## Activations of Deep Convolutional Neural Network are Aligned with Gamma Band Activity of Human Visual Cortex

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Previous work demonstrated a direct correspondence between the hierarchy of the human visual areas and layers of deep convolutional neural networks (DCNN) trained on visual object recognition. We used DCNNs to investigate which frequency bands carry feature transformations of increasing complexity along the ventral visual pathway. By capitalizing on intracranial depth recordings from 100 patients and 11293 electrodes we assessed the alignment between the DCNN and signals at different frequency bands in different time windows. We found that activity in low and high gamma bands was aligned with the increasing complexity of visual feature representations in the DCNN. These findings show that activity in the gamma band is not only a correlate of object recognition, but carries increasingly complex features along the ventral visual pathway. These results demonstrate the potential that modern artificial intelligence algorithms have in advancing our understanding of the brain.

#### **Significance Statement** 1

Recent advances in the field of artificial intelligence have revealed 2 principles about neural processing, in particular about vision. 3 Previous works have demonstrated a direct correspondence 4 between the hierarchy of human visual areas and layers of deep 5 convolutional neural networks (DCNNs), suggesting that DCNN 6 is a good model of visual object recognition in primate brain. 7 Studying intracranial depth recordings allowed us to extend pre-8 vious works by assessing when and at which frequency bands the 9 10 activity of the visual system corresponds to the DCNN. Our key finding is that signals in gamma frequencies along the ventral 11 visual pathway are aligned with the layers of DCNN. Gamma 12 frequencies play a major role in transforming visual input to 13 coherent object representations. 14

## Introduction

Biological visual object recognition is mediated by a hierarchy 2 of increasingly complex feature representations along the ventral visual stream (DiCarlo et al., 2012). Intriguingly, these transformations are matched by the hierarchy of transformations learned 5 by deep convolutional neural networks (DCNN) trained on natu-6 ral images (Güçlü and van Gerven, 2015). It has been shown that 7 DCNN provides the best model out of a wide range of neurosci-8 entific and computer vision models for the neural representation 9 of visual images in high-level visual cortex of monkeys (Yamins et 10 al., 2014) and humans (Khaligh-Razavi and Kriegeskorte, 2014). 11 Other studies have demonstrated with fMRI a direct correspon-12 dence between the hierarchy of the human visual areas and layers 13 of the DCNN (Güçlü and van Gerven, 2015; Eickenberg et al., 14 2016; Seibert et al., 2016; Cichy et al., 2016b). In sum, the 15 increasing feature complexity of the DCNN corresponds to the 16

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increasing feature complexity occurring in visual object recog nition in the primate brain (Kriegeskorte, 2015; Yamins and
 DiCarlo, 2016).

However, fMRI based studies only allow one to localize object 4 recognition in space, but biological visual object recognition is 5 also specific in time and frequency. With time-resolved magne-6 toencephalography (MEG) recordings it has been demonstrated 7 that the correspondence between the DCNN and neural signals 8 peaks in the first 200 ms (Cichy et al., 2016b; Seeliger et al., 9 2017). Here we test the remaining dimension: that biological 10 visual object recognition is also specific to certain frequencies. 11 In particular, there is a long-standing hypothesis that especially 12 gamma band (30 - 150 Hz) signals are crucial for object recog-13 nition (Singer and Gray, 1995; Singer, 1999; Fisch et al., 2009; 14 Tallon-Baudry et al., 1997; Tallon-Baudry and Bertrand, 1999; 15 Lachaux et al., 1999; Wvart and Tallon-Baudry, 2008; Lachaux 16 et al., 2005; Vidal et al., 2006; Herrmann et al., 2004; Hipp et 17 al., 2011; Gaillard et al., 2009; Srinivasan et al., 1999; Levy et 18 al., 2015). Hence, if DCNN capture biological object recognition 19 there should be a correspondence between the DCNN layers and 20 21 gamma signals along the ventral visual pathway.

22 To empirically evaluate the specific role of gamma frequency in visual object recognition we assessed the alignment between 23 the responses of layers of the DCNN and the neural signals in five 24 25 distinct frequency bands and three time windows along the areas 26 constituting the ventral visual pathway. Based on the previous findings we expected that: 1) mainly gamma frequencies should 27 be aligned to the DCNN; 2) the correspondence between the 28 DCNN and gamma should be confined to early time windows; 3) 29 the correspondence between gamma and the DCNN layers should 30 be restricted to visual areas. In order to test these predictions 31 we capitalized on direct intracranial depth recordings from 100 32 33 patients with epilepsy and a total of 11293 electrodes implanted 34 throughout the cerebral cortex.

35 Studying the alignment between the DCNN and gamma frequencies would also help to elucidate the role of gamma band 36 signals in object recognition. The classic view is that gamma 37 band activity signals the emergence of coherent object represen-38 tations (Singer and Gray, 1995; Singer, 1999; Fisch et al., 2009). 39 40 However, it is possible that gamma frequencies carry feature transformations of increasing complexity instead of reflecting 41 solely the final product of object recognition. Suggestive evidence 42 for this view is provided by studies demonstrating that feedfor-43 ward activity from lower to higher visual areas is carried by the 44 gamma frequencies along the ventral visual pathway (Van Kerko-45 46 erle et al., 2014; Bastos et al., 2015; Michalareas et al., 2016). The 47 existence of quantifiable increase of feature complexity along the layers of DCNN allows one to use the DCNN as a computational 48 model to assess whether signals in the gamma frequency indeed 49 reflect such gradual transformations. 50

We observed that activity in the gamma range along the ven-51 tral pathway is statistically significantly aligned with the activity 52 along the layers of DCNN: gamma (31 - 150 Hz) activity in the 53 early visual areas correlates with the activity of early layers of 54 DCNN, while the gamma activity of higher visual areas is bet-55 ter captured by the higher layers of the DCNN. We also found 56 that neural activity in the theta range (5 - 8 Hz) throughout the 57 visual hierarchy correlated with higher layers of DCNN. 58

