Memory retrieval recruits both innate and learned olfactory processing centres in *Drosophila*

Michael-John Dolan[1-2] *, Ghislain Belliart-Guérin[3]*, Yoshinori Aso[2], Alexander Shakeel Bates[1], Shahar Frechter[1], Allan Wong[2], Adnan Hammad[1], Gerald M. Rubin[2], Thomas Preat[3], Pierre-Yves Plaçais[3 #], Gregory S.X.E. Jefferis[1,4 #]

[1]Division of Neurobiology, MRC Laboratory of Molecular Biology, Cambridge CB2 0QH, UK

[2]Janelia Research Campus, Howard Hughes Medical Institute, United States

[3] Genes and Dynamics of Memory Systems, Brain Plasticity Unit, CNRS, ESPCI Paris, PSL Research University, 10 rue Vauquelin, 75005 Paris, France

[4]Department of Zoology, University of Cambridge, CB2 3EJ, UK

*Joint first author

#Senior and Corresponding authors

Contacts: jefferis@mrc-lmb.cam.ac.uk or pierre-yves.placais@espci.fr

Abstract

Many animals can show either learned or innate behavioural responses to a given stimulus. However, how innate and memory circuits interact to produce an appropriate behavioural response is unknown. The *Drosophila* olfactory system is an excellent model to study how sensory stimuli are processed, stored and transformed into behaviour. Its numerical simplicity, powerful genetics and similarity to mammalian olfactory circuits have enabled many insights into sensory processing and memory. Two higher brain regions, the lateral horn (LH) and the mushroom body (MB) are thought to mediate innate and learned olfactory behaviour respectively, although the function of the LH has not been directly tested. Here we identify a LH cell-type (PD2a1/b1) that receives input from the MB. In contrast to the above model that LH is only necessary for innate behaviour, this cell-type is required for memory retrieval. Moreover, the activity of this cholinergic cell-type is modulated by training, indicating that memory information is passed to the innate olfactory processing centre. PD2a1/b1 dendrites likely receive input from appetitive olfactory projection neurons while their axons project to convergence zones of the MB extrinsic neurons and interdigitate with MB input and output neurons. This work demonstrates that LH neurons play a role in memory retrieval and illustrates the extensive interconnectivity of these two higher brain regions.

Introduction

Animals can respond to stimuli of particular ethological relevance through the action of natural selection on their genome on evolutionary timescales. For the most other stimuli, animals need to learn how to respond adaptively. However, it remains unknown how memory recall interacts with innate sensory representations to produce the most appropriate behaviour. This study explores this very general issue using the *Drosophila* olfactory system. Olfaction is a shallow sense that has a privileged connection to memory systems in many species (Su et al., 2009). The genetic tractability of *Drosophila* and the numeric simplicity of its brain make it an ideal model to study how these processes interact at a neural circuit level (Masse et al., 2009), while the similarity of peripheral olfactory circuits across the animal kingdom makes it possible that circuit principles may also be shared deeper in the brain between insects and mammals (Su et al., 2009; Wilson, 2008).

In *Drosophila*, olfactory sensory neurons project to specific glomeruli in the antennal lobe (Masse et al., 2009; Su et al., 2009; Vosshall et al., 2000). Following local computations, projection neurons (PNs) make divergent connections to two higher processing regions, the lateral horn (LH) and the mushroom body (MB) (Keene and Waddell, 2007; Masse et al., 2009). The prevailing model of olfactory processing describes a clear functional division between these regions: the MB is required for learning, consolidation and retrieval of olfactory memories while the LH is thought to mediate innate behaviour (Keene and Waddell, 2007; Masse et al., 2009). Many studies have confirmed the necessity of the MB for associative memory, where a reward or

punishment (the unconditioned stimulus, US) is associated with one odour (the conditioned stimulus, CS+) but not with a second odour (CS-) (Keene and Waddell, 2007). The MB's function in olfactory memory appears conserved in other insects such as ants (Vowles, 1964) and honeybees (Erber et al., 1980). The role of the LH in innate behaviour has been inferred from experiments that silenced the MB and observed innate olfactory responses (Heimbeck et al., 2001; Parnas et al., 2013) (but see (Wang et al., 2003)). However, no studies to date have directly examined the behavioural functions of LH neurons in olfaction.

Brain mapping studies have shown that PNs from different glomeruli have stereotyped axonal projections in the LH (Datta et al., 2008; Jefferis et al., 2007; Kazama and Wilson, 2009; Marin et al., 2002; Wong et al., 2002) consistent with a role in innate olfactory behaviours. Anatomical and physiological analyses have shown a role for specific *Drosophila* LH neurons in processing pheromone cues likely relevant for sex-specific behaviours such as courtship and aggression (Jefferis et al., 2007; Kohl et al., 2013; Liang et al., 2013; Ruta et al., 2010). Recent results have shown that some LH neurons can also show stereotyped responses to general olfactory stimuli (Fişek and Wilson, 2014; Strutz et al., 2014) and are stereotypically connected to input PNs (Fişek and Wilson, 2014). In addition, new large scale data have confirmed response stereotypy and showed that different LHNs have wide variations in odour tuning and may encode odour quality (Fişek and Wilson, 2014; Kohl et al., 2013; Strutz et al., 2014) (SF and GSXEJ, personal communication).

In contrast to the LH, the neurons of the MB assembly are extremely well-characterised (Aso et al., 2014a). The dendrites of intrinsic MB neurons (Kenyon cells) are localised to a region called the calyx, where they sample incoming PN axons in an apparently random manner (Caron et al., 2013). Kenyon cells have parallel, axonal fibers which form five different lobes, with three distinct branching patterns that define as many Kenyon cell types (Aso et al., 2014a; Crittenden et al., 1998). A comprehensive anatomical analysis revealed that the lobes can be subdivided into 15 compartments, each innervated by specific dopaminergic input neurons (DANs) and MB output neurons (MBONs) (Aso et al., 2014a). These compartments are both anatomically and physiologically distinct (Cohn et al., 2015; Hige et al., 2015a), although each Kenyon cell axon forms synapses in all compartments of each lobe (Cohn et al., 2015).

Odours are sparsely represented in the Kenyon cell assembly so that in the lobes only a subset of axons will release neurotransmitter upon olfactory stimulation (Honegger et al., 2011; Perez-Orive et al., 2002; Szyszka et al., 2005). Electric shock, the US during aversive learning, activates a subset of DANs such that when US and CS+ are presented coincidentally, the subset of olfactory-driven Kenyon cells also receive dopaminergic input within specific compartments. This coincident input produces compartment-specific synaptic plasticity (Bouzaiane et al., 2015; Cohn et al., 2015; Hige et al., 2015a; Owald et al., 2015), changing the response of that compartment's MBON to the CS+. MBONs function in valence behaviours, and a modified response to the trained odour may bias the fly's behaviour towards avoidance or attraction depending on the compartment (Aso et al., 2014b; Owald et al., 2015). One of these output neurons, MBON-α2sc (also known as MB-V2α), projects from the MB to several brain regions,

including the LH (Séjourné et al., 2011). Previous work demonstrated this MBON is required for the retrieval of aversive olfactory memories across short, medium and long timescales (Séjourné et al., 2011), although this is not the case for appetitive memories (Felsenberg et al., 2017; Séjourné et al., 2011). Recordings from MBON-α2sc demonstrated it is broadly responsive to odour (Hige et al., 2015b) and depresses its response to the CS+ after training (Hige et al., 2015b; Séjourné et al., 2011). While stimulation of the entire V2 cluster (MBON-α2sc, MBON-α′3m and MBON-α′3ap) drives approach, activation of MBON-α2sc alone does not lead to any change in valence behaviour (Aso et al., 2014b). Given the presumed role of the LH in innate olfaction, the function of this MB to LH projection is unclear. Is memory information transmitted to the LH and if so, is this communication required for retrieval of the aversive memory?

In this study we examine the behavioural function of this connection between the presumed innate and learned olfactory processing centers. We use a combination of computational anatomy and high-resolution microscopy to identify a LH output neuron cell-type (PD2a1/b1) which is postsynaptic to MBON-α2sc. We then proceed to test the function of this cell-type in behaviour. Contrary to the model of olfactory processing described above, PD2a1/b1 is necessary for memory retrieval. To confirm our observations we generated split-GAL4 lines (Luan et al., 2006; Pfeiffer et al., 2010) specifically targeting these neurons and used these reagents to confirm their necessity for memory recall. We performed calcium imaging of olfactory responses and similarly to MBON-α2sc, PD2a1/b1 also depress their responses to the CS+ after training. Finally, we determine the likely PN input to PD2a1/b1 dendrites and show that PD2a1/b1 neurons

potentially synapse back onto the MB network by projecting to convergence zones containing MBON axons and DAN dendrites.

Results 1: Identifying neurons postsynaptic to MBON-α2sc in LH

In order to understand the role of information flow from the MB and LH, we first sought to identify the postsynaptic neurons in the LH which receive input from the MBON-α2sc. As no transsynaptic labelling system exists in flies, we developed a computational pipeline to find putative MBON-α2sc postsynaptic candidates in the LH. We used *in-silico* overlap of GAL4 expression patterns to narrow down the field of candidate postsynaptic cell-types. Using image registration (Jefferis et al., 2007) of MBON-α2sc expressing a presynaptically localised marker (Christiansen et al., 2011), we created a mask of the MBON axonal terminals in the LH within template brain space. We then compared pixel overlap with registered images (Manton et al., 2014) of published GAL4 lines (Jenett et al., 2012) and ranked the lines by an "overlap score", which for each brain was normalised to GFP signal within the MB peduncle to exclude lines with MB expression. This allowed us to identify lines with no Kenyon cell expression, which could complicate behavioural analysis. The distribution of scores for approximately 3,500 GAL4 lines shows that the majority of lines lie close to zero or have negative scores (Figure 1A). Negative scores are derived from lines with little or no LH overlap and strong peduncle expression. We chose to focus on the top ~100 lines, above the 0.97 quantile. After excluding lines driving expression in MBON-α2sc, examination of these top hits identified 5 cell-types putatively postsynaptic to the MBON-α2sc in the dorsal LH. As many lines were excluded during manual

annotation due to broad expression, there are certainly other LH neurons we did not analyse in this study.

We next generated a LexA line to orthogonally control MBON-α2sc (Figure S1A), and conducted double-labelling of MBON presynapses and various LH cell-types identified in the computational screen to narrow down the number of identified cell-types. Two of the five cell-types had potential synaptic sites identified using double-labelling and high-resolution confocal microscopy: Lateral Horn output neuron cell-types PD2a1/b1 (Figure 1B-1C, see below for single neuron data) and AV6a1 (Figure S2A, S2C). These names are defined by a hierarchical nomenclature, that considers cell body/primary neurite tract, axonal tract and arborisation zones, and detailed morphology that will be presented in a forthcoming manuscript (SF and GSXEJ, in preparation). We also repeated this analysis for a mask of MBON axonal processes in the superior intermediate protocerebrum (SIP, data not shown) which yielded one clear, candidate postsynaptic cell-type (Figure S2B) which was further supported by double labelling (Figure S2D).

Results 2: PD2a1/b1 is necessary for memory retrieval

We took the sparsest GAL4 lines for each of the three cell-types identified and screened for memory retrieval defects when the neurons were silenced in an aversive olfactory associative conditioning paradigm. LH cell-types expressed the temperature-sensitive neuronal silencer shibire^{ts1} (Kitamoto, 2001) which inhibits neuronal signalling at high temperatures (33℃, the restrictive temperature). By raising the temperature (from permissive 25℃ to restrictive 33℃)

during a memory test 3h after an aversive olfactory conditioning we could silence these neurons to probe their role specifically in memory recall (Séjourné et al., 2011).

