MODELLING OF THE CARDIOPULMONARY RESPONSES TO MAXIMAL AEROBIC EXERCISE IN PATIENTS WITH CYSTIC FIBROSIS

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ABSTRACT

Cystic fibrosis (CF) is a debilitating chronic condition, which requires complex and expensive

disease management. Exercise has now been recognised as a critical factor in improving health and

quality of life in patients with CF. Hence, cardiopulmonary exercise testing (CPET) is used to

determine aerobic fitness of young patients as part of the clinical management of CF. However, at

present there is a lack of conclusive evidence for one limiting system of aerobic fitness for CF

patients at individual patient level.

Here, we perform detailed data analysis that allows us to identify important systems-level factors that

affect aerobic fitness. We use patients' data and principal component analysis to confirm the

dependence of CPET performance on variables associated with ventilation and metabolic rates of

oxygen consumption. We find that the time at which participants cross the anaerobic threshold (AT)

is well correlated with their overall performance. Furthermore, we propose a predictive modelling

framework that captures the relationship between ventilatory dynamics, lung capacity and function

and performance in CPET within a group of children and adolescents with CF. Specifically, we show

that using Gaussian processes (GP) we can predict AT at the individual patient level with reasonable

accuracy given the small sample size of the available group of patients. We conclude by presenting

future perspectives for improving and extending the proposed framework.

Our modelling and analysis have the potential to pave the way to designing personalised exercise

programmes that are tailored to specific individual needs relative to patient's treatment therapies.

INTRODUCTION

Cystic fibrosis (CF) is the most common life shortening genetic disease in the Caucasian population,

affecting nearly 11,000 individuals in the United Kingdom (UK) (1). The pathology of the disease,

for which there is no cure, manifests itself throughout the respiratory, digestive and reproductive

systems of the human body. Currently, as there is no cure for CF, the management of the disease is a

key factor in the quality of care and health related quality of life factors. Part of the management of

this disease requires cardiopulmonary exercise testing (CPET) to determine aerobic fitness, as

represented by the maximal oxygen consumption ($\dot{V}O_{2max}$). This parameter provides a clinically

useful prognostic evaluation of a patient's functional capabilities. Even in mild to moderate severity

of CF, patients are known to demonstrate impairments in cardiac and respiratory functions leading to

exercise intolerance.

Enhanced aerobic fitness has been shown to improve quality of life in young patients with CF (6-18

years), with benefits including lower risk of hospitalisation, increased exercise tolerance, reduced

residual volume, increased endurance of the respiratory muscles, enhanced sputum expectoration and

decreased rate of decline in pulmonary function (2-6). It has also been shown from CPET that

individuals with CF possessing a higher $\dot{V}O_{2max}$ are shown to have a reduced mortality risk. Nixon *et*

al. (1992) reported that individuals with a $\dot{V}O_{2max}$ greater than 82% of their predicted value had an

83% 8-year survival rate, compared to just 28% 8-year survival rate for patients with a $\dot{V}O_{2max}$ less

than 58% of their predicted value (7). Furthermore, patients with a higher $\dot{V}O_{2max}$ also have

additional benefits in terms of improved fluid balance, retention of serum electrolytes through

increased plasma volume, and potential for impact on sweat gland function thus reducing thermal

strain and dehydration (8). These systemic changes are advantageous responses to exercise training,

and result in an enhanced quality of life, increased physical function and increased life expectancy (9,

10).

Effective management of the disease has become even more critical in recent years due to an aging

CF population group, with the median predicted survival of children born with CF in the UK now

being 45 years (1). As a consequence of an aging patient group and high medical care costs (11),

maintaining or enhancing fitness is crucial. Exercise has being widely acknowledged as a key

management strategy for CF, supported by some mechanistic data on the systemic effects of exercise

at the cellular level in vivo in young patients with CF (5-9). However, an integrated systems level

understanding of the limitations of aerobic fitness for CF patients is lacking. Measurement techniques

that do exist to quantify within-organ, real-time perfusion and intracellular oxygenation are invasive

and unethical for use with paediatric patients, and current animal model research provides limited

direct relevance to paediatric pathology. In clinical practice, there is significant interaction between

cardiac and pulmonary function and the behaviour of the systemic vasculature during exercise

training. This can result in the functional improvement in one part of the combined system, but

detrimental effects on others (9). Clinicians therefore inevitably have to adopt very imprecise

guidelines related to exercise prescription (12).

The use of modelling and simulation tools in clinical medicine is currently the subject of intense

research interest both in the UK and internationally (13-17), and the adoption of a systems

biomedicine approach to build and validate novel multi-scale, organ-level, integrated, re-usable and

re-deployable models represents a paradigm shift in biomedical modelling and simulation. There are

numerous organ level models in existence (18-21), however, to date there have been limited attempts

to either integrate these or to apply them to real clinical applications. There is ongoing basic science

and clinical trial work providing data on the micro (22) and macrovascular (23) changes associated

with exercise. These data, although important, have yet to be integrated quantitatively with other data

streams. In particular, there has been almost no previous work on the use of predictive modelling and

simulation technologies for developing treatment strategies for CF patients.

Here, we present detailed data analysis of responses to progressive exercise in patients with CF, with

a view of determining predictors of performance. We find that the time at which participants cross

the anaerobic threshold (AT), as measured by means of gas exchange threshold (GET) is well

correlated with overall performance. To gain further insight, we then develop a surrogate (statistical)

model that allows us to evaluate how CF impairs exercise tolerance relative to increasing ventilatory

and metabolic demands. Our modelling and analysis was based on data collected during cycling

exercise form a CPET at different work rates (from resting to voluntary exhaustion) in young patients

with CF. The outputs produced are discussed in this paper, with analyses focussing on pulmonary

parameters.

METHODS

Before turning our attention to modelling, we perform an exploratory analysis of the available data in

order to identify predictors of performance. This study was carried out in accordance with the

recommendations of the European Respiratory Society, written consent and assent was obtained from

parent(s)/guardian(s) and participants, respectively. All participants' parents gave written informed

consent in accordance with the Declaration of Helsinki. The protocol was approved by the South

West NHS Research Ethics Committee. Data were collected from 15 children and adolescents with

CF, who performed a valid (24) and reliable (25) combined ramp incremental and supramaximal

(Smax) CPET to determine $\dot{V}O_{2max}$ and the GET. This protocol was performed on an electronically

braked cycle ergometer, and required patients to perform an initial exhaustive ramp incremental test

at a pre-determined rate between 10–25 W·min^{-1}, in order to elicit exhaustion in approximately ten

minutes (26). After a 3-min warm-up at 10-20 W, participants completed this incremental test to the

point of volitional exhaustion, maintaining a cadence of 70–80 rpm throughout. Exhaustion was

defined as a 10 rpm drop in cadence for five consecutive seconds, despite strong verbal

encouragement. Active (5-min cycling at 20 W) and then passive seated recovery (10 min) then

preceded the Smax bout. Smax verification consisted of a 3-min warm-up (10-20 W), followed by a

''step'' transition to a constant work rate corresponding to 110% peak power output (27). Upon

volitional exhaustion (defined previously), a 5-min active recovery (slow cycling at 20 W) concluded

the combined CPET session.

