

Antibiotic resistance detection is essential for gonorrhoea point-of-care testing: A mathematical modelling study

Stephanie M. Fingerhuth^{1,2,*}, Nicola Low², Sebastian Bonhoeffer¹, Christian L. Althaus²

Abstract

Antibiotic resistance is threatening to make gonorrhoea untreatable. Point-of-care (POC) tests that detect resistance promise individually tailored treatment, but might lead to more treatment and higher levels of resistance. We investigate the impact of POC tests on antibiotic-resistant gonorrhoea. We used data about the prevalence and incidence of gonorrhoea in men who have sex with men (MSM) and heterosexual men and women (HMW) to calibrate a mathematical gonorrhoea transmission model. With this model, we simulated four clinical pathways for the diagnosis and treatment of gonorrhoea: POC test with (POC + R) and without (POC – R) resistance detection, culture, and nucleic acid amplification tests (NAATs). We calculated the proportion of resistant infections, cases averted after 5 years, and compared how fast resistant infections spread in the populations. The proportion of resistant infections after 30 years is lowest for POC + R (median MSM: 0.18%, HMW: 0.12%), and increases for culture (MSM: 1.19%, HMW: 0.13%), NAAT (MSM: 100%, HMW: 99.27%), and POC – R (MSM: 100%, HMW: 99.73%). NAAT leads to 36 366 (median MSM) and 1 228 (median HMW) observed cases after 5 years. When compared with NAAT, POC + R results in most cases averted after 5 years (median MSM: 3 353, HMW: 118 per 100 000 persons). POC tests that detect resistance with intermediate sensitivity slow down resistance spread more than NAAT. POC tests with very high sensitivity for the detection of resistance are needed to slow down resistance spread more than using culture. POC with high sensitivity to detect antibiotic resistance can keep gonorrhoea treatable longer than culture or NAAT. POC tests without reliable resistance detection should not be introduced because they can accelerate the spread of antibiotic-resistant gonorrhoea.

Keywords

gonorrhoea — bacterial drug resistance — point-of-care testing — mathematical model — sexually transmitted infection — epidemiology

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Introduction

Antibiotic resistance is a major challenge for the management of gonorrhoea globally: extended-spectrum cephalosporins are the last antibiotic class remaining for empirical treatment of gonorrhoea [1, 2], and 42 countries have already reported *Neisseria gonorrhoeae* strains with decreased susceptibility against them [2]. The first strain with high-level resistance to recommended combination therapy with ceftriaxone and azithromycin was recently described [3]. With an estimated 78 million new gonorrhoea cases each year [4], new control strategies are urgently needed before gonorrhoea becomes untreatable.

Conventional diagnostic tests for gonorrhoea, nucleic acid amplification tests (NAATs) and culture, are not sufficient to control antibiotic resistance. Commercially available NAATs, the most commonly used diagnostic gonorrhoea tests in high income countries, cannot detect

antibiotic resistance [5, 6]. Culture of *N. gonorrhoeae* can be used to determine antibiotic resistance profiles, but reliable results depend on stringent collection and transport of specimens [7]. Both tests need several days to deliver results in routine use. While symptomatic gonorrhoea patients usually receive empirical treatment at their first visit, asymptomatic patients might have to return for treatment. Loss to follow up and further spread of resistant infections can result.

Point-of-care (POC) tests promise to help control antibiotic resistance [8]. POC tests provide results rapidly and allow informed clinical decisions about treatment at the first visit of a patient. POC tests therefore reduce the time to treatment and avoid loss to follow up. A modelling study suggested that POC tests can reduce gonorrhoea prevalence if no antibiotic resistance is present in the population [9]. Though not yet commercially

available [8], POC tests that detect resistance promise to reduce the use of antibiotics [10] and to spare last-line antibiotics through individually tailored treatment [11, 12]. One modelling study illustrated that individualised treatment could slow down the spread of resistance as much as combination therapy [13]. However, reduced time to treatment and increased follow up with POC tests might increase the rate of gonorrhoea treatment. Since higher treatment rates can lead to faster spread of resistance [14, 15], POC tests might increase resistance levels. We extended a previously developed mathematical model of gonorrhoea transmission [15] to compare the effects of current conventional tests, culture and NAAT, with POC tests that reduce time to treatment and loss to follow up. We investigated the potential impact of POC tests on resistance and on the number of gonorrhoea cases for a population at high risk of infection [16], men who have sex with men (MSM), and a population at lower risk of infection, heterosexual men and women (HMW).

Methods

We developed a mathematical model that describes transmission of antibiotic-sensitive and -resistant gonorrhoea, clinical pathways for diagnostic testing with culture, NAAT or POC, and treatment with first- and second-line antibiotics (Supplementary Material: Section Model). Here we describe the model focusing on testing and treatment of gonorrhoea (Fig. 1, Table 1).

Basic model structure

The model is based on our previously published compartmental model of gonorrhoea transmission and resistance spread [15]. The model describes a population with two sexual activity classes $i \in C$, where $C = \{L, H\}$ indicates that there are two sexual activity classes L and H with low and high partner change rates. The model incorporates sexual mixing between the sexual activity classes, sexual behaviour change, migration in and out of the population, and gonorrhoea transmission. Individuals in the population can be susceptible to infection, S_i , infected with antibiotic-sensitive gonorrhoea, I_{Sen_i} , infected with gonorrhoea resistant to the first-line antibiotic, I_{Res_i} , or infected with gonorrhoea resistant to the first-line antibiotic and waiting for re-treatment, W_i . Depending on the parameters for sexual behaviour, transmission, and gonorrhoea natural history (Supplementary Material: Table S2), the model describes a population of men who have sex with men (MSM) or heterosexual men and women (HMW).

Gonorrhoea testing and treatment

Antibiotic-sensitive gonorrhoea

Individuals infected with antibiotic-sensitive gonorrhoea, I_{Sen_i} , (Fig. 1, left) can recover spontaneously at rate ν or seek care. Symptomatic care-seekers receive treatment

Table 1. Gonorrhoea testing and treatment parameters and their default values. Unless a value is set by definition, all values listed are default values and are varied in sensitivity analyses. Baseline: resistance-free scenario (corresponds to scenario where culture or nucleic acid amplification test (NAAT) is used; ξ_R can take any value since there is no resistance to detect). Culture, NAAT, Point-of-care (POC) with resistance detection (POC+R) and without resistance detection (POC-R) refer to scenarios after resistance is introduced. Sources for parameters: ¹Derived, ²[17], ³Assumption, ⁴by definition, ⁵[18, 19], ⁶[20].

