bioRxiv preprint doi: https://doi.org/10.1101/129536; this version posted April 24, 2017. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

A human huntingtin SNP alters post-translational modification and pathogenic proteolysis of the protein causing Huntington disease

Martin DDO†*, Kay C†, Collins JA, Nguyen YT, Slama RA, Hayden MR*

Centre for Molecular Medicine and Therapeutics, Department of Medical Genetics, University of British Columbia, Vancouver, BC, Canada

†These authors contributed equally to this work

***Corresponding authors:**

Dr. Dale Martin

dalem@cmmt.ubc.ca

Phone: +1 604 875 3809

Fax: +1604 875 3819

Dr. Michael R. Hayden

mrh@cmmt.ubc.ca

Phone: +1 604 875 3535

Fax: +1 604 875 3819

bioRxiv preprint doi: https://doi.org/10.1101/129536; this version posted April 24, 2017. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

Abstract

Post-translational modifications (PTMs) are key modulators of protein function. Huntington disease (HD) is a dominantly inherited neurodegenerative disorder caused by an expanded CAG trinucleotide repeat in the huntingtin (*HTT*) gene. A spectrum of PTMs have been shown to modify the normal functions of HTT, including proteolysis, phosphorylation and lipidation, but the full contribution of these PTMs to the molecular pathogenesis of HD remains unclear. In this study, we examine all commonly occurring missense mutations in *HTT* to identify potential human modifiers of HTT PTMs relevant to HD biology. We reveal a SNP that modifies post-translational myristoylation of HTT, resulting in downstream alterations to toxic HTT proteolysis in human cells. This is the first SNP shown to functionally modify a PTM in HD and the first validated genetic modifier of post-translational myristoylation. This SNP is a high-priority candidate modifier of HD phenotypes and may illuminate HD biology in human studies.

bioRxiv preprint doi: https://doi.org/10.1101/129536; this version posted April 24, 2017. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

Introduction

Huntington disease (HD) is a debilitating neurodegenerative disease with no treatment to delay progression of the disease. HD is caused by an expanded CAG repeat in the huntingtin gene (*HTT*) that translates into a polyglutamine tract at the N-terminus of the huntingtin protein (HTT)[1]. Wild-type HTT is critical for cell viability[2] and has been shown to play a role in a variety of pathways including orientation of the mitotic spindle[3], trafficking of autophagosomes in neurons[4], and in regulating autophagy[5]. Expanded polyglutamine in HTT leads to loss of function and a toxic gain of function in neurons of the striatum and cortex, causing behavioral changes, movement deficits and ultimately death. Age of disease onset is inversely proportional to the length of the CAG repeat[6]. However, age of onset may be accelerated or delayed by genetic modifiers, including a SNP at the transcription factor binding site of Nf-kB in the *HTT* promoter[7], and in genes related to DNA repair[8].

The HTT protein is a large monomeric protein whose function is intricately regulated by post-translational modifications including phosphorylation, acetylation, ubiquitination, proteolysis, and fatty acylation[9]. While some PTMs of HTT have been shown to be protective against toxicity of mutant HTT, such as phosphorylation at S13/16 and S421[10], others are crucial for HD pathogenesis or increase mutant HTT toxicity. In particular, caspase-mediated proteolysis of HTT at amino acid D586 has been shown to be necessary for the development of disease phenotypes in HD mouse models[11,12]. Consequently, modulating PTMs has become a focus of therapeutic strategies for HD. We sought to identify human SNPs that lead to missense mutations that may alter PTMs in HTT and, consequently, modify progression or pathogenic effects of the disease.

bioRxiv preprint doi: https://doi.org/10.1101/129536; this version posted April 24, 2017. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

Results

To identify SNPs that could alter HTT PTMs and potentially modify HTT function, all common missense mutations (≥0.1% minor allele frequency; MAF) within *HTT* were curated from Phase 3 of the 1000 Genomes Project (1KG) and from the Genome Aggregation Database (gnomAD). Nineteen common missense SNPs with ≥0.1% MAF were found in 1KG, and 19 common missense SNPs with ≥0.1% MAF were found in gnomAD (**Table 1**). The top 14 most common missense SNPs in 1KG and in gnomAD were shared in both data sets, highlighting convergent allele discovery by distinct methodologies.

