LETTER: DISCOVERIES

Cryptic variation in quantitative spatial gene expression in *Drosophila*

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ABSTRACT (150 words)

Robustness in development allows for the accumulation of cryptic variation, and this largely neutral

variation is potentially important for both evolution and complex disease phenotypes. However, it has

generally only been investigated as variation in the response to large genetic perturbations. Here we

use newly developed methods to quantify spatial gene expression patterns during development of the

Drosophila eye disc, and uncover cryptic variation in wildtype developmental systems. We focus on

four conserved morphogens, *hairy*, *atonal*, *hedgehog*, and *Delta*, that are involved in specifying

ommatidia in the developing eye. We find abundant cryptic variation within and between species,

genotypes, and sexes, as well as cryptic variation in the regulatory logic between *atonal* and *hairy* and

their regulators, *Delta* and *hedgehog*. This work paves the way for a synthesis between population

and quantitative genetic approaches with that of developmental biology.

RESULTS AND DISCUSSION (2,000 words)

Cryptic variation accumulates due to the robustness of developmental systems to mutational perturbation (Duveau and Felix 2012; Felix 2012; Felix and Barkoulas 2012; Kienle and Sommer 2013; Paaby and Rockman 2014; Paaby and Gibson 2016). Although the final phenotype is the same, the presence of this cryptic variation effects the response to mutational or environmental perturbation, and thus the evolutionary potential of the phenotype. In the past cryptic variation has been experimentally exposed through the use of synthesized genetic backgrounds with large perturbations that prevent buffering of variation (Braendle et al. 2010; Woodruff et al. 2010; Duveau and Felix 2012; Chandler et al. 2014; Taylor and Ehrenreich 2015). While this work provides valuable insights into the importance of cryptic variation, cryptic variation in developmental processes in natural genotypes has not been experimentally observed.

Indeed, natural genetic variation within populations has long been the purview of evolutionary and population geneticists, while developmental biologists focus on the effect of large mutations in otherwise isogenic backgrounds (Paaby and Gibson 2016). This dearth of work on cryptic developmental variation in wildtype genetic backgrounds is in part because developmental approaches have long been restricted to data that is at best semi-quantitative (i.e. *in situ* hybridization, antibody staining). Indeed, gene expression studies are generally spatial or quantitative, but not both. Without quantitative replication, there can be no rigorous statistical testing when developmental processes are compared among conditions, including health versus disease. This is especially important given the potential for cryptic variation to result in disease phenotypes, for example the complex disease phenotypes seen in humans after their recent, rapid change in lifestyle (Felix 2012). The goal of this paper is to provide a proof-of-principle for the presence of abundant cryptic developmental variation in gene expression that is both spatial and quantitative. We focus on the well-known morphogens driving ommatidia specification in *Drosophila* (Li et al. 1995; Raj et al. 2008; Tsachaki and Sprecher 2011; Atkins et al. 2013; Shah et al. 2016).

Here we use hybridization chain reaction (HCR) to quantitatively measure spatial gene expression in multiple genotypes from two sexes of two species (*Drosophila melanogaster* and *D. simulans*). We

do this in the eye imaginal disc, which is initially patterned by a wave of differentiation marked by a

visible indentation of the tissue, termed the morphogenetic furrow (MF), that passes from the posterior

to the anterior of the disc, giving each disc an element of both time and space in development (Fig 1).

The furrow is initiated by *hedgehog,* which both represses (short range) and activates (long range)

hairy (Fig 1) (Felsenfeld and Kennison 1995; Strutt and Mlodzik 1997). *hedgehog* also activates the

expression of *atonal*, driving the furrow anteriorly (Heberlein et al. 1993; Ma et al. 1993; Struhl 1999).

Delta/Notch is under the control of *hedgehog* and represses *atonal* (Firth and Baker 2005; Gavish et

al. 2016). We analyze the spatial quantitative expression of *hedgehog*, *hairy*, *atonal*, and *Delta* to

understand the evolving cryptic regulatory logic of the gene network and cryptic changes in spatial

dynamics between sexes and species.

