

1 **Orb-web spider color polymorphism through the eyes of**
2 **multiple prey and predators**

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28 **Summary statement:** We assessed the role of both prey and predator vision on the
29 evolution and maintenance of color polymorphism and color conspicuousness of the orb-
30 web spider *Gasteracantha cancriformis* (Araneidae).

31

32 **ABSTRACT**

33 The sensory drive theory predicts that signals, sensory systems, and signaling behavior
34 should coevolve. Variation in the sensory systems of prey and predators may explain the
35 diversity of color signals, such as color polymorphism. The spider *Gasteracantha*
36 *cancriformis* (Araneidae) possesses several conspicuous color morphs. The aim of the
37 present study was to assess whether the color polymorphism of *G. cancriformis* may be
38 maintained by pressure from multiple signal receivers, such as prey and predators with
39 distinct color vision systems. In orb-web spiders, the prey attraction hypothesis states that
40 conspicuous colors are prey lures that increase spider foraging success via flower mimicry.
41 However, in highly defended species, conspicuous colors could also be a warning signal
42 to predators. We used color vision modelling to estimate chromatic and achromatic
43 contrast of *G. cancriformis* morphs as perceived by potential prey and predator taxa. Our
44 results revealed that individual prey and predator taxa perceive the conspicuousness of
45 morphs differently. For instance, the red morph is perceived as quite conspicuous to
46 lepidopteran prey and avian predators, but not by dipteran prey and hymenopteran prey
47 and predators. Therefore, the multiple prey and multiple predator hypotheses may
48 explain the evolution of color polymorphism in *G. cancriformis*. However, the spider's
49 coloration did not resemble flower coloration, which suggests that the species'
50 conspicuousness is not the result of flower mimicry. Other parameters that are not
51 evaluated by color vision models, such as distance, shape, angle, pattern geometry, and
52 contour, could also affect the perception of color morphs by both prey and predators and
53 thereby influence morph survival.

54

55 **INTRODUCTION**

56 The evolution and maintenance of color polymorphism have traditionally been attributed
57 to apostatic selection (Clarke, 1979). Assuming that predators form a search image
58 (Tinbergen, 1960), the advantage of rarity promotes the coexistence of multiple prey types

59 and stabilizes polymorphisms (Bond 2007). Nonetheless, other adaptive and non-adaptive
60 explanations for the evolution and maintenance of color polymorphisms have been
61 proposed (Gray and McKinnon, 2007). For instance, gene flow between populations with
62 distinct phenotypes that are favored by natural selection could maintain polymorphism
63 within populations (Farkas et al., 2013; Gray and McKinnon, 2007).

64 In the context of visual signaling, the distinct visual systems of prey and predators
65 may play a role in the evolution and maintenance of color polymorphisms (Ruxton et al.,
66 2004; White and Kemp, 2015). Animal communication involves the generation, emission
67 and transmission of a signal, and processing of the signal by a receiver, in which an
68 appropriate response is elicited (Endler 1993). Any factors that affect these steps can
69 influence signal efficiency and, as a result, affect the direction of communication evolution
70 (Endler 1993). Thus, the diversity of signals can be attributed to variation in the sensory
71 systems of receivers. Considering that the same “color” may be perceived as cryptic or
72 conspicuous by different species (Endler and Mappes 2004), each color morph of
73 polymorphic populations may represent an adaptation to particular visual systems of prey
74 or predator species (Endler, 1992; Ruxton et al., 2004; White and Kemp, 2015).

75 Many orb-web spiders exhibit conspicuous coloration. The prey attraction
76 hypothesis states that the bright coloration of some spiders lures insects, possibly by
77 mimicking flower coloration (e.g. Craig and Ebert, 1994; Hauber, 2002). The hypothesis
78 has been empirically tested several times, and most studies have found support for it. The
79 polymorphic *Nephila pilipes* (Nephilidae) present a melanic and a bright colored morph
80 (Tso et al., 2004). The bright color patterns of this species are thought to resemble
81 symmetric flower patterns that may attract bees, owing to the innate preference of bees
82 for symmetry (Chiao et al., 2009). Moreover, yellow patches on the spider’s body may be
83 perceived as food resources by flowers visitors (Tso et al. 2004). Besides being attractive
84 to pollinators, the yellow patches on the species’ body also seems to attract hymenopteran
85 predators. Therefore, it is possible that there is a trade-off between foraging success and
86 predation risk in polymorphic populations in which some morphs are more cryptic than
87 others (Fan et al. 2009).

88 The predators of orb-web spiders possess very distinct visual systems. Birds, for
89 example, are tetrachromats, whose photoreceptors are most sensitive to ultraviolet-violet,

90 blue, green, and red (Hart 2001), whereas spider hunting wasps, such as members of the
91 Sphecidae, are trichromats, whose photoreceptors are most sensitive to ultraviolet, blue,
92 and green (Peitsch, 1992; Briscoe and Chittka, 2001). Similarly, the insect prey of orb-web
93 spiders also vary in their types of color vision. For example, bees are trichromats with
94 spectral sensitivities that are similar to those of sphecid wasps (Briscoe and Chittka, 2001),
95 whereas some lepidopterans are tetrachromats, and some dipterans possess
96 photoreceptors with five different sensitivity peaks (Schnaitmann et al., 2013). Therefore,
97 the maintenance of spider color polymorphism may result not only from a trade-off
98 between prey attraction and capture success but also from selective pressure from multiple
99 receivers (Endler, 1992; Ruxton et al., 2004; White and Kemp, 2015). Color perception
100 depends on both the signal reflectance and observer visual system, as well as on the
101 background reflectance spectrum and ambient light intensity (Endler 1990). Physiological
102 models of color vision include all these factors and have been effective for objectively
103 studying animal coloration (i.e., independent of human subjective assessment; Renoult,
104 2015).

