remains rare, but 280
minimum inhibitory concentration (MIC) levels have been steadily increasing [66]. If alternative treatment 281
options could be used, even for a minority of cases, then it would delay and maybe even prevent the 282
emergence of resistance to the dual-therapy antibiotics by reducing the fitness benefit it would confer. 283

For some previously used antibiotics, such as penicillin or ciprofloxacin, significant levels of resistance 284
remain in the gonococcal population (24% and 39% in 2015, respectively [21]), so that they could 285
not be recommended even for a small fraction of cases. These antibiotics could be prescribed only if 286
drug-sensitivity could be quickly established, for example using real-time PCR assays [70, 71], or whole 287
genome sequencing [64, 72], which both remain experimental. In contrast, the fact that resistance to 288
cefixime has become very low in England (around 1% in 2015, [21]) makes it a prime candidate for return 289
into action without the need for case-by-case susceptibility testing. Perhaps the greatest threat posed by 290
this proposed strategy would be the evolution of compensatory mutations which could reduce the fitness 291
cost of resistance. As previously mentioned, compensatory mutations do not seem to have emerged 292
during the 2007-2012 cefixime-resistant epidemic, but if they did the acceptable prescribing proportion 293
would be lowered, and the probability of persistence of cefixime resistance increased. Therefore, a 294
redeployment of cefixime would require close monitoring of resistance trends in England [68] and 295
beyond [73, 74]. 296

Figures

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Fig 1. Usage and resistance of cefixime in England and Wales. The proportion of gonococcal isolates in GRASP resistant to cefixime over time is compared with the proportion of gonorrhoea diagnoses treated with cefixime. Dashed lines show dates of treatment guideline changes.

Fig 2. Flow diagram of model compartments with rates of transition between infection states. The left hand side represents infection with the cefixime-susceptible strains and the right hand side infection with the resistant strains. The two sides are symmetric with the exception of the two arrows highlighted in red which correspond to the cost and benefit of resistance.

Fig 3. Posterior distributions of parameters Diagonal plots show histograms of posterior distributions for all sampled parameters. Blue lines show prior distributions, red lines indicate posterior mean and 95% credible intervals. Plots below the diagonal show scatter plots based on 1,000 samples from the posterior, illustrating the relationships between pairs of estimated parameters. An orange background indicates a correlation higher than 0.8, a yellow background indicates a correlation between 0.5 and 0.8, a green background between 0.3 and 0.5, and a white background indicates a correlation less than 0.3. Plots above the diagonal show corresponding correlation coefficients with (95% CI).

Fig 4. Comparison of simulated and observed cases of gonorrhoea. Panel A shows total number of cases and panel B shows only the cefixime-resistant cases. Observed data are shown in orange, with the shaded area showing the 95% posterior predictive interval (based on 1,000 simulations using samples from posterior distribution).

Fig 5. A. histogram of posterior estimate of R_0^{sus} **B.** 95% credible interval of $R_0^{\text{res}}(\pi)$ against π with dashed lines showing 95% credible interval for $\pi^{\{R_0^{\text{res}}=1\}}$ **C.** histogram of posterior estimate of $\pi^{\{R_0^{\text{sus}}=R_0^{\text{res}}\}}$: the threshold of cefixime prescriptions above which $R_0^{\text{res}} > R_0^{\text{sus}}$

Fig 6. Incidence of gonorrhoea in 2015 based on simulations from 2004 to 2015 with varying levels of cefixime prescribing. A. incidence of the cefixime-resistant strain, red lines show 95% credible interval for $\pi^{\{R_0^{\text{res}}=1\}}$. B. incidence of the cefixime-susceptible strain, red lines show 95% credible interval for $\pi^{\{R_0^{\text{sus}}=R_0^{\text{res}}\}}$. Shaded areas show the 95% posterior predictive intervals (based on 1,000 simulations using samples from posterior distribution).

Supporting information

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S1 Appendix. Model equations and stochastic simulations. We developed a simple susceptible-
 infected-susceptible (SIS)-type stochastic compartmental model that describes the transmission and
 natural history of gonorrhoea, as illustrated in Fig 1 with parameters summarised in Table 1. The
 population was assumed to be closed due to the short time period under consideration. A deterministic
 version of the model is described by the following differential equations.