HCR is unique in that it produces gene expression patterns that are both quantitative and spatial

(Supplementary Materials). The DNA probes were designed and synthesized by Molecular

Instruments (Choi et al. 2014) (Table S1). Four genes were multiplexed in each preparation as

orthogonally-designed hairpins allowed the simultaneous amplification of their target sequences (Fig

1, S1). Eye imaginal discs were dissected using standard procedures from three genotypes and both

sexes of previously published *D. melanogaster* and *D. simulans* inbred panels (Mackay et al. 2012;

Signor, et al. *in review*). In order to compare gene expression patterns between eye imaginal discs

gene expression was quantified and each image was registered to a common grid such that the same

regions are being compared despite variation in disc size and deformation (Supplementary Materials).

We interpret cryptic variation in quantitative spatial patterns of gene expression in light of

regulatory relationships among them. We approach a spatial and quantitative analysis of these gene

expression patterns in three ways, first by explicitly creating a spatial gene expression profile and

comparing between genotypes, sexes, and species. Second, we were interested in examining if the

regulatory relationship between these genes had evolved between species or harbors variation within

a species. Lastly, we investigated the possibility that the spatial relationship between these genes

relative to the MF had evolved or harbors variation within populations.

First, to characterize the spatiotemporal dynamics of transcriptional activity along the anterior-

posterior axis, we took the spatial average of signal across the dorsal-ventral axis and compared

between species, genotypes, and sexes (Fig 2). We fit curves to each gene expression profile with

mgcv in R, using a generalized additive model with integrated smoothness estimation (Fig 2,

Supplemental Materials). Smoothing terms are represented using penalized regression splines. The

curves were fit to the original range of values and down sampled to eight points to capture their overall

profile.

We found abundant cryptic spatial quantitative variation in expression profiles (Fig 2). The

expression profile of *hairy* around the morphogenetic furrow harbors cryptic variation between

genotypes and there is an interaction between genotype and sex (Fig 2B & D, $p = 2 \times 10^{-3}$, $p = 0.02$).

There is also been cryptic variation between species (Fig 2B & D, $p = 3 \times 10^{-4}$). While *atonal* has not

evolved between species, there is cryptic variation in expression profile between genotypes, sexes,

and there is an interaction between genotype and sex (Fig 2C & D, $p = 4 \times 10^{-4}$, $p = .02$, $p = .02$).

Surprisingly, given the conservation of *Delta* in general, *Delta* harbors variation in spatial quantitative

expression behind the furrow between genotypes and sexes (Fig 2D, $p = 2 \times 10^{-3}$, $p = 7 \times 10^{-4}$) and

there are significant interactions between genotype and sex (Fig 2D, $p = 2 \times 10^{-3}$). There has also

been evolution of *Delta* between species, and evolution of the interaction between species and sex (p

$= .03$, $p = 3 \times 10^{-4}$). *hedgehog* is not different between species but is significantly different between

genotypes, sexes, and there is an interaction between the two ($p = 5 \times 10^{-4}$, $p = .05$, $p = 1 \times 10^{-3}$).

There is also a significant interaction between species and sex ($p = .01$).

Thus, *hairy* and *Delta* have evolved different spatial quantitative expression patterns between

species, while *hairy*, *atonal*, *Delta*, and *hedgehog* harbor variation within species and sexes. Given

that there are regulatory relationships between these genes, it is interesting to see that they do not all

harbor variation for the same factors. This could potentially be due to the influence of other

unmeasured regulatory factors, or to variation in the relationship between these genes and other

components in the gene regulatory network.

In the previous analysis, we investigated variation in the cryptic spatial quantitative expression

pattern of genes in the morphogenetic furrow. Here, we will investigate the possibility that genes in the

MF have evolved, or harbor variation, for how they affect each other in particular cells, termed here

the 'cryptic variation in regulatory logic'. For example, is high *atonal* expression associated with high

expression of *hedgehog*, given that *hedgehog* activates *atonal*? We used the following equations to

determine the relationship between the expression of these genes:

$$hairy(i,s) = k^{Dl} \times Dl(i,s) + k^{hh} \times hh(i,s) + \alpha$$

$$atonal(i,s) = k^{Dl} \times Dl(i,s) + k^{hh} \times hh(i,s) + \alpha$$

The coefficient k and constant α were fit using standard methods for multiple regression. Here

$hairy(i,s)$ and $atonal(i,s)$ are the measured expression level of each gene in cell i in individual s.

