

1 **Catecholamines, not acetylcholine, alter**
2 **cortical and perceptual dynamics in line with**
3 **increased excitation-inhibition ratio**

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20 **Abbreviated title:** Neuromodulation and Cortical Excitation-Inhibition Balance

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34 **ABSTRACT**

35 The ratio between excitatory and inhibitory neurons (E/I ratio) is vital for cortical
36 circuit dynamics, computation, and behavior. This ratio may be under the
37 dynamic control of neuromodulatory systems, which are in turn implicated in
38 several neuropsychiatric disorders. In particular, the catecholaminergic
39 (dopaminergic and noradrenergic) and cholinergic systems have highly specific
40 effects on excitatory and inhibitory cortical neurons, which might translate into
41 changes in the local net E/I ratio. Here, we assessed and compared their net
42 effects on net E/I ratio in human cortex, through an integrated application of
43 computational modeling, placebo-controlled pharmacological intervention,
44 magnetoencephalographic recordings of cortical activity dynamics, and
45 perceptual psychophysics. We found that catecholamines, but not acetylcholine,
46 altered both the temporal structure of intrinsic activity fluctuations in visual and
47 parietal cortex, and the volatility of perceptual inference based on ambiguous
48 visual input. Both effects indicate that catecholamines increase the net E/I ratio in
49 visual and parietal cortex.

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60 INTRODUCTION

61 Cortical activity fluctuates continuously, even in the absence of changes in
62 sensory input or motor output (1). These intrinsic fluctuations in cortical activity
63 are evident from the level of single neurons to large-scale networks of distant
64 cortical areas (2–4). Fluctuations in cortical mass activity, specifically the
65 amplitude modulation of ongoing oscillations, exhibit temporal structure
66 characteristic of so-called “scale-free” behavior: Power spectra that scale as a
67 function of frequency according to a power law, $P(f) \propto f^{-\beta}$ (5,6), and long-range
68 temporal autocorrelations (7–10). This temporal structure of cortical activity varies
69 widely across individuals, is partly explained by genetics (11), and it exhibits
70 marked changes in brain disorders (12,13).

71 The large variability of cortical activity is not only due to the biophysics of
72 individual cells (1), but also due to the balance between excitatory and inhibitory
73 inputs to each neuron (2,14). The ratio between excitatory and inhibitory
74 interactions in local cortical circuits, henceforth referred to as E/I ratio, is also
75 essential for the characteristic structure of spontaneous cortical activity (15,16).
76 For example, structural variations of excitatory and inhibitory connectivity affect
77 the temporal structure of activity fluctuations in a model of a local cortical circuit
78 (15). Finally, the E/I ratio is also a key determinant of the computational
79 properties of individual cortical neurons (17,18) as well as the behavior of the
80 organism, as shown for perceptual categorization tasks (16,18–21).

81 This key property of cortical circuits, E/I ratio, might not be a fixed
82 property of cortex, but rather under dynamic control. One factor in particular
83 might be key for regulating cortical E/I ration and thus cortical variability as well
84 as behavior: dynamic variations in neuromodulatory tone (22). Modulatory
85 systems of the brainstem regulate cortical state through widespread ascending

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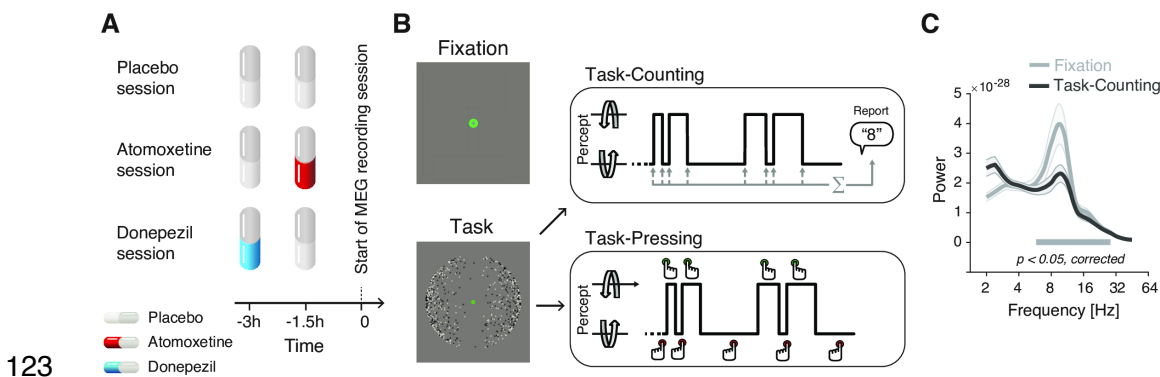
86 projections, and they are implicated in most of the major neuropsychiatric
87 disorders (17,23–26). The modulatory neurotransmitters released from these
88 systems, such as noradrenaline or acetylcholine, alter specific elements
89 (pyramidal cells or inhibitory interneurons) of cortical microcircuits (27,28) as well
90 as the variability of cortical neurons (27,29,30). Critically, whether and how
91 neuromodulatory systems change the net E/I ratio and ongoing activity
92 fluctuations within local populations of cortical neurons has remained unknown. A
93 systematic, empirical assessment of the net effects on cortical E/I ratio in human
94 cortex would be key for understanding how synaptic and cellular effects of
95 neuromodulation translate into changes in human cognition and behavior, as well
96 as into disturbances thereof in brain disorders. However, inferences on cortical
97 net E/I ratio based on standard “resting-state” measurements of human cortical
98 population activity have, so far, been challenging.

99 Here, we aimed to overcome this challenge through the integrated
100 application of computational modeling, magnetoencephalographic (MEG)
101 recordings of fluctuations in cortical population activity under different
102 pharmacological interventions and “steady-state” task conditions, and
103 psychophysical measurements of bistable perceptual dynamics that are sensitive
104 to cortical E/I ratio (21,31,32). This integrative approach enabled us to
105 systematically image and compare the effects on the cortical net E/I ratio of two
106 major groups of neuromodulatory systems: the catecholaminergic (noradrenergic
107 and dopaminergic) and cholinergic systems. Importantly, we read out their effects
108 on cortical net E/I ratio from two separate measurements: changes in the intrinsic
109 fluctuations in cortical activity and of bistable perceptual dynamics. Both yielded
110 convergent evidence for an increase of net E/I ratio in visual and parietal cortex
111 due to catecholamines, but not acetylcholine.

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113 RESULTS

114 We tested for changes in intrinsic perceptual and cortical dynamics under
115 placebo-controlled pharmacological manipulations of catecholamine (using
116 atomoxetine) and acetylcholine (using donepezil) levels (Fig 1A). Importantly,
117 intrinsic fluctuations in cortical activity were measured during two steady-state
118 conditions (Fig 1B): (i) fixation of an otherwise gray screen (Fixation), as in most
119 common studies of human “resting-state” activity; and (ii) silent counting of the
120 spontaneous perceptual alternations induced by a continuously presented,
121 ambiguous visual stimulus (Task-counting). In a third condition, subjects
122 immediately reported the perceptual alternations by button-press (Task-pressing).



124 **Fig 1.** Experimental design **(A, B)** Types and time course of experimental sessions. **(A)** Each
125 subject participated in three sessions, involving administration of placebo, atomoxetine, or
126 donepezil (session order randomized across subjects). Each session entailed the administration
127 of two pills, in the order depicted for the different session types. **(B)** Within each session,
128 subjects alternated between three conditions, Fixation, Task-Counting and Task-Pressing, during
129 which MEG was recorded (runs of 10 min each). See Materials and Methods for details. **(C)** Group
130 average power spectrum, averaged across all MEG sensors, for Rest and Task (Placebo condition
131 only).

132

133 This design capitalized on recent insights into the changes in cortical E/I-
134 ratio under sensory stimulation (33,34) and on the effects of cortical E/I-ratio on
135 bistable perceptual dynamics (21,31,32). These previous insights and our
136 experimental data combined, allowed for interpreting the latter in terms
137 alterations in net cortical E/I ratio under the pharmacological treatments.

138 To solidify our predictions about the impact on modulations of E/I ratio on
139 the intrinsic correlation structure of cortical population activity, we also simulated
140 the population activity of a simplified cortical circuit model made up of recurrently
141 connected excitatory and inhibitory neurons, under systematic variations of gain
142 modulation at different synapse types.

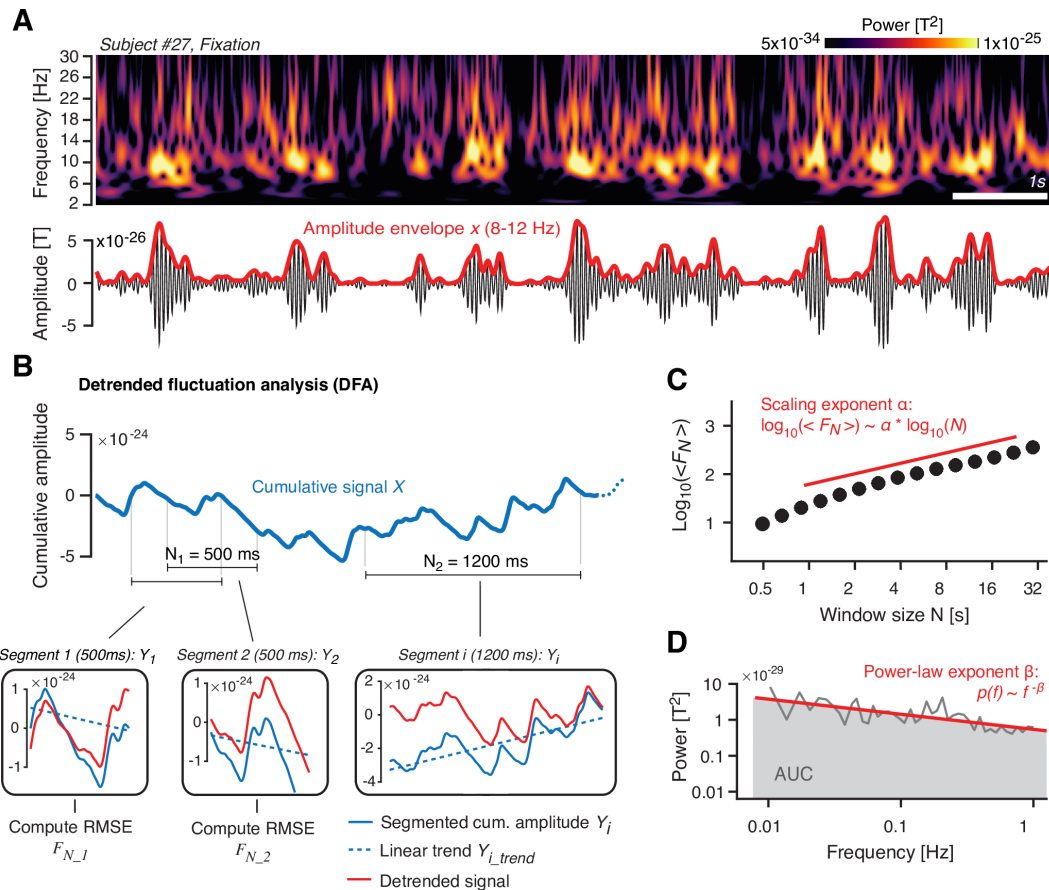
143 The Results section is organized as follows. We first present the effects of
144 the drugs on perceptual alternation rate. We then show how dynamic variations
145 of E/I ratio due to synaptic gain modulation alter intrinsic fluctuations in the
146 amplitude of cortical oscillations of a cortical circuit model. Next, we show how
147 manipulating catecholaminergic and cholinergic neuromodulation, affects
148 fluctuations in cortical activity—specifically, the temporal correlation structure of
149 intrinsic fluctuations in the amplitude of cortical oscillations (Fig 2), during both
150 steady-state conditions (Fixation and Task-counting). Finally, we discuss the drug
151 effects on other measures of cortical activity as well as peripheral signals. These
152 controls support the validity and specificity of our main conclusions.

153

154 **Atomoxetine increases the rate of perceptual alternations compared to**
155 **placebo and donepezil**

156 We used the rate of the reported alternations in perception of the ambiguous
157 visual structure-from-motion stimulus (Fig 1B) as a behavioral proxy for changes
158 in cortical E/I ratio in visual cortex. Current models of the neural dynamics
159 underlying bistable perception postulate that such perceptual alternations emerge
160 from the interplay between feedforward drive of stimulus-selective neural
161 populations in sensory cortex, mutual inhibition between them, adaptation, and
162 noise (31,32). Convergent evidence from model simulations (21) as well as
163 functional magnetic resonance imaging, magnetic resonance spectroscopy, and

164 pharmacological manipulation of GABAergic transmission (21,35) indicates that
 165 increases in the ratio between feedforward, excitatory input to, and mutual
 166 inhibition within the cortical circuit give rise to faster perceptual alternation rates.
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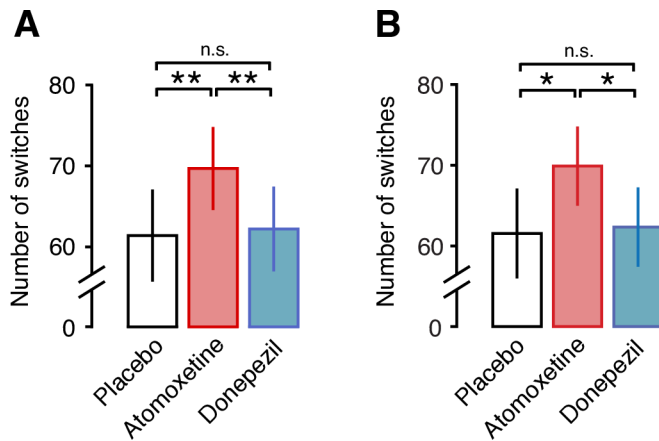
169 **Fig 2. Quantifying the temporal structure of fluctuations in oscillatory cortical activity**

170 **(A) Top.** Time-frequency representation of MEG power fluctuations during Rest (example subject).
 171 **Bottom.** Filtered signal (10 Hz; black) and the corresponding amplitude envelope (red). **(B)**
 172 Illustration of detrended fluctuation analysis. See main text (Materials and Methods) for details. **Top.**
 173 Cumulative sum of the amplitude envelope. **Bottom.** Detrending of cumulative sum within segments,
 174 shown for two different window lengths N ($N_1 = 500$ ms and $N_2 = 1200$ ms). **(C)** Root-mean-square
 175 fluctuation function $\langle F_N \rangle$. In log-log coordinates, $\langle F_N \rangle$ increases approximately linearly as a
 176 function of N , with a slope that is the scaling exponent α . **(D)** Illustration of power spectrum analysis
 177 of amplitude envelope. In log-log coordinates, the power spectrum can be approximated by a
 178 straight line, with a slope β (power-law exponent) and an area under the curve (gray) that quantifies
 179 the overall variance of the signal.

180

181 In this study, atomoxetine increased the rate of perceptual alternations compared
 182 to both, placebo and donepezil (Fig 3A; atomoxetine vs. placebo: $p = 0.007$; $t =$
 183 2.913 ; atomoxetine vs. donepezil: $p = 0.001$; $t = 3.632$; donepezil vs. placebo: $p =$
 184 0.966 ; $t = -0.043$; all paired t-tests, pooled across Task-counting and Task-

185 pressing). This atomoxetine effect on the perceptual dynamics was also
186 significant for Task-counting ($p = 0.045$; $t = 2.103$; paired t-test; Fig S1A) and
187 Task-pressing ($p = 0.018$; $t = 2.540$; paired t-test; Fig S1B) individually, and the
188 perceptual alternation rates were highly consistent across both conditions (Fig
189 S1C).