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$$\begin{aligned} \frac{dS(t)}{dt} = & -\theta \frac{I(t)}{N} S(t) + \nu(A_{\text{sus}}(t) + \alpha A_{\text{res}}(t)) \\ & + \rho(T_{\text{sus;cef}}(t) + T_{\text{sus;oth}}(t) + (1 - \phi)T_{\text{res;cef}}(t) + T_{\text{res;oth}}(t)) \end{aligned} \quad (\text{S1})$$

$$\frac{dU_s(t)}{dt} = \theta \frac{I_s(t)}{N} S(t) - \sigma U_s(t); s \in \{\text{sus}, \text{res}\} \quad (\text{S2})$$

$$\frac{dA_{\text{sus}}(t)}{dt} = (1 - \psi)\sigma U_{\text{sus}}(t) - \nu A_{\text{sus}}(t) \quad (\text{S3})$$

$$\frac{dA_{\text{res}}(t)}{dt} = (1 - \psi)\sigma U_{\text{res}}(t) - \alpha \nu A_{\text{res}}(t) + \phi \rho T_{\text{res;cef}}(t) \quad (\text{S4})$$

$$\frac{dE_s(t)}{dt} = \psi \sigma U_s(t) - \mu E_s(t); s \in \{\text{sus}, \text{res}\} \quad (\text{S5})$$

$$\frac{dT_{s;p}(t)}{dt} = \pi(t) \mu E_s(t) - \rho T_{s;p}(t); s \in \{\text{sus}, \text{res}\}, p \in \{\text{cef}, \text{oth}\} \quad (\text{S6})$$

We used a stochastic version of the model described in Eqs S1-S6. The process was initialised on 31 December 2007, with an initial number of asymptomatic infections $A_s(0)$ for $s \in \{\text{sus}, \text{res}\}$, and the remainder of the population being susceptible. Simulation proceeds through repeated iterations of the

steps below for each day. Transition variables d_1, \dots, d_{17} are drawn from the following distributions:

$$d_1, d_2 \sim \text{Multinom}\left(S(t), \theta \frac{I_{\text{sus}}(t)}{N}, \theta \frac{I_{\text{res}}(t)}{N}\right) \quad (\text{S7})$$

$$d_3, d_4 \sim \text{Multinom}(U_{\text{sus}}(t), \psi\sigma, (1 - \psi)\sigma) \quad (\text{S8})$$

$$d_5, d_6 \sim \text{Multinom}(U_{\text{res}}(t), \psi\sigma, (1 - \psi)\sigma) \quad (\text{S9})$$

$$d_7, d_8 \sim \text{Multinom}(E_{\text{sus}}(t), \pi(t)\mu, (1 - \pi(t))\mu) \quad (\text{S10})$$

$$d_9, d_{10} \sim \text{Multinom}(E_{\text{res}}(t), \pi(t)\mu, (1 - \pi(t))\mu) \quad (\text{S11})$$

$$d_{11} \sim \text{Binom}(A_{\text{sus}}(t), \nu) \quad (\text{S12})$$

$$d_{12} \sim \text{Binom}(A_{\text{res}}(t), \alpha\nu) \quad (\text{S13})$$

$$d_{13} \sim \text{Binom}(T_{\text{sus};\text{cef}}(t), \rho) \quad (\text{S14})$$

$$d_{14} \sim \text{Binom}(T_{\text{sus};\text{oth}}(t), \rho) \quad (\text{S15})$$

$$d_{15}, d_{16} \sim \text{Multinom}(T_{\text{res};\text{cef}}(t), \phi\rho, (1 - \phi)\rho) \quad (\text{S16})$$

$$d_{17} \sim \text{Binom}(T_{\text{res};\text{oth}}(t), \rho) \quad (\text{S17})$$