Silencing the AV6a1 and SIP cell-type GAL4 lines had no detectable effect on memory performance (Figure S2G-H). However, silencing PD2a1/b1 neurons with R37G11-GAL4 impaired memory retrieval relative to genotype (Figure 1D'') and permissive temperature (Figure S3B) controls in a 3h memory test assay. We extended these analyses of PD2a1/b1 to include immediate and long term memory which also require MBON-α2sc (Bouzaiane et al., 2015; Séjourné et al., 2011). Silencing PD2a1/b1 neurons attenuated memory retrieval for both phases of memory (Figure 1D' and 1D''') compared to genotype and permissive temperature controls (Figure S3A and S3C). Surprisingly, inhibition of PD2a1/b1 had no effect on naive olfactory avoidance to either of the two training odours at the same concentration used in our memory assay (Figure 1E), indicating that the observed phenotype was not a defect in innate olfactory processing, the presumed function of LH neurons. These results indicate that PD2a1/b1 signalling during the test phase is necessary for memory recall.

We confirmed that PD2a1/b1 is primarily an output cell-type by expressing HA-fused synaptotagmin (Syt::HA) to label presynapses (Robinson et al., 2002) (Figure 1F). We also observed some presynapses in the presumptive dendrites (Figure 1F), although this has been observed for many different types of LH output neurons (Dolan, unpublished). We next determined the neurotransmitter profile of these cells using immunohistochemistry. PD2a1/b1

was ChAT-positive (Figure 1G'-G''') but GABA- and dVGlut-negative (Figure S1B-C) indicating that neurons in cell-type are cholinergic LH outputs.

Results 3: Generation and characterization of cell-type specific split GAL4 lines

Although a relatively specific GAL4 line, R37G11-GAL4 contained several other cell-types that could confound our behavioural results. To confirm that PD2a1/b1 was indeed responsible for the memory retrieval deficit, we generated split-GAL4 lines (Luan et al., 2006; Pfeiffer et al., 2010) that specifically labelled PD2a1/b1 neurons in the central brain (Figure 2A-B). Two of the lines identified, LH989 and LH991, used the R37G11-DBD hemidriver, the same enhancer that drove the PD2a1/b1 GAL4 line tested above. We focused on these two split-GAL4 lines because they were most likely to contain identical LH neurons implicated in GAL4 behavioural analysis. Both of these split-GAL4 lines also labelled neurons in the ventral nerve cord (VNC), however these VNC cell-types were different between lines (Figure 2A-B).

We compared the number of PD2a1/b1 neurons labelled by each line. R37G11-GAL4 labelled 6.9±0.6 cells while LH989 and LH991 contained 5.25±0.5 and 5.67±0.8 neurons respectively. To understand the anatomy of the constituent cells within the PD2a1/b1 cell-type, we labelled single neurons in R37G11-GAL4 and the two split-GAL4 lines using the MultiColor FlpOut (Nern et al., 2015) technique (Figure S4A-C), isolating 22 single neurons from the PD2a1/b1 cell-type. 3/22 labelled neurons also projected to the MB calyx (and indeed this was visible in projection patterns of R37G11-GAL4, LH989 and LH991), while all other neurons appeared indistinguishable (Figure S4B-D). Therefore these lines label two distinct subtypes, PD2a1

(without calyx projections) and PD2b1 (with calyx projections). As we could not get specific genetic control of these subtypes we refer to these cells as PD2a1/b1. PD2a1/b1 neurons appear morphologically similar to some members of a broader class of Type I LH neurons identified in a previous study (Fişek and Wilson, 2014).

To confirm that PD2a1/b1 is involved in the retrieval of several memory phases, immediately after single-cycle training, on the middle-term time scale (~3h), and 24h after spaced training, we repeated our behavioural experiments with these sparse, split-GAL4 lines. When flies were tested at the restrictive temperature to silence PD2a1/b1, memory performance was impaired in each of these three conditions compared to genotype controls (Figure 2C-H). This ranged from mild attenuation immediately after training (Fig. 2C,D) to full impairment for LTM retrieval (Fig. 2G,H), similar to phenotypes observed by silencing MBON-α2sc (Séjourné et al., 2011). This defect was due to neuronal silencing, as genotypically identical flies tested at the permissive temperature had no memory recall deficits at any phase (Figure S5). Finally we confirmed with these sparse split-GAL4 lines that silencing these PD2a1/b1 neurons had no effect on innate olfactory avoidance for the two training odours in our behavioural assay (Figure 2I-J), indicating that the observed defect was specific to memory recall. Output from cell-type PD2a1/b1 is therefore necessary for the retrieval of aversive olfactory memory, with the same characteristics as MBON-α2sc.

Results 4: MBON-α2sc drives activity in PD2a1/b1

Our double labelling experiments suggested that MBON-α2sc is presynaptic to PD2a1/b1, but light microscopy does not have the resolution to confirm synaptic connectivity. We first used GRASP (Gordon and Scott, 2009) as an independent measure of the adjacency of PD2a1/b1 dendrites and MBON axons. The experimental genotype displayed clear GFP reconstitution in the dorsal LH (Figure 3A), indicating that the processes are proximal enough to form synapses, while no signal was detected in either of the control genotype brains (Figure 3B-C).

As MBON-α2sc is cholinergic (Aso et al., 2014a; Séjourné et al., 2011), we would expect that stimulation of this neuron would drive activity in PD2a1/b1 if these neurons are connected. Initial experiments with optogenetic tools failed, as targeted expression of CsChrimson (Klapoetke et al., 2014) caused either fatality or miswiring in the two MBON-α2sc LexA lines available, mostly likely due to overexpression (data not shown). In an alternative approach we expressed the heat activated ion channel dTRPA1 (Hamada et al., 2008) to activate MBON-α2sc (Figure 3D-3F) while recording calcium transients in PD2a1/b1. We used R37G11-GAL4 to drive expression of GCaMP6f (Chen et al., 2013) and our R71D08-LexA line to drive dTRPA1 (Figure 3D). We imaged the axons of PD2a1/b1 *in-vivo* to determine if driving MBON-α2sc could induce calcium transients in PD2a1/b1. In a control experiment, we observed a small temperature-dependent increase in calcium in the absence of the LexAop2-dTRPA1 transgene indicating that temperature alone may also stimulate these neurons (Figure 3E-F), consistent with the projection of thermosensory neurons to the MB and LH (Frank et al., 2015). Similarly, we also observed a small calcium increase in flies carrying only the LexAop-dTRPA1 transgene (Figure 3E-F). However, increasing temperature in flies expressing dTRPA1 in MBON-α2sc

yielded a larger increase in calcium indicating that these neurons are functionally connected (Figure 3E-F). We confirmed that dTRPA1 was expressing in MBON-α2sc by driving expression of a LexAop2-TdTomato reporter in the same landing site as the LexAop2-dTRPA1 transgene (Figure S6). This data, together with the double labelling and GRASP results suggest that MBON-α2sc directly connects to PD2a1/b1, a LH cell-type that is necessary for memory retrieval.

Results 5: PD2a1/b1 neurons have decreased responses to the CS+

After training, MBON-α2sc depresses its response to the conditioned stimulus (Hige et al., 2015a; Séjourné et al., 2011). Next we wanted to determine if PD2a1/b1, being downstream of MBON-α2sc, also modulates its response to the CS+. We drove expression of the calcium indicator GCaMP3 (Tian et al., 2009) in PD2a1/b1 axons (Figure 4A). First, we observed in naïve flies that PD2a1/b1 neurons were responsive to 3-Octanol (Oct) and 4-Methylcyclohexanol (Mch), the two odorants that are alternatively used as CS+ in our behavioural experiments (Figure 1D–E and Figure 2). PD2a1/b1 neurons displayed significantly higher response for Oct than for Mch. Since MBON-α2sc responds with similar magnitude to these two odours (Hige et al., 2015b; Séjourné et al., 2011), this likely reflects additional input from antennal lobe PNs to the LH.

We next looked for training-induced alterations of odour responses, by comparing olfactory responses in PD2a1/b1 neurons following either associative training or a control unpaired protocol, which matched the odour sequence of the associative training but temporally separated

electric shocks and odour delivery (See Figure S7 for full protocol). We performed these experiments either 3h after single-cycle training (Figure S7A-B), or 24h after spaced training (Figure S7C), and using either Oct or Mch as the CS+.

Our measurements revealed that the pairing between the CS+ and electric shocks during single-cycle training resulted in a decreased CS+ response in PD2a1/b1 axons 3h later, either compared to unpaired controls (Figure 4D' and E') or relative to the CS- response in the same fly (Figure 4D''and E''). Similar results were observed 24h after spaced training, i.e. upon LTM formation (Figure 4F-G). This data is consistent with PD2a1/b1 neurons receiving memory-relevant information (the decreased CS+ response), resulting from depression at Kenyon cell to MBON-α2sc synapses.

After a spaced protocol, either associative or unpaired, responses were globally lower than after single-cycle protocol or than in naïve flies (compare scale bar in Figure 4F-G and C–E). In addition, although responses to Oct were systematically higher than to Mch (black bars in Figure 4D'', E'', F'' and G''), the magnitude of this difference varied depending on the sequence of odour presentation during conditioning, being lower when Mch was presented first. This phenomenon occurred whether or not flies formed a memory, and may represent olfactory plasticity occurring in PNs or AL.

Results 6: PD2a1/b1 likely receives input from PNs encoding attractive odours and interdigitates with DAN dendrites and MBON axons

As noted earlier PD2a1/b1 neurons are likely to receive input from PNs as well as MBON-α2sc. Indeed their dendrites are poised to integrate inputs from these two sources. A previous study identified morphologically similar neurons which are known to receive PN input (Fişek and Wilson, 2014). Since the PN inputs to the LH have been mapped anatomically (Jefferis et al., 2007) and these maps can be predictive of functional connectivity (Fişek and Wilson, 2014), we sought to determine which PNs might provide input from the AL. We carried out an overlap analysis using a mask for PD2a1/b1 dendrites (Figure S9B) and uniglomerular PN axons from 47 different glomeruli (Figure 5A).

The dorsal LH, where PD2a1/b1 dendrites are located, has previously been associated with coding of food odours (Jefferis et al., 2007). We categorised the function of PNs in our dataset based on published data (see methods). The top eight PNs with the greatest PD2a1/b1 dendrite overlap were from glomeruli (VM3, DM3, DP1m, DM5, DM2, DM1, VM2 and VA2) responsive to food/attractive odours (Figure 5A). This includes DM1 and VA2, glomeruli required for approach behaviour to vinegar (Semmelhack and Wang, 2009). These data suggest that PD2a1/b1 receives predominantly appetitive olfactory stimulation from the AL. Preliminary data from EM reconstruction confirms that PD2a1/b1 neurons receive input from both PNs and the MBON-α2sc (A.S.B., D. Bock, and G.S.X.E.J., unpublished observations).

As a first insight into what role PD2a1/b1 may play in memory retrieval, we sought to identify potential downstream targets of this LH cell-type. Initial characterisation of PD2a1/b1 axons suggested it transmits information from the LH to the crepine (CRE) and the superior

intermediate protocerebrum (SIP) (Figure S8). The SIP and CRE have been identified as convergence zones that contain both the dendrites of DANs and the axons of MBONs (Aso et al., 2014a, 2014b; Owald et al., 2015). This raised the possibility that PD2a1/b1 may potentially interact with input and output neurons of the MB assembly.

We searched for potential sites of contact with MBONs and DANs by computational alignment of segmented axon and dendrite masks in the same brain space (Jefferis et al., 2007; Rohlfing and Maurer, 2003), comparing PD2a1/b1 axons (Figure S9A) with published MB data (Aso et al., 2014a). We calculated the percentage overlap between the mask of PD2a1/b1 axons and either the axon or dendrite of each MB cell-type. This coarse analysis revealed several potential contacts between PD2a1/b1 and DAN dendrites (Figure 5B) and MBON axons (Figure 5C). We further investigated all neurons above a 15% threshold of PD2a1/b1 overlap using double labelling with R37G11-LexA, a PD2a1/b1 LexA line (Pfeiffer et al., 2010) (www.janelia.org/gal4-gen1).

