Curriculum

The CURE for Cultivating Fastidious Microbes

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ABSTRACT

Course-based Undergraduate Research Experiences (CUREs) expand scientific educational benefits of research to large groups of students through a course setting. As part of an ongoing effort to integrate CUREs into freshman biology, we developed a microbiology CURE (mCURE) that uses a modified version of a dilution-to-extinction high throughput culturing protocol for isolating abundant yet fastidious aquatic bacterioplankton. Students learn to serially dilute and inoculate microbial cultures, perform DNA extractions and PCR, and identify microbial isolates via 16S rRNA gene sequences. The curriculum also includes exercises for learning to critically read and evaluate scientific literature, and emphasizes scientific communication through written and oral exercises that incorporate social media elements. In the first three semesters, the mCUREs achieved similar cultivability success as implementation of the protocol in a standard laboratory setting. The mCURE students have cultivated 43 unique bacterioplankton taxa, many of which occur as abundant taxa in the coastal environments from which the initial inoculum was sampled. Moreover, trends observed in the pre- and post-course survey data provide preliminary evidence that the mCURE format may improve a variety of scientific training objectives, based on the students' perceptions of the course. Our modular framework facilitates customization of the curriculum for use in multiple settings, and we provide classroom exercises, assignments, assessment tools, and examples of student output to assist with implementation.

INTRODUCTION

Undergraduate research experiences in STEM increase student retention in science majors; increase the proportion of students that go on to professional or graduate school; as well as improve critical thinking skills, data interpretation skills, content knowledge, and attitudes toward science (1-5). Typically, undergraduate research experiences are limited to relatively few students due to research lab size and funding, making these positions competitive, highly selective, and typically dominated by upperclassmen (4, 5). Course-based undergraduate research experiences (CUREs), in which students experience research as part of a course, can reach students early in their degree program and accommodate large numbers of students, thus increasing the diversity of students participating in research (4, 5). Despite these benefits, the time necessary to plan CURE projects and create assignments and rubrics can restrict their use (6). Fortunately, an increasing number of publications have shared CURE implementation strategies for a variety of settings (3, 7-9).

We recently outlined a flexible, modular CURE framework, including rubrics and course materials, that has facilitated conducting a variety of different research projects in the freshmen biology laboratory courses at Louisiana State University (LSU) (10). Using this framework, we have developed the microbiology CURE (mCURE) described herein that focuses on the cultivation of bacterioplankton from aquatic systems (**Figure 1**). Students execute a modified version of the protocol utilized by the Thrash Laboratory at LSU (11, 12) incorporating lessons on pipetting, serial dilution, sterile technique, DNA extraction, PCR, and basic sequence identification using BLAST. In addition, the mCURE emphasizes enhancing student skills in the areas of reading primary literature, communicating with a variety of audiences, and experimental design. The possibility of isolating new organisms provides a charismatic entry point into biological research and students experience a genuine excitement of discovery along with their laboratory and communication training.

Bacterioplankton occupy marine and freshwater environments at cell concentrations typically between 10^5-10^7 cells mL^{-1}. Within these mixed populations, taxa occupy a spectrum of frequencies, with some groups representing 10-20% of the total number of cells, and others occurring in much lower abundance. Regardless of their abundance, cultivation of many taxa from marine and fresh water has not been possible. Methods using traditional agar plating have only successfully cultivated 0.1-1% of the organisms present in a given sample (13), hampering our ability to understand the functions of a large majority of microorganisms. This Great Plate Count Anomaly (13) has led to the development of more sophisticated cultivation methods. One of these combined serial dilution of

samples with sterilized natural water as a medium (14). Many of the most abundant taxa in marine and freshwater systems have been successfully cultured using this approach. Examples included SAR11 *Alphaproteobacteria* (15-17), SUP05 *Gammaproteobacteria* (18), SAR116 *Alphaproteobacteria* (19, 20), and members of the so-called "Oligotrophic Marine *Gammaproteobacteria*" (21). Recently, a modification of these protocols demonstrated considerable success in cultivating many of the most abundant microorganisms in coastal systems using a set of artificial media (12). This advance was important because unlike sterilized natural water media, which requires access to and shipping of large volumes of water from environmental sources, artificial media can be made anywhere and easily modified (e.g., in salinity, carbon and nitrogen sources, etc.) to accommodate different environments. This advance also facilitated the adaptation of the protocol in the classroom laboratory.