$Dl(i,s)$ and $hh(i,s)$ are vectors containing the corresponding expression levels of *hairy* and *atonal*'s

regulators *Delta* and *hedgehog*. To determine if the regulatory logic is the same between genotypes

and species we can then use the regression coefficients from these models in a MANOVA. We note

that because we cannot parse overall changes in spatial expression pattern from cryptic variation in

regulatory logic, we focus on genes that are known to be regulated by the other genes in the pathway,

i.e. *hairy*, *Delta*, and *hedgehog*. We also cannot exclude the possibility that other unmeasured genes

are responsible for producing this variation.

There is cryptic variation in regulatory logic in *hairy* and its upstream regulators *Delta* and

hedgehog between species and sexes (Fig 3A-C, $p = .03$, $p = .03$). We illustrate this difference

between species in Figure 3, where a different relationship between *hairy* and *hedgehog* is visible

between *D. melanogaster* and *D. simulans*. The regulatory logic of *atonal* does not seem strictly to

have evolved between species, but there is a significant interaction between species and sex ($p = 1 \times$

10^{-3}). Furthermore, while there was no significant effect of genotype for *hairy*, there is for *atonal*,

indicating that there is cryptic variation segregating in the population effecting the relationship between

atonal, hedgehog, and *Delta* (p = 1.6 x 10^{-5}). There is also a significant interaction between genotype

and sex (p = 1 x 10^{-3}). Thus, the relationship between *hairy* and *atonal* and their regulators has

evolved between species and sexes in *hairy*, and between genotypes and sex in *atonal*. For example,

in *D. simulans* this appears as an approximately linear relationship between *hairy* and *hedgehog*

expression that is not present in *D. melanogaster* (Fig 3A).

Cryptic variation in regulatory logic between sexes and species is somewhat surprising. Eye size

does seem to vary both within and between species, but the exact nature of this variation is unclear.

By one account female eyes in *D. melanogaster* are smaller than expected based on allometry, while

other work finds that the opposite is the case in both *D. melanogaster* and *D. simulans* (Cowley and

Atchley 1988; Posnien et al. 2012). Body size is at least in part controlled by *Sex-lethal,* and sex

specific development in somatic tissues are governed by *doublesex* (Rideout et al. 2010; Mathews et

al. 2017). We have detected *doublesex* expression in the eye anterior to the morphogenetic furrow, in

cells prior to cell cycle arrest at the furrow (Fig S2). It is possible that an interaction between

doublesex and *hairy*, *atonal*, *Delta*, or *hedgehog* is mediating the change in cryptic variation in

regulatory logic between sexes. It is also possible that some of this variation is not cryptic, and in fact

has an effect on sex-specific development of the eyes. To date very little sexual dimorphism has been

documented in *Drosophila* eyes, other than a > 10% difference in the composition of ommatidial

subtypes in some genotypes (Hilbrant et al. 2014).

Lastly, we wanted to understand if there is cryptic variation for the relationship between the furrow

and gene expression, or if variation existed for the size of the furrow overall. For example, the furrow

was called as the position of overlap between *atonal* and *hairy* expression, but it is unclear how the

overall gene expression pattern of these genes relates to their overlap. For example, is the position of

maximum expression of each always the same relative to the furrow? Two processes occurred to

make the furrow comparable between samples, they were shifted to occupy the same position

depending upon the position of overlap of *hairy* and *atonal*, and they were scaled to occupy the same

total area. The amount required to scale will depend both on the size of the original disc and the width

of the furrow relative to the disc. To account for differences in size we include the number of rows in

the original disc prior to any transformations as a cofactor and perform ANOVA in R. The amount that

they were shifted is not significant for genotype, sex, or species, suggesting that the relationship of

maximum gene expression with the furrow does not vary. However, the amount that they were scaled

is, after accounting for original differences in size, between species ($p = 1.38 \times 10^{-6}$). This suggests

that the total relative width of the MF varies between species, but not between genotypes or sexes.