A manually curated list of known HTT PTMs was intersected with all common missense SNPs identified from 1KG and gnomAD (**Figure 1**). Among the 21 common missense SNPs identified from 1KG and gnomAD, two overlapped directly with known HTT PTMs; rs118005095 (G553E) and rs201646519 (S2076P) (**Table 1**, boxed; **Supplemental Table 1**). SNP rs201646519 intersects with the predicted phosphorylated residue S2076, resulting in a proline substitution thereby blocking phosphorylation at this site. Myristoylated G553[13,14] overlapped with the missense SNP rs118005095, which leads to a substitution of glycine to glutamic acid (G553E) and would therefore completely abrogate post-translational myristoylation at this site (**Figure 2a**). Myristoylation involves the covalent addition of the 14 carbon fatty acid, myristate, to N-terminal glycines either co-translationally on the nascent polypeptide as it is being translated on the ribosome following the removal of the initiator methionine or post-translationally following proteolysis that exposes an N-terminal glycine[15]. The fatty acid moiety promotes membrane binding of proteins. No other common missense SNPs directly intersected with a known HTT PTM residue. Due to the low frequency of rs201646519, we focused on rs118005095.

The G553E SNP rs118005095 shows pronounced ethnic differences in frequency, being most common in individuals of East Asian ancestry. In 1KG, rs118005095 is observed on 31 out of 5008 chromosomes

bioRxiv preprint doi: https://doi.org/10.1101/129536; this version posted April 24, 2017. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

from all populations, of which 30 instances occur in subjects of defined East Asian origin (n=504 subjects) at a genotypic frequency of 6.0% (30/504) and allelic frequency of 3.0% (30/1008) (**Figure 2b**). The one remaining 1KG chromosome with rs118005095 outside East Asian individuals occurs in a Bengali subject from Bangladesh, close to East Asia. In the gnomAD data, rs118005095 occurs in chromosomes from East Asian subjects at an allelic frequency of 2.914% (552/18942). In contrast, rs118005095 is observed in <0.1% of chromosomes from European, African, South Asian, and Latino subjects, reflecting its absence in similar reference populations from 1KG. Therefore rs118005095 is expected to occur in approximately 6.0% of individuals from the East Asian general population.

We have previously shown that *HTT* is characterized by a haplotype block of low recombination and that SNPs within the gene represent specific haplotypes[16]. *HTT* haplotype analysis in 1KG reveals that rs118005095 occurs on a specific A3b haplotype variant in the East Asian population, tagged by three additional haplotype-defining SNPs: rs116955486, rs80044337, and rs362332 (**Figure 2c**). Of note, previously published haplotypes of the HD mutation in East Asian patients do not include this haplotype, suggesting that this SNP is not found on mutant *HTT* chromosomes in the East Asian population. We thus predicted that rs118005095 may occur in wild-type *HTT* of East Asian HD patients, potentially impacting the normal function of myristoylation in the context of the disease.

We have previously shown that the expanded polyglutamine mutation dramatically reduces myristoylation of HTT at G553[14]. HD patients with the G553E mutation (rs118005095) would therefore be unable to compensate for deficient myristoylation with a wild-type copy bearing the rs118005095 variant. Genotyping of archived DNA from 33 East Asian HD patients in the UBC HD Biobank confirmed the presence of the SNP in two patients (**Figure 2b**). As expected, phasing of rs118005095 to the

bioRxiv preprint doi: https://doi.org/10.1101/129536; this version posted April 24, 2017. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

mutation revealed that this missense variant occurred on wild-type *HTT* in these patients. However, no clinical information was available to assess the potential modifying effects of the SNP.

To investigate the predicted cellular effects of the rs118005095 (G553E) variant, we generated the mutation in a biologically relevant form of HTT bearing the first 1-588 amino acids appended to YFP (HTT_{1-588}-YFP, **Figure 3a**), previously shown to be post-translationally myristoylated[14] and disease-causing in HD mouse models[17]. Myristoylation at G553 was first validated in WT and $mHTT_{1-588}$-YFP using an alkyne-myristate analog that can be covalently linked to biotin-azide via Click chemistry for detection (**Supplementary Figure 1**)[18]. Caspase-cleavage at D552 was promoted using staurosporine (STS) and cycloheximide (CHX) in order to increase post-translational myristoylation at G553, as previously described[14]. Post-translational myristoylation was detected in fully-cleavable (FC) HTT (HTT_{1-588}-YFP with no point mutations added) even in the absence of STS/CHX, likely due to basal levels of caspase activity (**Supplementary Figure 1**). Myristoylation was increased in the presence of STS/CHX. A point mutation at the primary site of caspase-cleavage in HTT at position D586 to glutamate was made in order to more easily measure myristoylation at G553 following caspase cleavage at D552 (D586E, **Supplementary Figure 1**). This led to a dramatic increase of post-translational myristoylation at G553.