## **Materials and Methods**

Our methodology involves four major steps described in the fol-2 lowing subsections. In "Patients and Recordings" we describe 3 the visual recognition task and data collection. In "Processing 4 of Neural Data" we describe the artifact rejection, extraction of 5 spectral features and the electrode selection processes. "Process-6 ing of DCNN Data" shows how we extract activations of artificial 7 neurons of DCNN that occur in responses to the same images as 8 were shown to human subjects. In the final step we map neural 9 activity to the layers of DCNN using representational similarity 10 analysis. See Figure 1 for the illustration of the analysis workflow. 11

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## Patients and Recordings

100 patients of either gender with drug-resistant partial epilepsy 13 and candidates for surgery were considered in this study and 14 recruited from Neurological Hospitals in Grenoble and Lyon 15 (France). All patients were stereotactically implanted with multi-16 lead EEG depth electrodes (DIXI Medical, Besançon, France). 17 All participants provided written informed consent, and the 18 experimental procedures were approved by local ethical commit-19 tee of Grenoble hospital (CPP Sud-Est V 09-CHU-12). Recording 20 sites were selected solely according to clinical indications, with 21 no reference to the current experiment. All patients had normal 22 or corrected to normal vision. 23

### Electrode Implantation

Eleven to 15 semi-rigid electrodes were implanted per patient. 25 Each electrode had a diameter of 0.8 mm and was comprised 26 of 10 or 15 contacts of 2 mm length, depending on the target 27 region, 1.5 mm apart. The coordinates of each electrode con-28 tact with their stereotactic scheme were used to anatomically 29 localize the contacts using the proportional atlas of Talairach 30 and Tournoux (Talairach and Tournoux, 1993), after a linear 31 scale adjustment to correct size differences between the patients 32 brain and the Talairach model. These locations were further 33 confirmed by overlaying a post-implantation CT scan (show-34 ing contact sites) with a pre-implantation structural MRI with 35 VOXIM<sup>®</sup> (IVS Solutions, Chemnitz, Germany), allowing direct 36 visualization of contact sites relative to brain anatomy. 37

All patients voluntarily participated in a series of short exper-38 iments to identify local functional responses at the recorded sites 39 (Vidal et al., 2010). The results presented here were obtained 40 from a test exploring visual recognition. All data were recorded 41 using approximately 120 implanted depth electrode contacts per 42 patient with a sampling rates of 512 Hz, 1024 Hz or 2048 Hz. For 43 the current analysis all recordings were downsampled to 512 Hz. 44 Data were obtained in a total of 11293 recording sites. 45

## Stimuli and Task

The visual recognition task lasted for about 15 minutes. Patients 47 were instructed to press a button each time a picture of a fruit 48 appeared on screen (visual oddball paradigm). Non-target stimuli consisted of pictures of objects of eight possible categories: 50 houses, faces, animals, scenes, tools, pseudo words, consonant 51 strings, and scrambled images. The target stimuli and last three categories were not included in this analysis. All the included 53

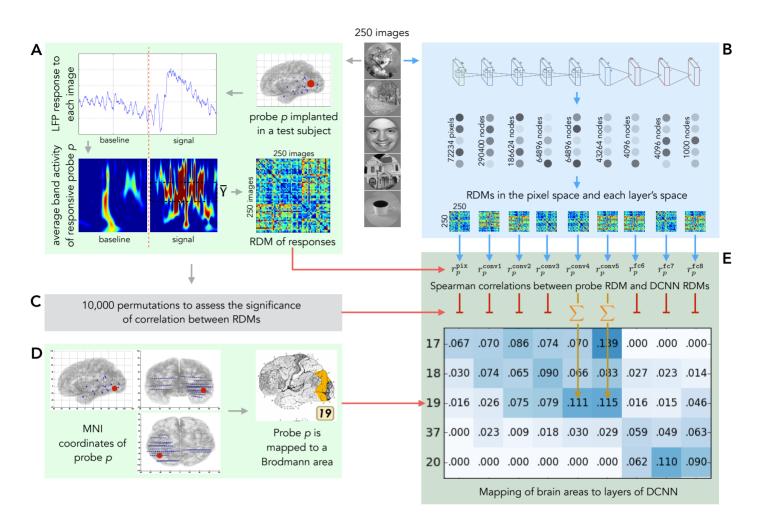


Figure 1 Overview of the analysis pipeline. 250 natural images are presented to human subjects (panel A) and to an artificial vision system (panel B). The activities elicited in these two systems are compared in order to map regions of human visual cortex to layers of deep convolutional neural networks (DCNNs). A: LFP response of each of 11293 electrodes to each of the images is converted into the frequency domain. Activity evoked by each image is compared to the activity evoked by every other image and results of this comparison are presented as a representational dissimilarity matrix (RDM). B: Each of the images is shown to a pre-trained DCNN and activations of each of the layers are extracted. Each layer's activations form a representation space, in which stimuli (images) can be compared to each other. Results of this comparison are summarized as a RDM for each DCNN layer. C: Subject's intracranial responses to stimuli are randomly reshuffled and the analysis depicted in panel A is repeated 10000 times to obtain 10000 random RDMs for each electrode. D: Each electrode's MNI coordinates are used to map the electrodes). E: Spearman's rank correlation is computed between the true (non-permuted) RDM of neural responses and RDMs of each layer of DCNN. Also 10000 scores are computed with the random RDM for each electrode-layer pair to assess the significance of the true correlation score. If the score obtained with the true RDM is significant (the value of p < 0.001 is estimated by selecting a threshold such that none of the probes would pass it on the permuted data), then the score is added to the mapping matrix. The procedure is repeated for each electrode and the correlation scores are summed and normalized by the number of electrodes per Brodmann area. The resulting mapping matrix shows the alignment between the consecutive areas of the ventral stream and layers of DCNN.