We examined three DANs using double labelling. Both PAM-β′1 and PAM-β′2m interdigitated and exhibited potential synaptic contacts with PD2a1/b1 axons (Figure 5D-E), suggesting potential synaptic contacts. PAM-β′2p had dendrites proximal to PD2a1/b1 axons but did not interdigitate (data not shown). Analysis of MBON axons and PD2a1/b1 axons revealed close co-projection for MBON-α′2, MBON-γ2α′1 and MBON−β′2mp (Figure 5F-H), indicating common postsynaptic partners or possibly axo-axonic synapses. This data indicates that

PD2a1/b1 projects to MB convergence zones, interdigitates with both DANs and MBONs, potentially feeding back onto the MB assembly (Aso et al., 2014a, 2014b).

Discussion

In both mice and *Drosophila*, there is thought to be a functional division between higher olfactory processing regions. Although many studies have manipulated the neurons of the MB (Aso et al., 2014b; Dubnau et al., 2001; Krashes et al., 2007; McGuire et al., 2001; Owald et al., 2015; Wang et al., 2003), the role of LH neurons in olfactory behaviour had not been directly tested. Previous studies inferred a role for the LH in innate behaviour based on comparing PN and MB silencing (Heimbeck et al., 2001; Strutz et al., 2014; Wang et al., 2003). For the first time, to our knowledge, we have directly interrogated the role of LH neurons in olfactory behaviour in adult *Drosophila*, discovering a LH cell-type that is necessary for memory retrieval.

Previous work (Aso et al., 2014a; Séjourné et al., 2011; Tanaka et al., 2008) identified MBON-α2sc as an MBON that connects the MB to the LH and other regions, raising the possibility that memory information flows from the MB to the LH. After associative conditioning, synaptic depression occurs at Kenyon cell to MBON-α2sc synapses, resulting in depression of its response to the trained odour (Hige et al., 2015a; Séjourné et al., 2011). MBON-α2sc output is necessary for memory recall across immediate memory, medium-term and long-term memory (Aso et al., 2014b; Bouzaiane et al., 2015; Séjourné et al., 2011) but dispensable for innate olfactory responses.

Labelling cells connected to genetically defined neuronal populations is challenging in *Drosophila*, in the absence of transsynaptic tracing tools. Some studies have used photoactivation of PA-GFP proximal to axon terminals, but this does not provide genetic access to postsynaptic neurons (Datta et al., 2008; Frank et al., 2015; Ruta et al., 2010). We developed a simple but novel approach to identify putative postsynaptic partners of MBON-α2sc. We screened *in silico* a database of 3500 confocal images of expression patterns, identifying genetic drivers expressed in candidate postsynaptic partners; we then generated intersectional drivers based on that information. The *in silico* approach is scalable, can be applied to any neuron type in *Drosophila* and depends only on co-registered images and open source software that we have made freely available. Recent developments in genetic labelling (Nern et al., 2015) and imaging of whole mount brain tissue (Aso et al., 2014a) should increase the availability of appropriate image data in flies other model organisms.

By combining this analysis with double labelling, GRASP and thermogenetic mapping we identify LH output neurons PD2a1/b1, a neuronal cell-type postsynaptic to MBON-α2sc. Strikingly, using specific split-GAL4 control of PD2a1/b1 neurons in the brain, we found that PD2a1/b1 signalling is necessary for memory retrieval across all phases tested but dispensable for innate olfactory responses. It is quite possible that PD2a1/b1 neurons do participate in some innate olfactory behaviours, but they are not critical for the stimuli used in our memory tests. We also observed that while most of the constituent cells in PD2a1/b1 appear homogenous, a subset of neurons send dendrites to the MB calyx (PD2b1). The calyx is the site of PN input to the MB,

upstream from the site of associative olfactory memory, arguing against a role for this connection in our memory retrieval phenotype.

We next examined the responses of PD2a1/b1 to the training odours using *in-vivo* calcium imaging. Although odour tuning varies between individual flies, on average MBON-α2sc responds equally to both 4-Methylcyclohexanol and 3-Octanol (Hige et al., 2015b; Séjourné et al., 2011). PD2a1/b1 was also odour-responsive but unlike MBON-α2sc responded more strongly to Oct than Mch. The higher response to Oct implies that PD2a1/b1 receives additional olfactory input, most likely from PNs given its extensive dendritic arbors in the LH.

We observed that associative olfactory training induced a decreased response to the CS+ relative to the CS- in PD2a1/b1 neurons, in a manner similar to MBON-α2sc (Séjourné et al., 2011), indicating that memory information is sent from the MB to LH. This trace was observed both 3h after single-cycle training, or 24h after spaced training, and using either Oct or Mch as the CS+. Overall our results indicate that MBON-α2sc modulates PD2a1/b1 during memory retrieval to produce the appropriate behavioural response to the trained odour.

We used a combination of image analysis and double labelling to identify potential PN input to PD2a1/b1. We examined overlap of PD2a1/b1 dendrites with uniglomerular PN axons and found that PD2a1/b1 dendrites in the dorsal LH have likely receive input from PNs encoding appetitive/food odours (Hallem and Carlson, 2006; Semmelhack and Wang, 2009), including uniglomerular PNs from the DM1 glomerulus, which is necessary for attraction to vinegar

(Semmelhack and Wang, 2009). Using the same approach we examined convergence and connectivity between PD2a1/b1 and the MB assembly. We identified PAM-β′1 and PAM-β′2m as potential postsynaptic neurons to PD2a1/b1, although we do not have the resolution to confirm this with light microscopy. PAM-β′1 mediates memory erasure (Shuai et al., 2015) and therefore a MBON-LH-PAM neuron circuit may play a role in forgetting or extinction. Interestingly, a recent study demonstrated that MBON-α2sc is necessary for the extinction of appetitive memories (Felsenberg et al., 2017). PAM-β′2m, together with PAM-β′2p can drive approach behaviour when stimulated (Lewis et al., 2015). PD2a1/b1 also interdigitates with the axons of MBON-α′2, MBON-γ2α′1 and MBON−β'2mp. Silencing of MBON-α′2 throughout training and testing abolishes appetitive memories (Aso et al., 2014b). MBON-γ2α′1 drives approach when stimulated (Aso et al., 2014b), and has been implicated in appetitive memory retrieval; it is also necessary and sufficient for memory reconsolidation (Aso et al., 2014b; Felsenberg et al., 2017). MBON−β'2mp receives input from the MB compartment that is innervated by PAM-β'2m and plays a role in appetitive and aversive memory retrieval (Owald et al., 2015). In aggregate this data suggests that PD2a1/b1 interacts or at least converges with MB neurons that drive approach and reappraisal of memories.

We speculate that if PD2a1/b1 neurons are stimulated by appetitive odours (via PNs projecting to the LH) and converge with MBONs and DANs that mediate attractive behaviour, PD2a1/b1 neurons may themselves play a role in olfactory-driven approach. Therefore their decreased response to the CS+ after training may result in decreased attraction to the trained odour, resulting in net avoidance of the CS+. To examine this, we attempted to stimulate PD2a1/b1

optogenetically in a place preference assay (Aso et al., 2014b) but due to off-target expression in the GAL4 and split-GAL4 lines we failed to observe a consistent phenotype (data not shown). To test this model, future studies will need to refine genetic control of PD2a1/b1 and clarify the connectivity between PD2a1/b1 and identified PNs, MBONs and DANs. New resources such as a full electron microscopy volume of the adult fly brain (Zheng et al., 2017) will allow for comprehensive mapping of synaptic connectivity.

What function could this MB-to-LH connectivity fulfill? One possibility is to provide a means for learned and innate behaviours to interact in conflicting situations. MB-to-LH connectivity may also constrain what flies can learn through hard-wired circuitry in the LH (Jefferis et al., 2007; Marin et al., 2002; Wong et al., 2002). One recent study has implicated the MB in the integration of context and internal state (Lewis et al., 2015) during CO_2 olfactory processing, implying a memory-independent role for the MB. In this study we also identify potential contacts between PD2a1/b1 and DANs and MBONs. Moreover, similar potential convergence has been identified previously for two other LH output neurons (Aso et al., 2014b), for which no functional attribution has been reported so far. These data and our new study provide evidence that the MB and LH are interconnected in multiple ways. Together these findings illustrate that the simple functional division of innate and learned olfactory behaviour in the fly higher brain between LH and MB is an oversimplification.

The olfactory systems of both flies and mice share the same basic blueprint (Su et al., 2009; Wilson, 2008). In mice, the piriform cortex is required for learning and memory (Choi et al.,

2011), responds sparsely to odours (Stettler and Axel, 2009) and samples from the whole olfactory bulb (Miyamichi et al., 2011), similar to the MB. In contrast the olfactory amygdala is necessary and sufficient to instruct innate olfactory behaviour (Root et al., 2014) and receives stereotyped input from the olfactory bulb (Miyamichi et al., 2011; Sosulski et al., 2011), drawing a comparison to the LH. Intriguingly, there are uncharacterised connections between the Piriform Cortex and olfactory amygdala (Schwabe et al., 2004) and we speculate that these connections may play a role in memory retrieval in the mammalian brain.

In summary, we have examined the function of connections between the olfactory center for learning and memory (the MB) and the innate processing center (the LH). By tracing MBON-α2sc into the LH we identified a cholinergic output neuron, PD2a1/b1, that receives direct input from the MB. In contrast to current models of the fly olfactory system (Keene and Waddell, 2007; Masse et al., 2009), this LH output neuron is necessary for memory retrieval but dispensable for innate avoidance behaviour. Finally we demonstrate that PD2a1/b1 potentially receives input from food-related PNs and interdigitates with behaviourally-relevant DANs and MBONs, implying interconnectivity between these two brain regions with distinct wiring principles and odour coding rules (Fişek and Wilson, 2014; Hige et al., 2015b; Honegger et al., 2011).

Acknowledgements

This work was supported by an MRC LMB Graduate Studentship, Boehringer Ingelheim Fonds PhD Fellowship and Janelia Graduate Research Fellowship (M-J.D.); ERC Starting (211089) and Consolidator (649111) grants and core support from the MRC (MC-U105188491) (to G.S.X.E.J); the Agence National de la Recherche funding of the MemoNetworks project (to P.-Y.P and T.P.) and the Labex Memolife for PhD fellowship to G.B.-G.; and the Howard Hughes Medical Institute (A.W. and G.R.). This work was also supported by the Janelia Visiting Scientist Program. The FlyLight Project Team performed brain dissections, histological preparations and confocal imaging for the PD2a1/b1 Split-GAL4 characterisation, Polarity and MCFO data. Heather Dionne cloned the R71D08-LexA construct and created the split-GAL4 hemidriver transgenic lines. We thank the Bloomington Stock Center (NIH P40OD018537) and Matthias Landgraf for fly lines. We thank Paavo Huoviala for contributing some PN tracings and curating the behavioural significances of different glomeruli. Finally we thank Marta Zlatic, Glenn Turner and his group, and the Jefferis and Préat groups for many insightful comments on the manuscript.

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Online Methods

1. Molecular Biology

The pBP-R71D08 gateway entry construct was a kind gift from Heather Dionne. The insert was transferred to the pBPLexA::p65Uw destination vector (Addgene) via a Gateway LR recombination (Invitrogen).

The enhancers used to create split-GAL4 hemidrivers were created based on annotations for PD2a1/b1 in a GAL4 expression pattern database (Jenett et al., 2012) as previously described (Aso et al., 2014a). The enhancer hemidriver lines were created as previously described (Pfeiffer et al., 2010).

All transgenic fly lines were generated by either Bestgene Inc or Genetic Services, Inc.

2. Drosophila Husbandry and Genetics

Standard techniques were used for fly stock maintenance and construction.For imaging and immunohistochemistry flies were raised at 25°C on standard Drosophila food. For MultiColor FlpOut (MCFO) experiments (Nern et al., 2015), the MCFO stock (see below) was crossed to either R37G11-GAL4, LH989 or LH991. Flies were collected after eclosion, transferred to a new food vial and incubated in a 37°C water bath for 20-25 minutes.