Learning time

We designed the mCURE for a semester timeline, with each single three-hour laboratory section meeting once a week for a minimum of 13 weeks. The project is divided into four major segments (color-coded in both **Figure 1** and **Table 1**). In weeks #2-4 (***orange***), students attempt to establish an initial culture of marine bacterioplankton using serial dilutions with the High-Throughput Culturing protocol (HTC; (11)). Transfer of the initial cultures to larger flasks for further growth occurs during weeks #5-6 (***green***). During weeks #7-9 (***blue***), students extract DNA from the successful cultures and amplify the 16S rRNA genes with PCR. The amplified products are then sequenced out of class for subsequent taxonomic identification of the microbe on week #10 (***yellow***). The remaining weeks (#11-13) are spent discussing poster construction and administering the final assessments for the semester. Note that the entire workflow does not require 13 weeks, but we have built in flexibility to allow for some elements of the protocol to be repeated in case of a failure in one or more of them (see Faculty Instructions below).

Intended audience and prerequisite student knowledge

This course has been offered in sections of freshman biology laboratory; no prerequisite high school courses are required. Students receive training in many of the basic biology skills that they will utilize in other contexts, both at the bench and in communication of their results. Given the success of the course with untrained freshman students, we anticipate that this course could be modified for upper class high school laboratories as well. We provide

suggestions for protocol modifications that could facilitate the use of this curriculum, albeit with likely variations in the types of organisms cultivated, in settings for which the type of equipment we utilize is unavailable.

Learning objectives

By the end of the semester, students should be able to:

1. Describe the relationship between the research objectives, the high-throughput culturing approach, and the experimental design;
2. Find, read, and interpret relevant articles from the primary literature;
3. Work in groups to isolate, culture, and identify bacterioplankton from aquatic environments;
4. Communicate the methods, results and implications of their research to both scientific and non-scientific audiences using posters, writing assignments, and social media.

In addition to the learning objectives outlined above, the format of the mCURE sections incorporated aspects of three high-impact practices: Undergraduate Research, Collaborative Assignments, and Writing Intensive (22).

PROCEDURE

A summary of basic approach for the CURE is shown in **Figure 1**, along with a week-by-week breakdown of activities in **Table 1**. Table 1 also includes a summary of all class materials made available with this manuscript, as well as prep notes for faculty.

Materials

The required equipment and chemicals are the same as those previously published (11). Briefly, because most of the highly abundant aquatic microorganisms have oligotrophic lifestyles, occur in low cell densities (< 10^7 cells/mL), are very small (< 1μm), and will not grow on solid media, the cultivation approach makes use of liquid media and cell growth is measured using a benchtop flow cytometer (Millipore "Guava"). Alternative media, including detailed recipes and preparation instructions were previously published (12). To avoid trace-metal contamination, all reusable cultivation vessels should be made of polycarbonate plastic. Other major items include a thermocycler and PCR reagents, electrophoresis equipment and a gel viewing system (e.g., Bio-Rad Gel Doc), a DNA quantification

system (e.g. Qubit, ThermoFisher), DNA extraction kits (MoBio PowerWater), and pipettes/tips. The only exceptions to the established protocol equipment (11) in the mCURE sections are a portable laminar flow hood (AirClean Systems or COY Laboratory Products) instead of biosafety cabinet, and disposable 2.1 mL 96-well plates (Thermo Nunc A/S) instead of those made from PTFE. For those without access to some or most of this equipment, we provide alternatives in the Discussion that can improve implementation flexibility.

Student instructions

Segment 1 (*orange* in **Figure 1, Table 1**): In first two weeks of class, students are introduced to the overall mCURE approach, and coached on how to use pipets and follow sterile technique. Each group of two students (a total of 12 per section) then dilutes and inoculates seven wells of a 96-well plate, containing low-nutrient medium, with the diluted seawater (**Appendix 1**). An eighth well is inoculated with sterile media as a contamination control. Thus, each section begins cultures in a 96-well plate, which is incubated at *in situ* temperature (depending on source water) for 2-3 weeks, prior to being checked for growth using flow cytometry. During the incubation weeks, student assignments focus on introducing effective reading of scientific literature, and experimental design and rationale.

Segment 2 (*green*): Each group selects 1-2 culture wells from those that were deemed positive via flow cytometry (growth over 10^4 cells/mL) for transfers into larger volume growth flasks, and creates cryostocks for culture preservation in 10% DMSO (**Appendix 2**). In our experience, most groups were able to perform at least one transfer. The 2-3 groups (out of 12) that had no growth in any of their wells select an "extra" well from another group (one that group isn't transferring). These flasks are then allowed to incubate for two weeks at the same temperature as the initial cultivation plates. During the interim, students are introduced to scientific writing and give "lightning talks" (see footnote #2 in **Table 1**).

