Gene flow between divergent cereal- and grass-specific lineages of the rice blast fungus *Magnaporthe oryzae*

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Abstract

Delineating species and epidemic lineages in fungal plant pathogens is critical to our understanding of disease emergence and the structure of fungal biodiversity, and also informs international regulatory decisions. *Pyricularia oryzae (syn. Magnaporthe oryzae)* is a multi-host pathogen that infects multiple grasses and cereals, is responsible for the most damaging rice disease (rice blast), and of growing concern due to the recent introduction of wheat blast to Bangladesh from South America. However, the genetic structure and evolutionary history of *M. oryzae*, including the possible existence of cryptic phylogenetic species, remain poorly defined. Here, we use whole-genome sequence information for 76 *M. oryzae* isolates sampled from 12 grass and cereal genera to infer the population structure of *M. oryzae*, and to reassess the species status of wheat-infecting populations of the fungus. Species recognition based on genealogical concordance, using published data or extracting previously-used loci from genome assemblies, failed to confirm a prior assignment of wheat blast isolates to a new species (*Pyricularia graminis tritici*). Inference of population subdivisions revealed multiple divergent lineages within *M. oryzae*, each preferentially associated with one host genus, suggesting incipient speciation following host shift or host range expansion. Analyses of gene flow, taking into account the possibility of incomplete lineage sorting, revealed that genetic exchanges have contributed to the makeup of multiple lineages within *M. oryzae*. These findings provide greater understanding of the eco-evolutionary factors that underlie the diversification of *M. oryzae* and highlight the practicality of genomic data for epidemiological surveillance in this important multi-host pathogen.

Importance

Infection of novel hosts is a major route for disease emergence by pathogenic micro-organisms. Understanding the evolutionary history of multi-host pathogens is therefore important to better predict the likely spread and emergence of new diseases. *Magnaporthe oryzae* is a multi-host fungus that causes serious cereal diseases, including the devastating rice blast disease, and wheat blast, a cause of growing concern due to its recent spread from South America to Asia. Using whole genome analysis of 76 fungal strains from different hosts, we have documented the divergence of *M. oryzae* into numerous lineages, each infecting a limited number of host species. Our analyses provide evidence that inter-lineage gene flow has contributed to the genetic makeup of multiple *M. oryzae* lineages within the same species. Plant health surveillance is therefore warranted to safeguard against disease emergence in regions where multiple lineages of the fungus are in contact with one another.

58 ## Introduction

59 Investigating population genetic structure in relation to life history traits such as
60 reproductive mode, host range or drug resistance is particularly relevant in pathogens [1,
61 2]. Knowledge of species, lineages, populations, levels of genetic variability and
62 reproductive mode is essential to answer questions common to all infectious diseases, such
63 as the tempo, origin, proximate (i.e. molecular) and ultimate (eco-evolutionary) causes of
64 disease emergence and spread [3]. Multilocus molecular typing schemes have shown that
65 cryptic species and lineages within species are often more numerous than estimated from
66 phenotypic data alone. Genomic approaches are emerging as a new gold standard for
67 detecting cryptic structure or speciation with increased resolution, allowing fine-grained
68 epidemiological surveillance and science-based regulatory decisions. The added benefit of
69 whole genomes approaches includes identifying the genetic basis of life history traits, and
70 better understanding of both the genomic properties that influence the process of
71 speciation and the signatures of (potentially incomplete) speciation that are observable in
72 patterns of genomic variability [4, 5].

73 Many plant pathogenic ascomycete fungi are host-specific, and some of their life history
74 traits have been shown to be conducive to the emergence of novel pathogen species
75 adapted to new hosts [6, 7]. Investigating population structure within multi-host
76 ascomycetes thus offers a unique opportunity to identify the genomic features associated
77 with recent host-range expansions or host-shifts. In this study, our model is *Magnaporthe*
78 *oryzae* (synonym of *Pyricularia oryzae*) [6-8], a fungal ascomycete causing blast disease on
79 a variety of grass hosts. *Magnaporthe oryzae* is well studied as the causal agent of the most

important disease of rice (*Oryza sativa*), but it also causes blast disease on more than 50 cultivated and wild monocot plant species [9]. This includes other cereal crops such as wheat (*Triticum aestivum*), barley (*Hordeum vulgare*), finger millet (*Eleusine coracana*), and foxtail millet (*Setaria italica*, *S. viridis*), as well as wild and cultivated grass hosts including goosegrass (*Eleusine indica*), annual ryegrass (*Lolium multiflorum*), perennial ryegrass (*L. perenne*), tall fescue (*Festuca arundinacea*) and St. Augustine grass (*Stenotaphrum secundatum*) [10]. Previous studies based on multilocus sequence typing showed that *M. oryzae* is subdivided into multiple clades, each found on only a limited number of host species, with pathogenicity testing revealing host-specificity as a plausible driver of genetic divergence [11, 12]. More recently, comparative genomics of eight isolates infecting wheat, goosegrass, rice, foxtail millet, finger millet and barley revealed deep subdivision of *M. oryzae* into three groups infecting finger millet or wheat, foxtail millet, and rice or barley [13, 14]. Subsequent analysis of genomic data from nine wheat-infecting isolates, two ryegrass infecting isolates, and one weeping lovegrass-infecting isolate subdivided lineages infecting wheat only on the one hand and wheat or ryegrass on the other hand, and revealed an additional lineage associated with the weeping lovegrass strain [15]. Together, these studies suggest a history of host-range expansion or host-shifts and limited gene flow between lineages within *M. oryzae*.

Magnaporthe oryzae isolates causing wheat blast represent a growing concern in terms of food security. This seed-borne pathogen can spread around the world through movement of seed or grain. Therefore, understanding the evolutionary origin and structure of populations causing wheat blast is a top priority for researchers studying disease emergence and for regulatory agencies. Wheat blast was first discovered in southern Brazil

103 in 1985 [16] and the disease subsequently spread to the neighboring countries of
104 Argentina, Bolivia and Paraguay [17-19] where it represents a considerable impediment to
105 wheat production [20, 21]. Until recently, wheat blast had not been reported outside South
106 America. In 2011, a single instance of infected wheat was discovered in the U.S., but
107 analysis of the isolate responsible revealed that it was genetically similar to a local isolate
108 from annual ryegrass and, therefore, unlikely to be an exotic introduction from South
109 America [22]. More recently, in 2016, wheat blast was detected in Bangladesh [23]. Unlike
110 the U.S. isolate, strains from this outbreak resembled South American wheat blast isolates
111 rather than ryegrass-derived strains [15, 23], thereby confirming the spread of wheat blast
112 from South America to Bangladesh.

113 It has recently been proposed that a subgroup of the wheat-infecting isolates, together with
114 some strains pathogenic on *Eleusine* spp. and other *Poaceae* hosts, belongs to a new
115 phylogenetic species, *Pyricularia graminis-tritici* (*Pgt*) that is well separated from other
116 wheat- and ryegrass-infecting isolates, as well as pathogens of other grasses [24]. However,
117 this proposed split was based on bootstrap support in a genealogy inferred from multilocus
118 sequence concatenation, and Genealogical Concordance for Phylogenetic Species
119 Recognition was not applied (GCPSR; [25, 26]). The observed lineage divergence appeared
120 to be mostly driven by genetic divergence at one of 10 sequenced loci, raising questions on
121 the phylogenetic support of this species.

122 The present study was designed to re-assess the hypothesis that *Pgt* constitutes a cryptic
123 species within *M. oryzae* and, more generally, to infer population structure in relation to
124 host of origin in this important plant pathogen. Using whole-genome sequences for 81

125 *Magnaporthe* isolates (76 *M. oryzae* from 12 host plant genera, four *M. grisea* from
126 crabgrass [*Digitaria* spp.], and one *M. pennisetigena* from *Pennisetum* sp.) we addressed the
127 following questions: do *M. oryzae* isolates form distinct host-specific lineages; and is there
128 evidence for relatively long-term reproductive isolation between lineages (i.e. cryptic
129 species) within *M. oryzae*? Our analyses of population subdivision and species
130 identification revealed multiple divergent lineages within *M. oryzae*, each preferentially
131 associated with one host plant genus, but refuted the existence of a novel cryptic
132 phylogenetic species named *P. graminis-tritici*. In addition, analyses of gene flow revealed
133 that genetic exchanges have contributed to the makeup of the multiple lineages within *M.*
134 *oryzae*.

135 Results

136 Re-assessing the validity of the proposed *P. graminis-tritici* species by analyzing the 137 original published data according to Phylogenetic Species Recognition by 138 Genealogical Concordance (GCPSR)

139 To test the previous delineation of a subgroup of wheat-infecting isolates as a new
140 phylogenetic species, we re-analyzed the Castroagudin et al. dataset [24], which mostly
141 included sequences from Brazilian isolates. However, instead of using bootstrap support in
142 a total evidence genealogy inferred from concatenated sequences for species delineation,
143 we applied the GCPSR test [25, 26]. This test identifies a group as an independent
144 evolutionary lineage (i.e. phylogenetic species) if it satisfies two conditions: (1)
145 Genealogical concordance: the group is present in the majority of the single-locus
146 genealogies, (2) Genealogical nondiscordance: the group is well-supported in at least one

147 single-locus genealogy and is not contradicted in any other genealogy at the same level of
148 support [25]. Visual inspection of the topologies and supports in each single-locus tree
149 revealed that GCPSR condition (1) was not satisfied since isolates previously identified as
150 belonging to the phylogenetic species *Pgt* grouped together in only one maximum
151 likelihood gene genealogy – the one produced using the *MPG1* locus (Figure S1A). The *Pgt*
152 separation was not supported by any of the nine other single-locus genealogies (Figures
153 S1B-S1J).

154 Next, we used the multilocus data as input to the program ASTRAL with the goal of
155 inferring a species tree that takes into account possible discrepancies among individual
156 gene genealogies [27-29]. The ASTRAL tree failed to provide strong support for the branch
157 holding the isolates previously identified as *Pgt* (Figure S2). Thus, analysis of the
158 Castroagudin et al. data according to GCPSR standards failed to support the existence of the
159 newly described *Pgt* species.

160 **Inferring population subdivision within *M. oryzae* using whole genome data**

161 We sought to test whether a phylogenomic study could provide better insight into the
162 possibility of speciation within *M. oryzae*. To this end, whole genome sequence data were
163 acquired for a comprehensive collection of 76 *M. oryzae* isolates from 12 host genera, four
164 *M. grisea* isolates from *Digitaria* spp. and one *M. pennisetigena* isolate from *Pennisetum*
165 (Table 1). The analysis included sequence data for strains collected on rice (*Oryza sativa*),
166 finger millet and goosegrass (*Eleusine spp.)*, wheat (*Triticum spp.*), tall fescue (*Festuca*
167 *arundinaceum)*, annual and perennial ryegrasses (*Lolium multiflorum and L. perenne,*
168 respectively), and barley (*Hordeum vulgare*). Representatives of previously unstudied host-

169 specialized populations from foxtails (*Setaria* sp.), St. Augustine grass (*Stenotaphrum*
170 *secundatum*), weeping lovegrass (*Eragrostis curvula*), signalgrass (*Brachiaria* sp.),
171 cheatgrass (*Bromus tectorum*) and oat (*Avena sativa*) were also included. SNPs identified in
172 aligned sequences of 2,682 orthologous single copy genes identified in all *M. oryzae*
173 genomes (in total ~6.6 Mb of sequence data), and from whole-genome SNPs identified from
174 pairwise blast alignments of repeat-masked genomes (average ~36 Mb aligned sequence).