As predicted, substitution of the essential glycine to alanine or serine (G553A/D586E and G553S/D586E) completely abrogated post-translational myristoylation of HTT at G553 following caspase-induced cleavage. Similarly, blocking caspase cleavage at D552 by substituting the aspartate to glutamate (D552E, **Supplementary Figure 1**) completely blocked post-translational myristoylation at G553. Inhibiting caspase-cleavage at D552 appeared to block the generation of both the myristoylated and non-myristoylated bands. Therefore, we determined that myristoylation status at G553 could also be determined by gel mobility. Non-myristoylated C-terminal (ct) $HTT_{553-588}$-YFP migrates at a slightly higher

bioRxiv preprint doi: https://doi.org/10.1101/129536; this version posted April 24, 2017. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

molecular weight than myristoylated-ctHTT$_{553-588}$-YFP (**Supplementary Figure 1**). This mobility shift was shown previously in a shorter myristoylated fragment [14]and mimics other lipidated proteins such as LC3[19].

Consequently, the G553E mutation was further characterized in the absence of the myristate analog in WT-HTT$_{1-588}$-YFP (**Figure 3**). As expected, substitution of the essential N-terminal glycine at position 553 completely abrogated post-translational myristoylation of the wild-type form (**Figure 3b**). Surprisingly, the G553E mutation also inhibited caspase cleavage of HTT at D552, which was associated with a concomitant increase in caspase cleavage of HTT at D513 (**Figure 3b**). Cleavage at D586 did not appear to be affected. Of note, blocking myristoylation conventionally by substituting glycine to alanine did not prevent cleavage at D552 (**Supplemental Figure 1**). Proteolysis at D513 was recently shown to be toxic in cells [11]. Due to the increased cleavage of HTT at the toxic site D513 in the presence of the G553E mutation, we hypothesized that the G553E variant may induce toxicity of wild-type HTT. Indeed, the G553E mutation significantly increased the toxicity of wild-type HTT to similar levels as mutant 138Q HTT alone (**Figure 3c**).

bioRxiv preprint doi: https://doi.org/10.1101/129536; this version posted April 24, 2017. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

Discussion

Previously, we have shown that proteolysis at D586 is essential to the pathogenesis of HD in a mouse model[12]. In addition, we identified post-translational myristoylation as a novel PTM in HTT that is decreased in the presence of the expanded polyQ suggesting it is a protective PTM[14]. Now, we have identified the first SNP leading to a missense mutation that alters multiple PTMs in HTT. This is also the first demonstration of a naturally occurring missense mutation that blocks post-translational myristoylation. The G553E mutation was shown to completely abrogate post-translational myristoylation of HTT and induce cellular toxicity of the protein *in cellulo*. Surprisingly, this mutation gives rise to a toxic fragment of wild-type HTT caused by proteolysis at D513.

We have shown the G553E mutation blocks caspase cleavage at D552 leading to increased cleavage of HTT at the toxic caspase site D513. It has recently been shown that proteolysis of either wild-type HTT or mutant HTT at more than one site, particularly caspase-cleavage at D513 and D586, is toxic[11]. This is mediated by a newly appreciated toxicity associated with the C-terminal HTT that is exacerbated by toxic shorter N-terminal fragments[11,20,]. It is thought that the N- and C-terminal fragments interact in the presence of single site cleavage, which is considered less toxic. However, increased cleavage at multiple sites leads to decreased interaction following proteolysis at more than one site, particularly caspase cleavage at D513 and D586 and calpain cleavage at R167[11]. This proteolysis is toxic in both wild-type and mutant HTT.

Consequently, increased proteolysis of wild-type HTT at D513 caused by the G553E mutation, in conjunction with cleavage at D586 and loss of protective myristoylation, is predicted to be toxic. Loss of function of the mutant HTT allele would likely exacerbate this toxicity. Remarkably, blocking cleavage at D586 in an HD mouse model has been shown to completely ameliorate the disease phenotype in mice[12].

bioRxiv preprint doi: https://doi.org/10.1101/129536; this version posted April 24, 2017. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

Therefore, we predict that the G553E mutation may be an important disease modifier and will lead to an earlier onset of disease in HD patients. It will be important to screen East Asian HD patients for the G553E mutation to validate the proteolytic consequences and phenotypic impact of the rs118005095 variant.

bioRxiv preprint doi: https://doi.org/10.1101/129536; this version posted April 24, 2017. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

Materials and Methods

Cloning

All HTT-YFP point mutations were generated using gBlocks, as described previously[21], except the D513E mutation was generated by Top Gene Technologies. All point mutations were confirmed by DNA sequencing (Operon Eurofins).