Segment 3 (*blue*): Groups select at least one flask that shows growth and extracts DNA from the culture (**Appendix 3**). In the last three semesters of conducting this mCURE, the majority of groups observed growth in at least one flask. The remaining 2-3 groups with no growth in any of their flasks use part of another group's culture for extraction. Note this introduces redundancy in the final identification results. Over the next two weeks, they amplify

the 16S rRNA genes from their isolates using PCR and confirm the amplification product with gel electrophoresis. Successful amplicons are then sequenced off campus at a sequencing facility (e.g., the RTSF Genomics Core at Michigan State University or similar).

Segment 4 (*yellow*): Students learn to assemble forward and reverse (via the reverse complement) PCR amplification sequence reads into a contig and identify their isolate sequences using NCBI BLASTN portal (**Appendix 4**). Briefly, reads from both the forward and reverse primer, as well as the contig (if any), are searched against the GenBank nt database with and without the exclusion of uncultured/environmental samples. The % ID, Query coverage, E-value, and GenBank # for the top five BLAST hits (from all the runs) are recorded for all isolates. Interpretation and contextualization of the results, including the similarity of isolates generated by the students to those in the database, occurs via discussion with knowledgeable faculty/teaching assistants (TAs). These results become part of the final data presented via posters.

Faculty instructions

Segment 1 (*grey*, *orange* in **Figure 1, Table 1**): Prior to the beginning of the course, instructors must prepare the following:

1) Collect seawater and measure the concentration of bacterioplankton using flow cytometry. The students use this initial concentration to calculate the dilution factor required to inoculate ~1-5 cells per well. Collection should occur as proximately to inoculation as possible to avoid microbial community change via bottle effects.
2) Prepare the low-nutrient media (12) required to grow the microbes. This can be stored for a few days at 4˚C if necessary. Keep wrapped in foil to reduce photodegradation of the vitamins. Aliquot ~1.7 mL of media into each well of the 96-well plate just before class and allow time for equilibration to incubation temperature.
3) Select ~12-15 scientific articles relevant to the project and create a reading guide for one of them for class discussion (sample: **Appendix 5** for Connon & Giovannoni, 2002). The students may select one of the

remaining papers for their lightning talks (Table 1, weeks #4-5), and use any number of them as references for their formal writing assignments.

Because of the considerably long incubation period (2-3 weeks) for the initial inoculations, we recommend that this step involve at least one "holiday week" for the lab (see **Supplemental Table 1**). At the end of the incubation period, instructors perform flow cytometry to check for growth in the 96-well plate and record the well numbers positive for growth.

Segment 2 (*green*): Prior to the start of this segment, instructors must prepare more of the low-nutrient medium and aliquot 50 mL into flasks, as well as prepare cryotubes with DMSO. At the end of the 2-week incubation, instructors perform flow cytometry again to check the flasks showing growth, and record cell concentrations for student use. For the scientific writing discussion, we have made an activity available (**Appendix 6**) that familiarizes students with the content in various sections of a paper (19).

Segment 3 (*blue*): We recommend that instructors aliquot the required amount of DNA extraction reagents (Power Water DNA Isolation Kit; Mo Bio Laboratories) for each group of students to prevent cross-contamination. For PCR, we propose that the instructor create a master mix with all PCR components (detailed in **Appendix 9**) as part of a class demo, then have students add 1 µL of the isolated DNA to 49 µL master mix in a 50 µL PCR reaction. For the gels, we suggest melting an appropriate amount of agarose in a flask for each section, and allowing it to solidify until class time. Then, prior to the start of class, the instructor can melt the agarose in the flask and have it ready for students to pour their own gels.

Segment 4 (*yellow*): Before the BLAST lab, instructors need to have all successful 16S rRNA gene amplicons sequenced, using both forward and reverse primers (we use 27F and 1492R, but this can be specified by the instructor- see (23) for additional options), and the data available where the students can access it. Each sequencing read needs to be labeled with the sample number and whether it is a "forward" or a "reverse" sequencing read. We recommend the "BLAST behind the scenes" activity (**Appendix 7**) to introduce students to the concept of sequence analysis.