Transgenic lines used for behaviour (with the exception of optogenetic activation experiments) were outcrossed for five generations to a *w1118* strain in a wild-type Canton-Special (CS) background. For behavioral experiments (with the exception of optogenetic activation experiments) flies were raised at 18 ℃ and 60% humidity under a 12-hr:12-hr light-dark cycle.

For optogenetic activation experiments, flies were raised on standard Iberian containing yeast cornmeal and agar at 22 ℃ under a 12-hr:12-hr light-dark cycle. Behaviour was performed essentially as described (Aso et al., 2014b) For each vial of experimental flies, between 3-5 each of males and females were crossed on normal fly food supplemented with 1/500 all-trans-retinal (Sigma-Aldrich, MO, USA). Parental flies were allowed oviposit for 2-4 days before being tipped into a new vial, to control for population density of offspring. Upon eclosion females of the correct genotype were sorted on a cold plate and kept on fresh 1/250 all-trans-retinal food. Experiments were performed less than 3 days after cold sorting. All lines were grown at 22 ℃ in a covered box in a 12 hour light:dark cycle incubator. Optogenetic activation experiments were carried out at 50% humidity and 25℃.

All *Drosophila* strains used in this study are listed below:

•*w; +; 10XUAS-IVS-mCD8::GFP (attP2)*

• *w; LexAop2-Brp(d3)::mCherry (su(hw)attP5)* For labelling MBON-α2sc axons. A gift from Mathias Landgraf. (Christiansen et al., 2011)
• *w; Insite0089-GAL4* For labelling Av6a1 cell-type neurons. (Gohl et al., 2011)
• *w; +; R58G03-GAL4* (attP2) For labelling SIP cell-type neurons. (Jenett et al., 2012)
• *w; +; R37G11-GAL4 (attP2)* For labelling PD2a1/b1 cell-type neurons. (Jenett et al., 2012)
• *13xLexAop2-mCD8::GFP(su(Hw)attP8)* Reporter for the LexA system. Bloomington Stock Number: 32204 (Pfeiffer et al., 2010)
• *w; +; 20XUAS-IVS-mCD8::GFP (attP2)* Bloomington Stock Number: 32194
• *20xUAS-IVS-csChrimson::mVenus (attP18)* For optogenetic stimulation and split-GAL4 screening. Bloomington Stock Number: 55134 (Klapoetke et al., 2014)
• *w; LexAop-spGFP11* For GRASP experiments. Bloomington Stock Number: 58755 (Gordon and Scott, 2009)
• *w; +; UAS-spGFP1-10* For GRASP experiments. (Gordon and Scott, 2009)
• *w; R73B12-ADp65 (attP40); R37G11-DBD (attP2)* (this study). LH989, for labelling cell-type PD2a1/b1 neurons.
• *w; R29G05-ADp65 (attP40); R37G11-DBD (attP2)* (this study). LH991, for labelling cell-type PD2a1/b1 neurons.
• *w; +; R71D08-GAL4 (attP2)* For labelling the MBON-α2sc neurons. (Pfeiffer et al., 2010; Séjourné et al., 2011)
• *w; + ; UAS-Shi^{ts1}* For silencing neurons with a temperature shift (Bouzaiane et al., 2015; Kitamoto, 2001)
• *w; +; LexAop2-dTRPA1 (VK5)* For activating neurons with a temperature shift (Hamada et al., 2008; Pfeiffer et al., 2010)
• *w; +; LexAop2-TdTomato (VK5)* For confirming MBON-α2sc expression in landing site VK5 (Hamada et al., 2008; Pfeiffer et al., 2010)
• *w; R37G11-LexA (attp40)* For controlling PD2a1/b1 with the LexA system (Pfeiffer et al., 2010)
• *w; R71D08-LexA (attp40)* For controlling MBON-α2sc with the LexA system (Pfeiffer et al., 2010)
• *w; +; UAS-GCaMP3 (VK5)* For calcium imaging of odor responses after conditioning (Tian et al., 2009). Bloomington Stock Number: 32237
• *w, UAS-GCaMP6f (attP18)* For calcium imaging in thermal activation experiments (Chen et al., 2013)
• *w; pBPp65ADZpUw (attP40); pBPZpGAL4DBDUw (attP2)* Enhancerless split-GAL4 hemidrivers for behavioural control (Hampel et al., 2015; Pfeiffer et al., 2010)
• *w; +; 3xUAS-Syt::smGFP-HA (su(Hw)attP1), 5xUAS-IVS-myr::smGFP-FLAG (VK5)* Polarity and membrane marker (Aso et al., 2014a)

• *hsFlp2::PEST (attP3);+ ; 10XUAS-FRT>STOP>FRT-myr::smGFP-HA (VK00005), 10XUAS-FRT>STOP>FRT-myr::smGFP-V5-THS-10XUAS-FRT>STOP>FRT-myr::smGFP-FLAG (su(Hw)attP1)/ TM3, Sb*
Reporter for MultiColor FlpOut (MCFO) (Nern et al., 2015)
• *w, LexAop2-CD8::GFP (su(Hw)attp8) , 10xUAS-CD8::RFP (attp18)* For double labelling the membranes of R37G11-LexA and different DAN and MBON split-GAL4 lines. Bloomington Stock Number: 32229 (Liu et al., 2012)
• *w; R24E12-ADp65 (attP40); R52H01-DBD (attP2)* MB025B, split-GAL4 control of PAM-β'1 (Aso et al., 2014a)
• *w; R30G08-ADp65 (attP40); RTH-DBD (attP2)* MB032B, split-GAL4 control of PAM-β'2m (Aso et al., 2014a)
• *w; R20G03-ADp65 (attP40); R19F09-DBD (attP2)* MB018B, split-GAL4 control of MBON-α'2 (Aso et al., 2014a)
• *w; R25D01-ADp65 (attP40); R19F09-DBD (attP2)* MB077B, split-GAL4 control of MBON-γ2α'1 (Aso et al., 2014a)
• *w; R12C11-ADp65 (attP40); R15B01-DBD (attP2)* MB002B, split-GAL4 control of MBON−β'2mp (Aso et al., 2014a)

Figure Number	**Genotype**
Figure 1C	w; *LexAop2-Brp(d3)::mCherry (su(hw)attP5), R71D08-LexA (attP40)/+; R37G11-GAL4 (attP2)/10XUAS-IVS-mCD8::GFP (attP2)*
Figure 1F	*w; R73B12-ADp65 (attP40)/+; R37G11-DBD (attP2)/3xUAS-Syt::smGFP-HA (su(Hw)attP1), 5xUAS-IVS-myr::smGFP-FLAG (VK5)*
Figure 1G	*w, 20xUAS-IVS-csChrimson::mVenus (attP18)/w; +*; R37G11-GAL4/+
Figure 2A	*w, 20xUAS-IVS-csChrimson::mVenus (attP18)/w; R73B12-ADp65 (attP40)/+; R37G11-DBD (attP2)/+*
Figure 2B	*w, 20xUAS-IVS-csChrimson::mVenus (attP18)/w; R29G05-ADp65 (attP40); R37G11-DBD (attP2)*

Figure 4	*w; +; UAS-GCaMP3 (VK5)/R37G11-GAL4 (attP2)*
Figure 5C	*w, LexAop2-CD8::GFP (su(Hw)attp8) , 10xUAS-CD8::RFP (attp18)/w; R24E12-ADp65 (attP40)/+; R52H01-DBD (attP2)/+*
Figure 5D	*w, LexAop2-CD8::GFP (su(Hw)attp8) , 10xUAS-CD8::RFP (attp18)/w; R30G08-ADp65 (attP40/+); RTH-DBD (attP2)+*
Figure 5E	*w, LexAop2-CD8::GFP (su(Hw)attp8) , 10xUAS-CD8::RFP (attp18)/w; R20G03-ADp65 (attP40)/+; R19F09-DBD (attP2)/+*
Figure 5F	*w, LexAop2-CD8::GFP (su(Hw)attp8) , 10xUAS-CD8::RFP (attp18)/w; R25D01-ADp65 (attP40)/+; R19F09-DBD (attP2)/+*
Figure 5G	*w, LexAop2-CD8::GFP (su(Hw)attp8) , 10xUAS-CD8::RFP (attp18)/w; R12C11-ADp65 (attP40)/+; R15B01-DBD (attP2)/+*
Figure S1A	*w, 13xLexAop2-mCD8::GFP(su(Hw)attP8)/ w; R71D08-LexA (attP40)/+*
Figure S1B	*w; +; R37G11-GAL4 (attP2)/10XUAS-IVS-mCD8::GFP (attP2)*
Figure S1C	*w, 20xUAS-IVS-csChrimson::mVenus (attP18)/w; R73B12-ADp65 (attP40)/+; R37G11-DBD (attP2)/+*
Figure S2C	w; *LexAop2-Brp(d3)::mCherry (su(hw)attP5), R71D08-LexA (attP40)/Insite0089-GAL4; +/10XUAS-IVS-mCD8::GFP (attP2)*

Figure S2D	w; *LexAop2-Brp(d3)::mCherry (su(hw)attP5), R71D08-LexA (attP40)/+; R58G03-GAL4 (attP2)/10XUAS-IVS-mCD8::GFP (attP2)*
Figure S2E	*w; Insite0089-GAL4/+; 10XUAS-IVS-mCD8::GFP/+*
Figure S6	*w; R71D08-LexAp65 (attP40)/+; LexAop2-TdTomato (VK5)/+*

Table 1: List of transgenic lines used for anatomical experiments. For behavioural experiments the genotypes are labelled in the figure.

3. Immunohistochemistry

Throughout this study we used two different immunohistochemistry (IHC) protocols. Figures 1F, 2A-B, and S4 used Protocol 2 while all other IHC data was processed using Protocol 1.

Protocol 1:

IHCs were performed as described (Ostrovsky et al., 2013). All specimens were mounted in Vectashield (H-1000) (Vector Laboratories, Burlingame, CA, USA).

Protocol 2:

These IHCs were performed as described (Aso et al., 2014a). A full step-by-step protocol can be found at https://www.janelia.org/project-team/flylight/protocols. Following the IHC protocol the brains were fixed again in 4% Paraformaldehyde (Electron Microscopy Sciences, Hatfield, PA) for four hours at room temperature. The brains were mounted on poly-L-lysine-coated cover slips and dehydrated through a series of ethanol baths (30%, 50%, 75%, 95%, and 3 × 100%) each for 10 min. Following dehydration they were submerged in 100% Xylene three times for 5 minutes each. Samples were embedded in DPX (DPX; Electron Microscopy Sciences, Hatfield, PA).

Antibody, Concentration	**Catalog Number**	**Purpose**
Chicken anti-GFP, 1/1600	Abcam, ab13970	GFP or mVenus
Mouse anti-Brp, 1/20-1/30	DSHB, University of Iowa, nc82	neuropil
Mouse anti-GFP, 1/400	Roche, 11814460001	Reconstituted GFP (GRASP)
Rabbit anti-GABA, 1/200	Sigma-Aldrich, A2052	GABAergic neurons

Mouse anti-ChaT, 1/400	DSHB, University of Iowa ,4B1	Cholinergic neurons
Rabbit anti-DvGlut, 1/500	Gift from Dion Dickman, University of Southern California, USA (Chen et al., 2017)	Glutamatergic neurons
Alexa-488 Goat anti-mouse IgG1, 1/1600 or 1/400	Thermofisher, A-21121	Secondary for GRASP (1/1600) and MCFO (1/400, V5)
Alexa-488 Goat anti-chicken, 1/1600	Invitrogen, A-11039	Secondary for GFP stain
Alexa-568 Goat anti-rabbit, 1/1600	Invitrogen, A-21069	Secondary for mCherry
Alexa-568 Goat anti-mouse, 1/1600	Invitrogen, A-11004	Secondary for nc82
Alexa-633 Goat anti-mouse IgG1, 1/1600	Invitrogen, A-21126	Secondary for nc82 (FigS6)
Rat anti-FLAG tag, 1/200	Novus Biologicals NBP1-06712SS	Primary for FLAG stain (MCFO and Polarity)
Rabbit anti-HA tag, 1/300	Cell Signalling Technologies, C29F4	Primary for HA stain (MCFO and Polarity)
Mouse anti-V5, 1/500	AbD Serotec, DL550	Primary for V5 (MCFO)
Cy5 Donkey anti-Rat, 1/150	Jackson Immuno Research, 712-175-150	Secondary for Polarity (FLAG)
Cy3 Goat anti-Rabbit, 1/1000	Jackson Immuno Research, 111-165-144	Secondary for Polarity and MCFO (HA)
Cy2 Goat anti-Mouse, 1/600	Jackson Immuno Research, 111-165-144	Secondary for Polarity (nc82)
Alexa-647 Donkey anti-Rat, 1/150	Jackson Immuno Research, 712-605-153	Secondary for MCFO (FLAG)

Table 2: List of all antibodies used in this study, their catalog numbers and purpose.

