

1 **Is it possible to keep the exoskeleton of the crab *Callinectes***
2 ***ornatus* soft for several days?**

3 Short Title: **Effects of water quality on the molt of crab**

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14 **Abstract**

15 Soft-shell crab is considered a gastronomic delicacy, reaching high values in the
16 international market. The process of hardening of the crab's exoskeleton after
17 moulting takes approximately two days to complete; however, the duration for which
18 the shell remains at the consistency of high commercial value is only 3 hours on
19 average. After this period, the shell assumes a consistency classified as "paper",
20 later becoming "hard" again. The goal of this work was to evaluate the use of the
21 crabs themselves to alter the chemical characteristics of the water and thereby
22 increase the amount of time during which they can be marketed as "soft-shell crab".
23 In this work, 241 individuals of *Callinectes ornatus* were used in two experiments. In
24 the first experiment, the animals were maintained in a collective system with filtration
25 and partial daily water renewal. In the second experiment, the crabs were maintained
26 in a collective system with filtration but no water renewal. In Experiment 1, the
27 chemical characteristics of the water remained unchanged over time ($p > 0.05$), and
28 the median time to hardening of the exoskeleton to the paper consistency after
29 moulting was 3 hours. Over the course of Experiment 2, there was a significant
30 reduction ($p < 0.05$) in pH and significant increases in the ammonia and nitrite
31 concentrations. When moulting occurred in water with a pH below 7.3 and total
32 ammonia concentrations above 6.0 mg/L, the crabs' shells did not harden, and it was
33 possible to keep them soft for up to 5 days.

34 **Introduction**

35 *Callinectes ornatus* Ordway, 1863 (Crustacea, Decapoda, Portunidae) is a
36 swimmer crab found from North Carolina (USA) to the Rio Grande do Sul (Brazil). It
37 occurs in areas with sand, mud or shell bottoms and inhabits estuarine to marine

38 areas at a depth of approximately 75 m (Carvalho and Couto, 2011; Melo-Filho,
39 1996).

40 Similar to other arthropods, *C. ornatus* grows through a process of periodic
41 exoskeleton changes; each shedding of the exoskeleton is known as ecdysis or
42 moult (Drach, 1939; Freeman and Perry, 1985; Newcombe et al., 1949). Immediately
43 after shedding its exoskeleton, the crab presents a soft and flexible integument that
44 has a low level of calcification. In this phase, the animals can be commercialised and
45 consumed whole as "soft-shell crab", a delicacy that is appreciated worldwide and
46 that reaches high market values (Gaudé and Anderson, 2011; Oesterling, 1995;
47 Perry et al., 2010). According to FAO (2013), the annual revenue generated from the
48 production and marketing of soft-shell crab in 2012 was more than US\$ 940 million.

49 Immediately after moult, CaCO_3 deposition begins on the protein matrix of the
50 new exoskeleton. This process involves a complex system of absorption of Ca^{2+} ,
51 CO_2 , and HCO_3^- and the synthesis of CaCO_3 and other elements (Greenaway, 1985;
52 Perry et al., 2001; Wheatly, 1999; Zanotto and Wheatly, 2002). The initially fragile
53 exoskeleton undergoes rapid hardening, providing rigidity and mechanical protection
54 for the animal. Under natural conditions, the hardening of the exoskeleton takes
55 about two days to complete (Cameron and Wood, 1985). During the hardening
56 process, the exoskeleton can be classified into four sequential levels of consistency:
57 soft, leather, paper and hard (Freeman et al., 1987). Only the first two are valued in
58 the international market of soft-shell crabs (Gaudé and Anderson, 2011; Oesterling,
59 1995; Perry et al., 2010). However, the combined duration of the soft and leathery
60 stages is very short in nature, rarely lasting more than 3 h (Cameron and Wood,
61 1985), which obliges commercial producers to inspect all of the animals stocked in

62 the pre-moulting phase every 4 h on average (Oesterling, 1995). Extending the
63 duration in which the crab shells remain at the consistencies of high market value
64 would significantly reduce production costs (Perry et al., 2001). Furthermore, it would
65 minimise the damage caused by rapid exoskeleton hardening, providing better
66 quality and uniformity regarding the softness of the product.

67 The goal of this work was to test the viability of using the crab *C. ornatus* to alter the
68 chemical characteristics of the water to extend the time during which the animals
69 could be marketed as soft-shell crab.

70 **Material and Methods**

71 **Crab collection and maintenance**

72 Specimens of *C. ornatus* were obtained via trawling by professional fishers at
73 the balneary of Shangri-la, municipality of Pontal do Paraná (25°37'S/48°25'O),
74 Paraná, Brazil. Shrimp trawls 12 m in length and with 20 mm mesh were used. In
75 each sampling campaign, on average, three trawls of approximately 50 min each
76 were made. Immediately after crab collection from the net, the crabs were separated
77 and transferred to two 70 L polyethylene tanks with lids, each containing 20 L of
78 seawater. The tanks received continuous aeration supplied via an 18 W air
79 compressor. Inside each tank were plastic screens with 2 mm mesh positioned to
80 reduce contact and prevent fights between the animals and minimise injuries and
81 deaths. Thereafter, 100% of the water was renewed every half hour during the
82 campaign.

83 Immediately after capture, the animals were transported to the Marine
84 Aquaculture and Restocking Center (CAMAR) of the Integrated Group of Aquaculture

85 and Environmental Studies (GIA), Federal University of Paraná (UFPR), at Pontal do
86 Paraná (25°41'29.94"S, 48°27'57.09"W). The time elapsed between animal capture
87 and arrival at the laboratory was consistently less than 4 h. Animals that were not
88 used were returned to the sea.

89 In the laboratory, the crabs were maintained in 1,000 L tanks containing 100 L
90 of seawater (30 psu) supplied with constant aeration for approximately 6 h. This
91 period was purposely short since a large proportion of the captured individuals were
92 very close to moult. Dead animals were discarded, and the live animals were
93 classified by sex. Then, the crabs were inspected to determine the phase of the
94 moulting cycle. Those individuals at the pre-ecdysis phase were selected for the
95 experiments based on macroscopic indicators (visualisation of an inner line along the
96 edges of the fifth pair of pleopods) (Drach, 1939; Drach and Tchernigovtzeff, 1967;
97 Wehrtmann and Mena-Castañeda, 2003). The selected individuals were weighed on
98 an analytical balance (Marte AL 500c, Brazil; accuracy of 0.01 g) and measured
99 (width of the carapace, measured as the distance between the base of the largest
100 lateral spines) with a pachymeter.