MODEL SIMULATIONS

Simulations are widely used in various fields of science and engineering because conducting physical

experiments is too costly, or highly time-consuming, or even impossible in some cases (28). In the

case of CPET in CF patients, there are also ethical considerations, since the test adds to the treatment

burden many children and adolescents with CF already face.

Often, a primary goal of using model simulations is to perform quantitative studies such as

uncertainty quantification or sensitivity analysis. Such studies are crucially important in biomedicine,

since there exists significant variation both between and within patient groups. Through

understanding and quantification of the uncertainty within the mathematical models, outcomes of

patient-specific interventions can be better predicted. However, such investigations require a large

number of runs that makes it impractical if each run takes more than a few seconds. To cope with this

difficulty, one can use *emulators*, also known as *surrogates*, or *metamodels* or *response surfaces*

(29). These provide a fast approximation of the input/output relation governed by the underlying

simulator. The most important classes of surrogate models have been described elsewhere (30-32).

The surrogate model employed in this study is based on Gaussian processes (GP), which have

become increasingly popular over the last decade (29). GPs have been used in a wide range of

applications from wireless communication, to obtain position estimates for a mobile user (33);

metallurgy, to model the development of microstructure (34); and in biology, to describe gene

regulatory processes and cell growth (35-37).

The data analysis was performed using Python (Anaconda Software Distribution. Version 2-2.4.0.

Continuum Analytics, 2016. URL https://continuum.io) and MATLAB and Statistics Toolbox

Release 2016b, The MathWorks, Inc., Natick, Massachusetts, United States. The GP model simulator

was implemented in R (R Core Team (2013). R: A language and environment for statistical

computing. R Foundation for Statistical Computing, Vienna, Austria.   URL http://www.R-

project.org/.)

RESULTS

Data Analysis

To facilitate understanding, we first plot in **Figure 1(a)**, raw data displaying the performance of the

participants. The work rate for each participant is increased at a rate that is either, a) dependent on

their performance in previous tests, or b) when a prior test is unavailable, at a rate that is predicted to

elicit exhaustion in approximately ten minutes (26). This is done in order to keep the expected

duration of the test comparable to other participants. Note that this means that the total energy

expended by a given participant is not based on the duration of the test alone. In **Figure 1(b)**, we

show how participant age affects overall test performance. We observe a correlation between the two:

the worst performing participants tend to be the youngest, but this effect is insignificant at older ages.

The colour coordination used in this figure (red-worst performance → blue-average performance

→green-best performance) will be used throughout the remainder of this section, where performance

is quantified by the total energy transferred during the test.

Figure 1: (a) The work rate for each participant is increased at a rate dependent on their past test performance. (b) Participant age is correlated with test performance for young participants, but not for older ones.

In **Figure 2**, we plot the ratio of $\dot{V}O_2$ over total ventilation ($\dot{V}E$) with respect to time. The markers on each of the time traces indicate the time of volitional exhaustion for that participant. There are two features that stand out from this figure. Firstly, participants who perform better have higher $\dot{V}O_2/\dot{V}E$ ratios, suggesting that their oxygen uptake is more efficient than their poorer performing counterparts. Secondly, in the recovery phase of the test (5 minutes following volitional exhaustion) better performing participants exhibit a sharp decrease in $\dot{V}O_2/\dot{V}E$, which is not observed in the poor performance group. Again, this suggests a more efficient utilisation amongst the former group and that exhalation of CO_2 is perhaps more significant to total breathing following the test.

Figure 2: Ratio of oxygen utilisation and total breathing throughout the test. Markers indicate the volitional exhaustion times for each participant.

We next examine the effect that breathing patterns have on participant performance. Two classical

prognostic measures used for patients with cystic fibrosis are the forced vital capacity (FVC), and the

forced expiratory volume in one second (FEV_1). These measures have been shown to be well

correlated with mortality and overall fitness of CF patient groups (38-40). In **Figure 3**, we

demonstrate how these metrics are correlated with performance in the CPET test. In **Figure 3(a)**, we

observe good correlation between FVC and the maximum tidal volume (TV) of breathing achieved

throughout the test. This is unsurprising since participants are likely to be trying to maximise their

breathing depth close to their exhaustion point. However, notice that, although the group with low

FVC performed poorly, this measure was unable to separate other participants. **Figure 3(b)** reiterates

this result and also highlights the high correlation between FEV_1 and FVC.

Figure 3: (a) Correlation of FEV_1 with the maximal tidal volume achieved throughout the test.

(b) Correlation between FEV_1 and FVC is high. Note that, although FVC and FEV_1 are good

predictors of poor test performance, they are unable to distinguish better performing

participants.

In order to better classify the performance of the participants, we must instead look for other factors.

In **Figure 4**, we present the total breathing rate against the oxygen consumption throughout the test.

In **Figure 4(a)**, we find a strong relationship between test performance and respiratory pattern. Note

that the curvature of the graphs suggests that an exponential fit, rather than a linear one, is most

appropriate for these data. In order to test this, we take logarithms of the data and perform a linear

regression, ignoring the first 180s of the test since participants are here in the warm up phase (work

rate is not increasing) and the final 60s of the data prior to volitional exhaustion, since participants

pass their respiratory compensation point, inducing hyperventilation and erratic breathing. The

results of the fit are shown in **Figure 4(b)** and we can see more clearly the association of

performance on breathing pattern.

Figure 4: (a) Total ventilation plotted against oxygen utilisation. We observe that breathing

pattern is strongly correlated with test performance. (b) Exponential curves are fitted through

the raw data, further highlighting this dependence.

From the fitted curves, we can further explore the dependence of breathing patterns on performance.

Firstly, in **Figure 5(a)**, we plot the slope of the fitted curve against the total energy transfer. We find

that the slope of the curve of $\log\dot{V}E$ against $\dot{V}O_2$ alone does not capture all of the variation in energy,

which is highlighted by the low R-squared value (0.68). Instead, we plot in **Figure 5(b)** the oxygen

consumption at a fixed rate of breathing against the total energy. Here, we find a good

characterisation of the overall performance, with a much higher R-squared value (0.86), confirming

that those who utilise oxygen more efficiently perform better.

Figure 5: Slope of the fitted curves ($\log\dot{V}E$ against $\dot{V}O_2$) from Fig. 4(b) plotted against the total

energy transfer during the test. We find a relatively poor characterisation of the variance

between performances. (b) By instead plotting the oxygen consumption at a fixed rate of
breathing, we better capture differences in performance.

Next, we examine the specific patterns of breathing exhibited by the participants, in particular,
focussing on breathing depth and frequency. Initial characterisations of these patterns seem to
provide little information, as indicated in **Figure 6(a)**. However, when we now also include
dependence of oxygen consumption, we find a near perfect classification of participants into the
lowest performing groups, the best performing groups and the middle group. These data are
displayed in **Figure 6(b)**. Note that in this figure, the trajectories appear to be evolving on a planar
manifold, suggesting significant co-dependence between these three variables.