175 First we employed the multivariate approach implemented in Discriminant Analysis of
176 Principal Components (DAPC; [30]) to examine population subdivision within *M. oryzae*.
177 Using the haplotypes identified from orthologous loci, the Bayesian Information Criterion
178 plateaued at K=10 in models varying K from 2 to 20 clusters, indicating that K=10 captures
179 the most salient features of population subdivision (Figure S3). Clusters identified at K=10
180 were as follows: (1) isolates from rice and two isolates from barley (dark green; referred to
181 as the *Oryza* lineage); (2) isolates from *Setaria* sp. (light green; referred to as the *Setaria*
182 lineage); (3) isolate Bm88324 from *Brachiaria mutica* (olive; referred to as the *Brachiaria1*
183 *lineage*); (4) isolate Bd8401 from *Brachiaria distachya* (brown; referred to as the
184 *Brachiaria2* lineage); (5) isolates from *Stenotaphrum* (red; referred to as the *Stenotaphrum*
185 lineage); (6) 17 of the 22 isolates from wheat and an isolate from *Bromus* (blue; referred to
186 as the *Triticum* lineage); (7) the remaining 3/22 isolates from wheat together with isolates
187 from *Lolium*, *Festuca*, oat and a second isolate from *Bromus* (purple; referred to as the
188 *Lolium* lineage); (8 & 9) isolates from *Eleusine* that formed two distinct clusters (light
189 orange and orange; referred to as the *Eleusine1* and *Eleusine2* lineages, respectively); and
190 (10) an isolate from *Eragrostis* (yellow; referred to as the *Eragrostis* lineage) (Figure 1).
191 Increasing K mostly resulted in further subdivision among the isolates from wheat, rice and

192 *Lolium* sp. The discovery of three wheat blast isolates that grouped with the *Festuca-Lolium*
193 pathogens was important because it supports the idea that wheat-infecting isolates belong
194 to at least two distinct populations.

195 Next, we inferred gene genealogies using maximum-likelihood and distance-based methods.
196 Both approaches produced trees that corresponded well with the subdivisions identified in
197 DAPC. The tree generated using maximum likelihood (ML) analysis of orthologous genes
198 displayed a topology with ten lineages (Figure 2) showing one-to-one correspondence with
199 the K clusters from DAPC (Figure 1; Figure S3). Nine of these lineages had >90% bootstrap
200 support. The lineage that corresponded to the "blue" DAPC cluster (including the 17
201 isolates from wheat and isolate P29 from *Bromus*) had poor bootstrap support (50%).

202 The neighbor-joining tree built using "total genome" pairwise distances resolved very
203 similar groupings to the DAPC (Figure 1; Figure S3) and ML ortholog tree (Figure 3). The
204 only major discrepancy between ML and NJ trees was the confident placement of 87-120 –
205 an isolate from rice – outside of the rice clade in the NJ tree (Figure 3).

206 **Levels of polymorphism within and divergence between lineages/species**

207 We compared levels of polymorphism within lineages to levels of divergence between
208 lineages or species to apprehend the relative evolutionary depth of the lineages within *M.*
209 *oryzae*. Genetic variability based on 2,682 orthologs was relatively low and one order of
210 magnitude higher in the rice and wheat lineages (0.1% differences per site) than in the
211 *Lolium* and *Setaria* lineages (other lineages not included in the calculations due to small
212 sample sizes – only lineages with n>6 included; Table 2). The null hypothesis of no

213 recombination could be rejected in the *Lolium*, wheat, rice and *Setaria* lineages using the
214 Pairwise Homoplasy Test implemented in the SPLITSTREE 4.13 program ([31]; p-value:0.0;
215 Table 2).

216 Genome-wide nucleotide divergence was one order of magnitude higher between *M. oryzae*
217 and its closest relatives, *M. grisea* and *M. pennisetigena*, than it was among isolates within
218 *M. oryzae.* The maximum pairwise distance (number of differences per kilobase) between
219 any two *M. oryzae* isolates was less than 1%, genome-wide (Figure S4; Table S1), compared
220 with *M. oryzae* vs *M. grisea*, *M. oryzae* vs. *M. pennisetigena,* or *M. grisea* vs *M. pennisetigena*,
221 all of which were consistently greater than 10%. The low level of genetic divergence among
222 *M. oryzae* isolates, compared with that observed when comparing *M. oryzae* isolates to
223 other established related species, provides good evidence against the existence of relatively
224 ancient cryptic species within *M. oryzae*. (Table S1).

225 **Re-assessment of Pgt as a novel species using whole genome data**

226 While the 10 loci utilized in the Castroagudin et al. [24] study do not support the *Pgt* split
227 based on GCPSR criteria, our DAPC and whole genome ML and NJ analyses supported the
228 partitioning of wheat blast isolates into two, genetically-distinct lineages: one consisting
229 almost exclusively of wheat-infecting isolates, the other comprising largely *Festuca*- and
230 *Lolium*-infecting isolates as well as few wheat-infecting isolates (Figure 2; Figure 3).
231 However, the Castroagudin et al. study did not include *Festuca*- and *Lolium*-infecting
232 isolates and genome sequences from this study are not available. Therefore, to test for
233 possible correspondence between the proposed *Pgt* species and the *Lolium* lineage (or
234 indeed the *Triticum* lineage), we extended the 10 loci analysis to the *M. oryzae* genome

235 sequences used in the present study. For reference, we included the multilocus data for 16
236 isolates from the Castroagudin et al., representing all the major clades from that study. Nine
237 of the 10 loci were successfully recovered from 68 of our *M. oryzae* genome sequences. The
238 remaining locus, CH7-BAC9, was absent from too many genome sequences and, as a result,
239 was excluded from the analysis.

240 The nine concatenated loci produced a total-evidence RAxML tree in which very few
241 branches had bootstrap support greater than 50% (Figure 4). All of the *Pgt* isolates from
242 the Castroagudin et al. study were contained in a clade with 80% support. Inspection of the
243 *MPG1* marker that was reported to be diagnostic for *Pgt* (Castroagudin et al. 2016)
244 revealed that all of the isolates in this clade contained the *Pgt*-type allele (green dots) and
245 should therefore be classified as *Pgt* (Figure 4). Critically, however, a few isolates outside
246 this clade also harbored the *Pgt*-type allele. Moreover, the clade also included isolates from
247 the present study which came from wheat, annual ryegrass, perennial ryegrass, tall fescue,
248 finger millet, and goosegrass – isolates that did not group together in the DAPC analysis
249 (Figure 1), or in the ML and NJ trees built using the orthologous genes or whole genome
250 SNP data (Figure 2; Figure 3). Isolates carrying *Pgt*-type allele were in fact distributed
251 among three genetically distinct and well-supported clades (Figure 2; Figure 3).
252 Furthermore, visual inspection of the topologies and bootstrap supports for each single-
253 locus tree revealed that GCPSR criteria were not satisfied for the clade including all of the
254 *Pgt* isolates from the Castroagudin et al. Thus, isolates characterized by Castroagudin et al.
255 [24] as *Pgt* fail to constitute a phylogenetically cohesive group based on total genome
256 evidence and, thus, the existence of the *Pgt* species is not supported by our new genome-
257 wide data and analyses.

258 The basis for the previous designation of *Pgt* as a novel species was clearly revealed when
259 *MPG1* alleles were mapped onto the ML and NJ trees. The distribution of *MPG1* alleles
260 among different *M. oryzae* lineages was discontinuous (Figure S5). As an example, isolates
261 from the *Triticum* lineage carried three different *MPG1* alleles. Two of these (including the
262 *Pgt*-type) were also present in the *Lolium* lineage, while the third *MPG1* (ACT17T-C-
263 6CAA140, Figure S5) was shared by distantly-related isolates from the *Stenotaphrum*
264 lineage (Figure S5). Isolates from the *Eleusine* lineage also carried *Pgt*-type *MPG1* allele and
265 two other variants, while isolates from the *Setaria* and *Oryza* lineages carried an *MPG1*
266 allele distinct from all the others (Figure S5). Overall, the distribution of *MPG1* alleles
267 points to the occurrence of incomplete lineage sorting and gene flow during *M. oryzae*
268 diversification. Importantly, seven markers studied by Castroagudin et al. – including *MPG1*
269 - showed discontinuities in their distributions among lineages defined using genome-wide
270 data and analyses (Figure S5). The two other markers (*ACT1* and *CHS1*) used by
271 Castroagudin et al. showed no sequence variations among the 68 *M. oryzae* isolates
272 analyzed in the present study (data not shown) and are not useful for phylogenetic
273 classification.

274 **Species tree inference and phylogenetic species recognition from genome-wide data**

275 The total evidence genealogies generated using sequence data from 76 *M. oryzae* genomes
276 using either distance-based (whole genomes) or maximum likelihood (2,682 single-copy
277 orthologs) phylogenetic methods were highly concordant in terms of lineage composition
278 and branching order (Figure 2; Figure 3). However, concatenation methods can be
279 positively misleading, as they assume that all gene trees are identical in topology and
280 branch lengths and they do not explicitly model the relationship between the species tree

281 and gene trees [32]. To estimate the "species tree" and to re-assess previous findings of
282 cryptic species within *M. oryzae*, we used a combination of species inference using the
283 multispecies coalescent method implemented in ASTRAL [27-29] and a new
284 implementation of the GCPSR that can handle genomic data.

285 The ASTRAL "species tree" with the local q1 support values on key branches is shown in
286 Figure 5. The four *M. grisea* isolates from crabgrass (*Digitaria* sp.) and the *M. pennisetigena*
287 isolate from fountaingrass (*Pennisetum* sp.) were included as outgroups, bringing the total
288 number of isolates to 81 and reducing the dataset to 2,241 single-copy orthologous genes.
289 The branches holding the clades containing the wheat blast isolates had q1 support values
290 of 0.49, 0.39 and 0.37 which means that, in each case, fewer than 50% of the whole set of
291 quartet gene trees recovered from the individual gene genealogies agreed with the local
292 topology around these branches in the species tree. The branches that separated *M. grisea*
293 and *M. pennisetigena* from *M. oryzae* had respective q1 values of 1, providing strong
294 support for relatively ancient speciation. In contrast, the highest q1 value on any of the
295 branches leading to the host-specialized clades was 0.8 for the *Setaria* pathogens,
296 indicating that approximately 20% of the quartets recovered from individual gene trees
297 were in conflict with the species tree around this branch. Together, these results indicate
298 high levels of incomplete lineage sorting within, and/or gene flow involving these groups,
299 and are thus inconsistent with the presence of genetically isolated lineages (i.e. species).