190

191 **Fig 3.** Atomoxetine, but not donepezil, increases the rate of perceptual alternations **(A)** Number of
192 perceptual alternations reported by the subjects per 10 min run, pooled across task conditions
193 (Task-counting and Task-pressing). **(B)** Same as (A), after removing blink and eye movement data
194 (with linear regression).
195

196 One potential concern is that atomoxetine might have increased the rates
197 of spontaneous eye blinks or fixational eye movements, inducing retinal
198 transients and thus fluctuations in visual cortical activity and perception, without
199 any change in intra-cortical E/I ratio. Three observations rule out this concern.
200 First, there was no significant increase during atomoxetine compared to placebo
201 in any of five different eye movement parameters measured here (Fig S2).
202 Second, none of the eye movement parameters correlated significantly with the
203 perceptual alternation rate (Fig S2). Third, and most importantly, the effect of
204 atomoxetine on the perceptual dynamics was also significant after removing (via
205 linear regression) the individual eye movement parameters (Fig 3B).

206 In sum, the psychophysical results are consistent with an atomoxetine-
207 induced increase in the net E/I ratio. This change should have occurred in cortical

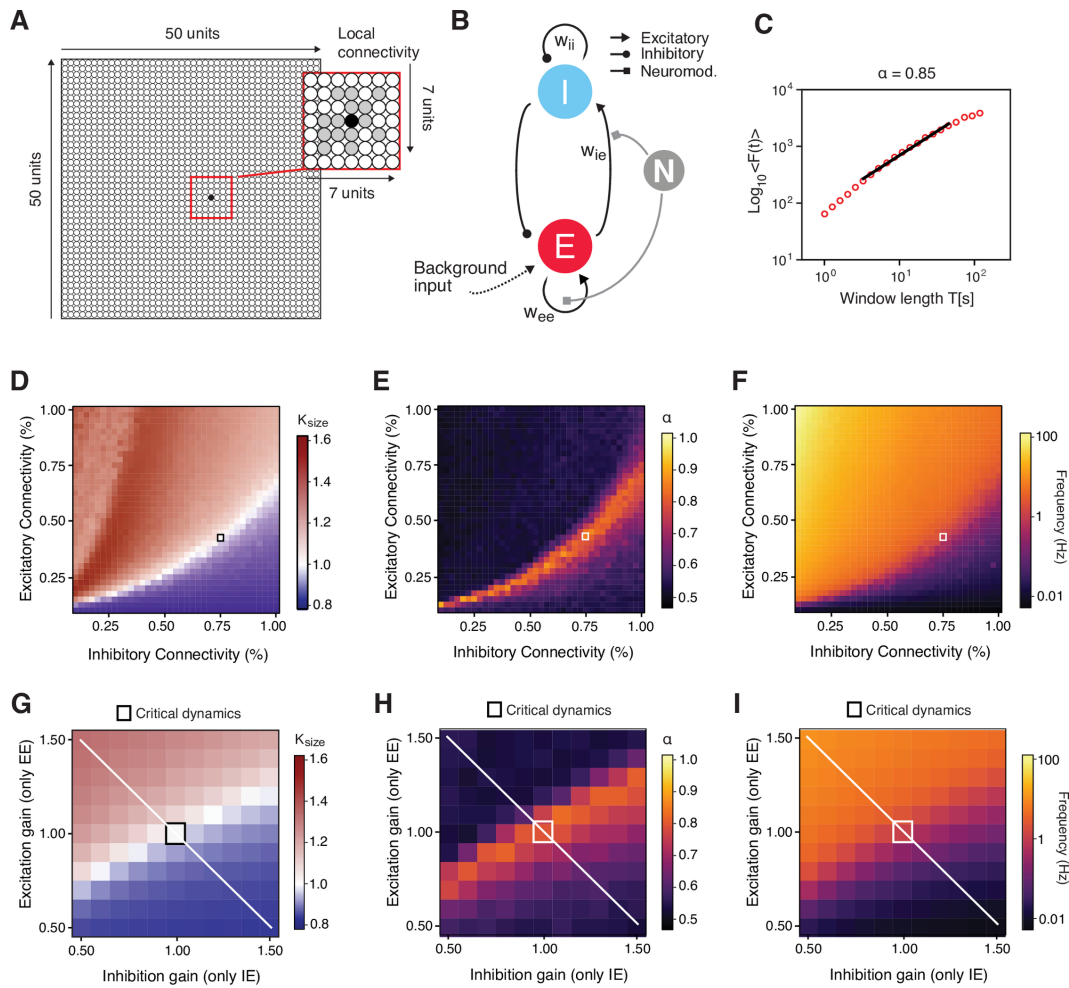
208 circuits within the dorsal visual stream that govern the perceptual dynamics of
209 ambiguous structure-from-motion signals (36).

210

211 **Effects of synaptic gain modulation on scaling behavior in a cortical circuit**
212 **model**

213 We used the temporal correlation structure of fluctuations in cortical activity as a
214 separate read-out of changes on cortical E/I ratio, guided by simulations of
215 cortical circuit models under neuromodulation. The models of bistable perception
216 discussed above are sufficient for generating perceptual time courses, but are not
217 sufficiently realistic to generate the features of cortical mass activity evident in
218 physiological recordings of local field potentials or MEG signals (e.g., alpha-band
219 oscillations, scale-free amplitude envelope fluctuations). We used a more
220 complex cortical circuit model that does exhibit these features (15) as a starting
221 point for our modeling work (Fig 4). The model has previously been used to show
222 that scale-free intrinsic fluctuations in cortical activity are highly sensitive to
223 variations in the structural E/I ratio (i.e., the percentage of excitatory and
224 inhibitory connections) in the circuit (15). This model accounts for the joint
225 emergence of two empirically established scale-free behaviors, which we
226 reproduced: (i) neuronal avalanches, activity patterns propagating through the
227 network as evident in recordings from microelectrode arrays, with an event size
228 distribution following a power-law (37); and (ii) long-range temporal correlations
229 of the amplitude envelope fluctuations of the model's local field potential, which
230 we assessed empirically through MEG recordings. The power-law scaling of
231 avalanche size distribution was quantified in terms of the kappa-index, which
232 quantifies the similarity between the measured avalanche size distribution and a

233 theoretical power-law distribution with an exponent of -1.5 (38); a kappa index of
 234 1 indicates perfect match between the two.



235

236 **Fig 4.** Dynamic modulation of excitation-inhibition ratio alters long-range temporal correlations in
 237 model of cortical patch. **(A)** Schematic of the computational model. The network consisted of 2500
 238 excitatory and inhibitory integrate-and-fire units and random, local (within an area of 7x7 units)
 239 connectivity (magnified within the red square). **(B)** Neuromodulation was simulated as a gain
 240 modulation term multiplied with excitatory synaptic weights (w_{ee} and w_{ie}). **(C)** Detrended fluctuation
 241 analysis of model simulation (scaling exponent α of 0.85). **(D)** κ as a function of excitatory and
 242 inhibitory connectivity (with a spacing of 2.5%; means across 10 simulations per cell). The region of
 243 $\kappa \sim 1$, overlaps with the region of $\alpha > 0.5$ and splits the phase space into an excitation-dominant
 244 ($\kappa > 1$) and an inhibition-dominant region ($\kappa < 1$). The black square depicts the network configuration
 245 that was chosen for assessing the effects of neuromodulation **(E)** Scaling exponent α as a function
 246 of excitatory and inhibitory connectivity. **(F)** Same as (D) and (E), but for mean firing rate. **(G)** κ as a
 247 function of independent synaptic gain modulation. **(H)** Same as (G), but for scaling exponent α . **(I)**
 248 Same as (G), but for firing rate. Red square, baseline state of critical network before
 249 neuromodulation was applied. White line, axis of parameter combinations corresponding to
 250 changes in excitation-inhibition ratio re-plotted schematically in Fig 8.

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The two phenomena unfold on different scales of spatial resolution (single
 253 neurons vs. mass activity summed across neurons) and different temporal scales
 254 (tens of milliseconds vs. several hundred seconds). Yet, both phenomena have

255 been found to emerge at the same ratio between structural excitatory and
256 inhibitory connectivity (15), and we replicated this finding here (Fig 4D-F).

257 Critically, we extended this model with a modulatory mechanism in order
258 to assess the impact of dynamic, multiplicative changes in cortical E/I ratio that
259 might result from catecholamines or acetylcholine. We first determined the
260 structural connectivity (small squares in Fig 4D-F) and the time scale parameters
261 such that the network generated intrinsic alpha-band oscillations with amplitude
262 fluctuations that exhibited robust long-range temporal correlations (with $\alpha \sim 0.85$,
263 Fig 4C), as well as neuronal avalanches with scale-free size distributions
264 (Materials and Methods). We then independently modulated synaptic connections
265 through multiplicative scaling of the weights (as illustrated in Fig 4B).

266 Two separate versions of the synaptic gain modulation yielded
267 qualitatively similar effects. In the first version shown in Fig 4, we modulated only
268 excitatory synapses, but independently on excitatory as well as inhibitory neurons
269 (EE and IE), thus producing asymmetries in the circuits net E/I ratio as in recent
270 modeling work on the effects of E/I ratio on a cortical circuit for perceptual
271 decision-making (18). In the second version (Fig S3A), we co-modulated EE and
272 IE and independently modulated inhibitory synapses on excitatory neurons (EI).
273 This was intended to simulate modulations of the GABA receptors in the former
274 case (mediating the effects of inhibitory neurons on others), as opposed (AMPA
275 or NMDA) glutamate receptors in both of the latter two cases (mediating the
276 effects of excitatory neurons on others). N_{EE} and N_{IE} were co-modulated by the
277 same factor for simplicity, because we did not assume that excitatory
278 (glutamatergic) synapses would be differentially modulated depending on
279 whether they were situated on excitatory or inhibitory target neurons.

280 In both versions of the model, changes in net E/I ratio altered κ (Fig 4G
281 and Fig S3B) as well as the scaling exponent α (Fig 4H and Fig S3C) and mean
282 firing rate (Fig 4I and Fig S3D). Importantly, the effect of changes in E/I ratio on
283 the scaling exponent α were non-monotonic, dependent on the starting point:
284 increases in excitation led to increases in α when starting from an inhibition-
285 dominant point, but to decreases in α when starting from an excitation-dominant
286 point (Fig 4G-I, white line).

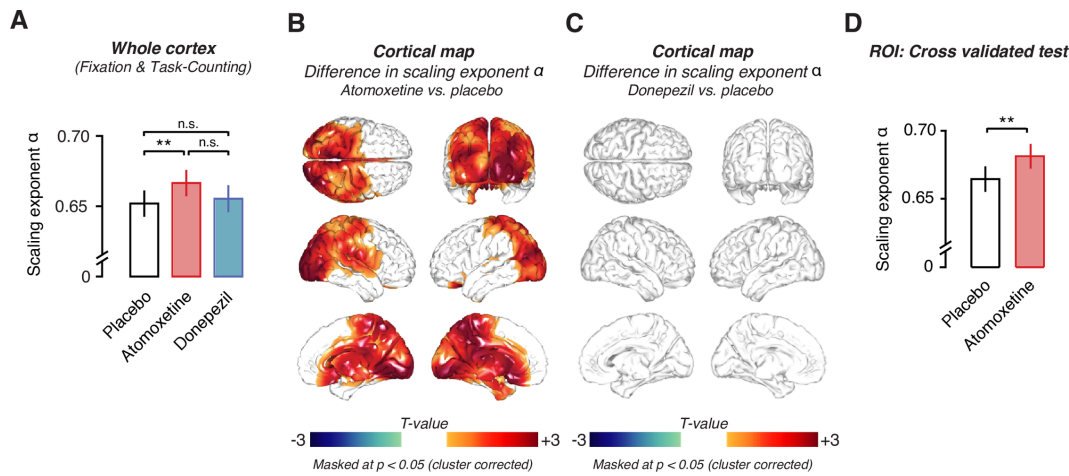
287 The effects of excitatory and inhibitory gain modulation on the temporal
288 correlation structure of the simulated activity were qualitatively similar to the
289 effects of (structural) changes in the fraction of excitatory and inhibitory synapses
290 simulated (as shown in Fig 4D-F). We conceptualize the latter as simulations of
291 individual differences in cortical anatomical microstructure, and the former as
292 simulations of within-subject, state-dependent changes in cortical dynamics,
293 which are the focus of the current study. The new simulation results provided a
294 solid foundation for the interpretation of the pharmacological effects on
295 fluctuations of alpha-band amplitude envelope signals in human MEG data,
296 described next.

297

298 **Atomoxetine, not donepezil, increases the scaling exponent of cortical** 299 **activity**

300 We found a subtle, but robust and highly consistent increase in the scaling
301 exponent α of fluctuations in human MEG under atomoxetine, but not donepezil
302 (Fig 5 and Fig 6). We focused our analyses on amplitude envelope fluctuations in
303 the 8–12 Hz frequency range (“alpha band”), for two reasons. First, as expected
304 from previous work (39), the cortical power spectra exhibited a clearly discernible
305 in this frequency range, which robustly modulated with task conditions

306 (suppressed under Task-counting, Fig 1C). Second, the parameters of the above
 307 model were tuned to produce oscillations in the same range (see above and (15)).



308
 309

310 **Fig 5.** Scaling exponent α for the pharmacological conditions, pooled across Fixation and Task-
 311 counting conditions. **(A)** Mean scaling exponent across all voxels ($N = 3000$) for all three
 312 pharmacological conditions. Compared to placebo, the exponent exhibits a significant increase
 313 under atomoxetine, but not under donepezil. **(B, C)** Spatial distributions of drug-induced changes
 314 (threshold: at $p = 0.05$, two-sided cluster-based permutation test). **(B)** atomoxetine vs. placebo; **(C)**
 315 donepezil vs. placebo. **(D)** Cross-validation approach, see Results for details.

316

317 The average scaling exponent across cortical patches and participants

318 during Fixation (placebo only) was $\alpha = 0.67$ ($\sigma = 0.09$) and during Task-counting

319 (placebo only) $\alpha = 0.64$ ($\sigma = 0.07$), indicative of robust long-range temporal

320 correlations during both behavioral contexts. Averaged across all cortical voxels

321 and across Fixation and Task-counting conditions, there was a highly significant

322 increase in α ($p = 0.0068$; $t = 2.93$; paired t-test) under atomoxetine ($\alpha = 0.67$, σ

323 $= 0.05$), compared to placebo ($\alpha = 0.65$, $\sigma = 0.05$; Fig 5A). There was no

324 evidence for any effect of donepezil ($\alpha = 0.66$, $\sigma = 0.05$) compared to placebo (p

325 $= 0.50$; $t = 0.68$; $bf = 0.68$; paired t-test; Fig 5A). The increase in scaling exponent

326 α under atomoxetine was widespread, but not homogenous across cortex,

327 comprising occipital and posterior parietal as well as a number of cortical regions

328 in the midline (Fig 5B, $p = 0.0022$; cluster-based permutation test).