Figure 6: (a) Breathing patterns subdivided into the breathing rate and tidal volume. These
data appear uninformative for predicting test performance. (b) With the additional inclusion of
the oxygen consumption at a fixed rate of breathing, we find that these variables now almost
perfectly capture variation in participant performance.

Given that there appears to be co-dependence between the variables used in **Figure 6(b)**, a sensible
next step is to use principal component analysis (PCA) to account for these dependencies. By
projecting the data onto their principal components, we show in **Figure 7** how well these capture the
variation in participant performance. Given that there are only three independent variables in our
analysis, it is natural to use spherical polar coordinates to show how these quantify performance. The
first of these components, θ, captures over 90% of the variation in performance (**Figure 7a**), as does

the normal component in the direction of breathing frequency (**Figure 7b**). These results further
indicate the importance of breathing frequency, together with co-variation of oxygen consumption
and tidal volume as predictors of test performance.

Figure 7: First of the principal components obtained via PCA accounts for over 90% of the
variation in test performance. (b) Similar levels of variance are accounted for by taking only
the normal component of the first principal component in the breathing frequency direction.

Gas Exchange Threshold

Under steady state levels of exercise, the metabolic rate of production of CO_2 is assumed to be
proportional to the utilisation rate of O_2 via the cellular respiratory quotient, since (after the initial
rest-work transition) adenosine triphosphate (ATP) is replenished primarily via aerobic metabolism
pathways. As the work rate increases, this pathway becomes unable to supply sufficient ATP to
satisfy the required amount of energy and anaerobic pathways have to contribute to overcome the
shortfall. In doing so, they increase the levels of waste products, such as lactate and also increase the
overall production rate in CO_2. The point at which this occurs is known as the anaerobic threshold
(AT) or sometimes referred to as the lactate threshold. However, this is difficult to directly measure
in vivo during exercise, and as such the GET is utilised as a non-invasive surrogate of the AT (41).

The time at which participants cross the anaerobic threshold is well correlated with overall
performance, as shown in **Figure 8(a)**, due to the fact that the anaerobic pathways are less efficient at
producing ATP and because build-up of lactate contributes significantly to fatigue. One of the major
contributing factors in defining AT is $\dot{V}O_{2max}$, since this is indicative of the limit of the rate of
oxidative phosphorylation. It thus comes as little surprise that $\dot{V}O_{2max}$ is the best single predictor of
CPET performance, as shown in **Figure 8(b)**.

Figure 8: (a) Quantifying the relationship between the anaerobic threshold and overall test
performance. Anaerobic thresholds were calculated using an automated procedure based on
previous methods (41) (b) $\dot{V}O_{2max}$ is the best single predictor of overall test performance.

Gaussian Processes-based Modelling

A Gaussian Process (GP) defines a probability distribution over functions where the true function is
considered as a particular sample path. In our modelling, we attempt to describe the influence of
breathing patterns and $\dot{V}O_2$ on the AT, since this is shown to correlate well with overall test
performance (see **Figure 8**). In mathematical terms, we treat AT as our scalar output variable, with
input variables comprising: baseline breathing rate and tidal volumes, and O_2 consumption rates at a
fixed ventilation rate, FVC, FEV_1, and the rate of changes in breathing rate and tidal at exercise
onset, using the slopes calculating in **Figure 4(b)**. Thus, we have an output variable dependent on
seven input variables, which we conveniently store in a vector $\mathbf{x} \in D \subset \mathbb{R}^d$, with $d=7$. We now
assume that there exists a 'true' function $f: D \to \mathbb{R}$, such that AT $= f(\mathbf{x})$.

A GP is fully specified by its mean, μ, and covariance $K,$ which are both functions of the input
variables: $\mu = \mu(\mathbf{x})$, $K = K(\mathbf{x}, \mathbf{x'})$. Specifically, if Y is a GP, then we write:

$$Y \sim GP(\mu, K): \quad \mu = \mathrm{E}[Y(\mathbf{x})], \quad K(\mathbf{x}, \mathbf{x'}) = \mathrm{Cov}(Y(\mathbf{x}), Y(\mathbf{x'})), \quad \mathbf{x}, \mathbf{x'} \in D$$

The above can be regarded as prior distribution over function spaces. This can be seen more clearly
in **Figure 9(a)**. In this subfigure, which shows a generic example of a GP, the bold red line is the
'true' function $f.$ Note that the true function is unknown – our aim is to construct a model that

approximates it. The thin grey lines are sample paths of a GP with a zero mean prior and a constant

prior covariance function. For exposition purposes, the example plot is restricted to the case $d=1$, but

the approach is unchanged for $d > 1$. In principle, μ could be any function, for practical purposes,

polynomial regressions are common choices. The choice of covariance function reflects our prior

belief about the structure of f (such as the level of smoothness) and therefore has a crucial influence

on our modelling. A typical choice for the covariance function is the squared-exponential kernel,

given by:

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$$K(\mathbf{x}, \mathbf{x'}) \;=\; \sigma^2 \prod_{i=1}^{d} exp\left(-\frac{|x_i - x_i'|^2}{2\Phi_i^2}\right)$$

Here, σ controls variability of Y along the y-axis while $\Phi_i > 0, i = 1, \dots, d$, scale the distance

measure for each input dimension. A more thorough exposition of common choices for the

covariance function may be found in (42).

Thus far, we have defined the prior distribution for the GP. It is clear from **Figure 9(a)** that, in

general, prior distributions are unlikely to provide a good approximation to the true function f. We

can incorporate data (or evaluation of f at specific points), with our prior distribution to give a

posterior distribution, following a Bayesian framework. The resulting posterior distribution of the

GP, conditioned on the data, will be much closer to the true function.

Let $\mathbf{y} = \{f(\mathbf{x}^1), \dots, f(\mathbf{x}^n)\}$ be a set of function evaluations at n locations $\mathbf{X} = \{\mathbf{x}^1, \dots, \mathbf{x}^n\}$. Here,

function evaluations correspond to the AT location for a set of patients during the CPET. Predicting

with GP is obtained by conditioning Y on sample points $\Omega = \{\mathbf{X}, \mathbf{y}\}$. For any (new) $\mathbf{z} \in D$, the

posterior distribution of $Y(\mathbf{z})|\Omega$ has a normal distribution with the following mean, m, and variance,

s^2:

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$$m(\mathbf{z}) = \mu(\mathbf{z}) + K(\mathbf{z}, \mathbf{X})^\top K(\mathbf{X}, \mathbf{X})^{-1}(\mathbf{y} - \mathbf{\mu}), \qquad (1)$$

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$$s^2(\mathbf{z}) = K(\mathbf{z},\mathbf{z}) - K(\mathbf{z}, \mathbf{X})^\top K(\mathbf{X}, \mathbf{X})^{-1} K(\mathbf{z}, \mathbf{X}), \qquad (2)$$

where $^\top$ denotes the transpose operator, $^{-1}$ is the inverse operator and $\mathbf{\mu} = \mu(\mathbf{X})$ is the vector of the

mean function at $\mathbf{X}$. In addition, $K(\mathbf{z},\mathbf{X})$ and $K(\mathbf{X},\mathbf{X})$ are the covariance vector between $Y(\mathbf{X})$ and $Y(\mathbf{z})$

and the covariance matrix between the observations. **Figure 9(b)** shows an example of incorporating

sample points to update the prior distribution shown in **Figure 9(a)**. In this generic example, the

function f is evaluated at five distinct values of **x,** and the mean and variance of the GP are updated

using (1)-(2).

