Genomics-Based Identification of Microorganisms in Human Ocular Body Fluid

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ABSTRACT

Advances in genomics have the potential to revolutionize clinical diagnostics. Here, we examine the microbiome of vitreous (intraocular body fluid) from patients who developed endophthalmitis following cataract surgery or intravitreal injection. Endophthalmitis is an inflammation of the intraocular cavity and can lead to a permanent loss of vision. As controls, we included vitreous from endophthalmitis-negative patients, balanced salt solution used during vitrectomy, and DNA extraction blanks. We compared two DNA isolation procedures and found that an ultraclean production of reagents appeared to reduce background DNA in these low microbial biomass samples. We created a curated microbial genome database (>5700 genomes) and designed a metagenomics workflow with filtering steps to reduce DNA sequences originating from: i) human hosts, ii) ambiguousness/contaminants in public microbial reference genomes, and iii) the environment. Our metagenomic read classification revealed in nearly all cases the same microorganism than was determined in cultivation- and mass spectrometry-based analyses. For some patients, we identified the sequence type of the microorganism and antibiotic resistance genes through analyses of whole genome sequence (WGS) assemblies of isolates and metagenomic assemblies. Together, we conclude that genomics-based analyses of human ocular body fluid specimens can provide actionable information relevant to infectious disease management.

Introduction

Genomics-based analyses of patient specimens have the potential to provide actionable information that could facilitate faster and possibly more precise clinical diagnoses and guide treatment strategies in infectious diseases. A medical condition where a faster and more precise diagnosis could make a difference in clinical outcomes is endophthalmitis. Endophthalmitis is an acute intraocular inflammation that can lead to a permanent loss of vision. It often develops in response to microorganisms (usually bacteria and fungi) that enter the eye following eye surgery such as cataract surgery and intravitreal injection. The treatment strategy as well as visual outcome depends in part on the identity of the causative agents. For example, endophthalmitis cases involving coagulase-negative staphylococci have a better prognosis than cases involving enterococci or streptococci [1]. Often, the involving bacteria appear to originate from the patients' own microbiota, but may also be introduced through contaminated solutions or instruments used during eye surgery [2,3]. Endophthalmitis is an acute emergency and therefore clinicians start with a treatment before obtaining information about the identity of the causing microbial agent. It is anticipated that in the future, a more rapid determination of the identity of the causing agents and their antimicrobial resistance profiles using diagnostic metagenomics could facilitate the application of more precise treatments and reduce blindness.

Cataract is a condition in which the lens of the eye becomes progressively opaque and is one of the major causes of reversible visual loss. It is estimated that every year 10 million cataract surgeries are performed around the world [4]. The risk of endophthalmitis after cataract surgery is 1.4-4 per 10,000 cataract surgeries in the US and Denmark, and can be higher in other countries [1,5,6]. About 1/3 of the eyes with endophthalmitis in cataract patients remain blind after treatment [7].

Intravitreal injection with anti-vascular endothelial growth factor (anti-VEGF) has revolutionized the treatment of wet age-related macular degeneration, as well as diabetic maculopathy, and retinal vein occlusions during the last decade. It is the fastest growing procedure in ophthalmology and it was estimated that the number of intravitreal injections in the US would reach nearly 6 million in 2016 [8]. The risk of endophthalmitis after intravitreal injection is approximately 4.9 per 10,000 intravitreal injections [9].

The diagnosis and treatment of endophthalmitis is performed by vitrectomy surgery or a vitreous tap [10]. A vitrectomy is a procedure in which the vitreous body of the eye, which is the immobile gel-like fluid that occupies the space between the lens and retina, is aspirated and replaced by balanced salt solution. A vitreous tap is a more simple procedure where the vitreous is aspirated without being replaced by balanced salt solution. In both procedures, antibiotics, such as vancomycin combined with ceftazidime, are being injected into the vitreous body to treat the infection. The vitreous is often examined for infectious agents in the clinical laboratory using cultivation-based techniques.