Parameter	Description (unit)	Baseline	Culture	NAAT	POC+R	POC-R
τ_S	Rate at which symptomatic individuals seek care (y^{-1})	variable ¹	see Baseline	see Baseline	see Baseline	see Baseline
τ_A	Rate at which asymptomatic individuals seek care (y^{-1})	variable ¹	see Baseline	see Baseline	see Baseline	see Baseline
ξ_G	Test sensitivity to detect gonorrhoea	99% ²	see Baseline	see Baseline	see Baseline	see Baseline
ξ_R	Test sensitivity to detect resistance against the first-line antibiotic	any value	99% ³	0% ⁴	99% ³	0% ⁴
η_1, η_2	Efficacy of first-line (1) or second-line (2) antibiotic	99% ⁵	see Baseline	see Baseline	see Baseline	see Baseline
δ	Average time after test individuals return for treatment (days)	7 ⁶	see Baseline	see Baseline	0 ⁴	0 ⁴
$1/\omega$	Average time individuals with resistant gonorrhoea wait for re-treatment (days)	7 ³	see Baseline	see Baseline	see Baseline	see Baseline
λ_A	Fraction of asymptomatic individuals who return for treatment	90% ³	see Baseline	see Baseline	100% ⁴	100% ⁴
λ_S	Fraction of symptomatic individuals who remain symptomatic after failed treatment	90% ³	see Baseline	see Baseline	see Baseline	see Baseline
ψ	Fraction of successfully treated individuals who were symptomatic at baseline	60% ⁶	—	—	—	—

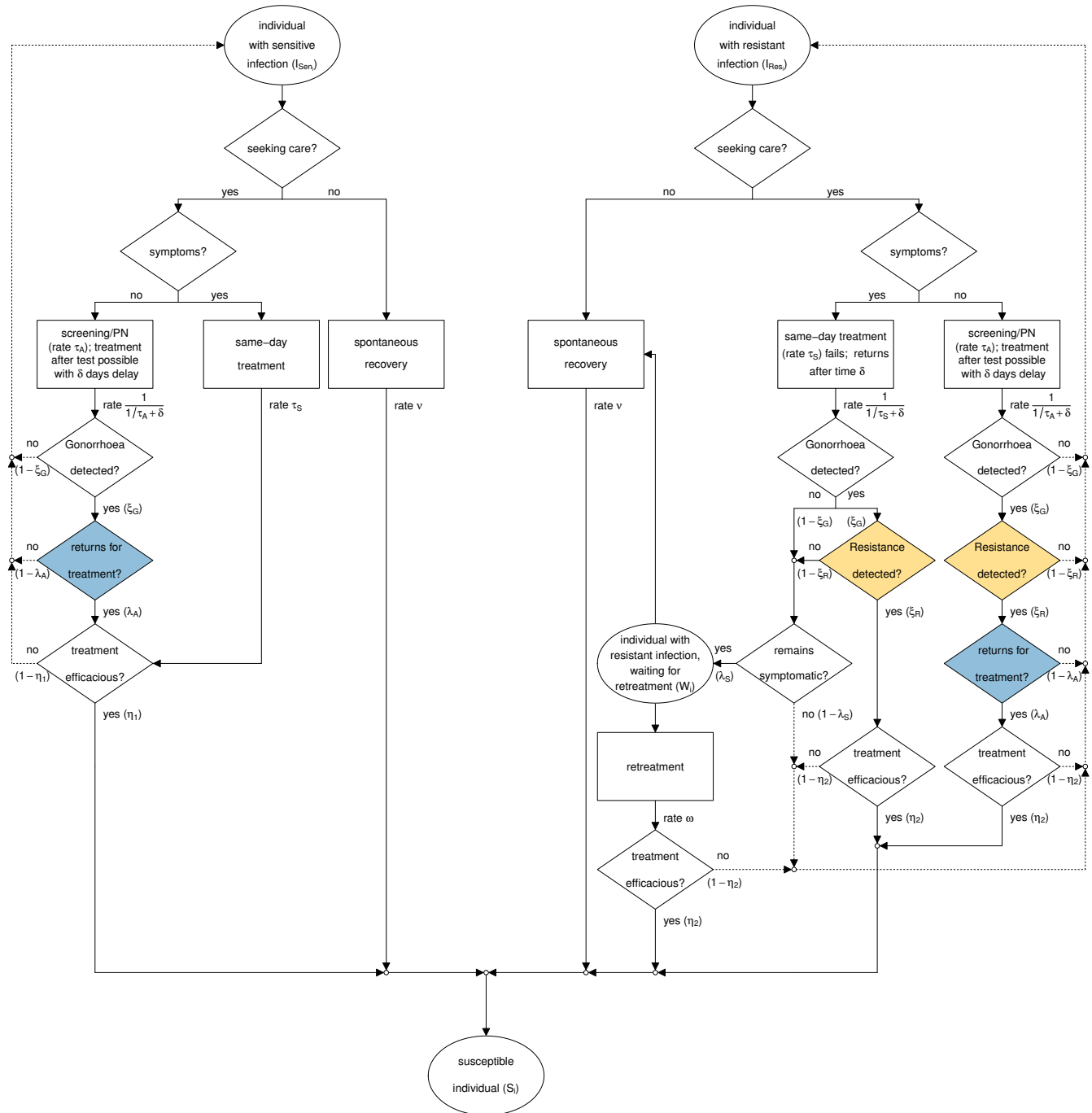


Figure 1. Testing and treatment of gonorrhoea infections. Dashed arrows indicate that individuals remain infected. In the nucleic acid amplification (NAAT) and point-of-care without resistance detection (POC – R) scenario, “Resistance detected?” (yellow) defaults to “no”. In all point-of-care scenarios, “returns for treatment?” (blue) defaults to “yes”. In the culture scenario, the flowchart is followed as shown. PN: partner notification.

on the same day at rate τ_S . Asymptomatic care-seekers, i.e. those who are screened for gonorrhoea or were notified through an infected partner, are tested at rate τ_A . Gonorrhoea is detected with sensitivity ξ_G . On average, a fraction λ_A of asymptomatic individuals returns for treatment after δ days. The treatment rate for asymptomatic individuals is approximated by $\frac{1}{1/\tau_A + \delta}$, the inverse of the average time until individuals are tested, $1/\tau_A$, and the time until they return for treatment, δ . Both symptomatic and asymptomatic individuals are treated with a first-line antibiotic that has treatment efficacy η_1 . We assumed that individuals whose treatment was inefficacious remain infected and do not seek care again immediately. This assumption reflects the notion that treatment failure of antibiotic-sensitive gonorrhoea is most likely to occur in pharyngeal infections, which are usually asymptomatic [21].

Antibiotic-resistant gonorrhoea

Individuals infected with gonorrhoea resistant to the first-line antibiotic, I_{Res} , (Fig. 1, right) can also recover spontaneously at rate ν . Asymptomatic care-seekers that return for treatment (fraction λ_A) receive treatment with the second-line antibiotic at rate $\frac{1}{1/\tau_A + \delta}$ if both gonorrhoea (sensitivity ξ_G) and resistance (sensitivity ξ_R) are detected. Symptomatic care-seekers receive the first-line antibiotic as treatment on the same day, but remain infected due to resistance and return for treatment after δ days. At their second visit, symptomatic care-seekers receive the second-line antibiotic if both gonorrhoea (sensitivity ξ_G) and resistance (sensitivity ξ_R) are detected. If either test fails, they do not receive the second-line antibiotic. If they remain symptomatic (fraction λ_S), they wait for re-treatment in compartment W_i , where they either receive re-treatment with the second-line antibiotic at rate ω or recover spontaneously at rate ν . The assumption that re-treatment occurs with the second-line antibiotic follows recommendations from the World Health Organization (WHO) [16] and the Centers for Disease Control (CDC) [22] to obtain a specimen for culture-based antibiotic resistance testing at a patient's second visit. The second-line antibiotic has efficacy η_2 ; individuals whose treatment is inefficacious remain infected and can recover spontaneously or seek care at a later point. De novo resistance to the first-line antibiotic or resistance to the second-line antibiotic are not considered in the model.