Figure 9: GP is a probability distribution on function spaces. The thick solid red line is the true

function f and the thick dashed black line is the GP mean. (a) Thin grey lines show sample

paths of a prior GP whose mean is zero. (b) Blue bullets indicate five data points sampled from

f. The GP mean and covariance are updated using these sample points. The thin grey dashed

lines show $m(x) \pm 2s(x)$.

At the evaluated points, indicated in blue (colour online), the true value of f is known and so the

variance of the GP at these points vanishes and $\mu(\mathbf{X}) = f(\mathbf{X})$. In between these points, the variance

increases, dependent on the distance from an evaluated point. The mean of the GP, shown by the

thick black dashed line now approximates the true function much more closely (recall that the prior

mean function in **Figure 9(a)** was zero everywhere), and matches exactly at the evaluated points. The

approximation can be further improved by incorporating more data (function evaluations),

particularly around those input values for which the variance is high. Thus, as more data becomes

available, the model is iteratively improved.

Justification of Variables

In our simulator, we have used the AT as our output (dependent) variable. Another choice for this

could be the performance in the CPET or $\dot{V}O_{2max}$, since these are the primary biomarkers for gauging

aerobic fitness. However, the use of the GET has been shown to have high agreement with the lactate

threshold (another surrogate for the AT), and related to disease severity in CF (43). Furthermore, as

reported in **Figure 8(a),** the AT location for a given patient correlates well with their overall

performance in this test. Furthermore, by constructing a predictive model to approximate the AT

values for a patient, we can hope to further extend this to identify contributions of aerobic and

anaerobic pathways in supplying ATP to meet the demand imposed during the exercise test.

The initial exploration of results highlighted that both ventilation parameters and metabolic rates of

O_2 consumption were the primary factors influencing test performance. It is clear that $\dot{V}O_2$ should

play a significant role in determining the AT location, since it is a proxy for oxidative

phosphorylation which is the main pathway for ATP synthesis in steady state exercise. As a measure

of oxygen uptake efficiency in our model, we use the oxygen consumption rate at a fixed total

ventilation rate (that being 0.822 L/s) as an input (independent) variable for each patient.

There are a number of ventilatory input variables incorporated in our simulator. Given their potential

importance as clinical biomarkers, highlighting the limitations of lung capacity and function, we

include FVC and FEV_1 as input (independent) variables. During the aerobic exercise test, participants

spend three minutes cycling at a minimal work rate, over which we quantify their baseline breathing

frequency and baseline tidal volume by taking the means of these variables over this period. To

capture the dynamics response associated with the exercise, the rates of change of breathing

frequency and tidal volume are calculated, based on the fits obtained in **Figure 4(b)**. The rates of

change of these ventilatory variables indicate how participants respond to changes in workload and

were shown in **Figure 6(b)** to discriminate between participant performances. Moreover, differences

in rates of change of breathing frequency and tidal volume have previously been shown to be

significantly different between control groups and CF groups (44), suggesting that these are

potentially key biomarkers for assessing aerobic fitness in patients with CF.

GP Emulator Performance

The GP emulator was constructed using the data presented above, with AT calculated using

previously described methods (41). These data were used to train the emulator. For the prior

distributions, a first order polynomial regression was used for the mean, whilst a squared-exponential

kernel with σ and Φ provided via maximum likelihood estimation was used as the prior covariance.
Since in this pilot study, we have a small number of participants, we use leave-one-out cross-
validation to assess the accuracy of our emulator, that is, for each patient, we use the emulator trained
against the remainder of the sample points to approximate the AT value for that patient, given their
input variables. The results of this are presented in **Figure 10(a)**, in which we also plot the 95%
confidence intervals, with relative percentage errors demonstrated in **Figure 10(b)**.

Figure 10: (a) Predictions (asterisks) vs. exact values (bullets). Red bars show the 95%
confidence intervals, $m_i(x_i) \pm 1.96\ s_i(x_i)$ around the predicted value, where m_i and s_i are GP
prediction mean and standard deviation based on all but the *i*'th data point. (b) Relative
prediction error expressed as a percentage of the true value $f(x_i)$.

The emulator has reasonable accuracy, in spite of the small sample size. In general, for high accuracy
in GP emulator, the number of sample points (corresponding to the number of patients in our case)
should be at least ten times larger than the number of input variables (45). Clearly, there is a need to
acquire further data points to improve the predictive capabilities of the emulator. For each patient, the
95% confidence interval around the predicted point contains the true value. Moreover, the relative
errors for many patients is small (considering the low sample size), though we note that some
patients, (3 and 7), the error is high. This highlights a need to extend this study to include more data
to refine estimates around these points, particularly to deal with the high variability of lung function
parameters in this patient group (46).

DISCUSSION
Our primary modelling aim is to eventually use the model to evaluate how CF impairs exercise
tolerance relative to increasing ventilatory and metabolic demands. Our predictive model could also
be used to evaluate therapies and their effect on exercise performance. Ultimately, we hope that this
will form a series of steps to design better exercise treatment that is tailored to specific individual
needs relative to patient's treatment therapies, a treatment modality that is affordable, and
personalised (47).

The data analysis and modelling results have highlighted the dependence of CPET performance on
variables associated with ventilation and metabolic rates of O_2 consumption. Whilst these
observations are, in themselves, not novel, we believe that this is the first attempt to mathematically
model the relationship between ventilatory dynamics, $\dot{V}O_2$ and performance in CPET within a group
of children and adolescents with CF. Whilst it is clear that there is much work to be done in this area,
we hope that this will serve as a starting point for improved modelling, not only in the arena of GP
emulators, but also in the domain of mechanistic modelling, which we shall describe briefly.

Perspectives for GP improvements

At present, the GP model is conditioned on specific data points for each patient. An improvement to

the GP could be made by instead conditioning with respect to distributions. Given that repeated tests

are often performed for the same individual, so that multiple sample points are provided for each

participant, we can consider a fit to a probability distribution capturing the variability in the identified

variables. This approach has advantages compared to standard GP models, such as avoiding

problems associated with overfitting and regularisation (which is important for the inverting ill-

conditioned covariance matrices that often arise during the application of (1)-(2), (48).

Perspectives for mechanistic modelling

In order to better understand and characterise the difference between performances, it would be

extremely useful to construct and simulate a mechanistic mathematical model, based upon on

ordinary differential equation (ODE) framework, describing the relationship between the

cardiopulmonary system and the metabolic dynamics of skeletal muscle. By describing the

relationships between different organ-level systems, the model would be able to identify the patient-

specific rate-limiting factors defining aerobic fitness. Moreover, analysis of the model could be used

to suggest treatment strategies to improve these factors and thus predict how patients will improve

under such regimes.