The compartments of the model are then updated as follows:

$$S(t+1) := S(t) - d_1 - d_2 + d_9 + d_{12} + d_{13} + d_{14} + d_{15} + d_{17} \quad (\text{S18})$$

$$U_{\text{sus}}(t+1) := U_{\text{sus}}(t) + d_1 - d_3 - d_4 \quad (\text{S19})$$

$$U_{\text{res}}(t+1) := U_{\text{res}}(t) + d_2 - d_5 - d_6 \quad (\text{S20})$$

$$E_{\text{sus}}(t+1) := E_{\text{sus}}(t) + d_3 - d_7 - d_8 \quad (\text{S21})$$

$$E_{\text{res}}(t+1) := E_{\text{res}}(t) + d_5 - d_{10} - d_{11} \quad (\text{S22})$$

$$A_{\text{sus}}(t+1) := A_{\text{sus}}(t) + d_4 - d_9 \quad (\text{S23})$$

$$A_{\text{res}}(t+1) := A_{\text{res}}(t) + d_6 - d_{12} + d_{16} \quad (\text{S24})$$

$$T_{\text{sus};\text{cef}}(t+1) := T_{\text{sus};\text{cef}}(t) + d_7 - d_{13} \quad (\text{S25})$$

$$T_{\text{sus};\text{oth}}(t+1) := T_{\text{sus};\text{oth}}(t) + d_8 - d_{14} \quad (\text{S26})$$

$$T_{\text{res};\text{cef}}(t+1) := T_{\text{res};\text{cef}}(t) + d_{10} - d_{15} - d_{16} \quad (\text{S27})$$

$$T_{\text{res};\text{oth}}(t+1) := T_{\text{res};\text{oth}}(t) + d_{11} - d_{17} \quad (\text{S28})$$

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References

1. Public Health England. Infection Report: HIV - STIs. Public Health England; 2015. 22.
2. Lewis DA. The Gonococcus fights back: is this time a knock out? *Sexually Transmitted Infections*. 2010;86(6):415–421. doi:10.1136/sti.2010.042648.
3. Grad YH, Kirkcaldy RD, Trees D, Dordel J, Harris SR, Goldstein E, et al. Genomic epidemiology of *Neisseria gonorrhoeae* with reduced susceptibility to cefixime in the USA: A retrospective observational study. *The Lancet Infectious Diseases*. 2014;14(3):220–226. doi:10.1016/S1473-3099(13)70693-5.
4. Bignell C, FitzGerald M. UK national guideline for the management of gonorrhoea in adults, 2011. *International Journal of STD & AIDS*. 2011;22(10):541–547. doi:10.1258/ijsa.2011.011267.
5. Chisholm SA, Wilson J, Alexander S, Tripodo F, Al-Shahib A, Schaefer U, et al. An outbreak of high-level azithromycin resistant *Neisseria gonorrhoeae* in England. *Sexually Transmitted Infections*. 2015;doi:10.1136/sextrans-2015-052312.
6. GRASP Steering Group. Annual Report 2000. London: Health Protection Agency; 2001.
7. GRASP Steering Group. The Gonococcal Resistance to Antimicrobials Surveillance Programme 2001. London: Health Protection Agency; 2002.
8. GRASP Steering Group. The Gonococcal Resistance to Antimicrobials Surveillance Programme (GRASP) 2002. London: Health Protection Agency; 2003.
9. GRASP Steering Group. Annual Report Year 2003. London: Health Protection Agency; 2004.
10. GRASP Steering Group. The Gonococcal Resistance to Antimicrobials Surveillance Programme (GRASP) Year 2004 report. London: Health Protection Agency; 2005.
11. GRASP Steering Group. The Gonococcal Resistance to Antimicrobials Surveillance Programme (GRASP) 2005 report. London: Health Protection Agency; 2006.
12. GRASP Steering Group. Annual Report 2006. London: Health Protection Agency; 2007.
13. GRASP Steering Group. Annual Report 2007. London: Health Protection Agency; 2008.
14. GRASP Steering Group. The Gonococcal Resistance to Antimicrobials Surveillance Programme (GRASP) Year 2008 report. London: Health Protection Agency; 2009.