Testing scenarios

The model allowed us to simulate clinical pathways for gonorrhoea detection with culture, NAAT, and POC tests by adapting the parameters δ , λ_A , and ξ_R (Table 2). For culture, test results are not available immediately ($\delta_{\text{culture}} > 0$), resistance can be detected ($\xi_{R, \text{culture}} > 0$), and asymptomatic infected individuals might not return for treatment ($\lambda_{A, \text{culture}} < 1$). For NAAT, test results are

not available immediately ($\delta_{\text{NAAT}} > 0$), resistance cannot be detected ($\xi_{R, \text{NAAT}} = 0$), and asymptomatic infected individuals might not return for treatment ($\lambda_{A, \text{NAAT}} < 1$). For POC, test results are available immediately ($\delta_{\text{POC}} = 0$), all individuals are followed up ($\lambda_{A, \text{POC}} = 1$), and thus all individuals are treated at the first visit. We explore the impact of a POC test with ($\xi_{R, \text{POC}} > 0$, POC+R) and without resistance detection ($\xi_{R, \text{POC}} = 0$, POC-R); we use the term ‘‘POC’’ alone when $\xi_{R, \text{POC}}$ is variable.

Table 2. Conditions on parameters for different testing scenarios. NAAT: nucleic acid amplification test, POC: point-of-care test (with or without resistance detection), POC+R: POC test with resistance detection, POC-R: POC test without resistance detection.

	δ	λ_A	ξ_R
Culture	> 0	< 1	> 0
NAAT	> 0	< 1	$= 0$
POC	$= 0$	$= 1$	≥ 0
POC+R	$= 0$	$= 1$	> 0
POC-R	$= 0$	$= 1$	$= 0$

Impact measures

We evaluated the impact of a testing scenario by calculating the proportion of resistant infections among all infections, observed cases averted, and the rate at which resistance spreads, compared with another testing scenario. We measured the proportion of resistant infections up to 30 years after introduction of resistance into the resistance-free baseline scenario. If applicable, we also calculated the time until resistance levels reached 5%, the level above which an antibiotic should not be used for empirical gonorrhoea treatment [18]. We defined observed cases averted as the difference between the cumulative incidence of observed (i.e. diagnosed and successfully treated at baseline; fraction ϕ [15]) cases using NAAT and the cumulative incidence of observed cases using culture or POC tests. We calculated the observed cases averted 5 years after the introduction of resistance. The rate at which resistance spreads describes how fast resistant infections replace sensitive infections in a human population [15]. We calculated the ratio of the rate of resistance spread between POC with different test sensitivities to detect resistance ($\xi_{R, \text{POC}}$) and culture or NAAT scenarios (Supplementary Material: Section Rate of resistance spread and ratio of resistance spread). If the ratio of the rate of resistance spread is > 1 , resistance spreads faster when using POC tests compared with other tests. If the ratio is < 1 , resistance spreads slower when using POC tests compared with other tests.

Parameters

We used the parameters describing sexual behaviour, gonorrhoea transmission, natural history, and treatment from our previous model [15]. There, we estimated sexual behaviour parameters from the second British National

Survey of Sexual Attitudes and Lifestyles (Natsal-2), which is a nationally representative population-based survey [23]. We calibrated all other parameters to yield prevalence and incidence rates within empirically observed ranges (Table 3 and 4). For this study, we used a subset of 1000 calibrated parameter sets from the previous study. For each calibrated parameter set, we derived the care seeking rate of asymptomatic (τ_A) and symptomatic (τ_S) individuals using the fraction of successfully treated individuals who were symptomatic at baseline ϕ (Supplementary Material: Section Derivation of τ_A and τ_S). We set default values for the testing and treatment parameters ψ , ξ_G , ξ_R , η_1 , η_2 , δ , ω , λ_A and λ_S guided by literature (Table 1).

Sensitivity Analyses

We performed sensitivity analysis to confirm that our model results are robust in scenarios with different properties of tests (ξ_G , ξ_R), antibiotics (η_1 , η_2), and populations and clinics (δ , ω , λ_A , λ_S). First, we performed sensitivity analyses of the number of observed cases averted with regard to changes in both the fraction of asymptomatic individuals who return for treatment at baseline (λ_A) and fraction of successfully treated individuals who were symptomatic at baseline (ψ) (Fig. 3), as well as to changes in single testing and treatment parameters (ξ_G , ξ_R , λ_A , λ_S , ψ , δ , ω , Supplementary Material: Figures S3-S9). Second, we evaluated the sensitivity of the ratio of resistance spread with regard to changes in the test sensitivity to detect resistance against the first-line antibiotic when using POC ($\xi_{R, POC}$), the fraction of asymptomatic individuals who return for treatment at baseline ($\lambda_{A, baseline}$) and the fraction of successfully treated individuals who were symptomatic at baseline (ψ , Fig. 4 and 5). Third, we tested the sensitivity of our model results to the assumption that the test sensitivity to detect *N. gonorrhoeae* (ξ_G) is 99% for culture testing. For this, we simulated an alternative baseline scenario where only culture, with a test sensitivity to detect *N. gonorrhoeae* (ξ_G) of 90%, is used (all other parameters as in Table 1, Supplementary Material: Figures S10-S12).

Simulation

For each parameter set, we first simulated a resistance-free baseline scenario where either culture or NAAT is used ($\delta > 0$, $\lambda_A < 1$). We simulated the baseline scenario until it reached equilibrium using the function *runsteady* from the R language and software environment for statistical computing [27] package *rootSolve* [28]. Next, we introduced resistant strains by converting 0.1% of all sensitive infections into resistant infections. We then set the parameter ξ_R to reflect the different testing scenarios (culture, NAAT, POC + R or POC - R). For POC tests, we additionally set $\delta = 0$ and $\lambda_A = 1$. Finally, we simulated the model using the function *lsoda* from the R package *deSolve* [29].

Results

Proportion of resistant infections

We determined the proportion of gonorrhoea infections resistant to the first-line antibiotic for up to 30 years after the introduction of resistance (Fig. 2). The proportion of resistant infections remains lowest when POC + R is used (MSM: median 0.18% after 30 years, interquartile range (IQR) 0.17 – 0.21%; HMW: 0.12%, 0.11 – 0.12%). The proportion of resistant infections also remains low with culture (MSM: 1.19%, 0.68 – 3.59%, HMW: 0.13%, 0.12 – 0.15%). In contrast, resistant infections largely replace sensitive infections after 30 years using NAAT (MSM: 100%, 100 – 100%, HMW: 99.27%, 88.54 – 99.97%) and POC - R (MSM: 100%, 100 – 100%, HMW: 99.73%, 94.30 – 99.99%). The proportion of resistant infections exceeds the 5% resistance threshold (Fig. 2, dashed line) marginally earlier when POC - R is used (MSM: median < 2.42, IQR 2.00 – 2.92 years, HMW: < 9.25, 7.25 – 12.25 years) than when NAAT is used (MSM: < 2.58, 2.08 – 3.08 years, HMW: < 10.08, 7.83 – 13.33 years). Overall, POC + R performs best in keeping the proportion resistant infections low and POC - R performs worst.