4. IHC Image Acquisition

All images for IHC were acquired using a Zeiss 710 Confocal Microscope, essentially as described (Aso et al., 2014a; Cachero et al., 2010; Kohl et al., 2013). We used three modes of imaging: 20x, 40x and 63x.

For 20x imaging, whole mount brain and VNCs were imaged using a Plan-Apochromat 20x/0.8 M27 objective (voxel size = 0.56 × 0.56 × 1.0 µm; 1024 × 1024 pixels per image plane).
20x imaging was used for Figures 2A-B.

For 40x imaging, whole mount brains were imaged using an EC Plan-Neofluar 403/1.30 oil objective with 768 x 768 pixel resolution at each 1 µm, 0.6-0.7 zoom factor.
40x imaging was used for Figures 3A-C, S1A-B, S2E, S6.

For 63x imaging, whole mount brains were imaged using a Plan-Apochromat 63x/1.40 oil immersion objective (voxel size = 0.19 × 0.19 × 0.38 µm; 1024 × 1024 pixels). For certain images, tiles of regions of interest were stitched together into the final image (Yu and Peng, 2011).
63x imaging was used for Figures 1C, 1F-1G’’’, 5C-G, S1C, S4A.

5. Image Processing and analysis

To accurately label the presynapses of the LH-projecting MBONs, the 71D08-LexA driver was crossed to LexAop2-Brp(d3)::mCherry resulting in axon-specific labelling. For the region of the MBON under investigation (MBON-a2sc axons in the dorsal LH) a mask was created. Eight 71D08>Brp(d3)::mCherry brains were immunostained and registered onto a common template brain (JFRC2, http://www.virtualflybrain.org) using the nc82 counterstain. Image registration was carried out as described (Cachero et al., 2010) using the CMTK registration suite (https://www.nitrc.org/projects/cmtk). The boundary of the overlaid neurites for each region of interest from each brain was segmented manually as a mask in Fiji (https://fiji.sc/), using the Segmentation Editor function. The overlap was calculated against a large database of GAL4 expression patterns (Gohl et al., 2011; Jenett et al., 2012) also registered against JFRC2 (Manton et al., 2014) using the `cmtk.statistics` function in the open source nat (NeuroAnatomy Toolbox) package (https://github.com/jefferis/nat) for R (https://cran.r-project.org/). To control for the background signal of the brain we created a mask of the peduncle and performed the same overlap calculation for each GAL4 line. This peduncle overlap score was used normalise the MBON axon masks to produce the final overlap score for each GAL4 line. This allowed us to select lines with high signal-to-noise within the MBON masks and excluded expression patterns with Kenyon Cell expression which would confound behavioural analysis. The stacks GAL4 line expression patterns in the top 0.97 quartile were further analysed manually to identify Lateral Horn neurons.

For counting the number of cells in each line, images of R37G11-GAL4, LH989 and LH991 crossed to 20xUAS-csChrimson::mVenus (attp18) were used and cells manually counted.

MCFO brains were imaged in 63x mode (see above) and the stitched final image registered to the JFRC2013 template brain. Single neurons were manually annotated and segmented in 3D using Fluorender. Segmented single neurons were skeletonized as described (Costa et al., 2016) and their anatomy was analysed using the "nat.nblast" package for R (https://github.com/jefferislab/nat.nblast). We isolated 5 cells from R37G11-GAL4, 13 cells from LH989 and 5 cells from LH991. All lines contained neurons which projected to calyx.

To examine overlap between PD2a1/b1 axons and MB neurons or PD2a1/b1 dendrites and PN axons, high resolution images (63x) of PD2a1/b1 split-GAL4 lines driving both a membrane and presynapse markers were segmented using Fluorender (http://www.sci.utah.edu/...are/127-fluorender.html). For PN data we used published PN skeleton data (Grosjean et al., 2011; Jefferis et al., 2007; Silbering et al., 2011), a spreadsheet of glomeruli and published behavioural significance/functions are available upon request. We compared this to published segmentations of the DANs and MBONs (Aso et al., 2014a). All data were registered to the JFRC2013 template brain (Aso et al., 2014a) . For all cell-types in addition to the entire membrane stain, the axons and dendrites were segmented separately for at least n=2 well-registered brains. For each category of segmentation (dendrite-only, axon-only) we created a mask from their different samples by overlaying all the examples of each line. This was followed by contrast enhancement, gaussian blurring and auto-thresholding to create a mask. All image processing was performed using Fiji. Overlap comparisons for pairs of masks were compared in R using the cmtk.statistics function in the "nat" package.

Double labelling images performed with R37G11-LexA and different MBON and DAN Split-GAL4 lines were processed with a median filter using the despeckle command in Fiji. This was necessary to remove background due to the weak expression levels of the R37G11-LexA line.

6. Generation of split-GAL4 lines

Each split-GAL4 line consists of two hemidrivers, the p65ADZp in attP40 and the ZpGAL4DBD in attP2. The lines were screened by combining these two hemidrivers with a copy of *20xUAS-IVS-csChrimson::mVenus (attP18).* The brains of females from each line were dissected and screened with an epifluorescence microscope. Split-GAL4 combinations with favourable expression patterns (sparse expression of PD2a1/b1) were double balanced to make a stable stock.

7. Behaviour: Olfactory Assays

For all behavior experiments, 0–2 day-old flies were transferred to fresh food vials the day before conditioning. Conditioning and tests of memory performance and of olfactory acuity were performed as described previously (Bouzaiane et al., 2015). Groups of 40-50 flies were trained with either one cycle of aversive training (single-cycle training), or five cycles spaced by 15 min inter-trial intervals (spaced training). During one cycle of training, flies were first exposed to an odorant (the CS+) for 1 min while 12 pulses of 5s-long 60V electric shocks were delivered; flies were then exposed 45 s later to a second odorant without shocks (the CS–) for 1 min. The odorants 3-octanol and 4-methylcyclohexanol, diluted in paraffin oil at 0.360mM and 0.325mM respectively, were alternately used as conditioned stimuli. The test of memory performance was performed in a T-maze. Flies were placed at the convergence point of two airflows interlaced with octanol or methylcyclohexanol from either arm of the T-maze. After 1 min in the dark, flies were collected from both arms of the T-maze for subsequent counting, yielding a score calculated as $(N_{CS+} – N_{CS-})/ (N_{CS+} + N_{CS-})$. A single value of the performance index is the average of two scores obtained from two groups of genetically identical flies conditioned in two reciprocal experiments, using either odorant as CS+, and tested consecutively in the T-maze. Flies were maintained on food at all times, with the exception of during conditioning and memory test. Memory test occurred 10±5 minutes after conditioning, 3h±30 minutes after conditioning, and 24±1.5 h after conditioning to assay immediate memory, 3-h memory and long-term memory, respectively. For experiments involving neuronal blockade with Shi^{ts}, the time courses of the temperature shifts are provided alongside each graph of memory performance, and periods of neurotransmission blockade are highlighted in red. Flies were transferred to the restrictive temperature (33°C) 30 min before the targeted time, to allow for acclimatization to the new temperature.

To measure innate odor avoidance towards 3-octanol or 4-methylcyclohexanol, naïve flies were placed at the convergence point of two airflows, one interlaced with 3-octanol or 4-methylcyclohexanol and the other from a bottle with paraffin oil only. The odor-interlaced side was alternated for successive groups that were tested. Odor concentrations used in this assay were the same as for memory assays. At these concentrations, both odorants are innately repulsive. The avoidance index was calculated the same way as the performance index in memory assays.

Memory scores are displayed as means ± SEM. A single value of the performance index is the average of two scores obtained from two groups of genetically identical flies conditioned in two reciprocal experiments, using either odorant as CS+, and tested consecutively in the T-maze. The indicated 'n' is the number of independent values of the performance index or avoidance index for each genotype. Memory graphs were subjected to statistical analysis using 1-way ANOVA followed by Newman-Keuls pairwise comparisons between the experimental group and its controls. ANOVA is robust against slight deviations from normal distributions or the inequality

of variances if the sample sizes are similar between groups which was the case in our experiments. Therefore, we did not systematically test our data for normality or verify variance homogeneity prior to statistical tests, but we rather adopted a uniform analysis strategy for all our data ANOVA results are given as the value of the Fisher distribution F(x,y) obtained from the data, where x is the number of degrees of freedom between groups and y is the total number of degrees of freedom of the distribution. Statistical tests were performed using the GraphPad Prism 5 software. In the figures, asterisks illustrate the significance level of the t-test, or of the least significant pairwise comparison following an ANOVA, with the following nomenclature: *: $p<0.05$; **: $p<0.01$; ***: $p<0.001$; NS: not significant, $p>0.05$).

8. Calcium Imaging: Functional Connectivity

The genetically encoded GCaMP6f calcium reporter (Chen et al., 2013) (*UAS-GCaMP6f* in *attp18*) was driven by *R37G11 GAL4 (attP2)*. The thermosensitive cation channel dTrpA1 (Hamada et al., 2008) (*LexAop2-dTrpA1 VK00005*) was expressed in the V2 neurons by the 71D08-LexA driver (attP40). Female flies of the indicated genotypes were prepared for in vivo imaging as described above, and mounted on a custom-made chamber with controlled temperature through a Peltier cell and an analog electronic PID circuit. The baseline setpoint for the temperature was 20°C. Imaging was performed on the same setup as for olfactory responses, images were acquired at a rate of one image every 640 ms. During an acquisition with thermal activation, the setpoint of the temperature control circuit was shifted to 31°C for 30 s after a baseline recording of 10 s, and then back to 20°C. The measured risetime of the temperature from 20°C to 29°C was ~8 s, and temperature reached 31°C within ~11 s. For the temperature decrease was slower, taking ~ 15s from 31°C to 22°C and ~ 25s in total to decrease down to 20°C. For each fly, three such acquisitions were recorded, and the resulting time traces from visible hemispheres and from all these recordings were pooled and averaged. In R71D08LexA>LexAop2-TrpA1 flies, acquisitions with activation were alternated with acquisitions without activation as a permissive temperature control. The magnitude of activation was calculated as the mean of the time trace over a 20 s-time windows starting 5s after the change in temperature setpoint.

9. Calcium Imaging: Olfactory Responses

To monitor the olfactory responses in PD2a1/b1 neurons, the genetically encoded GCaMP3 calcium reporter (Tian et al., 2009) was driven by R37G11 GAL4 driver. We used a transgenic line carrying the UAS-IVS-GCaMP3-p10 construct inserted on the third chromosome in VK00005 (Bouzaiane et al., 2015; Tian et al., 2009). For *in-vivo* imaging, one female fly was prepared essentially as described previously (Séjourné et al., 2011). The fly was then placed under the objective lens (25x, 0.95 NA) of a confocal microscope under a constant airflow of 1.5 L·min-1. Images were acquired at a rate of one image every 128 ms. The emitted light was collected from transverse sections of the brain showing presynaptic terminals of PD2a1/b1

neurons. In general, both hemispheres could be recorded simultaneously. Olfactory stimuli were triggered by switching a valve to direct 30% of the total flow for 2 s through bottles containing odorants diluted in paraffin oil. Final dilution in the airflow was 1:2000. We recorded two series of responses to octanol and methylcyclohexanol, in alternating order, each separated by a 2 min interval, but only the first response to each odorant was kept for analysis. Data analysis was performed with Matlab software. For each recording, a $\Delta F/F0$ time trace trace was calculated from an ROI surrounding the PD2a1/b1 projections. The baseline F0 value was calculated from the 2 s period preceding the switch of the valve. The response integral was calculated as the integral of the time trace during 10 consecutive timepoints following the onset of odor response (approx.. 2 s). The comparison of the response to a given odor between two groups, and of the response difference (Oct–Mch or Mch–Oct) between two groups, was performed using unpaired t-test.

