

# **GFF3sort: an efficient tool to sort GFF3 files for tabix indexing**

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# 1 Abstract

2 **Motivation:** The traditional method of visualizing gene annotation data in JBrowse is  
 3 converting GFF3 files to JSON format, which is time-consuming. The latest version  
 4 of JBrowse supports rendering sorted GFF3 files indexed by tabix, a novel strategy  
 5 that is more convenient than the original conversion process. However, current tools  
 6 available for GFF3 file sorting have bugs that would lead to erroneous rendering in  
 7 JBrowse.

8 **Results:** We developed GFF3sort, a script to sort GFF3 files for tabix indexing.  
 9 GFF3sort can properly deal with the order of features that have the same chromosome  
 10 and start position. Based on our test datasets from multiple species, GFF3sort  
 11 produced accurate sorting results while taking significantly less running time  
 12 compared with currently available tools. We anticipate that GFF3sort will be a useful  
 13 tool to help with genome annotation data processing and visualization.

14 **Availability:** <https://github.com/billzt/gff3sort>

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16 **Keywords:** GFF3, JBrowse, Visualization, Tabix

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# 1   **Implementation**

## 2   **Introduction**

3   As a powerful genome browser based on HTML5 and JavaScript, JBrowse has been  
4   widely used since released in 2009[1, 2]. According to its configuration document[3],  
5   it works by first converting genome annotation data in GFF3 file formats to JSON  
6   files by a built-in script “flatfile-to-json.pl”, and then rendering visualized element  
7   models such as genes, transcripts, repeat elements, etc. The main problem, however, is  
8   that this step is extremely time-consuming. The time is proportional to the number of  
9   feature elements in GFF3 files (Figure 1A and Additional file 1). Even for small  
10   genomes like yeast (*Saccharomyces cerevisiae*), it takes ~10 seconds to finish the  
11   conversion. For large and deeply annotated genomes such as that of humans, the time  
12   increases to more than 15 minutes. In addition, through the conversion process, a  
13   single GFF3 file is converted to thousands of piecemeal JSON files, thus putting a  
14   heavy burden on the ability to back up and store data.

15   In the recently released JBrowse version (v1.12.3), support for indexed GFF3  
16   files has been added[4]. In this strategy, the GFF3 file is compressed with bgzip and  
17   indexed with tabix[5], which generates only two data files: a compressed file (.gz) and  
18   an index file (.tbi). Compared with the traditional processing protocol, the whole  
19   compression and index process could be finished within a few seconds even for large  
20   datasets such as the human genome annotation data (Figure 1A and Additional file 1).  
21   The tabix tool requires GFF3 files to be sorted by chromosomes and positions, which  
22   could be performed in the GNU sort program or the GenomeTools[6] package (see  
23   [7]). However, when dealing with feature lines in the same chromosome and position,  
24   both of the tools would sort them in an ambiguous way that usually results in parent  
25   features being placed behind their children (Figure 1B), causing erroneous rendering  
26   in JBrowse[8] (Figure 1B). An alternative sorting tool is needed to resolve this  
27   problem.

1        Here, we present GFF3sort, an efficient tool to sort GFF3 files for tabix indexing.  
2        Compared with GNU sort and GenomeTools, GFF3sort produces sorting results that  
3        could be correctly rendered by JBrowse while saving a significant amount of time. We  
4        anticipate that GFF3sort will be a useful tool to help with processing and visualizing  
5        genome annotation data.

## 6        **Methods**

7        GFF3sort is a script written in Perl. It uses a single hash table to store the input GFF3  
8        annotation data (Figure 1C). For each feature, the chromosome ID and the start  
9        position are stored in the primary and secondary key, respectively. Features with the  
10       same chromosome and start position are grouped in an array in the same order of their  
11       appearance in the original GFF3 data. After sorting the hash table by chromosome IDs  
12       and start positions, GFF3sort implemented two modes to sort features within the array:  
13       the default mode and the precise mode (Figure 1C). In most situations, the original  
14       GFF3 annotations produced by genome annotation projects have already placed  
15       parent features before their children. Therefore, GFF3sort returns the feature lines in  
16       their original order, which is the default behavior. In some situations where orders in  
17       the input file has already been disturbed (for example, by GNU sort or GenomeTools),  
18       GFF3sort would sort them according to the parent-child topology using the sorting  
19       algorithm of directed acyclic graph[9], which is the most precise behavior but costs  
20       more computational source.