300 As a formal test for the presence of cryptic species within *M. oryzae*, we applied the
301 phylogenetic species recognition criteria to the set of 2,241 single-copy orthologous genes
302 using an implementation of the GCPSR scalable to any number of loci. Applying the GCPSR

303 following the non-discordance criterion of Dettman et al. (a clade has to be well supported
304 by at least one single-locus genealogy and not contradicted by any other genealogy at the
305 same level of support; [25]) resulted in the recognition of no species within *M. oryzae*.

306 **Historical gene flow between lineages**

307 The existence of gene flow and/or incomplete lineage sorting was also supported by
308 phylogenetic network analysis. We used the network approach neighbor-net implemented
309 in SPLITSTREE 4.13 [25] to visualize evolutionary relationships, while taking into account the
310 possibility of recombination within or between lineages. The network inferred from
311 haplotypes identified using the 2,682 single-copy orthologs in the 76 *M. oryzae* strains,
312 showed extensive reticulation connecting all lineages, consistent with recombination or
313 incomplete lineage sorting (Figure 6).

314 To disentangle the role of gene flow versus incomplete lineage sorting in observed network
315 reticulations, but also to gain insight into the timing and extent of genetic exchanges, we
316 used ABBA/BABA tests, which compare numbers of two classes of shared derived alleles
317 (the ABBA and BABA classes). For three lineages P1, P2 and P3 and an outgroup with
318 genealogical relationships (((P1,P2),P3),O), and under conditions of no gene flow, shared
319 derived alleles between P2 and P3 (ABBA alleles) and shared derived alleles between P1
320 and P3 (BABA alleles) can only be produced by incomplete lineage sorting, and should be
321 equally infrequent [33]. Differences in numbers of ABBA and BABA alleles are interpreted
322 as gene flow assuming no recurrent mutation and no deep ancestral population structure
323 within lineages. We computed *D*, which measures the imbalance between numbers of ABBA
324 and BABA sites and is used to test for admixture in ((P1,P2),P3) triplets, with D>0 implying

325 gene flow between P2 and P3, and D<0 implying gene flow between P1 and P3 [34, 35]. We
326 also made use of the heterogeneity in divergence time between members of ((P1,P2),P3)
327 triplets to examine gene flow across three time periods [33], following these principles: (i)
328 triplets including the most recently diverged lineages as P1 and P2 (i.e. the *Triticum* and
329 *Lolium* lineages, the two *Eleusine* lineages, or the *Oryza* and *Setaria* lineages) carried
330 information about gene flow across relatively recent times, (ii) triplets including, as P1 and
331 P2, two lineages from the same main group of lineages (i.e.
332 *Eragrostis*/*Eleusine*1/*Eleusine*2/*Triticum*/*Lolium* or *Brachiaria2*/*Setaria*/*Oryza*, excluding
333 (P1,P2) pairs already used in (i)) carried information about gene flow across intermediate
334 times, and (iii) triplets including, as P1 and P2, two lineages from different main groups of
335 lineages (i.e. *Eragrostis* / *Eleusine*1 / *Eleusine*2 / *Triticum* / *Lolium* and *Brachiaria2* /
336 *Setaria* / *Oryza*) and *Stenotaphrum* or *Brachiaria1* as P3 carried information about gene
337 flow across a relatively long time period (Figure S6).

338 The D statistic measuring differences in counts of ABBA and BABA alleles was significantly
339 different from zero (Z-score>3) in 104 of 120 lineage triplets, consistent with a history of
340 gene flow between lineages within *M. oryzae* (Table S2). Given that a (P1,P2) pair can be
341 represented as multiple ((P1,P2),P3) triplets, and that the sign of D indicates what is the
342 pair involved in gene flow within each triplet, the 104 triplets with significant D values in
343 fact represented 35 pairs connected by gene flow, spanning the three time scales defined
344 by the phylogenetic affiliation of lineages (Figure S6). Lineages were equally represented in
345 triplets deviating from null expectations assuming no gene flow, no ancient structure and
346 no recurrent mutations. Consistent with historical gene flow, searches for private allele
347 found no gene, among the 2241 gene surveyed, carrying mutations exclusive to a single

348 lineage. Together, these results indicate that gene flow was widespread, both across
349 historical times and lineages, but it cannot be excluded that much of the signal was caused
350 by events that happened prior to lineage splitting.

351 **Recent admixture and gene flow between lineages**

352 We then used the program STRUCTURE [36-38] to detect possible recent admixture between
353 lineages (Figure S3). STRUCTURE uses Markov chain Monte Carlo simulations to infer the
354 assignment of genotypes into K distinct clusters, minimizing deviations from Hardy–
355 Weinberg and linkage disequilibria within each cluster. The patterns of clustering inferred
356 with STRUCTURE were largely similar to those inferred with DAPC. STRUCTURE analysis
357 provided evidence for admixture at all K values (Figure S3), suggesting that recent
358 admixture events have recently shaped patterns of population subdivision within *M. oryzae*.
359 'Chromosome painting', a probabilistic method for reconstructing the chromosomes of each
360 individual sample as a combination of all other homologous sequences [39], also supported
361 the lack of strict genetic isolation between lineages (Text S1).

362 **Discussion**

363 **Population subdivision but no cryptic phylogenetic species**

364 Using population- and phylogenomic analyses of single-copy orthologous genes and whole-
365 genome SNPs identified in *M. oryzae* genomes from multiple cereal and grass hosts, we
366 provide evidence that *M. oryzae* is subdivided in multiple lineages preferentially associated
367 with one host plant genus. Neither the re-analysis of previous data, nor the analysis of new
368 data using previous phylogenetic species recognition markers, supports the existence of a
369 wheat blast-associated species called *P. graminis-tritici* [24]. Marker *MPG1*, which holds

370 most of the divergence previously detected, does not stand as a diagnostic marker of the
371 wheat-infecting lineage of *M. oryzae* when tested in other lineages. Previous conclusions
372 about the existence of cryptic species *P. graminis-tritici* also stem from the fact that
373 available information on *M. oryzae* diversity had been insufficiently taken into account. In
374 particular, isolates from the lineages most closely related to wheat strains (i.e. isolates from
375 the *Lolium* lineage; [11, 12, 15, 22]) were not represented in previous species identification
376 work [24]. Using phylogenetic species recognition by genealogical concordance we could
377 not identify cryptic phylogenetic species and thus *M. oryzae* is not, strictly speaking, a
378 species complex. As a consequence, *Pyricularia graminis-tritici* cannot - and should not –
379 be considered as a valid name for wheat-infecting strains, because (1) it refers to a subset
380 of wheat-infecting strains, and quarantine on *P. graminis-tritici* alone would not prevent
381 introduction of aggressive wheat blast pathogens (2) it groups very aggressive wheat
382 pathogens from South America and South Asia with *Eleusine*-infecting strains that are
383 largely distributed in the world. Given the devastating potential of wheat blast disease, it is
384 vital that accurate strain identification and species assignment can be carried out by plant
385 health agencies in order to safeguard against importation and spread of the
386 disease. Correct species assignment is therefore a critical consideration. Hence, although
387 the formal rules of taxonomy would imply treating *P. graminis-tritici* as synonym of
388 *Magnaporthe oryzae*, we strongly recommend dismissal of *P. graminis-tritici* as a
389 valid name to refer to wheat-infecting strains of *M. oryzae*.

390 **Incipient speciation by ecological specialization following host-shifts**

391 Several features of the life cycle of *M. oryzae* are conducive to speciation by ecological
392 specialization following host shifts, suggesting that the observed pattern of population

393 subdivision in *M. oryzae* actually corresponds to ongoing speciation. Previous experimental
394 measurements of perithecia formation and ascospore production – two important
395 components of reproductive success- suggested inter-fertility in vitro between most pairs
396 of lineages with high levels of ascospore viability [40-43]. This suggests that intrinsic pre-
397 or post-mating reproductive barriers, such as assortative mating by mate choice or gametic
398 incompatibility, and zygotic mortality, are not responsible for the relative reproductive
399 isolation between lineages – which creates the observed pattern of population subdivision.
400 Instead, the relative reproductive isolation between lineages could be caused by one or
401 several pre- or post-mating barriers (Table 1 in [44]), such as mating-system isolation or
402 hybrid sterility (intrinsic barrier), or difference in mating times, difference in mating sites,
403 immigrant inviability, or ecologically-based hybrid inviability (extrinsic barriers).

404 Previous pathogenicity assays revealed extensive variability in the host range of *M. oryzae*
405 isolates, and both in terms of pathogenicity towards a set of host species or pathogenicity
406 towards a set of genotypes from a given host [41, 45]. Indeed, extensive genetic analyses
407 show that host species specificity in *M. oryzae*, similar to rice cultivar specificity, could be
408 controlled by a gene-for-gene relationship in which one to three avirulence genes in the
409 fungus prevent infection of particular host species [43, 46, 47]. Loss of the avirulence genes
410 would allow infection of novel hosts to occur. Additionally, host species specificity is not
411 strictly maintained. Under controlled conditions, most lineages have at least one host in
412 common [45], and strains within one lineage can still cause rare susceptible lesions on
413 naive hosts [21, 48]. Moreover, a single plant infected by a single genotype can produce
414 large numbers of spores in a single growing season [49], allowing the pathogen to persist
415 on alternative host even if selection is strong, and promoting the rapid and repeated

416 creation of genetic variation [6]. Although some of these features appear to be antagonistic
417 with the possibility of divergence by host-specialization within *M. oryzae*, our finding that
418 the different lineages within *M. oryzae* tend to be sampled on a single host suggests that
419 ecological barriers alone may in fact contribute to reduce gene flow substantially between
420 host-specific lineages. Differences in the geographic distribution of hosts, for which the
421 level of sympatry has varied -and still varies- in space and time, might also contribute to
422 reduced gene flow between lineages infecting different hosts, although some level of
423 sympatry at some time is required so that new hosts could become infected, triggering
424 host-range expansion or host-shifting.

425 Mating within host (i.e. reproduction between individuals infecting the same host), and to a
426 lesser extent mating system isolation (i.e. lack of outcrossing reproduction), may
427 contribute to further reduce gene flow between *M. oryzae* lineages. The fact that mating in
428 *M. oryzae* likely occurs within host tissues, such as dead stems [50], may participate in the
429 maintenance of the different lineages by decreasing the rate of reproduction between
430 isolates adapted to different hosts [6]. Loss of sexual fertility also appears to have a role in
431 lineage maintenance. The rice lineage, in particular, is single mating-type and female-sterile
432 throughout most of its range, which would reduce the chance of outcrossing sex with
433 members of other lineages [51]. Our analyses rejected the null hypothesis of clonality in all
434 lineages, but they provided no time frame for the detected recombination events.
435 Population-level studies and experimental measurements of mating type ratios and female
436 fertility are needed to assess the reproductive mode of the different lineages within *M.*
437 *oryzae* in the field.