10. Data Availability

The data supporting the findings in this study are available upon request.

11. Code Availability

The custom code can be made available upon request.

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Figure 1: PD2a1/b1 is postsynaptic to MBON-α2sc and necessary for memory retrieval

(A) Summary distribution of overlap scores for the MBON-α2sc axon mask in the LH against all the GAL4 lines in the Janelia GMR collection. Lines that scored above the .97 quantile are labelled in red. Note that the Y axis is clipped below -2000 for clarity. (B) The sparsest GAL4 line labelling cell-type PD2a1/b1, R37G11-GAL4. Image stack is from the FlyLight website (www.janelia.org/gal4-gen1). The Scale bar is 30µm. (C) Z-projection of double labelling. Note that this LexA line contains both MBON-α2sc (dorsal) and MBON-α′3ap (ventral) axons are labelled in magenta while PD2a1/b1 is labelled with membrane-bound GFP (in green). The Scale bar is 5µm. Image is representative sample from n=4. (D) Flies expressing Shi^{ts} driven by R37G11-GAL4, and their genotypic controls, were trained and tested according to the illustrated protocols (periods at the restrictive temperature are indicated in red). Silencing PD2a1/b1 neurons impaired immediate memory after single-cycle (1x) training (D': n = 12–13, $F_{(2,36)}$ = 3.79, p = 0.033), 3h-memory after single-cycle training (D'': n = 9, $F_{(2,26)}$ = 12.07, p = 0.0002), and long-term memory after spaced training (D''': n = 9, $F_{(2,26)}$ = 6.28, p = 0.0064). (E) Flies expressing Shi^{ts} driven by 37G11-GAL4 driver showed normal olfactory avoidance to octanol (Oct) and methyl-cyclohexanol (Mch) compared to their genotypic controls at the restrictive temperature (Oct: n = 14, $F_{(2,41)}$ = 2.41, p = 0.10; Mch: n = 14, $F_{(2,41)}$ = 0.23, p = 0.79). Data are presented as mean±SEM. (F) Confocal z-projection of PD2a1/b1 driving both membrane-bound GFP (green) and Synaptotagmin-HA (grey). PD2a1/b1 has been manually segmented in both channels. The image is registered with the neuropil marker nc82 (magenta) to the JFRC2013 template brain. Image is representative of n=5. (G'-G''') Representative single confocal slices of ChAT immunohistochemistry demonstrating that PD2a1/b1 neurons are cholinergic. Image is representative slice of n=4 stacks. The Scale bar is 5µm.

Figure 2: Specific control with the split-GAL4 system confirms PD2a1/b1's role in memory retrieval but not innate behaviour

(A-B) Confocal z-projections of the two split-GAL4 lines generated to target PD2a1/b1 neurons, (A) LH989 and (B) LH991. Images contain both the brain and ventral nerve chord. Green is the membrane stain (mVenus) while the neuropil immunolabelling is in magenta. Image is representative of n=2 for each genotype. Flies expressing Shi^{ts} through the split-GAL4 lines LH989 or LH991 were trained and tested according to the illustrated protocols along with their respective genotypic controls (periods at the restrictive temperature are indicated in red). (C-D) Silencing PD2a1/b1 neurons using LH989 (C: n = 14–15, $F_{(2,42)}$ = 4.13, p = 0.02) or LH991 (D: n = 18, $F_{(2,53)}$ = 7.27, p = 0.0017) impaired immediate memory after 1x training. (E)-(F) Silencing PD2a1/b1 neurons during the retrieval phase 3h after 1x training using LH989 (E: n = 14, $F_{(2,42)}$ = 6.73, p = 0.0031) or LH991 (F: n = 11–13, $F_{(2,35)}$ = 8.23, p = 0.0013) caused a memory defect. (G-H) Silencing PD2a1/b1 neurons during the retrieval phase 24h after spaced training using

LH989 (G: n = 7–9, $F_{(2,23)}$ = 9.79, p = 0.0010) or LH991 (H: n = 19–23, $F_{(2,72)}$ = 10.83, p < 0.0001) abolished performance. (I-J) Silencing PD2a1/b1 neurons using LH989 (I: Oct: n = 8–12, $F_{(2,29)}$ = 0.63, p = 0.54; Mch: n = 10, $F_{(2,29)}$ = 0.44, p = 0.65) or LH991 (J: Oct: n = 7–8, $F_{(2,22)}$ = 0.25, p = 0.78; Mch: n = 7, $F_{(2,20)}$ = 0.068, p = 0.93) had no effect on naive avoidance of Oct or Mch. Data are presented as mean±SEM.

Figure 3: MBON-α2sc is functionally connected to PD2a1/b1

(A-C) GFP reconstitution across synaptic partners (GRASP) signal in the dorsal LH (indicated by green circle) for the experimental genotype (A), and two controls. (B-C) Genotypes are indicated and missing components are represented in the schematics above each figure. Images are representative of n=3 for controls and experimental genotypes. (D) GCaMP6f was expressed in PD2a1/b1 neurons with the R37G11-GAL4 driver (scale bar: 10µm). Fluorescence of the sensor was recorded in vivo from the axonal compartment of PD2a1/b1 neurons while the temperature of the imaging chamber was shifted from 20°C to 31°C (dashed line on F, except for the blue trace). The heat-sensitive channel dTrpA1 was expressed in V2 MBONs with 71D08-LexA. (E) The calcium increase of PD2a1/b1 neurons due to thermal activation of V2 MBONs (red trace) was stronger than that due to temperature shift only in the genotypic controls (green and purple trace). (F) Quantification of calcium increase from the traces shown in F (n = 10 flies per condition except 71D08-LexA/+ (n = 8), $F_{(3,37)}$ = 9.09, p = 0.0001). Data are presented as mean±SEM.

Figure 4: PD2a1/b1 decreases its response to the CS+ after training

(A) GCaMP3 was expressed in PD2a1/b1 with the R37G11-GAL4 driver. Olfactory responses to Oct and Mch were recorded in vivo from the axonal compartment of PD2a1/b1 neurons. (B-C) In naïve flies, the calcium increase in PD2a1/b1 neurons in response to Oct was larger than to Mch (average traces from n=6 flies; t-test, p = 0.015). (D) Odor responses were recorded 3h after 1x training using Oct as CS+ (n = 19 flies), or after the corresponding unpaired control protocol (n = 20 flies) (Fig. S7A).The integral of the odor responses (D': t-test, p = 0.023) and the calculation of the difference between Oct and Mch responses (D'': t-test, p = 0.024) revealed a decreased response to the CS+ after the associative protocol. (E) Odor responses were recorded 3h after 1x training using Mch as CS+ (n = 22 flies) , or after the corresponding unpaired control protocol (n = 21 flies) (Fig. S7B). The integral of the odor responses (E': p = 0.047) and the calculation of the difference between Mch and Oct responses (E'': t-test, p = 0.041) revealed a decreased response to the CS+ after the associative protocol. (F) Odor responses were recorded 24h after 5x spaced training using Oct as CS+ (n = 9 flies), or after the corresponding unpaired control protocol (n = 11 flies) (Fig. S7A). The integral of the odor responses (F': t-test, p = 0.036) and the calculation of the difference between Oct and Mch responses (F'': t-test, p = 0.035) revealed a decreased response to the CS+ after the associative protocol. (G) Odor responses were recorded 24h after 5x spaced training using Mch as CS+ (n = 9 flies), or after the corresponding unpaired

control protocol (n = 9 flies) (Fig. S7B). The integral of the odor responses (G': t-test, p = 0.047) and the calculation of the difference between Mch and Oct responses (G'': t-test, p = 0.010) revealed a decreased response to the CS+ after the associative protocol. Data are presented as mean±SEM. The grey bar indicates the period of olfactory stimulation. The delay between the switch of the valve and the onset of calcium response corresponds to the time for the odorant to flow from the valve output to the fly's antennae.

Figure 5: PD2a1/b1 axons converge with a subset of MBONs and DANs

(A) Histogram of overlap between a mask of PD2a1/b1 dendrites and the axons of uniglomerular PNs from 47 glomeruli (along the x-axis, note the names of the glomeruli are used). The bars are colored by function assigned by previously published data. (B) Histogram of overlap between a mask of PD2a1/b1 axons and masks of the dendrites of most DANs (along the x-axis). (C) Histogram of overlap between a mask of PD2a1/b1 axons and masks of the axons of most MBONs (along the x-axis). We analysed all MBONs and DANs with overlap of more than 15%. (D-H) High resolution double labelling of PD2a1/b1 axons (labelled with GFP, green) and DAN dendrites or MBON axons (labelled with RFP, magenta). (D) PAM-β'1 dendrites (E) PAM-β'2m dendrites (F) MBON-α'2 axons (G) MBON-y2α'1 axons and (H) MBON−β'2mp axons. Scale bar is 5µm. All double labelling are representative slices from n=2 brains.