15. GRASP Steering Group. Health Protection Report: Gonococcal Resistance to Antimicrobials Surveillance Programme in England and Wales (GRASP) report of 2009 data. London: Health Protection Agency; 2010. 34.
16. GRASP Steering Group. GRASP 2010 Report. London: Health Protection Agency; 2011.
17. GRASP Steering Group. GRASP 2011 Report: The Gonococcal Resistance to Antimicrobials Surveillance Programme. London: Health Protection Agency; 2012.
18. GRASP Steering Group. GRASP 2012 Report: The Gonococcal Resistance to Antimicrobials Surveillance Programme. London: Public Health England; 2013.
19. GRASP Steering Group. GRASP 2013 Report The Gonococcal Resistance to Antimicrobials Surveillance Programme (England and Wales). 2014;.
20. GRASP Steering Group. The Gonococcal Resistance to Antimicrobials Surveillance Programme (GRASP) 2014 report. London: Public Health England; 2015.
21. GRASP Steering Group. Surveillance of antimicrobial resistance in *Neisseria gonorrhoeae* Key findings from the Gonococcal Resistance to Antimicrobials Surveillance Programme (GRASP). 2016;(October). doi:2016433.
22. Bignell C. National Guideline on the Diagnosis and Treatment of Gonorrhoea in Adults 2005; 2005.
23. World Health Organization. Global action plan to control the spread and impact of antimicrobial resistance in *Neisseria gonorrhoeae*. World Health Organization; 2012.
24. Ison C, Hussey J, Sankar K, Evans J, Alexander S. Gonorrhoea treatment failures to cefixime and azithromycin in England, 2010. *Eurosurveillance*. 2011;16(14):1–4.
25. Forsyth S, Penney P, Rooney G. Cefixime-resistant *Neisseria gonorrhoeae* in the UK: a time to reflect on practice and recommendations. *International Journal of STD and AIDS*. 2011;22(5):296–297. doi:10.1258/ijisa.2009.009191.
26. Grad YH, Goldstein E, Lipsitch M, White PJ. Improving Control of Antibiotic-Resistant Gonorrhea by Integrating Research Agendas Across Disciplines: Key Questions Arising From Mathematical Modeling. *Journal of Infectious Diseases*. 2016;213(6):883–890. doi:10.1093/infdis/jiv517.

27. Bissessor M, Tabrizi SN, Fairley CK, Danielewski J, Whitton B, Bird S, et al. Differing *Neisseria gonorrhoeae* bacterial loads in the pharynx and rectum in men who have sex with men: Implications for gonococcal detection, transmission, and control. *Journal of Clinical Microbiology*. 2011;49(12):4304–4306. doi:10.1128/JCM.05341-11.
28. Kent CK, Chaw JK, Wong W, Liska S, Gibson S, Hubbard G, et al. Prevalence of rectal, urethral, and pharyngeal chlamydia and gonorrhea detected in 2 clinical settings among men who have sex with men: San Francisco, California, 2003. *Clinical Infectious Diseases*. 2005;41(1):67–74. doi:10.1086/430704.
29. Norris Turner A, Carr Reese P, Ervin M, Davis JA, Fields KS, Bazan JA. HIV, rectal chlamydia and rectal gonorrhea in men who have sex with men attending an STD clinic in a midwestern US city. *Sex Transm Dis*. 2013;40(6). doi:10.1097/OLQ.0b013e31828fd163.HIV.
30. Didelot X, Dordel J, Whittles L, Collins C, Bilek N, Bishop C, et al. Genomic analysis and comparison of two gonorrhoea outbreaks. *mBio*. 2016;7:e00525-16.
31. Hosenfeld CB, Workowski Ka, Berman S, Zaidi A, Dyson J, Mosure D, et al. Repeat infection with Chlamydia and gonorrhea among females: a systematic review of the literature. *Sexually transmitted diseases*. 2009;36(8):478–489. doi:10.1097/OLQ.0b013e3181a2a933.
32. Schmidt K, Schneider H, Lindstrom J, Boslego J, Warren R, Van de Verg L, et al. Homologous Gonococci in Male Volunteers. *Sexually Transmitted Diseases*. 2001; p. 555–564.
33. Office for National Statistics. United Kingdom population mid-year estimate; 2016. Available from: www.ons.gov.uk/peoplepopulationandcommunity/populationandmigration/populationestimates/timeseries/ukpop/pop.
34. Mercer CH, Tanton C, Prah P, Erens B, Sonnenberg P, Clifton S, et al. Changes in sexual attitudes and lifestyles in Britain through the life course and over time : findings from the National Surveys of Sexual Attitudes and Lifestyles (Natsal). *The Lancet*. 2013;382(9907):1781–1794. doi:10.1016/S0140-6736(13)62035-8.
35. Prah P, Hickson F, Bonell C, McDaid LM, Johnson AM, Wayal S, et al. Men who have sex with men in Great Britain: comparing methods and estimates from probability and convenience sample surveys. *Sexually Transmitted Infections*. 2016; p. sextrans—2015—052389. doi:10.1136/sextrans-2015-052389.