Click chemistry and fragment analysis

Post-translational myristoylation of HTT_{1-588}-YFP was detected as described previously[14]. Briefly, HeLa cells were transiently transfected using X-tremeGene (Roche) as per the manufacturer's directions. After 20h post-transfection, the media was exchanged with DMEM containing charcoal filtered FBS and cells were incubated with alkyne-myristate (Cayman Chemicals) for 30 mins prior to the addition of 1 µM staurosporine (STS) and 5 µg/mL of cycloheximide (CHX) to promote caspase activity and prevent synthesis of new protein, respectively. After 4h, cells were lysed in modified RIPA buffer. HTT_{1-588}-YFP was immunoprecipitated using goat anti-GFP (Eusera) and subjected to Click chemistry using azido biotin (Cayman Chemicals). Following SDS-PAGE on large 10% gels, post-translational myristoylation was detected by Western blot analysis using streptavidin Alexa680 (Invitrogen) and Licor. YFP was detected using rabbit anti-GFP (Eusera).

MTT assay

Cell viability was measured using the MTT assay (Sigma) as per the manufacturer's directions. Briefly, 20 h post-transfection. MTT was added to the cells for an additional 4 h. OD readings were normalized to WT fully cleavable HTT. The experiment was performed 6 times with 3 technical replicates per experiment for each condition.

bioRxiv preprint doi: https://doi.org/10.1101/129536; this version posted April 24, 2017. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

Acknowledgements

This work was supported by the CHDI Foundation and the Canadian Institutes of Health Research (CIHR 20R90174). DDOM was supported by postdoctoral fellowships from CIHR, Michael Smith Foundation for Health Research and the Bluma Tischler Fellowship from UBC. CK was supported by a doctoral scholarship from CIHR.

bioRxiv preprint doi: https://doi.org/10.1101/129536; this version posted April 24, 2017. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

References

1. Collaborative, T. H. D. & Group, R. A novel gene containing a trinucleotide repeat that is expanded and unstable on Huntington's disease chromosomes. *Cell* **72,** 971–983 (1993).

2. Dragatsis, I., Levine, M. S. & Zeitlin, S. Inactivation of Hdh in the brain and testis results in progressive neurodegeneration and sterility in mice. *Nat. Genet.* **26,** 300–306 (2000).

3. Godin, J. D. *et al.* Huntingtin Is Required for Mitotic Spindle Orientation and Mammalian Neurogenesis. *Neuron* **67,** 392–406 (2010).

4. Wong, Y. C. & Holzbaur, E. L. F. The Regulation of Autophagosome Dynamics by Huntingtin and HAP1 Is Disrupted by Expression of Mutant Huntingtin, Leading to Defective Cargo Degradation. *J. Neurosci.* **34,** 1293–1305 (2014).

5. Martin, D. D. O., Ladha, S., Ehrnhoefer, D. E. & Hayden, M. R. Autophagy in Huntington disease and huntingtin in autophagy. *Trends Neurosci.* **38,** 26–35 (2015).

6. Langbehn, D. R., Brinkman, R. R., Falush, D., Paulsen, J. S. & Hayden, M. R. A new model for prediction of the age of onset and penetrance for Huntington's disease based on CAG length. *Clin. Genet.* **65,** 267–277 (2004).

7. Bečanović, K. *et al.* A SNP in the HTT promoter alters NF-κB binding and is a bidirectional genetic modifier of Huntington disease. *Nat. Neurosci.* **18,** 807–816 (2015).

8. Consortium, G. M. of H. D. (GeM-H. Identification of Genetic Factors that Modify Clinical Onset of Huntington's Disease. *Cell* **162,** 516–526 (2015).

9. Ehrnhoefer, D. E., Sutton, L. & Hayden, M. R. Small Changes, Big Impact: Posttranslational Modifications and Function of Huntingtin in Huntington Disease. *Neurosci.* **17,** 475–492 (2011).

10. Warby, S. C. *et al.* Huntingtin phosphorylation on serine 421 is significantly reduced in the striatum and by polyglutamine expansion in vivo. *Hum. Mol. Genet.* **14,** 1569–1577 (2005).