stimuli had the same average luminance. All categories were pre-1 sented within an oval aperture (illustrated on Figure 1). Stimuli 2 were presented for a duration of 200 ms every 1000 - 1200 ms in 3 series of 5 pictures interleaved by 3-s pause periods during which 4 5 patients could freely blink. Patients reported the detection of a 6 target through a right-hand button press and were given feedback of their performance after each report. A 2-s delay was 7 placed after each button press before presenting the follow-up 8

9 stimulus in order to avoid mixing signals related to motor action

with signals from stimulus presentation. Altogether, we measured responses to 250 natural images. Each image was presented only once.

### **Processing of Neural Data**

The final dataset consists of 2823250 local field potential (LFP) recordings – 11293 electrode responses to 250 stimuli.

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To remove the artifacts the signals were linearly detrended 1 and the recordings that contained values  $\geq 10\sigma_{images}$ , where 2  $\sigma_{images}$  is the standard deviation of responses (in the time win-3 dow from -500 ms to 1000 ms) of that particular probe over 4 all stimuli, were excluded from data. All electrodes were re-5 referenced to a bipolar reference. The signal was segmented in 6 the range from -500 ms to 1000 ms, where 0 marks the moment 7 8 when the stimulus was shown. The -500 to -100 ms time window served as the baseline. There were three time windows in g which the responses were measured: 50 - 250 ms, 150 - 350 ms 10 and 250 - 450 ms. 11

We analyzed five distinct frequency bands:  $\theta$  (5 – 8 Hz),  $\alpha$ 12  $(9 - 14 \text{ Hz}), \beta (15 - 30 \text{ Hz}), \gamma (31 - 70 \text{ Hz}) \text{ and } \Gamma (71 - 150 \text{ Hz}).$ 13 To quantify signal power modulations across time and frequency 14 we used standard time-frequency (TF) wavelet decomposition 15 (Daubechies, 1990). The signal s(t) is convoluted with a complex 16 Morlet wavelet  $w(t, f_0)$ , which has Gaussian shape in time  $(\sigma_t)$ 17 and frequency  $(\sigma_f)$  around a central frequency  $f_0$  and defined 18 by  $\sigma_f = 1/2\pi\sigma_t$  and a normalization factor. In order to achieve 19 good time and frequency resolution over all frequencies we slowly 20 21 increased the number of wavelet cycles with frequency  $(\frac{f_0}{\sigma_f})$  was set to 6 for high and low gamma, 5 for beta, 4 for alpha and 3 for 22 theta). This method allows obtaining better frequency resolution 23 24 than by applying a constant cycle length (Delorme and Makeig, 2004). The square norm of the convolution results in a time-25 varying representation of spectral power, given by:  $P(t, f_0) =$ 26  $|w(t, f_0)s(t)|^2$ . 27

Further analysis was done on the electrodes that were respon-28 sive to the visual task. We assessed neural responsiveness of 29 an electrode separately for each region of interest - for each 30 frequency band and time window we compared the average 31 post-stimulus band power to the average baseline power with 32 a Wilcoxon signed-rank test for matched-pairs. All p-values from 33 this test were corrected for multiple comparisons across all elec-34 trodes with a false discovery rate (FDR) procedure (Genovese et 35 al., 2002). In the current study we deliberately kept only pos-36 itively responsive electrodes, leaving the electrodes where the 37 post-stimulus band power was significantly weaker than the aver-38 age baseline power for future work. Table 1 contains the numbers 39 of electrodes that were used in the final analysis in each of 15 40 regions of interest across the time and frequency domains. 41

	$\theta$		1.	/	
50-250  ms	1299	709	269	348	504
$150-350~\mathrm{ms}$	1689	783	260	515	745
50 - 250  ms 150 - 350  ms 250 - 450  ms	1687	802	304	555	775

Table1Number of positively responsive electrodes in each of the15 regions of interest in a time-resolved spectrogram.

Each electrode's MNI coordinates were mapped to a corre-42 sponding Brodmann brain area (Brodmann, 1909) using Brod-43 mann area atlas contained in MRICron (Rorden, 2007) software. 44 To summarize, once the neural signal processing pipeline is 45 complete, each electrode's response to each of the stimuli is rep-46 resented by one number – the average band power in a given 47 time window normalized by the baseline. The process is repeated 48 49 independently for each time-frequency region of interest.

#### Processing of DCNN Data

We feed the same images that were shown to the test subjects to a deep convolutional neural network (DCNN) and obtain activations of artificial neurons (nodes) of that network. We use Caffe (Jia et al., 2014) implementation of AlexNet (Krizhevsky et al., 2012) architecture (see Figure 7) trained on ImageNet (Russakovsky et al., 2015) dataset to categorize images into 1000 classes. Although the image categories used in our experiment are not exactly the same as the ones in the ImageNet dataset, they are a close match and DCNN is successful in labelling them.