438 **Inter-lineage gene flow**

439 Several potential barriers contribute to reduce genetic exchanges between *M. oryzae*
440 lineages (see previous paragraph), but not completely so, as evidenced by signal of gene
441 flow and admixture detected in our genomic data. We hypothesize that the lack of strict
442 host specialization of the different lineages is a key driver of inter-lineage gene flow. Many
443 of the grass or cereal species that are hosts to *M. oryzae* are widely cultivated as staple
444 crops or widely distributed as pasture or weeds, including "universal suscepts" such as
445 barley, Italian ryegrass, tall fescue and weeping lovegrass [40], increasing the chance for
446 encounters and mating between isolates with overlapping host ranges. These shared hosts
447 may act as a platform facilitating encounters and mating between fertile and compatible
448 isolates from different lineages, thereby enabling inter-lineage gene flow [52]. Plant health
449 vigilance is therefore warranted for disease emergence via recombination in regions where
450 multiple lineages are in contact and shared hosts are present. This is particularly so, given
451 that once infection of novel host has taken place (i.e. host shift or host range expansion),
452 the fungus has the capacity to build inoculum levels very rapidly, facilitating spread of the
453 disease over considerable distances. It is striking, for example, that wheat blast has, within
454 a year, spread from Bangladesh into the West Bengal region of India where it emerged in
455 2017 (openwheatblast.org).

456 **Conclusion**

457 Using a population genomics framework, we show that *M. oryzae* is subdivided into
458 multiple lineages with limited host range and present evidence of genetic exchanges
459 between them. Our findings provide greater understanding of the eco-evolutionary factors
460 underlying the diversification of *M. oryzae* and highlight the practicality of genomic data for

461 epidemiological surveillance of its different intraspecific lineages. Reappraisal of species
462 boundaries within *M. oryzae* refuted the existence of a novel cryptic phylogenetic species
463 named *P. graminis-tritici,* underlining that the use of node support in total evidence
464 genealogies based on a limited dataset in terms of number of loci and of range of variation
465 in origin (geography and host) of isolates can lead to erroneously identify fungal cryptic
466 species. Our work illustrates the growing divide between taxonomy that 'creates the
467 language of biodiversity' [53] based on limited sets of characters, and genomic data that
468 reveals more finely the complexity and continuous nature of the lineage divergence process
469 called speciation.

470 Materials and Methods

471 Fungal strains

472 Thirty-eight newly sequenced genomes were analyzed together with 43 published
473 genomes [13, 14, 22, 54-56] resulting in a total of 81 *Magnaporthe* strains, including 76 *M.*
474 *oryzae* genomes representing 12 different hosts available for analysis (Table 1). We also
475 included as outgroups one strain of *Pyricularia pennisetigena* from *Pennisetum* sp. and four
476 strains of *Pyricularia grisea* (syn. *Magnaporthe grisea*) from crabgrass (*Digitaria*
477 *sanguinalis*). All newly sequenced strains were single-spored prior to DNA extraction.
478

479 Genome sequencing and assembly

480 New genome data were produced by an international collaborative effort. Characteristics of
481 genome assemblies are summarized in Table S3. For newly sequenced genomes provided
482 by MF and BV, sequences were acquired on a MiSeq machine (Illumina, Inc.). Sequences

483 were assembled using the paired-end mode in NEWBLER V2.9 (Roche Diagnostics,
484 Indianapolis, IN). A custom perl script was used to merge the resulting scaffolds and
485 contigs files in a non-redundant fashion to generate a final assembly. Newly sequenced
486 genomes BR130 and WHTQ provided by TM were sequenced using an Illumina paired-end
487 sequencing approach at >50X depth. Short reads were assembled *de-novo* using VELVET 1.2.10
488 [57] resulting in a 41.5Mb genome for BR130 with N50 44.8Kb, and 43.7Mb for WHTQ with
489 N50 36.2Kb. For newly sequenced genomes provided by DS and NT, DNA was sequenced on
490 the Illumina HiSeq 2500 producing 100 base paired-end reads, except in the case of VO107
491 which was sequenced on the Illumina Genome Analyzer II producing 36 base paired-end
492 reads. Reads were filtered using FASTQ-MCF and assembled 'de novo' using VELVET 1.2.10
493 [57].
494

495 **Orthologous genes identification in genomic sequences**

496 Protein-coding gene models were predicted using AUGUSTUS V3.0.3 [58]. Orthologous genes
497 were identified in the 76-genomes *M. oryzae* or in the dataset including outgroups using
498 PROTEINORTHO [59]. The v8 version of the 70-15 *M. oryzae* reference genome [60] was
499 added at this step in order to validate the predicted sets of orthologs. Only orthologs that
500 were single-copy in all genomes were included in subsequent analyses. Genes of each
501 single-copy orthologs sets were aligned using MACSE [61]. Sequences from the lab strain
502 70-15 were removed and not included in further analyses due to previously shown hybrid
503 origin [13]. Only alignments containing polymorphic sites within *M. oryzae* strains were
504 kept for further analyses. This resulted in 2,241 alignments for the whole dataset, and
505 2,682 alignments for the 76 *M. oryzae* strains.

506

507 **Population subdivision, summary statistics of polymorphism and divergence**

508 Population subdivision was analyzed using DAPC and STRUCTURE [30, 36-38], based on
509 multilocus haplotype profiles identified from ortholog alignments using a custom Python
510 script. DAPC was performed using the ADEGENET package in R [13]. We retained the first 30
511 principal components, and the first 4 discriminant functions. Ten independent STRUCTURE
512 runs were carried out for each number of clusters K, with 100,000 MCMC iterations after a
513 burn-in of 50,000 steps.

514 Polymorphism statistics were computed using EGGLIB 3.0.0b10 [62] excluding sites with
515 >30% missing data. Divergence statistics were computed using a custom perl script.

516 To infer total evidence trees within the 76 *M. oryzae* strains (respectively within the 81
517 *Magnaporthe* strains), all sequences from the 2,682 (respectively 2,241) orthologous
518 sequences were concatened. The maximum-likelihood tree was inferred using RAxML [63]
519 with the GTR-gamma model, and bootstrap supports were estimated after 1000 replicates.

520 **Retrieval of loci used in the Castroagudin et al. (2016) study**

521 The 10 loci used by Castroagudin et al. [24], i.e. actin (ACT), beta-tubulin1 (βT-1),
522 calmodulin (CAL), chitin synthase 1 (CHS1), translation elongation factor 1-alpha (EF1-α),
523 hydrophobin (MPG1), nitrogen regulatory protein 1 (NUT1), and three anonymous
524 markers (CH6, CH7-BAC7, CH7-BAC9), were search in all genomes using BLASTn. Due to
525 heterogeneity in the quality of assemblies, 9 of the 10 loci could be full-length retrieved
526 without ambiguity in 68 out of the 81 available genomes, still representative of the
527 diversity of host plants.

528

529 **Secondary data analysis**

530 Species recognition based on multiple gene genealogies as described by Castroagudin et al.
531 [24] was repeated following the reported methods. The robustness of the Pgt species
532 inference was tested by re-iterating the study, omitting one marker at a time. Individual
533 genealogies were built using RAxML with the GTR-gamma model and 100 bootstrap
534 replicates.

535

536 **Inference of "species tree" using ASTRAL**

537 The ASTRAL method [27, 29] is based on the multi-species coalescent and allows taking
538 into account possible discrepancies among individual gene genealogies to infer the "species
539 tree". Individual genealogies inferred using RAxML with the GTR-gamma model and 100
540 bootstrap replicates, were used as input data for ASTRAL analysis. Local supports around
541 branches were evaluated with 100 multilocus bootstrapping using the bootstrap replicates
542 inferred from each individual gene tree as input data, and with local quartet supports (q1,
543 obtained using the –t option of ASTRAL) that represent the proportion of quartets
544 recovered from the whole set of individual gene trees that agree with the local topology
545 around the branch in the species tree.

546

547 **MPG1-based classification**

548 The *MPG1* hydrophobin sequence is described as being diagnostic for the
549 *P. graminis-tritici/M. oryzae* species split [24]. *MPG1* sequences from one of each species

550 (GIs: KU952644.1 for *P. gramini-triticis*, KU952661.1 for *M. oryzae*) were used as BLAST
551 [64] queries to classify isolates as either *P. graminis-tritici* or *M. oryzae.*
552
553 **Signatures of gene flow and/or incomplete lineage sorting**
554 A phylogenetic network was built using SPLITSTREE 4.13 [65], based on the concatenation of
555 sequences at single-copy orthologs identified in *M. oryzae*, excluding sites with missing
556 data, sites with gaps, singletons, and monomorphic sites. The null hypothesis of no
557 recombination was tested using the PHI test implemented in SPLITSTREE.
558
559 **ABBA/BABA tests**
560 ABBA/BABA tests were performed using custom python scripts. The D statistic measuring
561 the normalized difference in counts of ABBA and BABA sites was computed using equation
562 (2) in ref. [66]. Significance was calculated using block jackknife approach (100 replicates,
563 1k SNPs blocks), to account for non-independence among sites.
564
565 **Probabilistic chromosome painting.**

566 We used CHROMOPAINTER program version 0.0.4 for probabilistic chromosome painting. This
567 analysis was based on biallelic SNPs without missing data identified in the set of 2,682
568 single-copy orthologs, ordered according to their position in the reference genome of the
569 rice-infecting strain 70-15. We initially estimated the recombination scaling constant N_e
570 and emission probabilities (μ) by running the expectation-maximization algorithm with
571 200 iterations for each lineage and chromosome. Estimates of N_e and μ were then
572 computed as averages across lineages, weighted by chromosome length, rounded to the

573 nearest thousand for N_e (N_e=5000; μ=0.0009). The file *recom_rate_infile* detailing the
574 recombination rate between SNPs was built using the INTERVAL program in LDHAT version
575 2.2 [67] based on the whole dataset combining isolates from all lineages, with 10 repeats
576 by chromosome to check for convergence. Estimated N_e and μ values and the per-
577 chromosome recombination maps estimated using LDHAT were then used to paint the
578 chromosomes of each lineage, considering the remaining lineages as donors, using 200
579 expectation-maximization iterations. For each lineage and each chromosome,
580 CHROMOPAINTER was run thrice to check for convergence.