11. El-Daher, M.-T. *et al.* Huntingtin proteolysis releases non-polyQ fragments that cause toxicity through dynamin 1 dysregulation. *EMBO J.* **34,** 2255–71 (2015).

12. Graham, R. K. *et al.* Cleavage at the Caspase-6 Site Is Required for Neuronal Dysfunction and Degeneration Due to Mutant Huntingtin. *Cell* **125,** 1179–1191 (2006).

13. Martin, D. D. O. *et al.* Tandem reporter assay for myristoylated proteins post-translationally (TRAMPP) identifies novel substrates for post-translational myristoylation: PKCε, a case study. *FASEB J.* **26,** 13–28 (2012).

14. Martin, D. D. O. *et al.* Identification of a post-translationally myristoylated autophagy-inducing domain released by caspase cleavage of huntingtin. *Hum. Mol. Genet.* **23,** 3166–3179 (2014).

15. Martin, D. D. O. & Hayden, M. R. Post-translational myristoylation at the cross roads of cell death, autophagy and neurodegeneration. *Biochem. Soc. Trans.* **43,** 229–234 (2015).

16. Kay, C. *et al.* Huntingtin Haplotypes Provide Prioritized Target Panels for Allele-specific Silencing in Huntington Disease Patients of European Ancestry. *Mol. Ther.* **23,** 1759–1771 (2015).

bioRxiv preprint doi: https://doi.org/10.1101/129536; this version posted April 24, 2017. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

17. Zuleta, A., Vidal, R. L., Armentano, D., Parsons, G. & Hetz, C. AAV-mediated delivery of the transcription factor XBP1s into the striatum reduces mutant Huntingtin aggregation in a mouse model of Huntington's disease. *Biochem. Biophys. Res. Commun.* **420,** 558–563 (2012).

18. Yap, M. C. *et al.* Rapid and selective detection of fatty acylated proteins using omega-alkynyl-fatty acids and click chemistry. *J. Lipid Res.* **51,** 1566–80 (2010).

19. Klionsky, D. J. *et al.* Guidelines for the use and interpretation of assays for monitoring autophagy (3rd edition). **8627,**

20. Ochaba, J. *et al.* Potential function for the Huntingtin protein as a scaffold for selective autophagy. *Proc. Natl. Acad. Sci.* **111,** 16889–16894 (2014).

21. Wong, B. K. Y. *et al.* Partial rescue of some features of Huntington Disease in the genetic absence of caspase-6 in YAC128 mice. *Neurobiol. Dis.* **76,** 24–36 (2015).

22. Choi, Y., Sims, G. E., Murphy, S., Miller, J. R. & Chan, A. P. Predicting the Functional Effect of Amino Acid Substitutions and Indels. *PLoS One* **7,** (2012).

23. Sim, N.-L. *et al.* SIFT web server: predicting effects of amino acid substitutions on proteins. *Nucleic Acids Res.* **40,** W452-7 (2012).

24. Hecht, M., Bromberg, Y. & Rost, B. Better prediction of functional effects for sequence variants. *BMC Genomics* **16,** S1 (2015).

bioRxiv preprint doi: https://doi.org/10.1101/129536; this version posted April 24, 2017. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

Table 1: Functional SNPs in gnomAD and 1000 Genomes Phase 3.