Finally, instructors need to prepare for a departmental poster session at the end of the semester. Therefore, the space for poster boards, display tables, and printing facilities need to be organized. However, as far as the graded poster presentation is concerned, we recommend that the student groups present their posters electronically in class. During this time, other students and the instructor can offer constructive criticism for the students to incorporate into the final printed version of the poster.

Based on our experience performing this mCURE for the past three semesters, we anticipate at least 1-2 protocol failures per semester; hence, flexibility is built into the framework (**Table 1**, **Supplemental Table 1**). Despite our anticipation of some failures and correcting these in subsequent semesters (e.g., students failing to properly transfer and freeze their samples), each new semester has presented us with new and different failures (e.g., flow cytometer reagents on back-order, failed PCRs due to old reagents). Many non-experimental activities, such as the lightning talks, can be easily performed during different time points in the course, amended to take less time, or even completely eliminated. Similarly, other related activities may be added, such as peer review of initial formal writing drafts and using social media for science outreach (*e.g.* we use the Twitter hashtag #LSUCURE for all CURE efforts in the Department of Biological Sciences at LSU; **Supplemental Table 1**). If feasible, we recommend adding the following enhancements to further engage students in the course: (i) taking students on a field trip, such as a one-day research cruise to collect water samples; (ii) demonstrating the use of "behind the scenes" equipment, such as the flow cytometer, capillary sequencer, and/or modern microscopes used to image bacteria.

Suggestions for determining student learning

We used quizzes throughout the semester to gauge student learning, along with a set of written assignments (**Table 1**). While the quizzes and final exam were intended to test the students' knowledge about the course materials at various levels of Bloom's taxonomy, the written and oral assignments were meant to hone their communication skills. The students received instructor feedback for each informal and formal draft of their papers before submitting the next one (**Appendices 8-11**). Finally, the course has a cumulative final exam (**Appendix 12**) and a poster presentation by the student groups.

In addition to the course materials outlined above, we used pre/post surveys to assess self-evaluated student research efficacy and perceptions of the course. The survey instrument consisted of 21 questions measuring student attitudes

toward science and biology labs (written by EWW and collaborators; 5 point Likert scale), twelve questions from Lopatto's (24) instrument (5 point Likert scale), 27 questions from the Clinical Research Appraisal Inventory (CRAI; 11 point Likert scale) (25), and demographic questions (**Appendix 13**). These surveys were emailed to students in all research and traditional lab sections during the first week of the semester and again during the last week of the semester. Students were allowed two weeks to complete the surveys; two reminder emails were sent to students that had not yet completed the survey. Pre- and post-survey data were paired. We determined significance in the pre/post semester differential (post response minus pre response) between research and traditional students using the Mann-Whitney *U*-test implemented in R (26). Here we present a preliminary analysis of the data; we will present a more thorough analysis of research and traditional lab student outcomes, perceptions, and self-efficacy for all CUREs in our department in a future contribution.

Sample data

Examples of formal writing assignment 1, final lab report, Lightning Talk, and student posters are included in **Appendix 14** and are shared with permission from the students.

Safety issues

Aquatic microbes that grow in oligotrophic media are not usually pathogenic to humans. Hence, there are only a few minor safety issues to consider.

1) Wear gloves and eye protection to protect from chemicals. Gloves and lab coats also help prevent introduction of contaminating skin microbiota into the cultures and are therefore should also be worn throughout the protocol.
2) If using a Bunsen burner as an alternative to the portable laminar flow hood (see *Possible modifications* below), tie long hair back to protect from potential fire-related accidents.

Additional safety measures must be taken for faculty during washing and preparation of medium mixture bottles and growth flasks. See (11) for more details.

DISCUSSION

Field testing

Thus far we have offered the mCURE during the fall 2015 and 2016 semesters in Biology 1207 (Honors: Biology Laboratory for Science Majors) and spring 2016 in Biology 1208 (Biology Laboratory for Science Majors I). There have been four sections per semester taught by two graduate TAs (two sections per TA), with up to 28 students per section. Biology 1207 is only offered in the fall semester and consists of a total of four sections. Multiple (12-50) sections of Biology 1208 are offered every semester, a few of which are typically offered as CUREs as outlined in our previous publication (10); students do not know when they register for this course if their section will be in a CURE or traditional format. We have plans to continue offering this mCURE at LSU for the foreseeable future.