581

582 **Phylogenetic species recognition**

583 We used an implementation of the GCPSR scalable to genomic data (https://github.com/b-
584 brankovics/GCPSR). The method works in two steps: (1) Concordance and non-
585 discordance analysis produces a genealogy that has clades that are both concordant and
586 non-discordant across single gene genealogies, with support value for each of the clades
587 being the number of single gene genealogies harboring the given clade at bootstrap support
588 above 95%; (2) Exhaustive subdivision places all the strains into the least inclusive clades,
589 by removing clades that would specify a species within potential phylogenetic species. We
590 kept only two outgroup sequences per gene (BR29, *M. grisea*; Pm1, *M. pennisetigena*) to
591 make sure to have the same isolate at the root of all genealogies (Pm1 isolate). Majority-
592 rule consensus trees were produced from 100 outgrouped RAxML bootstrap replicates for
593 all 2241 genes. The concordance and non-discordance analysis was carried out assuming
594 95 as the minimum bootstrap support value, and a discordance threshold of 1. Exhaustive
595 subdivision was carried out using a concordance threshold of 1121.

596

597 **Whole genome alignment & tree building**

598 A custom perl script was used to mask sequences that occur in multiple alignments when
599 the genome is BLASTed against itself. The masked genomes were then aligned in a pairwise
600 fashion against all other genomes using BLAST [64]. Regions that did not uniquely align in
601 each pair at a threshold of $1e^{-200}$, were excluded. SNPs were then identified for each
602 pairwise comparison and scaled by the total number of nucleotides aligned after excluding
603 repetitive and duplicate regions. This produced a distance metric of SNPs/Mb of uniquely
604 aligned DNA. The pairwise distances were used to construct phylogenetic trees with the
605 neighbor-joining method as implemented in the R package, Analyses of Phylogenetics and
606 Evolution (APE) [68].

607 Because alignments are in pairwise sets as opposed to a single orthologous set, assessment
608 of confidence values by traditional bootstrapping by resampling with replacement is not
609 possible. Instead, confidence values were assigned by creating 1,000 bootstrap trees with
610 noise added from a normal distribution with a mean of zero, and the standard deviation
611 derived from the pairwise distances between or within groups.

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Tables

Table 1. *Magnaporthe oryzae*, *M. grisea* and *M. pennisetigena* strains used in this study

Isolate ID	Synonym	Host	Year	Locality	NCBI accession	Sequence source	References
BdBar	BdBar16-1	*Triticum aestivum*	2016	Barisal, Bangladesh		[23]	
BdJes	BdJes16-1	*Triticum aestivum*	2016	Jessore, Bangladesh		[23]	
BdMeh	BdMeh16-1	*Triticum aestivum*	2016	Mehepur, Bangladesh		[23]	
B2		*Triticum aestivum*	2011	Bolivia	XXXXXXXX	[69]	
B71		*Triticum aestivum*	2012	Bolivia		[23]	[69]
Br7		*Triticum aestivum*	1990	Parana, Brazil	XXXXXXXX	[22]	[69]
BR0032	BR32	*Triticum aestivum*	1991	Brazil		[13]	[15]
Br48		*Triticum aestivum*	1990	Mato Grosso do Sul, Brazil		[14]	[22]
Br80		*Triticum aestivum*	1991	Brazil	XXXXXXXX	[22]	[69]
Br130		*Triticum aestivum*	1990	Mato Grosso do Sul, Brazil	XXXXXXXX	[22]	
P3		*Triticum durum*	2012	Canindeyu, Paraguay	XXXXXXXX	[69]	
PY0925		*Triticum aestivum*	2009	Predizes, Brazil		[15]	
PY36-1	PY36.1	*Triticum aestivum*	2007	Brasilia, Brazil		[15]	
PY5003	PY05003	*Triticum aestivum*	2005	Londrina, Brazil		[15]	
PY5010	PY05010	*Triticum aestivum*	2005	Londrina, Brazil		[15]	[69]
PY5033	PY05033	*Triticum aestivum*	2005	Londrina, Brazil		[15]	
PY6017	PY06017	*Triticum aestivum*	2006	Coromandel, Brazil		[15]	
PY6045	PY06045	*Triticum aestivum*	2006	Goiania, Brazil		[15]	
PY86-1	PY86.1	*Triticum aestivum*	2008	Cascavel, Brazil		[15]	
T25		*Triticum aestivum*	1988	Parana, Brazil	XXXXXXXX	[22]	[69]
WHTQ		*Triticum aestivum*	ND	Brazil	XXXXXXXX	[22]	
WBKY11	WBKY11-15	*Triticum aestivum*	2011	Lexington, KY, USA	XXXXXXXX	[22]	[69]
P28	P-0028	*Bromus tectorum*	2014	Paraguay	XXXXXXXX	[69]	
P29	P-0029	*Bromus tectorum*	2014	Paraguay	XXXXXXXX	[69]	
CHRF		*Lolium perenne*	1996	Silver Spring, MD, USA	XXXXXXXX	[69]	
CHW		*Lolium perenne*	1996	Annapolis, MD, USA	XXXXXXXX	[69]	
FH		*Lolium perenne*	1997	Hagerstown, MD, USA	XXXXXXXX	[22]	[69]
GG11		*Lolium perenne*	1997	Lexington, KY, USA	XXXXXXXX	[22]	
HO		*Lolium perenne*	1996	Richmond, PA, USA	XXXXXXXX	[22]	
LpKY97	LpKY97-1	*Lolium perenne*	1997	Lexington, KY, USA	XXXXXXXX	[22]	[69]

PgKY	PgKY4OV2.1	*Lolium perenne*	2000	Lexington, KY, USA		[15]	
PGPA	PgPA18C-02, PgPA	*Lolium perenne*	1998	Pennsylvania, USA		[15]	
PL2-1		*Lolium multiflorum*	2002	Pulaski Co., KY, USA	XXXXXXXX	[22]	
PL3-1		*Lolium multiflorum*	2002	Pulaski Co., KY, USA	XXXXXXXX	[22]	[69]
Pg1213-22		*Festuca arundinaceum*	1999/ 2000	GA	XXXXXXXX	[69]	
TF05-1		*Festuca arundinaceum*	2005	Lexington, KY, USA	XXXXXXXX	this study	
IB33		*Oryza sativa*	ND	Texas, USA	XXXXXXXX	A. Marchetti	
FR13	FR0013	*Oryza sativa*	1988	France		[13]	[15]
GY11	GY0011, Guy11	*Oryza sativa*	1988	French Guyana		[13]	[15, 22]
IA1	ARB114	*Oryza sativa*	2009	Arkansas, USA	XXXXXXXX	[69]	
IB49	ZN61	*Oryza sativa*	1992	Arkansas, USA	XXXXXXXX	[69]	
IC17	ZN57	*Oryza sativa*	1992	Arkansas, USA	XXXXXXXX	[69]	
IE1K	TM2	*Oryza sativa*	2003	Arkansas, USA	XXXXXXXX	[69]	
INA168	Ina168	*Oryza sativa*	1958	Aichi, Japan		[14]	[70]
KEN53-33	Ken53-33	*Oryza sativa*	1953	Aichi, Japan		[14]	
ML33		*Oryza sativa*	1995	Mali	XXXXXXXX	this study	
P131		*Oryza sativa*	ND	Japan		[55]	[22, 56]
Y34		*Oryza sativa*	1982	China, Yunnan		[55]	[22, 56]
P-2	P2	*Oryza sativa*	1948	Aichi, Japan		[14]	
PH0014-rn	PH0014, PH14	*Oryza sativa*	ND	The Philippines		[13]	[15]
TH3		*Oryza sativa*	ND	Thailand		[14]	[14]
87-120		*Oryza sativa*	ND		XXXXXXXX	this study	
TH0012-rn	TH0012, TH12	*Hordeum vulgare*	ND	Thailand		[13]	[15]
TH0016	TH16	*Hordeum vulgare*	ND	Thailand		[13]	[15]
Arcadia		*Setaria viridis*	1998	Lexington, KY, USA	XXXXXXXX	[22]	[69]
US0071	US71	*Setaria spp.*	ND	USA		[13]	[15]
GrF52		*Setaria viridis*	2001	Lexington, KY, USA	XXXXXXXX	this study	
KANSV1-4-1	KNSV	*Setaria viridis*	1975	Kanagawa, Japan		[14]	
SA05-43		*Setaria viridis*	2005	Nagasaki, Japan		[14]	
Sv9610		*Setaria viridis*	1996	Zhejian, China		[56]	
Sv9623		*Setaria viridis*	1996	Zhejian, China		[56]	
GFSI1-7-2	GFSI	*Setaria italica*	1977	Gifu, Japan		[14]	

B51		*Eleusine indica*	2012	Quirusillas, Bolivia	XXXXXXXX	[22]	[56, 69]
BR62		*Eleusine indica*	1991	Brazil		[15]	
CD156	CD0156	*Eleusine indica*	1989	Ferkessedougou, Ivory Coast		[13]	[15]
EI9411		*Eleusine indica*	1990	Fujian, China		[56]	
EI9064		*Eleusine indica*	1996	Fujian, China		[56]	
G22	WGG-FA40	*Eleusine coracana*	1976	Japan	XXXXXXXX	this study	
Z2-1		*Eleusine coracana*	1977	Kagawa, Japan		[14]	[70]
PH42		*Eleusine indica*	1983	The Philippines	XXXXXXXX	[69]	
SSFL02		*Stenotaphrum secundatum*	2002	Disneyworld, FL, USA	XXXXXXXX	[69]	
SSFL14-3		*Stenotaphrum secundatum*	2014	New Smyrna, FL, USA	XXXXXXXX	this study	
G17	K76-79	*Eragrostis curvula*	1976	Japan	XXXXXXXX	[15]	
Br58		*Avena sativa*	1990	Parana, Brazil		[14]	[70]
Bd8401		*Brachiaria distachya*	1984	The Philippines	XXXXXXXX	this study	
Bm88324		*Brachiaria mutica*	1988	The Philippines	XXXXXXXX	this study	
PM1		*Pennisetum americanum*	1990	Georgia, USA	XXXXXXXX	this study	
BR29	BR0029	*Digitaria sanguinalis*	1989	Brazil		[13]	
Dig41		*Digitaria sanguinalis*	ND	Hyogo, Japan		[14]	[70]
DsLIZ		*Digitaria sanguinalis*	2000	Lexington, KY, USA	XXXXXXXX	[69]	
VO107		*Digitaria sanguinalis*	1981	Texas, USA	XXXXXXXX	this study	

ND, no data. "References" lists studies that used the sequencing data, besides present study. Isolates Br116.5, Br118.2, TP2, MZ5-1-6, and Br35 sequenced by Inoue et al. [70], Bangladeshi isolates and isolates PY05002, PY06025, PY06047, PY25.1, PY35.3, PY05035 sequenced by Islam et al. [15], isolate SA05-144 sequenced by Yoshida et al. [14], and isolates DS9461 and DS0505 sequenced by Zhong et al. [56] were not included in the study.

