

1 ORIGINAL ARTICLE

2 **Running head:** COMPARING COLOURS

3 **Comparing colours using visual models**

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11 **Keywords:** vision, dimorphism, polymorphism, mimicry, crypsis, multivariate
12 statistics

¹³ **Lay Summary**

¹⁴ An outstanding challenge for the study of colour traits is how best to use "colour
¹⁵ spaces" to represent their visual perception, particularly when asking questions
¹⁶ of colour-difference (e.g. the (dis)similarity of males and females, mimics and
¹⁷ models, or sister species, to a given viewer). We use simulations to show that
¹⁸ existing methods fail to statistically and biologically estimate the separation of
¹⁹ groups in colour space, and we suggest a flexible, robust, alternative that avoids
²⁰ those pitfalls.

21 **Abstract**

22 Colour in nature presents a striking dimension of variation, though understanding
23 its function and evolution largely depends on our ability to capture the perspec-
24 tive of relevant viewers. This goal has been radically advanced by the development
25 and widespread adoption of colour spaces, which allow for the viewer-subjective
26 estimation of colour appearance. Most studies of colour in camouflage, aposema-
27 tism, sexual selection, and other signalling contexts draw on these models, with
28 the shared analytical objective of estimating how similar (or dissimilar) colour
29 samples are to a given viewer. We summarise popular approaches for estimating
30 the separation of samples in colour space, and use a simulation-based approach
31 to test their efficacy with common data structures. We show that these meth-
32 ods largely fail to estimate the separation of colour samples by neglecting (i) the
33 statistical distribution and within-group variation of the data, and/or (ii) the dis-
34 criminability of groups relative to the observer's visual capabilities. Instead, we
35 formalize the two questions that must be answered to establish both the statistical
36 presence and theoretical magnitude of colour differences, and propose a two-step,
37 permutation-based approach that achieves this goal. Unlike previous methods,
38 our suggested approach accounts for the multidimensional nature of visual model
39 data, and is robust against common colour-data features such as heterogeneity
40 and outliers. We demonstrate the pitfalls of current methods and the flexibility of
41 our suggested framework using an example from the literature, with recommen-
42 dations for future inquiry.

43 Introduction

44 The study of colour in nature has driven fundamental advances in ecology and
45 evolutionary biology (Cuthill *et al.*, 2017). Colour is a subjective experience, how-
46 ever, so substantial effort has been dedicated to measuring and representing colours
47 “objectively” (Garcia *et al.*, 2014; Johnsen, 2016) through visual models that con-
48 sider the perspective of ecologically relevant viewers (Kemp *et al.*, 2015; Renoult
49 *et al.*, 2017). These models have significantly advanced the study of colour traits
50 by allowing researchers to account for the factors influencing the generation and
51 reception of visual information, such as the structure of signals and viewing back-
52 grounds, the properties of veiling and incident light, and the attributes of visual
53 systems (Chittka, 1992; Endler & Mielke, 2005; Kelber *et al.*, 2003; Vorobyev &
54 Osorio, 1998).

55 Several forms of visual models are currently used, which vary in their assump-
56 tions about the nature of visual processing (Chittka, 1992; Endler & Mielke, 2005;
57 Vorobyev & Osorio, 1998). These models function by delimiting a colour space
58 informed by the number and sensitivity of photoreceptors in an animal’s retina
59 (Renoult *et al.*, 2017). Individual colours are then represented in this space as
60 points, with their location determined by the differential stimulation of the view-
61 ers’ receptors.

62 This colour space representation is convenient for several reasons. It offers an
63 intuitive way of analysing phenotypes that we cannot measure directly: we can
64 estimate how animals with different visual systems “see” different colours by rep-
65 resenting them in a Cartesian coordinate system, producing a receiver-dependent
66 morphospace (Kelber *et al.*, 2003; Renoult *et al.*, 2017). Further, it allows estimating
67 how similar or dissimilar colours are *to a given observer*, by measuring the distance
68 between colour points in its colour space (Endler & Mielke, 2005; Vorobyev *et al.*,
69 1998; Vorobyev & Osorio, 1998). Crucially, we can test and refine these mod-
70 els using psychophysical data (e.g. Dyer & Neumeyer, 2005; Garcia *et al.*, 2017;

71 Maier, 1992; Vorobyev *et al.*, 2001), to estimate the magnitude of colour-differences
72 and ultimately predict whether an observer could effectively discriminate pairs
73 of colours (Chittka, 1992; Vorobyev & Osorio, 1998). This final point is critical to
74 many tests of ecological and evolutionary hypotheses, such as the efficacy of cam-
75 ouflage (Pessoa *et al.*, 2014; Troscianko *et al.*, 2016), the presence of polymorphism
76 or dichromatism (Schultz & Fincke, 2013; Whiting *et al.*, 2015), the accuracy of
77 mimicry (O’Hanlon *et al.*, 2014; White *et al.*, 2017), the extent of signal variability
78 among populations or species (Delhey & Peters, 2008; Rheindt *et al.*, 2014), or the
79 effect of experimental manipulations (Barry *et al.*, 2015; White & Kemp, 2017). At
80 the heart of these inquiries lies the same question: *how different are these colours to*
81 *the animal viewing them?*

82 *Challenges in estimating the discriminability of colour samples*

83 The receptor noise-limited model of Vorobyev & Osorio (1998) has proven partic-
84 ularly useful for addressing questions of discriminability and colour-difference.
85 The model is focused on receptor-level processes, and assumes that chromatic
86 and achromatic channels operate independently (which does not necessarily hold
87 beyond the receptor level in some species, such as humans; Nathans, 1999), that
88 colour is coded by $n - 1$ unspecified opponent mechanisms (where n is the number
89 of receptor channels), and that the limits to colour discrimination are set by noise
90 arising in receptors (Vorobyev *et al.*, 1998; Vorobyev & Osorio, 1998). This noise
91 is dependent on the receptor type and abundance on the retina which, along with
92 Weber’s law ($k = \Delta I / I$) more generally, ultimately establishes the unit of Just No-
93 ticeable Differences (JND Vorobyev *et al.*, 2001). Distances calculated in this man-
94 ner correspond to the Mahalanobis Distance D_M , and represent distances between
95 points standardized by the Weber fraction; i.e. $\frac{\text{signal}}{\text{noise}}$ (Clark *et al.*, 2017). It follows
96 that values lower than 1 JND ($\frac{\text{signal}}{\text{noise}} < 1$) are predicted to be indistinguishable,
97 while values greatly above this threshold are likely distinct. This provides a use-
98 ful standard for estimating the similarity of groups of points in colour space: the

99 greater the distance between colours, the less alike they are. If differences are,
100 on average, above an established threshold, then we can consider the groups dif-
101 ferent: sexes dichromatic, mimetism imperfect, crypsis ineffective. This offers a
102 clear link between variation and classification within a sensory framework, and
103 has been widely used for this purpose (Barry *et al.*, 2015; Delhey & Peters, 2008;
104 O’Hanlon *et al.*, 2014; Schultz & Fincke, 2013; White *et al.*, 2017; White & Kemp,
105 2017).