105 The orb-web spider *G. cancriformis* constructs large webs and rests in the web
106 hub during the day (Levi, 1978). Females of the species possess a hard abdomen with
107 three pairs of spines and vary in color, with some morphs quite conspicuous to human
108 observers (Levi, 1978; Gawryszewski and Motta, 2012). The ventral side of females are
109 mostly black, sometimes with small bright spots. In one studied population, the dorsal
110 side of females possessed black or reddish spines and four different color patterns: yellow,
111 white, red, and a combination of black and white (Gawryszewski 2007; Gawryszewski and
112 Motta, 2012). Adult females measure from 5 to 7 mm in length and 10 to 13 mm in width
113 (Muma, 1971), whereas the males are brownish, small, and do not exhibit chromatic
114 variation (Levi, 1978). The prey attraction hypothesis does not seem to explain the
115 coloration of the orb-web spider *Gasteracantha cancriformis* (Araneidae), since both
116 naturally bright morphs and yellow-painted individuals failed to capture more prey than
117 either naturally cryptic morphs or black-painted individuals (Gawryszewski and Motta,
118 2012). Although evidence is still needed, Edmunds and Edmunds (1983) suggested that
119 the conspicuous body coloration of *Gasteracantha* spiders might serve as a warning signal
120 to predators. The aim of the present study was to investigate three hypotheses for the

121 evolution and maintenance of color polymorphism, using *G. cancriformis* as a model.
122 Two derivations from the prey attraction hypothesis include (1) the *multiple prey*
123 *hypothesis*, which posits that color morphs are perceived differently by different prey taxa
124 and that each color morph is adapted to lure a specific type of prey, and (2) the *multiple*
125 *mimic model* hypothesis, which posits that the spiders attract prey *via* aggressive mimicry
126 of flower colors and that each color morph mimics a different flower color. In addition,
127 (3) the *multiple predator hypothesis* posits that the conspicuous colors found in spiders
128 could serve as warning signals to predators and that color polymorphism could evolve
129 and be maintained if each color morph is adapted to the vision of a specific predator.

130

131 MATERIALS AND METHODS

132 Color vision model

133 To estimate the perception of *G. cancriformis* color morphs by distinct predators and
134 prey groups, we used the color vision model proposed by Chittka (1992). Although this
135 model has been only validated with behavioral experiments on honeybees, its general
136 form allow us to apply it for other taxa (e.g. They and Casas 2002). The model requires
137 four inputs: (1) the irradiance reaching the observed object, (2) the observer
138 photoreceptor excitation curves, (3) the background reflectance to which photoreceptors
139 are adapted to, and (4) the reflectance curve of the observed object. First, the sensitivity
140 factor R was determined for each photoreceptor, as follows:

$$R = 1 / \int_{700}^{300} I_B(\lambda) S(\lambda) D(\lambda) d\lambda \quad (1)$$

141 where $I_B(\lambda)$ is the spectral reflectance function of the background, $S(\lambda)$ is the spectral
142 sensitivity function of each photoreceptor, and $D(\lambda)$ is the illuminant irradiance spectrum.
143 Secondly, the quantum flux P (relative amount of photon catch) is calculated, as follows:

$$P = R \int_{700}^{300} I_S(\lambda) S(\lambda) D(\lambda) d\lambda \quad (2)$$

144 where $I_S(\lambda)$ is the spectral reflectance function of the stimulus. Assuming that the maximum
145 excitation of a photoreceptor is 1, the phototransduction process is determined by:

$$E = P / (P + 1) \quad (3)$$

146 Stimuli spectra are projected in specific color spaces. The coordinates of each
147 spectrum are calculated using photoreceptor excitations, as follows (Chittka et al. 1994):

$$X1 = \sin 60^\circ (E_3 - E_1) \quad (4)$$

$$X2 = E_2 - 0.5 (E_1 + E_3) \quad (5)$$

148 For tetrachromat organisms (Théry and Casas, 2002):

$$X1 = \frac{2\sqrt{2}}{3} \cos 30^\circ (E_3 - E_4) \quad (6)$$

$$X2 = E_1 - \frac{1}{3} (E_2 + E_3 + E_4) \quad (7)$$

$$X3 = \frac{2\sqrt{2}}{3} [\sin 30^\circ (E_3 + E_4) - E_2] \quad (8)$$

149 We extended the model of Chittka (1992) to accommodate pentachromatic organisms, as
150 follows:

$$X1 = \frac{5}{2\sqrt{2}\sqrt{5}} (E_2 - E_1) \quad (9)$$

$$X2 = \frac{5\sqrt{2}}{2\sqrt{3}\sqrt{5}} [E_3 - \left(\frac{E_1 + E_2}{2}\right)] \quad (10)$$

$$X3 = \frac{5\sqrt{3}}{4\sqrt{5}} [E_4 - \left(\frac{E_1 + E_2 + E_3}{3}\right)] \quad (11)$$

$$X4 = E_4 - \left(\frac{E_1 + E_2 + E_3 + E_4}{4}\right) \quad (12)$$

151 Chromatic contrast between a color stimulus and background, or between two
152 color stimuli, is calculated as the Euclidean distance (ΔS) between two points in color
153 space, as follows:

$$S = \sqrt{\sum_{i=1}^n (X_{a_i} - X_{b_i})^2} \quad (13)$$

154 where X_i ($i = 1, 2, 3, \dots, n$) represents the coordinate in the color space.

155 The color spaces are chromaticity diagrams and, thus, do not estimate the
156 achromatic contrast between objects. Nonetheless, achromatic contrasts can be important

157 visual cues used by both prey and predators. In bees, achromatic contrast is more
158 important than chromatic cues for objects that subtend a visual angle smaller than $\sim 15^\circ$,
159 which means that bees have to be very close to flowers in order to use their color vision
160 for discrimination tasks (Giurfa et al., 1997). Similarly, birds use achromatic contrast in
161 detection of small objects (Osorio et al., 1999). We estimated the achromatic contrast as
162 the excitation (Eq. 3) of the photoreceptor responsible for achromatic discrimination in
163 each organism (Chittka and Kevan 2005).

164 For our modeling, we used the reflectance data of *G. cancriformis* color morphs
165 that was collected during a previous study (for reflectance curves see figure 1.8 in
166 Gawryszewski, 2007, and figure 5 in Gawryszewski and Motta 2012). These data have
167 already been used to estimate the visual contrast of the yellow and white morphs and the
168 black and white morphs from the perspective of *Apis mellifera* (Gawryszewski and Motta
169 2012). The spiders were collected from a Brazilian savanna physiognomy, namely
170 Cerrado *sensu stricto*, which is characterized by shrubs and trees of 3 to 8 m tall that are
171 contorted and possess thick, fire-resistant bark, a crown cover of $>30\%$, and additional
172 herbaceous vegetation (Oliveira-Filho and Ratter 2002). The background reflectance was
173 estimated from the average reflectance of leaves, leaf litter, bark, and grasses that were
174 collected from the same area as the spiders (see figure 5 in Gawryszewski and Motta,
175 2012). To avoid issues with negative values and unrealistic positive values we adjusted the
176 reflectance data by subtracting the reflectance values by the minimum value of each
177 measurement. As illuminant spectrum, we used the International Commission on
178 Illumination (CIE) standard illuminant of D65, which is comparable to open areas, such
179 as the Brazilian savanna (Chittka, 1996).