Table 2. Summary of population genetic variation at 2682 single-copy orthologous genes in wheat, lolium, rice and setaria lineages of *Magnaporthe oryzae*

Lineage	n	S	K	H_e	θ_w	π	PHI test (p-value)
Wheat	20	5.8	1.9	0.17	1.28E-03	1.24E-03	0
Lolium	17	3.1	1.5	0.10	7.02E-04	6.54E-04	0
Rice	18	5.3	2.3	0.12	1.55E-03	7.75E-04	0
Setaria	8	2.6	1.8	0.18	9.10E-04	7.68E-04	0

Other lineages were not included in calculations because of too small a sample size (n<6); n is sample size; θ_w is Watterson's θ per bp; π is nucleotide diversity per bp; H_e is haplotype diversity; K is the number of haplotypes. PHI test is the Pairwise Homoplasy Test. The PHI test is implemented in SPLITSTREE. The null hypothesis of no recombination was tested, for the PHI test using random permutations of the positions of the SNPs based on the expectation that sites are exchangeable if there is no recombination

Figure legends

Figure 1. Discriminant Analysis of Principal Components, assuming K=10 clusters

Each isolate is represented by a thick vertical line divided into K segments that represent the isolate's estimated membership probabilities in the K=10 clusters (note that all isolates have high membership probabilities in a single cluster, hence only a single segment is visible). The host of origin of samples is shown below the barplot, and lineage IDs are shown above the barplot.

Figure 2. Maximum likelihood tree based on the concatenation of 2,682 orthologous coding sequences extracted from 76 *M. oryzae* genome. Nodes with bootstrap support >90% are indicated by dots (100 bootstrap replicates).

Figure 3. Total evidence neighbor-joining distance tree using pairwise distances (number of differences/kb) calculated from analysis of pairwise blast alignments between repeat-masked genomes. Only nodes with confidence > 80% (see methods) are labeled. Gray ovals are drawn around the main host-specialized populations for clarity.

Figure 4. Maximum likelihood tree based on concatenated dataset comprising nine loci used in Castroagudin et al. [24], **retrieved from 76 *M. oryzae* genomes.** Numbers above branches represent bootstrap supports after 100 bootstrap replicates. Only nodes with bootstrap support > 50 are labelled. Representatives of isolates used by Castroagudin et al. [24] in their study were included in the analysis and are colored in light grey. Green dots mark the strains containing the *Pgt*-type allele according to Castroagudin et al. [24].

Figure 5. ASTRAL analysis to test for incomplete lineage sorting/gene flow among 81 *Magnaporthe* genomes, using 2,241 single-copy orthologous sequence loci. Thicker

branches represent branches that have a bootstrap support > 50 after multilocus bootstrapping. Number above branches represent q1 local support (i.e. the proportion of quartet trees in individual genealogies that agree with the topology recovered by the ASTRAL analysis around the branch), with q1 values showed on black background for branches holding wheat blast isolates.

Figure 6. Neighbor-Net network built with SPLITSTREE. The figure shows relationships between haplotypes identified based on the full set of 25,078 SNPs identified in 2,682 single-copy orthologs, excluding sites missing data, gaps and singletons.

Supplementary Tables legends

Table S1. Pairwise distances measured in SNPs per megabase of uniquely aligned DNA

Table S2. (A) Gene flow signatures from ABBA/BABA tests. P1, P2, P3 refer to the three lineages used for the tests. The D statistic tests for an overrepresentation of ABBA versus BABA patterns. SE is the standard error. Z-score and P-value for the test of whether D differs significantly from zero, calculated using 1000 block jackknifes of 100 SNPs. Analyses were based on 354,848 biallelic SNPs identified in 2241 single-copy orthologous genes, with *M. grisea* as the outgroup. Brachiaria1: Bm88324. Brachiaria2: Bd8401. Eleusine1 and Eleusine2 are, respectively, the light orange and orange clusters in DAPC analysis. Boldface labels indicate pairs connected by gene flow, as indicated by the sign of D. Time period represents the time scale over which gene flow is measured, as described in Figure S6. **(B) Timing of gene flow.** For all pairs of lineages belonging to triplets that yielded D values significantly different from zero, the corresponding time period over which gene flow was measured (as defined in Figure S6) is indicated. Each pair belongs to multiple triplets, spanning different time periods, and the reported time period is therefore the consensus of the corresponding time scales. For instance, the pair (brachiaria2, eleusine1) was included in triplets measuring gene flow at both intermediate (time periods 1 and 2) and recent (time period 1) time scales, and the consensus time period is therefore "1+2".

Table S3. Summary statistics of genome assemblies used in this study.

Supplementary Figure legends

Figures S1A-S1J Maximum likelihood tree based on *MPG1, ACT1, b-tubulin1, BAC6, CAL, CH7BAC7, CH7BAC9, CHS, Ef1a* and *NUT1* marker, respectively. Trees are represented as unrooted cladograms. Dark branches represent branches with bootstrap support > 50 after 100 bootstrap replicates (corresponding support are indicated). Clades are labeled according to the convention used by Castroagudin et al. [24]. Green dots: representatives of Pgt (*Pyricularia graminis-tritici sp. nov.*). Red dots: representatives of Pot (*Pyricularia oryzae* pathotype Triticum) clade 1. Blue dots: representatives of Pot clade 2. Orange dots: representatives of Poo (*Pyricularia oryzae* pathotype Oryza).

Figure S2. Species tree inference based on the dataset of Castroagudin et al. [24] using ASTRAL. The tree is represented as an unrooted cladogram.

Multilocus bootstrap supports above 50 are indicates above branches. Dark branches represent branches with bootstrap support > 50 after 100 bootstrap replicates (corresponding support are indicated). Number in brackets are q1 local quartet supports (i.e. the proportion of quartet trees in individual genealogies that agree with the topology recovered by the ASTRAL analysis around the branch). Clades are labeled according to the convention used by Castroagudin et al. [24] as in S1-S10 Figures.

Figure S3. Analyses of population subdivision using clustering algorithms. (A) Bayesian Information Criterion vs number of clusters assumed in DAPC analysis. The Bayesian Information Criterion assesses the fit of models of population structure assuming

different K values. **(B) DAPC analysis of population subdivision, assuming K=2 to K=15 clusters.** Each isolate is represented by a thick vertical line divided into K segments that represent the isolate's estimated membership probabilities in the K clusters. The host of origin of samples is shown below the barplot. **(C) Log likelihood of data vs number of clusters assumed in STRUCTURE analysis.** Error bars are standard deviations of likelihood across STRUCTURE repeats. **(D) STRUCTURE analysis of population subdivision, assuming K=2 to K=15 clusters.** Each isolate is represented by a thick vertical line divided into K segments that represent the isolate's estimated membership proportions in the K clusters (note that two to seven clusters are empty, i.e. represented by no isolates, for models with K>9). The host of origin of samples is shown below the barplot.

Figure S4. Neighbor joining tree showing the genetic distance separating the *M. oryzae* strains from *M. grisea* and *M. pennisetigena.* Distances are in SNPs/kb.

Figure S5. Distribution of MPG1, BAC6, β-tubulin1, CAL, CH7BAC7, CH7BAC9, EF1α and NUT1 alleles among *M. oryzae* isolates as indicated by mapping onto the neighbor-joining tree built using whole genome SNP data. Alleles were identified by using a reference marker sequence for each gene, to search all the genomes using BLAST. Sequence variants are noted above each tree using the BLAST backtrace operations (BTOP) format.

Figure S6. Time periods for gene flow covered by different triplets of lineages in ABBA/BABA tests. Heterogeneity in divergence time between members of ((P1,P2),P3) triplets allows to examine gene flow at three time scales [33]: (A) triplets including the most recently diverged lineages as P1 and P2 (i.e. the *Triticum* and *Lolium* lineages, the two

Eleusine lineages, or the *Oryza* and *Setaria* lineages) carry information about gene flow across relatively recent times, (B) triplets including, as P1 and P2, two lineages from the same main group of lineages (i.e. *Eragrostis* / *Eleusine*1 / *Eleusine*2 / *Triticum* / *Lolium* or *Brachiaria2* / *Setaria* / *Oryza*, excluding (P1,P2) pairs already used in (A)) carry information about gene flow across intermediate times, and (C) triplets including, as P1 and P2, two lineages from different groups of lineages (i.e. *Eragrostis* / *Eleusine*1 / *Eleusine*2 / *Triticum* / *Lolium* and *Brachiaria2* / *Setaria* / *Oryza*) and *Stenotaphrum* or *Brachiaria1* as P3 carry information about gene flow across a relatively long time period. All three graphs correspond to hypothetical cases in which the D statistic that measures imbalance between ABBA and BABA types indicates gene flow between P2 and P3 (i.e. positive D values). In (A) and (B) multiple possible topologies are shown, as P1, P2 and P3 can either belong to the same group of lineages or to different groups of lineages.

Supplementary Text legends

Text S1. Probabilistic "chromosome painting" analyses.