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The architecture of the AlexNet artificial network can be seen 11 on Figure 7. It consists of 9 layers. The first is the input layer, 12 where one neuron corresponds to one pixel of an image and acti-13 vation of that neuron on a scale from 0 to 1 reflects the color 14 of that pixel: if a pixel is black, the corresponding node in the 15 network is not activated at all (value is 0), while a white pixel 16 causes the node to be maximally activated (value 1). After the 17 input layer the network has 5 convolutional layers referred to 18 as conv1-5. A convolutional layer is a collection of filters that 19 are applied to an image. Each filter is a small image that rep-20 resents a particular visual pattern. A filter is applied to every 21 possible position on an input image and if the underlying patch 22 of an image coincides with the pattern that the filter repre-23 sents, the filter becomes activated and translates this activation 24 to the artificial neuron in the next layer. That way, nodes of 25 conv1 tell us where on the input image each particular visual 26 pattern occurred. Hierarchical structure of convolutional layers 27 gives rise to the phenomenon we are investigating in this work 28 increase of complexity of visual representations in each sub-29 sequent layer of the visual hierarchy: in both the biological and 30 artificial systems. Convolutional layers are followed by 3 fully-31 connected layers (fc6-8). Each node in a fully-connected layer 32 is, as the name suggests, connected to every node of the previous 33 layer allowing the network to decide which of those connections 34 are to be preserved and which are to be ignored. 35

For each of the images we store the activations of all nodes of 36 DCNN. As the network has 9 layers we obtain 9 representations 37 of each image: the image itself (referred to as layer 0) in the pixel 38 space and the activation values of each of the layers of DCNN. 39 See pane B of figure 1 for the cardinalities of those feature spaces. 40

#### Mapping Neural Activity to Layers of DCNN

Once we extracted the features from both neural and DCNN 42 responses our next goal was to compare the two and use a simi-43 larity score to map the brain area where a probe was located to a 44 layer of DCNN. By doing that for every probe in the dataset we 45 obtained cross-subject alignment between visual areas of human 46 brain and layers of DCNN. There are multiple deep neural net-47 work architectures trained to classify natural images. Our choice 48 of AlexNet does not imply that this particular architecture cor-49 responds best to the hierarchy of visual layers of human brain. It 50 does, however, provide a comparison for hierarchical structure of 51 human visual system and was selected among other architectures 52 due to its relatively small size and thus easier interpretability. 53

Recent studies comparing the responses of visual cortex with the activity of DCNN have used two types of mapping methods. The first type is based on linear regression models that predict 55

neural responses from DCNN activations (Güçlü and van Gerven,
 2015). The second is based on representational similarity analy sis (RSA) (Kriegeskorte et al., 2008). We used RSA to compare

4 distances between stimuli in the neural response space and in the

5 DCNN activation space (Cichy et al., 2016a).

#### 6 Representational Dissimilarity Matrices

We built a representation dissimilarity matrix (RDM) of size 7 number of stimuli  $\times$  number of stimuli (in our case  $250 \times 250$ ) 8 g for each of the probes and each of the layers of DCNN. Given a matrix  $RDM^{feature space}$  a value  $RDM^{feature space}_{iii}$  in the *i*th 10 row and *j*th column of the matrix shows the Euclidean distance 11 between the vectors  $\mathbf{v}_i$  and  $\mathbf{v}_i$  that represent images *i* and *j* 12 respectively in that particular feature space. Note that the pre-13 14 processed neural response to an image in a given frequency band and time window is a scalar, and hence correlation distance is not 15 applicable. Also, given that DCNNs are not invariant to the scal-16 ing of the activations or weights in any of its layers, we preferred 17 18 to use closeness in Euclidean distance as a more strict measure of similarity. In our case there are 10 different features spaces 19 in which an image can be represented: the original pixel space, 20 8 feature spaces for each of the layers of the DCNN and one 21 space where an image is represented by the preprocessed neural 22 response of probe p. For example, to analyze region of interest 23 of high gamma in 50 - 250 ms time window we computed 504 24 25 RDM matrices on the neural responses – one for each positively 26 responsive electrode in that region of interest (see Table 1), and 27 9 RDM matrices on the activations of the layers of DCNN. A pair frequency band and a time window, such as "high gamma 28 in 50-250 ms window" is referred to as region of interest in this 29 30 work.

#### 31 Representational Similarity Analysis

The second step was to compare the  $RDM^{probe p}$  of each probe p to RDMs of layers of DCNN. We used Spearman's rank correlation as measure of similarity between the matrices:

$$\rho_{\text{layer }l}^{\text{probe }p} = \text{Spearman}(\text{RDM}^{\text{probe }p}, \text{RDM}^{\text{layer }l}).$$
(1)

As a result of comparing RDM<sup>probe p</sup> with every RDM<sup>layer l</sup> we obtain a vector with 9 scores:  $(\rho_{\text{pixels}}, \rho_{\text{conv1}}, \ldots, \rho_{\text{fc8}})$  that serves as a distributed mapping of probe p to the layers of DCNN (see pane E of Figure 1). The procedure is repeated independently for each probe in each region of interest.

## 40 Statistical significance and controls

To assess the statistical significance of the correlations between 41 the RDM matrices we run a permutation test. In particular, we 42 reshuffled the vector of brain responses to images 10000 times, 43 each time obtaining a dataset where the causal relation between 44 the stimulus and the response is destroyed. On each of those 45 datasets we ran the analysis and obtained Spearman's rank cor-46 relation scores. To determine score's significance we compared 47 the score obtained on the original (unshuffled) data with the dis-48 tribution of scores obtained with the surrogate data. If the score 49 obtained on the original data was bigger than value obtained on 50 51 the surrogate sets with p < 0.001 significance we considered the

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Figure 2 Mapping of the activity in Brodmann areas to DCNN layers. Underlying data comes from the activity in low gamma (31-70 Hz, subfigures A and C) and high gamma (71-150 Hz, subfigures B and D) bands in 150-350 ms time window. C and D are subselection of the areas that constitute ventral stream: 17, 18, 19, 37, 20. There are two important observations to made out of this plot: a) statistically significant neural responses are specific to visual areas b) the alignment between the ventral stream and layer of DCNN is clearly visible. Area 0 contains the regions of the brain not mapped by the atlas. The numbers on the left of each panel show the number of significantly correlating probes in each area out of the total number of responsive probes in that area.