In the clinical setting it is challenging to distinguish between infectious endophthalmitis and non-infectious ("sterile") endophthalmitis. Studies have shown that the proportion of culture-positive cases can be as low as 39% after cataract surgery and 52% after intravitreal injection [9,11]. Polymerase chain reaction can increase the rate of identifying the microorganisms by 20% [12], but in many endophthalmitis cases a causative agent cannot be identified. It is also unclear, whether the vitreous in endophthalmitis may contain multiple microorganisms that are not all being detected with the current methods. Furthermore, from a clinical perspective it is of importance to have a

method that facilitates the identification of the cases of non-infectious endophthalmitis. Non-infectious endophthalmitis can present as a variant of TASS (toxic anterior segment syndrome), and these patients may benefit from steroid instead of antibiotic treatment to obtain a better visual outcome [13].

Genomics approaches have the potential to revolutionize clinical diagnostic and therapeutic approaches in particular in the area of infectious diseases. Using shotgun metagenomic sequencing, a range of microorganisms and possible causing agents (e.g. bacteria, archaea, fungi, protozoa, viruses) can be identified [14,15]. In addition, upon cultivation-based isolation of microorganisms from the patient specimen, these can be subjected to whole genome sequencing (WGS) and *in silico*-determination of their taxonomic affiliation, phylogenetic relationships, potential antibiotic resistance genes, and virulence-associated genes [16,17].

Here, we perform metagenomic sequencing of vitreous specimens obtained from patients with endophthalmitis and a range of control samples. We evaluate two DNA isolation procedures for vitreous, and describe a bioinformatics workflow for data analysis and identification of potential infectious agents. The workflow includes *in silico* filtering steps for the removal of human DNA sequences, ambiguous and contaminant sequences in reference genomes from public repositories, and background DNA detected in control samples. We compare the metagenomics-based results with the results from the routine clinical cultivation- and mass spectrometry-based analysis, as well as to WGS-based identification of isolates obtained from the vitreous. Our findings suggest that metagenomics analysis together with WGS-based analysis is suitable for the identification of the potential infectious agents from human ocular body fluid, and in the future could guide therapeutic strategies including targeted antimicrobial therapy and the choice of steroids.

Results

Study design and metagenomic sequencing

To evaluate the use of shotgun metagenomic sequencing for the identification of potential disease-causing agents in postoperative endophthalmitis, we collected vitreous during vitrectomy from 14 patients with endophthalmitis (7 post cataract surgery, 7 post intravitreal injection) (Figure 1, Supplementary Table S1). As control, we obtained vitreous from 7 patients without endophthalmitis during macula hole surgery. Additional controls included 6 balanced salt solution (BSS) aliquots, of which 3 originated from individual bottles (BSS-B), and 3 from the vitrectomy BSS infusion lines (to be inserted into the eye) after the bottle had been connected to the vitrectomy system (BSS-S) (Figure 1). As there exist no standard procedure for the isolation of DNA from vitreous, we examined two procedures using the QIAamp DNA Mini Kit (QIA) and QIAamp UCP Pathogen Mini kit (UCP), and 4 extraction (blank) controls were included per kit (Figure 1).

The 62 samples were sequenced using Illumina MiSeq sequencing technology and a total of 90.6 million raw read-pairs were obtained. The average number of read-pairs after quality control for the endophthalmitis patients were 2.1/2.3 million read-pairs (QIA/UCP), and for the endophthalmitis-negative vitreous samples 1.0/0.6 million read-pairs (QIA/UCP). The average number of read-pairs for the BSS samples were 52,899/6,067 (QIA/UCP), and for the DNA extraction controls 20,931/3,134 (QIA/UCP). Overall, more read-pairs were obtained on average for the control samples when extracted with the QIA kit, while more read-pairs were obtained for the vitreous from the endophthalmitis patients when extracted with the UCP kit (Supplementary Fig. S1, Supplementary Table S2).