This is also suggestive of evolving interrelationships among genes that could result in broader or

narrower areas in which they enhance or suppress expression of one another.

In the past, the developmental approach to understanding gene networks has been to analyze

large effect mutations and their qualitative downstream effects. This results in robust data on

perturbed regulatory networks, but obfuscates information on more normal interrelationships among

genes. Similarly, the evolutionary approach to development generally targets large changes that have

occurred over broad phylogenetic distances (Signor et al. 2016). Here we take an entirely different

approach by focusing on small, quantitative variation between genotypes and closely related species.

Using this approach, we were able to quantify the regulatory relationships among genes and to

observe how these relationships are altered between sexes and how they evolve between sister taxa.

Reconciling the speed of this cryptic developmental evolution with the patterns of molecular evolution

in the underlying genes will yield new insights on the rules governing microevolution of gene

regulatory networks.

AUTHOR CONTRIBUTIONS: S. A. performed the staining and imaging of the eye discs, K. K.

processed the imaging data, S.S. conceived of the experiment, analyzed the processed data and

wrote the paper, S. N. conceived of the experiment, coordinated the research, and co-wrote the paper.

ACKNOWLEDGEMENTS: The authors thank S. Restrepo, M. Samsonova, P. Marjoram, J. Butler for

help with experimental procedures and manuscript preparation. This work was supported by grants

U01GM103804 and RO1GM102227 to S.V.

Figure 1. **a)** A summary of the eye patterning genes and pathway explored in this paper. The position

of the morphogenetic furrow is shown in pink, and its direction of movement indicated below.

Regulatory relationships are illustrated either as repression (bar) or activation (arrow). Shown at right

is a composite image of all four genes (*hairy* (white), *atonal* (red), *Delta* (green), and *hedgehog*

(blue)). Please note that this is a very simplified illustration, intended as a summary between the

genes that are the subject of this paper rather than an exhaustive illustration of the eye patterning

gene network. **b)** Example gene expression patterns of each gene. *hedgehog* and *Delta* were

recolored in Photoshop (Adobe, v. 21.1) without altering any aspects of expression level or contrast.

d)

hairy

Effect	numDF	denDF	F-value	p-value
Species	8	35	5.1	3×10^{-4}
Genotype	32	152	2.07	2×10^{-3}
Sex	8	35	1.08	0.32
Species x Sex	8	35	1.94	0.08
Genotype x Sex	32	152	1.73	**0.02**

atonal

Effect	numDF	denDF	F-value	p-value
Species	8	35	1.53	0.18
Genotype	32	152	2.31	4×10^{-4}
Sex	8	35	2.6	**0.02**
Species x Sex	8	35	1.91	0.09
Genotype x Sex	32	152	1.73	**0.02**

Delta

Effect	numDF	denDF	F-value	p-value
Species	8	35	2.43	**0.03**
Genotype	32	152	2.07	2×10^{-3}
Sex	8	35	4.55	7×10^{-4}
Species x Sex	8	35	5.03	3×10^{-4}
Genotype x Sex	32	152	2.03	2×10^{-3}

hedgehog

Effect	numDF	denDF	F-value	p-value
Species	8	35	0.87	0.55
Genotype	32	152	2.28	5×10^{-4}
Sex	8	35	2.23	**0.05**
Species x Sex	8	35	2.88	**0.01**
Genotype x Sex	32	152	2.17	1×10^{-3}

Figure 2. **a)** An example of the data-processing used to produce gene expression profiles. *hairy*

expression is shown on the left for an imaginal disc, while on the right is the same disc after it has

been segmented and registered. The bottom is an example of a curve being fitted to the gene

expression profiles, though note that the curve corresponds to the average in a given row (*x*-axis). **b)**

An illustration of variation in hairy expression between species and sexes. Curves shown are fitted to

all genotypes within a sex and species with confidence intervals indicated in gray. Only the effect of

species was large enough to be significant, though it is clear that perhaps with a larger sample size an

interaction effect would also have been significant. **c)** An illustration of variation in *atonal* expression

between genotypes and sexes. Curves shown are fitted to all samples within a genotype and sex with

confidence intervals indicated in gray. The effect of genotype, sex, and genotype x sex were

significant. All *D. melanogaster* genotypes are shown in shades of purple, *D. simulans* in blue. **d)** The

results of the full model for each gene, significant *p*-values are indicated and bold, with gray shading.