106 To adequately compare samples of colours, however, it is necessary to deter-
107 mine if the average distance between them is both statistically and biologically
108 meaningful (i.e. above-threshold; Endler & Mielke, 2005). Commonly, an “average
109 colour” for each group is derived by taking a mean reflectance spectrum or by
110 averaging their position in colour space. In either case, the colour distance be-
111 tween groups is then calculated from these mean quantum catches per-receptor
112 per-group — their centroids in multivariate space (Fig. 1, bold arrow). However,
113 the centroid obtained from arithmetic means of receptor coordinates is not an
114 appropriate measure of location for this purpose, since colour distances are per-
115 ceived in a ratio scale (Cardoso & Gomes, 2015). Instead, the geometric mean must
116 be used. Further, since the result is a single value representing the multivariate
117 distance between group means, there is no associated measure of uncertainty or
118 precision that would allow for the statistical testing of differences between samples
119 (e.g. Avilés *et al.*, 2011; Burns & Shultz, 2012; Maia *et al.*, 2016).

120 An alternative approach calculates the pairwise distances between all points
121 in group A and group B, then averages these distances to obtain a mean distance
122 between groups (Fig. 1, thin arrows; e.g. Barry *et al.*, 2015; Dearborn *et al.*, 2012).
123 In cluster analyses, this is called the “average linkage” between groups (Hair *et al.*,
124 1998). This is an appealing method, providing measures of variation among dis-
125 tances, and thus a t-test or equivalent can be used to test if differences are greater
126 than a given threshold. The average linkage, however, is also inadequate because
127 it conflates within- and among-group variation. This is because Euclidean dis-

128 tances (and by extension JND's) are *translation-invariant*: they ignore the position
129 of points in colour space and the direction of the distance vectors, reflecting only
130 the magnitude of differences between two points. Therefore, the average linkage
131 reduces to a measure of spread, and will scale with both within- and between-
132 group distances (Fig. 1, insert).

133 As these issues show, hypotheses of discriminability and colour-difference have
134 primarily focused on testing whether the difference between samples is above a
135 theoretical threshold. However, the convenience of such thresholds belies fact that
136 simply comparing means between groups is not sufficient to infer, statistically,
137 whether samples are different. To answer if two groups are different, one must
138 compare the variation between- and within-groups. This is particularly problem-
139 atic in the case of colours that function as signals in social interactions (e.g. [Kemp](#)
140 [& Rutowski, 2011](#)). For a trait to function in this context, the observer must be
141 able to tell signals of 'low' and 'high' quality apart. This means that, by defi-
142 nition, *most pairs of individuals should be readily distinguishable*. The trait must be
143 highly variable and colour distances should be above the threshold of discrimina-
144 tion ([Delhey et al., 2017](#)), otherwise no information can be extracted by an observer
145 comparing phenotypes.

146 Consider a hypothetical species that uses colour in mate choice, but is not sex-
147 ually dichromatic (Fig. 1). In this species colour is highly variable and, on average,
148 pairs of individuals are discriminable, but there is no *consistent* male-female dif-
149 ference. Therefore, if a researcher sampled this species and calculated the average
150 distance between all pairs of individuals, regardless of sex, these should be largely
151 greater than 1 JND. However, if they took separate samples of males and females,
152 then all pairwise distances (the average linkage) between sexes will be also greater
153 than 1 JND, despite them being sampled from the same (statistical) population.

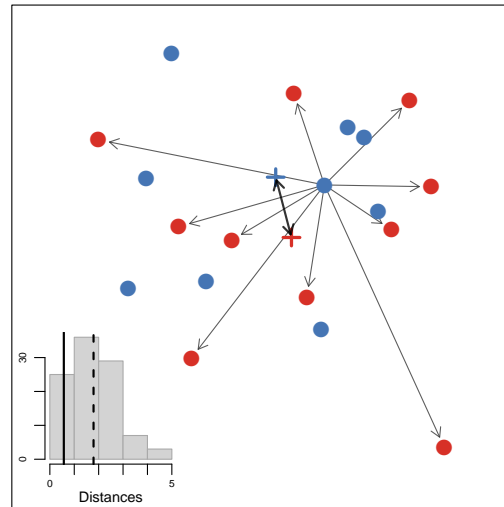


Figure 1: The link distance (i.e. average pairwise distance between groups) conflates within- and among-group variation. Here, two samples were drawn from the same simulated distribution. Thin arrows represent distances between a random point in the first sample (blue) and all points from the second sample (red), all of which are greater than the distance between the geometric means of the two samples (“x”, bold arrows). Inset shows the histogram of pairwise distances among groups, and how their average (dashed line) is greater than the mean distance (bold line).

154 *The limitations of current methods for comparing colour space distributions*

155 Several methods have been proposed to avoid the aforementioned issues by ac-
156 counting for the relative distributions of samples in colour space. Eaton (2005),
157 for example, noted that within-group variation influenced the conclusions on the
158 extent of avian dichromatism, and thus tested for intersexual differences in pho-
159 ton catches separately for each receptor. However, this ignores the multivariate
160 nature of visual model data by failing to account for multiple comparisons and
161 correlations among receptor catches (which are critical, since any n -receptor vi-
162 sual system can be represented in $n - 1$ dimensions; Kelber *et al.*, 2003).

163 An alternative, multivariate metric suggested by Stoddard & Prum (2008) is
164 the volume overlap. In this approach, the volume occupied by a sample of colours
165 is estimated from its enveloping convex hull, and separation between samples is

166 inferred from their overlap. [Stoddard & Stevens \(2011\)](#) used this metric to show
167 that a greater overlap in colour volume between cuckoo and host eggs is associ-
168 ated with lower rejection of parasitic eggs. This approach is appealing because it
169 considers the distribution of colour points in multivariate space, though there are
170 limits to its interpretation: (i) there is a lower bound to group separation (i.e. if
171 samples do not overlap, there is no distinction between cases where samples are
172 near or far apart) and (ii) it is unclear how variation in volume overlap should be
173 interpreted biologically (e.g. how biologically relevant is the difference between
174 20% or 40% overlap?). It is also particularly sensitive to outliers, because the vol-
175 ume defined by a convex hull does not lend itself to a probabilistic interpretation,
176 leading to the often unacknowledged assumption that the sampled data reflects
177 the true boundaries of the population (however, “loose wrap” hypervolumetric
178 methods exist; to our knowledge, these have not been applied to colour stud-
179 ies; [Blonder *et al.*, 2017](#)). Finally, in its original implementation this method does
180 not consider receptor noise or discrimination thresholds (but incorporating this is
181 straightforward; see below).