RSID	Chromosome Position (GRCh37.p13)	Reference	Alternate	Amino acid position (NP_002102.4)	Mutation	Allele Frequency (gnomAD)	Allele Frequency (1000 G)	PROVEAN[22] PREDICTION	SIFT[23] PREDICTION	SNAP[24] PREDICTION
rs362331	3215835	T	C	2311	Y/H	0.43680	0.44129	Neutral	Tolerated	Neutral
rs362272	3234980	G	A	2788	V/I	0.28390	0.21026	Neutral	Tolerated	Neutral
rs363125	3189547	C	A	1722	T/N	0.12750	0.18910	Neutral	Tolerated	Neutral
rs149109767	3230410	AGAG	A	2645	E/-	0.06067	0.02656	Deleterious	NA	----
rs35892913	3148570	G	A	1066	V/I	0.04200	0.02356	Neutral	Tolerated	Neutral
rs363075	3137674	G	A	895	G/R	0.04069	0.02376	Deleterious	Damaging	Effect
rs1143646	3148653	T	G	1093	I/M	0.03962	0.02236	Neutral	Damaging	Neutral
rs3025837	3174845	A	C	1387	N/H	0.01489	0.04153	Neutral	Tolerated	Neutral
rs34315806	3162034	C	T	1262	T/M	0.00837	0.02756	Neutral	Damaging	Neutral
rs151106561	3132038	A	G	627	Q/R	0.00523	0.00280	Neutral	Tolerated	Neutral
rs150003356	3189510	T	C	1710	C/R	0.00281	0.00100	Deleterious	Tolerated	Effect
rs3025843	3156038	A	G	1175	T/A	0.00256	0.00679	Neutral	Tolerated	Neutral
rs118005095	3129240	G	A	553	G/E	0.00242	0.00619	Neutral	Tolerated	Neutral
rs34389685	3133113	G	A	698	G/E	0.00190	0.00200	Deleterious	Damaging	Effect
rs186355914	3109095	C	T	233	A/V	0.00170	0.00020	Neutral	Damaging	Neutral
rs201702168	3221998	A	T	2446	E/D	0.00154	0.00140	Neutral	Tolerated	Neutral
rs375957195	3230428	C	A	2647	D/E	0.00153	0.00319	Neutral	Tolerated	Neutral
rs1065746	3148624	G	C	1084	D/H	0.00135	0.00020	Deleterious	Damaging	Effect
rs190593027	3144528	C	G	996	T/R	0.00110	0.00140	Neutral	Damaging	Neutral
rs201646519	3210567	T	C	2076	S/P	0.00018	0.00100	Neutral	Tolerated	Neutral
rs201760820	3134411	G	A	789	V/M	0.00047	0.00100	Neutral	Tolerated	Neutral

bioRxiv preprint doi: https://doi.org/10.1101/129536; this version posted April 24, 2017. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

FIGURES

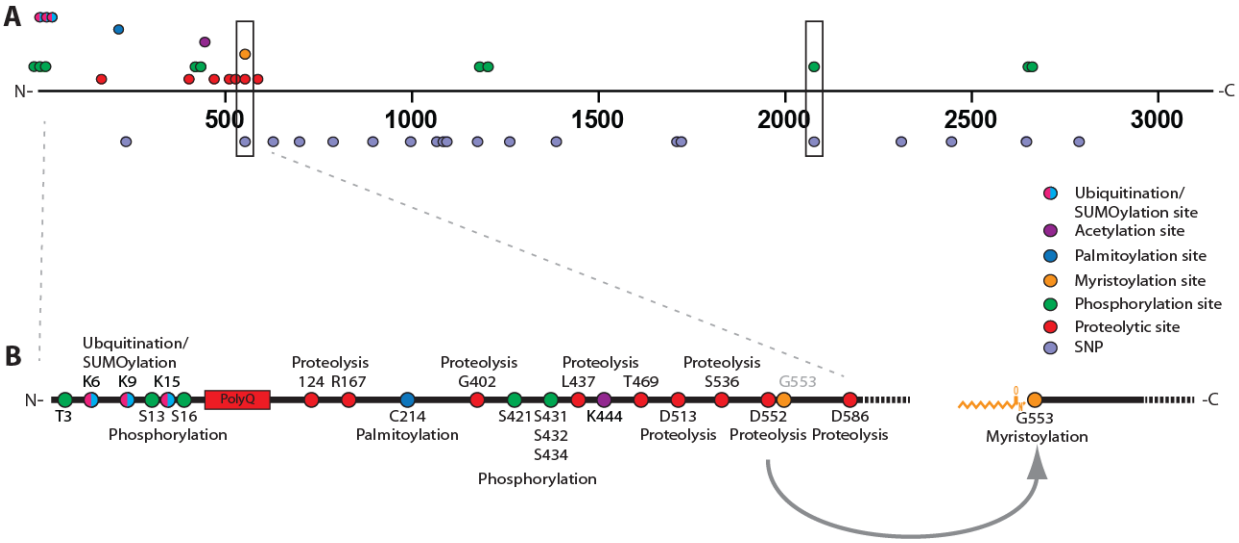

Figure 1 – Schematic representing where HTT PTMs and *HTT* missense SNPs intersect. (**A**) Linear map of *HTT* SNPs leading to missense mutations mapped to known HTT PTMs. The two missense mutations that directly intersect with myristoylation at G553 and phosphorylation at S2076, G553E and S2076P, are boxed. (**B**) PTMs within the first 586 amino acids of HTT are highlighted. Proteolytic caspase sites are indicated on the bottom while non-caspase mediated proteolytic sites are displayed on top. G553 is myristoylated following caspase cleavage at D552.

bioRxiv preprint doi: https://doi.org/10.1101/129536; this version posted April 24, 2017. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