21       In order to test the performance of GFF3sort, the GFF3 annotation files of seven  
22       species (see the Data Sources in Table 1) were downloaded from the ENSEMBL  
23       database [10]. All the tests were conducted on a SuperMicro® server equipped with  
24       80 Intel® Xeon® CPUs (2.40GHz), 128 GB RAM, and running the CentOS 6.9  
25       system.

# 1    **Functionality and Performance**

2    GFF3sort takes a GFF3 file as its input data and returns a sorted GFF3 file as output.  
3    An optional parameter is used to turn on the precise mode. It outperforms the GNU  
4    sort program and GenomeTools in two aspects: correctness and running speed. In our  
5    seven test datasets, GFF3sort produces the sorted GFF3 files with a correct rate of 100%  
6    (measured by the percentage of parent features correctly placed before their children),  
7    compared with the <50% correct rate for GNU sort or GenomeTools (Table 1). It is  
8    able to fix the order of GFF3 files that has been incorreced sorted. Element models  
9    sorted by GFF3sort can be correctly rendered by JBrowse (Figure 1D). In addition,  
10    GFF3sort runs faster than both of those tools (Figure 1E and Additional file 1). In the  
11    default mode, GFF3sort saves ~70% running time. The precise mode takes longer  
12    time but still runs faster than traditional tools, especially for large annotation data.

13        In conclusion, GFF3sort is an efficient tool to sort GFF3 files for tabix indexing  
14    and therefore can be used to visualize annotation data. It has a high correct rate and a  
15    fast running speed compared with similar, existing tools. We anticipate that GFF3sort  
16    will be a useful tool to simplify data processing and visualization.

# 17    **Figure Legends**

18    Figure 1. **The motivation for, outlines of, and performance of GFF3sort.** A)  
19    Comparison of the running time of GFF3-to-JSON conversion and the bgzip-tabix  
20    process based on seven GFF3 annotation datasets: *Saccharomyces cerevisiae*  
21    (R64-1-1), *Aspergillus nidulans* (ASM1142v1), *Chlamydomonas reinhardtii* (INSDC  
22    v3.1), *Drosophila melanogaster* (BDGP6), *Arabidopsis thaliana* (Araport11), *Rattus*  
23    *norvegicus* (Rnor\_6.0), and *Homo sapiens* (GRCh38). Feature numbers are measured  
24    by counting lines in the GFF3 file. B) An example of incorrectly sorted GFF3 data  
25    and its snapshots in JBrowse. The two lines (mRNA) marked in red were placed after  
26    their sub-features (exon or UTR). Such incorrect placement leads to losing the first

1 exon in JBrowse rendering results. C) Outlines of GFF3sort. D) An example of  
2 correctly sorted data by GFF3sort and its snapshots in JBrowse. In this example, the  
3 two lines (mRNA) marked in red were correctly placed before their sub-features,  
4 allowing JBrowse to render them properly. E) Comparison of the running time of  
5 GFF3sort (including the default mode and the precise mode) and other sorting tools.

## 6 Tables

7 Table 1. The correct rate\* of GFF3 sorting results using different tools.

Data Source	Parent Feature Number	GNU sort	GenomeTools	GFF3sort
<i>Saccharomyces cerevisiae</i> (R64-1-1)	14,252	47.21%	50.08%	100%
<i>Aspergillus nidulans</i> (ASM1142v1)	21,653	48.94%	37.75%	100%
<i>Chlamydomonas reinhardtii</i> (INSDC v3.1)	29,047	49.68%	36.96%	100%
<i>Drosophila melanogaster</i> (BDGP6)	52,308	27.54%	23.26%	100%
<i>Arabidopsis thaliana</i> (Araport11)	86,846	36.20%	29.70%	100%
<i>Rattus norvegicus</i> (Rnor_6.0)	73,961	34.32%	41.17%	100%
<i>Homo sapiens</i> (GRCh38)	257,467	28.09%	17.41%	100%

8 \*The correct rate is measured by the percentage of correctly sorted features that have  
9 children ones. Such feature includes coding or non-coding genes and transcripts. If a  
10 feature line is placed before all its children features, then it is considered as correctly  
11 sorted.