180

181 **Multiple prey hypothesis**

182 Using the model described above, we estimated the chromatic and achromatic
183 conspicuousness of the *G. cancriformis* morphs (yellow, white, red, and white parts of the
184 black and white morph) to a suit of potential prey: *A. mellifera* (Hymenoptera, Apidae),
185 *Drosophila melanogaster* (Diptera, Drosophilidae), and *Fabriciana adippe* (Lepidoptera,
186 Nymphalidae). We selected these insect orders because they are commonly intercepted
187 by orb-webs in field experiments (Craig and Ebert 1994; Tso et al. 2002) and represent

188 the diversity of visual systems among insects (Briscoe and Chittka, 2001). For *A. mellifera*
189 and *D. melanogaster*, we used photoreceptor sensitivity curves from the literature (Peitsch
190 et al., 1992; Schnaitmann et al., 2013). It was recently shown that, together with R7-R8
191 photoreceptors, R1-R6 photoreceptors contribute to color vision in *D. melanogaster*
192 (Kelber and Henze, 2013; Schnaitmann et al., 2013). Therefore, we included the R1-R6
193 photoreceptor curve, treating *D. melanogaster* as a pentachromat. The graphical curves
194 were extracted directly from the figures of relevant publications using DataThief III
195 version 1.7 (Tummers, 2006). For *Fabriciana adippe*, however, no photoreceptor
196 sensitivity curves are available, so electrophysiological measurements of photoreceptor
197 sensitivity peaks (λ_{\max} ; Eguchi et al., 1982) were used to estimate the photoreceptor curves
198 (for details see Govardovskii et al., 2000).

199 For achromatic contrast, bees only use the green photoreceptor (Giurfa et al.,
200 1996), whereas flies only use the outer photoreceptors (R1-R6; Kelber & Henze, 2013).
201 Because the exact mechanism used by lepidopterans for achromatic discrimination is
202 incompletely understood, we assumed that they employ the same mechanism as in bees.
203 The color hexagon model assumes that photoreceptors respond to half their maximum
204 for the background they are adapted to, so that the photoreceptor excitation for
205 background is equivalent to 0.5 units (Chittka, 1992).

206 The multiple prey hypothesis predicts that different prey taxa perceive color
207 morphs differently. To assess whether each spider morph was perceived differently by
208 prey species, we constructed two linear mixed models, one for chromatic contrast and
209 one for achromatic contrast. Either chromatic (ΔS) or achromatic contrast were used as
210 the dependent variable, and spider morph and prey taxon were used as the independent
211 variables (contrast = spider morph \times observer). The spider morph was defined as yellow,
212 white, red, or black and white, and the observers were defined as hymenopteran,
213 dipteran, or lepidopteran. Individual spiders were used as random effects. Normality and
214 homogeneity were verified by visual inspection of quantile-quantile and residuals vs. fitted
215 values plots. We computed all nested models and used the Akaike Information Criterion
216 to select the best model.

217 As a reference point, we used a color discrimination threshold of $\Delta S = 0.11$, which
218 is the threshold value below which trained bees are unable to distinguish different flower

219 colors (Chittka, 1996). However, discrimination thresholds are variable and can change
220 depending on the study species, learning conditions, previous experience, background
221 coloration, whether the task involves discrimination between colors or detection against
222 the background, and whether objects are compared sequentially or simultaneously
223 (Avarguès-Weber & Giurfa, 2014). It should also be noted that threshold values were not
224 behaviorally validated for other taxa. Therefore, values originated here using the Chittka
225 model (1992) must be taken carefully.

226

227 **Multiple mimic models hypothesis**

228 To test the multiple mimic models hypothesis, we compared how flowers and spider
229 morphs are perceived by prey. We gathered all flower reflectance data available in the
230 Floral Reflectance Database (FReD; Arnold et al., 2010), excluding reflectance data from
231 lower flower parts, leaves, bracts, stamens, the inner parts of bell-shaped flowers, and
232 unknown items, as well as spectrum files that did not cover 300 to 700 nm. Most species
233 in the database have only one reflectance spectrum, and for species with multiple
234 reflectance spectra, we randomly selected a single spectra. We did not average the
235 reflectance of these species because there was no information available on whether these
236 measurements referred to different individuals or different parts of single flowers. In total,
237 we gathered reflectance data from 859 plant species.

238 We calculated a matrix of chromaticity distances between average color loci of
239 each spider color morph and each individual flowers species and plotted these values
240 using a Kernel density estimation. If spiders are mimicking some group of flower colors,
241 we would expect to find at least one peak near or below the theoretical detection threshold
242 of 0.11. We also followed the same procedure for the absolute achromatic difference
243 between individual flowers and the mean achromatic value for each color morph. If
244 spiders and flowers are similar in the achromatic dimension, we would expect to find one
245 peak near zero.

246

247 **Multiple predator hypothesis**

248 The methodology used to investigate the multiple predator hypothesis methodology was
249 very similar to that used for the multiple prey hypothesis, except that we used predator

250 species in our models. As predators, we considered the bird *Parus caeruleus* (Paridae)
251 and the wasp *Philanthus triangulum* (Sphecidae), since birds and wasps are the main
252 predators of orb-web spiders (Rayor, 1996; Foelix, 2010), are visually guided hunters, and
253 have distinct color vision systems. For *P. caeruleus*, we used photoreceptor sensitivity
254 curves available in the literature (Hart, 2001), and for *P. triangulum*, we used
255 photoreceptor sensitivity peaks to estimate photoreceptor sensitivity curves (data available
256 in Briscoe and Chittka, 2001; see Govardovskii et al. 2000 for estimation of sensitivity
257 curves from sensitivity peaks).

258 The multiple predator hypothesis predicts that different predator taxa perceive
259 color morphs differently. To assess this prediction, we established two linear mixed
260 models, one for chromatic contrast and one for achromatic contrast. Either chromatic (Δ
261 S) or achromatic contrast were used as the dependent variable, and spider morph and
262 predator taxon were used as the independent variables (contrast = spider morph \times
263 observer). The spider morph was defined as yellow, white, red, or black and white, and
264 individual spiders were used as random effects. Normality and homogeneity were verified
265 by visual inspection of quantile-quantile and residuals vs. fitted values plots. We computed
266 all nested models and used the Akaike Information Criterion to select the best model.