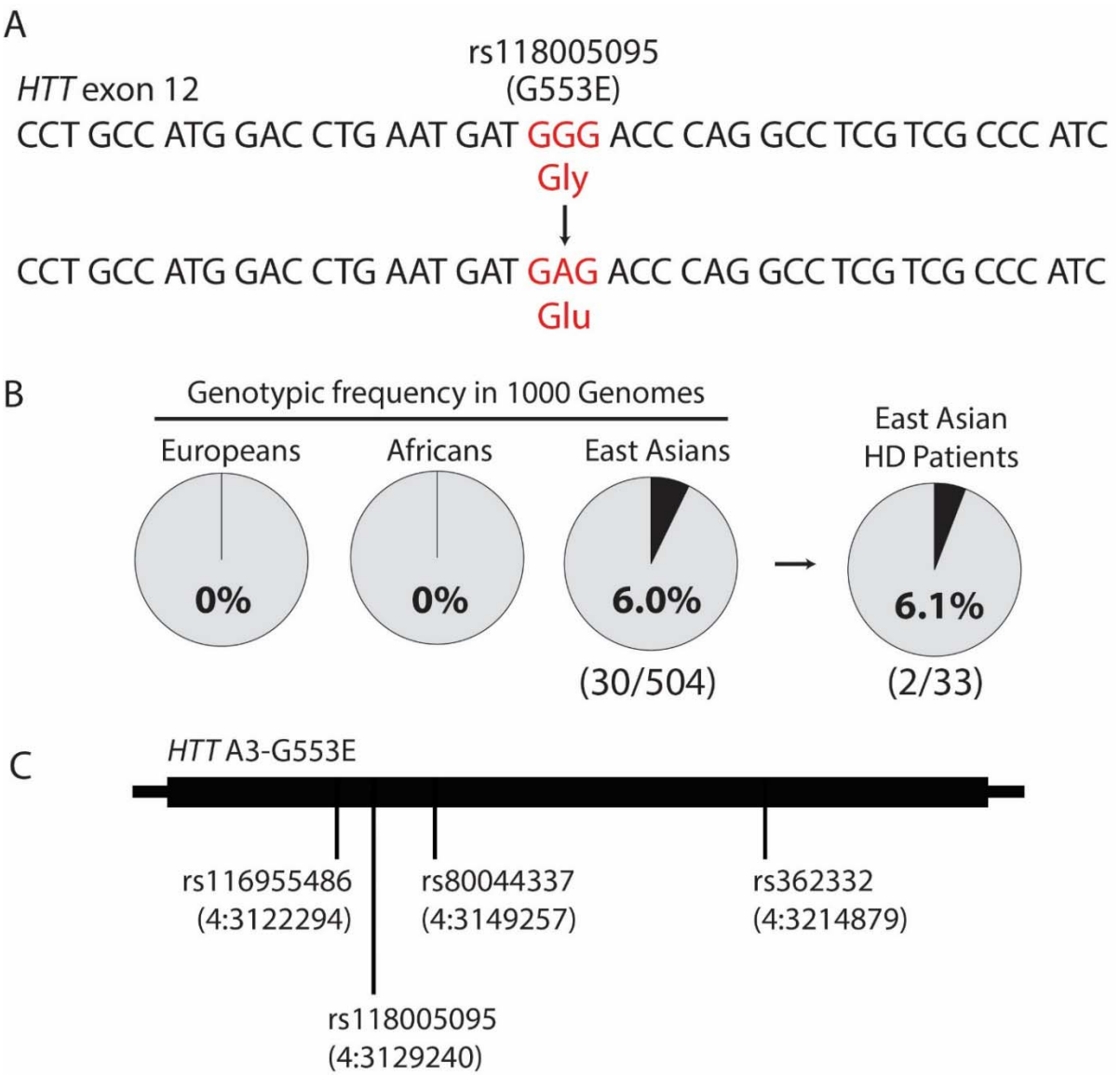

Figure 2 - The rs118005095 missense variant is a naturally occurring human SNP that alters the HTT amino acid sequence. (**A**) rs118005095 results in mutation of the HTT 553 glycine residue to glutamic acid. (**B**) rs118005095 occurs exclusively in populations of East Asian ancestry. (**C**) The rs118005095 variant is one of four SNPs defining a gene-spanning *HTT* haplotype in the East Asian population.

bioRxiv preprint doi: https://doi.org/10.1101/129536; this version posted April 24, 2017. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