Figure 1

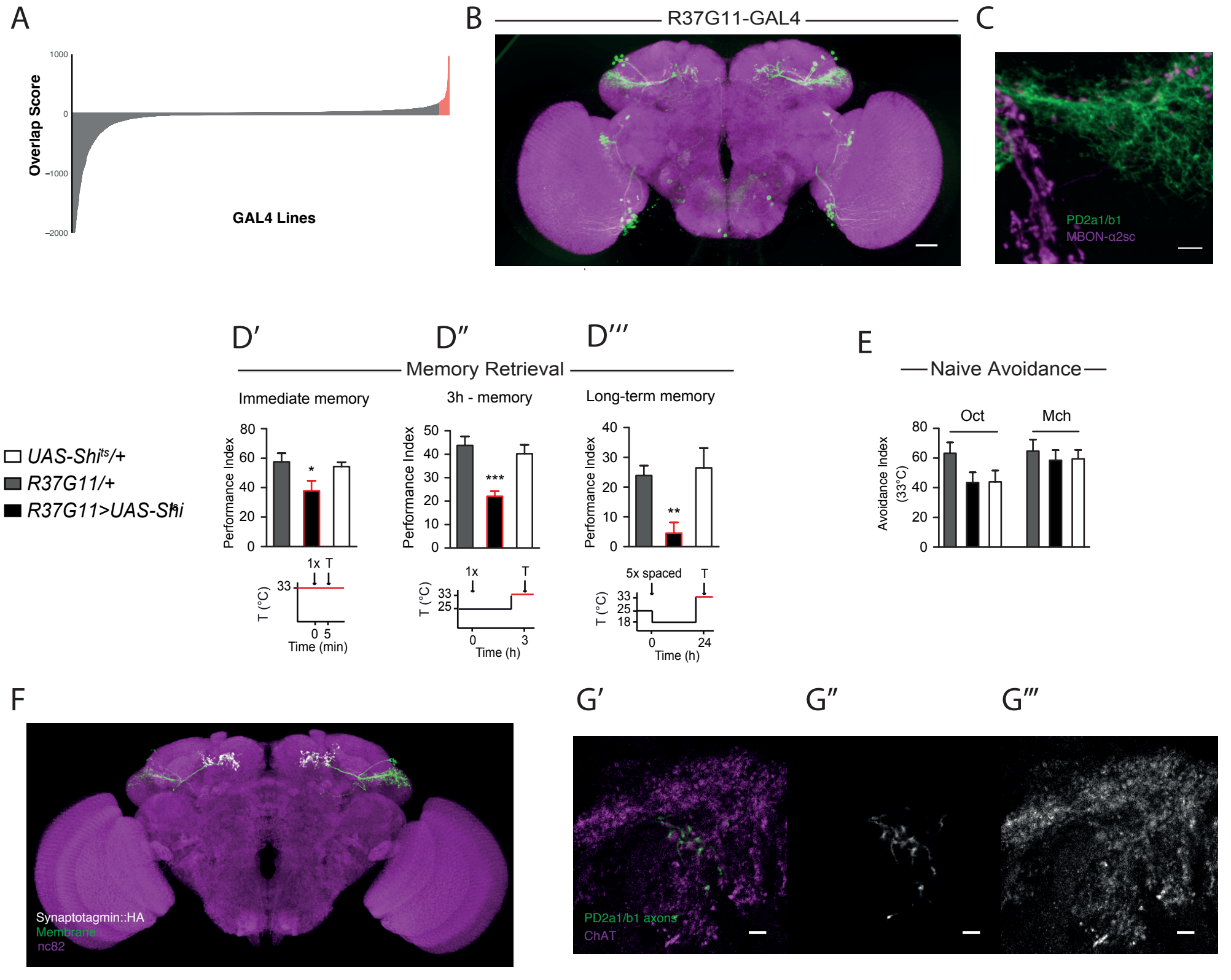

A
Overlap Score
1000
0
−1000
−2000
GAL4 Lines
B
R37G11-GAL4
C
PD2a1/b1
MBON-α2sc
D′
D″
D‴
Memory Retrieval
Immediate memory
3h - memory
Long-term memory
UAS-Shi^ts/+
R37G11/+
R37G11>UAS-Shi^ts
Performance Index
*
***
**
1x T
T (°C)
33
0 5
Time (min)
1x
T
33
25
0
3
Time (h)
5x spaced
T
33
25
18
0
24
Time (h)
E
Naive Avoidance
Oct
Mch
Avoidance Index (33°C)
F
Synaptotagmin::HA
Membrane
nc82
G′
G″
G‴
PD2a1/b1 axons
ChAT

Figure 2

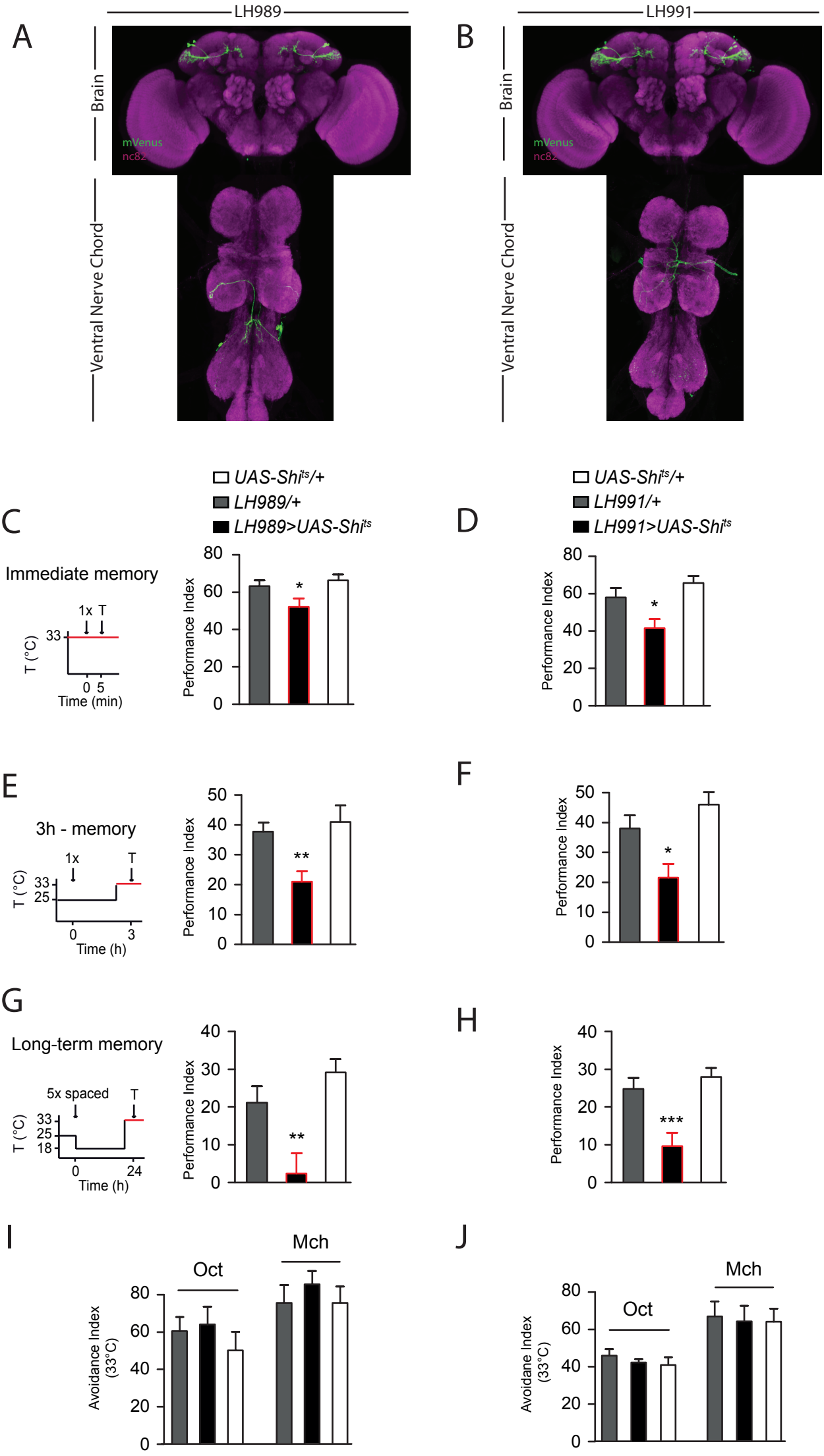
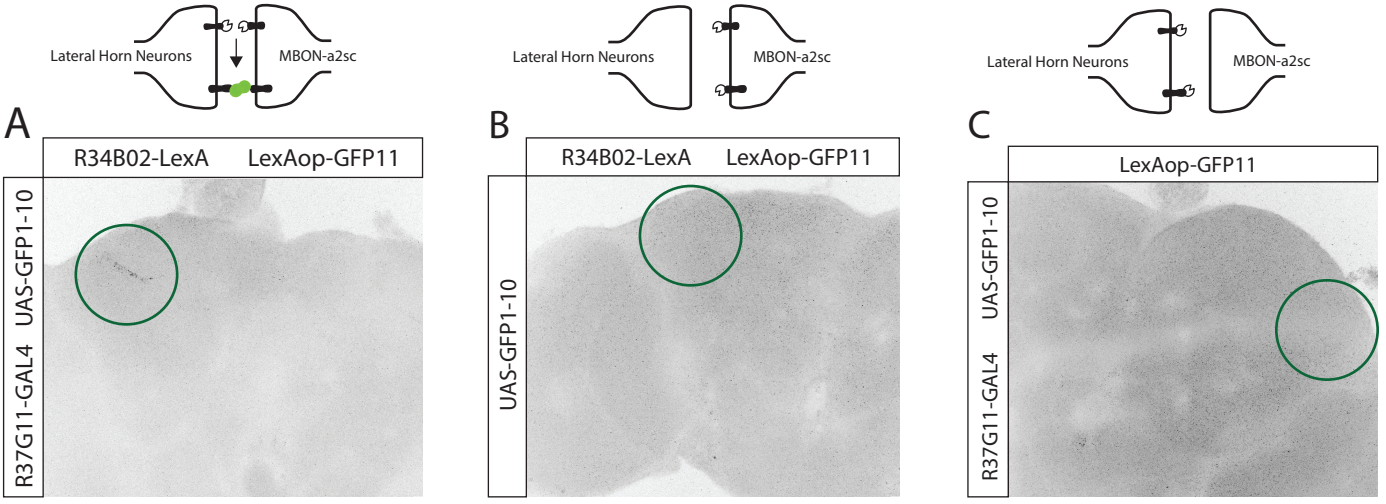

A
LH989
Brain
mVenus
nc82
Ventral Nerve Chord
B
LH991
Brain
mVenus
nc82
Ventral Nerve Chord
C
UAS-Shi^ts/+
LH989/+
LH989>UAS-Shi^ts
Immediate memory
1x T
T (°C)
33
0 5
Time (min)
Performance Index
80
60
40
20
0
*
D
UAS-Shi^ts/+
LH991/+
LH991>UAS-Shi^ts
Performance Index
80
60
40
20
0
*
E
3h - memory
1x
T
T (°C)
33
25
0
3
Time (h)
Performance Index
50
40
30
20
10
0
**
F
Performance Index
50
40
30
20
10
0
*
G
Long-term memory
5x spaced
T
T (°C)
33
25
18
0
24
Time (h)
Performance Index
40
30
20
10
0
**
H
Performance Index
40
30
20
10
0
***
I
Oct
Mch
Avoidance Index (33°C)
80
60
40
20
0
J
Oct
Mch
Avoidane Index (33°C)
80
60
40
20
0