101 **Pilot experiments**

102 Two pilot experiments were carried out. The first experiment tested the
103 influence of fasting on animal survival under laboratory conditions. The animals only
104 began to die after 50 days without access to food. Based on this result and to
105 potential feeding effects on water quality or the process of moult and hardening, the
106 crabs were not fed during the 12 days of each of the main experiments.

107 The second experiment tested the influence of the non-renewal of water on
108 hardening time. The time elapsed between moult and shell hardening was

109 significantly higher under water non-renewal than under the periodic renewal of
110 water. In addition, a higher frequency of moult was observed at night (between 18:00
111 and 06:00); this information informed the design of the experimental methodology
112 described below.

113 **Experimental Design**

114 In both experiments, the saltwater had been previously chlorinated and
115 maintained under constant aeration for 24 h. After this period, residual chlorine was
116 neutralised (with 50% sodium thiosulfate), and the water was stored in the dark in
117 25,000 L tanks. Before use in the experiments, the water was passed through
118 mechanical filters of 5 and 25 µm mesh and a UV filter for disinfection. Two
119 experiments were performed and are represented schematically in Figure 1.

120 **Experiment 1: Crab maintenance in a collective system with filtration** 121 **and partial daily water renewal**

122 Sixty-six *C. ornatus* crabs were individually placed in perforated pet bottles
123 (600 mL) and distributed in a system consisting of 20 polyethylene tanks (71.0 x 35.5
124 x 35.0 cm, containing 25 L of seawater each). The tanks were interconnected via a
125 skimmer and a mechanical/biological filter system and were under constant aeration,
126 continuous water recirculation and controlled photoperiod (14L:10D).

127 The animals were separated into two groups: A1, pre-ecdysis animals (n =
128 46), and AC, control animals (at the inter-ecdysis stage) (n = 20). Each day
129 throughout the experimental period (12 days), 1/3 of the total water volume of the
130 system (333 L) was added, promoting mixing with the water already present, and an

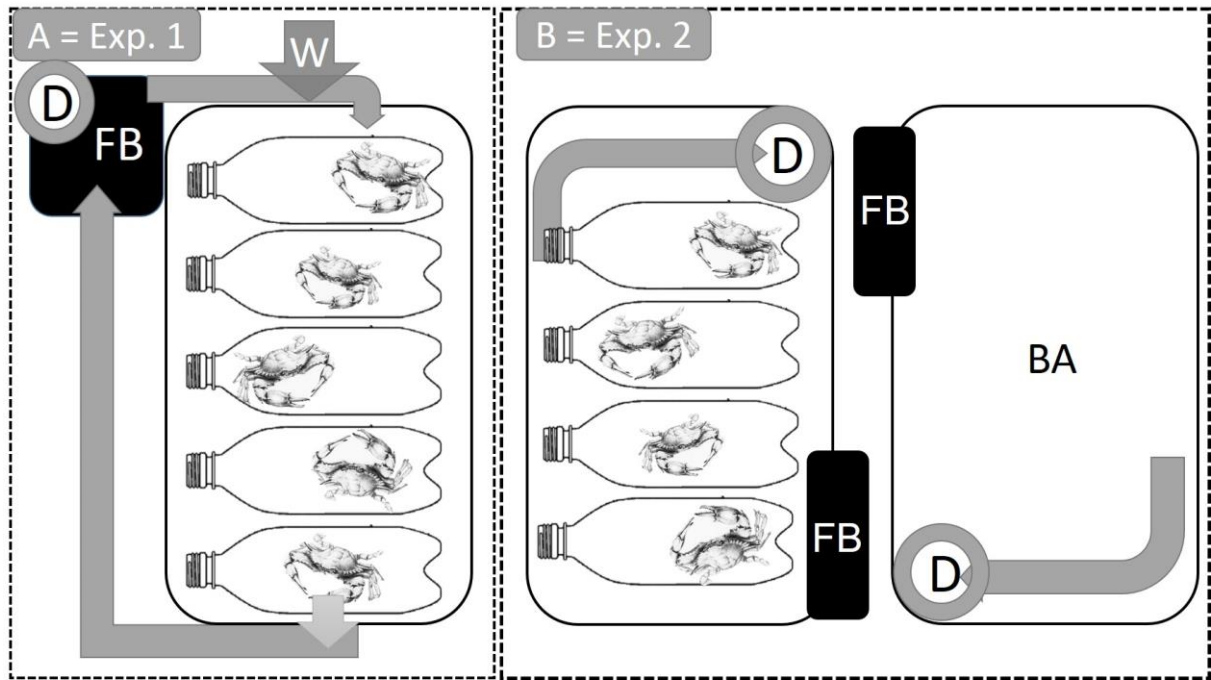
131 approximately equivalent amount was removed, keeping the total water volume in the
132 system constant.

133 Experiment 2: **Crab maintenance in a collective system with filtration**
134 **and without water renewal**

135 One hundred and seventy-six *C. ornatus* crabs were individually placed in
136 perforated (600 mL) pet bottles and distributed among 12 polyethylene tanks (71.0 x
137 35.5 x 35.0 cm, containing approximately 30 L of water each). Each tank contained a
138 protein skimmer and a mechanical/biological filter and was subjected to constant
139 aeration, continuous water recirculation and controlled photoperiod (14L:10D).

140 The animals were subdivided into 3 groups: B1, pre-ecdysis animals (n = 83);
141 B2, pre-ecdysis animals (n = 40); and BC, control inter-ecdysis animals (n = 52). In
142 addition, three tanks containing water only were maintained throughout the
143 experimental period (12 days) for comparison of physical and chemical water
144 variables between these tanks and the 3 treatment groups. There was no water
145 renewal during the experiment.

146 The animals in the B2 group were housed in the same tanks used to house
147 group B1 and maintained in the same water used for group B1.



148

149 Figure 1. Schematic representations of the experimental systems. A: Experiment 1 (Exp. 1): 66 crabs
 150 (46 in pre-ecdysis stage and 20 in inter-ecdysis stage) maintained in a collective system with filtration
 151 and partial daily water renewal. B: Experiment 2 (Exp. 2): 175 crabs (123 in pre-ecdysis stage and 52
 152 in inter-ecdysis stage) maintained in a collective system with filtration but without water renewal. A =
 153 external water supply; D = protein skimmer; FB = mechanical / biological filter; BA = Water control.
 154

155 Table 1 provides summary information on the subject animals and design of
 156 the two experiments.

157 Table 1. Summary of the general conditions of the experiments performed to evaluate the effects of
 158 water quality on the hardening time of the exoskeleton in *Callinectes ornatus* and width and weight
 159 data of the animals (mean \pm SD). NA: not applicable.