329 The atomoxetine effect was, although subtle, highly reproducible across

330 runs. We tested this using a cross-validation approach. We first obtained a set of

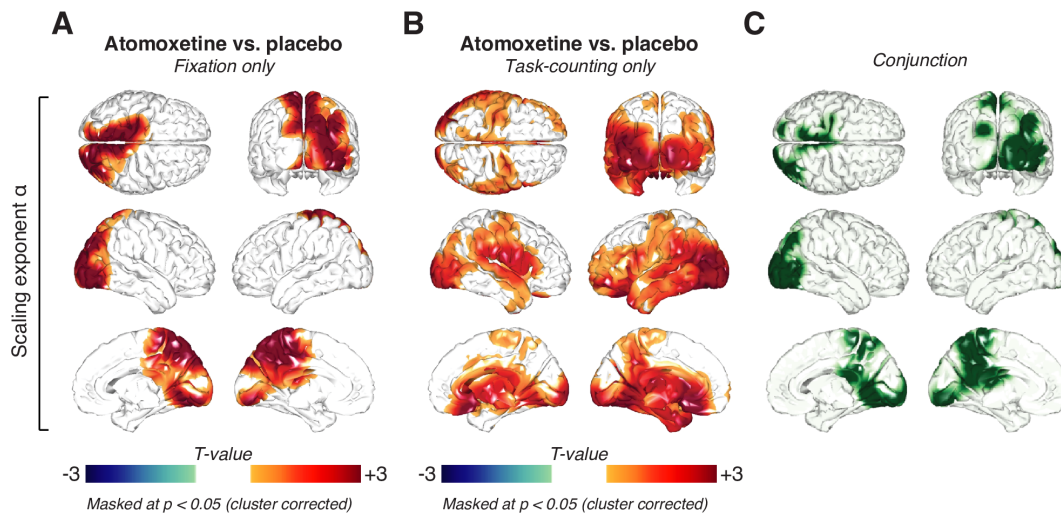
331 voxels that were significantly increased under atomoxetine compared to placebo

332 (paired t-test, $p < 0.05$) during run 1 (averaged across the two behavioral
333 contexts, Fixation and Task-counting). Next, we extracted the average scaling
334 exponents across subjects for both conditions (atomoxetine and placebo) from
335 run 2. We repeated the procedure with a set of voxels obtained from run 2 and
336 extracted the scaling exponents from run 1. This unbiased approach reveals a
337 highly significant increase in scaling exponent α after the administration of
338 atomoxetine compared to placebo ($p = 0.0023$; $t = 3.365$; Fig 5D).

339 Repeating the spatial comparison separately for Fixation and Task-
340 counting yielded significant effects of atomoxetine on α during both behavioral
341 contexts (Fig 6A, Fixation: $p = 0.0245$; Fig 6B, Task-counting: $p = 0.0035$; cluster-
342 based permutation test). The significant atomoxetine effects occurred in largely
343 overlapping posterior cortical regions (Fig 6C). Conversely, we found no evidence
344 for a significant interaction between the effects of atomoxetine and task anywhere
345 in cortex: A direct comparison of the atomoxetine vs. placebo contrast maps
346 between Fixation and Task-counting yielded no significant clusters ($p > 0.081$ for
347 all clusters; cluster-based permutation test). Taken together, these results
348 indicate that the effects of atomoxetine were largely independent of sensory drive
349 and behavioral context.

350
351 By contrast, we found no significant effect of donepezil on α in any
352 cortical region ($p > 0.22$ for all clusters; cluster-based permutation test; Fig 5C).
353 Further, no effects were evident for donepezil, when splitting by task conditions
354 (Fig S4). The control analyses presented below establish clear effects of
355 donepezil on both cortical activity as well as markers of peripheral nervous
356 system activity, thus ruling out concerns that the drug may have been less
357 effective overall than atomoxetine (see Discussion).

358



359

360 **Fig 6.** Atomoxetine increases long-range temporal correlations irrespective of behavioral condition.
361 Spatial distribution of the atomoxetine-induced changes in scaling exponent α during **(A)** Fixation
362 and **(B)** Task-counting. **(C)** Conjunction of maps in (A) and (B), highlighting (in green) voxels with
363 significant increases in both conditions.
364

365 **Decreased scaling exponent of cortical activity during Task-counting**

366 The cortex-wide scaling exponent α was significantly larger during Fixation than
367 during Task-counting ($p = 0.0062$; $t = 2.97$; paired t-test; placebo condition only).

368 This difference was significant across large parts of cortex ($p < 0.05$; cluster-
369 based permutation test; Fig 7A). The task-related decrease was also observed

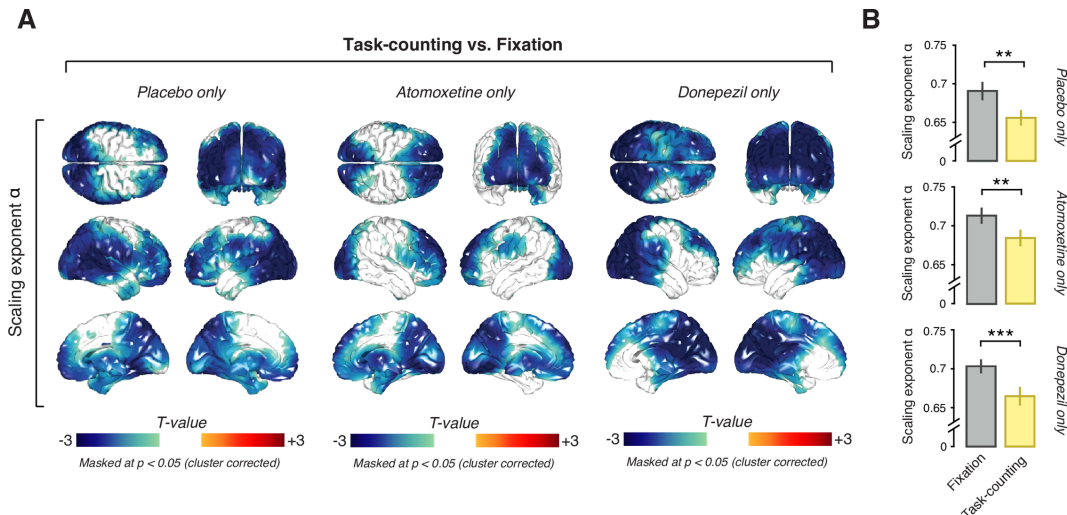
370 consistently across all pharmacological conditions (Fig 7A). Importantly, the
371 regions exhibiting significant decreases during Task-counting included the

372 occipital and parietal regions that were driven by the moving stimulus and
373 exhibited atomoxetine-induced changes in scaling behavior. Indeed, when testing

374 for the task-dependent change in scaling exponent specifically in those regions
375 showing a significant atomoxetine effect, the reduction during Task-counting was

376 also highly significant (Fig 7B).

377



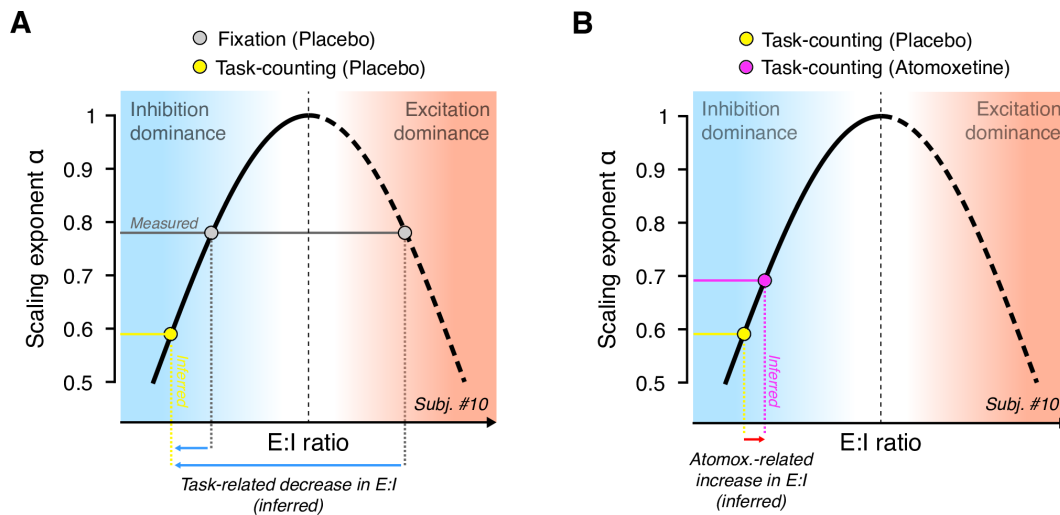
378

379 **Fig 7.** Decreased long-range temporal correlations under Task-counting **(A)** Difference in scaling
 380 exponent α between Task-counting and Fixation. *Left:* Contrast only for Placebo condition. *Middle.*
 381 Contrast only for Atomoxetine condition. *Right.* Contrast only for Donepezil condition. **(B)** Scaling
 382 exponent α for Fixation (purple) and Task-counting (yellow) conditions, averaged across voxels
 383 comprising the conjunction cluster depicted in Fig. 6C for placebo only (*Top*), atomoxetine only
 384 (*Middle*) and donepezil only (*Bottom*).
 385

386 **Change in scaling exponent under atomoxetine is consistent with increase**
 387 **in net cortical E/I ratio**

388 In our model, the scaling exponent α exhibited a non-monotonic dependence on
 389 excitation-inhibition ratio (see the white diagonal line in Fig 4G-I and schematic
 390 depiction in Fig 8). Consequently, without knowing the baseline state, any change
 391 in α is ambiguous with respect to the direction of the change in E/I ratio (i.e.,
 392 towards excitation- or inhibition-dominance). Thus, the observed increase in α
 393 under atomoxetine during Fixation could have been due to either an increase or a
 394 decrease in E/I ratio. However, recent insights into the changes in visual cortical
 395 E/I ratio during sensory drive in rodents help constrain the baseline state during
 396 the Task-counting condition: In the awake state, counter-intuitively, sensory drive
 397 decreases E/I ratio in primary visual cortex (33,34). Assuming that the same
 398 holds in human cortex during the Task-counting condition this insight enabled us
 399 to infer the change in net cortical E/I ratio induced by atomoxetine during Task-
 400 counting.

401 The rationale is illustrated in Fig 8. The observed decrease in α during
 402 Task-counting compared to Fixation (Fig 7A) was likely due to a shift towards
 403 inhibition-dominance (yellow point in Fig 8A). Then, the atomoxetine-induced
 404 increase in α during this condition was likely due to an increase in net E/I ratio
 405 during Task-counting (Fig 8B) – the same conclusion inferred from the increase
 406 in the rate of perceptual alternations above. Because the effects of atomoxetine
 407 on α were the same during Task-counting and Fixation, it is likely that the same
 408 mechanism was at play during Fixation, where the baseline state was unknown.

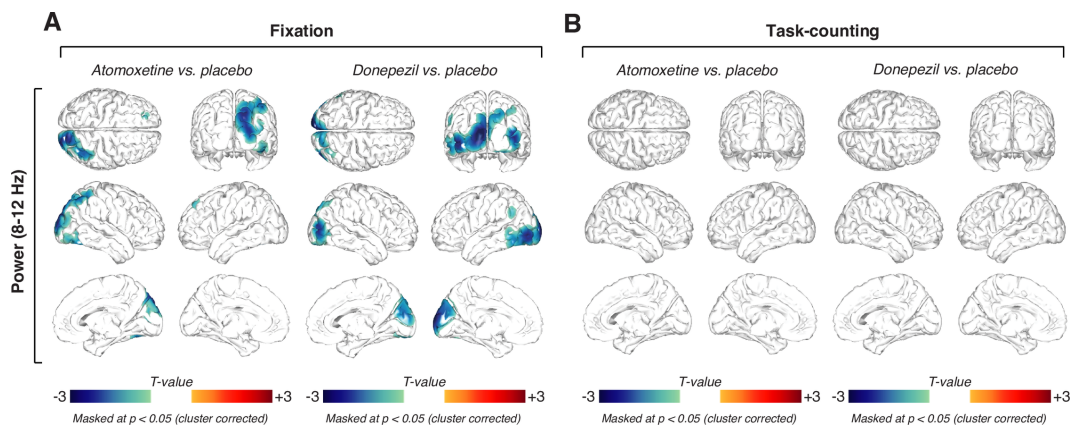


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 411 **Fig 8.** Inferring net E/I ratio from changes in scaling exponent α . Schematic illustration of the
 412 inference from observed change in exponent to (hidden) change in net E:I ratio (see main text for
 413 details). The non-monotonic dependence of scaling exponent α on E:I ratio (white line in Fig 4H) is
 414 replotted schematically. **(A)** The measured scaling exponent α during Fixation (gray) can result
 415 from both, inhibition- or excitation-dominant regimes; the baseline is unknown. We assume that
 416 external drive (Task-counting; yellow dot) does not increase E:I ratio (Shadlen & Newsome, 1998).
 417 Thus, the observed decrease in scaling exponent during Task-counting (yellow) must reflect a shift
 418 towards the inhibition-dominance (blue arrows), consistent with animal physiology (34). **(B)** This
 419 constrains the baseline state for the interpretation of the atomoxetine-induced increase in scaling
 420 exponent during Task-counting (red): The latter increase likely reflects an increase in E:I ratio (red
 421 arrow).
 422

423 Distinct, or absent, drug effects on other features of cortical dynamics

424 The absence of a consistent change in the scaling behavior of cortical activity
 425 fluctuations under donepezil (Fig 5C) was not simply due to a lack of effect on
 426 cortical dynamics per se. During Fixation, atomoxetine and donepezil both
 427 significantly reduced MEG power in the 8-12 Hz range, relative to placebo, in

428 posterior cortical regions (Fig 9 A/B; $p < 0.05$ for all clusters; two-sided cluster-
429 based permutation test). This suppression in cortical 8-12 Hz power due to both
430 catecholamines and acetylcholine during Fixation is largely consistent with
431 previous pharmacological work (30,40), as well as with correlations of cortical
432 activity with pupil diameter (41–44), a marker of neuromodulatory brainstem
433 activity underlying the release of noradrenaline and, to some extent, acetylcholine
434 (45–48).



435

436 **Fig 9.** Similar effects of atomoxetine and donepezil on 8-12 Hz power. **(A)** Spatial distribution of
437 drug-related alpha power changes during Fixation, thresholded at $p = 0.05$ (two-sided cluster-based
438 permutation test). *Left.* Power changes after the administration of atomoxetine. *Right.* Power
439 changes after the administration of donepezil. **(B)** Same as (A), but for Task-counting.

440

441 The atomoxetine-induced changes on 8-12 Hz power during Fixation had
442 a different spatial pattern than those of the atomoxetine-induced changes in the
443 scaling exponent α : within the cluster of the significant main effect of atomoxetine
444 on α , power did not significantly correlate with the changes in α (group average
445 spatial correlation between pooled difference maps within cluster; $r = 0.073$; $p =$
446 0.129 , $bf = 1.065$).

447 During Task-counting, neither drug significantly altered MEG-power (Fig
448 9B, $p > 0.05$ for all clusters; two-sided cluster-based permutation test),
449 presumably due to the already suppressed power in the 8-12 Hz range in that
450 condition.