Observed cases averted

We calculated the observed cases averted (per 100 000 persons) after 5 years using culture, POC + R or POC - R in comparison with NAAT (Fig. 3). For the default values ($\lambda_{A, baseline} = 90\%$, $\psi = 60\%$), using NAAT leads to a median of 36366 (IQR 33789 – 39692) observed cases after 5 years for MSM and 1228 (927 – 1610) for HMW. Culture averts 1876 (740 – 4919) cases in MSM and 3 (1 – 7) in HMW compared with NAAT. POC + R averts even more cases than culture in both MSM (3353, 1697 – 7259) and HMW (118, 69 – 198). POC - R averts less cases than culture in MSM (772, 452 – 1119), but about the same as POC + R in HMW (115, 68 – 190).

For culture, increasing the fraction of asymptomatic individuals who return for treatment at baseline ($\lambda_{A, baseline}$) and decreasing the fraction of successfully treated individuals who were symptomatic at baseline (ψ) increases the median number of observed cases averted. For POC + R, decreasing $\lambda_{A, baseline}$ and decreasing ψ leads to an increase in the median observed cases averted. For POC - R, decreasing $\lambda_{A, baseline}$ and the intermediate value of ψ results in an increase in median averted cases. For all combinations of $\lambda_{A, baseline}$ and ψ in both MSM and HMW, POC + R averts more cases at the median than culture. This result is robust to changes in single testing and treatment parameters (Supplementary Material: Figures S3-S9).

Ratio of resistance spread

We determined the ratio of the rate of resistance spread between POC and culture (Fig. 4) and POC and NAAT

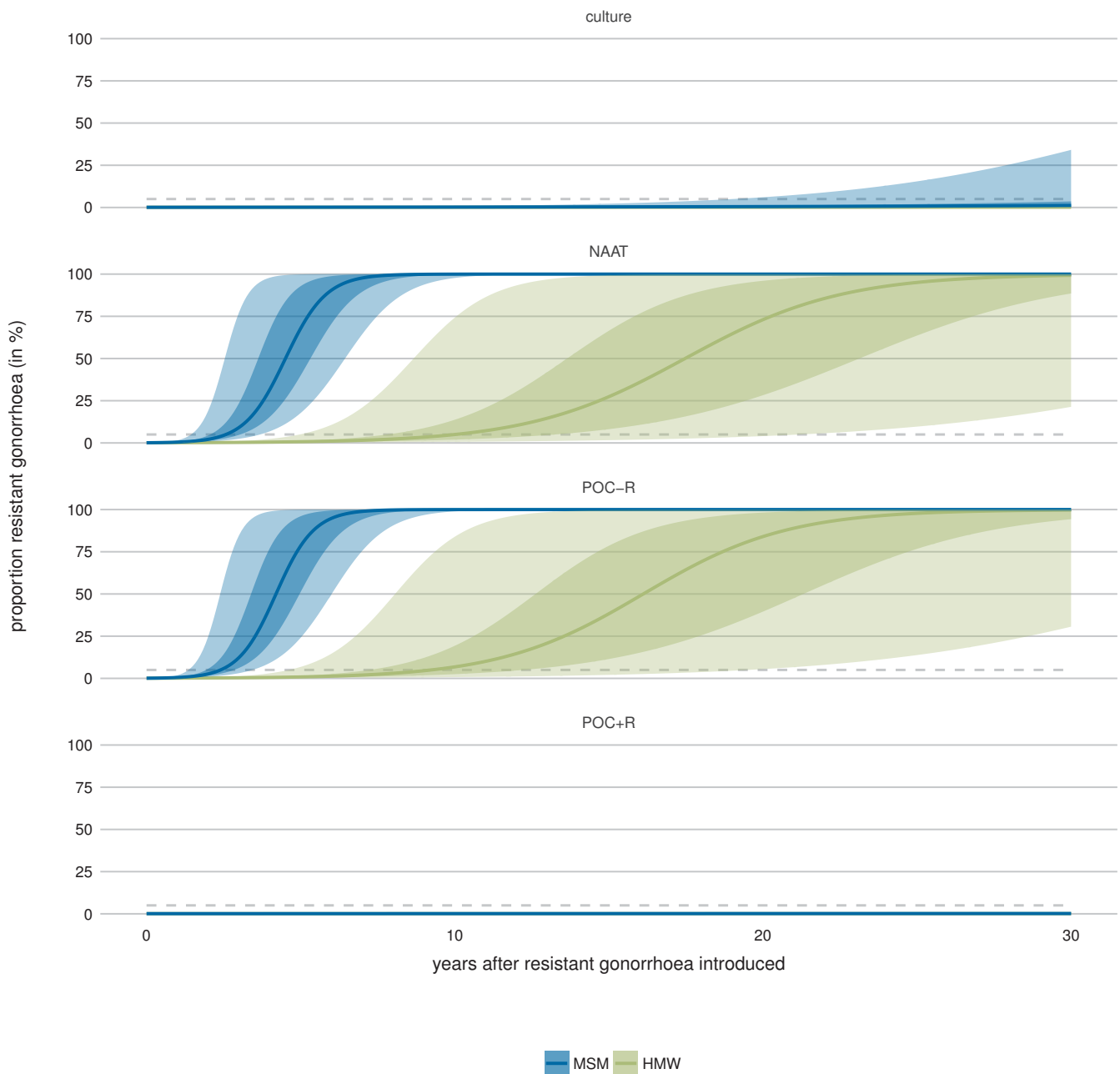


Figure 2. Proportion of resistant gonorrhoea infections for each testing scenario. The continuous lines give the median proportion of resistant infections over all simulations. Shaded areas indicate that 50% or 95% of all simulations lie within this range. MSM: men who have sex with men, HMW: heterosexual men and women. The proportion of resistant infections remains lowest when point-of-care with resistance detection (POC + R) is used, followed by culture. The proportion of resistant infections exceeds the 5% threshold (dashed lines) marginally earlier with point-of-care without resistance detection (POC – R) than with the nucleic acid amplification test (NAAT).

Table 3. Gonorrhoea prevalence and incidence in baseline scenario (before resistance introduced) for MSM. The prevalence and incidence ranges used for calibration for men who have sex with men (MSM) were based on the Health in Men Study in Australia [24]. The baseline median and interquartile range (IQR) are based on the simulation results of 1000 calibrated parameter sets. The upper and lower bound of the calibration range for the low and high sexual activity class were set to the lower and upper bound for the total population. The calibration is detailed in [15].

	Range used for calibration	Baseline median (IQR)
Prevalence low activity class (%)	0 – 2.79	0.59 (0.42 – 0.79)
Prevalence high activity class (%)	1.19 – 100	27.64 (23.25 – 31.91)
Prevalence total population (%)	1.19 – 2.79	2.09 (1.69 – 2.43)
Incidence total population (100000 persons ⁻¹ y ⁻¹)	5880 – 7190	6493.49 (6192.89 – 6842.70)

Table 4. Gonorrhoea prevalence and incidence in baseline scenario (before resistance introduced) for HMW. The prevalence and incidence ranges used for calibration for Heterosexual men and women (HMW) were based on the National Health and Nutrition Examination Survey [25] and surveillance data [26], both from CDC. The baseline median and interquartile range (IQR) are based on the simulation results of 1000 calibrated parameter sets. The upper and lower bound of the calibration range for the low and high sexual activity class were set to the lower and upper bound for the total population. The calibration is detailed in [15].