Experimental results

Fall 2015, spring 2016, and fall 2016 average cultivability (14) was 9.9, 2.8, and 12%, respectively. These cultivability numbers generally match the success rate of other HTC experiments (12), and demonstrate a significant improvement over plate-based methods (13). The number of unique pure cultures that survived successive transfers and were positively identified at the end of each course was 28 (fall 2015), 13 (spring 2016), and 23 (fall 2016). In total, mCURE sections isolated 43 unique bacterioplankton during the first three semesters. Some courses isolated taxa identified in a previous mCURE, so the overall total was smaller than the sum of the individual semesters. Many of these have close relationships to organisms previously isolated using HTC techniques in the Thrash lab and other labs, as indicated by taxonomic affiliations to strains with “LSUCC”, “HTCC”, “HIMB”, or “IMCC” strain numbers (**Table 3**). Importantly, many isolates represent abundant marine clades (12), and thus the results validate the mCURE approach to produce valuable cultures with similar efficacy as HTC experiments conducted under more typical laboratory settings.

Our curriculum development also provided anecdotal information that generated testable hypotheses about cultivation conditions. During the first semester (fall 2015), the mCURE protocol utilized glass tubes instead of the Thermo plates indicated above. This semester was also the only time we isolated numerous *Vibrio* spp., which are rarely cultivated in HTC experiments with the same media and inoculated from the same source waters (12). Although the lack of cultivation-independent data precludes us from knowing if the particular sample had a bloom of *Vibrio* spp., thus increasing the probability of their isolation, the results of this experiment present the possibility

that glass preferentially selects for certain bacterioplankton. The finding corroborates our understanding of *Vibrio* spp. as commonly having a surface-associated lifestyle (27). More importantly, the hypothesis that cultivation vessel material selects for specific taxa can be tested experimentally in the mCURE setting, thus adding value to the course design. During the fall 2016 semester, mCURE sections also experimented with using media of two different salinities for the same inoculum. Sample site salinity was 17. Students inoculated cultures in our MHW2 and MHW3 media with salinities of 23.2 and 11.6, and achieved 8.3 and 15.7% cultivability, respectively. Although these results are inconclusive due to variability in student success with downstream protocol elements, future experiments of this kind will provide additional data points and eventually strengthen the relationship between media salinity, cultivability, and taxonomic identity of the isolates. Other examples of comparative cultivation experiments easily facilitated with multiple concurrent mCURE sections include testing alternative carbon substrates and alternative incubation temperatures while using the same inoculation source. Ideally, cultivation independent data like 16S rRNA gene amplicons would also be collected to evaluate cultivation success (e.g., see (12)), but this requires additional cost and analysis expertise.

Evidence of student learning

Sample size and demographics of survey respondents who completed both pre- and post-surveys are outlined in **Table 2**. We should note that these survey results reflect self-reported student perceptions of their confidence and skills and do not necessarily reflect actual changes in student skills or competencies. Most research (mCURE) students were freshmen with a few sophomores, whereas respondents in the traditional 1208 labs included students at all undergraduate academic levels (**Table 2**). Despite these differences, mCURE students were demographically similar to traditional lab students. In general, we found that students in both the traditional lab sections and research lab sections had decreased confidence in their abilities at the end of the semester. However, we observed a non-significant trend whereby research students lost confidence less and had less negative perceptions of the lab relative to traditional sections (**Figure 2** and **Appendix 15**). These results are largely consistent with other similar studies comparing freshmen level CUREs to traditional labs (3). Although all students in freshmen level biology labs at our institution work in groups, we found that CURE students had significantly more positive (or at least less negative) perceptions of collaboration and in their ability to work independently compared to traditional students (**Figure 2**).

In addition, research students increased confidence in their writing skills more than their traditional lab counterparts (**Figure 2**).

Possible modifications

We appreciate that many instructors may wish to implement the mCURE design but do not have access to some of the more expensive equipment used in our protocol. Thus, we mention a few modifications to circumvent some of these restrictions. Bunsen burners can serve in place of a biosafety cabinet or laminar flow hood. Keep in mind this also introduces additional safety considerations (discussed in *Safety issues* above). Although a flow-cytometer works best for rapid counting of small bacterioplankton cells at typical low growth densities, instructors can replace flow cytometry with direct microscopic counts, e.g., as in some of the earlier iterations of the HTC protocol (19). For those without access to either a flow cytometer or a fluorescence microscope, the protocol can still be completed using traditional agar-plate based methods. Our media can be prepared with agar (11), or a replaced with a classic marine medium like Difco 2216 (BD). Although solid media generally select for different taxa than liquid media, for the purposes of a basic biology laboratory, this may not matter. After streaking a seawater sample on plates, individual colonies can be picked, grown up in liquid culture to increase cellular mass, or directly processed through DNA extraction. Colony PCR (28) may also be an attractive alternative identification method, particularly because this also eliminates the time and cost associated with DNA extraction. These last two steps may also help adapt the overall protocol for shorter time frames, e.g., academic quarters instead of semesters. Finally, for those interested in freshwater environments, the same protocol can be conducted with freshwater media, either artificial (29) or natural (30).