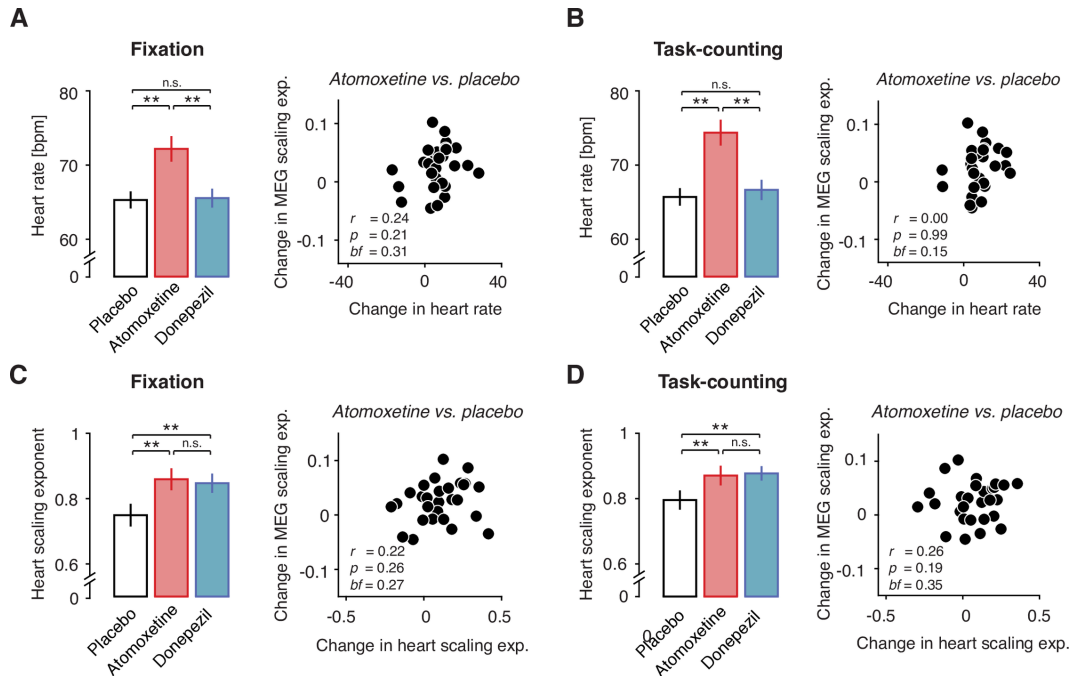
451 In sum, the effects of the drugs on cortical power during both conditions
452 showed that both were, at the dosages selected for our study, were equally
453 effective on cortical dynamics, consistently suppressing the power of low-
454 frequency oscillations during Fixation. This, as well as the lack of spatial
455 correlation of the atomoxetine-induced effects on power and scaling exponent α
456 further supports the specificity of the atomoxetine effect on cortical scaling
457 behavior.

458

459 **Atomoxetine effect on fluctuations in cortical activity is not due peripheral**
460 **confounds**

461 We also controlled for changes in peripheral physiological signals under the
462 drugs as potential confounds of the effect on cortical scaling behavior (Fig 10). As
463 expected, atomoxetine increased average heart rate (Fig 10A,B). Donepezil had
464 no significant effect on average heart rate, during neither Fixation ($p = 0.8676$; $t =$
465 0.16 ; paired t-test; $bf = 0.8676$; Fig 10A) nor Task-counting ($p = 0.3274$; $t = 1.0$;
466 paired t-test; $bf = 0.3139$; Fig 10B). Both drugs, however, significantly altered
467 heart rate scaling behavior, increasing the scaling exponent α (computed on
468 inter-heartbeat-interval time series, see Methods) in both behavioral contexts
469 (Fixation/atomoxetine: $p = 0.0012$, $t = 3.62$; Task-counting/atomoxetine: $p =$
470 0.0167 ; $t = 2.55$; Fig 10C; Fixation/donepezil: $p = 0.0076$, $t = 2.88$; Task-
471 counting/donepezil: $p = 0.0049$, $t = 3.06$; Fig 10D; all paired t-tests). Critically, the
472 atomoxetine-induced changes in heart rate showed no (Task-counting: $r = 0.00$; p
473 $= 0.99$; Person correlation; $bf = 0.15$) or only weak and statistically non-significant
474 (Fixation: $r = 0.24$; $p = 0.21$; Person correlation; $bf = 0.31$) correlations with the
475 changes in cortical activity (Fig 10A/B, right). Similarly, the atomoxetine-related
476 changes in the scaling behavior of inter-heartbeat intervals were only weakly (and

477 not significantly) correlated with the changes in cortical scaling behavior (Fixation:
 478 $r = 0.22$; $p = 0.26$; $bf = 0.27$; Task-counting: $r = 0.26$; $p = 0.19$; $bf = 0.35$; Fig
 479 10C/D, right).



480

481 **Fig 10.** Drug effect on cortical scaling behavior is not explained by systemic drug effects. **(A) Left.**
 482 Heart rate for atomoxetine, placebo and donepezil during Fixation. **Right.** Correlation of
 483 atomoxetine-related changes in heart rate (x-axis) with atomoxetine-related changes in MEG
 484 scaling exponent a (y-axis) (within significant cluster during Fixation). **(B) As (A),** but during Task-
 485 counting **(C) Right.** Scaling behavior of inter-heartbeat intervals (heart scaling exponent). **Left.**
 486 Heart scaling exponent for all pharmacological conditions during Fixation. **Right.** Correlation of
 487 atomoxetine-related changes in heart scaling exponent (x-axis) with atomoxetine-related changes
 488 in MEG scaling exponent a (y-axis). **(D) Same as (C),** but during Task-counting.

489

490 Atomoxetine, but not donepezil, significantly decreased spontaneous blink
 491 rate during Fixation ($p = 0.034$; $t = 2.24$; paired t-test), but not during Task-
 492 counting ($p = 0.112$; $t = 1.645$; $bf = 1.130$; paired t-test; Fig S2B). However, again
 493 there was no significant correlation between changes in blink-rate and changes in
 494 cortical scaling behavior due to atomoxetine (Fixation: $r = -0.26$; $p = 0.19$; $bf =$
 495 0.35 ; Task-counting: $r = -0.09$; $p = 0.64$; $bf = 0.16$).

496 In sum, drug-induced changes in peripheral physiological signals under
 497 the drugs, if present, did not account for the atomoxetine-induced changes in the
 498 scaling behavior of the fluctuations in cortical activity (Figs 5 and 6). These
 499 controls support our interpretation in terms of a specific effect on cortical net E/I

500 ratio rather than non-specific secondary effects due to the systemic drug effects
501 or changes in retinal input due to blinks.

502

503 **DISCUSSION**

504 Cortical circuits maintain a tight balance between excitation and inhibition. The
505 E/I ratio shapes the computational properties of cortical neurons and circuits (49),
506 and thereby the behavior of the organism (18–20). Deviations from this balance
507 have been linked to schizophrenia and autism and might also be at play in
508 various other neuropsychiatric disorders (50–53). Even in the absence of
509 changes in sensory input, the ratio between excitation and inhibition changes
510 continuously in cortex (17,54), presumably due to the effects of neuromodulators,
511 such as noradrenaline and acetylcholine (20,27–29,55,56). Neuromodulators also
512 regulate ongoing changes in the operating mode of behavior (23,25,57,58). Here,
513 we unraveled the effect of neuromodulatory-controlled microcircuit level changes
514 on the net cortical E/I ratio, as manifest in perception and behavior as well as in
515 local cortical population dynamics. Catecholamines, but not acetylcholine, altered
516 both, the dynamics of perceptual inference in the face of ambiguous input, and
517 intrinsic fluctuations in cortical activity. Both effects provided independent and
518 convergent evidence for an increase in E/I ratio due to catecholamines.

519

520 **Convergent evidence for catecholaminergic disinhibition in cortical circuits**

521 Our simulations indicated that the long-range temporal correlation of neural
522 population activity, as measured with the scaling exponent α , was highly
523 sensitive to changes in E/I ratio, produced through different regimes of
524 asymmetric synaptic gain modulation (see the white line in Fig 4H). In both
525 versions of our model, the neuromodulatory effects were not perfectly symmetric

526 (see the deviations of peak scaling exponents from main diagonal in Fig 4H).
527 While the latter effect was small and may be specific to the particular details of
528 the model, it remains possible that the subtle changes in scaling exponents we
529 observed were produced through symmetric gain modulations that maintained
530 the net E/I balance (i.e., along the main diagonal). However, two additional lines
531 of evidence converge on our conclusion that catecholamines (in particular
532 noradrenaline) boosted the cortical E/I ratio.

533 The first line of evidence is the specific and consistent effect of the
534 catecholaminergic manipulation on perceptual switch rate in same group of
535 participants. Building on a well-documented link between the volatility of
536 perceptual inference on cortical net E/I-balance (21,31,32), this behavioral effect
537 sits well with the notion of an effective net disinhibition in the circuits of visual
538 cortex that determine the dynamics of perceptual inference in the face of
539 ambiguous motion signals.

540 Second, a mounting body of evidence from recent invasive rodent work
541 also supports an overall increase in net cortical E/I ratio due to catecholamines,
542 specifically noradrenaline (17). One study established that noradrenaline
543 decreases tonic, ongoing inhibition of neurons in auditory cortex, with the
544 excitatory inputs unaffected (56). Another study showed that noradrenaline (but
545 not acetylcholine) mediated a locomotion-related, tonic depolarization of visual
546 cortical neurons (including pyramidal cells) (27). Both studies indicated a non-
547 selective (i.e. broadband) gain increase of neuronal responses, irrespective of the
548 features of presented stimuli, which is different from the more subtle disinhibitory
549 effects of acetylcholine (17,55).

550

551 **Cortical distribution of catecholaminergic effects on activity fluctuations**

552 The atomoxetine effects on the scaling exponent were widespread across cortex,
553 but not entirely homogenous. They were pronounced across occipital and parietal
554 cortex, but not robust in frontal cortex (see Fig 5B). This distribution might point to
555 a noradrenergic, rather than dopaminergic origin. Atomoxetine increases the
556 levels of both catecholamines, noradrenaline and dopamine (59), but the
557 dopaminergic system mainly projects to prefrontal cortex (60) but only sparsely
558 projects to occipital areas (61), whereas the noradrenergic projections are more
559 widespread and strong to occipito-parietal cortex (62). Alternatively, this
560 distribution may reflect the different receptor composition of across cortical
561 regions (63,64): The relative frequency of different adrenoceptors (α 1-, α 2 or β -
562 adrenoceptor) differs strongly between frontal and posterior cortex, which, in turn,
563 can result in distinct effects of noradrenaline on the dynamics of neural activity in
564 these different cortical regions (63), in particular persistent activity. Future studies
565 should investigate whether the observed differences of noradrenergic effects on
566 long-range temporal correlations in cortical activity are due to these differences in
567 adrenoceptor composition across cortex.

568

569 **No evidence for cholinergic effects on net E/I ratio**

570 In contrast to atomoxetine, we observed no robust effect of increased
571 acetylcholine levels on cortical long-range temporal correlations. This absence of
572 an effect was unlikely due to an ineffective pharmacological manipulation through
573 donepezil: the latter had equally strong effects as atomoxetine on alpha-band
574 power in some cortical regions, as well as on heart rate variability. Rather, the
575 absence of robust donepezil effects might reflect specific properties of cholinergic
576 action, which may leave the cortical net excitation-inhibition ratio largely
577 unchanged. Substantial evidence points to the rapid disinhibition of (excitatory)

578 pyramidal cells by acetylcholine, by activating a circuit made up of a chain of two
579 inhibitory interneurons (VIP+ and SOM+) (28,65,66). The cholinergic activation of
580 this disinhibitory circuit would be expected to shift the net excitation-inhibition
581 ratio towards excitation, just as we inferred for catecholamines. However, this
582 disinhibitory circuit seems to mainly affect transient, stimulus-evoked responses
583 (55), whereas noradrenaline also alters the tonic levels of inhibition (56). This
584 may explain the relative lack of donepezil effects during the steady-state
585 conditions (blank fixation and continuous task drive) employed in our present
586 study. In general, cholinergically mediated disinhibitory effects on cortical
587 neurons might be subtler as well as more selective than the ones mediated by
588 noradrenaline (17).

589

590 **Decrease of long-range temporal correlations during task and sensory**
591 **drive**

592 Consistent with our current results, previous studies also found a decrease in
593 temporal autocorrelations of cortical activity due to external drive, even during
594 intermittent presentation of stimuli and tasks, entailing more external transients
595 than the steady-state task condition used here (8,67). The observation is
596 consistent with the insight from intracellular recordings of cortical neurons in
597 animals, that cortical responses to sensory stimulation in the awake state are
598 dominated by inhibition (33,34). One candidate source of this sensory-driven
599 state change is thalamocortical inhibition (68), but intracortical feedback inhibition
600 might also contribute (69).

601 Simulations of large-scale biophysical models of cortical networks show
602 that the driven state is associated with shortened temporal autocorrelations as
603 well as a decrease in the entropy of activity states in the network (70).

604 Correspondingly, the increase in long-range temporal autocorrelations under
605 catecholaminergic modulation observed presently may be associated with an
606 increase in entropy, in other words, a tendency of the cortex to explore a larger
607 set of activity states. This greater exploration of cortical state space may in turn
608 be linked to a prominent idea about the function of noradrenaline, which
609 postulates that high tonic noradrenaline levels promote exploratory, and more
610 distractible, behavior (23).

611

612 **Functional consequences of changes in net cortical E/I ratio**

613 We observed a selective increase in the rate of spontaneous perceptual
614 alternations under catecholaminergic but not cholinergic boost, adding to
615 evidence that these dynamics are under neuromodulatory control (71). Such a
616 change could be due to an increase in cortical “noise” defined as the amplitude of
617 spontaneous fluctuations in activity (31). Future invasive studies should relate
618 catecholaminergic changes in the variability of spiking activity (72) to bistable
619 perception.

620 The selective increase of perceptual alternation rate under atomoxetine is
621 consistent with the relative decrease of intra-cortical inhibition (21) that was also
622 inferred from the changes in the long-range temporal correlation structure of
623 cortical activity. A net increase in excitation will likely have particularly strong
624 effects on the dynamics of parietal and prefrontal cortical circuits involved in
625 working memory and decision-making (19). These circuits are characterized by
626 slow intrinsic fluctuations of activity (73–75). The catecholaminergic increase in
627 long-range temporal correlations of intrinsic activity fluctuations in parietal circuits
628 that we observed in the current study may reflect a relative increase specifically
629 in the recurrent excitation in ‘accumulator’ circuits. Recurrent excitation, in turn, is

630 essential for both the computational capacities (76) as well as the timescale of
631 intrinsic activity fluctuations of these circuits (74,75). Simulations of synaptic gain
632 modulation of such ‘accumulator’ circuits indicate that the most robust behavior
633 emerges from co-modulation of both excitatory and inhibitory synapses, but with
634 different factors (20). It will be important to test these predictions in future work,
635 using tasks tailored to probing into these circuits of association cortex.

636

637 **Catecholamines: a control parameter for critical network dynamics**

638 Long-range temporal correlations in the fluctuations of neural mass activity (i.e.,
639 activity summed across the entire local network) (7) and avalanches within the
640 neuronal network (37) jointly emerge at the same ratio between excitatory and
641 inhibitory connectivity in the simplified cortical patch model used here. Both
642 phenomena, long-range temporal correlations and neuronal avalanches, are
643 commonly interpreted as hallmarks of “criticality” (7,10,37,77). Criticality refers to
644 a complex dynamical system poised between order and chaos (78–80).

645 The cortex might operate in a narrow regime around this critical point
646 (80,81). This operating mode, in turn, might yield computational modes superior
647 to those of the “sub-“ or “supercritical” modes (38,77,82–84). A number of recent
648 reports have indicated that cortical dynamics may fluctuate around the critical
649 state (85–88), but these fluctuations have, so far, been spontaneous. Here, we
650 identified two key factors (task drive and catecholaminergic neuromodulation) to
651 bring these changes under experimental control. Complex systems can self-
652 organize towards criticality (78), e.g., through plasticity and/or feedback
653 connections. However, critical dynamics can also be achieved through an
654 external control parameter that fine-tunes the system. The tuning of temperature

655 in the Ising model of spin magnetization is a common example (80).

656 Noradrenaline may serve as such a control parameter in the cerebral cortex.

657 In sum, combining measurements of perceptual dynamics as well as

658 intrinsic fluctuations in cortical population activity under steady-state perceptually

659 ambiguous stimulation provides a novel non-invasive read-out of pharmacological

660 effects on cortical net E/I ratio in humans. This read-out might be useful for

661 addressing fundamental questions about the state dependence of cortical

662 computation and for inferring changes in cortical E/I ratio in neuropsychiatric

663 disorders, or pharmacological treatments of these disorders.