1 score to be significantly different. The threshold of p = 0.001 is 2 estimated by selecting such a threshold that on the surrogate

 $_{3}$   $\,$  data none of the probes would pass it.

To size the effect caused by training artificial neural network on natural images we performed a control where the whole analysis pipeline depicted on figure 1 is repeated using activations of a network that was not trained – its weights are randomly

sampled from a Gaussian distribution  $\mathcal{N}(0, 0.01)$ .

## 9 Quantifying properties of the mapping

<sup>10</sup> To evaluate the results quantitatively we devised a set of mea-<sup>11</sup> sures specific to our analysis. *Volume* is the total sum of <sup>12</sup> significant correlations (see Equation 1) between the probes in a <sup>13</sup> subset of brain areas A and DCNN layers L:

$$V_{\text{layers }L}^{\text{areas }A} = \sum_{a \in A} \sum_{l \in L} \sum_{p \in S_l^a} \rho_{\text{layer }l}^{\text{probe }p}, \qquad (2)$$

where A is a subset of brain areas, L is a subset of layers, and  $S_l^a$  is the set of all probes in area a that significantly correlate with layer l.

17 We express volume of visual activity as

$$V_{\text{all layers}}^{\{17,18,19,37,20\}},$$
 (3)

which shows the total sum of correlation scores between all layers
of the network and the Brodmann areas that are located in the
ventral stream: 17, 18, 19, 37, and 20.

Visual specificity of activity is the ratio of volume in visual
areas and volume in all areas together, for example visual specificity of all of the activity in the ventral stream that significantly
correlates with any of layers of DCNN is

$$S_{\text{all layers}}^{\{17,18,19,37,20\}} = \frac{V_{\text{all layers}}^{\{17,18,19,37,20\}}}{V_{\text{all layers}}^{\text{all layers}}}$$
(4)

The measures so far did not take into account hierarchy of the ventral stream nor the hierarchy of DCNN. The following two measures are the most important quantifiers we rely on in presenting our results and they do take hierarchical structure into account.

The ratio of complex visual features to all visual features is defined as the total volume mapped to layers conv5, fc6, fc7 divided by the total volume mapped to layers conv1, conv2, conv3, conv5, fc6, fc7:

$$C^{\text{areas }A} = \frac{V_{\text{conv5,fc6,fc7}}^{\text{areas }A}}{V_{\text{conv1,conv2,conv3,conv5,fc6,fc7}}^{\text{areas }A}}.$$
(5)

Note that for this measure layers conv4 and fc8 are omitted: layer conv4 is considered to be the transition between the layers with low and high complexity features, while layer fc8 directly represents class probabilities and does not carry visual representations of the stimuli (if only on very abstract level).

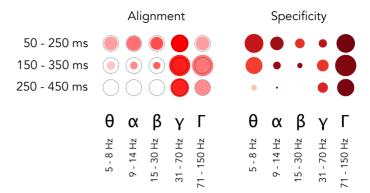


Figure 3 Overall relative statistics of brain responses across frequency bands and time windows. The left panel shows the alignment between visual brain areas and DCNN layers (see Equation 6). The color indicates the correlation value ( $\rho$ ) while the size of the marker shows the logarithm (so that not significant results are still visible on the plot) of inverse of the statistical significance of the correlation, dotted circle indicates p = 0.0003(3) – the Bonferroni-corrected significance threshold level of 0.005. The right panel shows whether activity in a region of interest is specific to visual areas (see Equation 4): intensive red means that most of the activity in that band and time window happened in visual areas, size of the marker indicates total volume (Equation 2) of activity in all areas. The maximal size of a marker is defined by the biggest marker on the figure.

Finally, the *alignment* between the activity in the visual areas and activity in DCNN is estimated as Spearman's rank correlation between the vector of electrode assignments to visual areas and the vector of electrode assignments to DCNN layers:

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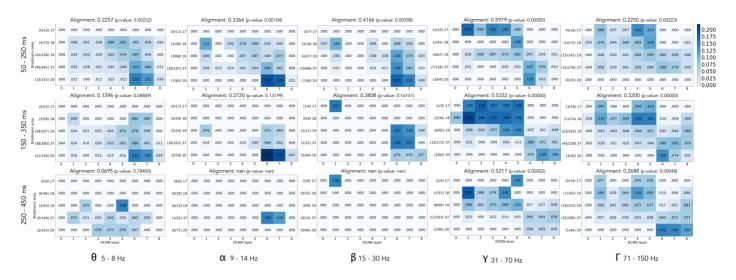
$$\rho = \text{Spearman} \begin{pmatrix}
\text{Brodmann areas with sig-} & \text{DCNN} & \text{layers where} \\
\text{nificantly correlating probes} & \text{significantly correlating} \\
\text{ordered by the hierarchy of,} & \text{probes are mapped,} \\
\text{the ventral stream: BA17, ordered by the hierarchy} \\
\text{BA18, BA19, BA37, BA20} & \text{of DCNN architecture}
\end{cases}$$
(6)

We note that although the hierarchy of the ventral stream is usu-5 ally not defined through the progression of Brodmann areas, such 6 ordering nevertheless provides a reasonable approximation of the 7 real hierarchy (Lerner et al., 2001; Grill-Spector and Malach, 8 2004). As both the ventral stream and the hierarchy of layers a in DCNN have an increasing complexity of visual representa-10 tions, the relative ranking within the biological system should 11 coincide with the ranking within the artificial system. Based on 12 the recent suggestion that significance levels should be shifted 13 to 0.005 (Dienes et al., 2017) and after Bonferroni-correcting for 14 15 time-frequency windows we accepted alignment as significant 15 when it passed p < 0.0003(3). 16

## Results

## Increasing complexity of visual representations is captured by activity in gamma band

We tested the hypothesis that gamma activity carries increasingly complex features along the ventral stream. To that end we assessed the alignment of neural activity in different frequency bands and time windows to the activity of layers of a DCNN. 23



**Figure 4** Mapping of activity in visual areas to activations of layers of DCNN across five frequency bands and three time windows. The alignment score is computed as Spearman's rank correlation between electrode assignment to Brodmann areas and electrode assignment to DCNN layers (Equation 6). The numbers on the left of each subplot show the number of significantly correlating probes in each area out of the total number of responsive probes in that area.