132
133 **Identification of human-affiliated DNA sequences**
134 In a first-pass analysis, in which we mapped the reads against a set of reference genomes, we
135 detected a high number of reads affiliated with human DNA sequences, which was anticipated in
136 particular in the endophthalmitis cases that can experience an infiltration of immune cells into the
137 vitreous chamber. Hence, we implemented a 2-step filtering process to remove the reads affiliated
138 with human genome sequences (Figure 2). In the first step we removed the reads that mapped to the
139 human reference genome (GRCh8.p10). Due to the genetic individuality of humans some reads
140 might not map to this reference genome, and therefore we removed in a second step all reads that
141 aligned to any human DNA sequence entry in the NCBI nt database (Supplementary Fig. S2,
142 Supplementary Table S2).
143
144 **Identification of ambiguous and contaminant DNA sequences in genomes from public**
145 **repositories**
146 In the initial first-pass analysis involving mapping of reads against reference genomes, we observed
147 that some genomes recruited particular high numbers of reads. These included *Hammondia*
148 *hammondi* strain H.H.34, *Alcanivorax hongdengensis* Strain A-11-3, *Toxoplasma gondii* ME49,
149 and *Arthrobacter* sp. Soil736. Upon inspection of these genomes we found that the reads mapped
150 only to specific genome sequence fragments such as short contigs and scaffolds (Supplementary
151 Fig. S3). To examine why specific contigs and scaffolds recruited high numbers of reads, we
152 aligned these against the nucleotide collection nt (NCBI). We found that the Top10 matches for
153 most of these contigs and scaffolds included several human DNA sequence entries that are not part
154 of the human reference genome GRCh8.p10 (Supplementary Table S3). While a few scaffolds of
155 *Hammondia hammondi* strain H.H.34 aligned with human DNA sequences (e.g. scaffold
156 NW_008644893.1), many aligned to *Bradyrhizobium* spp. genomes in the nt database
157 (Supplementary Table S3), indicating that human as well as microbial sequence contamination can
158 be found in public genome assemblies.
159
160 **Construction of a curated microbial genome database**
161 Our analysis suggested that some microbial reference genomes contain ambiguous/contaminant
162 sequences and we aimed at constructing a curated microbial genome database, devoid of these
163 sequences to the extent possible. Removing these sequences could reduce the number of false
164 positive hits that are the result of either contaminant sequences in the (incomplete) genome
165 assemblies, or because highly similar sequence regions naturally exist across genera that result in
166 the classification of reads to a different genus. We examined 5715 of the microbial reference and
167 representative genomes (archaea, bacteria, fungi, protozoa) (Supplementary Table S4) and aligned
168 all sequences ≤10 kb against the nucleotide collection nt (for a detailed description, see
169 Supplementary Methods). A total of 70,478 ambiguous sequences (contigs and scaffolds) were
170 identified, of which the majority were detected in incomplete microbial genomes. A total of 62% of
171 all incomplete microbial genomes had sequences flagged as ambiguous (range: 1 - 10,590; average:
172 28 sequence fragments). Ambiguous sequences were identified in 43% of all bacterial and 72% of
173 all protozoan genomes, and on average comprised 0.36% and 0.84% of the total genome sequence,
174 respectively (Table 1, and https://figshare.com/s/a282670f1405eae232df,
175 https://figshare.com/s/045b1252bd7555b50ef0, https://figshare.com/s/c42158cdee23f25489cd) [18].
176 The ambiguous sequences were removed and the resulting reference microbial genome database
177 contained a total of 5,751 genomes with 34 Tb (including 3.1 Tb for the human genome). The code
178 for the creation of the curated microbial reference genome database is accessible from Github