182 The most robust attempt at comparing distributions of colours was proposed
183 by [Endler & Mielke \(2005\)](#), who devised a non-parametric rank distance-based
184 approach based on the least sum of Euclidean distances, compared through multi-
185 response permutation procedures (LSED-MRPP). This multivariate approach is
186 powerful because it calculates an effect size based on the relationship of between-
187 and within-group distances. However, this single statistic captures differences be-
188 tween samples not only in their means, but also in their dispersion and correlation
189 structure (i.e. shape; [Endler & Mielke, 2005](#)). Like other distance-based methods,
190 it is sensitive to confounding heterogeneity among samples when testing for dif-
191 ferences in *location* ([Anderson & Walsh, 2013](#); [Warton *et al.*, 2012](#)). Despite its
192 considerable strengths, this method has seen little adoption over the last decade,
193 largely due to limitations in implementation and accessibility.

194 The shortcomings of these methods reflect the fundamental fact that the ques-

195 tion of discriminability actually represents a test of two hypotheses that are seldom
196 formally distinguished: (i) that the focal samples are statistically distinct, and (ii)
197 that the magnitude of their difference is greater than a psychophysical threshold
198 of detection. Most approaches will test one, but not both, of these hypotheses
199 through their respective nulls, and often with no estimate of variation or uncer-
200 tainty in estimates. We explore these issues using a simulation-based approach
201 by testing the efficacy of popular methods in detecting the separation of groups
202 in colour space. We then propose a flexible solution that avoids these problems,
203 demonstrating its utility using an example from the literature.

204 **Methods**

205 **Simulation procedures**

206 To compare methods for detecting group separation in colour space, we simu-
207 lated data analogous to that obtained from applying an avian visual model to
208 spectral reflectance data. Birds are tetrachromatic (Hart, 2001), and colours will
209 thus be represented by the quantum catches of its four photoreceptors (though
210 the procedure followed here can be applied to visual systems with any number
211 of receptors). For each replicate, we simulated two samples defined by four vari-
212 ables (*USML* photoreceptors) taken from log-normal distributions (since quantum
213 catches are non-negative and noise-corrected distances follow a ratio scale, as de-
214 fined by the Weber fraction, described above). We generated samples following
215 two different scenarios: first, we simulated varying degrees of separation (i.e. ef-
216 fect sizes) to evaluate the power and Type I error rates of the approaches tested.
217 Second, we simulated threshold conditions to evaluate the performance of differ-
218 ent approaches in correctly classifying whether samples are above-threshold.

219 For the first set of simulations (power and error-rates) we simulated the quantal
220 catch of each photoreceptor i for the first sample (group A) by drawing from a log-
221 normal distribution with mean μ_{iA} seeded from a uniform distribution $\mathcal{U}(0, 10)$,

222 and standard deviation proportional to the mean: $\sigma_i = a_i \mu_{iA}$, with $a_i \sim \mathcal{U}(0, 0.5)$
223 (note that, for these simulations, μ and σ refer to the mean and standard deviation
224 of the random variable itself, not in log scale). To generate two samples with
225 varying degrees of separation proportional to the within-group variance, we used
226 a multivariate effect size S obtained by calculating a constant $d_i = \frac{S}{\sqrt{n}} \bar{\sigma}_i$, where n
227 is the number of photoreceptors (in this case, 4) and $\bar{\sigma}_i$ is the standard deviation
228 of the sample. We then drew a second sample (group B) defined by $\mu_{iB} = \mu_{iA} + d_i$
229 and σ_i . Thus, our simulations effectively produced two samples with Mahalanobis
230 Distance $D_M \sim S$ (calculated as the distance between centroids of the two groups
231 weighted by their pooled variance-covariance matrix). We simulated data for $S =$
232 $\{0, 0.1, 0.25, 0.5, 0.75, 1.0, 1.5, 2, 2.5, 3.0\}$ (Fig. 2), replicated 200 times for sample
233 sizes $N = \{10, 20, 50, 100\}$ each.

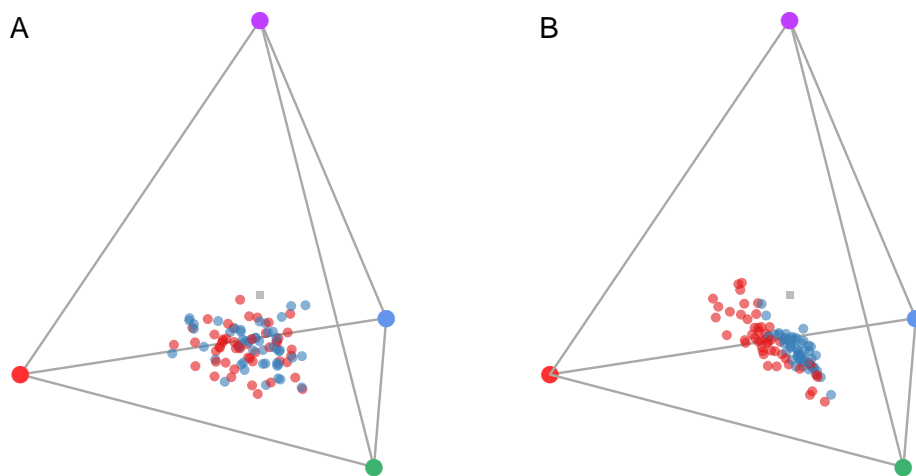


Figure 2: Example simulated data for the two groups (red, blue) in a tetrahedral colourspace. Shown here are data with sample size $N = 50$ and effect size (A) $S = 0$ and (B) $S = 3$.

234 For the second set of simulations (threshold conditions across a range of within-
235 sample variation), we followed a similar procedure. Group A was sampled from a
236 log-normal distribution with $\mu_{iA} \sim \mathcal{U}(0, 10)$, while σ_i was taken from an exponen-

237 tial distribution $\sigma_i \sim \text{Exp}(\lambda = 1)$. To obtain a second sample, group B, that was
238 separated from group A with an average approximate distance of ~ 1 JND given a
239 Weber fraction of 0.1 for the long-wavelength photoreceptor (Vorobyev *et al.*, 1998),
240 we drew from log-normal distributions with $\mu_{iB} = d_i \mu_{iA}$, where $d_i \sim \mathcal{U}(0.88, 1.12)$,
241 resulting in an average distance between geometric means (hereafter, “mean dis-
242 tance”) of 1.11 (95% quantiles: 0.35 – 2.77 JND) and within-group average pairwise
243 distance of 4.46 (95% quantiles: 1.03 – 11.10 JND) after 1000 replicates.