Experiment	Group	Experimental Unit	Stage	N	Width (mm) (\pm sd)	Weight (g) (\pm sd)	Water Volume (L)	Water Sampling Frequency
1	A1	Tank	Pre-ecdysis	46	45.4 (\pm 6.16)	15.04 (\pm 6.11)	0.6	24 h
	AC	Tank	Inter-ecdysis	20	61.55 (\pm 10.53)	35.57 (\pm 16.9)	0.6	24 h
2	B1	Tank (Replicate 1)	Pre-ecdysis	27	50.7 (\pm 7.78)	16.64 (\pm 7.03)	32.5	12 h
		Tank (Replicate 2)		27	3.10 (\pm 7.36)	14.36 (\pm 7.47)	32.5	12 h
		Tank (Replicate 3)		29	47.5 (\pm 5.12)	15.11 (\pm 4.92)	32.5	12 h
	B2	Tank (Replicate 1)	Pre-ecdysis	14	48.1 (\pm 8.25)	16.46 (\pm 8.08)	30	12 h
		Tank (Replicate 2)		13	46.4 (\pm 7.02)	15.40 (\pm 6.7)	30	12 h
		Tank (Replicate 3)		13	47.0 (\pm 6.64)	14.75 (\pm 6.3)	30	12 h
	BC	Tank (Replicate 1)	Inter-ecdysis	17	56.2 (\pm 2.9)	26.51 (\pm 5.07)	32.5	12 h
		Tank (Replicate 2)		17	59.7	28.31	32.5	12 h

Experiment	Group	Experimental Unit	Stage	N	Width (mm) (\pm sd)	Weight (g) (\pm sd)	Water Volume (L)	Water Sampling Frequency
		2)			(\pm 3.8)	(\pm 6.1)		
		Tank (Replicate 3)		18	56.65 (\pm 4.93)	26.54 (\pm 7.95)	32.5	12 h
	BA	Tank (3 Replicates)	NA	NA	NA	NA	32.5	12 h

160 **Experimental Procedures**

161 During the experiments, the crabs were monitored every three hours on the
162 first four days, every six hours on the following five days, and every 12 hours on the
163 last three days of experimentation, preferably between 18:00 and 06:00 h. These
164 times were selected based on the results of the pilot experiments.

165 Monitoring consisted of identifying animals undergoing the moulting process,
166 removing any moulted exoskeletons (to prevent the animals from obtaining calcium
167 by feeding on them), evaluating the consistency of the carapace of those animals
168 that had moulted, and removing any dead animals. Evaluating the consistency of the
169 exoskeleton was performed by pressing the carapace with an index finger. Sufficient
170 pressure was applied to deform the carapace but not injure the animal or break the
171 carapace when rigid. Based on the resistance to pressure and texture of the
172 exoskeleton, its consistency (Co) was classified by the evaluator as follows: hard -
173 before ecdysis (1), soft (2), leather (3), soft paper (4), hard (5) or hard paper - after
174 ecdysis (6). To reduce and standardise the error, a single evaluator performed the
175 consistency assessments in both experiments.

176 **Water analysis**

177 In both experiments, salinity (refractometer; Instrutemp, Brazil), temperature
178 (digital thermometer), pH (AZ pH/mV/TDS /Temperature Meter 86505, Taiwan), and
179 dissolved oxygen concentration (Oximeter YSI 550A, USA) were monitored daily in

180 all experimental units. Water samples were collected from the units, labelled and
181 immediately frozen (-20 ° C) for later evaluation of the physical and chemical
182 variables. For group A1, water collection was performed every 24 hours before the
183 new water was added to the system. For groups B1, B2, BC and BA, 50 mL of water
184 was collected every 12 h.

185 At the end of the experiments, the frozen water samples were analysed with
186 respect to the following parameters: Na⁺, K⁺, Ca²⁺ and NO₃ (electrodes of the
187 LAQUAtwin series, Horiba Scientific®, Japan) and total ammonia (NH₃+NH₄⁺) and
188 NO₂⁻ (SpectraMax® m2 spectrophotometer, USA). Measurements were performed
189 following APHA (2005) and Büldt and Karst (1999). The determinations of Mg₂⁺ and
190 Cl⁻ were performed using colourimetry (Labtest®, Brazil) at a wavelength of 540 nm
191 and 470 nm, respectively (SpectraMax® m2, USA), according to the method
192 described by Clarke (1950).

193 **Statistical analyses**

194 The survival of the animals during the experiments was analysed through
195 Kaplan-Meier curves. The data were grouped by treatment (groups), and the
196 normality of the distribution of each variable was tested by using Shapiro-Wilk test.
197 Where the normality hypothesis was rejected, non-parametric Mann-Whitney or
198 Kruskal-Wallis tests were used.

199 Multiple linear regression analysis was performed to model the influences of
200 the physical and chemical variables that determine water quality on exoskeleton
201 hardening time. The assumption of the independence of the physical and chemical
202 variables was upheld, the hypothesis of autocorrelation and collinearity (using

203 Durbin-Watson and the serial error correlation tests) was rejected, and the normality
204 of the error was confirmed.

205 To limit the number of variables and thereby minimise the complexity of the
206 models without a significant loss of the information offered by the total set of original
207 variables, we select only those variables that: 1) were statistically significant ($p <$
208 $0,05$) and; 2) contributed more than 5% to the coefficient of determination (R^2) of the
209 model or that caused the R^2 value to move into a higher category when it was
210 included in the model, following the classification proposed by Mukaka (2012): very
211 weak:: $R^2 < 0,19$; weak: $0,20 > R^2 < 0,39$; moderate: $0,40 > R^2 < 0,69$; strong: $0,70 >$
212 $R^2 < 0,89$; very strong: $R^2 > 0,90$.

213 **Results**

214 **Ecdysis**

215 Significant effects of sex on survival rate, ecdysis, or exoskeleton post-ecdysis
216 hardening time were not observed. Therefore, the data from males and females were
217 pooled. In addition, water temperature (27.0 ± 1.1 ° C), salinity (31.0 ± 2.1 ups) and
218 dissolved oxygen concentration (5.0 ± 0.52 mg/L) remained largely stable and did not
219 significantly influence any of the dependent variables.