At the individual organ level, there are a plethora of models describing individual dynamics of the

level of the heart (49-52), lung (53-56) and systemic metabolic demand (57-63). There also exist a

number of models describing such interactions between cardiopulmonary and metabolic systems (64-

68) in a variety of settings, including heart failure and mechanical ventilation. A core feature in all of

these models is the nonlinear interaction between the constituent model compartments that

encompass the distinct tissues. An important consequence of this is that the model must be studied as

whole, in an integrated fashion, to truly understand the body's response to exercise.

With respect to the present question, there are a number of limitations of the existing modelling

approaches. Most significantly, none have been designed with either an adolescent, or a CF patient

group in mind, and the nuances of these patient groups will have to be factored into to any model

development. In particular, these models have relatively simple, empirical models to describe

changes in ventilation, which may not capture breathing dynamics of our patient group well.

Moreover, to the best of our knowledge, no model considers the changes in ventilation separated into

breathing frequency and depth that have been shown by us and others (44) to be critical to overall test

performance.

In our analysis, we have demonstrated that the AT or GET location is a critical factor in determining

overall patient aerobic fitness. Many of the mathematical exercise models describe only steady-state

exercise, in which aerobic pathways meet most of the ATP demand (64-68). As such, these models

are inadequate to capture the dynamics we describe here. Another common topic of study is the

dynamic response at exercise onset, which again, does not meet the current need to describe the AT

crossing point (69-72).

Of the mathematical models that describe the contribution of anaerobic pathways to ATP production,

some assume that the shortfall in meeting ATP demand via oxidative phosphorylation is met entirely

by anaerobic pathways (73), yet this is clearly not so, since ATP levels in skeletal muscle post-

exercise may be up to 30% lower than pre-exercise values (74). Mathematical models that factor in

fatigue brought about by anaerobic metabolism are generally phenomenological in nature, and it is

difficult to quantify these models against real patient data (73, 75, 76), and these models mostly fall

outside the arena of ODE-based modelling and so dynamical properties are difficult to infer from

them.

Developing a mathematical model to describe the integrated behaviour of all of the relevant organs,

whilst remaining biophysically plausible, but without requiring excessive or invasive

parameterisation is a difficult task. The proposed model should include descriptions of the

cardiovascular system, the ventilatory system and simple models of metabolism at the tissue level.

Specifically, dynamic variables should include alveolar, arterial, venous and tissue level partial

pressures/concentrations of O_2 and CO_2, cardiac output, ventilation and metabolic rates oxygen

utilisation and CO_2 production. Partial alveolar gas pressures can be linked to data collected during

the test, and the work rate can then be provided as inputs to the model. Note that these variables are

similar to those included in previously defined models (64-68) and the aim is to extend these to

describe the dynamics observed in patients with CF. The proposed model schematic is displayed in

Figure 11.

Of critical importance to the overall model construction is the development of a simple, yet realistic

model of cellular metabolism, to overcome the issues discussed earlier. The model should respect the

different metabolic processes that occur in the muscle tissue, in particular: glycolysis,

phosphocreatine breakdown and synthesis and oxidative phosphorylation, in a simplistic fashion that

is amenable to being fit to CPET data. Whilst there are models that describe the biochemical

reactions associated with these processes, and importantly, their stoichiometry (58, 61, 63, 67, 77),

quantifying their associated rate constants *in vivo* is a near-impossible task, and so efforts must be

made to develop a model that incorporates the relevant metabolic dynamics whilst being simple

enough to be fit to data.

With knowledge of the integrated system, attempts can also be made to describe other important

exercise-based processes, such as lactate buffering and recycling (as a fuel source) (61, 78, 79) and

the overall muscle fatigue brought about by the combination of all of these factors. Only by

systematically exploring the dependence of aerobic fitness of all of the factors described in this

section can we begin to understand the system in an integrated fashion.

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Figure 11: Schematic of the variables and processes in the proposed ODE-based mathematical
model. Adapted from Timischl (1998) (66) and Batzel _et al._ (2005) (80).

LIMITATIONS

A limitation with the current study is the utilisation of a relatively small sample size, and this may be
contributing towards aforementioned errors. Future studies should seek to utilise CPET collected
annually in CF centres, to develop larger, multi-centre, samples whereby a uniform exercise protocol
is utilised. Given that utilisation of CPET is now recommended and endorsed for regular use by
international medical societies (81), and individual CF centres are reporting upon experiences of
using CPET (82), large-scale utilisation of such data is a feasible target.

The findings presented may be derived from a smaller sample, and therefore the models presented are
only preliminary results for this patient cohort; however, the study provides a unique examination
into the aerobic and anaerobic signatures of individual patients with CF in response to progressive
exercise.

Finally, whilst this study provides an insight into metabolic process during exercise, future research

and models must account for additional variables predictive of function and mortality (e.g. genotype,

body composition, pancreatic sufficiency, infection status, exacerbations (83, 84)) and co-morbidities

(CF-related diabetes (85), pulmonary arterial hypertension (86)) existent within CF, notably those

that may affect exercise tolerance.

CONCLUSIONS

Benefits of modelling include the ability to utilise existing data sets at a time when there are limited

resources. There is also a call to reduce the burden on sick patients (EU directive). As models

improve and the quality of fits to data are improved, the models can be used in a prognostic setting to

predict potential improvements in aerobic fitness that may arise due to therapeutic intervention.

Moreover, with proper mechanistic modelling of the primary organs affected in CF, there exists the

potential to optimise treatment for this patient group by identifying the limiting factors of aerobic

fitness.

CONFLICT OF INTEREST

The authors declare that the research was conducted in the absence of any commercial or financial

relationships that could be construed as a potential conflict of interest.

AUTHOR CONTRIBUTIONS

CAW conception, design of the work, data collection, analysis, interpretation, manuscript drafting,

final approval of the version to be published, agreement to be accountable for all aspects of the work

in ensuring questions related to the accuracy and integrity of any part of the work are appropriately

investigated and resolved; KTA data interpretation, critical review of the paper, final approval of the

version to be published, agreement to be accountable for all aspects of the work in ensuring questions

related to the accuracy and integrity of any part of the work are appropriately investigated and

resolve; OWT data collection, analysis, interpretation, final approval of the version to be published,

agreement to be accountable for all aspects of the work in ensuring questions related to the accuracy

and integrity of any part of the work are appropriately investigated and resolved; KW supervision of

work organization and development, data analysis, interpretation, manuscript drafting, final approval

of the version to be published, agreement to be accountable for all aspects of the work in ensuring

questions related to the accuracy and integrity of any part of the work are appropriately investigated

and resolved. HM construction of GP emulator, final approval of the version to be published,
agreement to be accountable for all aspects of the work in ensuring questions related to the accuracy
and integrity of any part of the work are appropriately investigate and resolved.

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Supplementary Material

Supplementary Material should be uploaded separately on submission, if there are Supplementary
Figures, please include the caption in the same file as the figure. Supplementary Material templates
can be found in the Frontiers Word Templates file.