244 After the two groups were simulated, we used the R package *pavo* (Maia *et al.*,
245 2013) to calculate colour distances using relative receptor densities of $\{U, S, M, L\} =$
246 $\{1, 2, 2, 4\}$ and Weber fraction for $L = 0.1$. We calculated the within-group average
247 pairwise distance, as well as the distance between sample geometric means.

248 We then used four procedures to test for differences between groups. First,
249 we used a distance-based PERMANOVA (hereafter “distance PERMANOVA”) us-
250 ing the *adonis* function from the R package *vegan* (Oksanen *et al.*, 2007). This
251 non-parametric approach uses distances to calculate a pseudo-F statistic, simulat-
252 ing a null distribution by randomizing distances between observations (Anderson,
253 2005). We recorded if the analysis was significant ($\alpha = 0.05$) using 999 permuta-
254 tions for the null, as well as the R^2 as an effect size estimate. Second, we obtained
255 XYZ Cartesian coordinates based on “perceptually-scaled” (i.e. noise-corrected)
256 distances (Pike, 2012; functionally and mathematically equivalent to the receptor-
257 noise limited space of de Ibarra *et al.*, 2001) and applied a MANOVA test on these
258 coordinates (hereafter “Cartesian MANOVA”). For simplicity, we used a sum of
259 squares and cross-products matrix approach and calculated Pillai’s trace and its
260 associated P-value (but see discussion for extensions of this approach). Third, we
261 calculated the volume overlap between the two samples (relative to their combined
262 volumes) in a tetrahedral colour space defined by the receptors’ relative quantum
263 catches (thus disregarding receptor noise; Stoddard & Prum, 2008). Finally, we
264 calculated the volume overlap for the XYZ Cartesian coordinates based on noise-
265 corrected distances, generating a colour volume overlap that accounts for receptor

266 noise.

267 **Simulation results**

268 **Power and error rates**

269 Both the distance PERMANOVA and the Cartesian MANOVA showed appropriate
270 Type-I error rates, with about 5% of our simulations producing significant results
271 when $S = 0$, even for small sample sizes (Fig. 3). As expected, the power to detect
272 small effects steadily increased as a function of sample size, with the distance
273 PERMANOVA being overall more conservative than the Cartesian MANOVA (Fig.
274 3,4).

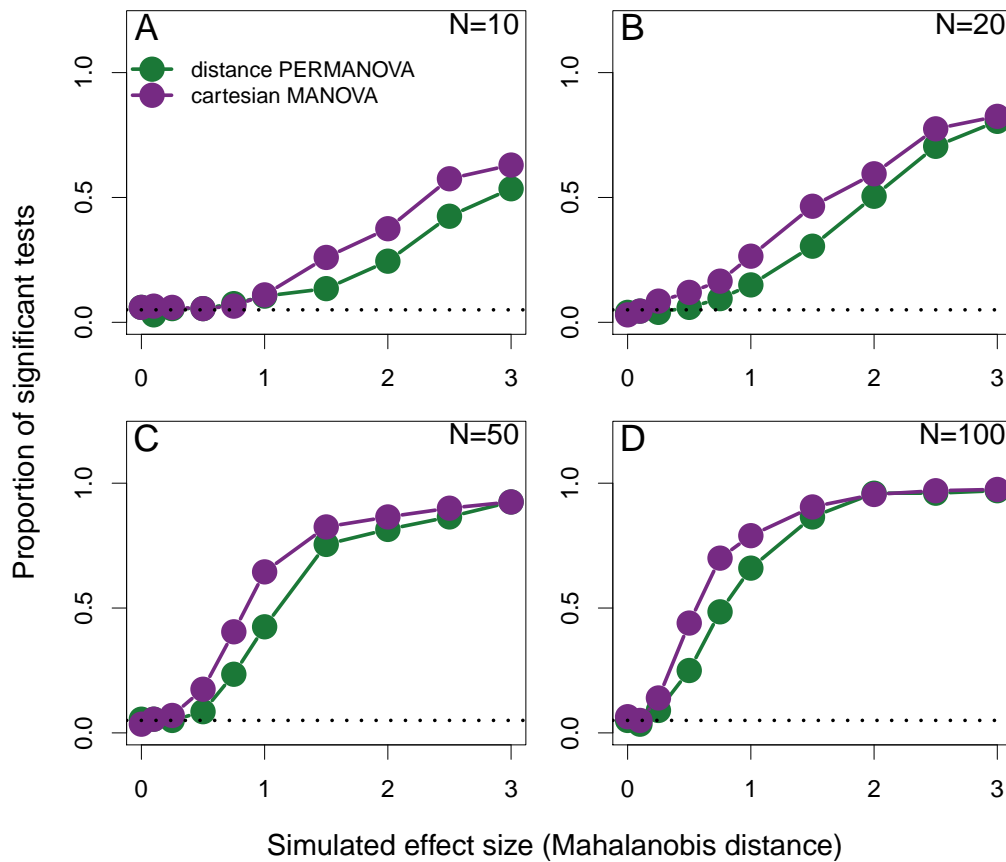


Figure 3: Power and Type I error rate of the distance PERMANOVA (green) and Cartesian MANOVA (purple). Panels show the proportion of simulations yielding significant results for each approach under different sample and effect sizes.

275 The two approaches showed some disagreement, with between 10 – 15% of
276 the simulations significant only in one of the two tests (Fig. 4). This disagreement
277 was not random, with the Cartesian MANOVA being more likely to be significant
278 when the distance PERMANOVA was not than vice-versa (Fig. 4a), at an approx-
279 imately constant rate across sample sizes, and disagreement being concentrated
280 at smaller effect sizes with increasing sample sizes (Fig. 4b).