36. Marcus U, Hickson F, Weatherburn P, Schmidt AJ. Estimating the size of the MSM populations for 38 European countries by calculating the survey-surveillance discrepancies (SSD) between self-reported new HIV diagnoses from the European MSM internet survey (EMIS) and surveillance-reported HIV diagnoses am. *BMC public health*. 2013;13(1):919. doi:10.1186/1471-2458-13-919.
37. Diekmann O, Heesterbeek JAP, Metz JAJ. On the definition and the computation of the basic reproduction ratio R_0 in models for infectious diseases in heterogeneous populations. *Journal of Mathematical Biology*. 1990;28(4):365–382. doi:10.1007/BF00178324.
38. Del Moral P, Doucet A, Jasra A. Sequential Monte Carlo samplers. *Journal of the Royal Statistical Society Series B: Statistical Methodology*. 2006;68(3):411–436. doi:10.1111/j.1467-9868.2006.00553.x.
39. Andrieu C, Doucet A, Holenstein R. Particle Markov chain Monte Carlo methods. *Journal of the Royal Statistical Society Series B-Statistical Methodology*. 2010;72(3):269–342. doi:10.1111/j.1467-9868.2009.00736.x.
40. King AA, Nguyen D, Ionides EL. Statistical Inference for Partially Observed Markov Processes via the R Package pomp. *Journal of Statistical Software*. 2016;69(12):1–43. doi:10.18637/jss.v069.i12.
41. Plummer M, Best N, Cowles K, Vines K. CODA: convergence diagnosis and output analysis for MCMC. *R News*. 2006;6(March):7–11. doi:10.1159/000323281.
42. Gelman A, Rubin DB. Inference from Iterative Simulation Using Multiple Sequences. *Statistical Science*. 1992;7(4):457–472.
43. Brooks SP, Gelman AG. General methods for monitoring convergence of iterative simulations. *Journal of computational and graphical statistics*. 1998;7(4):434–455. doi:10.2307/1390675.
44. Public Health England. STI diagnoses & rates in England by gender, 2006 - 2015; 2015. Available from: https://www.gov.uk/government/uploads/system/uploads/attachment_data/file/534517/2015_Table_1_STI_diagnoses_rates_in_England_by_gender_2006-2015.pdf.
45. Harrison WO, Hooper RR, Wiesner PJ, Campbell AF, Karney WW, Reynolds GH, et al. A trial of minocycline given after exposure to prevent gonorrhoea. *N Engl J Med*. 1979;300(19):1074–1078. doi:10.1056/NEJM197905103001903.