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REFERENCES

1.   Cystic Fibrosis Trust. UK Cystic Fibrosis Registry 2015 Annual Data Report. 2016.

2.   Pérez M, Groeneveld IF, Santana-Sosa E, Fiuza-Luces C, Gonzalez-Saiz L, Villa-Asensi JR,
et al. Aerobic fitness is associated with lower risk of hospitalization in children with cystic fibrosis.
Pediatric Pulmonology. 2014;49(7):641-9.

3.   Andréasson B, Jonson B, Kornfält R, Nordmark EVA, Sandström S. Long-term Effects of
Physical Exercise on Working Capacity and Pulmonary Function in Cystic Fibrosis. Acta Paediatrica.
1987;76(1):70-5.

4.   Orenstein DM. Exercise conditioning and cardiopulmonary fitness in cystic fibrosis. The
effects of a three-month supervised running program. Chest. 1981;80(4):392.

5.   Salh W, Bilton D, Dodd M, Webb AK. Effect of exercise and physiotherapy in aiding sputum
expectoration in adults with cystic fibrosis. Thorax. 1989;44(12):1006-8.

6.   Schneiderman-Walker J, Pollock SL, Corey M, Wilkes DD, Canny GJ, Pedder L, et al. A
randomized controlled trial of a 3-year home exercise program in cystic fibrosis. The Journal of
Pediatrics. 2000;136(3):304-10.

7.   Nixon PA, Orenstein DM, Kelsey SF, Doershuk CF. The prognostic value of exercise testing
in patients with cystic fibrosis. New England Journal of Medicine. 1992;327(25):1785-8.

8.   Wheatley CM, Wilkins BW, Snyder EM. Exercise is medicine in cystic fibrosis. Exercise and
Sport Sciences Reviews. 2011;39(3):155-60.

9.   Stevens D, Oades PJ, Armstrong N, Williams CA. Exercise metabolism during moderate-
intensity exercise in children with cystic fibrosis following heavy-intensity exercise. Applied
Physiology, Nutrition, and Metabolism. 2011;36(6):920-7.

10.   Williams CA, Stevens D. Physical activity and exercise training in young people with cystic
fibrosis: Current recommendations and evidence. Journal of Sport and Health Science. 2013;2(1):39-
46.

11.   Angelis A, Kanavos P, Lopez-Bastida J, Linertova R, Nicod E, Serrano-Aguilar P, et al.
Social and economic costs and health-related quality of life in non-institutionalised patients with
cystic fibrosis in the United Kingdom. BMC Health Services Research. 2015;15(1):428.

12.   Stevens D, Oades PJ, Armstrong N, Williams CA. A survey of exercise testing and training in
UK cystic fibrosis clinics. Journal of Cystic Fibrosis. 2010;9(5):302-6.

13.   Goodson HV, Gregoretti IV. Using Computational Modeling to Understand Microtubule
Dynamics. Methods in Cell Biology. 2010;95:175-88.

14.   Hardman JG, Ross JJ. Modelling: a core technique in anaesthesia and critical care research.
British Journal of Anaesthesia. 2006;97(5):589-92.

15.   Holford N, Ma SC, Ploeger BA. Clinical trial simulation: a review. Clinical Pharmacology
and Therapeutics. 2010;88(2):166-82.

16.   Kapellos GE, Alexiou TS, Payatakes AC. Theoretical modeling of fluid flow in cellular
biological media: an overview. Mathematical Biosciences. 2010;225(2):83-93.

17.   Seethala RR, Esposito EC, Abella BS. Approaches to improving cardiac arrest resuscitation
performance. Current Opinion in Critical Care. 2010;16(3):196-202.

18.   McCahon RA, Columb MO, Mahajan RP, Hardman JG. Validation and application of a high-
fidelity, computational model of acute respiratory distress syndrome to the examination of the indices
of oxygenation at constant lung-state. British Journal of Anaesthesia. 2008;101(3):358-65.

19.   Mintun MA, Lundstrom BN, Snyder AZ, Vlassenko AG, Shulman GL, Raichle ME. Blood
flow and oxygen delivery to human brain during functional activity: Theoretical modeling and
experimental data. Proceedings of the National Academy of Sciences of the United States of
America. 2001;98(12):6859-64.

20.   Noble D. Modeling the heart--from genes to cells to the whole organ. Science.
2002;295(5560):1678-82.

21.   Moppett IK, Hardman JG. Development and Validation of an Integrated Computational
Model of Cerebral Blood Flow and Oxygenation. Anesthesia & Analgesia. 2007;105(4):1094-103.

22.   Murkin JM, Arango M. Near-infrared spectroscopy as an index of brain and tissue
oxygenation. British Journal of Anaesthesia. 2009;103(Supplement 1):i3-i13.

23.   Hermans MP, Brotons C, Elisaf M, Michel G, Muls E, Nobels F. Optimal type 2 diabetes
mellitus management: the randomised controlled OPTIMISE benchmarking study: baseline results
from six European countries. European Journal of Preventive Cardiology. 2013;20(6):1095-105.

24.   Saynor ZL, Barker AR, Oades PJ, Williams CA. A protocol to determine valid VO2max in
young cystic fibrosis patients. Journal of Science and Medicine in Sport. 2013;16(6):539-44.

25.   Saynor ZL, Barker AR, Oades PJ, Williams CA. Reproducibility of maximal
cardiopulmonary exercise testing for young cystic fibrosis patients. Journal of Cystic Fibrosis.
2013;12(6):644-50.

26.   Hulzebos HJ, Werkman MS, van Brussel M, Takken T. Towards an individualized protocol
for workload increments in cardiopulmonary exercise testing in children and adolescents with cystic
fibrosis. Journal of Cystic Fibrosis. 2012;11(6):550-4.

27.   Barker AR, Williams CA, Jones AM, Armstrong N. Establishing maximal oxygen uptake in
young people during a ramp cycle test to exhaustion. British Journal of Sports Medicine.
2011;45(6):498-503.

28.   Sacks J, Welch WJ, Mitchell TJ, Wynn HP. Design and analysis of computer experiments.
Statistical Science. 1989:409-23.

29.   Kleijnen JP. Kriging metamodeling in simulation: A review. European Journal of Operational
Research. 2009;192(3):707-16.

30.   Jin R, Chen W, Simpson TW. Comparative studies of metamodelling techniques under
multiple modelling criteria. Structural and Multidisciplinary Optimization. 2001;23(1):1-13.

31.   Forrester AI, Keane AJ. Recent advances in surrogate-based optimization. Progress in
Aerospace Sciences. 2009;45(1):50-79.

32.   Chen VC, Tsui K-L, Barton RR, Meckesheimer M. A review on design, modeling and
applications of computer experiments. IIE Transactions. 2006;38(4):273-91.

33.   Schwaighofer A, Grigoras M, Tresp V, Hoffmann C, editors. GPPS: A Gaussian process
positioning system for cellular networks. Advances in Neural Information Processing Systems; 2004.