220 The moulting rate of the animals in pre-ecdysis at the beginning of the
221 experiments ranged from 40 to 95%. Most moulting events occurred during the night,
222 and 50% of the animals moulted between 52 and 80 hours after the beginning of the
223 experiments. There was a significant effect of moulting on final mortality rate and on
224 survival time after ecdysis (Table 2).

225 Table 2. General results of laboratory experiments to evaluate ecdysis in *Callinectes ornatus*.

Exp.	Group	Stage	n ₁	Weight Gain (%)	Ecdysis Performed			Time to Ecdysis (h)			Mortality Rate (%)	Survival Time (h)		
					Period	n ₂	%	25%	50%	75%		25%	50%	75%
1	A1	Pre	46	56	day night	14 ^a 24 ^b	83	44.5	80	124	24 ^a	230	-	-
	AC	Inter	20	NA	NA	NA	NA	NA	NA	NA	0 ^b	-	-	-
2	B1	Pre	83	69	day night	19 ^a 59 ^b	94	36	52	78	78 ^c	107	168	212
	B2	Pre	40	46	day night	3 ^a 13 ^b	40	102	-	-	78 ^c	174	228	413
	BC	Inter	53	NA	NA	NA	NA	NA	NA	NA	26 ^a	255	-	-

226 *Exp.*: experiment number; *Pre*: pre-ecdysis; *Inter*: Inter-ecdysis; *n*₁: number of individuals; *Weight Gain*: increase
 227 *in post-ecdysis weight (%)*; *Period*: the period in which moult occurred; *n*₂: number and percentage of crabs that
 228 *performed ecdysis*; *Time to Ecdysis*: time (h) at which 25, 50 and 75% of the animals had moulted; *Mortality rate*
 229 *(%)*; *Survival Time*: time (h) at which 25, 50 and 75% of the animals survived after ecdysis; *NA*: Not Applicable.
 230 *Different letters indicate significant differences (p < 0.05) between the groups according to the Kruskal-Wallis test.*
 231 *Experiment 1 (collective treatment with filtration and partial daily water renovation). A1: pre-ecdysis organisms;*
 232 *AC (Control): organisms in inter-ecdysis. Experiment 2 (collective treatment with filtration but no water renewal).*
 233 *B1: pre-ecdysis organisms; B2: tanks containing water previously used for group B1, with organisms in pre-*
 234 *ecdysis; BC (Control): tanks with organisms in inter-ecdysis.*
 235

236 Physical and chemical water parameters

237 In Experiment 1, the total ammonia (NH₃+NH₄⁺) concentrations remained
 238 below the limit of analytical detection, and the median pH was 8.5, with variation
 239 between 8.1 and 8.5. The remaining physical and chemical parameters were
 240 relatively stable throughout the experimental period (Table 3).

241 Table 3. Median and 1st and 3rd quartiles of the water quality
 242 parameters in Experiment 1 (collective treatment with filtration and
 243 partial daily water renewal).

Parameter	Median	25-75%
pH	8.50	8.4-8.5
K ⁺ (mg/L)	380	370-390
Ca ²⁺ (mg/L)	350	330-430
Mg ²⁺ (mg/L)	589.5	573.3-602.6
Na ⁺ (mg/L)	11,000	9,900-12,000
Cl ⁻ (mg/L)	16,830	16,059-17,668
TA (mg/L)	0.0	0.0
NH ₃ (mg/L)	0.0	0.0
NO ₂ ⁻ (mg/L)	1.30	1.29-1.30
NO ₃ (mg/L)	180	170-200

TA: Total ammonia (NH₃+NH₄⁺)

244 In Experiment 2, only potassium and sodium concentrations presented
 245 differences between the groups B1 and B2. There was a reduction in pH and
 246 increases in total ammonia and nitrite concentrations in the experimental treatments
 247 (pre-ecdysis organisms, B1 and B2) in relation to the control (BA, tanks containing

248 water only). The variables monitored in the BC tanks (inter-ecdysis organisms)
 249 presented intermediate values relative to the other groups (Table 4).

250 Table 4. Median and 1st and 3rd quartiles of the water parameters in Experiment 2
 251 (collective treatment with filtration but without water renewal).

Parameter	Groups			
	B1	B2	BA	BC
	Median 25-75%	Median 25-75%	Median 25-75%	Median 25-75%
pH	6.7 ^b (6.3-7)	6.5 ^b (6.2-7)	8.4 ^a (8.3-8.4)	7.8 ^{ab} (6.8-8.1)
K ⁻ (mg/L)	420.00 ^a (370-540)	280.00 ^b (230-330)	330.00 ^{ab} (260-370)	390.00 ^{ab} (290-420)
Ca ²⁺ (mg/L)	430.00 ^a (400-480)	350.00 ^a (320-420)	430.00 ^a (360-490)	450.00 ^a (390-480)
Mg ²⁺ (mg/L)	545.6 ^b (472.3-585.8)	551.2 ^b (531.8-622.5)	586.3 ^{ab} (573.3-597.4)	592.0 ^a (555.2-607.9)
Na ⁺ (mg/L)	12,000 ^a (10,000-14,000)	7,500 ^b (6,500-9,000)	11,500 ^{ab} (10,000-13,000)	12,000 ^{ab} (10,000-14,000)
Cl ⁻ (mg/L)	18279 ^a (16,507-20,245)	11995 ^a (9,158-15,782)	15830 ^a (14,705-17,473)	17602 ^a (16,122-19,278)
TA (mg/L)	6.8 ^b (5.7-10.5)	9.5 ^b (7.4-11.91)	0.0 ^a 0.0	1.1 ^{ab} (0.0-3.1)
NH ₃ (mg/L)	0.02 ^b (0.01-0.05)	0.02 ^b (0.01-0.05)	0.00 ^a 0	0.01 ^{ab} (0-0.08)
NO ₂ ⁻ (mg/L)	4.7 ^b (2.8-6.5)	5.4 ^b (2.15-7.8)	1.3 ^a (1.3-1.4)	5.2 ^b (4.0-6.1)
NO ₃ (mg/L)	230 ^a (190-310)	260 ^a (200-340)	210 ^a (120-260)	230 ^a (150-280)

252 *B1: organisms in pre-ecdysis. B2: tanks containing water previously used for group B1 and pre-ecdysis crabs. BA*
 253 *(Control): tanks containing water only. BC (Control): tanks with crabs in inter-ecdysis. Different letters indicate*
 254 *significant differences (p < 0.05) between groups according to the Kruskal-Wallis test. TA: Total ammonia*
 255 *(NH₃+NH₄⁺).*

256 Influence of the physical and chemical water parameters on the 257 survival and moulting of *C. ornatus*

258 As expected, crab survival time was influenced by moulting regardless of the
 259 experiment. The organisms of group B1 that underwent moult in the first 36 h showed
 260 rapid exoskeleton hardening and low mortality rates. Therefore, the B1 data were
 261 divided into two categories: M1, animals that moulted within the first 36 h, and M2,
 262 those that moulted after 36 h. The survival of M1 animals was strongly influenced by
 263 pH and total ammonia and nitrite concentrations, whereas the survival of the M2
 264 animals was moderately influenced by the same variables.