46. McCutchan JA. Epidemiology of venereal urethritis: comparison of gonorrhoea and nongonococcal urethritis. *Reviews of infectious diseases*. 1984;6(5):669–688.
47. Korenromp EL, Sudaryo MK, de Vlas SJ, Gray RH, Sewankambo NK, Serwadda D, et al. What proportion of episodes of gonorrhoea and chlamydia becomes symptomatic? *International journal of STD & AIDS*. 2002;13(2):91–101. doi:10.1258/0956462021924712.
48. Barbee LA, Khosropour CM, Dombrowski JC, Manhart LE, Golden MR. An estimate of the proportion of symptomatic gonococcal, chlamydial and non-gonococcal non-chlamydial urethritis attributable to oral sex among men who have sex with men: a case-control study. *Sexually transmitted infections*. 2016;92(2):155–160. doi:10.1136/sextrans-2015-052214.
49. Jin F, Prestage GP, Mao L, Kippax SC, Pell CM, Donovan B, et al. Incidence and risk factors for urethral and anal gonorrhoea and chlamydia in a cohort of HIV-negative homosexual men: the Health in Men Study. *Sexually Transmitted Infections*. 2007;83(2):113–119. doi:10.1136/sti.2006.021915.
50. Bachmann LH, Johnson RE, Cheng H, Markowitz LE, Papp JR, Edward IWH. Nucleic acid amplification tests for diagnosis of *Neisseria gonorrhoeae* oropharyngeal infections. *Journal of Clinical Microbiology*. 2009;47(4):902–907. doi:10.1128/JCM.01581-08.
51. Handsfield HH, Lipman TO, Harnisch JP, Tronca E, Holmes KK. Asymptomatic Gonorrhoea in Men. *New England Journal of Medicine*. 1974;290(3):117–123.
52. Yorke J, Hethcote H, Nold A. Dynamics and control of the transmission of gonorrhoea.. vol. 5; 1978.
53. Garnett GP, Mertz KJ, Finelli L, Levine WC, St Louis ME. The transmission dynamics of gonorrhoea: modelling the reported behaviour of infected patients from Newark, New Jersey. *Philosophical transactions of the Royal Society of London Series B, Biological sciences*. 1999;354(1384):787–97. doi:10.1098/rstb.1999.0431.
54. White PJ, Ward H, Cassell JA, Mercer CH, Garnett GP. Vicious and virtuous circles in the dynamics of infectious disease and the provision of health care: gonorrhoea in Britain as an example. *The Journal of infectious diseases*. 2005;192(5):824–836. doi:10.1086/432004.
55. Fairley CK, Chen MY, Bradshaw CS, Tabrizi SN. Is it time to move to nucleic acid amplification tests screening for pharyngeal and rectal gonorrhoea in men who have sex with men to improve gonorrhoea control? *Sexual Health*. 2011;8(1):9–11. doi:10.1071/SH10134.

56. Fairley CK, Hocking JS, Zhang L, Chow EPF. Frequent Transmission of Gonorrhoea in Men Who Have Sex with Men. *Emerging Infectious Diseases*. 2017;23(1):102 – 104.
57. Gelman A, Meng XL, Stern H. Posterior predictive assessment of model fitness via realized discrepancies. Vol.6, No.4. *Statistica Sinica*. 1996;6(4):733–807. doi:10.1.1.142.9951.
58. Dietz K. The estimation of the basic reproduction number for infectious diseases. *Statistical Methods in Medical Research*. 1993;2(1):23–41. doi:10.1177/096228029300200103.
59. Melnyk AH, Wong A, Kassen R. The fitness costs of antibiotic resistance mutations. *Evolutionary Applications*. 2015;8(3):273–283. doi:10.1111/eva.12196.
60. Savage EJ, Mohammed H, Leong G, Duffell S, Hughes G. Improving surveillance of sexually transmitted infections using mandatory electronic clinical reporting: the genitourinary medicine clinic activity dataset, England, 2009 to 2013. *Euro surveillance : bulletin Europeen sur les maladies transmissibles = European communicable disease bulletin*. 2014;19(48):20981. doi:10.2807/1560-7917.ES2014.19.48.20981.
61. Lipsitch M, Bergstrom CT, Levin BR. The epidemiology of antibiotic resistance in hospitals: paradoxes and prescriptions. *Proceedings of the National Academy of Sciences of the United States of America*. 2000;97(4):1938–1943. doi:10.1073/pnas.97.4.1938.
62. Kouyos RD, zur Wiesch PA, Bonhoeffer S. On being the right size: The impact of population size and stochastic effects on the evolution of drug resistance in hospitals and the community. *PLoS Pathogens*. 2011;7(4). doi:10.1371/journal.ppat.1001334.
63. Didelot X, Walker AS, Peto TE, Crook DW, Wilson DJ. Within-host evolution of bacterial pathogens. *Nat Rev Micro*. 2016;14(3):150–162. doi:10.1038/nrmicro.2015.13.
64. Grad YH, Harris SR, Kirkcaldy RD, Green AG, Marks DS, Bentley SD, et al. Genomic epidemiology of gonococcal resistance to extended-spectrum cephalosporins, macrolides, and fluoroquinolones in the United States, 2000–2013. *J Infect Dis*. 2016;214(10):1579–87. doi:10.1093/infdis/jiw420.
65. Fifer H, Natarajan U, Jones L, Alexander S, Hughes G, Golparian D, et al. Failure of Dual Antimicrobial Therapy in Treatment of Gonorrhoea. *The New England Journal of Medicine*. 2016;374(25):2504–2506. doi:10.1056/NEJMc1514294.