	Range used for calibration	Baseline median (IQR)
Prevalence low activity class (%)	0 – 0.38	0.12 (0.09 – 0.15)
Prevalence high activity class (%)	0.16 – 100	2.14 (1.71 – 2.60)
Prevalence total population (%)	0.16 – 0.38	0.25 (0.21 – 0.3)
Incidence total population (100000 persons ⁻¹ y ⁻¹)	120 – 360	222.13 (172.19 – 283.54)

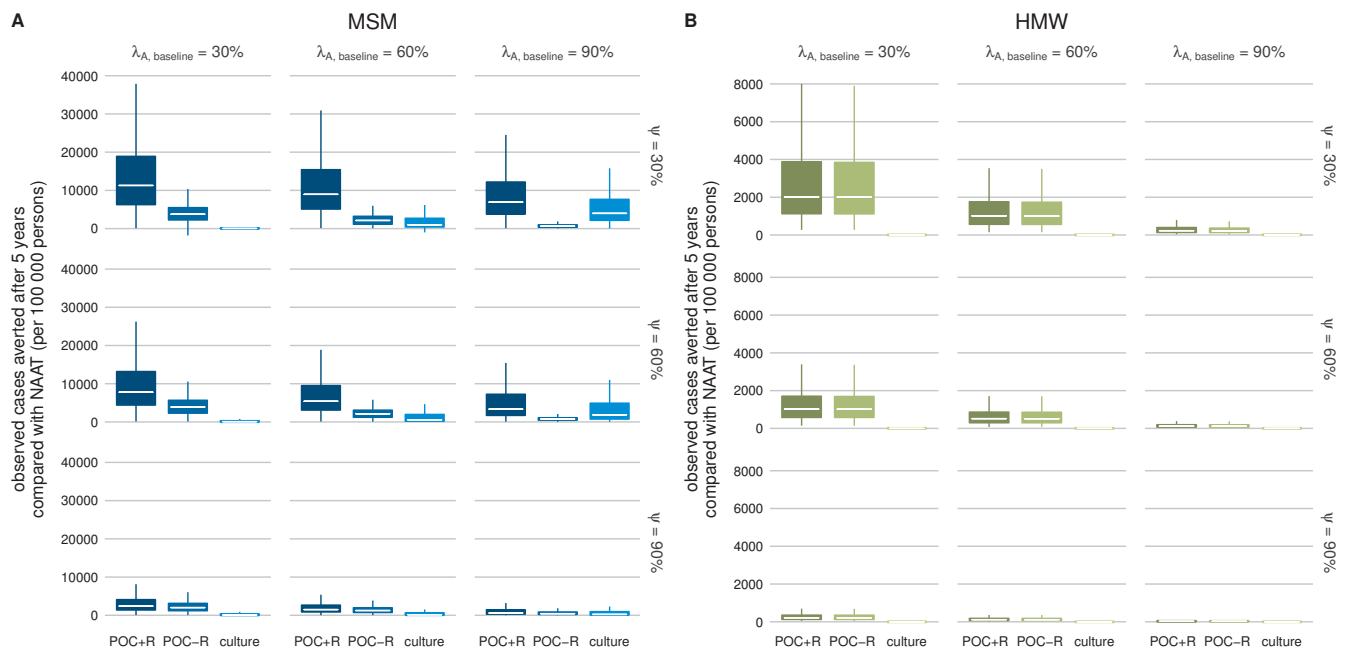


Figure 3. Two-dimensional sensitivity analysis of observed cases averted (per 100 000 persons) after 5 years. The sensitivity analysis is performed with respect to the fraction of asymptomatic individuals who return for treatment at baseline ($\lambda_{A, \text{baseline}}$) and the fraction of successfully treated individuals who were symptomatic at baseline (ψ), for (A) men who have sex with men (MSM) and (B) heterosexual men and women (HMW). The central right plot of each panel shows the default scenario ($\lambda_{A, \text{baseline}} = 90\%$, $\psi = 60\%$). NAAT: nucleic acid amplification test, POC + R: point-of-care test (POC) with resistance detection, POC – R: POC without resistance detection. Lower/upper bound of the box indicate first/third quartiles, bar in box indicates median, whiskers span 1.5 times interquartile range. Outliers not shown for more clarity.

(Fig. 5). For the default values ($\xi_{R, \text{culture}} = 99\%$, $\xi_{R, \text{NAAT}} = 0\%$, $\xi_{R, \text{POC}} = 99\%$, $\lambda_{A, \text{baseline}} = 90\%$, $\psi = 60\%$), resistance spreads more slowly with POC compared with culture or NAAT. Decreasing the test sensitivity to detect resistance of POC ($\xi_{R, \text{POC}}$) can result in a faster spread of resistance with POC. A slight decrease in $\xi_{R, \text{POC}}$ to 80-95% already leads to faster resistance spread with POC compared with culture. In contrast, only very low values of $\xi_{R, \text{POC}}$ result in a faster resistance spread for POC compared with NAAT.

Discussion

Using a mathematical transmission model, we compared the expected impact of POC tests on gonorrhoea cases and antibiotic resistance with conventional tests, culture and NAAT. We found that POC tests that detect antibiotic resistance avert more gonorrhoea cases than any other test across all simulated settings. Compared with culture and NAAT, POC tests with high sensitivity to detect resistance slow the spread of resistant infections. POC tests with no or low sensitivity to detect resistance accelerate the spread of resistant infections.

We captured the basic principles of the gonorrhoea testing and treatment process for culture, NAAT and POC in a single model structure. The parameters describing the sexual behaviour and the natural history of gonorrhoea were estimated and calibrated in a previous study [15]. The default parameters that describe testing and treatment of gonorrhoea were based on literature values and are measurable. The model results are robust in sensitivity analyses (Fig. 3 and 4, 5, Supplementary Material: Figures S3-S12).

We managed the complexity of our model with the following assumptions: First, we did not consider test specificity. A low test specificity to detect resistance against the first-line antibiotic would result in increased use of the second-line antibiotic, and thus simultaneously decrease the level of resistance against the first-line antibiotic and increase the level of resistance against the second-line antibiotic. Since we focused on resistance against the first-line antibiotic, we could not capture the impact of test specificity appropriately. Second, our model does not include a change in antibiotic recommendations: undetected resistant infections are always treated with the first-line antibiotic, even if all infections in the population are resistant. This clinical pathway increases the average duration of resistant infections and possibly the observed cases. Whilst this is unlikely in high income countries with good antibiotic resistance surveillance, it is not an unrealistic scenario in resource poor settings without surveillance where 71-100% of gonococcal strains are resistant to fluoroquinolones [30]. In our model, MSM have a substantial level of resistant gonorrhoea infections after 5 years using NAAT. We expect that our model overestimates the observed cases

using NAAT and the observed cases averted using culture and POC + R compared with a model including antibiotic recommendation change. Third, we considered only treatment with a single antibiotic although current treatment guidelines recommend combination therapy with two antibiotics simultaneously [1, 7]. The model results are fully applicable to treatment with combination therapy if antibiotic-resistant gonorrhoea is interpreted as resistance against both antibiotics used for combination therapy. Fourth, we investigated the effects of one test at a time and did not consider the effects of mixed testing. Our results therefore only show what the ideal effects of each test could be. Fifth, we simplified the testing and treatment process. To better compare the testing scenarios, we did not model care seeking and returning for treatment as separate processes, but approximated the overall treatment rates. In accordance with WHO [16] and CDC recommendations [22], we assumed that re-treatment of resistant infections occurs with the second-line antibiotic because a resistance profile has been determined after the second visit. Finally, for better comparability we assumed that culture, NAAT and POC tests have the same sensitivity to detect gonorrhoea, even though culture has lower sensitivity to detect rectal or pharyngeal gonorrhoea than molecular tests [31]. A lower test sensitivity to detect gonorrhoea, ξ_G , requires a higher care-seeking rate of asymptomatic individuals, τ_A , to obtain the same prevalence and incidence rates. We simulated an alternative scenario (Supplementary Material: Figures S10-S12) where only culture is used at baseline (with $\xi_G = 90\%$ for culture and all other values as in Table 1). In this scenario, the proportion of resistant infections after 30 years using culture is higher in MSM (median 3.18%, IQR 1.51 – 11.33%) and the observed cases averted after 5 years using POC + R compared with NAAT is larger (median 4236, IQR 2161 – 8839 per 100000 persons). Overall the effect of lower test sensitivity to detect gonorrhoea with culture was small.