34.   Bailer-Jones CAL, Sabin TJ, MacKay DJC, Withers PJ, editors. Prediction of deformed and
annealed microstructures using Bayesian neural networks and Gaussian processes. Proceedings of the
Australasia Pacific Forum on Intelligent Processing and Manufacturing of Materials; 1997.

35.   Kalaitzis AA, Lawrence ND. A simple approach to ranking differentially expressed gene
expression time courses through Gaussian process regression. BMC Bioinformatics. 2011;12(1):180.

36.   Lawrence ND, Sanguinetti G, Rattray M. Modelling transcriptional regulation using Gaussian
processes. Advances in Neural Information Processing Systems. 2007;19:785.

37.   Swain PS, Stevenson K, Leary A, Montano-Gutierrez LF, Clark IB, Vogel J, et al. Inferring
time derivatives including cell growth rates using Gaussian processes. Nature Communications.
2016;7.

38.   Taylor-Robinson D, Whitehead M, Diderichsen F, Olesen HV, Pressler T, Smyth RL, et al.
Understanding the natural progression in% FEV1 decline in patients with cystic fibrosis: a
longitudinal study. Thorax. 2012;67(10):860-6.

39.   Vilozni D, Bentur L, Efrati O, Minuskin T, Barak A, Szeinberg A, et al. Spirometry in early
childhood in cystic fibrosis patients. Chest. 2007;131(2):356-61.

40.   Kerem E, Reisman J, Corey M, Canny GJ, Levison H. Prediction of mortality in patients with
cystic fibrosis. New England Journal of Medicine. 1992;326(18):1187-91.

41.   Beaver WL, Wasserman K, Whipp BJ. A new method for detecting anaerobic threshold by
gas exchange. Journal of Applied Physiology. 1986;60(6):2020-7.

42.   Rasmussen C, Williams C. Gaussian processes for machine learning (Adaptive computation
and machine learning series) Cambridge, MA: The MIT Press; 2005.

43.   Thin AG, Linnane SJ, McKone EF, Freaney R, FitzGerald MX, Gallagher CG, et al. Use of
the gas exchange threshold to noninvasively determine the lactate threshold in patients with cystic
fibrosis. Chest. 2002;121(6):1761-70.

44.   Thin AG, Dodd JD, Gallagher CG, Fitzgerald MX, McLoughlin P. Effect of respiratory rate
on airway deadspace ventilation during exercise in cystic fibrosis. Respiratory Medicine.
2004;98(11):1063-70.

45.   Jones DR, Schonlau M, Welch WJ. Efficient global optimization of expensive black-box
functions. Journal of Global Optimization. 1998;13(4):455-92.

46.   Cooper PJ, Robertson CF, Hudson IL, Phelan PD. Variability of pulmonary function tests in
cystic fibrosis. Pediatric Pulmonology. 1990;8(1):16-22.

47.   Balfour-Lynn IM. Personalised medicine in cystic fibrosis is unaffordable. Paediatric
Respiratory Reviews. 2014;15 Suppl 1:2-5.

48.   Titsias MK, editor Variational Learning of Inducing Variables in Sparse Gaussian Processes.
AISTATS; 2009.

49.   Mazzoleni MJ, Battaglini CL, Martin KJ, Coffman EM, Mann BP. Modeling and predicting
heart rate dynamics across a broad range of transient exercise intensities during cycling. Sports
Engineering. 2016;19(2):117-27.

50.   Stirling JR, Zakynthinaki M, Refoyo I, Sampedro J. A Model of Heart Rate Kinetics in
Response to Exercise. Journal of Nonlinear Mathematical Physics. 2008;15(sup3):426--36.

51.   Ursino M. Interaction between carotid baroregulation and the pulsating heart: a mathematical
model. American Journal of Physiology Heart and Circulatory Physiology. 1998;275(5):H1733-H47.

52.   Zakynthinaki MS. Modelling heart rate kinetics. PLoS ONE. 2015;10(4):1--26.

53.   Gutierrez G. A mathematical model of tissue–blood carbon dioxide exchange during hypoxia.
American Journal of Respiratory and Critical Care Medicine. 2004;169(4):525-33.

54.   Bates JHT. Lung mechanics: an inverse modeling approach. Cambridge: Cambridge
University Press; 2009.

55.   Reynolds A, Ermentrout GB, Clermont G. A mathematical model of pulmonary gas exchange
under inflammatory stress. Journal of Theoretical Biology. 2010;264(2):161-73.

56.   Crooke P, Hotchkiss J, Marini J. Linear and nonlinear mathematical models for noninvasive
ventilation. Mathematical and Computer Modelling. 2002;35(11-12):1297-313.

57.   Kek K, Miyakawa T, Yoneyama S, Kudo N, Yamamoto K, editors. Simulation of exercise-
dependent difference in metabolism with a mathematical model for analyses of measurements using
near-infrared spectroscopy. Engineering in Medicine and Biology Society, 2006 EMBS'06 28th
Annual International Conference of the IEEE; 2006: IEEE.

58.   Krauss M, Schaller S, Borchers S, Findeisen R, Lippert J, Kuepfer L. Integrating cellular
metabolism into a multiscale whole-body model. PLoS Computational Biology.
2012;8(10):e1002750.

59.   Li Y, Lai N, Kirwan JP, Saidel GM. Computational model of cellular metabolic dynamics in
skeletal muscle fibers during moderate intensity exercise. Cellular and Molecular Bioengineering.
2012;5(1):92-112.

60.   Lai N, Camesasca M, Saidel GM, Dash RK, Cabrera ME. Linking pulmonary oxygen uptake,
muscle oxygen utilization and cellular metabolism during exercise. Annals of Biomedical
Engineering. 2007;35(6):956-69.

61.   Cloutier M, Wellstead P. The control systems structures of energy metabolism. Journal of The
Royal Society Interface. 2009:rsif20090371.

62.   Dash RK, Dibella Ja, Cabrera ME. A computational model of skeletal muscle metabolism
linking cellular adaptations induced by altered loading states to metabolic responses during exercise.
Biomedical Engineering Online. 2007;6:14.

63.   Kim J, Saidel GM, Cabrera ME. Multi-scale computational model of fuel homeostasis during
exercise: effect of hormonal control. Annals of Biomedical Engineering. 2007;35(1):69-90.

64.   Fresiello L, Meyns B, Di Molfetta A, Ferrari G. A Model of the Cardiorespiratory Response
to Aerobic Exercise in Healthy and Heart Failure Conditions. Frontiers in Physiology. 2016;7.

65.   Fresiello L, Ferrari G, Di Molfetta A, Zieliński K, Tzallas A, Jacobs S, et al. A cardiovascular
simulator tailored for training and clinical uses. Journal of Biomedical Informatics. 2015;57:100-12.

66.   Timischl S. A Global Model for the Cardiovascular and Respiratory System: University of
Graz; 1998.

67.   D'Angelo C. Multiscale modelling of metabolism and transport phenomena in living tissues:
École Polytechnique Fédérale de Lausanne; 2007.