265 Table 5 shows the multiple linear regression results. Crab survival rate was
 266 significantly influenced by pH, nitrite and total ammonia in all of the experiments. The
 267 remaining parameters had no significant influence ($p < 0.05$) on the crab survival
 268 time.

269 As expected, crab survival time was influenced by moulting regardless of the
 270 experiment. The organisms of group B1 that underwent moult in the first 36 h showed
 271 rapid exoskeleton hardening and low mortality rates. Therefore, the B1 data were
 272 divided into two categories: M1, animals that moulted within the first 36 h, and M2,
 273 those that moulted after 36 h. The survival of M1 animals was strongly influenced by
 274 pH and total ammonia and nitrite concentrations, whereas the survival of the M2
 275 animals was moderately influenced by the same variables.

276 Table 5. General results of multiple linear regression analysis of the influences of physical and
 277 chemical parameters on crab survival time.

Experiment	Group	Correlated Parameters	Cases n	p	Adjusted R ²	Correlation
1	A1	pH (max.)	384	0.000	0,372	Weak
		TA (min.)		0.000		
		NO ₂ ⁻ (min.)		0.000		
	AC	NA		NA	NA	NA
3	B1	pH (max.)	233	0.000	0,724	Strong
		TA (min.)		0.000		
		NO ₂ ⁻ (max)		0.000		
	M2	pH (max.)	318	0.000	0,681	Moderate
		TA (min.)		0.000		
		NO ₂ ⁻ (med.)		0.000		
3	B2	pH (max.)	495	0.000	0,430	Moderate
		TA (min.)		0.000		
		NO ₃ (med.)		0.000		
	BC	pH (max.)	452	0.000	0,743	Strong
TA (max.)	0.000					
NO ₂ ⁻ (max.)	0.000					

278 *Experiment 1 (collective treatment with filtration and partial daily water renovation). A1: pre-ecdysis crabs; AC*
 279 *(Control): inter-ecdysis crabs. Experiment 2: Collective treatment with filtration but no water renewal. B1: pre-*
 280 *ecdysis crabs; B2: tanks containing water previously used for the B1 group, with pre-ecdysis crabs; BC (Control):*
 281 *tanks with crabs in inter-ecdysis; M1: crabs of B1 Group that performed ecdysis within the first 36 hours; M2:*
 282 *crabs that moulted after 36 hours. NA: Not applicable. TA: Total ammonia (NH₃+NH₄⁺).*
 283

284 The results of the multiple linear regression analysis of the effects of the
 285 physical and chemical parameters on the time until either the shells fully hardened

286 (reached Co 6) or death are presented in Table 6. In Experiment 1, only pH had an
 287 influence (weak) on the results. In experiment 2, pH, ammonia and nitrite had
 288 moderate influences on the results.

289 Table 6. General results of multiple linear regression analysis of the influences of physical and
 290 chemical parameters on the time until shell hardening or death after ecdysis.

Experiment	Group	Correlated parameters	Cases (n)	p	Adjusted R ²	Correlation
1	A1	pH (max.)	68	0,000	0,196	Weak
	M1	pH (max.)	116	0,000	0,559	Moderate
2	B1	pH (max.)	136	0,000	0,410	Moderate
		AT (min.)		0,000		
	B2	AT (min.) NO ₂ ⁻ (min.)	203	0,000 0,001	0,657	Moderate

291 *Experiment 1 (collective treatment with filtration and partial daily water renovation). A1: crabs in pre-ecdysis.*
 292 *Experiment 2 (collective treatment with filtration but no water renewal). B1: crabs in pre-ecdysis; B2: tanks*
 293 *containing water previously used for the B1 group, with crabs also in pre-ecdysis; M1: crabs that moulted within*
 294 *36 hours; M2: crabs that moulted after 36 hours. TA: Total ammonia (NH₃+NH₄⁺).*

295 The duration at which the shell was at consistency 2 (i.e., the consistency with
 296 the highest market value) was significantly higher in the M2 animals than in the M1
 297 animals. Furthermore, none of the M2 individuals that moulted after 36 h achieved
 298 Co 6 (hard) shells, whereas in the M1 group, more than half of the individuals had
 299 shells that reached this consistency. In addition, 68% of individuals with shells that
 300 hardened remain alive. Among those that did not achieve shell hardening, the
 301 survival rate was only 15% (Table 7).

302 Table 7. Duration of *Callinectes ornatus* at shell consistencies 2 and 3 (Co 2 and Co 3) and associated
 303 hardening, ecdysis and mortality data in group B1 (organisms initially in pre-ecdysis) of Experiment 2
 304 (collective treatment with filtration but no water renewal). Different letters indicate a significant
 305 difference (p < 0.05) between the groups (within a column) according to the Kruskal-Wallis or Mann-
 306 Whitney test.
 307

Group	Co 2/3 (h) (min-max)	n	Hardening (%)		Ecdysis (n (%))	Dead (n (%))
M1	3 ^a (1-18)	32	Yes	(54%) ^a	19 (23%)	8 (42%) ^a
			No	(37%) ^a	13 (16%)	11 (85%) ^b
M2	61 ^b (3-129)	46	Yes	-	0	-
			No	(100%) ^b	46 (55%)	43 (93%) ^b

308 *Individuals who moulted before (M1) or after (M2) 36 hours.*

309 Table 8 shows the median, minimum and maximum values of the physical and
 310 chemical parameters of water quality that most influenced crab survival time and
 311 exoskeleton consistency: pH, ammonia and nitrite. The duration at each level of

312 consistency was highly related to the pH and concentration of total ammonia at the
 313 time of moulting. When the levels of the physical and chemical parameters favoured
 314 hardening, the time until the organisms reached Co 4 (soft paper) was short
 315 (between two and four hours). In contrast, when the pH was below 7.3 and the total
 316 ammonia concentration remained above 6 mg/L, the median time to Co 4 was 60 h.