Figure 3



Naive flies

R37G11GAL4>UAS-GCaMP6f

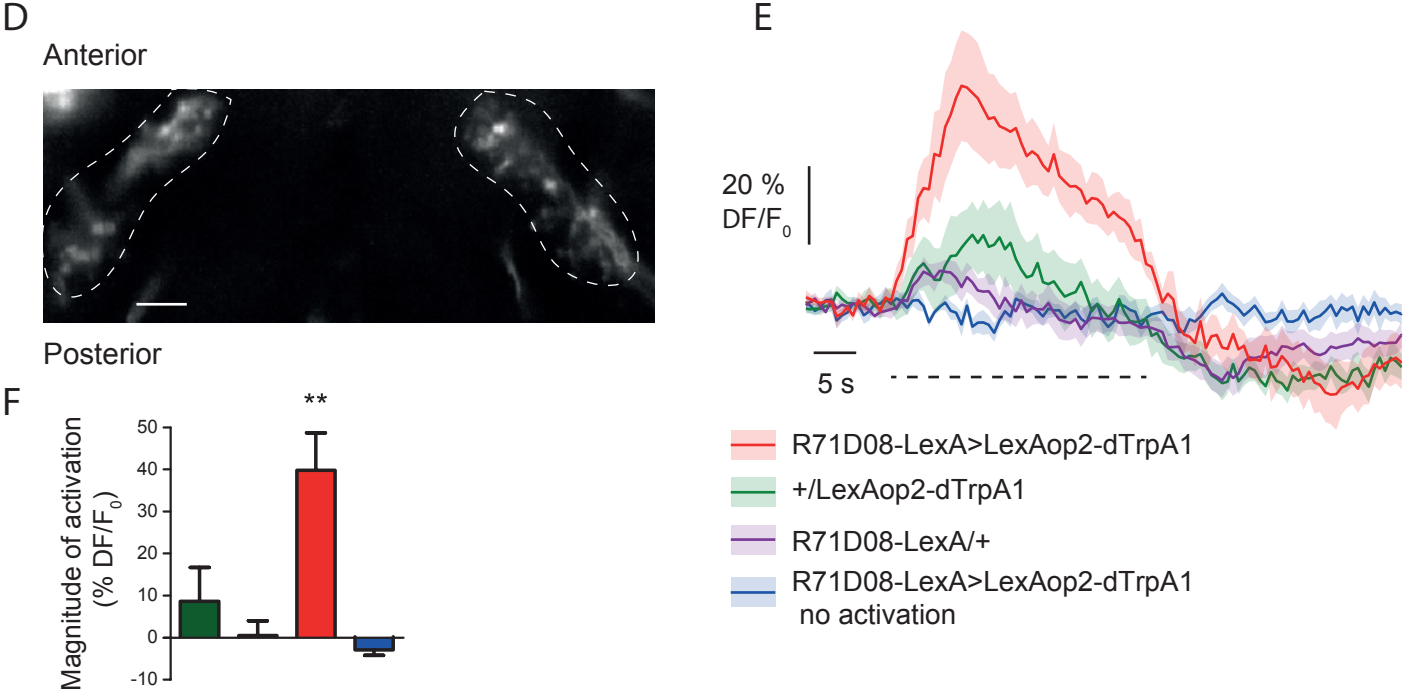


Figure 4

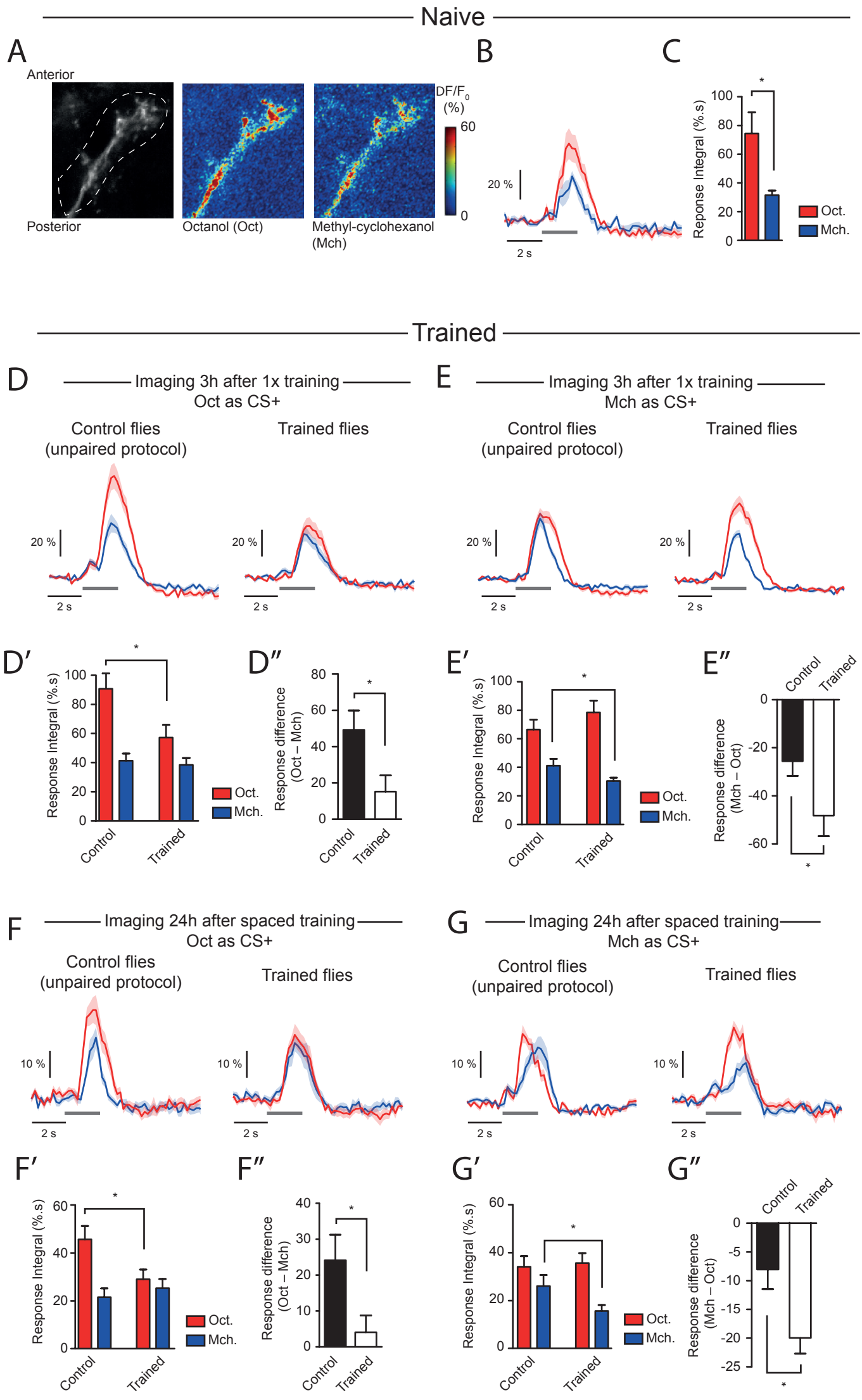
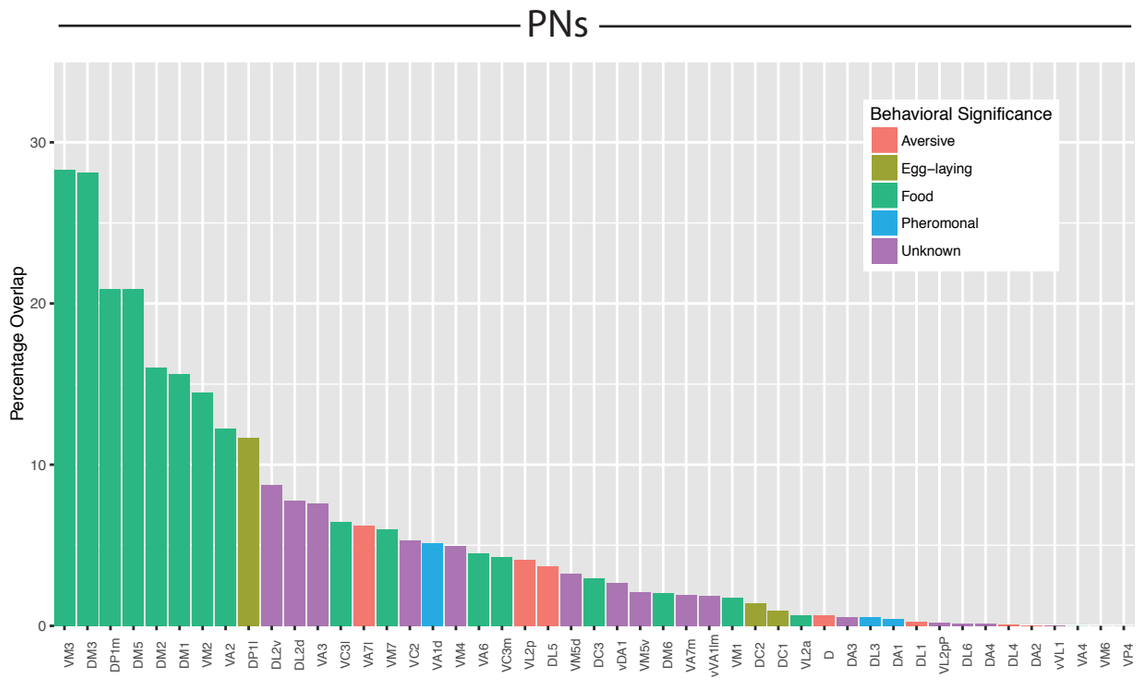

Naive
A
Anterior
Posterior
Octanol (Oct)
Methyl-cyclohexanol (Mch)
DF/F0 (%)
60
0
B
20 %
2 s
C
Reponse Integral (%.s)
Oct.
Mch.
Trained
D
Imaging 3h after 1x training
Oct as CS+
Control flies (unpaired protocol)
Trained flies
D′
Response Integral (%.s)
Control
Trained
D″
Response difference (Oct – Mch)
E
Imaging 3h after 1x training
Mch as CS+
E′
E″
Response difference (Mch – Oct)
F
Imaging 24h after spaced training
Oct as CS+
10 %
F′
F″
G
Imaging 24h after spaced training
Mch as CS+
G′
G″

Figure 5

A



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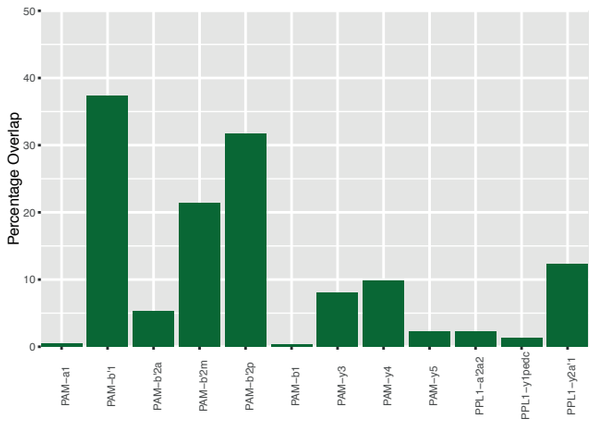

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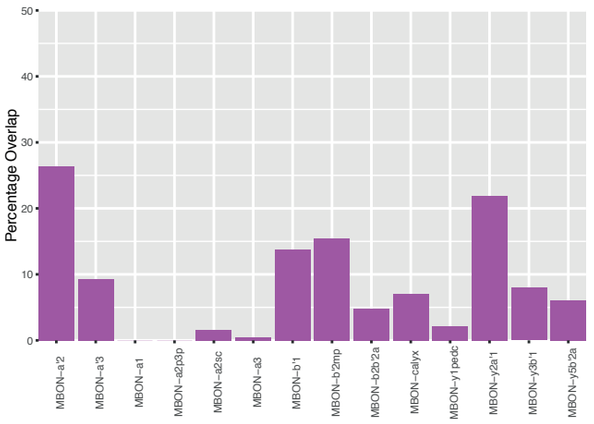

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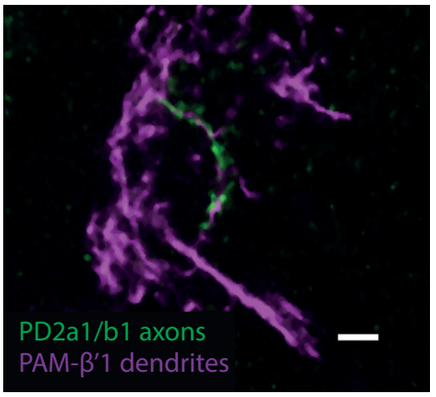

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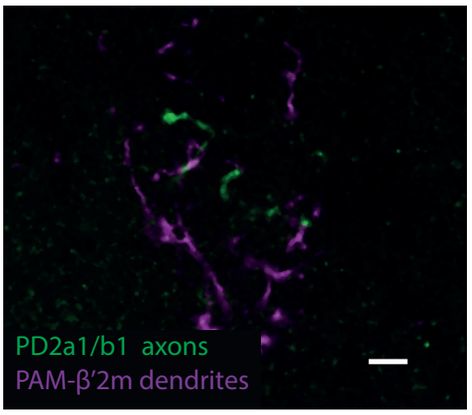

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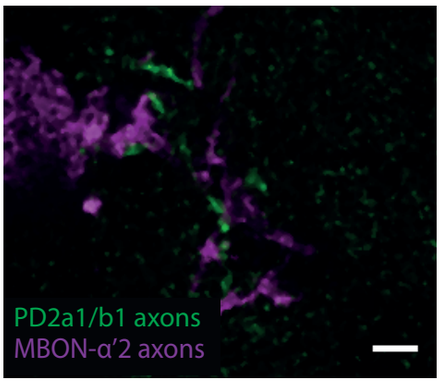

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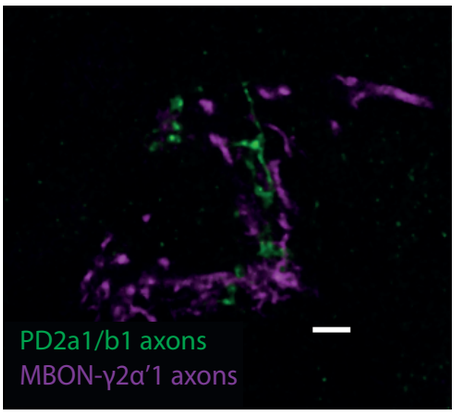
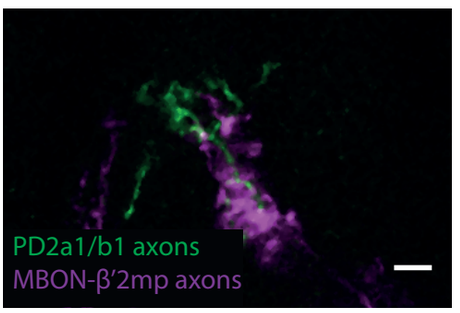
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