179 (https://github.com/philDTU/endoPublication), and the curated microbial reference genomes can be
180 downloaded from ftp://ftp.cbs.dtu.dk/public//CGE/databases/CuratedGenomes.
181
182
183 **Identification of contaminant (environmental background) DNA sequences in samples**
184 From the sequencing of DNA extraction (blank) control samples we obtained sequencing data,
185 albeit at a lower frequency compared to the patient specimens (Supplementary Fig. S1). The *in*
186 *silico* identification and removal of background DNA sequences are of critical importance,
187 particularly from specimens where the potential infectious agent may be present in low abundance.
188 We carefully examined the eight DNA extraction control samples and devise a list of the most
189 abundant and frequent environmental contaminant taxa in these samples (Supplementary Table S5,
190 Supplementary Fig. S4). We did not include taxa in the list that were occasionally observed in
191 endophthalmitis-positive patients and that were detected at a higher abundance in these samples
192 than in the respective DNA extraction controls. These non-contaminant taxa include *Enterococcus*
193 *faecalis*, *Escherichia coli*, *Micrococcus luteus*, *Staphylococcus aureus*, and *Staphylococcus*
194 *epidermidis* (Figure 3, and https://figshare.com/s/a4fd9d84260e8456ab72). The microbial
195 composition patterns in the DNA extraction control samples appeared to be influenced by the
196 choice of DNA isolation kit, the day of DNA extraction, and sequencing run (Supplementary Fig.
197 S4). The contaminant taxa (Supplementary Table S5) were removed from the datasets of all
198 endophthalmitis patients.
199
200 **The microbial composition in endophthalmitis-negative and balanced salt solution samples**
201 **is similar to DNA extraction controls**
202 The contaminant taxa that were identified in the DNA extraction controls were often present at
203 similar abundances in the endophthalmitis-negative (vitreous control) and balanced salt solution
204 samples (Figure 3, Supplementary Fig. S5). We found certain taxa to be specific for the DNA
205 isolation method (QIA or UCP) in round C of DNA extractions (Supplementary Fig. S5,
206 Supplementary Table S2). Samples processed using the QIA method contained *Pseudomonas* spp.,
207 *Acinetobacter* spp., and *Janthinobacterium* spp. among others, and samples processed with the UCP
208 method included mainly *Bradyrhizobium* spp. Other organisms appeared to be present across all
209 samples (Supplementary Fig. S5). For example, *Cutibacterium acnes* and *Propionibacterium*
210 *humerusii* were detected in most samples and they might represent environmental bacteria
211 originating from the staff handling the samples or fomites such as the laboratory equipment and
212 supplies.
213
214 **Microorganisms in endophthalmitis-positive patients as determined by metagenomics**
215 For 12 out of 14 endophthalmitis patients a dominant microorganism was identified in the vitreous
216 (for all UCP-extracted, and most QIA-extracted specimens) using the read classification approach
217 (Figure 4 and 5). These organisms included *Staphylococcus epidermidis* (six patients),
218 *Enterococcus faecalis* (two patients), *Serratia marcescens* (one patient), *Paenibacillus* spp. (one
219 patient), and *Staphylococcus hominis* (one patient). In one patient (C5), a number of different
220 organisms were identified, most dominantly *E. coli* in the UCP-extracted specimen (>3000 reads),
221 *Moraxella catarrhalis* (11 reads) in the QIA-extracted specimen, and *Micrococcus luteus* with 9
222 and 45 reads in QIA and UCP-extracted samples, respectively (Figure 4 and 5, and
223 https://figshare.com/s/5feabfad1d8c495bf7a3). For two additional patients, *Commamonas*
224 *testosteronii* and *Escherichia coli*, or *Caulobacter* spp. were identified as the most dominant
225 organisms respectively (C1, I7), however, these were only represented by <25 reads. In the seven
226 patients that contracted endophthalmitis following cataract surgery, the most frequent bacteria were