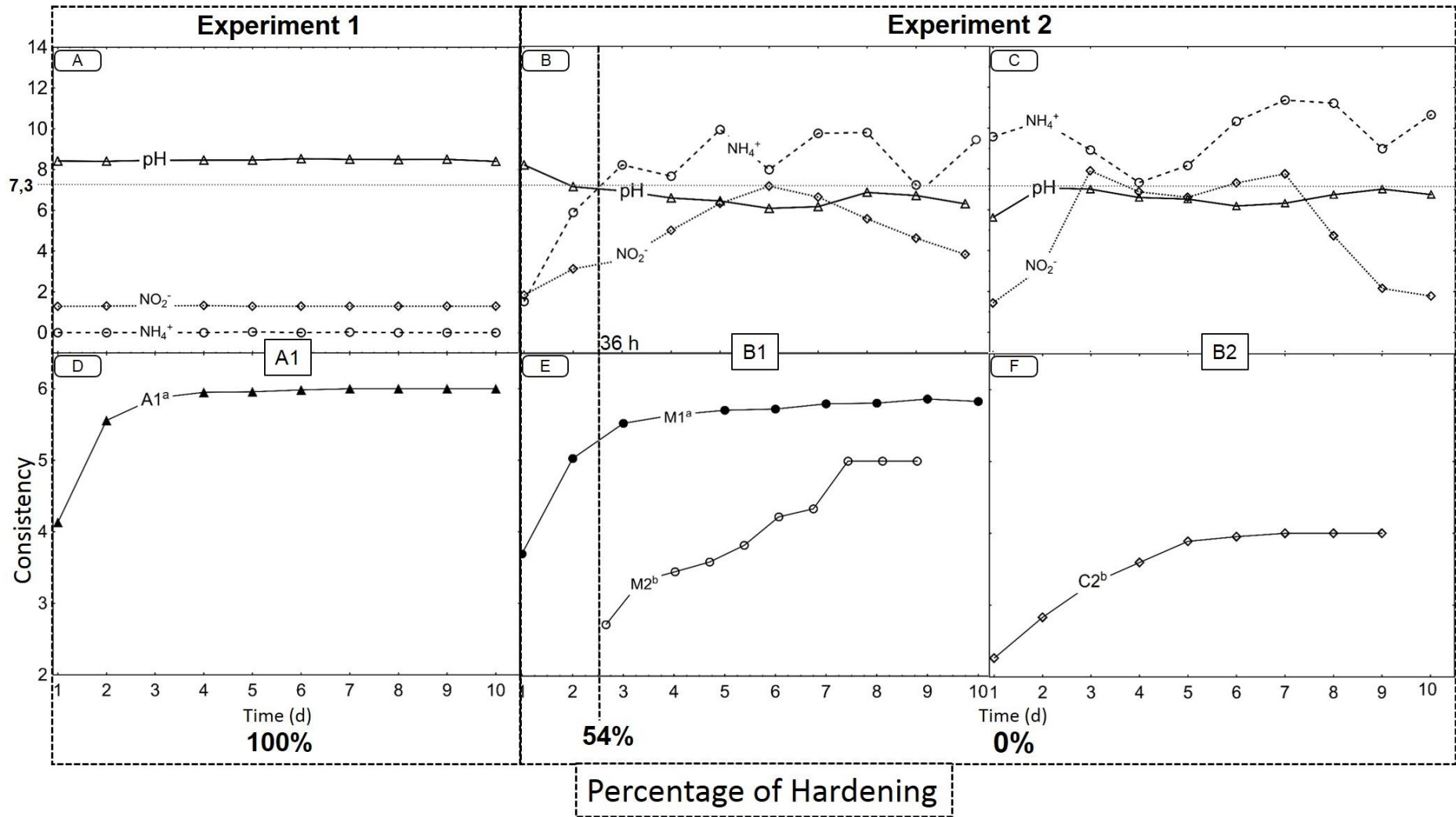
317 When water renewal was not performed (Experiment 2), the pH and total
 318 ammonia and nitrite concentrations varied significantly (Figure 2). As a result, there
 319 was an increase in the carapace hardening time and a decrease in the number of
 320 individuals reaching Co 6. The crabs of Experiment 1 (A1) and the animals that
 321 moulted within the first 36 hours of Experiment 2 (M1) spent significantly less time at
 322 Co 2 and Co 3 than did those that moulted after the first 36 hours (M2) in group B1
 323 and those in group B2.

324 Table 8. Water quality parameter measurements and crab survival data registered in the experimental
 325 units in which the crabs (*Callinectes ornatus*) moulted.

Exp.	Group	Co	Median (min-max)				Mortality (%)	Survival (h)		
			pH	TA (mg/L)	NO ₂ ⁻ (mg/L)	Time (h)		25 %	50%	75 %
1	A 1	1	8.4 ^{aA} (8.4-8.6)	0 ^{aA} (0-0.3)	1.31 ^{aA} (1.28-1.50)	61 ^{aAD} (2-245)	15			
		2-3	8.4 ^{abA} (8.4-8.5)	0 ^{aA} (0-0.3)	1.33 ^{abA} (1.29-1.33)	4 ^{bA} (1-9)	2			
		4-5	8.5 ^{abA} (8.3-8.6)	0 ^{aA} (0-0.3)	1.30 ^{abA} (1.28-1.50)	26 ^{cA} (3-137)	4	0	0	0
		6	8.5 ^{bA} (8.5-8.6)	0 ^{aA} (0-0.3)	1.3 ^{bA} (1.28-1.50)	155 ^{dA} (17-209)	2			
		1	8.4 ^{aA} (7.6-8.4)	0 ^{aA} (0-4.8)	1.7 ^{aB} (1.3-2.72)	30 ^{aC} (0-36)	9			
2	B 1	2-3	7.6 ^{bA} (6.7-8.4)	4.8 ^{bA} (0-9.8)	2.7 ^{bA} (1.3-3.22)	3 ^{bA} (1-18)	3	69	150	-
		4-5	7.0 ^{cD} (5.7-8.3)	6.1 ^{cD} (1.1-16.5)	3.2 ^{cC} (1.67-7.85)	42 ^{cC} (21-234)	31			
		6	6.5 ^{dB} (5.5-7.7)	8.2 ^{dB} (4.5-16.5)	5.6 ^{dB} (2.27-8.02)	183 ^{CA} (55-240)	20			
		1	7.1 ^{aB} (5.7-8.4)	6.0 ^{aB} (0-16.5)	2.8 ^{aC} (1.3-7.85)	60 ^{aBD} (36-192)	2			
		2-3	6.6 ^{bC} (5.5-7.5)	7.7 ^{bD} (3.9-16.5)	5.6 ^{bB} (2.27-8.02)	61 ^{aC} (3-129)	52	66	91	107
B 2	M2	4-5	6.5 ^{bC} (5.5-7.3)	9.1 ^{cC} (4.5-16.5)	5.6 ^{bD} (2.27-8.02)	45 ^{aAC} (3-222)	35			
		1	6.5 ^{aC} (5.3-8.1)	9.3 ^{aC} (3.3-14.7)	5.2 ^{aD} (1.35-9.40)	177 ^{aB} (0-186)	38	58	94	144
		2-3	6.5 ^{aC} (5.3-8.1)	10.1 ^{abC} (3.3-14.7)	6.3 ^{aBC} (1.35-9.40)	64 ^{bBC} (0-186)	20			