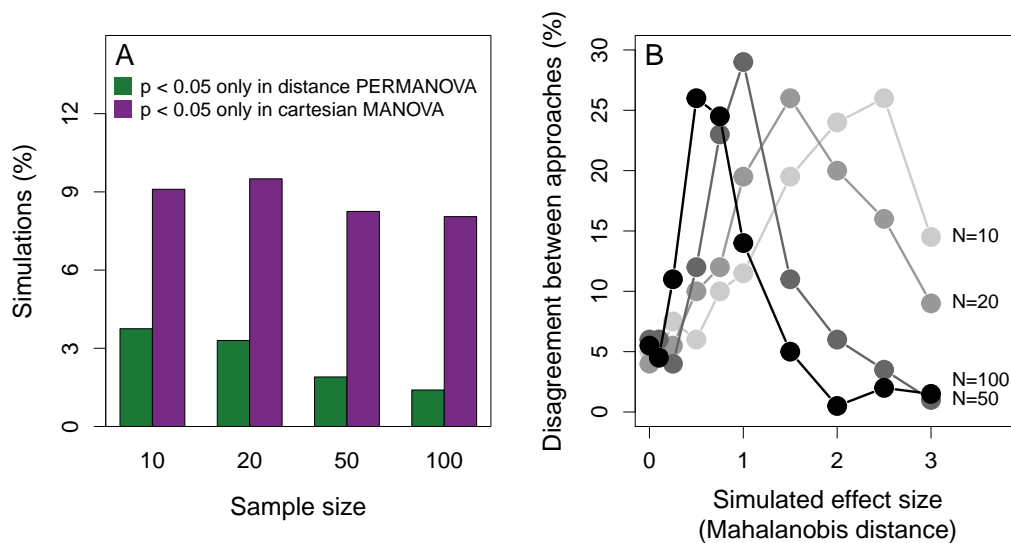


Figure 4: The disagreement between multivariate statistical approaches when testing for separation between samples in colour space in relation to sample size (A) and effect size (B).

281 Focusing on $N = 50$ simulations, our results show that mean distance was
282 positively associated with the effect size, and the threshold of significance using
283 the distance PERMANOVA fell approximately at the $1JND$ mark (Fig. 5A; equiv-
284 alent results are observed with the Cartesian MANOVA, not shown). Still, even
285 around that threshold, significance is variable, showing that large within-group
286 variation can lead to non-significant differences between groups despite among-
287 group distances being above the theoretical perceptual threshold. Volume overlap
288 also showed a (negative) association with effect size, but no specific threshold
289 for significance is observed (e.g. both significant and non-significant results are
290 observed for 20 – 60% overlap; Figure 5B).

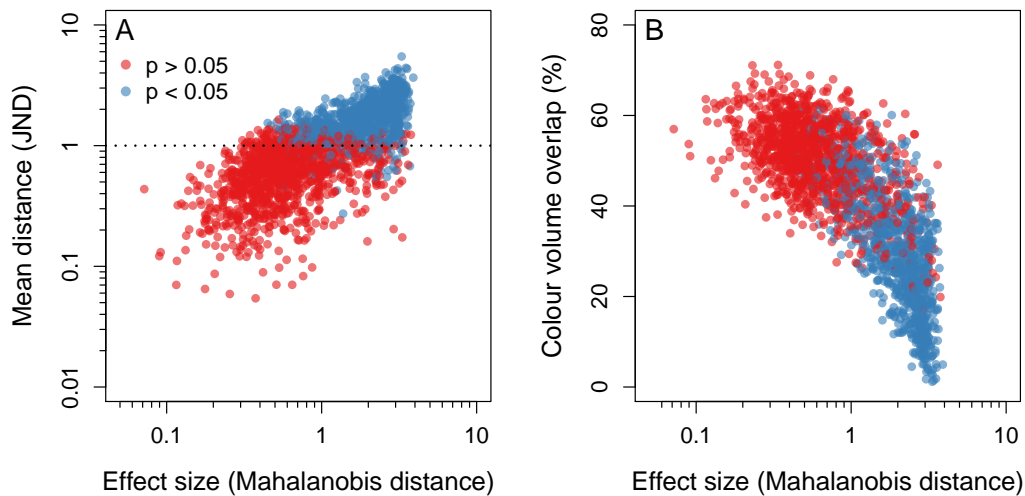


Figure 5: The association between effect size and (A) mean distance and (B) colour volume overlap. Significant distance PERMANOVA results are in blue, whereas non-significant results are in red. Dotted line indicates the threshold of 1 JND.

291 **Threshold scenarios**

292 Since results from the distance PERMANOVA and the Cartesian MANOVA were
293 comparable, we focus on the former due to the convenience of the R^2 statistic
294 describing among-group separation (but see Discussion for comments on the use
295 of these approaches). Simulations produced a wide range of outcomes, with non-
296 significant and significant tests both above and below the theoretical threshold of 1
297 JND (Fig. 6). In contrast with the power simulations above (Fig. 5), the significance
298 threshold did not match the theoretical perceptual threshold. As in the hypothet-
299 ical example from the introduction, 20.2% of the simulated cases were statistically
300 indistinguishable despite having mean above-threshold distances (Fig. 6, dark
301 red). Likewise, 15.1% of the simulations produced samples that were statistically
302 different, but where this difference was below threshold and was therefore likely
303 undetectable to its observer (Fig. 6, dark blue points). These results highlight the
304 importance of considering both statistical separation and theoretical perceptual
305 thresholds when testing the hypothesis that samples are discriminable.