Currently, there are no commercial POC tests that can detect antibiotic-resistant *N. gonorrhoeae* [8] and there remain challenges for their development. First, molecular POC test that detect resistance need molecular markers that reliably predict phenotypic resistance. So far only markers that predict resistance against some antibiotics are known [8, 32, 33]. Second, diagnostic tests need to deliver results fast to be considered point-of-care. The fastest molecular diagnostic test for gonorrhoea that is commercially available takes 90 minutes [34, 35] which might be too long to wait for some patients. Finally, costs and training requirements for molecular tests have hindered their availability in low income countries so far [36].

This study addresses two key questions for gonorrhoea control and resistance [37]. First, we investigated the potential impact of a POC test that detects antibiotic

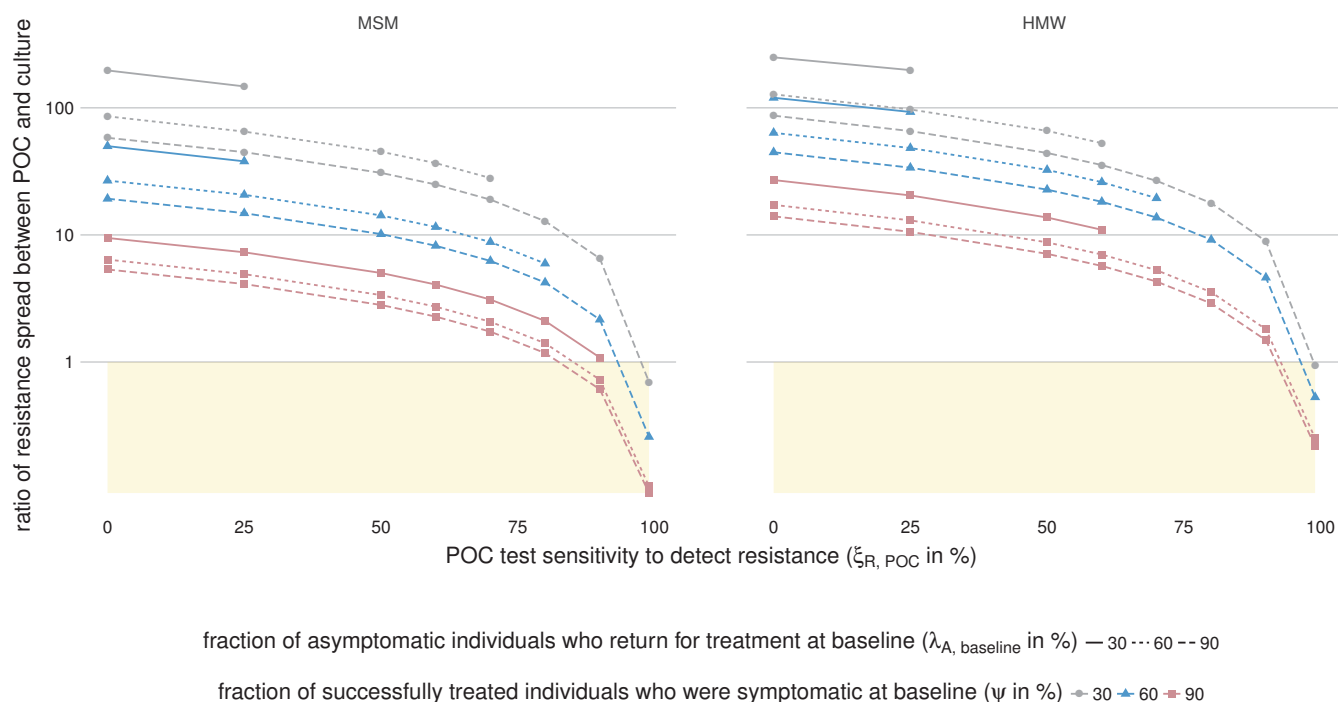
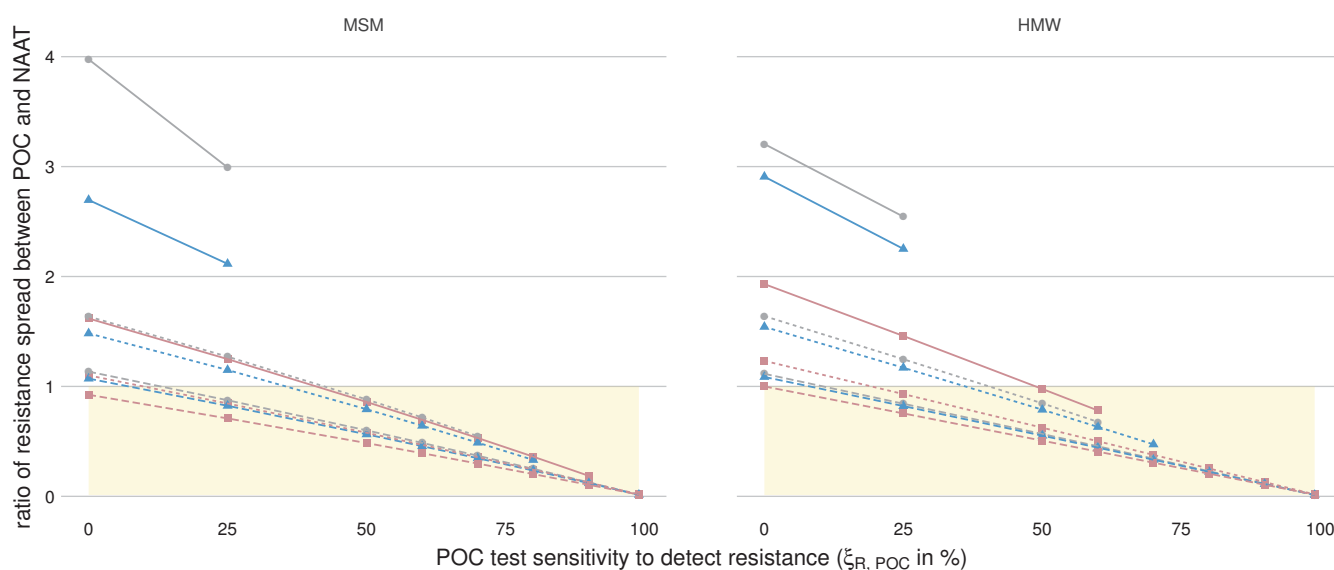