66. Town K, Obi C, Quaye N, Chisholm S, Hughes G. Drifting towards ceftriaxone treatment failure in gonorrhoea: risk factor analysis of data from the Gonococcal Resistance to Antimicrobials Surveillance Programme in England and Wales. *Sexually Transmitted Infections*. 2016; p. sextrans–2016–052583. doi:10.1136/sextrans-2016-052583.
67. Fingerhuth SM, Bonhoeffer S, Low N, Althaus CL. Antibiotic-Resistant *Neisseria gonorrhoeae* Spread Faster with More Treatment, Not More Sexual Partners. *PLOS Pathogens*. 2016;12(5):e1005611. doi:10.1371/journal.ppat.1005611.
68. Ison CA, Town K, Obi C, Chisholm S, Hughes G, Livermore DM, et al. Decreased susceptibility to cephalosporins among gonococci: Data from the Gonococcal Resistance to Antimicrobials Surveillance Programme (GRASP) in England and Wales, 2007-2011. *The Lancet Infectious Diseases*. 2013;13(9):762–768. doi:10.1016/S1473-3099(13)70143-9.
69. Johnson LF, Dorrington RE, Bradshaw D. The role of immunity in the epidemiology of gonorrhoea, chlamydial infection and trichomoniasis: insights from a mathematical model. *Epidemiology and Infection*. 2011;139(12):1875–1883. doi:10.1017/S0950268811000045.
70. Pond MJ, Hall CL, Miari VF, Cole M, Laing KG, Jagatia H, et al. Accurate detection of *Neisseria gonorrhoeae* ciprofloxacin susceptibility directly from genital and extragenital clinical samples: Towards genotype-guided antimicrobial therapy. *Journal of Antimicrobial Chemotherapy*. 2016;71(4):897–902. doi:10.1093/jac/dkv432.
71. Buckley C, Trembizki E, Donovan B, Chen M, Freeman K, Guy R, et al. Real-time PCR detection of *Neisseria gonorrhoeae* susceptibility to penicillin. *Journal of Antimicrobial Chemotherapy*. 2016;(July):dkw291. doi:10.1093/jac/dkw291.
72. Eyre DW, De Silva D, Cole K, Peters J, Cole MJ, Grad YH, et al. WGS to predict antibiotic MICs for *Neisseria gonorrhoeae*. *Journal of Antimicrobial Chemotherapy*. 2017;doi:10.1093/jac/dkx067.
73. Kirkcaldy RD, Hook EW, Olusegun O, del Rio C, Kubin G, Zenilman JM, et al. Trends in *Neisseria gonorrhoeae* susceptibility to cephalosporins in the United States, 2006–2014. *Jama*. 2015;314(17):1869–1871. doi:10.1038/nbt.3121.ChIP-nexus.
74. Kirkcaldy RD, Schlanger K, Papp JR, Torrone EA. Considerations for Strengthening Surveillance of *Neisseria gonorrhoeae* Antimicrobial Resistance and Interpreting Surveillance Data. *Sex Transm Dis*. 2017;44(3):154–156.