	(5.8-8.1)	(3.4-14.7)	(1.53-9.40)	(18-126)	
	6.5 ^{aC}	11.2 ^{bC}	4.2 ^{aBC}	80 ^{bABC}	20
4-5	(5.8-7.6)	(4.2-14.7)	(1.53-9.40)	(12-180)	

326 *Exp.: Experiment 1 (collective treatment with filtration and partial daily water renovation), Experiment 2 (collective*
327 *treatment with filtration but no water renewal). A1 and B1: water not used previously; B2: tanks containing water*
328 *used previously for the group B1; M1: crabs that moulted within the first 36 hours; M2: crabs that moulted after 36*
329 *hours. Co: consistency. TA: total ammonia. Time: Length of stay at a given consistency. Different letters indicate*
330 *significant differences (p < 0.05) according to the Kruskal-Wallis test. Lowercase letters indicate differences in*
331 *carapace consistency within the same group. Uppercase letters indicate differences in carapace consistency*
332 *among groups.*



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Figure 2. Median pH, total ammonia (mg/L), nitrite (NO_2^-) (mg/L) (A, B and C) and carapace consistency (2 - soft, 3- leather, 4 - soft paper, 5 - hard, paper 6 - hard) over time (in days) (D, E and F). Experiment 1: collective treatment with filtration and partial daily water renewal (66 crabs tested, 46 in pre-ecdysis and 20 in inter-ecdysis stage). Experiment 2: collective treatment with filtration but without water renewal (175 crabs tested, 123 in pre-ecdysis and 52 in inter-ecdysis stage). A1 (46 crabs in pre-ecdysis stage) and B1 (86 crabs in pre-ecdysis stage divided in three replicas): previously unused water and organisms in pre-ecdysis. B2 (40 crabs in

339 pre-ecdysis stage): organisms maintained in the reused water of group B1. M1: 32 crabs that moulted within the first 36 hours. M2:
340 46 crabs that moulted after 36 hours. Different letters indicate significant differences ($p < 0.05$) among groups according to the
341 Kruskal-Wallis test.

342 Discussion

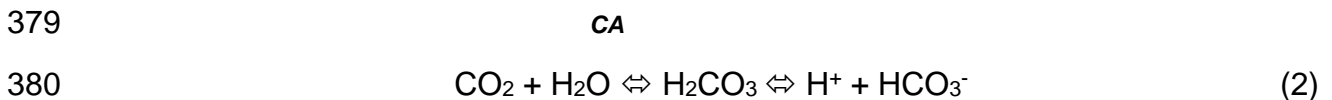
343 An issue repeatedly debated among those who investigate the shedding and
344 hardening process in crustaceans is the importance of calcium, the main constituent
345 element of the exoskeleton (Greenaway, 1985), in this process (Cameron, 1985;
346 Cameron and Wood, 1985; Clarke and Wheeler, 1922; Freeman and Perry, 1985;
347 Granado e Sá et al., 2010; Greenaway, 1983; Mangum et al., 1985; Middlemiss et
348 al., 2016; Neufeld and Cameron, 1992; Pan et al., 2006; Perry et al., 2001;
349 Robertson, 1960; Welinder, 1974; Wheatly et al., 2001; Wheatly, 1997; Wheatly,
350 1999; Wheatly et al., 2002; Zanotto and Wheatly, 2002). The lack of significant
351 correlations between the concentrations of Ca^{2+} and Mg^{2+} in water and either
352 carapace hardening or *C. ornatus* survival does not indicate that calcium is not
353 important in this process. On the contrary, it indicates that certain processes can
354 directly interfere with the physiology of the absorption and immobilisation of Ca_2^+ in
355 the exoskeleton and thereby significantly increase the time that these animals remain
356 soft after moult.

357 The organisms of Experiment 1 (subjected to daily water renewal) that
358 underwent ecdysis hardened rapidly, achieving paper consistency (Co 4) a median of
359 4 h after moult. This finding is consistent with studies conducted with *C. sapidus*
360 (Cameron and Wood, 1985; Freeman et al., 1987). Similar results were observed
361 among the crabs in Experiment 2 that moulted in water with a pH above 7.6 and a
362 total ammonia concentration below 4.8 mg/L, with Co 4 achieved after a median time
363 of 2 to 3 h. However, among the animals that began moulting in water with a pH
364 below 7.3 and a total ammonia concentration above 6 mg/L, up to 129 h (median of
365 more than 60 h) elapsed before either reaching Co 4 or death.

366 To understand this result, it is necessary to understand the chemical
367 processes involved in the calcification of the crab exoskeleton. In a closed system
368 with water recirculation, it is expected that over time there will be a reduction in the
369 concentration of free Ca_2^+ , due mainly to the immobilisation of Ca_2^+ in the form of
370 CaCO_3 during exoskeleton hardening (Perry et al., 2001). This immobilisation can be
371 represented by the following equation:



373 With the increased demands for Ca_2^+ and HCO_3^- , crabs begin to consume
374 both metabolic and external CO_2 . CO_2 reaches its highest internal concentrations at
375 moulting time (Mangum et al., 1985), increasing the availability of internal HCO_3^-
376 (Cameron and Wood, 1985). As soon as moulting occurs, the enzyme carbonic
377 anhydrase (CA), present mainly in the epithelium and the gills, is activated (Mangum
378 et al., 1985), accelerating the reaction:



381 As explained by Detours et al. (1968) and Zeebe and Wolf-Gladrow (2001),
382 the formed carbonic acid tends to be buffered by the carbonate-bicarbonate system.
383 This process results in an increase in the fraction of CO_3^- and acidification of the
384 medium (Greenaway, 1974; Mangum et al., 1985; Wheatly, 1997). However, over
385 time, the natural acid neutralisation capacity of the system becomes compromised,
386 and the medium tends to acidify as a result, increasingly compromising the crab's
387 capacity to deposit CaCO_3 in its exoskeleton. According to Cameron and Wood
388 (1985), the calcification process can be compromised if the pH outside the body is
389 less than 0.3 to 0.5 above the internal pH.