Figure 4. Ratio of resistance spread between point-of-care test (POC) and culture. Shown are the ratios of resistance spread for men who have sex with men (MSM) and heterosexual men and women (HMW) for $\xi_{R, \text{culture}} = 99\%$ and different values of $\xi_{R, \text{POC}}$, $\lambda_{A, \text{baseline}}$ and ψ (POC – R: $\xi_{R, \text{POC}} = 0$, POC + R: $\xi_{R, \text{POC}} > 0$). The shaded areas indicate that resistance spread is slower when using POC than when using culture. For the default values ($\xi_{R, \text{POC}} = 99\%$, $\lambda_{A, \text{baseline}} = 90\%$, $\psi = 60\%$), resistance spread is slower when using POC than when using culture. For most other shown values using POC accelerates resistance spread. Each data point gives the median value over 1000 simulations (one per calibrated parameter set). Some calibrated parameter sets lead to extinction of gonorrhoea in the simulation (Supplementary Material: Figure S2). In these simulations, resistance did not spread and the ratio of resistance spread could not be calculated. Data points that would include such simulations were excluded from this figure since they would show the median ratio of resistance spread over less than 1000 simulations. Note that the y-axis is shown in logarithmic scale.

resistance (POC + R). We found that POC + R can slow resistance spread and reduce the gonorrhoea cases compared with culture or NAAT. The impact of POC + R is particularly strong when the fraction of asymptomatic individuals who return for treatment ($\lambda_{A, \text{baseline}}$) and the fraction of successfully treated individuals who were symptomatic (ψ) were low before POC + R is introduced. However, when the POC test cannot detect resistance (POC – R) the benefits of POC are outweighed by accelerated resistance evolution: because fewer patients are lost to follow up, more patients are treated and more antibiotic treatment selects more strongly for antibiotic resistance. Since resistance cannot be detected, resistance levels increase and fewer cases are averted. Second, we investigated the impact of POC tests in two populations with different levels of risk of gonorrhoea, MSM and HMW. We found that in both populations, POC tests with reliable resistance detection (POC + R) slow down the spread of resistance and avert the most cases. POC tests without resistance detection (POC – R) avert about as many cases as POC + R in HMW, but clearly

fewer cases than POC + R in MSM. Since resistance usually spreads faster in MSM [15], the faster spread of resistance caused by POC – R already impacts the cases averted after 5 years in MSM, but not yet in HMW. POC tests that detect resistance reliably are crucial for both populations and both populations need culture-based surveillance of resistance to keep molecular markers for POC resistance detection updated.

This modelling study addresses clinically relevant situations, can be used to help design trials comparing different test strategies and guide the introduction of POC tests in the future. POC tests with high sensitivity to detect resistance can replace culture-based diagnosis in clinical settings, as long as culture-based surveillance of antibiotic resistance is maintained to monitor resistance levels and to determine molecular markers for POC tests. POC tests with lower sensitivities to detect resistance should not replace culture-based diagnosis, but might have some advantages over NAAT. POC test with low or no sensitivity to detect resistance should not be introduced, because POC tests without reliable resistance



fraction of asymptomatic individuals who return for treatment at baseline ($\lambda_{A, \text{baseline}}$ in %) — 30 — 60 — 90

fraction of successfully treated individuals who were symptomatic at baseline (ψ in %) — 30 — 60 — 90

Figure 5. Ratio of resistance spread between point-of-care test (POC) and nucleic acid amplification test (NAAT). Shown are the ratios of resistance spread for men who have sex with men (MSM) and heterosexual men and women (HMW) for $\xi_{R, \text{NAAT}} = 0\%$ and different values of $\xi_{R, \text{POC}}$, $\lambda_{A, \text{baseline}}$ and ψ (POC – R: $\xi_{R, \text{POC}} = 0$, POC + R: $\xi_{R, \text{POC}} > 0$). The shaded areas indicate that resistance spread is slower when using POC than when using NAAT. For the default values ($\xi_{R, \text{POC}} = 99\%$, $\lambda_{A, \text{baseline}} = 90\%$, $\psi = 60\%$) and most other shown values resistance spread is slower when using POC than when using NAAT. Each data point gives the median value over 1000 simulations (one per calibrated parameter set). Some calibrated parameter sets lead to extinction of gonorrhoea in the simulation (Supplementary Material: Figure S2). In these simulations, resistance did not spread and the ratio of resistance spread could not be calculated. Data points that would include such simulations were excluded from this figure since they would show the median ratio of resistance spread over less than 1000 simulations.

detection can accelerate the spread of antibiotic-resistant gonorrhoea.

thors contributed to and approved the final version of manuscript.

Competing interests

NL, SB and CLA received funding for RaDAR-GO from SwissTransMed, Platforms for Translational Research in Medicine. RaDAR-GO is a project that aims to develop a point-of-care test to detect antibiotic-resistant gonorrhoea. SMF and CLA are funded by RaDAR-GO.

Funding

SMF and CLA are funded by SwissTransMed, Platforms for Translational Research in Medicine. SB is funded by the European Research Council.

Authors' contributions

SMF, NL, SB, CLA designed the study. SMF simulated the model. SMF, NL, SB, CLA interpreted the data. SMF wrote the first version of the manuscript. All au-

References

- [1] C Bignell, M Unemo, and European STI Guidelines Editorial Board. 2012 European guideline on the diagnosis and treatment of gonorrhoea in adults. *Int J STD AIDS*, 24(2):85–92, feb 2013.
- [2] World Health Organization. Report on global sexually transmitted infection surveillance 2013, 2014.
- [3] Helen Fifer, Usha Natarajan, Lucy Jones, Sarah Alexander, Gwenda Hughes, Daniel Golparian, and Magnus Unemo. Failure of dual antimicrobial therapy in treatment of gonorrhoea. *N Engl J Med*, 374(25):2504–6, jun 2016.
- [4] Lori Newman, Jane Rowley, Stephen Vander Hoorn, Nalinka Saman Wijesooriya, Magnus Unemo, Nicola Low, Gretchen Stevens, Sami Gottlieb, James Kiarie, and Marleen Temmerman. Global estimates of the prevalence and incidence of four curable sex-