## 1    **Additional files**

2    Additional file 1: Benchmark data. This file displays: 1) the detailed running time of  
 3    GFF3-to-JSON conversion and the bgzip-tabix process on our test datasets; 2) the  
 4    detailed running time of GFF3sort, GNU sort, and GenomeTools on our test datasets.  
 5    (DOCX)  
 6

## 7    **Availability and requirements**

8    Project name: GFF3sort  
 9    Project home page: <https://github.com/billzt/gff3sort>  
 10    Operating system(s): Linux  
 11    Programming language: Perl  
 12    Other requirements: No  
 13    License: No restrictions for academic users.  
 14    Any restrictions to use by non-academics: license needed

## 15    **Declarations**

## 16    **List of abbreviations**

17    JBrowse: JavaScript-based genome browser  
 18    GFF3: General Feature Format, version 3  
 19    JSON: JavaScript Object Notation  
 20    HTML5: HyperText Markup Language, version 5

## 21    **Ethics approval and consent to participate**

22    Not applicable.

## 1    **Consent for publication**

2    Not applicable.

## 3    **Competing interests**

4    The authors declare that they have no competing interests.

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## 8    **Authors' contributions**

9    SG, RZ, and TZ initiated the idea of the tool and conceived the project. TZ designed  
10   the tool and analyzed the data. CL and ZM helped to test the tool. TZ wrote the paper.  
11   All authors read and approved the final manuscript.

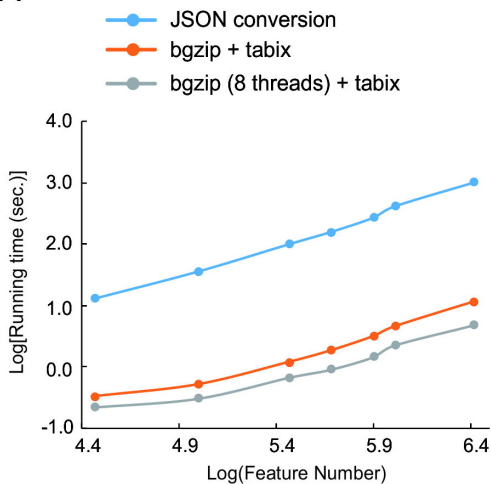
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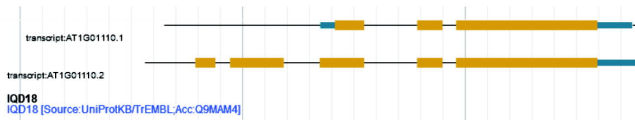
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- 7

A



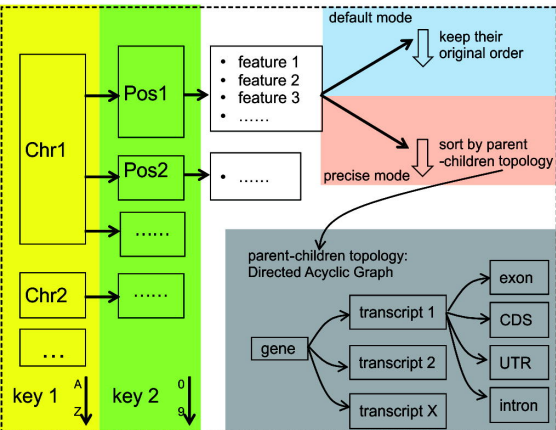
B

1	araport11	exon	51953	52346	+	Parent=AT1G01110.2
1	araport11	five_prime_UTR	51953	52238	+	Parent=AT1G01110.2
1	araport11	gene	51953	54737	+	ID=AT1G01110;
1	araport11	mRNA	51953	54737	+	ID=AT1G01110.2;Parent=AT1G01110
1	araport11	exon	52061	52730	+	Parent=AT1G01110.1
1	araport11	five_prime_UTR	52061	52730	+	Parent=AT1G01110.1
1	araport11	mRNA	52061	54689	+	ID=AT1G01110.1;Parent=AT1G01110



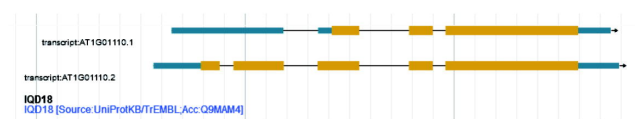
C

## Hash Table



D

1	araport11	gene	51953	54737	+	ID=AT1G01110;
1	araport11	mRNA	51953	54737	+	ID=AT1G01110.2;Parent=AT1G01110
1	araport11	five_prime_UTR	51953	52238	+	Parent=AT1G01110.2
1	araport11	exon	51953	52346	+	Parent=AT1G01110.2
1	araport11	mRNA	52061	54689	+	ID=AT1G01110.1;Parent=AT1G01110
1	araport11	exon	52061	52730	+	Parent=AT1G01110.1
1	araport11	five_prime_UTR	52061	52730	+	Parent=AT1G01110.1



E