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409 **Table 1.** The mCURE framework. Activities, associated assessments, faculty instructions, and the relevant
410 supporting documents are detailed week-by-week. The various segments of the course are color coded (grey, orange,
411 green, blue, and yellow), consistently with the flowchart in Figure 1.
412

Week	Topic	Quiz	Quiz Topic	In-class (Group) Activity	Assignments & Reminders	Instructor Prep Notes	Supporting Documents	Other Supporting Documents [a]
#1	Meet & greet; Basic intro to research question				Research paper (#1) reading assigned	Collect seawater & measure conc.; prep media and aliquot into 96-well plate	Course outline flowchart (Fig. 1)	
#2	Expt. design; pipetting & sterile technique; Scientific record-keeping			Research paper (#1) introduced; Dilute seawater & inoculate 96-well plates		Incubate cultures under optimum conditions for 2-3 weeks.	Inoculation protocol; Reading guide	
#3	How to read scientific literature	1	Dilutions, pipetting & sterile technique	Decode & understand main points in a scientific paper; Other research papers introduced as part of in-class activity	HW based on research paper (#1) assigned[1]			How to Read a Scientific Paper
#4	Experimental design	2	Decoding scientific literature	Interactive discussion re: experimental design (compare & contrast methodology with paper #1)	HW based on research paper (#1) due; Student groups choose one from a pool of papers for the "Lightening talks" [2]	Perform flow-cytometry to determine positive cultures[3]; prep media in flasks; prep cryostocks		
#5	Transfer & back-up; Discuss importance of back-ups; "Lightening talks" [2]	3	Expt. design	Transfer isolates to larger volume; prepare cryostocks	Formal Writing #1 assigned; Informal Writing #1 assigned	Incubate flasks under optimum conditions for ~2 weeks	Formal Writing #1 guide & rubric; Informal Writing #1 guide & rubric	Writing Rubric
#6	Scientific Writing	4	Expt. design (contd.) – focus on transfers & back-ups	Order the sections of a scientific paper; Evaluate excerpts of scientific papers based on guide & rubric	Informal Writing #1 due	Perform flow-cytometry to determine positive cultures[3]; Prep aliquots of DNA extraction reagents	Order a scientific paper	Scientific Writing
#7	DNA extraction			Perform DNA extraction of isolates	Feedback on Informal Writing #1 returned	Design & purchase 16S rDNA primers; aliquot PCR reagents		
#8	PCR; Primer choice & design	5	DNA extraction	Perform PCR of isolates	Formal Writing #1 due	Prep agarose gels		

#9	Gel electrophoresis & Sanger sequencing	6	PCR & primer design	Perform gel electrophoresis to confirm PCR products	Formal Writing #2 assigned; Informal Writing #2 assigned; Feedback on Formal Writing #1 returned	Perform Sanger Sequencing on positive samples	Formal Writing #2 guide & rubric; Informal Writing #2 guide & rubric	
#10	BLAST sequences & identify microbes	7	Electrophoresis & sequencing	Manually compare a set of sequences to identify the most closely related organisms; Identify the cultured microbes	Informal Writing #2 due; HW on poster critique assigned [4]		BLAST: How-to guide; BLAST behind the scenes	Poster Critique
#11	Elements of poster design; Poster development & critique	8	BLAST	Design rough drafts of posters; Peer poster critique session	HW on poster critique due; Feedback on Informal Writing #2 returned			Designing Scientific Posters
#12	Final Exam				Formal Writing #2 due			
#13	Poster Presentations; Peer Evaluation & Reflections							Poster Rubric; Peer Evaluation

[a] Available as Supplemental Materials from Bakshi, A., *et al*., *A Highly Scalable General Framework for Implementing Course-based Undergraduate Research Experiences (CUREs) in Freshman Biology Labs.* American Biology Teacher, 2016. **78**(6): p. 1-7.