664

665 **METHODS**

666 **Pharmacological MEG experiment**

667 *Participants*

668 30 healthy human participants (16 females, age range 20-36, mean 26.7)

669 participated in the study after informed consent. The study was approved by the

670 Ethical Committee responsible for the University Medical Center Hamburg-

671 Eppendorf. Two participants were excluded from analyses, one due to excessive

672 MEG artifacts, the other due to not completing all 3 recording sessions. Thus, we

673 report results from N=28 participants (15 females).

674

675 *General design*

676 We pharmacologically manipulated the levels of catecholamines (noradrenaline

677 and dopamine) and acetylcholine in a double-blind, randomized, placebo-

678 controlled, and cross-over experimental design (Fig 1A, B). Each participant

679 completed three experimental sessions, consisting of drug (or placebo) intake at

680 two time points, a waiting period of 3 hours, and an MEG recording. During each

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681 MEG session, participants were seated on a chair inside a magnetically shielded
682 MEG chamber. Each session consisted of 6 runs of different tasks, each of which
683 was 10 minutes long and followed by breaks of variable duration.

684

685 *Pharmacological intervention*

686 We used the selective noradrenaline reuptake inhibitor atomoxetine (dose: 40
687 mg) to boost the levels of catecholamines, specifically noradrenaline and (in
688 prefrontal cortex) dopamine (59). We used the cholinesterase inhibitor donepezil
689 (dose: 5 mg) to boost acetylcholine levels. A mannitol-aerosil mixture was
690 administered as placebo. All substances were encapsulated identically in order to
691 render them visually indistinguishable. Peak plasma concentration are reached
692 ~3-4 hours after administration for donepezil (89) and 1-2 hours after
693 administration for atomoxetine (90), respectively. We adopted the following
694 procedure to account for these different pharmacokinetics (Fig 1A): participants
695 received two pills in each session, one 3 h and another 1.5 h before the start of
696 MEG recording. In the Atomoxetine condition, they first received a placebo pill (t
697 = -3 h) followed by the atomoxetine pill (t = -1.5 h). In the Donepezil condition,
698 they first received the donepezil pill (t = -3 h), followed by placebo (t = -1.5 h). In
699 the Placebo condition, they received a placebo at both time points. The half-life is
700 ~ 5 h for atomoxetine (90) and ~ 82 h for donepezil, respectively (89). In order to
701 allow plasma concentration levels to return to baseline, the three recording
702 sessions were scheduled at least 2 weeks apart. This design ensured maximum
703 efficacy of both pharmacological manipulations, while effectively blinding
704 participants as well as experimenters.

705

706 *Stimuli and behavioral tasks*

707 In each session, participants alternated between three different task conditions (2
708 runs à 10 minutes per condition) referred to as Fixation, Task-counting, and
709 Task-pressing in the following (Fig 1B). All conditions entailed overall constant
710 sensory input. Fixation and Task-counting also entailed no overt motor responses
711 and are, therefore, referred to as “steady-state” conditions in the following. We
712 used these steady-state conditions to quantify intrinsic fluctuations in cortical
713 activity. Task-pressing entailed motor responses and was used for reliable
714 quantification of perceptual dynamics. All instructions and stimuli were projected
715 onto a screen (distance: 60 cm) inside the MEG chamber. The individual
716 conditions are described as follows.

717 *Fixation.* Participants were asked to keep their eyes open and fixate a
718 green fixation dot (radius = 0.45° visual angle) presented in the center of an
719 otherwise gray screen. This is analogous to eyes-open measurements of
720 “resting-state” activity widely used in the literature on intrinsic cortical activity
721 fluctuations.

722 *Task-counting.* Participants viewed a seemingly rotating sphere giving rise
723 to the kinetic depth effect (91,92): spontaneous changes in the perceived rotation
724 direction (Fig 1B). The stimulus subtended 21° of visual angle. It consisted of
725 1000 dots (500 black and 500 white dots, radius: 0.18° of visual angle) arranged
726 on a circular aperture presented on a mean-luminance gray background, with the
727 green fixation dot in the center. In order to minimize tracking eye movements, the
728 sphere rotation was along the horizontal axis, either “forward” (towards the
729 observer) or “backward” (away from the observer), and the dot density decreased
730 along the horizontal axis towards the center of the stimulus. Participants were
731 instructed to count the number of perceived changes in rotation direction and
732 report the total number of perceived transitions at the end of the run. Just like

733 during Fixation, Task-counting minimized any external (sensory or motor)
734 transients. Subjects silently counted the alternations in perceived rotation
735 direction and verbally reported the total count after the end of the 10 min run.

736 *Task-pressing.* This condition was identical to Task-counting, except that
737 participants were instructed to press and hold one of two buttons with their index
738 finger to indicate the perceived rotation direction of the sphere. Thus, each
739 perceptual alternation was accompanied by a motor response leading to change
740 in the button state. This allowed for a more reliable quantification of participants'
741 perceptual dynamics. On two sessions (atomoxetine condition), button presses
742 were not registered. Hence, the corresponding analyses were performed on 26
743 participants.

744

745 **Data acquisition**

746 MEG was recorded using a whole-head CTF 275 MEG system (CTF Systems,
747 Inc., Canada) at a sampling rate of 1200 Hz. In addition, eye movements and
748 pupil diameter were recorded with an MEG-compatible EyeLink 1000 Long
749 Range Mount system (SR Research, Osgoode, ON, Canada) at a sampling rate
750 of 1000 Hz. In addition, electrocardiogram (ECG) as well as vertical, horizontal
751 and radial EOG were acquired using Ag/AgCl electrodes (sampling rate 1200 Hz).

752

753 **Data analysis**

754 *Eye data*

755 Eye blinks were detected using the manufacturer's standard algorithm with
756 default settings. Saccades and microsaccades were detected using the saccade
757 detection algorithm described in (93), with a minimum saccade duration of 4

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758 samples (= 4 ms) and a threshold velocity of 6. For 18 out of 28 participants, only
759 horizontal eye movements were recorded.

760

761 *EOG data*

762 EOG events (blinks and saccades) were extracted using semi-automatic artifact
763 procedures as implemented in FieldTrip (94). In short, EOG traces were
764 bandpass filtered using a third-order butterworth filter (1 – 15 Hz) and the
765 resulting signal was z-scored. All time points where the resulting signal exceeded
766 a z-score of 4 were marked as an EOG event.

767

768 *MEG data*

769 *Preprocessing.* First, all data were cleaned of strong transient muscle artifacts
770 and squid jumps through visual inspection and manual as well as semi-automatic
771 artifact rejection procedures, as implemented in the FieldTrip toolbox for MATLAB
772 (94). To this end, data segments contaminated by such artifacts (+/- 500 ms)
773 were discarded from the data (across all channels). Subsequently, data were
774 downsampled to 400 Hz split into low (2-40 Hz) and high (>40 Hz) frequency
775 components, using a 4th order (low- or high-pass) Butterworth filter. Both signal
776 components were separately submitted to independent component analysis (95)
777 using the FastICA algorithm (96). Artifactual components (eye blinks/movements,
778 muscle artifacts, heartbeat and other extra-cranial artifacts) were identified based
779 on three established criteria (97): power spectrum, fluctuation in signal variance
780 over time (in bins of 1s length), and topography. Artifact components were
781 reconstructed and subtracted from the raw signal and low- and high frequencies
782 were combined into a single data set. On average, 20 (+/- 14) artifact

783 components were identified for the low frequencies and 13 (+/- 7) artifactual
784 components were identified for the high frequencies.

785

786 *Spectral analysis.* Sensor-level spectral estimates (power spectra and cross
787 spectral density matrices) were computed by means of the multi taper method
788 using a sequence of discrete prolate Slepian tapers (98). For the power spectrum
789 shown in Fig 1C, power spectra were computed using a window length of 5s and
790 a frequency smoothing of 2 Hz, yielding 19 orthogonal tapers. The focus of this
791 paper was on the fluctuations of the amplitude envelopes, rather than on the
792 (oscillatory) fluctuations of the carrier signals *per se*. The temporal correlation
793 structure of the amplitude envelope fluctuations of cortical activity seems similar
794 across different carrier frequency bands (10). We focused on amplitude envelope
795 fluctuations in the alpha-band because (i) the cortical power spectra exhibited a
796 clearly discernible alpha-peak, which robustly modulated with task, as expected
797 from previous work (39) (Fig 1C); and (ii) the computational model used to study
798 the effect of synaptic gain modulation on cortical activity fluctuations was tuned to
799 produce alpha-band oscillations (see above and (15)).

800

801 *Source reconstruction: general approach.* The cleaned sensor level signals (N
802 sensors) were projected onto a grid consisting of $M = 3000$ voxels covering the
803 cortical surface (mean distance: 6.3 mm) using the exact low-resolution brain
804 electromagnetic tomography (eLORETA; (99) method. The magnetic leadfield
805 was computed, separately for each subject and session, using a single shell head
806 model constructed from the individual structural MRI scans and the head position
807 relative to the MEG sensors at the beginning of the run (100). In case no MRI
808 was available (4 subjects), the leadfield was computed from a standard MNI

809 template brain transformed to an estimate of the individual volume conductor
810 using the measured fiducials (located at the nasion, the left and the right ear).

811

812 *Source level estimates of amplitude envelopes and power.* For comparing
813 amplitude envelope and power estimates between experimental conditions in
814 source space we aimed to select a single direction of the spatial filter for each
815 voxel across pharmacological conditions (i.e., MEG sessions), but separately for
816 Fixation and Task-Counting conditions. The rationale was to avoid filter-induced
817 biases in the comparisons between the pharmacological conditions, while
818 allowing that external task drive might systematically change the dipole
819 orientations.

820 To this end, we first computed the mean source-level cross-spectral
821 density matrix $C(r, f)$ for each frequency band, f , averaged across the three
822 MEG sessions, as follows:

$$823 \quad C(r, f) = \frac{1}{3} \sum_{i=1}^3 \left(A_i^T(r) C_i(f) A_i(r) \right) \quad (1)$$

824 whereby i indicated the MEG session, $C_i(f)$ was the (sensor-level) session- and
825 frequency-specific cross-spectral density matrix and A_i is the spatial filter for
826 session i . We then extracted the first eigenvector $u_1(r, f)$ of the session-average
827 matrix $C(r, f)$ and computed the unbiased filter selective for the dominant dipole
828 orientation, $B_i(r, f)$, as:

$$829 \quad B_i(r, f) = A_i(r) u_1(r, f) \quad (2)$$

830 Please note that this filter was now frequency-specific, whereas the
831 previous filters, $A_i(r)$, were not. To obtain instantaneous estimates of source-
832 level amplitudes, the sensor-level signal for session i , $X_i(t)$, was band-pass
833 filtered (using a finite impulse response filter) and Hilbert-transformed, yielding a
834 complex-valued signal $H_i(f, t)$ for each frequency band. This signal was

835 projected into source space through multiplication with the unbiased spatial filter,
836 $B_i(r, f)$, and the absolute value was taken:

$$837 \quad Env_i(r, f, t) = |(H_i(f, t) B_i(r, f))| \quad (3)$$

838 where $Env_i(r, f, t)$ was the estimated amplitude envelope time course of source
839 location r and frequency f . Next, for each session, unbiased source-level cross
840 spectral density estimates were obtained from the sensor-level cross-spectral
841 density matrix $C_i(f)$ and the frequency-specific, unbiased spatial filter $B_i(f)$. The
842 main diagonal of the resulting matrix contains source-level power estimates for all
843 source locations:

$$844 \quad S_i(f) = diag(B_i^T(f) C_i(f) B_i(f)) \quad (4)$$

845 These computations were repeated separately for the Task-counting and
846 Fixation conditions, session by session. The differences in amplitude envelope
847 fluctuations and power estimates between pharmacological and task conditions
848 reported in this paper were robust with respect to the specifics of the analysis
849 approach. In particular, we obtained qualitatively similar pharmacological effects
850 in sensor space, as reported in an earlier conference abstract (101).

851

852 *Detrended fluctuation analysis.* The source-level amplitude envelopes
853 $Env_i(r, f, t)$ were submitted to detrended fluctuation analysis (102,103) in order
854 to quantify long-range temporal correlations. Detrended fluctuation analysis
855 quantifies the power law scaling of the fluctuation (root-mean-square) of a locally
856 detrended, cumulative signal with time-window length. Different from the analysis
857 of the more widely known autocorrelation function (73,74), detrended fluctuation
858 analysis provides robust estimates of the autocorrelation structure for stationary
859 and non-stationary time series. The procedure of the detrended fluctuation
860 analysis is illustrated in Fig 2.

861 For simplicity, in the following, we re-write the amplitude envelope
862 $Env_i(r, f, t)$ as x of length T . First, we computed the cumulative sum of the
863 demeaned x , (Fig 2B):

$$864 \quad X(t) = \sum_{t'=1}^t (x(t') - \langle x \rangle) \quad (5)$$

865 where t' and t denote single time points up to length T . The cumulative signal X
866 was then cut into $i = 1 \dots k$ segments Y_i of length N (overlap: 50%), where $k =$
867 $\text{floor}[(T - N)/(0.5 N)]$ (Fig 2B, top). Within each segment Y_i of equal length N ,
868 the linear trend Y_{i_trend} (least squares fit) was subtracted from Y_i (Fig 2B, bottom,
869 blue vs. red lines), and the root-mean-square fluctuation for a given segment was
870 computed as:

$$871 \quad F_{N_i} = \left[\frac{1}{N} \sum_{n=1}^N (Y_i(n) - Y_{i_trend}(n))^2 \right]^{\frac{1}{2}} \quad (6)$$

872 where n indicates the individual time points. The fluctuation was computed for all
873 k segments of equal length N and the average fluctuation was obtained through:

$$874 \quad \langle F_N \rangle = \frac{1}{k} \sum_{i=1}^k F_{N_i} \quad (7)$$

875 The procedure was repeated for 15 different logarithmically spaced window
876 lengths N , ranging from 3 s to 50 s, which yields a fluctuation function (Fig 2C).
877 As expected for scale-free time series (103), this fluctuation function follows a
878 power-law of the form:

$$879 \quad \langle F_N \rangle \propto N^\alpha \quad (8)$$

880 The “scaling exponent” α was computed through a linear regression fit in log-log
881 coordinates (Fig 2C). The longest and shortest window lengths were chosen
882 according to guidelines provided in (103).

883 A scaling exponent of $\alpha \sim 0.5$ indicates a temporally uncorrelated (“white
884 noise”) process. Scaling exponents between $0.5 < \alpha < 1$ are indicative of scale-
885 free behavior and long-range temporal correlations (103), whereas exponents of
886 $\alpha < 0.5$ indicate long-range anti-correlations (“switching behavior”) and $\alpha > 1$ are

887 indicative of an unbounded process (103). The scaling exponents for alpha-band
888 MEG amplitude envelopes estimated in this study ranged (across experimental
889 conditions, MEG sensors and participants) from 0.40 and 1.04, with 99.4% of all
890 estimates in the range from 0.5 to 1. This is indicative of scale-free behavior and
891 consistent with previous human MEG work (7–10,12,13).

892

893 *Relationship between measures of cortical variability.* Scale-free behavior of
894 neural time series has also been quantified via analysis of the power spectrum
895 (5,6,73). There is a straightforward relationship between both approaches, which
896 we explain below, to help appreciate our results in the context of these previous
897 studies. The power spectrum of the amplitude envelope of cortical activity is
898 typically well approximated by the power law $p(f) \propto f^{-\beta}$, where β is referred to
899 as the power-law exponent (Fig 2D). For power-law decaying autocorrelations,
900 the relationship between the power-law exponent β and the scaling exponent α
901 (estimated through DFA) of a time series is:

902
$$\beta = 2\alpha - 1 \quad (9)$$

903

904 *Analysis of ECG data.* ECG data were used to analyze two measures of
905 peripheral autonomic activity: average heart rate and heart rate variability. For
906 both measures, we used an adaptive threshold to detect the R-peak of each
907 QRS-complex in the ECG. Heart rate was then computed by dividing the total
908 number of R-components by time. Heart rate variability was quantified by means
909 of the detrended fluctuations analysis described for MEG above, but now applied
910 to the time series of the intervals between successive R-peaks (9,10). In line with
911 the MEG analyses, we used windows ranging from 3 to 50 heartbeats (roughly
912 corresponding to 3–50 s).