227 *Enterococcus faecalis* (two patients), *Staphylococcus epidermidis* (two patients), and *Serratia*
228 *marcescens* (one patient). In the seven patients with endophthalmitis following intravitreal
229 injection, the most frequent bacteria were *Staphylococcus epidermidis* (four patients), *Paenibacillus*
230 spp. (one patient), and *Staphylococcus hominis* (one patient) (Figure 5). Overall, potential causing
231 agents where identified with 58 reads (*Paenibacillus* spp.) as a lower bound in patient I2, and
232 2,999,838 reads as the highest detected read count (*Staphylococcus epidermidis*) in patient I4. The
233 presence of the two most frequently detected bacteria, *Staphylococcus epidermidis* and
234 *Enterococcus faecalis*, in the vitreous fluid was also verified using targeted PCR assays (See
235 Supplementary Methods, and https://figshare.com/s/0e8a98f436f07efc4dd5).
236
237 In addition to the read classification approach, we constructed metagenomic assemblies for the
238 individual samples and characterized these according to a number of taxonomic and functional
239 categories, including bacterial species affiliation, sequence type, genomic MLST, resistance genes,
240 virulence-associated genes, and plasmids. For the three patients for whom we obtained high
241 numbers of classified reads using the read classification approach described above (C6, C7, I4)
242 (Figure 5), we obtained information in nearly all categories using the metagenomics assembly
243 approach (Supplementary Table S6). The taxonomic information that we obtained using the
244 metagenomic assembly approach was in agreement in all cases with the taxonomic information we
245 obtained using the metagenomic read classification approach. Furthermore, using metagenomics
246 assembly analysis we detected a number of antimicrobial resistance genes in the specimens for
247 which we also obtained taxonomic information. In addition, we detected a streptogramin B
248 resistance gene (vat(B)) in sample C3_UCP, an aminoglycoside resistance gene (aadD) in sample
249 C7_QIA, and a Col plasmid origin of replication in sample C5_UCP. Of note, five out of the seven
250 total samples, for which we obtained information using the metagenomics assembly approach, were
251 ocular body fluid samples that had been processed using the UCP DNA isolation protocol
252 (Supplementary Table S6).
253
254 **Bacterial isolates from endophthalmitis patients have in most cases the same identity as**
255 **the most abundant organism determined by metagenomics**
256 At the hospital microbiology laboratory, bacteria could be isolated from the vitreous right after
257 vitrectomy for 12 out of 14 patients. The identity of the isolates was determined by MALDI-TOF
258 mass spectrometry (MS), and in nine cases the same agent was identified as in the metagenomic
259 analysis (Figure 5). In addition, *Micrococcus luteus* was isolated from patient C5 in both QIA- and
260 UCP-extracted samples (9/45), but this organism was not the most abundant one identified using the
261 metagenomics sequencing-based method (Figure 5). Using the cultivation-based method a *Bacillus*
262 sp. (Order: Bacillales) was determined for patient I2, and reads classified as *Paenibacillus* spp.
263 (Order: Bacillales) were identified using the metagenomics analysis. A *Staphylococcus aureus*
264 culture was obtained in the hospital for patient I3, and *S. aureus* was also represented with 22/18
265 (QIA/UCP) reads in the metagenomics analysis in this patient, even though *S. epidermidis* was the
266 most abundant organisms identified using this approach (150(QIA)/162(UCP) reads). In the two
267 cases for which the culture-based approach was negative (C1, I7), only fewer than 25 reads were
268 classified using the metagenomics approach.
269
270 At the research laboratory, we attempted a recultivation of microorganisms from frozen vitreous
271 and successfully obtained isolates for six patients. Different colony morphotypes on the agar plates
272 were obtained and analysed using whole genome sequencing (WGS) and MALDI-TOF mass
273 spectrometry (MS). These isolates had the same species affiliation according to WGS and MS
274 analyses as the isolates obtained at the hospital for the same patient, and as identified in the vitreous

275 using metagenomics analysis (Figure 5). The presence of the isolates in the vitreous samples was
276 further verified by mapping the shotgun metagenomic reads originating from the vitreous to the
277 genomes of the isolates, and an even breath of coverage was observed for all isolates
278 (https://figshare.com/s/c2ce2d32daf25db54904).
279 Using WGS we found that for multiple morphotypes the same organism and sequence type was
280 identified, with one exception. For patient I6, we obtained three *Staphylococcus epidermidis*
281 isolates and of which two belonged to sequence type ST-88 and one to ST-487. The *Staphylococcus*
282 *epidermidis* isolates obtained from other patients (C3, C7, I4) belonged to different sequence types
283 (Figure 5, Supplementary Table S7), suggesting that they have different origins. Each
284 *Staphylococcus epidermidis* sequence type exhibited its own set of antibiotic resistance genes,
285 including genes facilitating resistance to macrolides, β-lactams, aminoglycosides, and tetracyclines.
286 The *Enterococcus faecalis* from two patients (C2, C6) belonged to different sequence types, and
287 both sequence types shared a gene facilitating resistance towards macrolides (Figure 5,
288 Supplementary Table S7). Several resistance genes that were identified in the sequenced isolates
289 were also identified in the metagenomic assembly analysis (Figure 5, Supplementary Table S6).
290 Some of the resistance genes and their predicted functions identified using the genomics approaches
291 were also in alignment with results from the phenotypic antibiotic susceptibility testing of the
292 isolates obtained during the 1st culturing at the hospital
293 (https://figshare.com/s/e579abea97dfc8c77a6a).
294
295 **Detection of bacteriophages and human DNA viruses**
296 As we did not identify a dominant microorganism in two endophthalmitis patients (C1, I7) we
297 examined whether these or any of the other specimens contained DNA viruses not represented in
298 our microbial genome database. We added an additional 7,180 virus genome sequences to the 35
299 RefSeq virus genomes (https://figshare.com/s/b040289827b79d3a60df) in our database and
300 classified our metagenomic sequencing data using kraken. We identified several *Enterococcus*,
301 *Staphylococcus* and *Propionibacterium* bacteriophages among others in specimens that also were
302 identified to contain the respective bacterial host (https://figshare.com/s/ff0527509828d1529ad9).
303 To evaluate whether our metagenomics approach (Figure 2) would facilitate the identification of
304 human DNA viruses, we analysed metagenomic data obtained from patients with uveitis in which
305 human DNA viruses had been detected previously [19]. We obtained similar results as described by
306 Doan and colleagues, including the identification of herpes simplex virus 1 (HSV-1) in subject 1,
307 and rubella virus in subject 6 (Supplementary Table S8). In the previous study *Hammondia*
308 *hammondi* was identified in subject 3 as the second most abundant organism after *Toxoplasma*
309 *gondii*. We also detected *Toxoplasma gondii* as the most abundant organism in this specimen
310 (represented by 4,410 reads), but did only detect 4 reads for *Hammondia hammondi*; most likely
311 because we had removed DNA sequences from the *Hammondia hammondi* genome in the database
312 that were flagged as ambiguous. In addition, in subject 5 we detected *Ochrobactrum anthropi*, an
313 agent that had been identified previously in eye infections such as endophthalmitis and keratitis
314 [20,21]. However, we detected *Ochrobactrum anthropi* in high abundance in the water control sample,
315 and therefore it may here rather represent an environmental contaminant.