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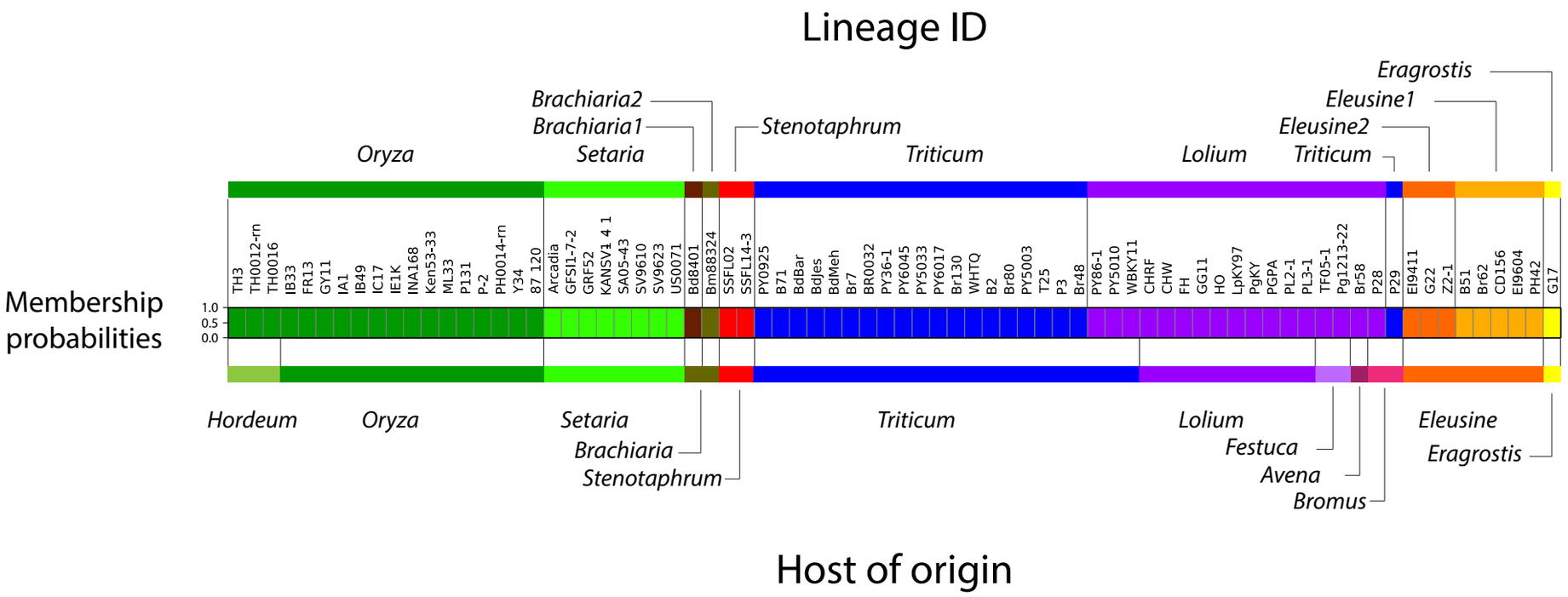
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Setaria
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Lolium
Triticum
TH3
TH0012-rn
TH0016
IB33
FR13
GY11
IA1
IB49
IC17
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Y34
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Arcadia
GFSIL-7-2
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EI9411
G22
Z2-1
B51
Br62
CD156
EI9604
PH42
G17
Membership probabilities
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0.5
0.0
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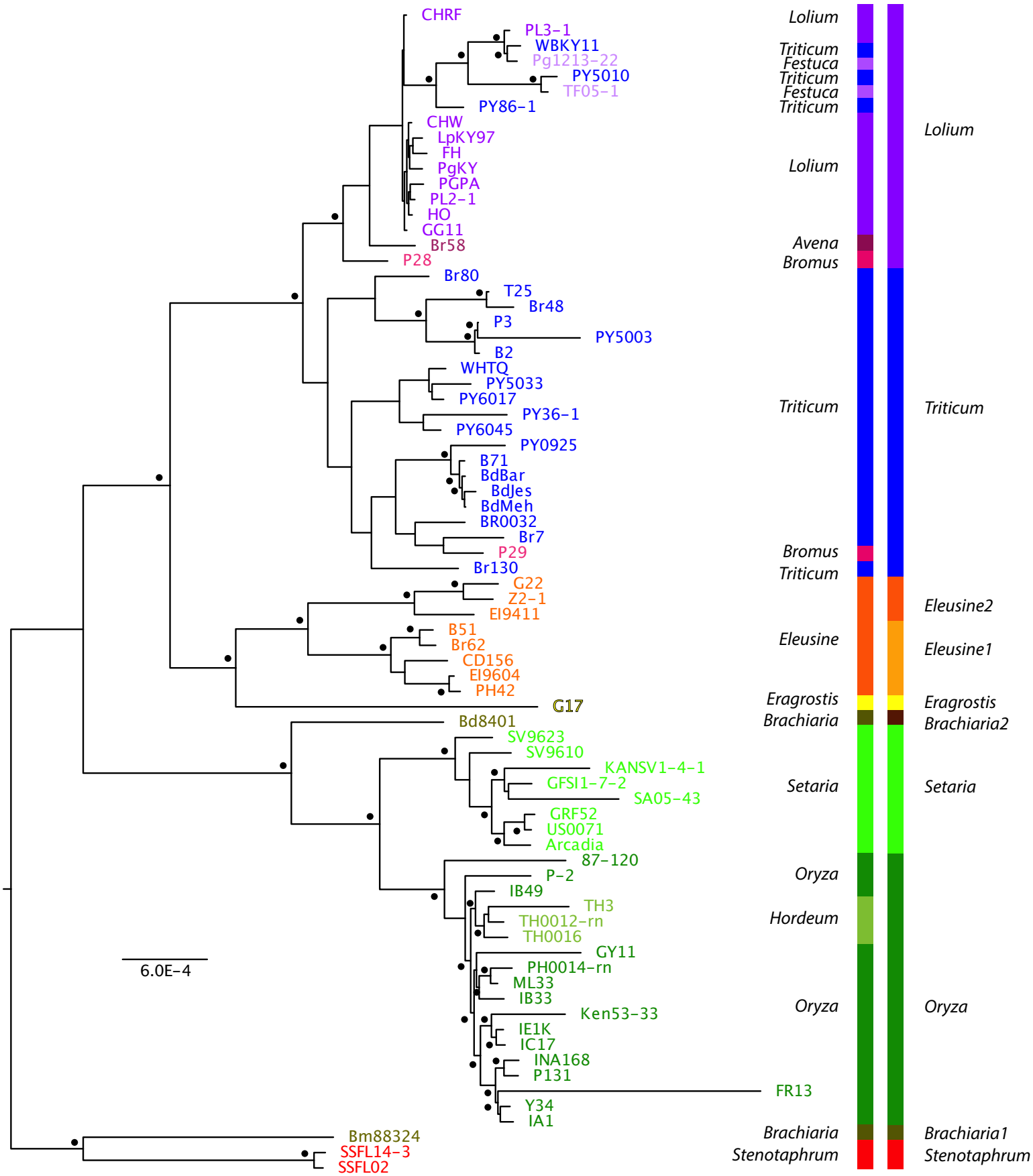
Lolium
Eleusine
Brachiaria
Stenotaphrum
Festuca
Avena
Bromus
Eragrostis
Host of origin


Host of origin
Lineages
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Br80
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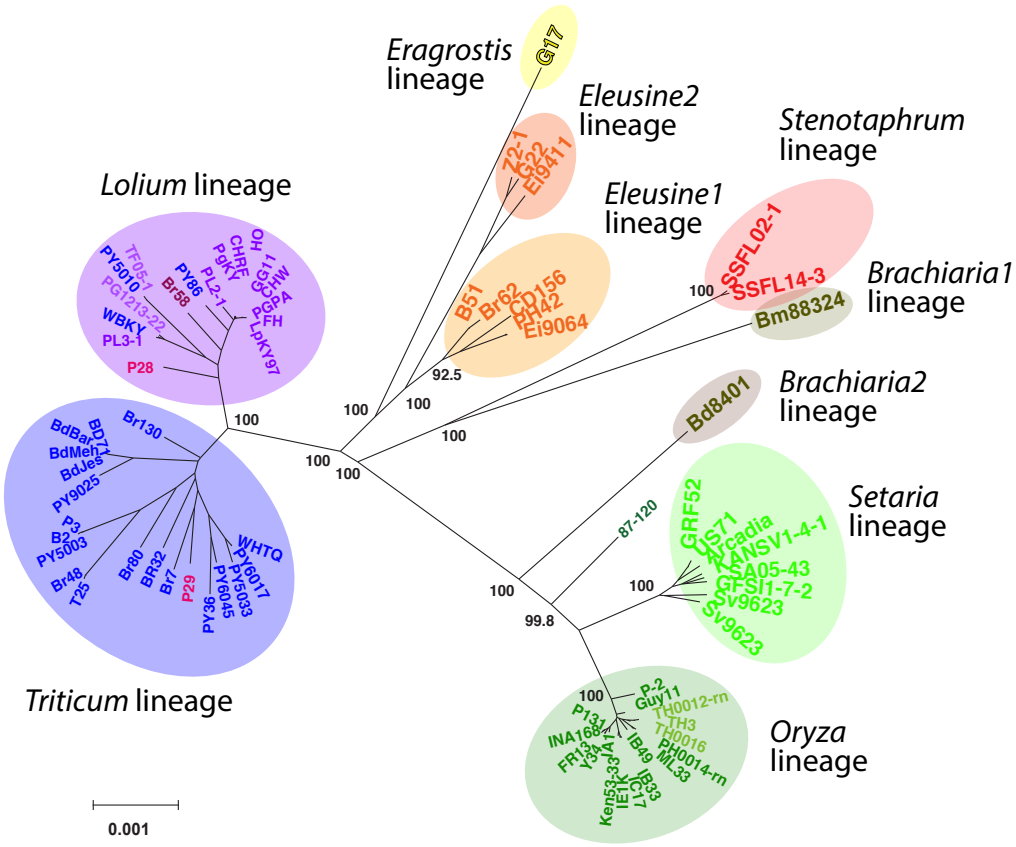
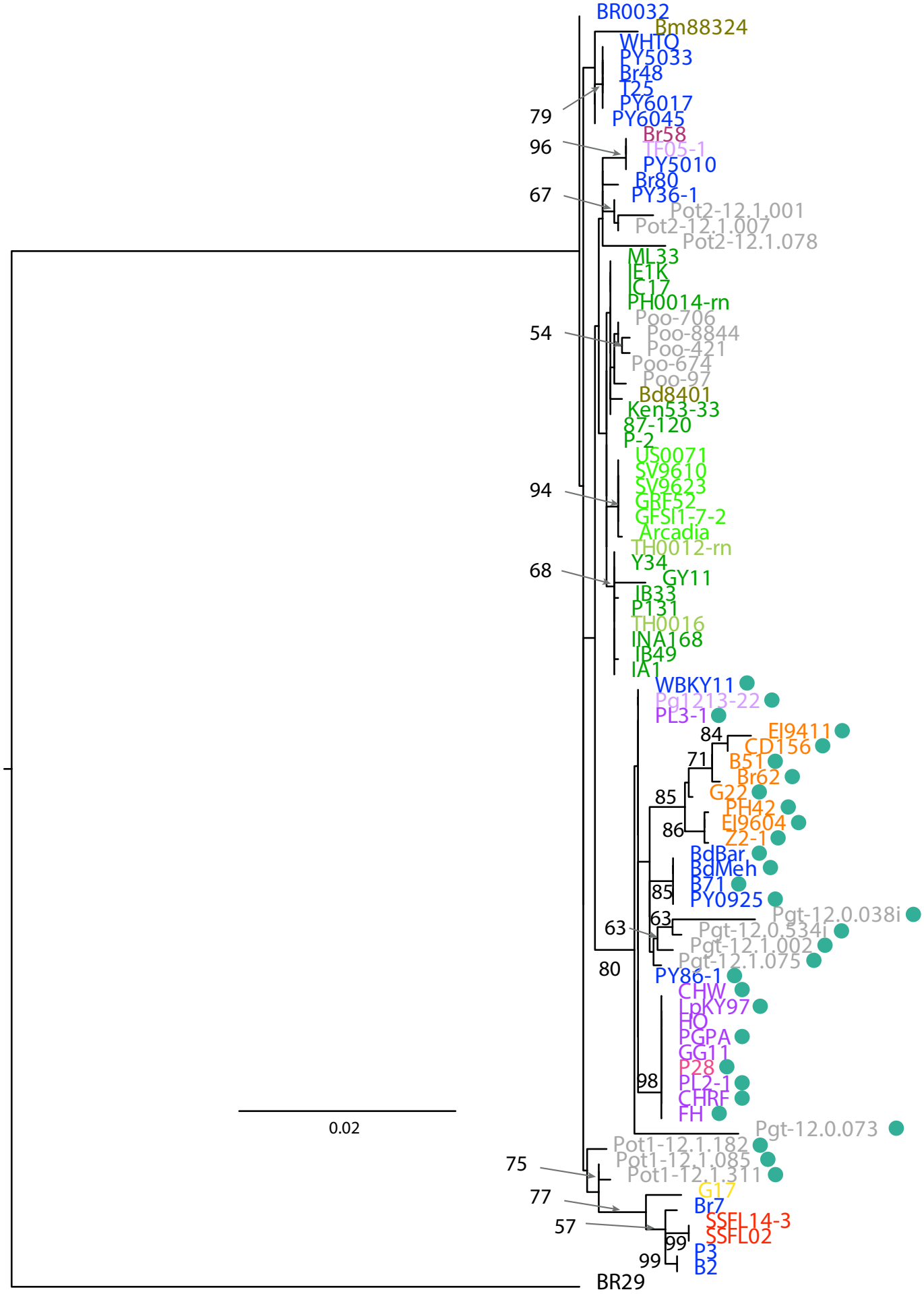
BdBar
BdJes
BdMeh
BR0032
Br7
P29
Br130
G22
Z2-1
EI9411
B51
Br62
CD156
EI9604
PH42
G17
Bd8401
SV9623
SV9610
KANSV1-4-1
GFSI1-7-2
SA05-43
GRF52
US0071
Arcadia
87-120
P-2
IB49
TH3
TH0012-rn
TH0016
GY11
PH0014-rn
ML33
IB33
Ken53-33
IE1K
IC17
INA168
P131
FR13
Y34
IA1
Bm88324
SSFL14-3
SSFL02
6.0E-4
Lolium
Triticum
Festuca
Triticum
Festuca
Triticum
Lolium
Lolium
Avena
Bromus
Triticum
Bromus
Triticum
Eleusine
Eragrostis
Brachiaria
Setaria
Oryza
Hordeum
Oryza
Brachiaria
Stenotaphrum
Lolium
Triticum
Eleusine2
Eleusine1
Eragrostis
Brachiaria2
Setaria
Oryza
Brachiaria1
Stenotaphrum