913

914 *Statistical tests*

915 Statistical comparisons of all dependent variables between conditions were,
916 unless stated otherwise, performed using paired t-tests.

917 Null effects are difficult to interpret using regular null hypothesis
918 significance testing. The Bayes Factor addresses this problem by quantifying the
919 strength of the support for the null hypothesis over the alternative hypothesis
920 provided by the data, taking effect size into account. Wherever null effects were
921 conceptually important, results obtained from a regular (paired) t-test (104) and
922 Pearson correlations (105) were converted into corresponding Bayes Factors.

923 To map significant changes of scaling exponents α on the cortical surface,
924 we computed a non-parametric permutation test based on spatial clustering
925 (106,107). This procedure has been shown to reliably control for Type I errors
926 arising from multiple comparisons. First, a paired t-test was performed to identify
927 voxels with significant changes (voxel with $p < 0.05$). Subsequently, significant
928 voxels are combined into clusters based on their spatial adjacency. Here, a voxel
929 was only included into a cluster when it had at least two significant neighbors.
930 Subsequently, the t-values of all voxels comprising a cluster were summed,
931 which yields a cluster statistic (i.e., a cluster t-value) for each identified cluster.
932 Next, a randomization null distribution was computed using a permutation
933 procedure ($N = 10,000$ permutations). On each permutation, the experimental
934 labels (i.e., the pharmacological conditions) were randomly re-assigned within
935 participants and the aforementioned procedure was repeated. For each iteration,
936 the maximum cluster statistic was determined and a distribution of maximum
937 cluster statistics was generated. Eventually, the cluster statistic of all empirical
938 clusters was compared to the values obtained from the permutation procedure.

939 All voxels comprising a cluster with a cluster statistic smaller than 2.5% or larger
940 than 97.5% of the permutation distribution were labeled significant, corresponding
941 to a corrected threshold of $\alpha = 0.05$ (two-sided).

942

943 **Model simulations**

944 To simulate the effects of synaptic gain modulation on cortical activity fluctuations,
945 we extended a previously described computational model of a local cortical patch
946 (15) by means of multiplicative modulation of synaptic gain. All features of the
947 model were identical to those of the model by (15), unless stated otherwise. The
948 model consisted of 2500 integrate-and-fire neurons (75% excitatory, 25%
949 inhibitory) with local connectivity within a square (width = 7 units) and a
950 connection probability that decayed exponentially with distance (Fig 4A). The
951 dynamics of the units were governed by:

$$952 \quad I_i = I_0 + \sum_j N_{ij} W_{ij} S_j \quad (10)$$

$$953 \quad \tau_i \frac{dI_i}{dt} = I_0 - I_i \quad (11)$$

954 where subscripts i, j indicated different units, N_{ij} was a multiplicative gain factor,
955 W_{ij} were the connection weights between two units, and S_j a binary spiking
956 vector representing whether unit j did or did not spike on the previous time step,
957 and $I_0 = 0$. The connection weights were $W_{EE} = 0.0085$, $W_{IE} = 0.0085$, $W_{EI} =$
958 -0.569 and $W_{II} = -2$ whereby subscript E indicated excitatory, subscript I
959 indicated inhibitory, and the first and second subscript referred to the receiving
960 and sending unit, respectively.

961 On each time step ($dt = 1$ ms), I_i was updated for each unit i , with the
962 summed input from all other (connected) units j and scaled by a time constant
963 $\tau_i = 9$ ms, which was the same for excitatory and inhibitory units. The probability
964 of a unit generating a spike output was given by:

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965
$$P_{si} = P_{si} + I_i \quad (12)$$

966
$$\tau_P \frac{dP_{si}}{dt} = P_0 - P_{si} \quad (13)$$

967 with the time constant for excitatory units $\tau_P = 6 \text{ ms}$ and for inhibitory $\tau_P = 12 \text{ ms}$.

968 P_0 was the background spiking probability, with $P_0(\text{exc.}) = 0.000001 [1/\text{ms}]$ and

969 $P_0(\text{inh.}) = 0 [1/\text{ms}]$. For each time step, it was determined whether a unit did or

970 did not spike. If it did, the probability of that unit spiking was reset to

971 $P_r(\text{excitatory}) = -2 [1/\text{ms}]$ and $P_r(\text{inhibitory}) = -20 [1/\text{ms}]$.

972 We used this model to analyze the dependency of two quantities on E/I

973 ratio: (i) the power-law scaling of the distributions of the sizes of neuronal

974 avalanches (37) estimated in terms of the kappa-index κ which quantifies the

975 difference between an empirically observed event size distribution and a

976 theoretical reference power-law distribution with a power-law exponent -1.5 (38),

977 and (ii) the scaling behavior (scaling exponent α) of the amplitude envelope

978 fluctuations of the model's local field potential. To this end, we summed the

979 activity across all (excitatory and inhibitory) neurons to obtain a proxy of the local

980 field potential. We band-pass filtered the local field potential in the alpha-band (8–

981 12 Hz) and computed long-range temporal correlations in the alpha-band

982 amplitude envelopes following the procedure described above (see *Detrended*

983 *fluctuation analysis of MEG data*), using windows sizes ranging from 5 s to 30 s.

984 For all simulations reported in this paper, we optimized the connection weights

985 using Bonesa, a parameter tuning algorithm (108), such that the network

986 exhibited alpha-band oscillations, long-range temporal correlations, and neuronal

987 avalanches (see Discussion).

988 In order to assess the influence of structural excitatory and inhibitory

989 connectivity on network dynamics (Figs 4D-F), we varied the percentage of units

990 (excitatory and inhibitory) a given excitatory or inhibitory unit connects to within a

991 local area (7 units x 7 units; Fig 4A). These percentages were varied
992 independently for excitatory and inhibitory units with a step size of 2.5%.

993 The gain factor N_{ij} was the main difference to the model described by
994 (15). It was introduced to simulate the effects of neuromodulation on synaptic
995 interactions in the cortical network (20). With all the above parameters fixed
996 (42.5% excitatory connectivity, 75% inhibitory connectivity; small square in Figs
997 4D-F), we systematically varied the synaptic gain factors, in two different ways. In
998 the first version, we only varied N_{EE} and N_{IE} to dynamically modulate the circuit's
999 net E/I ratio (Fig 4B), in a way consistent with recent modeling of the effects of E/I
1000 ratio on a cortical circuit for perceptual decision-making (18). In the second
1001 version, we varied N_{EE} , N_{IE} , and N_{EI} (Fig S3A). Here, N_{EI} was modulated
1002 independently from N_{EE} , and N_{IE} , which in turn were co-modulated by the same
1003 factor.

1004 Per parameter combination, we ran 10 simulations, using the Brian2
1005 spiking neural networks simulator (109). Each simulation was run for 1000
1006 seconds, with a random initialization of the network structure and the probabilistic
1007 spiking. In this paper, we focus on the effects of neuromodulation on the scaling
1008 exponent α , which served as a reference for interpretation of the MEG effects.

1009

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1019

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1021 Conceptualization: T.P., A.K.E., and T.H.D.; Experimental design: T.P. and
1022 T.H.D.; Model design: T.P., A-E.A., K.L-H., and T.H.D.; Investigation: T.P.;
1023 Formal analysis: T.P.; Model simulations: A.-E.A.; Writing - Original draft: T.P.
1024 and T.H.D.; Writing – Review & Editing: T.P., A-E.A., G.N., A.K.E., K.L-H., and
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1027

1028 **COMPETING FINANCIAL INTERESTS**

1029 The authors declare no competing financial interests.

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1033 **REFERENCES**

- 1034 1. Faisal AA, Selen LPJ, Wolpert DM. Noise in the nervous system. *Nat Rev*
1035 *Neurosci.* 2008 Apr;9(4):292–303.
- 1036 2. Shadlen MN, Newsome WT. The variable discharge of cortical neurons:
1037 implications for connectivity, computation, and information coding. *J*
1038 *Neurosci Off J Soc Neurosci.* 1998 May 15;18(10):3870–96.
- 1039 3. Fox MD, Snyder AZ, Vincent JL, Corbetta M, Van Essen DC, Raichle ME.
1040 The human brain is intrinsically organized into dynamic, anticorrelated
1041 functional networks. *Proc Natl Acad Sci U S A.* 2005 Jul 5;102(27):9673–8.
- 1042 4. Deco G, Jirsa VK, McIntosh AR. Emerging concepts for the dynamical
1043 organization of resting-state activity in the brain. *Nat Rev Neurosci.* 2011
1044 Jan;12(1):43–56.
- 1045 5. Miller KJ, Sorensen LB, Ojemann JG, den Nijs M. Power-law scaling in the
1046 brain surface electric potential. *PLoS Comput Biol.* 2009
1047 Dec;5(12):e1000609.

Pfeffer et al: Neuromodulation and Cortical Excitation-Inhibition Ratio

- 1048 6. He BJ, Zempel JM, Snyder AZ, Raichle ME. The temporal structures and
1049 functional significance of scale-free brain activity. *Neuron*. 2010 May
1050 13;66(3):353–69.
- 1051 7. Linkenkaer-Hansen K, Nikouline VV, Palva JM, Ilmoniemi RJ. Long-range
1052 temporal correlations and scaling behavior in human brain oscillations. *J*
1053 *Neurosci Off J Soc Neurosci*. 2001 Feb 15;21(4):1370–7.
- 1054 8. He BJ. Scale-Free Properties of the Functional Magnetic Resonance
1055 Imaging Signal during Rest and Task. *J Neurosci*. 2011 Sep
1056 28;31(39):13786–95.
- 1057 9. Palva JM, Zhigalov A, Hirvonen J, Korhonen O, Linkenkaer-Hansen K,
1058 Palva S. Neuronal long-range temporal correlations and avalanche
1059 dynamics are correlated with behavioral scaling laws. *Proc Natl Acad Sci*.
1060 2013 Feb 26;110(9):3585–90.
- 1061 10. Zhigalov A, Arnulfo G, Nobili L, Palva S, Palva JM. Relationship of fast-
1062 and slow-timescale neuronal dynamics in human MEG and SEEG. *J*
1063 *Neurosci Off J Soc Neurosci*. 2015 Apr 1;35(13):5385–96.
- 1064 11. Linkenkaer-Hansen K, Smit DJA, Barkil A, van Beijsterveldt TEM,
1065 Brussaard AB, Boomsma DI, et al. Genetic contributions to long-range
1066 temporal correlations in ongoing oscillations. *J Neurosci Off J Soc*
1067 *Neurosci*. 2007 Dec 12;27(50):13882–9.
- 1068 12. Linkenkaer-Hansen K. Breakdown of Long-Range Temporal Correlations
1069 in Theta Oscillations in Patients with Major Depressive Disorder. *J*
1070 *Neurosci*. 2005 Nov 2;25(44):10131–7.
- 1071 13. Montez T, Poil S-S, Jones BF, Manshanden I, Verbunt JPA, van Dijk BW,
1072 et al. Altered temporal correlations in parietal alpha and prefrontal theta
1073 oscillations in early-stage Alzheimer disease. *Proc Natl Acad Sci*. 2009
1074 Feb 3;106(5):1614–9.
- 1075 14. van Vreeswijk C, Sompolinsky H. Chaos in neuronal networks with
1076 balanced excitatory and inhibitory activity. *Science*. 1996 Dec
1077 6;274(5293):1724–6.
- 1078 15. Poil S-S, Hardstone R, Mansvelder HD, Linkenkaer-Hansen K. Critical-
1079 State Dynamics of Avalanches and Oscillations Jointly Emerge from
1080 Balanced Excitation/Inhibition in Neuronal Networks. *J Neurosci*. 2012 Jul
1081 18;32(29):9817–23.
- 1082 16. Deco G, Ponce-Alvarez A, Hagmann P, Romani GL, Mantini D, Corbetta M.
1083 How local excitation-inhibition ratio impacts the whole brain dynamics. *J*
1084 *Neurosci Off J Soc Neurosci*. 2014 Jun 4;34(23):7886–98.
- 1085 17. Froemke RC. Plasticity of Cortical Excitatory-Inhibitory Balance. *Annu Rev*
1086 *Neurosci*. 2015 Jul 8;38(1):195–219.

Pfeffer et al: Neuromodulation and Cortical Excitation-Inhibition Ratio

- 1087 18. Lam NH, Borduqui T, Hallak J, Roque AC, Anticevic A, Krystal JH, et al.
1088 Effects of Altered Excitation-Inhibition Balance on Decision Making in a
1089 Cortical Circuit Model. *bioRxiv*. 2017 Jan;
- 1090 19. Wang X-J. Decision making in recurrent neuronal circuits. *Neuron*. 2008
1091 Oct 23;60(2):215–34.
- 1092 20. Eckhoff P, Wong-Lin KF, Holmes P. Optimality and Robustness of a
1093 Biophysical Decision-Making Model under Norepinephrine Modulation. *J*
1094 *Neurosci*. 2009 Apr 1;29(13):4301–11.
- 1095 21. van Loon AM, Knapen T, Scholte HS, St. John-Saaltink E, Donner TH,
1096 Lamme VAF. GABA Shapes the Dynamics of Bistable Perception. *Curr*
1097 *Biol*. 2013 May;23(9):823–7.
- 1098 22. Marder E. Neuromodulation of neuronal circuits: back to the future. *Neuron*.
1099 2012 Oct 4;76(1):1–11.
- 1100 23. Aston-Jones G, Cohen JD. An integrative theory of locus coeruleus-
1101 norepinephrine function: adaptive gain and optimal performance. *Annu Rev*
1102 *Neurosci*. 2005;28:403–50.
- 1103 24. Berridge CW. Noradrenergic modulation of arousal. *Brain Res Rev*. 2008
1104 Jun;58(1):1–17.
- 1105 25. Harris KD, Thiele A. Cortical state and attention. *Nat Rev Neurosci*. 2011
1106 Aug 10;12(9):509–23.
- 1107 26. Lee S-H, Dan Y. Neuromodulation of brain states. *Neuron*. 2012 Oct
1108 4;76(1):209–22.
- 1109 27. Polack P-O, Friedman J, Golshani P. Cellular mechanisms of brain state-
1110 state-dependent gain modulation in visual cortex. *Nat Neurosci*. 2013
1111 Sep;16(9):1331–9.
- 1112 28. Fu Y, Tucciarone JM, Espinosa JS, Sheng N, Darcy DP, Nicoll RA, et al. A
1113 cortical circuit for gain control by behavioral state. *Cell*. 2014 Mar
1114 13;156(6):1139–52.
- 1115 29. Eggermann E, Kremer Y, Crochet S, Petersen CCH. Cholinergic Signals in
1116 Mouse Barrel Cortex during Active Whisker Sensing. *Cell Rep*. 2014
1117 Dec;9(5):1654–60.
- 1118 30. Chen N, Sugihara H, Sur M. An acetylcholine-activated microcircuit drives
1119 temporal dynamics of cortical activity. *Nat Neurosci*. 2015 Jun;18(6):892–
1120 902.
- 1121 31. Moreno-Bote R, Rinzel J, Rubin N. Noise-induced alternations in an
1122 attractor network model of perceptual bistability. *J Neurophysiol*. 2007
1123 Sep;98(3):1125–39.
- 1124 32. Noest AJ, van Ee R, Nijs MM, van Wezel RJA. Percept-choice sequences
1125 driven by interrupted ambiguous stimuli: A low-level neural model. *J Vis*.
1126 2007 Jun 21;7(8):10.