390 However, in addition to consuming HCO_3^- post-ecdysis, the organism excretes
391 H^+ or an equivalent ion such as NH_4^+ (Cameron, 1985; Middlemiss et al., 2016),
392 which is dissociated into NH_3 and H^+ . The rate of H^+/NH_4^+ excretion increases after
393 moulting (Cameron and Wood, 1985) and may increase further during bacterial
394 denitrification (Rijn et al., 2005). Under these conditions, the metabolism of excretion
395 also contributes to the acidification of the medium, further reducing the capacity for
396 calcium mobilisation by the crab, as observed in experiment 2. There is evidence that
397 water acidification is more critical for the hardening process of marine crustaceans
398 than for that of freshwater crustaceans. Unlike freshwater crustaceans, marine
399 crustaceans have almost no internal reserves of calcium (gastroliths) and depend
400 exclusively on the environment to supply the demand for Ca^{2+} (Greenaway, 1985;
401 Passano, 1960; Wheatly, 1997).

402 In a similar manner, acidification might affect the deposition of magnesium in
403 the crustacean exoskeleton. Although magnesium concentrations in water are
404 relatively lower than those of calcium, magnesium also plays an important role in the
405 hardening of the exoskeleton, and it is also obtained through water (Cameron and
406 Wood, 1985; Clarke and Wheeler, 1922; Welinder, 1974) in a process that might be
407 affected by pH (Tao et al., 2009).

408 In addition to Ca^{2+} and Mg^{2+} concentrations, the concentrations of Na^+ and K^+
409 were monitored in this study. These two ions directly participate in important
410 enzymatic activities that occur post-ecdysis (Towle and Mangum, 1985). Studies
411 have shown that if the relative proportions of these two ions are altered, ammonia
412 toxicity can occur due to the retention of ammonia by the organism and potentially
413 compromise the animal's survival (Pan et al., 2006; Romano and Zeng, 2011; Zanotto

414 and Wheatly, 1993). However, were observed no significant effects of these ions in
415 our experiments. It is possible that" the factors described above were much more
416 important in influencing exoskeleton hardening and the probability of survival in *C.*
417 *ornatus*.

418 There was also a direct relationship between the time to exoskeleton
419 hardening and the mortality rate. However, the mortality rate was only 25% among
420 those crabs that moulted after approximately 60 h. Those that did not moult died or
421 remained alive until the end of the experiment. In addition, in all of the groups except
422 those receiving periodic water renewal, there was an increase in mortality in the post-
423 ecdysis phase. In this case, the analyses again indicated the influences of pH and
424 total ammonia.

425 It is known that crabs (notably *C. sapidus*, the most studied species of the
426 genus *Callinectes*) can tolerate a pH range of 6.5 to 8.5 (Hochheimer, 1988).
427 Nevertheless, in artificial environments, it is recommended that pH be maintained
428 between 7.0 and 8.0 (Oesterling, 1995). It is also known that there are behavioural
429 and tolerance differences between young and adult animals in relation to pH
430 (Laughlin et al., 1978). In Experiment 2 of the present study, pH values of 5.5 and 5.3
431 were recorded in groups B1 and B2, respectively. In addition to having a direct effect
432 on the organisms, a reduction of pH causes an increase in the nitrous acid fraction
433 (HNO_2) present in water; HNO_2 is toxic to aquatic organisms (Ary and Poirrier, 1989;
434 Lin and Chen, 2003; Russo et al., 1981; Seneriches-Abiera et al., 2007).

435 The toxicity of ammonia, in turn, is directly proportional to pH and NH_3
436 concentrations. Romano and Zeng (2007) estimated an LC_{50} for juveniles of *Scylla*
437 *serrata* of 6.81 mg/L $\text{NH}_3\text{-N}$. Koo et al. (2005) reported that at least 50% of juveniles

438 of *Orithyia sinica* survived for 30 days at approximately 2.33 mg/L NH₃-N. Lakshmi
439 (1984) reported a mortality rate of 20% in *C. sapidus* in pre-ecdysis at 1.41 mg/L
440 NH₃, which increased to 100% at 2.31 mg/L NH₃. In our experiments, a pH reduction
441 was observed over time, which indicated that the NH₃ concentrations remained
442 sufficiently low as to rule out any toxic effects of ammonia on *C. ornatus*.

443 Regarding nitrite, there is no consensus regarding the concentrations at which
444 this compound is toxic to crabs. Lakshmi (1984) and Ary and Poirrier (1989) reported
445 that the survival of *C. sapidus* was only affected at NO₂⁻ concentrations above 10
446 mg/L. According to those authors, crab mortality reached 100% only after 96 h of
447 exposure to concentrations between 50 and 150 mg/L in water with a pH close to 8.
448 In contrast, Manthe et al. (1984) found that the moulting efficiency of *C. sapidus* was
449 affected by nitrite concentrations close to 2 mg/L. In the present study, the nitrite
450 concentration reached 7.6 mg/L. Thus, it is possible that the observed mortality might
451 have been influenced by both pH and nitrite levels during the experiments and that
452 they had a cumulative effect. Moreover, a long hardening time, which exposed the
453 animals to unfavourable physiological conditions, appears to have significantly
454 increased the risk of death.

455 **Conclusion**

456 Moulting in *C. ornatus* exhibited strong relationships with the characteristics of
457 the crab's aquatic medium. The crabs drastically altered the physical and chemical
458 characteristics of the water, mainly through processes related to acidification and
459 ammonification. These alterations, in turn, directly interfered with exoskeleton
460 hardening, causing the exoskeletons of the animals to remain at soft or paper
461 consistency for periods of up to 5 days. Commercially, the establishment of such

462 periods would allow crabs to be marketed as soft-shell crabs within a time window
463 more than 20 times longer than that typically observed. If the results observed here
464 can be replicated at the commercial scale, large reductions in workload and
465 operational costs could be obtained, increasing the efficiency and viability of large-
466 scale crab production.

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475 **Competing interests**

476 No competing interests declared.

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