In particular, we used RSA to compare the representational 1 geometry of different DCNN layers and the activity patterns of 2 different frequency bands of single electrodes (see Figure 1). We 3 consistently found that signals in low gamma (31 - 70 Hz) fre-4 quencies across all time windows and high gamma (71 - 150 Hz)5 frequencies in 150 - 350 ms window are aligned with the DCNN 6 in a specific way: increase of the complexity of features along the 7 layers of the DCNN was matched by the transformation in the 8 representational geometry of responses to the stimuli along the g ventral stream. In other words, the lower and higher layers of 10 11 the DCNN explained gamma band signals from earlier and later 12 visual areas, respectively.

Figure 2 illustrates assignment of neural activity in low 13 gamma band (panel A) and high gamma band (panel B) to Brod-14 mann areas and layers of DCNN. As one can see most of the 15 activity was assigned to visual areas (areas 17, 18, 19, 37, 20). 16 Focusing on visual areas (panels C, D) revealed a diagonal trend 17 that illustrated the alignment between ventral stream and layers 18 of DCNN. Our findings across all subjects, time windows and 19 frequency bands are presented in table 2 and summarized on the 20 left panel of figure 3. The results in table 2 show the comparison 21 of alignment between DCNN and brain areas with both random 22 and pre-trained networks. We can see that training a network to 23 24 classify natural images drastically increases the alignment score 25  $\rho$  and its significance. We note that the alignment in the gamma bands is also present at the single-subject level as can be seen in 26 Figure 6. 27

Apart from the alignment we looked at the total amount of 28 correlation and its specificity to visual areas. On the right panel 29 of Figure 3 we can see that the volume of significantly correlating 30 activity was highest in the high gamma range. Remarkably, 97% 31 of that activity was located in visual areas, which is confirmed 32 by figure 2 where we see that in the gamma range only a few 33 electrodes were assigned to Brodmann areas that are not part of 34 35 the ventral stream.

		Alignment	with	Alignme		
		layers of r	andomly	layers o		
		initialized		trained o		
Band	Window	ho	p-value	$\rho$	p-value	
$\theta$	$50\text{-}250~\mathrm{ms}$	0.0632	0.71	0.2257	0.00231575	*
$\theta$	$150\text{-}350~\mathrm{ms}$	-0.1013	0.59	0.1396	0.08848501	
$\theta$	$250\text{-}450~\mathrm{ms}$	0.1396	0.59	0.0695	0.78400416	
$\alpha$	$50-250 \mathrm{\ ms}$	-0.2411	0.32	0.3366	0.00103551	*
$\alpha$	$150\text{-}350~\mathrm{ms}$	0.0000	1.00	0.2720	0.13199463	
$\alpha$	$250\text{-}450~\mathrm{ms}$	-	-	-	-	
$\beta$	50-250  ms	_	-	0.4166	0.00397929	
$\beta$	$150\text{-}350~\mathrm{ms}$	_	-	0.3808	0.16141286	
$\beta$	$250\text{-}450~\mathrm{ms}$	—	-	-	-	
$\gamma$	$50-250 \mathrm{\ ms}$	0.1594	0.62	0.5979	0.00004623	***
$\gamma$	$150\text{-}350~\mathrm{ms}$	-0.1688	0.34	0.5332	0.00000059	***
$\gamma$	$250\text{-}450~\mathrm{ms}$	-0.1132	0.56	0.5217	0.00001624	***
Г	$50-250 \mathrm{\ ms}$	0.0869	0.42	0.2259	0.00222940	*
Г	$150\text{-}350~\mathrm{ms}$	-0.0053	0.96	0.3200	0.00000051	***
Г	$250\text{-}450~\mathrm{ms}$	-0.1361	0.33	0.2688	0.00047999	*

**Table 2** Alignment  $\rho$  score and significance for all 15 regions of interest. \* indicates the alignments that pass p-value threshold of 0.05 Bonferroni-corrected to 0.003(3) and \*\*\* the ones that pass 0.005 (Dienes et al., 2017) Bonferroni-corrected to 0.0003(3). Note how the values differ between random (control) network and a network trained on natural images. Visual representation of alignment and significance is given on the left pane of Figure 3.

#### Activity in other frequency bands

To test the specificity of gamma frequency in visual object recognition, we assessed the alignment between the DCNN and other frequencies. The detailed mapping results for all frequency bands

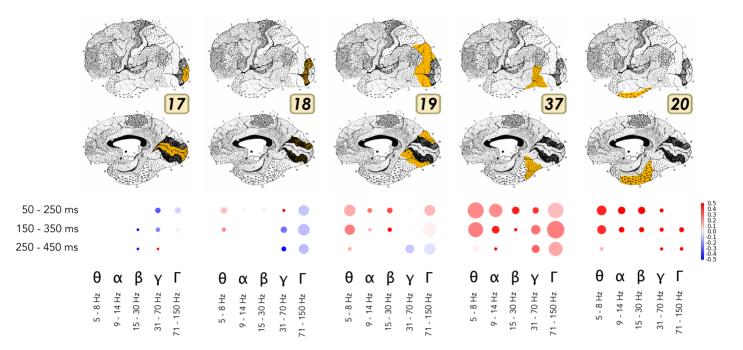


Figure 5 Area-specific analysis of volume of neural activity and complexity of visual features represented by that activity. Size of the marker shows the sum of correlation coefficients between the area and DCNN for each particular band and time window. Color codes the ratio of complex visual features to simple visual features, i.e. the comparison between the activity that correlates with the higher layers (conv5, fc6, fc7) of DCNN to the lower layers (conv1, conv2, conv3). Intensive red means that the activity was correlating more with the activity of higher layers of DCNN, while the intensive blue indicates the dominance of correlation with the lower areas. If the color is close to white then the activations of both lower and higher layers of DCNN were correlating with the brain responses in approximately equal proportion.

and time windows are are presented in figures 3 and 4. We can
 see that weaker alignment (that does not survive the Bonferroni
 correction) is present in early time window in theta and alpha
 frequency range. No alignment is observed in the beta band.