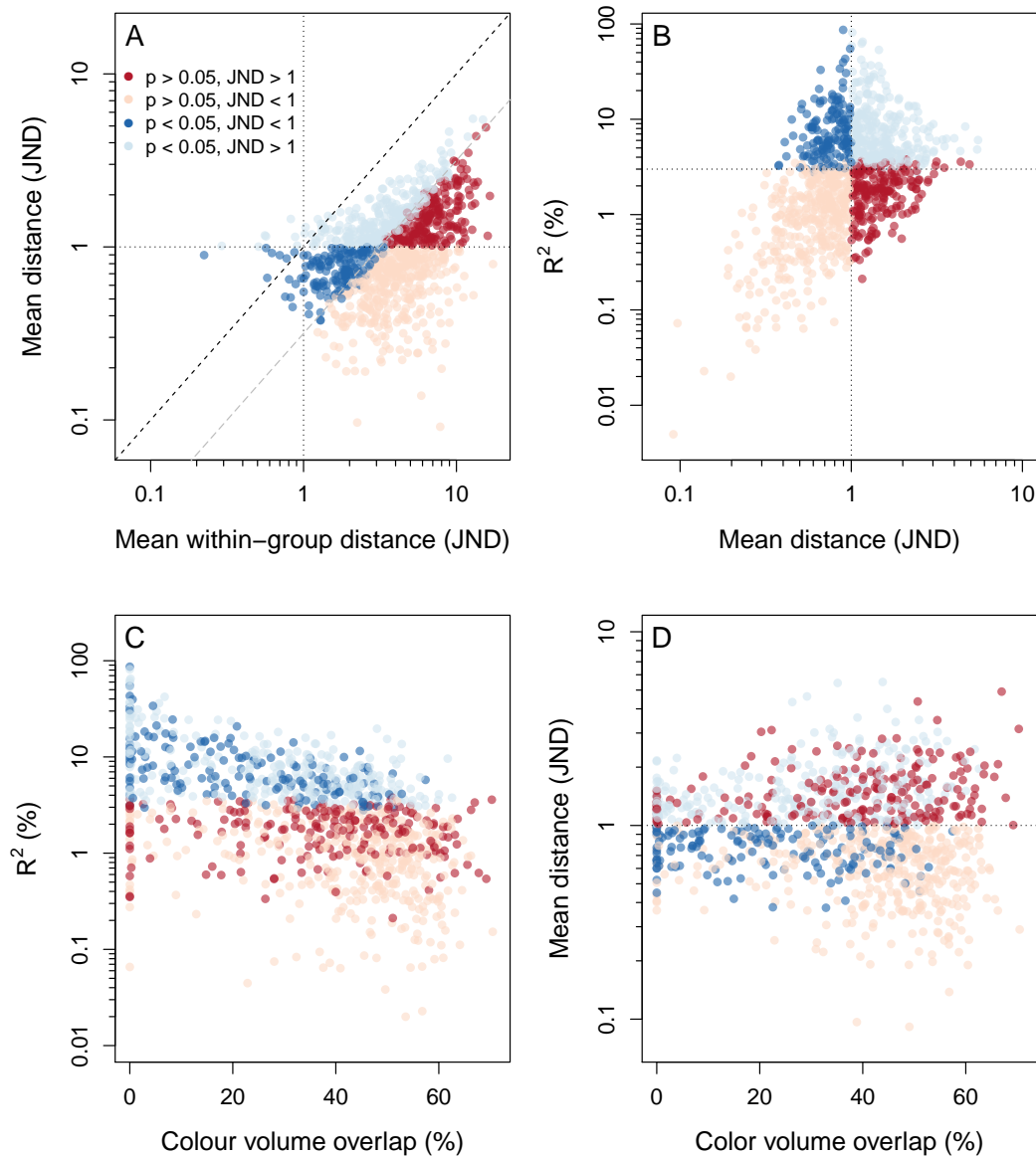


Figure 6: Results from threshold simulation. Red and blue denote non-significant and significant PERMANOVA tests, respectively, and light colours denote when that approach would yield the same inference as comparing mean distances to a threshold of 1JND. Thus, dark blue points indicate a significant statistical test that does not reach the threshold of discriminability of 1 JND, whereas dark red points indicate a non-significant statistical test that nonetheless has a mean distance greater than 1 JND.

306 Figure 6A shows that, intuitively, tests were significant when within-group
307 differences were small relative to among-group differences. However, nearly all
308 simulations — including most significant results — fell below the 1:1 line when

309 using the average link distance (i.e. the average pairwise distance) to describe
310 intragroup variation. Significant results are obtained when the mean difference is
311 up to 0.5 JND smaller than the within-group average link distance (Fig. 6A, grey
312 line intercept). Similarly, we can see that significant results can be obtained for
313 fairly low levels of among-group separation, with R^2 as small as 3 or 4% (Fig. 6B,
314 horizontal line at 3%).

315 Though there is a negative association between R^2 and volume overlap (Fig.
316 6C), results show low overall consistency between approaches: for any given value
317 of volume overlap, all possible outcomes of significance/threshold occur — even
318 when the overlap between samples is zero (Fig. 6C). In other words, even complete
319 separation in colour volumes can result in non-significant, below-threshold cases,
320 since samples can be contiguous without overlapping in noise-corrected colour
321 space. Likewise, samples can have high overlap but be entirely distinguishable
322 statistically and perceptually. Further, there is no association between volume
323 overlap and mean distance between groups (Fig. 6D). These results were unaltered
324 by considering receptor noise in the volume overlap calculation, since these are
325 still strongly and positively correlated with their non-noise-corrected counterparts
326 (Electronic Supplementary Material).

327 **A two-step approach to estimate statistical and perceptual separation**

328 As described previously, questions of discriminability and colour-difference re-
329 quire testing two distinct hypotheses: if samples are (i) statistically and (ii) ‘percep-
330 tually’ distinct. We therefore propose a two-step answer to such questions, which
331 explicitly formalizes these hypotheses. For the first question — are the samples
332 *statistically separate* in colour space? — we show that both a PERMANOVA us-
333 ing noise-corrected colour distances (Anderson, 2005; Cornuault *et al.*, 2015), and
334 a MANOVA using noise-calibrated Cartesian coordinates (de Ibarra *et al.*, 2001;
335 Delhey *et al.*, 2015; Pike, 2012) are well suited. Both exclude achromatic variation
336 and properly account for the multivariate nature of the data. There is also mini-

337 mal discrepancy between the two (Fig. 3,4), so the decision between them may be
338 informed by convenience and the structure of the data at hand.

339 Once the separation of samples is established statistically, a second question
340 must be answered: is this separation predicted to be *perceptually discriminable*? The
341 statistics calculated above cannot answer this, since effect sizes account for both
342 among- and within-group variance. We therefore suggest this be tested indepen-
343 dently, by estimating the distance in colour space between group geometric means
344 rather than through the average pairwise distance or volume-overlap based met-
345 rics, which fail to accurately estimate group separation (Figs. 1,6). One limitation
346 to this statistic is the lack of any measure of uncertainty. To circumvent that, we
347 suggest a bootstrap procedure in which new samples are produced through re-
348 sampling (with replacement) of individuals of each group, from which geometric
349 means and their distance are calculated. Repeating this procedure generates a dis-
350 tribution of mean distances, from which a confidence interval can be estimated. If
351 the groups being compared are statistically different and this bootstrapped con-
352 fidence interval does not include the theoretical threshold of adequate biological
353 significance, one can conclude that the samples being compared are distinct and
354 likely discriminable.

355 **Empirical example: Sexual dichromatism in the leaf-nosed** 356 **lizard *Ceratophora tennentii***

357 Visually signalling animals often use distinct body parts for different purposes,
358 such as social signalling to mates or warning predators (Barry *et al.*, 2015; Grether
359 *et al.*, 2004; Johnstone, 1995). The nature of intraspecific variation in colour can
360 thus inform their putative function, since selection may act differentially on signals
361 used in different contexts. For example, traits subject to strong sexual selection in
362 one of the sexes are often dimorphic, with one sex (typically males) expressing a
363 conspicuous colour pattern that is reduced or absent in the other (Bell & Zamudio,

364 2012; Kemp & Rutowski, 2011).