Pfeffer et al: Neuromodulation and Cortical Excitation-Inhibition Ratio

- 1127 33. Crochet S, Poulet JFA, Kremer Y, Petersen CCH. Synaptic Mechanisms
1128 Underlying Sparse Coding of Active Touch. *Neuron*. 2011 Mar;69(6):1160–
1129 75.
- 1130 34. Haider B, Häusser M, Carandini M. Inhibition dominates sensory
1131 responses in the awake cortex. *Nature*. 2013 Jan 3;493(7430):97–100.
- 1132 35. Donner TH, Sagi D, Bonnefante YS, Heeger DJ. Retinotopic Patterns of
1133 Correlated Fluctuations in Visual Cortex Reflect the Dynamics of
1134 Spontaneous Perceptual Suppression. *J Neurosci*. 2013 Jan
1135 30;33(5):2188–98.
- 1136 36. Brouwer GJ, van Ee R. Visual cortex allows prediction of perceptual states
1137 during ambiguous structure-from-motion. 2007;27(5):1015–23.
- 1138 37. Beggs JM, Plenz D. Neuronal avalanches in neocortical circuits. *J*
1139 *Neurosci Off J Soc Neurosci*. 2003 Dec 3;23(35):11167–77.
- 1140 38. Shew WL, Yang H, Petermann T, Roy R, Plenz D. Neuronal avalanches
1141 imply maximum dynamic range in cortical networks at criticality. *J Neurosci*
1142 *Off J Soc Neurosci*. 2009 Dec 9;29(49):15595–600.
- 1143 39. Donner TH, Siegel M. A framework for local cortical oscillation patterns.
1144 *Trends Cogn Sci*. 2011 May;15(5):191–9.
- 1145 40. Bauer M, Kluge C, Bach D, Bradbury D, Heinze HJ, Dolan RJ, et al.
1146 Cholinergic enhancement of visual attention and neural oscillations in the
1147 human brain. 2012;22(5):397–402.
- 1148 41. Reimer J, Froudarakis E, Cadwell CR, Yatsenko D, Denfield GH, Tolias AS.
1149 Pupil fluctuations track fast switching of cortical states during quiet
1150 wakefulness. *Neuron*. 2014 Oct 22;84(2):355–62.
- 1151 42. McGinley MJ, David SV, McCormick DA. Cortical Membrane Potential
1152 Signature of Optimal States for Sensory Signal Detection. *Neuron*. 2015
1153 Jul 1;87(1):179–92.
- 1154 43. Vinck M, Batista-Brito R, Knoblich U, Cardin JA. Arousal and locomotion
1155 make distinct contributions to cortical activity patterns and visual encoding.
1156 *Neuron*. 2015 May 6;86(3):740–54.
- 1157 44. Meindertsma T, Kloosterman NA, Nolte G, Engel AK, Donner TH. Multiple
1158 Transient Signals in Human Visual Cortex Associated with an Elementary
1159 Decision. *J Neurosci*. 2017 May 11;38:35–16.
- 1160 45. Murphy PR, G O Redmond, Michael O, Robertson IH, Balsters JH. Pupil
1161 diameter covaries with BOLD activity in human locus coeruleus. *Hum Brain*
1162 *Mapp*. 2014;35(8):4140–54.
- 1163 46. Joshi S, Li Y, Kalwani RM, Gold JI. Relationships between Pupil Diameter
1164 and Neuronal Activity in the Locus Coeruleus, Colliculi, and Cingulate
1165 Cortex. *Neuron*. 2016 Jan 6;89(1):221–34.

Pfeffer et al: Neuromodulation and Cortical Excitation-Inhibition Ratio

- 1166 47. Reimer J, McGinley MJ, Liu Y, Rodenkirch C, Wang Q, McCormick DA, et
1167 al. Pupil fluctuations track rapid changes in adrenergic and cholinergic
1168 activity in cortex. *Nat Commun.* 2016 Nov 8;7:13289.
- 1169 48. de Gee JW, Colizoli O, Kloosterman NA, Knapen T, Nieuwenhuis S,
1170 Donner TH. Dynamic modulation of decision biases by brainstem arousal
1171 systems. *eLife* [Internet]. 2017 Apr 11 [cited 2017 May 16];6. Available
1172 from: <http://elifesciences.org/lookup/doi/10.7554/eLife.23232>
- 1173 49. Denève S, Machens CK. Efficient codes and balanced networks. *Nat*
1174 *Neurosci.* 2016 Feb 23;19(3):375–82.
- 1175 50. Yizhar O, Fenno LE, Prigge M, Schneider F, Davidson TJ, O’Shea DJ, et al.
1176 Neocortical excitation/inhibition balance in information processing and
1177 social dysfunction. *Nature.* 2011 Jul 27;477(7363):171–8.
- 1178 51. Lisman J. Excitation, inhibition, local oscillations, or large-scale loops: what
1179 causes the symptoms of schizophrenia? *Curr Opin Neurobiol.* 2012
1180 Jun;22(3):537–44.
- 1181 52. Nelson SB, Valakh V. Excitatory/Inhibitory Balance and Circuit
1182 Homeostasis in Autism Spectrum Disorders. *Neuron.* 2015 Aug
1183 19;87(4):684–98.
- 1184 53. Fuchs T, Jefferson SJ, Hooper A, Yee P-H, Maguire J, Luscher B.
1185 Disinhibition of somatostatin-positive GABAergic interneurons results in an
1186 anxiolytic and antidepressant-like brain state. *Mol Psychiatry.* 2017
1187 Jun;22(6):920–30.
- 1188 54. Isaacson JS, Scanziani M. How inhibition shapes cortical activity. *Neuron.*
1189 2011 Oct 20;72(2):231–43.
- 1190 55. Froemke RC, Merzenich MM, Schreiner CE. A synaptic memory trace for
1191 cortical receptive field plasticity. *Nature.* 2007 Nov 15;450(7168):425–9.
- 1192 56. Martins ARO, Froemke RC. Coordinated forms of noradrenergic plasticity
1193 in the locus coeruleus and primary auditory cortex. *Nat Neurosci.* 2015 Aug
1194 24;18(10):1483–92.
- 1195 57. Sara SJ. The locus coeruleus and noradrenergic modulation of cognition.
1196 *Nat Rev Neurosci.* 2009 Mar;10(3):211–23.
- 1197 58. Dayan P. Twenty-Five Lessons from Computational Neuromodulation.
1198 *Neuron.* 2012 Oct;76(1):240–56.
- 1199 59. Robbins TW, Arnsten AFT. The neuropsychopharmacology of fronto-
1200 executive function: monoaminergic modulation. *Annu Rev Neurosci.*
1201 2009;32:267–87.
- 1202 60. Montague PR, Hyman SE, Cohen JD. Computational roles for dopamine in
1203 behavioural control. *Nature.* 2004 Oct 14;431(7010):760–7.

Pfeffer et al: Neuromodulation and Cortical Excitation-Inhibition Ratio

- 1204 61. Haber SN, Knutson B. The reward circuit: linking primate anatomy and
1205 human imaging. *Neuropsychopharmacol Off Publ Am Coll*
1206 *Neuropsychopharmacol*. 2010 Jan;35(1):4–26.
- 1207 62. Morrison JH, Foote SL. Noradrenergic and serotonergic innervation of
1208 cortical, thalamic, and tectal visual structures in Old and New World
1209 monkeys. *J Comp Neurol*. 1986 Jan 1;243(1):117–38.
- 1210 63. Ramos BP, Arnsten AFT. Adrenergic pharmacology and cognition: focus
1211 on the prefrontal cortex. *Pharmacol Ther*. 2007 Mar;113(3):523–36.
- 1212 64. Salgado H, Treviño M, Atzori M. Layer- and area-specific actions of
1213 norepinephrine on cortical synaptic transmission. *Brain Res*. 2016 Jun
1214 15;1641(Pt B):163–76.
- 1215 65. Pfeffer CK, Xue M, He M, Huang Z, Scanziani M. Inhibition of inhibition in
1216 visual cortex: the logic of connections between molecularly distinct
1217 interneurons. *Nat Neurosci*. 2013;16(8):1068–76.
- 1218 66. Pakan JM, Lowe SC, Dylka E, Keemink SW, Currie SP, Coutts CA, et al.
1219 Behavioral-state modulation of inhibition is context-dependent and cell type
1220 specific in mouse visual cortex. *Elife*. 2016;5.
- 1221 67. Linkenkaer-Hansen K, Nikulin VV, Palva S, Ilmoniemi RJ, Palva JM.
1222 Prestimulus oscillations enhance psychophysical performance in humans.
1223 *J Neurosci Off J Soc Neurosci*. 2004 Nov 10;24(45):10186–90.
- 1224 68. Swadlow HA. Thalamocortical control of feed-forward inhibition in awake
1225 somatosensory “barrel” cortex. *Philos Trans R Soc B Biol Sci*. 2002 Dec
1226 29;357(1428):1717–27.
- 1227 69. Kepecs A, Fishell G. Interneuron cell types are fit to function. *Nature*. 2014
1228 Jan 15;505(7483):318–26.
- 1229 70. Ponce-Alvarez A, He BJ, Hagmann P, Deco G. Task-Driven Activity
1230 Reduces the Cortical Activity Space of the Brain: Experiment and Whole-
1231 Brain Modeling. Graham LJ, editor. *PLOS Comput Biol*. 2015 Aug
1232 28;11(8):e1004445.
- 1233 71. Carter OL, Pettigrew JD, Hasler F, Wallis GM, Liu GB, Hell D, et al.
1234 Modulating the rate and rhythmicity of perceptual rivalry alternations with
1235 the mixed 5-HT_{2A} and 5-HT_{1A} agonist psilocybin. *Neuropsychopharmacol*
1236 *Off Publ Am Coll Neuropsychopharmacol*. 2005 Jun;30(6):1154–62.
- 1237 72. Noudoost B, Moore T. Control of visual cortical signals by prefrontal
1238 dopamine. *Nature*. 2011 May 15;474(7351):372–5.
- 1239 73. Honey CJ, Thesen T, Donner TH, Silbert LJ, Carlson CE, Devinsky O, et al.
1240 Slow Cortical Dynamics and the Accumulation of Information over Long
1241 Timescales. *Neuron*. 2012 Oct;76(2):423–34.
- 1242 74. Murray JD, Bernacchia A, Freedman DJ, Romo R, Wallis JD, Cai X, et al.
1243 A hierarchy of intrinsic timescales across primate cortex. *Nat Neurosci*.
1244 2014 Nov 10;17(12):1661–3.

Pfeffer et al: Neuromodulation and Cortical Excitation-Inhibition Ratio

- 1245 75. Chaudhuri R, Knoblauch K, Gariel M-A, Kennedy H, Wang X-J. A Large-
1246 Scale Circuit Mechanism for Hierarchical Dynamical Processing in the
1247 Primate Cortex. *Neuron*. 2015 Oct 21;88(2):419–31.
- 1248 76. Wang X-JJ. Probabilistic decision making by slow reverberation in cortical
1249 circuits. 2002;36(5):955–68.
- 1250 77. Beggs JM. The criticality hypothesis: how local cortical networks might
1251 optimize information processing. *Philos Transact A Math Phys Eng Sci*.
1252 2008 Feb 13;366(1864):329–43.
- 1253 78. Bak P, Tang C, Wiesenfeld K. Self-organized criticality: An explanation of
1254 the 1/f noise. *Phys Rev Lett*. 1987 Jul 27;59(4):381–4.
- 1255 79. Bak P. How nature works: the science of self-organized criticality [Internet].
1256 New York, NY, USA: Copernicus; 1996 [cited 2017 Apr 20]. Available from:
1257 <http://catalog.hathitrust.org/api/volumes/oclc/34623628.html>
- 1258 80. Chialvo DR. Emergent complex neural dynamics. *Nat Phys*. 2010
1259 Oct;6(10):744–50.
- 1260 81. Hesse J, Gross T. Self-organized criticality as a fundamental property of
1261 neural systems. *Front Syst Neurosci*. 2014;8:166.
- 1262 82. Kinouchi O, Copelli M. Optimal dynamical range of excitable networks at
1263 criticality. *Nat Phys*. 2006 May;2(5):348–51.
- 1264 83. Shew W, Yang H, Yu S, Roy R. Information capacity and transmission are
1265 maximized in balanced cortical networks with neuronal avalanches. *J*
1266 2011;
- 1267 84. Shriki O, Yellin D. Optimal Information Representation and Criticality in an
1268 Adaptive Sensory Recurrent Neuronal Network. *PLoS Comput Biol*. 2016
1269 Feb;12(2):e1004698.
- 1270 85. Priesemann V, Valderrama M, Wibral M, Le Van Quyen M. Neuronal
1271 avalanches differ from wakefulness to deep sleep--evidence from
1272 intracranial depth recordings in humans. *PLoS Comput Biol*.
1273 2013;9(3):e1002985.
- 1274 86. Arviv O, Goldstein A, Shriki O. Near-Critical Dynamics in Stimulus-Evoked
1275 Activity of the Human Brain and Its Relation to Spontaneous Resting-State
1276 Activity. *J Neurosci Off J Soc Neurosci*. 2015 Oct 14;35(41):13927–42.
- 1277 87. Fagerholm ED, Lorenz R, Scott G, Dinov M, Hellyer PJ, Mirzaei N, et al.
1278 Cascades and Cognitive State: Focused Attention Incurs Subcritical
1279 Dynamics. *J Neurosci*. 2015 Mar 18;35(11):4626–34.
- 1280 88. Shew WL, Clawson WP, Pobst J, Karimipanah Y, Wright NC, Wessel R.
1281 Adaptation to sensory input tunes visual cortex to criticality. *Nat Phys*.
1282 2015 Jun 22;11(8):659–63.