hairy

Effect	numDF	denDF	F-value	p-value
Species	2	41	4.03	**0.03**
Genotype	8	84	1.2	0.31
Sex	2	41	3.95	**0.03**
Species x Sex	2	41	0.78	0.46
Genotype x Sex	8	84	1.11	0.37

atonal

Effect	numDF	denDF	F-value	p-value
Species	2	41	2.64	0.08
Genotype	8	84	5.43	**1.6×10^{-5}**
Sex	2	41	0.67	0.52
Species x Sex	2	41	8.09	**1×10^{-3}**
Genotype x Sex	8	84	2.41	**0.02**

Figure 3. **a)** An example of cryptic variation in regulatory logic between *D. simulans* and *D.*

melanogaster for *hairy* and *hedgehog*. The heat map illustrates the density of points, and thus reflects

the frequency of a given co-expression profile between *hairy* and *hedgehog*. Gene expression values

were log-transformed to better illustrate lower values and split between anterior to the MF and

posterior to the MF. The split between the two regions was to investigate the possibility that *hedgehog*

had a different regulatory relationship with *hairy* depending upon its relationship to the furrow, given

that *hedgehog* is thought to activate *hairy* long range and repress *hairy* short range. Indeed, anterior

to the MF *D. simulans* has an almost linear relationship between *hairy* and *hedgehog*, while in *D.*

melanogaster almost any value of *hedgehog* can be associated with high *hairy* expression. **b)** An

illustration of the change in the relationship between the expression of *hedgehog* and *hairy* between

D. simulans and *D. melanogaster* anterior to the MF. The dotted, darker line is meant to indicate less

of a regulatory relationship than the thick red line. c) The results of the full model investigating

changes in the regulatory relationship between *hairy*, *Delta*, and *hedgehog*, and *atonal*, *Delta*, and

hedgehog.

Atkins M, Jiang Y, Sansores-Garcia L, Jusiak B, Halder G, Mardon G. 2013. Dynamic rewiring of the
Drosophila retinal determination network switches its function from selector to differentiation.
PLoS Genet. 9:e1003731.

Braendle C, Baer CF, Felix M-A. 2010. Bias and evolution of the mutationally accessible phenotypic
space in a developmental system. PLoS Genet. 6:e1000877.

Chandler CH, Chari S, Tack D, Dworkin I. 2014. Causes and consequences of genetic background
effects illuminated by integrative genomic analysis. Genetics 196:1321–1336.

Choi HMT, Beck VA, Pierce NA. 2014. Next-generation in situ hybridization chain reaction: higher
gain, lower cost, greater durability. ACS Nano 8:4284–4294.

Cowley DE, Atchley WR. 1988. Quantitative genetics of *Drosophila melanogaster*. II. Heritabilities and
genetic correlations between sexes for head and thorax traits. Genetics 119:421–433.

Duveau F, Felix M-A. 2012. Role of pleiotropy in the evolution of a cryptic developmental variation in
Caenorhabditis elegans. PLoS Biol. 10:e1001230.

Felix M-A, Barkoulas M. 2012. Robustness and flexibility in nematode vulva development. Trends
Genet. 28:185–195.

Felix M-A. 2012. Evolution in developmental phenotype space. Curr Opin Genet Dev. 22:593–599.

Felsenfeld AL, Kennison JA. 1995. Positional signaling by *hedgehog* in *Drosophila* imaginal disc
development. Development 121:1–10.

Firth LC, Baker NE. 2005. Extracellular signals responsible for spatially regulated proliferation in the
differentiating *Drosophila* eye. Dev Cell 8:541–551.

Gavish A, Shwartz A, Weizman A, Schejter E, Shilo B-Z, Barkai N. 2016. Periodic patterning of the
Drosophila eye is stabilized by the diffusible activator *Scabrous*. Nat. Comm. 7:1–10.