267 As in the multiple prey hypothesis, we used discrimination thresholds as reference
268 points. For the chromatic contrast, we considered color discrimination thresholds of $\Delta S =$
269 0.11 and $\Delta S = 0.06$ for the wasp (Dyer and Chittka, 2004) and bird (Théry et al., 2005),
270 respectively. For the achromatic contrast, we considered double cones in birds (Hart,
271 2001), and assumed green photoreceptors for wasps, as in bees.

272 Visual modeling calculations were conducted using the ‘colourvision’ R package
273 (Gawryszewski, 2017). Linear mixed models were performed using the ‘nlme’ package
274 (Pinheiro et al., 2016), and graphs were plotted using the ‘ggplot2’, ‘ggExtra’, and
275 ‘gridExtra’ packages (Wickham, 2009; Attali, 2016; Auguie, 2016; R Core Team, 2015).

276

277 **RESULTS**

278 **Multiple prey hypothesis**

279 For chromatic contrast, the model that included the interaction between spider morph
280 and prey taxon presented the lowest AIC value (Table 1). The yellow morph presented

281 the highest ΔS value for *A. mellifera* and *F. adippe* vision, whereas the white spider
282 presented the highest ΔS value for *D. melanogaster*, followed by the yellow morph (Fig.
283 1). The white patch of the black and white spiders presented a ΔS value that was very
284 close to the theoretical discrimination threshold for all prey species (Fig. 1). The red
285 spiders presented ΔS values near the theoretical discrimination threshold for *A. mellifera*
286 and *D. melanogaster*, but not for *F. adippe* (Fig. 1). For prey achromatic contrast, the
287 model that included the interaction between variables presented the lowest AIC value
288 (Table 1). For all prey groups, the white morph had the highest excitation value, followed
289 by the black and white, yellow, and red morphs, respectively (Fig. 1). The model
290 coefficients are provided in the supplementary material (Table S1 and S2).

291

292 **Multiple mimic models hypothesis**

293 For all three prey species, only the white patch of the black and white morph had a density
294 peak near the chromatic theoretical discrimination threshold of 0.11 when compared to
295 all flower reflectance spectra: 44.5% for *A. mellifera*, 16.8% for *D. melanogaster*, and 35%
296 for *F. addipe*. (Fig. 2). For the other spider morphs only a small proportion of the
297 Euclidean distances between flowers and morphs presented values < 0.11 . For *A. mellifera*
298 only 1.6% of yellow morphs presented values lower than 0.11, 3.4% of white morphs, and
299 4.8% of red morphs (Fig. 2). For *D. melanogaster* only 2.4% of yellow morphs had values
300 lower than 0.11, 4.0 % of white spiders, and 3.0% of red morphs (Fig. 2). For *F. addipe* this
301 values were 0.4%, 0.2%, and 0.5% respectively (Fig. 2). For all three prey species a large
302 proportion of flowers had similar achromatic values to the yellow morph, white morph,
303 and the white patch of the black and white morph, but not to the red morph (Fig. 3).

304

305 **Multiple predator hypothesis**

306 For the chromatic contrast, the model with interaction between variables presented the
307 lowest AIC value (Table 1). The black and white morph presented the lowest ΔS value
308 for both predators (Fig. 4A,B; Table S3). The yellow morph was the one with highest ΔS
309 value for *P. caeruleus*, though white and red morphs presented similar values (Fig. 4A).
310 For *P. triangulum*, the white spider morph presented the highest ΔS value, followed by
311 the yellow and red morphs, and the red morph was near the theoretical discrimination

312 threshold of 0.11 (Fig. 4B). For the achromatic contrast, the model that included the
313 interaction between variables presented the lowest AIC value (Table 1), even though the
314 values of the two predator species were very similar. For *P. caeruleus*, the white morph
315 had the highest excitation value, followed by the yellow, black and white, and red morphs,
316 respectively (Fig. 4C). The white morph also had the highest excitation value for *P.*
317 *triangulum*, followed by the black and white, yellow, and red morphs, respectively (Fig.
318 4D). The model coefficients are provided in the supplementary material (Table S3 and
319 S4).

320

321 DISCUSSION

322 Our statistical analyses suggested that the multiple prey and multiple predator hypotheses
323 are plausible explanations for the evolution of color polymorphism in *G. cancriformis*,
324 but provided poor support to the flower mimicry hypothesis. If conspicuous colors in orb-
325 web spider attracts prey, then polymorphism may occur in luring systems by multiple
326 mimic models that act as sensory traps for particular prey taxa. Conversely, polymorphism
327 may occur as a result of multiple receiver biases, where each morph lures specific prey
328 taxa that possess specific sensory, neural, or perceptual biases (White and Kemp, 2015).
329 In *G. cancriformis*, spider morphs conspicuousness is perceived differently by prey
330 species. The yellow and white morphs were the most conspicuous to all prey taxa. The
331 former being more contrasting from the background for honeybee color vision, and the
332 latter, for flies. Whereas, the red morph, although inconspicuous for honeybee and flies,
333 showed high detectability for butterflies. Yellow spider morphs could experience higher
334 foraging success as a result of insects preference for yellow signals. Yellow pollen and
335 yellow flower centers, both of which are likely adapted to insect visual systems, are among
336 the most common traits of floral color patterns (Papiorek et al., 2016; Lunau, 2000). On
337 the other hand, red morphs are conspicuous to lepidopterans and may be effective for
338 attracting them. Another recent study also using color vision models showed that insects
339 prey perceive coloration of *Verrucosa arenata* morphs differently, and they may be a
340 selective pressure on the evolution of these color patterns variation (Ajuria-Ibarra et al.
341 2017). Therefore, the distinct perceptions of insect taxa support the multiple prey

342 hypothesis as a mechanism for the evolution of color polymorphism in stationary
343 predators.