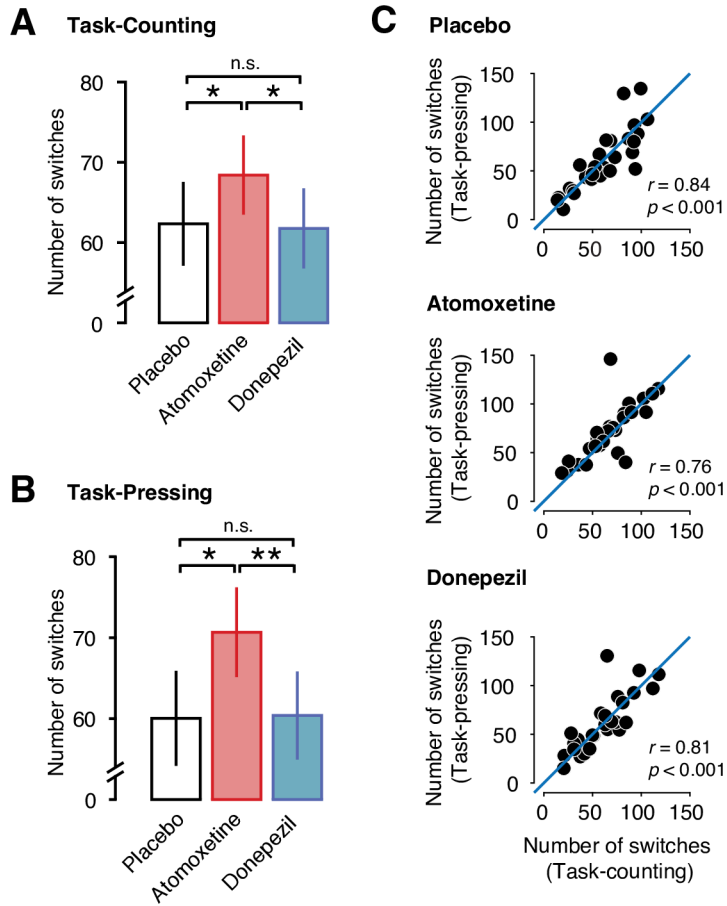
Pfeffer et al: Neuromodulation and Cortical Excitation-Inhibition Ratio

- 1283 89. Tiseo, Rogers, Friedhoff. Pharmacokinetic and pharmacodynamic profile of
1284 donepezil HCl following evening administration: Evening administration of
1285 donepezil HCl. *Br J Clin Pharmacol*. 1998 Jan 4;46(S1):13–8.
- 1286 90. Sauer J-M, Ring BJ, Witcher JW. Clinical pharmacokinetics of atomoxetine.
1287 *Clin Pharmacokinet*. 2005;44(6):571–90.
- 1288 91. Wallach H, O’connell DN. The kinetic depth effect. *J Exp Psychol*. 1953
1289 Apr;45(4):205–17.
- 1290 92. Sperling G, Doshier BA, Landy MS. How to study the kinetic depth effect
1291 experimentally. *J Exp Psychol Hum Percept Perform*. 1990
1292 May;16(2):445–50.
- 1293 93. Engbert R, Kliegl R. Microsaccades uncover the orientation of covert
1294 attention. *Vision Res*. 2003 Apr;43(9):1035–45.
- 1295 94. Oostenveld R, Fries P, Maris E, Schoffelen J-M. FieldTrip: Open source
1296 software for advanced analysis of MEG, EEG, and invasive
1297 electrophysiological data. *Comput Intell Neurosci*. 2011;2011:156869.
- 1298 95. Bell AJ, Sejnowski TJ. An information-maximization approach to blind
1299 separation and blind deconvolution. *Neural Comput*. 1995 Nov;7(6):1129–
1300 59.
- 1301 96. Hyvarinen A. Fast and robust fixed-point algorithms for independent
1302 component analysis. *IEEE Trans Neural Netw*. 1999 May;10(3):626–34.
- 1303 97. Hipp JF, Siegel M. Dissociating neuronal gamma-band activity from cranial
1304 and ocular muscle activity in EEG. *Front Hum Neurosci [Internet]*. 2013
1305 [cited 2017 Apr 20];7. Available from:
1306 <http://journal.frontiersin.org/article/10.3389/fnhum.2013.00338/abstract>
- 1307 98. Mitra PP, Pesaran B. Analysis of dynamic brain imaging data. *Biophys J*.
1308 1999 Feb;76(2):691–708.
- 1309 99. Pascual-Marqui RD, Lehmann D, Koukkou M, Kochi K, Anderer P, Saletu
1310 B, et al. Assessing interactions in the brain with exact low-resolution
1311 electromagnetic tomography. *Philos Transact A Math Phys Eng Sci*. 2011
1312 Oct 13;369(1952):3768–84.
- 1313 100. Nolte G. The magnetic lead field theorem in the quasi-static approximation
1314 and its use for magnetoencephalography forward calculation in realistic
1315 volume conductors. *Phys Med Biol*. 2003 Nov 21;48(22):3637–52.
- 1316 101. Pfeffer T, Linkenkaer-Hansen K, Avramiea A-E, Engel AK, Donner TH.
1317 Noradrenaline increases long-range temporal correlations of neuronal
1318 alpha oscillations in the human cortex. In 2015. p. 393.27.
- 1319 102. Peng CK, Buldyrev SV, Havlin S, Simons M, Stanley HE, Goldberger AL.
1320 Mosaic organization of DNA nucleotides. *Phys Rev E Stat Phys Plasmas
1321 Fluids Relat Interdiscip Top*. 1994 Feb;49(2):1685–9.

Pfeffer et al: Neuromodulation and Cortical Excitation-Inhibition Ratio

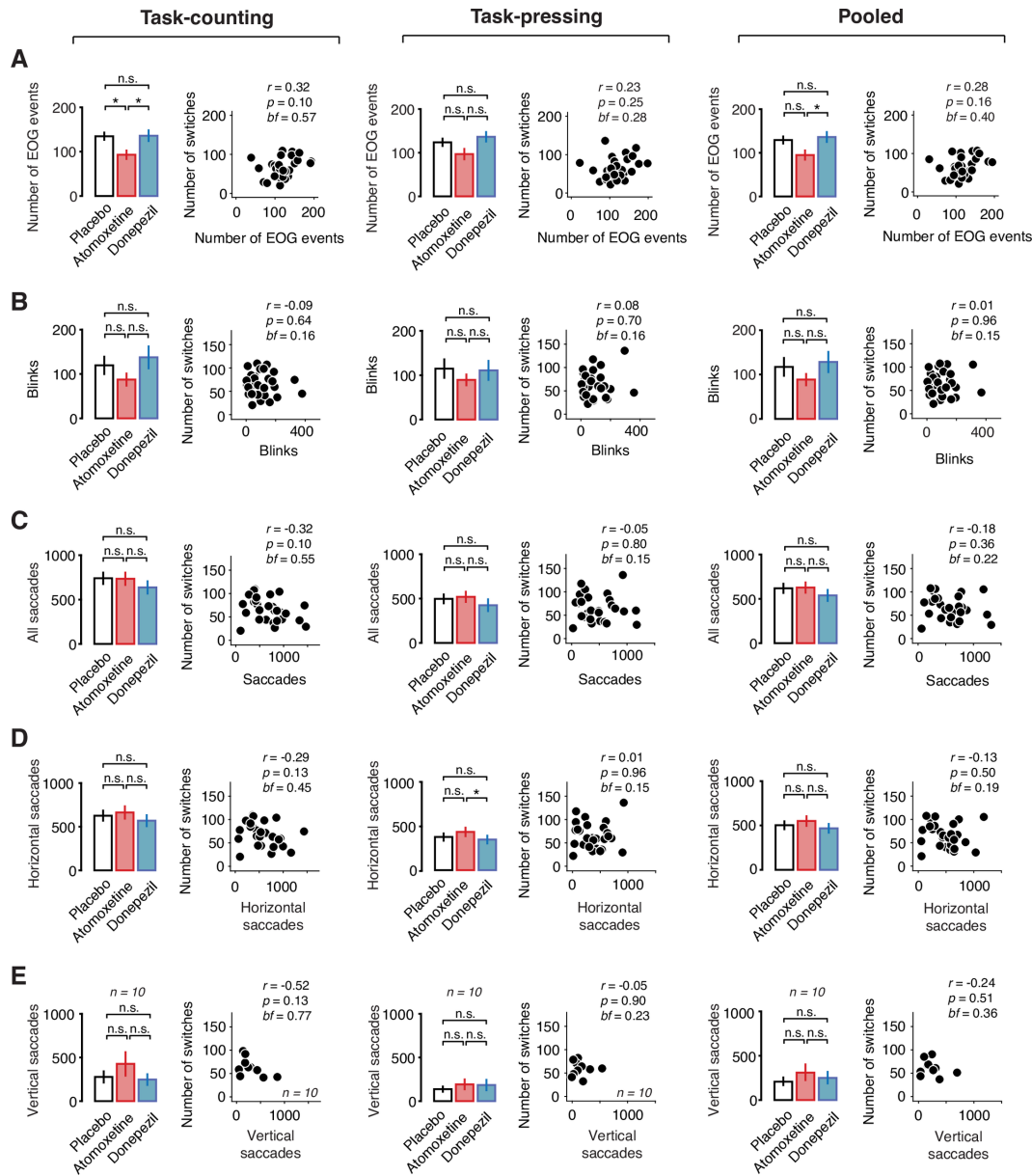
- 1322 103. Hardstone R, Poil S-S, Schiavone G, Jansen R, Nikulin VV, Mansvelder
1323 HD, et al. Detrended Fluctuation Analysis: A Scale-Free View on Neuronal
1324 Oscillations. *Front Physiol* [Internet]. 2012 [cited 2017 Apr 20];3. Available
1325 from:
1326 <http://journal.frontiersin.org/article/10.3389/fphys.2012.00450/abstract>
- 1327 104. Rouder JN, Speckman PL, Sun D, Morey RD, Iverson G. Bayesian t tests
1328 for accepting and rejecting the null hypothesis. *Psychon Bull Rev*. 2009
1329 Apr;16(2):225–37.
- 1330 105. Wetzels R, Wagenmakers E-J. A default Bayesian hypothesis test for
1331 correlations and partial correlations. *Psychon Bull Rev*. 2012
1332 Dec;19(6):1057–64.
- 1333 106. Nichols TE, Holmes AP. Nonparametric permutation tests for functional
1334 neuroimaging: A primer with examples. *Hum Brain Mapp*. 2002
1335 Jan;15(1):1–25.
- 1336 107. Maris E, Oostenveld R. Nonparametric statistical testing of EEG- and
1337 MEG-data. *J Neurosci Methods*. 2007 Aug 15;164(1):177–90.
- 1338 108. Eiben AE, Smit SK. Parameter tuning for configuring and analyzing
1339 evolutionary algorithms. *Swarm Evol Comput*. 2011 Mar;1(1):19–31.
- 1340 109. Goodman DF, Stimberg M, Yger P, Brette R. Brian 2: neural simulations on
1341 a variety of computational hardware. *BMC Neurosci*. 2014;15(Suppl
1342 1):P199.
- 1343

SUPPLEMENTARY FIGURES



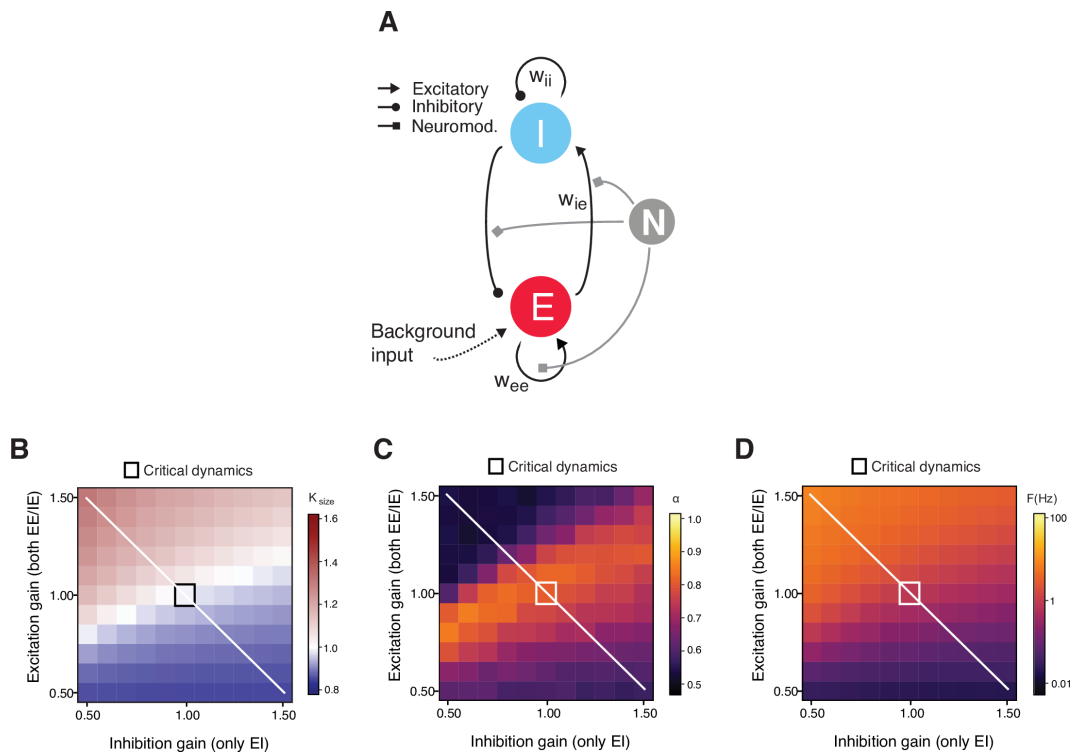
S1 Fig. Similar atomoxetine-related effects in both Task-counting and Task-resting conditions. **(A)** Number of perceptual alternations reported by the subjects per 10 min run for Task-counting condition. **(B)** Same as (A), but for Task-pressing condition. **(C)** Relation between the number of reported alternations during Task-counting (x-axis) and Task-pressing (y-axis). The blue line depicts a linear relation with slope 1 as a reference.

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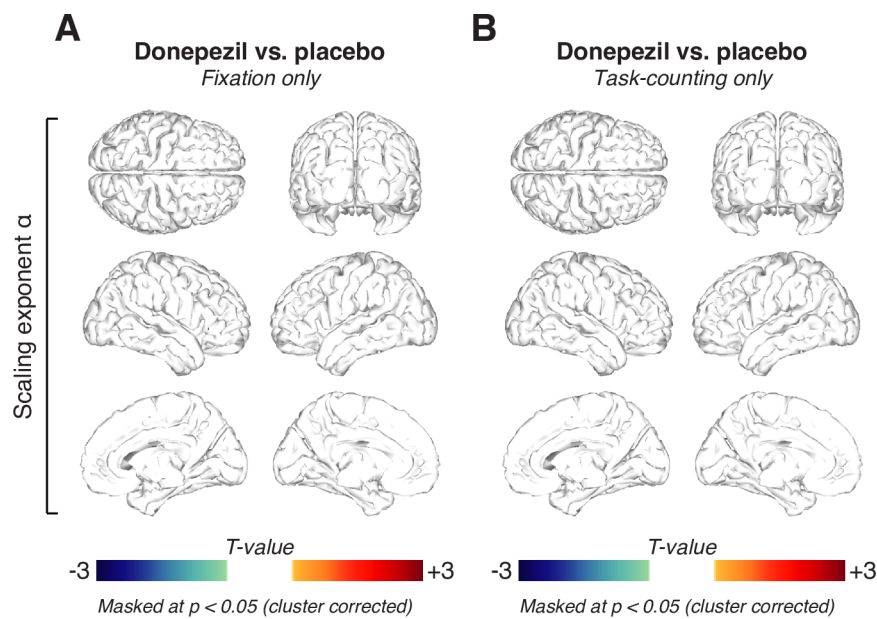


S2 Fig. Change in perceptual alternation rate is not due to change in blinks or fixational eye movements. **(A)** Number of EOG events for during Task-counting (left), Task-pressing (middle) and pooled across both conditions (right). Scatter plots depict the relation between the number of EOG events (x-axis) and the number of reported perceptual alternations (y-axis). **(B)** Same as (A), but for the number of detected eye blinks. **(C)** Same as (A) and (B), but for the number of saccades (horizontal and vertical). **(D)** Same as (C), but for horizontal saccades only. **(E)** Same as (D), but for vertical saccades only.

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S3 Fig. Different version of modulation of E/I ratio in cortical patch model **(A)** Neuromodulation was simulated as a gain modulation term multiplied with excitatory (EE and IE) and/or inhibitory (EI only) synaptic weights. **(B)** κ as a function of excitatory and inhibitory connectivity (with a spacing of 2.5%; means across 10 simulations per cell). The region of $\kappa \sim 1$, overlaps with the region of $\alpha > 0.5$ and splits the phase space into an excitation-dominant ($\kappa > 1$) and an inhibition-dominant region ($\kappa < 1$). **(C)** Same as (B), but for scaling exponent α . **(D)** Same as (B) and (C), but for firing rate.



S4 Fig. No donepezil-related changes in scaling exponent in neither behavioral contexts. **(A)** Spatial distribution of donepezil-induced changes in scaling exponent α during Fixation, thresholded at $p = 0.05$ (two-sided cluster-based permutation test). **(B)** As (A), but for Task-counting.