[1] Basic questions to engage students in background information and the major take-home points from the research article

[2] Students give 5-minute presentations on a relevant research article of their choice from a pool of papers made available by the instructor (these papers are to be then used in the future as references in Formal Writings)

[3] Students are encouraged to make an appointment with the instructor to observe how the flow cytometer works.

[4] Evaluate publicly displayed posters within the department for clarity and style; designed to familiarize students with various poster designs

Abbreviations: HW – Homework; Expt. = Experimental; Prep = Prepare (for student use); Conc. = concentration

Table 2. Demographic data (percentages) for research (mCURE) lab and traditional lab survey respondents.

		Research (n=53)	Traditional (n=287)
Gender	Female	75.5	79.4
	Male	24.5	20.2
Class standing	Freshman	90.5	88.1
	Sophomore	9.5	6.6
	Junior	0	2.8
	Senior	0	1.4
	Other	0	0.7
Ethnicity	White	81.1	81.2
	Other	18.9	18.8

Table 3. Bacteria cultured by mCURE students

Closest unique cultured relative	**Major Taxonomic Group**
Arthrobacter sp. 210_2	Actinomycetales; Actinobacteria
Marinobacterium sp. IMCC1424	Actinomycetales; Actinobacteria
Microbacterium esteraromaticum strain V45.13	Actinomycetales; Actinobacteria
Microbacterium sp. Ni17	Actinomycetales; Actinobacteria
Nocardioides exalbidus strain DS1-2B	Actinomycetales; Actinobacteria
Nocardioides hwasunensis strain XH199	Actinomycetales; Actinobacteria
Alteromonadales bacterium 3tb13	Alteromonadales; Gammaproteobacteria
Alteromonas macleodii	Alteromonadales; Gammaproteobacteria
Alteromonas tagae	Alteromonadales; Gammaproteobacteria
Marinomonas sp. SS8	Alteromonadales; Gammaproteobacteria
Porticoccus hydrocarbonoclasticus	Alteromonadales; Gammaproteobacteria
Pseudoalteromonas phenolica	Alteromonadales; Gammaproteobacteria
Pseudoalteromonas sp. A-3	Alteromonadales; Gammaproteobacteria
Shewanella sp. 49WBP	Alteromonadales; Gammaproteobacteria
Bacillus sp. L1(2012)	Bacillales; Firmicutes
Burkholderiales bacterium LSUCC0118	Burkholderiales; Betaproteobacteria
Limnobacter sp. MYOU6	Burkholderiales; Betaproteobacteria
Halieaceae bacterium LSUCC0247	Halieaceae; Gammaproteobacteria
Gamma proteobacterium SF293	OM182; Gammaproteobacteria
Gamma proteobacterium IMCC15037	OM252; Gammaproteobacteria
Gammaproteobacteria bacterium LSUCC0258	OM252; Gammaproteobacteria

Gammaproteobacteria bacterium LSUCC0272	OM252; Gammaproteobacteria
Marine gamma proteobacterium HTCC2080	OM60/NOR5; Gammaproteobacteria
Agrobacterium sp. TSH97	Rhizobiales; Alphaproteobacteria
Anderseniella baltica	Rhizobiales; Alphaproteobacteria
Anderseniella baltica strain BA141	Rhizobiales; Alphaproteobacteria
Rhizobium sp. MSSRF QS100	Rhizobiales; Alphaproteobacteria
Bacterium HIMB11	Rhodbacterales; Alphaproteobacteria
Rhodobacteraceae bacterium LSUCC0246	Rhodbacterales; Alphaproteobacteria
Rhodobacteraceae bacterium LSUCC0259	Rhodbacterales; Alphaproteobacteria
Roseobacter sp. strain WM2	Rhodbacterales; Alphaproteobacteria
Altererythrobacter ishigakiensi	Sphingomonadales; Alphaproteobacteria
Erythrobacteraceae bacterium LSUCC0210	Sphingomonadales; Alphaproteobacteria
Erythrobacteraceae bacterium LSUCC0236	Sphingomonadales; Alphaproteobacteria
Erythrobacteraceae bacterium LSUCC0240	Sphingomonadales; Alphaproteobacteria
Erythrobacteraceae bacterium LSUCC0267	Sphingomonadales; Alphaproteobacteria
Bacterium MH1	Vibrionales; Gammaproteobacteria
Vibrio chagasii	Vibrionales; Gammaproteobacteria
Vibrio pelagius	Vibrionales; Gammaproteobacteria
Vibrio proteolyticus	Vibrionales; Gammaproteobacteria
Vibrio sp. 0208F3	Vibrionales; Gammaproteobacteria
Vibrio sp. PaH3.31d	Vibrionales; Gammaproteobacteria
Vibrio sp. TP187	Vibrionales; Gammaproteobacteria