344 For the achromatic dimension, the statistical analyses also suggested an interaction
345 between spider morph and prey taxon. However, there was very little difference between
346 the morphs when viewed by different prey. Therefore, the differences in achromatic
347 contrast between prey perception may be statistically significant but not biologically
348 relevant. A recently proposed scenario for the evolution of color polymorphism is that
349 different morphs exploit different visual channels in prey. The yellow morphs of *G.*
350 *fornicata* would benefit from stimulating the dipteran chromatic channel, whereas white
351 morphs would benefit from stimulating the achromatic channel (White and Kemp, 2016).
352 However, this is inconsistent with *G. cancriformis* because the spider morphs presented
353 similar levels of achromatic detectability by all the prey taxa examined.

354 Several authors have proposed flower mimicry as a mechanism of prey attraction
355 by orb-web spiders *via* conspicuous body coloration (e.g. Craig and Ebert, 1994; Hauber,
356 2002). However, the hypothesis has seldom been tested. Our density graphs show that the
357 color of most of the *G. cancriformis* morphs was not similar to any group of flowers, which
358 suggests that the coloration of the *G. cancriformis* did not result from flower mimicry.
359 Conversely, a study of various orb-web spider species that also used color vision models
360 found that, as perceived by dipterans and hymenopterans, the colors of spiders are very
361 similar to those of flowers (White et al., 2016). It is tempting to use correlational data to
362 infer causal relationships, however, this finding may only reflect the diversity of flower
363 colors and spider colors. Color vision modelling of the prey of the orchid mantis
364 (*Hymenopus coronatus*) suggested that pollinators are unable to distinguish the colors of
365 the mantis and flowers, and a field experiment showed that the mantis actually attracts
366 more pollinators than flowers (O'Hanlon et al., 2013). However, the modification of non-
367 color mantis traits suggested that other types of flower mimicry, such as symmetry,
368 contrasting color patterns, and morphological similarities to flower petals, had no effect
369 on prey attraction (O'Hanlon, 2014). Therefore, mantis coloration is effective in attracting
370 prey, but may not result from flower mimicry. Instead, it may result from sensory
371 exploitation (O'Hanlon, 2014).

372 The results of the present study also support the multiple predator hypothesis for
373 the maintenance of color polymorphism in *G. cancriformis*, as the spider morphs
374 conspicuousness was perceived differently by predator species in both the chromatic and
375 achromatic dimensions. The red morph could be targeting bird predators but would
376 appear relatively inconspicuous to a hymenopteran predator. In contrast, the white and
377 yellow morphs are highly detectable by both predator taxa. The colors of two of the four
378 *G. cancriformis* morphs (yellow and red) are typical of aposematic species (Endler and
379 Mappes, 2004). Conspicuous coloration is especially advantageous when it increases the
380 mismatch with the background and facilitates predator learning (Endler and Greenwood,
381 1988). Spiders of the genus *Gasteracantha* possess spines and a hard abdomen. Moreover,
382 the hunting wasp *Sceliphron laetum* avoids provisioning initial instars with *Gasteracantha*
383 spiders (Elgar and Jebb, 1999). Morphological and behavioral defenses that make
384 ingestion difficult may also constitute an aposematic signal, along with the species' bright
385 colors, which enhance predator memory and learning (Endler and Greenwood, 1988;
386 Ruxton et al., 2004). Though aposematism is not commonly reported in spiders (Oxford
387 and Gillespie, 1998), Brandley et al. (2016) conducted an experiment with black widow
388 models and found that models with red markings were more likely to be avoided by birds
389 than all black models. If spider morphs are conspicuous to prey but do not resemble
390 flowers, it is possible that the conspicuousness of *G. cancriformis* reflects a trade-off
391 between an antipredatory strategy and foraging success, in which common spiders
392 morphs would be avoided by prey.

393 Thermoregulation is another possible explanation for color polymorphism in orb-
394 web spiders. In *Verrucosa arenata* (Araneidae), yellow and white morphs exhibited
395 different thermal responses when exposed to artificial heat. The white morphs absorbed
396 less visible radiation and shorter wavelengths than the yellow morphs, which could protect
397 the spiders' internal tissue from the harmful effects of UV radiation (Rao and Mendoza-
398 Cuenca, 2016). However, absorbing pigments are likely to offer protection against UV
399 damage (Gawryszewski et al., 2015), which is analogous to the melanin found in the
400 human epidermis (Brenner and Hearing, 2008) and commercial sunscreen lotions
401 (Antoniou et al., 2008).

402 We only considered chromatic and achromatic discrimination, but color pattern
403 geometry, shape, contour, size, angle, texture, and distance of visual detection
404 (Troscianko et al., 2009) may also influence the behavior of both prey and predators
405 toward spiders since different species use distinct visual cues for stimuli detection and
406 recognition (Théry and Gomez, 2010). Furthermore, color vision models do not include
407 other perceptual mechanisms, such as cognition, color categorization, past experiences,
408 or memory imprecision (Renoult et al., 2015), even though these factors may affect
409 detectability and, consequently, influence the survival rate of morphs in different ways
410 (Théry and Gomez, 2010). Additionally, non-adaptive explanations, such as
411 overdominance and allele equilibrium in absence of selection, are often ignored when
412 studying polymorphisms in an ecological perspective. Finally, predation experiments and
413 field experiments that evaluate the prey taxa caught by the different spider morphs are
414 paramount to validate and complement the findings of the present study.

415

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419

420 **Competing interests**

421 No competing interests declared.

422

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426

427 **References**

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601

602 Tables

603

604 Table 1. Delta Akaike Information Criterion (AIC) of Linear Mixed Models of the
605 chromatic and achromatic contrasts of prey and predators.

Model	df	Δ AIC
<i>Multiple prey hypothesis</i>		
Chromatic dimension		
$\Delta S \sim \text{morph} * \text{observer}$	17	0.0
$\Delta S \sim \text{morph} + \text{observer}$	11	23.9
$\Delta S \sim \text{observer}$	8	52.4
$\Delta S \sim \text{morph}$	9	61.5
$\Delta S \sim 1$	6	90.6
Achromatic dimension		
excitation $\sim \text{morph} * \text{observer}$	17	0.0
excitation $\sim \text{morph} + \text{observer}$	11	57.6
excitation $\sim \text{morph}$	9	72.2

excitation ~ observer	8	84.7
excitation ~ 1	6	100.2

Multiple predator hypothesis

Chromatic dimension

$\Delta S \sim \text{morph*observer}$	13	0.0
$\Delta S \sim \text{morph+observer}$	10	6.9
$\Delta S \sim \text{observer}$	7	30.6
$\Delta S \sim \text{morph}$	9	54.5
$\Delta S \sim 1$	6	74.9