68.   Hardman J, Bedforth N, Ahmed A, Mahajan R, Aitkenhead A. A physiology simulator:
validation of its respiratory components and its ability to predict the patient's response to changes in
mechanical ventilation. British Journal of Anaesthesia. 1998;81(3):327-32.

69.   Lai N, Saidel GM, Grassi B, Gladden LB, Cabrera ME. Model of oxygen transport and
metabolism predicts effect of hyperoxia on canine muscle oxygen uptake dynamics. Journal of
Applied Physiology. 2007;103(4):1366-78.

70.   Benson AP, Grassi B, Rossiter HB. A validated model of oxygen uptake and circulatory
dynamic interactions at exercise onset in humans. Journal of Applied Physiology. 2013;115(5):743-
55.

71.   Bell C, Paterson DH, Kowalchuk JM, Padilla J, Cunningham DA. A Comparison of
Modelling Techniques used to Characterise Oxygen Uptake Kinetics During the on-Transient of
Exercise. Experimental Physiology. 2001;86(5):667-76.

72.   Elstad M, Toska K, Walløe L. Model simulations of cardiovascular changes at the onset of
moderate exercise in humans. The Journal of Physiology. 2002;543(2):719-28.

73.   Ng LJ, Sih BL, Stuhmiller JH. An integrated exercise response and muscle fatigue model for
performance decrement estimates of workloads in oxygen-limiting environments. European Journal
of Applied Physiology. 2012;112(4):1229-49.

74.   Walter G, Vandenborne K, Elliott M, Leigh JS. In vivo ATP synthesis rates in single human
muscles during high intensity exercise. The Journal of Physiology. 1999;519(3):901-10.

75.   Eriksson A, Holmberg H-C, Westerblad H. A numerical model for fatigue effects in whole-
body human exercise. Mathematical and Computer Modelling of Dynamical Systems. 2016;22(1):21-
38.

76.   Liu JZ, Brown RW, Yue GH. A dynamical model of muscle activation, fatigue, and recovery.
Biophysical Journal. 2002;82(5):2344-59.

77.   Dash RK, Li Y, Kim J, Saidel GM, Cabrera ME. Modeling cellular metabolism and
energetics in skeletal muscle: large-scale parameter estimation and sensitivity analysis. IEEE
Transactions on Biomedical Engineering. 2008;55(4):1298-318.

78.   Moxnes JF, Sandbakk Ø. The kinetics of lactate production and removal during whole-body
exercise. Theoretical Biology and Medical Modelling. 2012;9(1):7.

79.   Péronnet F, Aguilaniu B. Lactic acid buffering, nonmetabolic CO 2 and exercise
hyperventilation: a critical reappraisal. Respiratory Physiology & Neurobiology. 2006;150(1):4-18.

80.   Batzel JJ, Kappel F, Timischl-Teschl S. A cardiovascular-respiratory control system model
including state delay with application to congestive heart failure in humans. Journal of Mathematical
Biology. 2005;50(3):293-335.

81.   Hebestreit H, Arets HG, Aurora P, Boas S, Cerny F, Hulzebos EH, et al. Statement on
Exercise Testing in Cystic Fibrosis. Respiration. 2015;90(4):332-51.

82.   Weir E, Burns PD, Devenny A, Young D, Paton JY. Cardiopulmonary exercise testing in
children with cystic fibrosis: one centre's experience. Archives of disease in childhood.
2017;102(5):440-4.

83.   Liou TG, Adler FR, Fitzsimmons SC, Cahill BC, Hibbs JR, Marshall BC. Predictive 5-Year
Survivorship Model of Cystic Fibrosis. American Journal of Epidemiology. 2001;153(4):345-52.

84.   Aaron SD, Stephenson AL, Cameron DW, Whitmore GA. A statistical model to predict one-
year risk of death in patients with cystic fibrosis. Journal of Clinical Epidemiology.
2015;68(11):1336-45.

85.   Middleton PG, Matson AG, Robinson PD, Jane Holmes-Walker D, Katz T, Hameed S. Cystic
Fibrosis Related Diabetes: Potential pitfalls in the transition from paediatric to adult care. Paediatric
Respiratory Reviews. 2014;15(3):281-4.

86.   Hayes D, Jr., Tobias JD, Mansour HM, Kirkby S, McCoy KS, Daniels CJ, et al. Pulmonary

hypertension in cystic fibrosis with advanced lung disease. American Journal of Respiratory and

Critical Care Medicine. 2014;190(8):898-905.

<u>**Figure Titles**</u>

Figure 1: (a) The work rate for each participant is increased at a rate dependent on their past

test performance. (b) Participant age is correlated with test performance for young

participants, but not for older ones.

Figure 2: Ratio of oxygen utilisation and total breathing throughout the test. Markers indicate

the volitional exhaustion times for each participant.

Figure 3: (a) Correlation of FEV_1 with the maximal tidal volume achieved throughout the test.

(b) Correlation between FEV_1 and FVC is high. Note that, although FVC and FEV_1 are good

predictors of poor test performance, they are unable to distinguish better performing

participants.

Figure 4: (a) Total ventilation plotted against oxygen utilisation. We observe that breathing

pattern is strongly correlated with test performance. (b) Exponential curves are fitted through

the raw data, further highlighting this dependence.

Figure 5: Slope of the fitted curves ($\log\dot{V}E$ against $\dot{V}O_2$) from Fig. 4(b) plotted against the total

energy transfer during the test. We find a relatively poor characterisation of the variance

between performances. (b) By instead plotting the oxygen consumption at a fixed rate of

breathing, we better capture differences in performance.

Figure 6: (a) Breathing patterns subdivided into the breathing rate and tidal volume. These

data appear uninformative for predicting test performance. (b) With the additional inclusion of

the oxygen consumption at a fixed rate of breathing, we find that these variables now almost

perfectly capture variation in participant performance.

Figure 7: First of the principal components obtained via PCA accounts for over 90% of the

variation in test performance. (b) Similar levels of variance are accounted for by taking only

the normal component of the first principal component in the breathing frequency direction.

Figure 8: (a) Quantifying the relationship between the anaerobic threshold and overall test
performance. Anaerobic thresholds were calculated using an automated procedure based on
previous methods (41) (b) $\dot{V}O_{2max}$ is the best single predictor of overall test performance.

Figure 9: GP is a probability distribution on function spaces. The thick solid red line is the true
function f and the thick dashed black line is the GP mean. (a) Thin grey lines show sample
paths of a prior GP whose mean is zero. (b) Blue bullets indicate five data points sampled from
f. The GP mean and covariance are updated using these sample points. The thin grey dashed
lines show $m(x) \pm 2s(x)$.

Figure 10: (a) Predictions (asterisks) vs. exact values (bullets). Red bars show the 95%
confidence intervals, $m_i(x_i) \pm 1.96 \, s_i(x_i)$ around the predicted value, where m_i and s_i are GP
prediction mean and standard deviation based on all but the i'th data point. (b) Relative
prediction error expressed as a percentage of the true value $f(x_i)$.

Figure 11: Schematic of the variables and processes in the proposed ODE-based mathematical
model. Adapted from Timischl (1998) (66) and Batzel *et al.* (2005) (80).