- ually transmitted infections in 2012 based on systematic review and global reporting. *PLoS One*, 10(12):e0143304, 2015.
- [5] Sepehr N Tabrizi, Magnus Unemo, Daniel Golparian, Jimmy Twin, Athena E Linnios, Monica Lahra, and Rebecca Guy. Analytical evaluation of GeneXpert CT/NG, the first genetic point-of-care assay for simultaneous detection of *Neisseria gonorrhoeae* and *Chlamydia trachomatis*. *J Clin Microbiol*, 51(6):1945–7, 2013.
- [6] Nicola Low, Magnus Unemo, J J Skov, Judith Breuer, and Judith M. Stephenson. Molecular diagnostics for gonorrhoea: implications for antimicrobial resistance and the threat of untreatable gonorrhoea. *PLoS Med*, 11(2):e1001598, 2014.
- [7] World Health Organization. WHO guidelines for the treatment of *Neisseria gonorrhoeae*, 2016.
- [8] Nicola Low and Magnus Unemo. Molecular tests for the detection of antimicrobial resistant *Neisseria gonorrhoeae*: when, where, and how to use? *Curr Opin Infect Dis*, 29(1):45–51, feb 2016.
- [9] Ben B Hui, David P Wilson, James S Ward, Rebecca J Guy, John M Kaldor, Matthew G Law, Jane S Hocking, and David G Regan. The potential impact of new generation molecular point-of-care tests on gonorrhoea and chlamydia in a setting of high endemic prevalence. *Sex Health*, 10(4):348–56, 2013.
- [10] Sze-Ann Woon and Dale Fisher. Antimicrobial agents - optimising the ecological balance. *BMC Med*, 14:114, aug 2016.
- [11] Catherine A Ison, Carolyn Deal, and Magnus Unemo. Current and future treatment options for gonorrhoea. *Sex Transm Infect*, 89:iv52–iv56, dec 2013.
- [12] Magnus Unemo. Current and future antimicrobial treatment of gonorrhoea - the rapidly evolving *Neisseria gonorrhoeae* continues to challenge. *BMC Infect Dis*, 15:364, 2015.
- [13] C H Chan, C J McCabe, and D N Fisman. Core groups, antimicrobial resistance and rebound in gonorrhoea in North America. *Sex Transm Infect*, 88(3):200–4, apr 2012.
- [14] M Xiridou, L C Soetens, F D H Koedijk, M A B van der Sande, and J Wallinga. Public health measures to control the spread of antimicrobial resistance in *Neisseria gonorrhoeae* in men who have sex with men. *Epidemiol Infect*, 143(8):1575–84, oct 2015.
- [15] Stephanie M. Fingerhuth, Sebastian Bonhoeffer, Nicola Low, and Christian L. Althaus. Antibiotic-resistant *Neisseria gonorrhoeae* spread faster with more treatment, not more sexual partners. *PLoS Pathog*, 12(5):e1005611, 2016.
- [16] World Health Organization. Global action plan to control the spread and impact of antimicrobial resistance in *Neisseria gonorrhoeae*, 2012.
- [17] David M. Whiley, John W. Tapsall, and Theo P. Sloots. Nucleic acid amplification testing for *Neisseria gonorrhoeae*: an ongoing challenge. *J Mol Diagn*, 8(1):3–15, 2006.
- [18] John Tapsall and World Health Organization. Antimicrobial resistance in *Neisseria gonorrhoeae*, 2001.
- [19] H Hunter Handsfield, Z A Dalu, David H Martin, John M Douglas, James M McCarty, David Schlossberg, and Azithromycin Gonorrhea Study Group. Multicenter trial of single-dose azithromycin vs. ceftriaxone in the treatment of uncomplicated gonorrhoea. *Sex Transm Dis*, 21(2):107–11, 1994.
- [20] G Brook. The performance of non-NAAT point-of-care (POC) tests and rapid NAAT tests for chlamydia and gonorrhoea infections. An assessment of currently available assays. *Sex Transm Infect*, 91:539–44, 2015.
- [21] S Dudareva-Vizule, K Haar, A Sailer, H Wisplinghoff, F Wisplinghoff, U Marcus, and PARIS study group. Prevalence of pharyngeal and rectal *Chlamydia trachomatis* and *Neisseria gonorrhoeae* infections among men who have sex with men in Germany. *Sex Transm Infect*, 90(1):46–51, 2014.
- [22] Centers for Disease Control and Prevention. Cephalosporin-resistant *Neisseria gonorrhoeae* public health response plan, 2012.
- [23] Anne M. Johnson, Catherine H. Mercer, Bob Erens, Andrew J. Copas, Sally McManus, Kaye Wellings, Kevin A. Fenton, Christos Korovessis, Wendy Macdowall, Kiran Nanchahal, Susan Purdon, and Julia Field. Sexual behaviour in Britain: partnerships, practices, and HIV risk behaviours. *Lancet*, 358(9296):1835–42, 2001.
- [24] F Jin, G P Prestage, L Mao, S C Kippax, C M Pell, B Donovan, P H Cunningham, D J Templeton, J M Kaldor, and A E Grulich. Incidence and risk factors for urethral and anal gonorrhoea and chlamydia in a cohort of HIV-negative homosexual men: the Health in Men Study. *Sex Transm Infect*, 83(2):113–9, 2007.
- [25] S Deblina Datta, Maya Sternberg, Robert E Johnson, Stuart Berman, John R Papp, Geraldine McQuillan, and Hillard Weinstock. Gonorrhoea and chlamydia in the United States among persons 14 to 39 years of age, 1999 to 2002. *Ann Intern Med*, 147(2):89–96, 2007.
- [26] Centers for Disease Control and Prevention. Sexually transmitted disease surveillance 2013, 2014.
- [27] R Core Team. R: a language and environment for statistical computing, 2013.

- [28] K Soetaert. rootSolve: Nonlinear root finding, equilibrium and steady-state analysis of ordinary differential equations, 2009.
- [29] K Soetaert, T Petzoldt, and R W Setzer. Solving differential equations in R: package deSolve. *J Stat Softw*, 33(9):1–25, 2010.
- [30] World Health Organization. Monitoring gonococcal antimicrobial susceptibility, 2017.
- [31] Julius Schachter, Jeanne Moncada, Sally Liska, Clara Shayevich, and Jeffrey D Klausner. Nucleic acid amplification tests in the diagnosis of chlamydial and gonococcal infections of the oropharynx and rectum in men who have sex with men. *Sex Transm Dis*, 35(7):637–42, 2008.
- [32] Ella Trembizki, Rebecca Guy, Basil Donovan, John M Kaldor, Monica M Lahra, David M Whitley, and GRAND study investigators. Further evidence to support the individualised treatment of gonorrhoea with ciprofloxacin. *Lancet Infect Dis*, 16(9):1005–6, sep 2016.
- [33] Yonatan H Grad, Simon R Harris, Robert D Kirkcaldy, Anna G Green, Debora S Marks, Stephen D Bentley, David Trees, and Marc Lipsitch. Genomic epidemiology of gonococcal resistance to extended-spectrum cephalosporins, macrolides, and fluoroquinolones in the United States, 2000-2013. *J Infect Dis*, 214(10):1579–87, nov 2016.
- [34] Charlotte A Gaydos. Review of use of a new rapid real-time PCR, the Cepheid GeneXpert® (Xpert) CT/NG assay, for Chlamydia trachomatis and Neisseria gonorrhoeae: results for patients while in a clinical setting. *Expert review of molecular diagnostics*, 14(2):135–7, 2014.
- [35] Sam Ratnam, Dan Jang, Jodi Gilchrist, Marek Smieja, Andre Poirier, Todd Hatchette, Jean-Frederic Flandin, and Max Chernesky. Workflow and maintenance characteristics of five automated laboratory instruments for the diagnosis of sexually transmitted infections. *J Clin Microbiol*, 52(7):2299–304, jul 2014.
- [36] Iruka N Okeke, Rosanna W Peeling, Herman Goossens, Raymond Auckenthaler, Stuart S Olmsted, Jean-Francois de Lavison, Barbara L Zimmer, Mark D Perkins, and Katarina Nordqvist. Diagnostics as essential tools for containing antibacterial resistance. *Drug Resist Updat*, 14(2):95–106, 2011.
- [37] Yonatan H Grad, Edward Goldstein, Marc Lipsitch, and Peter J White. Improving control of antibiotic-resistant gonorrhea by integrating research agendas across disciplines: key questions arising from mathematical modeling. *J Infect Dis*, 213(6):883–90, 2016.