Achromatic dimension

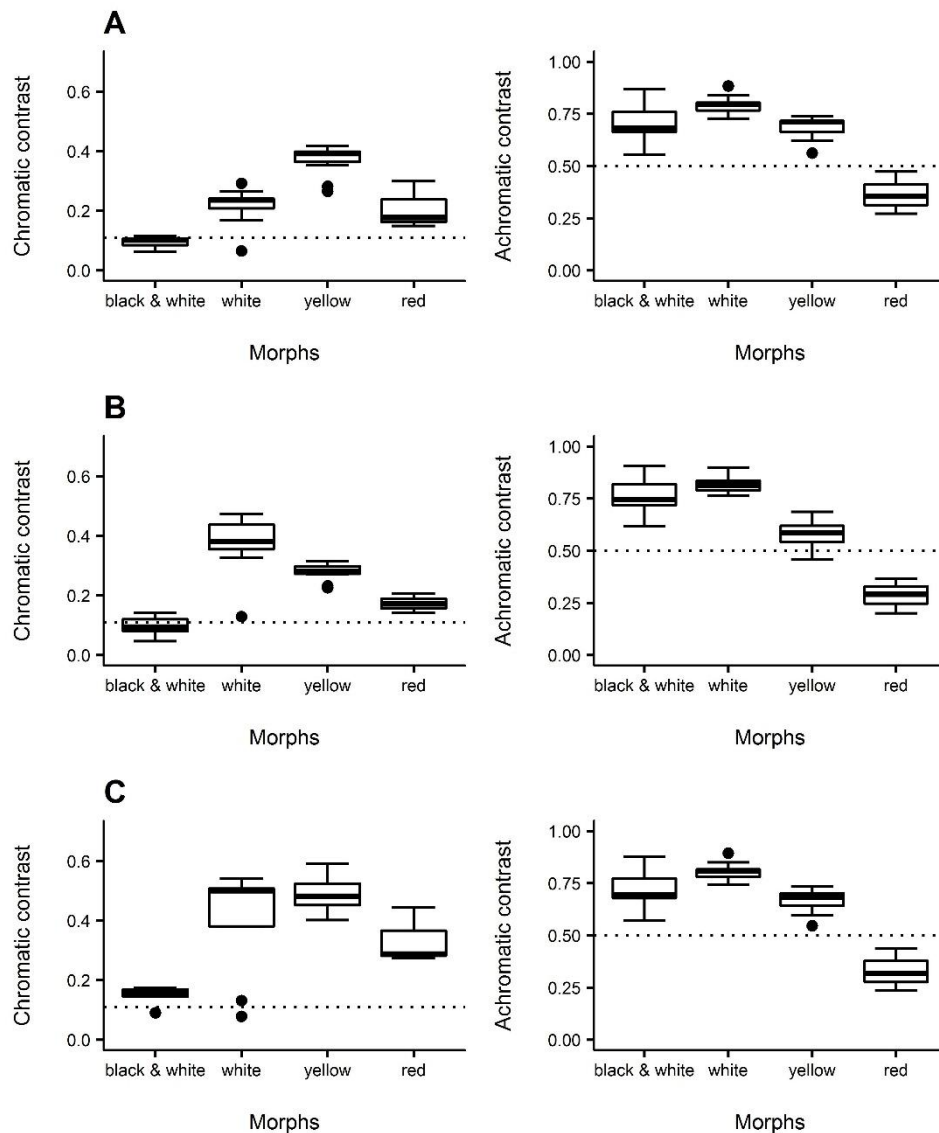
excitation ~ morph*observer	14	0.0
excitation ~ morph+observer	10	14.4
excitation ~ observer	7	21.1
excitation ~ morph	9	36.9
excitation ~ 1	6	43.9