365 Dragon lizards (Agamidae) are known for variable colouration used in both
366 social and anti-predator contexts (Johnston *et al.*, 2013; Somaweera & Somaweera,
367 2009). The leaf-nosed lizard *Ceratophora tennentii* has multiple discrete colour
368 patches, with apparent sex differences between body parts (Fig. 7). Here we
369 draw on the data of Whiting *et al.* (2015), who recorded the spectral reflectance of
370 29 male and 27 female *C. tennentii* from four body regions (throat, labials, mouth-
371 roof, and tongue). We used a tetrachromatic model of agamid vision to test for
372 dichromatism among body regions from the perspective of conspecifics.

373 Following standard calculations for the log-linear receptor-noise model, we
374 used the spectral sensitivity of *Ctenophorus ornatus* ($\lambda_{max} = 360, 440, 493, 571$ nm)
375 as modelled according to a vitamin A1 template (Barbour *et al.*, 2002; Govardovskii
376 *et al.*, 2000). We assumed a relative photoreceptor abundance of 1 : 1 : 3.5 : 6,
377 and a coefficient of variation of noise yielding a Weber fraction of 0.1 for the
378 long-wavelength cone (Fleishman *et al.*, 2011; Loew *et al.*, 2002). We tested each
379 body region separately using PERMANOVA. As above, we used the R package
380 pavo for visual modelling, and the adonis function in the R package vegan for
381 PERMANOVAs.

382 We found a statistical difference between male and female throats (PERMANOVA:
383 $F_{1,58} = 14.84$, $P < 0.01$) and labials ($F_{1,57} = 13.96$, $P < 0.01$; Fig. 7A,B), but not for
384 tongues ($F_{1,58} = 1.63$, $P = 0.22$) or mouth-roofs ($F_{1,55} = 0.52$, $P = 0.50$; Fig. 7C,D).
385 However, bootstraps of group separation suggest that intersexual differences in
386 labial colour are likely imperceptible to conspecifics (Fig. 7E; though like all such
387 predictions this requires behavioural validation). Our results therefore suggest
388 the absence of dichromatism in all but throat colour from the lizard perspective,
389 despite statistical significance for the labial region. These results thus do not im-
390 plicate sexual selection as a strong driver of intersexual colour differences in these
391 few body regions of *C. ornatus*.

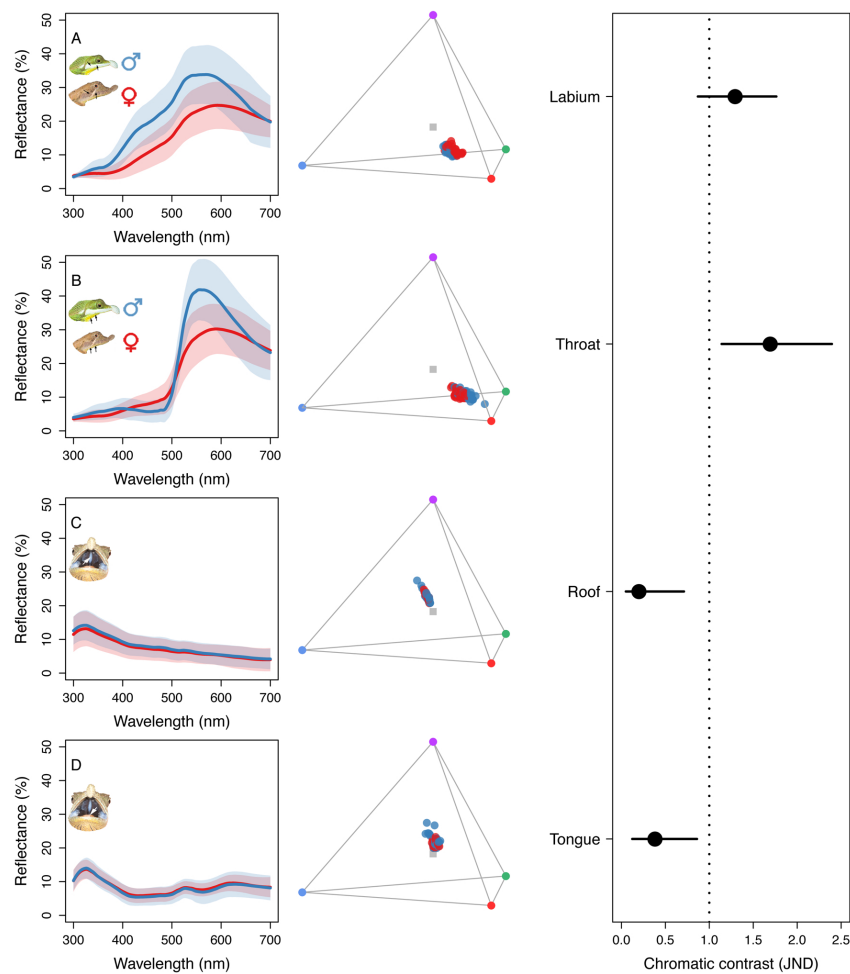


Figure 7: The mean (\pm SD) spectral reflectance of female (red) and male (black) (A) labial, (B) throat, (C) mouth-roof, and (D) tongue (left panels), and their colourspace distribution according in a tetrachromatic model of agamid vision (middle panels). Inset images indicate approximate sampling regions. The bootstrapped 95 % C.I.'s for mean distances between groups in colour space (right panels). Partly reproduced, with permission, from [Whiting *et al.* 2015](#).

392 Discussion

393 Visual models offer a useful tool for quantifying the subjective perception of
 394 colour, which — as the ultimate canvas for colour-signal evolution — can offer
 395 valuable insight into a breadth of biological phenomena. It is therefore essential
 396 that statistical considerations of biological hypotheses take into account both nat-
 397 ural variation in the compared samples as well as the limits to observer perception
 398 (as ultimately informed by behavioural and physiological data; [Kemp *et al.*, 2015](#)).

399 Here, we show that most methods typically fail to consider these aspects, and pro-
400 pose a flexible alternative that explicitly addresses both.