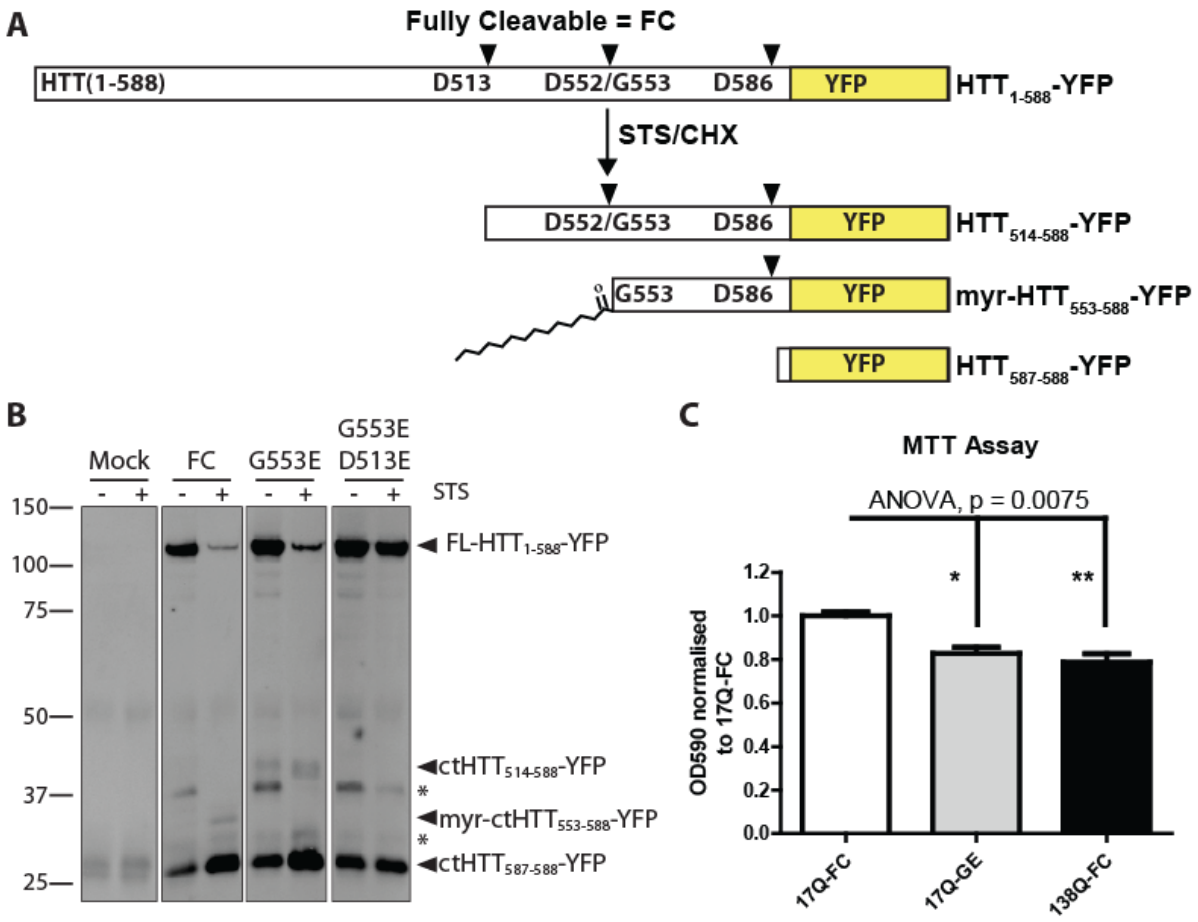

Figure 3 – The G553E substitution blocks post-translational myristoylation and alters proteolysis of wtHTT leading to toxicity. (**A**) Schematic representation of proteolytic cleavage of WT fully cleavable (FC) HTT_{1-588}-YFP. HTT is proteolysed by caspases at D586 and D552 and post-translationally myristoylated at G553. (**B**) HeLa cells were transfected with WT HTT_{1-588}-YFP bearing the indicated point mutations. Proteolysis was induced with staurosporine (STS) and cycloheximide. In the presence of the G553E mutation, a higher molecular weight band was detected that was blocked when D513 was mutated to E, confirming the D513 caspase cleavage site. Similarly, the D586E mutation completely abrogated the generation of the ~26 kDa band corresponding to caspase cleavage at D586. * denotes unknown or non-specific bands. (**C**) HeLa cells were transfected with the indicated constructs and cell viability was measured using the MTT assay (Kruskal-Wallis ANOVA, error bars represent SEM. *-p<0.05, ** - p<0.01).