606

607 **Figures**

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610

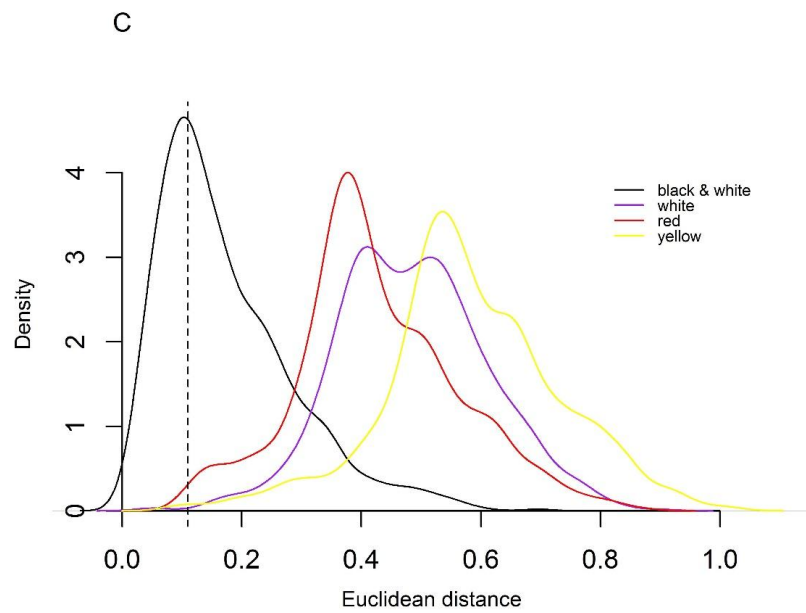
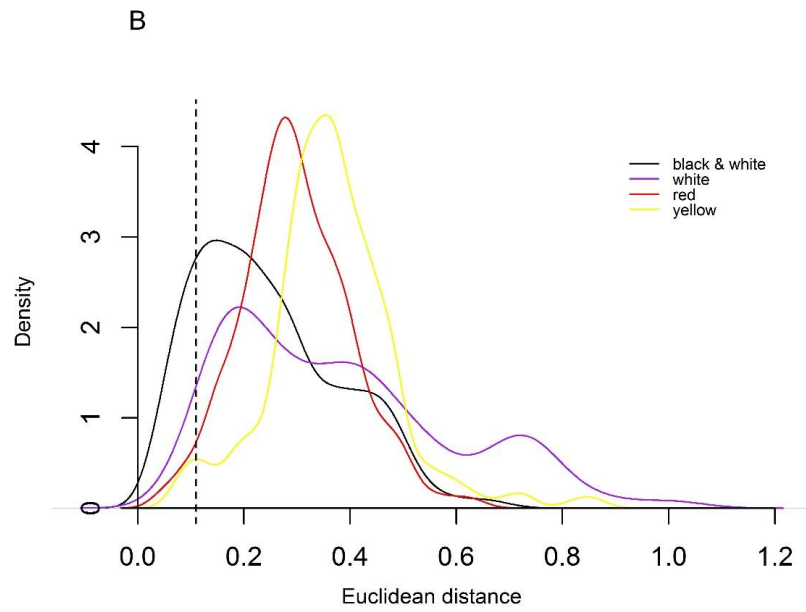
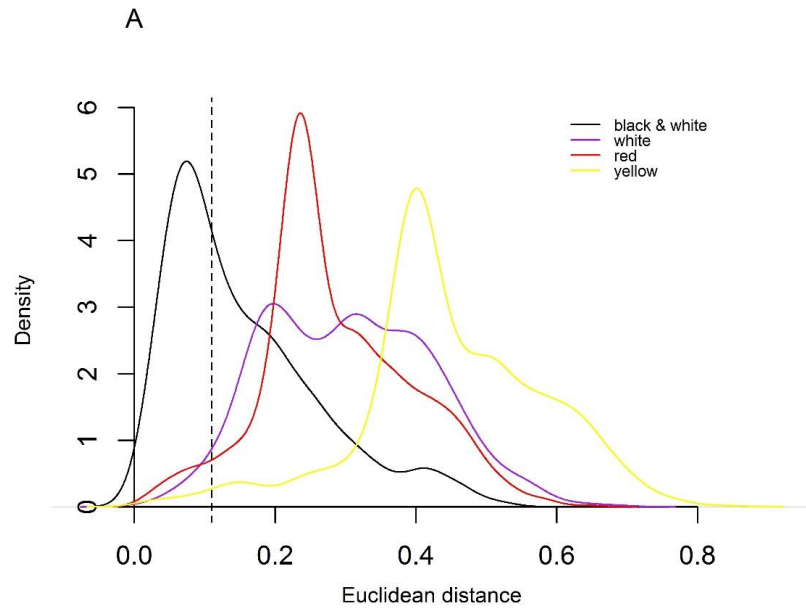
611 **Fig. 1.** Chromatic (left) and achromatic (right) contrasts of four *Gasteracantha cancriformis*
612 morphs (black and white, N=6; white, N=10; yellow, N=13; and red, N=3) when viewed
613 against a Brazilian savanna background by prey with distinct visual systems. (A) *Apis*
614 *mellifera* (Hymenoptera). (B) *Drosophila melanogaster* (Diptera). (C) *Fabriciana adippe*
615 (Lepidoptera). Dotted vertical lines represent the discrimination thresholds for chromatic
616 contrast (0.11) and photoreceptor excitation for background in achromatic contrast (0.5).