To investigate the involvement of each frequency band more 5 6 closely we analyzed each visual area separately. Figure 5 shows the volume of activity in each area (size of the marker on the 7 figure) and whether that activity was more correlated with the 8 complex visual features (red color) or simple features (blue color). g In our findings the role of the earliest area (17) was minimal, how-10 ever that might be explained by a very low number of electrodes 11 in that area in our dataset (less that 1%). One can see from figure 12 5 that activity in theta frequency in time windows 50 - 250 ms 13 and 150 - 350 ms had large volume and is correlated with the 14 higher layers of DCNN in higher visual areas (19, 37, 20) of the 15 ventral stream. This hints at the role of theta activity in visual 16 object recognition. In general, in areas 37 and 20 all frequency 17 18 bands carried information about high level features in the early 19 time windows. This implies that already at early stages of processing the information about complex features was present in 20 those areas. 21

# Gamma activity is more specific to convolutional layers, while the activity in lower frequency bands is

#### <sup>24</sup> more specific to fully connected layers

25 We analysed volume and specificity of brain activity that corre-

<sup>26</sup> lates with each layer of DCNN separately to see if any bands or

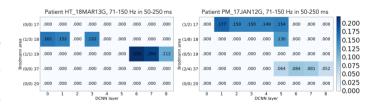


Figure 6 Single subject results from two different subjects. The numbers show the sum of correlations normalized by the number of probes in an area. On the left plot we see how a probe in Brodmann area 18 is mapped to the layers 0, 1, and 3 DCNN, while the activity in Brodmann area 19, which is located further along the ventral stream, is mapped to the higher layers of DCNN: 6, 7, 8. Similar trend is seen on the right plot. The numbers on the left of each subplot show the number of significantly correlating probes in each area out of the total number of responsive probes in that area.

time windows are specific to particular level of hierarchy of visual processing in DCNN. Figure 7 presents a visual summary of this analysis. In the "Methods" section we have defined total volume of visual activity in layers *L*. We used this measure to quantify the activity in low and high gamma bands. We noticed that while the fraction of gamma activity that is mapped to convolutional layers is high  $\left(\frac{\bar{V}_{\{conv1...conv5\}}^{\gamma,\Gamma}}{V_{all \ bands}} = 0.71\right)$ , this fraction diminished in fully connected layers fc6 and fc7  $\left(\frac{\bar{V}_{\{cof, fc7\}}^{\gamma,\Gamma}}{V_{all \ bands}} = 0.39\right)$ . Note that fc8 was excluded as it represents class label probabilities and

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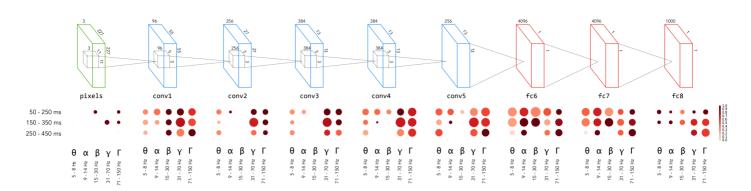


Figure 7 Specificity of neural responses across frequency bands and time windows for each layer of DCNN. Size of a marker is the total activity mapped to this layer and the intensity of the color is the specificity of the activity to visual areas.

does not carry information about visual features of the objects. 1 On the other hand the activity in lower frequency bands (theta, 2 3 alpha, beta) showed the opposite trend – fraction of volume in convolutional layers was 0.29, while in fully connected it growed 4 to 0.61. This observation highlighted the fact that visual features 5 extracted by convolutional filters of DCNN carry the signal that 6 is more similar to the signal carried by gamma frequency bands, 7 while the fully connected layers that do not directly correspond 8 to intuitive visual features, carry information that has more in g common with the activity in the lower frequency bands. 10

## 11 Discussion

The recent advances in artificial intelligence research have been 12 breathtaking. Not only do the deep neural networks match 13 14 human performance in visual object recognition, they also pro-15 vide the best model for how biological object recognition happens 16 (Kriegeskorte, 2015; Yamins and DiCarlo, 2016). Previous work 17 has established a correspondence between hierarchy of the DCNN and the fMRI responses measured across the human visual areas 18 19 (Güçlü and van Gerven, 2015; Eickenberg et al., 2016; Seibert et al., 2016; Cichy et al., 2016b). Further research has shown that 20 21 the activity of the DCNN matches the biological neural hierarchy in time as well (Cichy et al., 2016b; Seeliger et al., 2017). 22 Studying intracranial recordings allowed us to extend previous 23 findings by assessing the alignment between the DCNN and corti-24 cal signals at different frequency bands. As there is a quantifiable 25 increase of the complexity of features along the layers of the 26 27 DCNN, any signal that is aligned to the DCNN has to carry sim-28 ilarly increasingly complex features built-up during visual object recognition. We observed that the lower layers of the DCNN 29 explained gamma band signals from earlier visual areas, while 30 higher layers of the DCNN, responsive for more complex features, 31 matched with the gamma band signals from higher visual areas. 32 Correspondence between layers of DCNN and visual hierarchy of 33 human brain was present not only at the extremes, but also at the 34 intermediate layers of the hierarchy. Hence, one can conclude that 35 gamma band carries increasingly complex features required for 36 object recognition along the ventral visual pathway. This finding 37 confirms previous work that has given a central role for gamma 38 band activity in visual object recognition (Singer and Gray, 1995; 39 40 Singer, 1999; Fisch et al., 2009) and feedforward communication (Van Kerkoerle et al., 2014; Bastos et al., 2015; Michalareas et al., 2016). However, importantly, our results show that gamma activity reflects not only object recognition per se but also the feature transformations that are computed on the way towards explicit object representations. Our work demonstrates that the correlation between the DCNN and the biological counterpart is specific not only in space and time, but also in frequency.