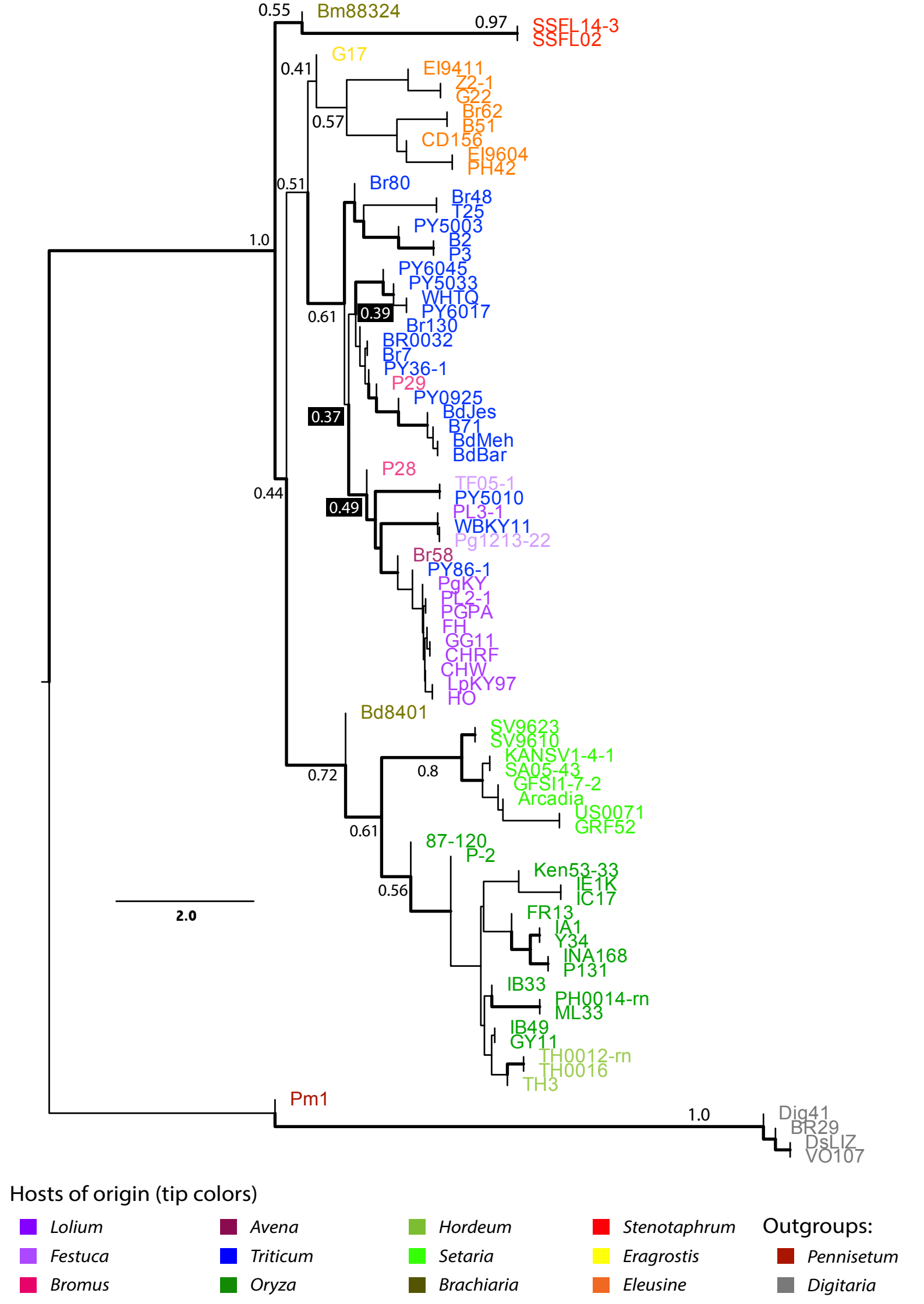
Hosts of origin (tip colors)

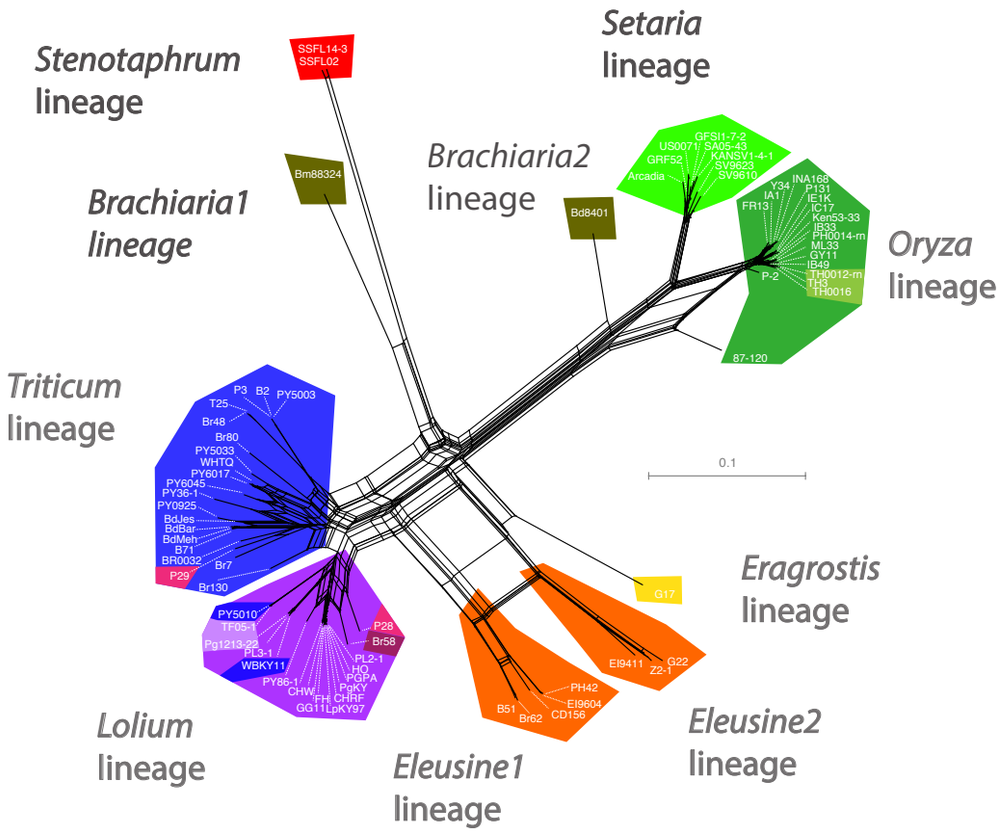
Lolium	*Avena*	*Hordeum*	*Stenotaphrum*
Festuca	*Triticum*	*Setaria*	*Eragrostis*
Bromus	*Oryza*	*Brachiaria*	*Eleusine*



Hosts of origin (tip colors; samples from Castroagudin et al. in grey)

- *Lolium*
- *Festuca*
- *Bromus*
- *Avena*
- *Triticum*
- *Oryza*
- *Hordeum*
- *Setaria*
- *Brachiaria*
- *Stenotaphrum*
- *Eragrostis*
- *Eleusine*


0.55
Bm88324
0.97
SSFL14-3
SSFL02
G17
0.41
EI9411
Z2-1
G22
Br62
B51
0.57
CD156
EI9604
PH42
0.51
Br80
Br48
T25
PY5003
B2
P3
1.0
PY6045
PY5033
WHTQ
PY6017
0.61
0.39
Br130
BR0032
Br7
PY36-1
P29
PY0925
BdJes
B71
BdMeh
BdBar
0.37
P28
TF05-1
PY5010
PL3-1
WBKY11
Pg1213-22
0.44
0.49
Br58
PY86-1
PgKY
PL2-1
PGPA
FH
GG11
CHRF
CHW
LpKY97
HO
Bd8401
SV9623
SV9610
KANSV1-4-1
SA05-43
GFSI1-7-2
Arcadia
US0071
GRF52
0.72
0.8
0.61
87-120
P-2
Ken53-33
IE1K
IC17
0.56
FR13
IA1
Y34
INA168
P131
IB33
PH0014-rn
ML33
IB49
GY11
TH0012-rn
TH0016
TH3
2.0
Pm1
1.0
Dig41
BR29
DsLIZ
VO107
Hosts of origin (tip colors)
Lolium
Avena
Hordeum
Stenotaphrum
Outgroups:
Festuca
Triticum
Setaria
Eragrostis
Pennisetum
Bromus
Oryza
Brachiaria
Eleusine
Digitaria



Hosts of origin (color of shaded areas)

Lolium	*Avena*	*Hordeum*	*Stenotaphrum*
Festuca	*Triticum*	*Setaria*	*Eragrostis*
Bromus	*Oryza*	*Brachiaria*	*Eleusine*