Heberlein U, Wolff T, Rubin GM. 1993. The TGFß homolog *dpp* and the segment polarity gene
hedgehog are required for propagation of a morphogenetic wave in the *Drosophila* retina. Cell
75:913–926.

Hilbrant M, Almudi I, Leite DJ, Kuncheria L, Posnien N, Nunes MDS, McGregor AP. 2014. Sexual
dimorphism and natural variation within and among species in the *Drosophila* retinal mosaic. BMC
Evol Biol 14:240.

Kienle S, Sommer RJ. 2013. Cryptic variation in vulva development by cis-regulatory evolution of a
HAIRY-binding site. Nat. Comm. 4:1714.

Li W, Ohlmeyer JT, Lane ME, Kalderon D. 1995. Function of protein kinase A in *hedgehog* signal
transduction and *Drosophila* imaginal disc development. Cell 80:553–562.

Ma C, Zhou Y, Beachy PA, Moses K. 1993. The segment polarity gene *hedgehog* is required for
progression of the morphogenetic furrow in the developing *Drosophila* eye. Cell 75:927–938.

Mackay TFC, Richards S, Stone EA, Barbadilla A, Ayroles JF, Zhu D, Casillas S, Han Y, Magwire
MM, Cridland JM, et al. 2012. The *Drosophila melanogaster* Genetic Reference Panel. Nature
482:173–178.

Mathews KW, Cavegn M, Zwicky M. 2017. Sexual dimorphism of body size is controlled by dosage of
the X-chromosomal gene *Myc* and by the sex-determining gene *tra* in *Drosophila*.
Genetics.205:1215-1228.

Paaby A, Gibson G. 2016. Cryptic genetic variation in evolutionary developmental genetics. Biology
5:28–13.

Paaby AB, Rockman MV. 2014. Cryptic genetic variation: evolution's hidden substrate. Nat Rev
Genet. 15:247–258.

Posnien N, Hopfen C, Hilbrant M, Ramos-Womack M, Murat S, Schönauer A, Herbert SL, Nunes
MDS, Arif S, Breuker CJ, et al. 2012. Evolution of eye morphology and rhodopsin expression in
the *Drosophila melanogaster* species subgroup. PLoS ONE 7:e37346.

Raj A, van den Bogaard P, Rifkin SA, van Oudenaarden A, Tyagi S. 2008. Imaging individual mRNA
molecules using multiple singly labeled probes. Nat Meth. 5:877–879.

Rideout EJ, Dornan AJ, Neville MC, Eadie S, Goodwin SF. 2010. Control of sexual differentiation and
behavior by the *doublesex* gene in *Drosophila melanogaster*. Nat Neurosci. 13:458–466.

Shah S, Lubeck E, Schwarzkopf M, He TF. 2016. Single-molecule RNA detection at depth via
hybridization chain reaction and tissue hydrogel embedding and clearing. Development.
143:2862-2867.

Signor SA, Li Y, Rebeiz M, Kopp A. 2016. Genetic convergence in the evolution of male-specific color
patterns in *Drosophila*. Curr Biol. 26: 2423-2433.

Signor SA, New F, Nuzhdin S. *in review*. An abundance of high frequency variance uncovered in a
large panel of *Drosophila simulans*. Genome Biol Evol.

Struhl SGAG. 1999. Progression of the morphogenetic furrow in the *Drosophila* eye: the roles of
Hedgehog, *Decapentaplegic* and the Raf pathway. Development. 126:5795-5808.

Strutt DI, Mlodzik M. 1997. *Hedgehog* is an indirect regulator of morphogenetic furrow progression in
the *Drosophila* eye disc. Development 124:3233–3240.

Taylor MB, Ehrenreich IM. 2015. Transcriptional derepression uncovers cryptic higher-order genetic
interactions. PLoS Genet. 11:e1005606.

Tsachaki M, Sprecher SG. 2011. Genetic and developmental mechanisms underlying the formation of
the *Drosophila* compound eye. Dev Dyn. 241:40–56.

Woodruff GC, Eke O, Baird SE, Felix M-A, Haag ES. 2010. Insights into species divergence and the
evolution of hermaphroditism from fertile interspecies hybrids of *Caenorhabditis* nematodes.
Genetics 186:997–1012.

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