## Feedforward and feedback computations in object recognition

Visual object recognition in the brain involves both feedforward 10 and feedback computations (DiCarlo et al., 2012; Kriegeskorte, 11 2015). What do our results reveal about the nature of feedfor-12 ward and feedback components in visual object recognition? We 13 observed that the DCNN corresponds to the biological processing 14 hierarchy even in the latest analysed time-window (Figure 3). In 15 a directly relevant previous work Cichy and colleagues compared 16 DCNN representations to millisecond resolved MEG data from 17 humans (Cichy et al., 2016b). There was a positive correlation 18 between the layer number of the DCNN and the peak latency of 19 the correlation time course between the respective DCNN layer 20 and MEG signals. In other words, deeper layers of the DCNN 21 predicted later brain signals. As evidenced on Figure 3 in (Cichy 22 et al., 2016b), the correlation between DCNN and MEG activity 23 peaked between ca 100 and 160 ms for all layers, but significant 24 correlation persisted well beyond that time-window. However, in 25 the work of (Cichy et al., 2016b) the correlation decreased over 26 time, while in our data we evidenced no such clear drop in the 27 later time windows: even between 250-450 ms the alignment in 28 low gamma was strong and significant. 29

How could this late alignment be interpreted? In particular, 30 feedforward object recognition is thought to be finished in ca 31 200-250 milliseconds after stimulus onset (DiCarlo et al., 2012). 32 Hence, one could think that the correspondence between the 33 DCNN, which is a feedforward network and biological visual 34 object recognition should be confined to early time windows. 35 However, although the DCNN is a purely feedforward network 36 it is important to notice that the alignment between electro-37 physiological signals and the DCNN does not imply that the 38 respective signals have to reflect feedforward computations. Such 39

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alignment only means that the progressive changes in representa-1 tional geometry along the processing hierarchy are similar to the 2 DCNN. In other words, it is possible that the activity patterns 3 observed are a result of recurrent computations, but their out-4 come representational geometry resembles that of the DCNN. 5 Therefore, the present results together with previous findings 6 (Cichy et al., 2016b) demonstrate that the DCNN is a good model 7 8 not only for feedforward object recognition, but also for the later 9 phases, which most likely include feedback computations. This fits with the predictive coding framework where the feedback 10 activity is not an unspecific modulatory signal but rather has to 11 signal specific contents from higher to lower levels of the process-12 ing hierarchy (Bastos et al., 2012). Hence, within this theoretical 13 framework, a specific representational geometry is expected even 14 from a feedback channel. 15

## 16 Low vs high gamma in object recognition

We observed significant alignment to the DCNN in both low 17 and high gamma bands. However, for high gamma this align-18 ment was more restricted in time, surviving correction only in 19 the middle time window. Previous studies have shown that low 20 21 and high gamma frequencies are functionally different: while low gamma is more related to classic narrow-band gamma oscilla-22 tions, high frequencies seem to reflect local spiking activity rather 23 than oscillations (Manning et al., 2009; Ray and Maunsell, 2011), 24 the distinction between low and high gamma activity has also 25 implications from cognitive processing perspective (Vidal et al., 26 2006; Wyart and Tallon-Baudry, 2008). In the current work we 27 approached the data analysis from the machine learning point of 28 view and remained agnostic with respect to the oscillatory nature 29 of underlying signals. Importantly, we found that numerically the 30 alignment to the DCNN was stronger and persisted for longer in 31 low gamma frequencies. However, high gamma was more promi-32 33 nent when considering volume and specificity to visual areas. The 34 most striking difference between the low and high gamma with regard to specificity was in the earliest time window 50-250 ms 35 where the correlation between the DCNN and high gamma was 36 almost exclusive to visual areas. 37

#### 38 Limitations

The present work relies on data pooled over the recordings 39 from 100 subjects. Hence, the correspondence we found between 40 responses at different frequency bands and layers of DCNN is 41 distributed over many subjects. While it is expected that single 42 43 subjects show similar mappings (see also Figure 6), the variability in number and location of recording electrodes in individual sub-44 jects makes it difficult a full single-subject analysis with this type 45 of data. We also note that the mapping between electrode loca-46 tions and Brodmann areas is approximate and the exact mapping 47 would require individual anatomical reconstructions and more 48 refined atlases. 49

#### 50 Future work

51 Intracranial recordings are both precisely localized in space and 52 time, thus allowing us to explore phenomena not observable with fMRI. In this work we investigated the correlation of DCNN activity with five broad frequency bands and three time windows. Our next steps will include the analysis of the activity on a more granular temporal and spectral scale. Replacing representation similarity analysis with a predictive model (such as regularized linear regression) will allow us to explore which visual features elicited the highest responses in the visual cortex.

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## Acknowledgements

IK, RV and JA thank the financial support from the Estonian Research Council through the personal research grants PUT438 and PUT1476. This work was supported by the Estonian Centre of Excellence in IT (EXCITE), funded by the European Regional Development Fund.

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