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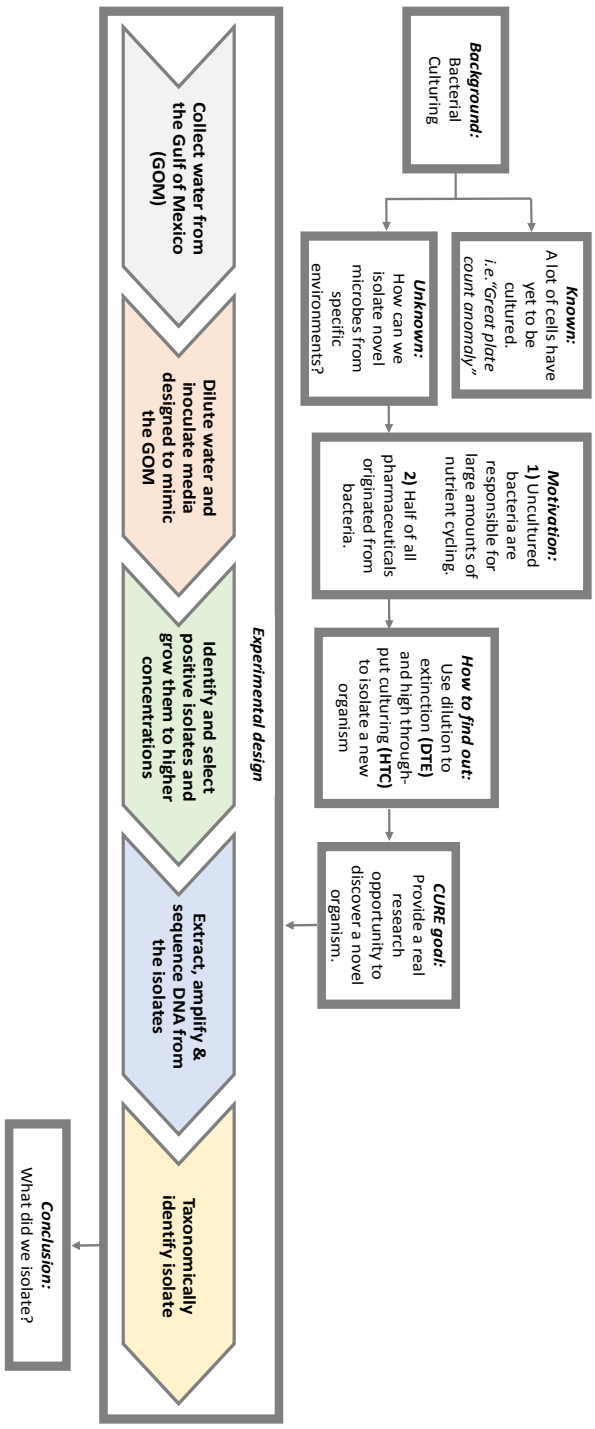

Figure 1. Flowchart of the mCURE background and experimental design. Using this flowchart, students are guided through the scientific process to gain an understanding of the relevance and importance of the project. Various segments of the course are color-coded (grey, orange, green, blue and yellow) corresponding to Table 1 where the week-by-week activities for each of these segments are described. This flowchart may be modified as needed to suit alternative projects using a similar protocol.

Figure 2. Survey results for questions with a significant difference (p-value ≤ 0.05) between research (mCURE) and traditional lab student pre/post perceptions. Note that the y-axis for "Reporting- Write results" is on a different scale than the other graphs.

Supplemental Materials

Supplemental Table 1. Example of a real implementation schedule of the idealized template in Table 1.

Appendix 1. Inoculation Protocol

Appendix 2. Transfer and Cryostock Protocol

Appendix 3. DNA Extraction Protocol

Appendix 4. BLAST How-to Guide

Appendix 5. Reading Guide

Appendix 6. Ordering a Scientific Paper

Appendix 7. BLAST Behind the Scenes

Appendix 8. Informal Writing 1

Appendix 9. Informal Writing 2

Appendix 10. Formal Writing 1

Appendix 11. Formal Writing 2

Appendix 12. Sample Final Exam

Appendix 13. Survey Instrument

Appendix 14. Example Student Assignments

Appendix 15. All Survey Instrument Data