401 The use of models that do not explicitly consider discriminability, such as
402 the volume-overlap and segment-based analyses, is often justified on the basis of
403 simplifying and relaxing assumptions about colour perception, since intricate em-
404 pirical work is required to estimate model parameters (Kelber *et al.*, 2017; Olsson
405 *et al.*, 2015; Vorobyev & Osorio, 1998). However, we contend that, on the contrary,
406 some of these ‘simpler’ methods actually make very strong latent assumptions,
407 which are not supported by the empirical evidence. This includes the assumption
408 that all cones contribute equally to colour perception, that colour discrimination is
409 unequivocal (i.e. the magnitude of colour-difference does not affect discriminabil-
410 ity) and that colour differences follow an interval scale (as opposed to a ratio scale).
411 Thus, we suggest that considering detectability relative to a threshold is essential
412 for tests of discriminability. We emphasise, however, that this does not necessitate
413 the use of the receptor-noise model specifically. Although we have focused on this
414 popular approach here, particularly due to its utility for non-model organisms, a
415 breadth of available modelling tools are capable of offering similar, and in some
416 cases superior, insight (Kemp *et al.*, 2015; Price & Fialko, 2017; Renoult *et al.*, 2017).
417 The hexagon model of Chittka (1992), for example, has been extensively tested
418 and validated in honeybees, and may outperform the receptor-noise model when
419 suitably parameterised (Garcia *et al.*, 2017). It too offers a psychophysiologically-
420 informed measure of perceptual distance, as well as discrimination thresholds,
421 and so may be readily applied within our suggested framework. The two-step
422 approach we propose can be easily and directly extended to these models.

423 Our simulations show that both the distance PERMANOVA and Cartesian
424 MANOVA perform similarly well in statistically differentiating colours in percep-
425 tual space (Fig. 3). Studies have pointed out that distance-based methods perform
426 poorly when the experimental design is unbalanced or when there is heteroscedas-
427 ticity (though, among distance-based methods, PERMANOVA outperforms other

428 approaches; Anderson & Walsh, 2013; Warton *et al.*, 2012). It is important to note
429 that these are often common features of, and applicable to, colour data (Endler
430 & Mielke, 2005), and that these assumptions should be considered and verified.
431 However, this might still be the most robust option for high-dimensional visual
432 systems (e.g. Arikawa *et al.*, 1987; Cronin & Marshall, 1989), by reducing data
433 to a single metric of distance. Recently, Delhey *et al.* (2015) advocated a similar
434 MANOVA approach, by applying a Principal Component Analysis (PCA) to the
435 noise-corrected Cartesian coordinates prior to the test. However, if all the princi-
436 pal components are used in the MANOVA, results will be numerically identical to
437 directly using the XYZ coordinates (which is preferable, since it is often tempting
438 to discard PC axes of low variance, which could be problematic given that those
439 axes may be involved in group differentiation). While we have focused on tests
440 of differences in the location of colours in colour space, we recognise that other
441 characteristics — such as differences in dispersion and correlation structure, and
442 to identify the direction of variation among groups — might themselves be of
443 biological interest, for which a PCA approach may be particularly useful.

444 The MANOVA approach can be extended to multivariate generalizations of
445 generalized linear models by using the noise-corrected Cartesian coordinates as
446 response variables (Hadfield, 2010). These models can also relax the assumptions
447 of heteroscedasticity by estimating the variance-covariance of the responses (Had-
448 field, 2010), and can be extended to include various error and model structures,
449 such as hierarchical and phylogenetic models (Hadfield & Nakagawa, 2010). Still,
450 these approaches will only test for the statistical separation in colour space, so
451 estimating the magnitude of that separation is still necessary. The bootstrapped
452 distance provides an easy to interpret measure of uncertainty to the mean dis-
453 tance estimate. Under a Bayesian approach, the mean distance bootstrap can be
454 substituted by estimating credible intervals for the distance between perceptually-
455 corrected Cartesian centroids from the posterior distribution, though this will be
456 influenced by the priors adopted (Hadfield, 2010, see Electronic Supplementary

457 Material for an example analysis).

458 Irrespective of the method used, it is essential to parametrize the underly-
459 ing visual model appropriately (Garcia *et al.*, 2017; Olsson *et al.*, 2017). The Weber
460 fraction and receptor densities chosen will strongly affect noise-corrected distances
461 since they directly scale with the JND unit (Bitton *et al.*, 2017). Further, even under
462 adequate values of the Weber fraction it is important to realize that the unit JND
463 usually reflects psychophysical performance under extremely controlled condi-
464 tions (Kelber *et al.*, 2003; Olsson *et al.*, 2015), and that more conservative estimates
465 of 2-4+ JND may be more appropriate for ecological and evolutionary questions
466 (Osorio *et al.*, 2004; Schaefer *et al.*, 2007). Sensitivity analyses are also useful for
467 exploring the robustness of conclusions against parameter variation, particularly
468 in the case of non-model systems where such values are often assumed or drawn
469 from related species (Bitton *et al.*, 2017; Olsson *et al.*, 2017). More broadly, we af-
470 firm recent (and ongoing) calls for pragmatism when drawing inferences from
471 any such model (Marshall & Simmons, 2017; Olsson *et al.*, 2017; Vasas *et al.*, 2017).
472 Colour spaces are valuable tools, but ultimately demand ongoing feedback from
473 physiological and behavioural tests to improve our understanding of complex bi-
474 ological phenomena.

475 Our results show that insight into the biology of colour and its role in commu-
476 nication is best achieved by disentangling the implicit assumptions in questions
477 of discriminability. By bringing these assumptions to light, our two-step approach
478 offers a flexible procedure for examining the statistical presence and theoretical
479 magnitude of differences between colour samples. We expect it will bring exciting
480 new perspectives on the role of colour in intra- and interspecific interactions, and
481 provide an efficient analytical framework for the study of colour in nature.

482 **Implementation and data accessibility**

483 Analyses and simulations can be found in <https://github.com/rmaia/msdichromatism/>,
484 and the described methods are fully implemented in the R package `pavo` as of ver-
485 sion 1.3.1, available via CRAN. Key functions include `bootcoldist` which calcu-
486 lates the bootstrapped confidence intervals for mean distances, while `jnd2xyz`
487 converts chromatic distances in JNDs to noise-corrected Cartesian coordinates
488 . Multi-dimensional plotting options for noise-converted coordinates are also
489 available. Lizard colour data from Whiting *et al.* 2015 are available at [http:](http://dx.doi.org/10.6084/m9.figshare.1452908)
490 [//dx.doi.org/10.6084/m9.figshare.1452908](http://dx.doi.org/10.6084/m9.figshare.1452908).

491 **Acknowledgments**

492 We would like to thank Ruchira Somaweera for leaf-nosed lizard photographs,
493 and Dan Noble, John Endler and two anonymous reviewers for valuable discus-
494 sion and comments on earlier drafts of the manuscript. This work was partially
495 supported by a Junior Fellow award from the Simons Foundation to R.M., and an
496 Australian Research Council grant (DP140140107) to T.E.W.

497 **Author contributions**

498 RM and TEW conceived the ideas, designed methodology, analysed the data, and
499 wrote the manuscript.

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