617

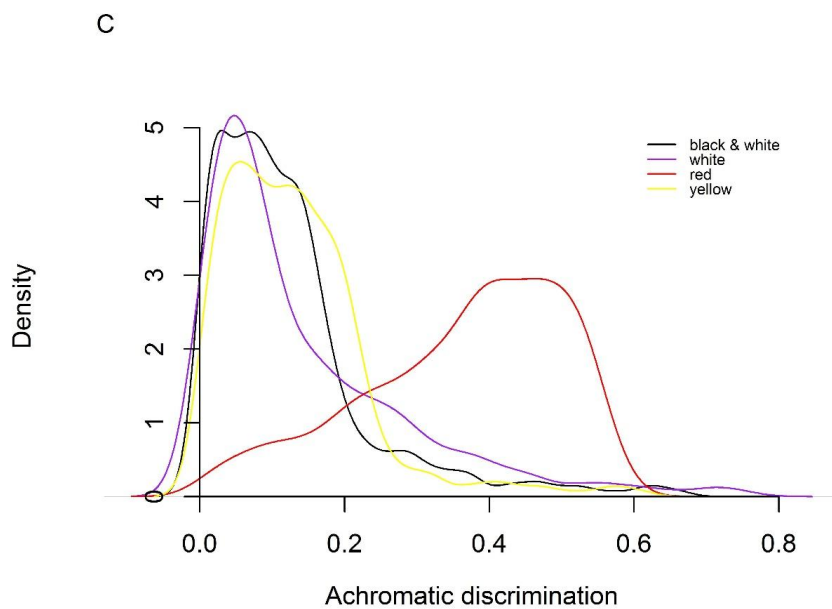
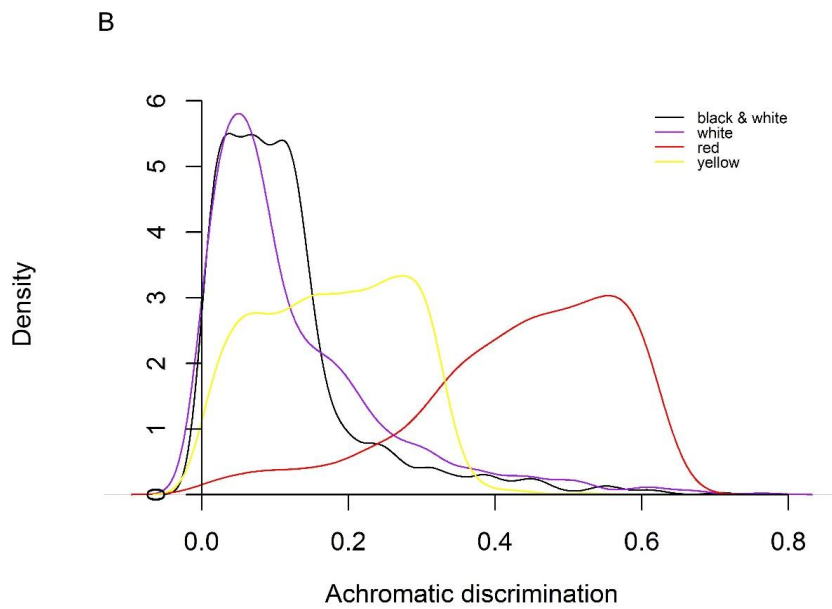
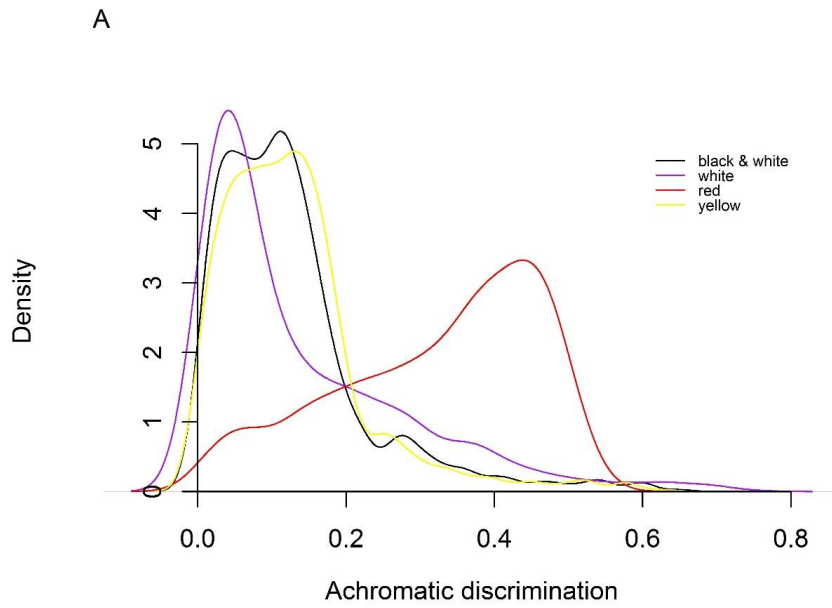
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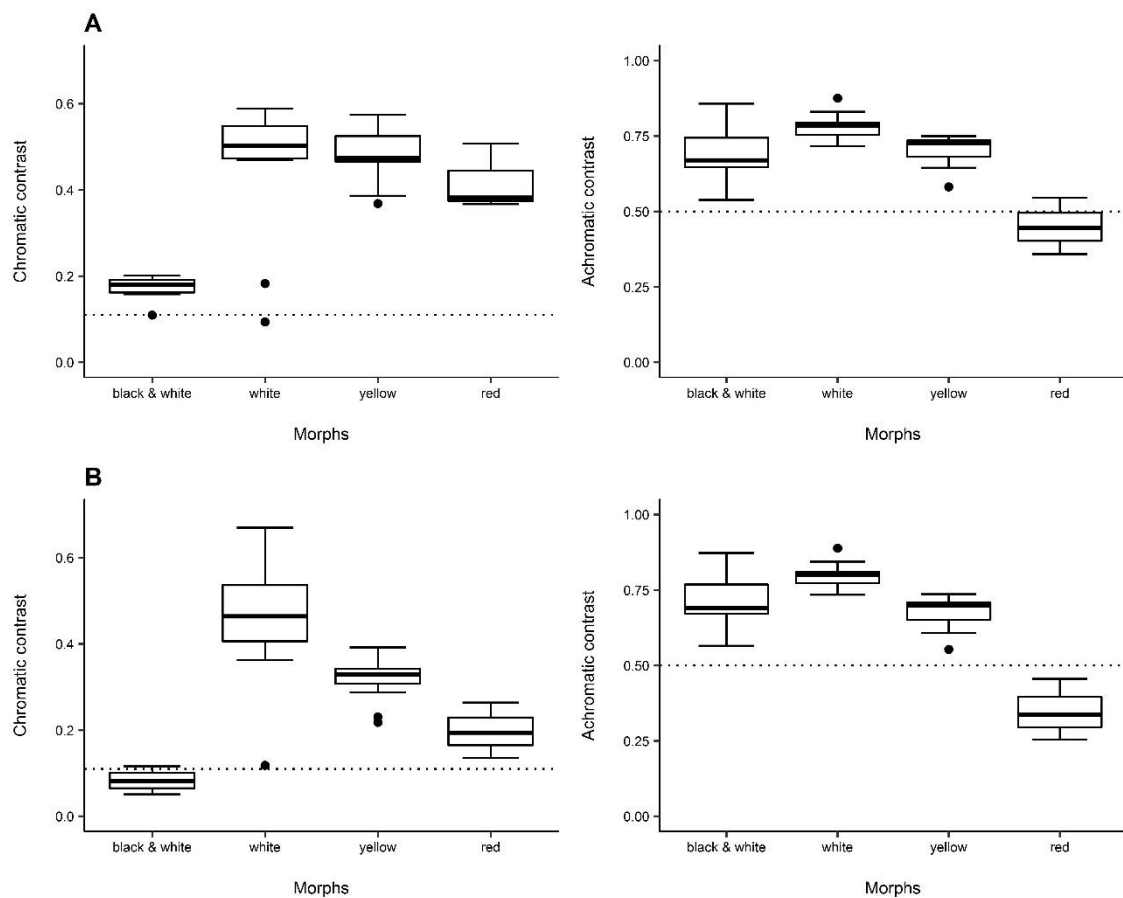


622 **Fig. 2.** Kernel density estimation of the Euclidean distance between four *Gasteracantha*
623 *cancriformis* morphs (black and white, N=6; white, N=10; yellow, N=13; and red, N=3)
624 and flower reflectance (N=859). Estimations were based on the photoreceptors of (A) *Apis*
625 *mellifera* (Hymenoptera), (B) *Drosophila melanogaster* (Diptera), and (C) *Fabriciana*
626 *adippe* (Lepidoptera). Dotted vertical line represents the crude approximation of the color
627 discrimination threshold.
628



630 **Fig. 3.** Kernel density estimation of achromatic contrast between four *Gasteracantha*
631 *cancriformis* morphs (black and white, N=6; white, N=10; yellow, N=13; and red, N=3)
632 and flower reflectance (N=859). Estimations were based on the photoreceptors of (A) *Apis*
633 *mellifera* (Hymenoptera), (B) *Drosophila melanogaster* (Diptera), and (C) *Fabriciana*
634 *adippe* (Lepidoptera). Dotted vertical line represents photoreceptor excitation for the
635 background.

636



637

638 **Fig. 4.** Chromatic (left) and achromatic (right) contrasts of four *Gasteracantha cancriformis*
639 morphs (black and white, N=6; white, N=10; yellow, N=13; and red, N=3) when viewed
640 against a Brazilian savanna background by predators with distinct visual systems. (A)
641 *Parus caeruleus* (Passeriformes). (B) *Philanthus triangulum* (Hymenoptera). Dotted
642 vertical lines represent the discrimination thresholds for chromatic contrast (0.06) and
643 photoreceptor excitation for background in achromatic contrast (0.5).
644