316 ## Discussion

317

318 Metagenomic sequencing-based analyses of complex patient specimens and whole genome
319 sequencing (WGS) of microbial isolates will advance clinical diagnostics and treatment strategies in
320 infectious diseases [22-24]. One example, for which this strategy may be advantageous is postoperative
321 endophthalmitis as currently a causing microbial agent can only be identified in a fraction of these
322 cases [1]. Immediate diagnosis and treatment of endophthalmitis is required to prevent vision loss of
323 the affected eye, and it would be helpful to be able to distinguish between infectious and non-
324 infectious ("sterile") endophthalmitis. Challenges in clinical metagenomics remain at several levels,
325 from specimen collection and processing to the generation of actionable information. We examine
326 here vitreous samples from endophthalmitis patients together with a variety of control samples,
327 evaluate two DNA isolation procedures, create a curated microbial reference database, and present a
328 workflow for metagenomic sequencing data analysis. We compare the results from metagenomic
329 read analysis to WGS and MALDI-TOF mass spectrometry identification of isolates obtained for
330 several patients, as well as to results from metagenomic assembly analysis.

331

332 Vitreous samples were collected from 14 patients with endophthalmitis. Seven patients developed
333 endophthalmitis post cataract surgery, in which the natural intraocular lens was exchanged with an
334 artificial one, without the introduction of surgical instruments into the vitreous body. Another seven
335 patients developed endophthalmitis post intravitreal injection, a procedure in which drugs were
336 introduced into the vitreous body using surgical instruments to treat retinal diseases such as age-
337 related macular degeneration. As controls, we included i) vitreous samples from seven
338 endophthalmitis-negative patients, ii) balanced salt solution used during vitrectomy from both,
339 individual bottles and the vitrectomy system after the solution had passed the vitrectomy infusion
340 lines, and iii) DNA extraction (blank) controls (Figure 1).

341

342 We investigated two DNA isolation procedures, QIAamp DNA Mini Kit (QIA) and QIAamp UCP
343 Pathogen Mini kit (UCP), for metagenomics analysis to determine possible infectious agents in the
344 vitreous fluid. We obtained more reads (total and classified) on average for endophthalmitis-
345 positive specimens when vitreous fluid was extracted with the UCP kit compared to the QIA
346 procedure (Supplementary Figure S1 and Figure 5). In contrast, lower numbers of reads on average
347 were obtained from the three types of control samples when they were extracted with the UCP kit
348 compared to the QIA procedure (Supplementary Figure S1). Our analysis revealed that UCP-
349 extracted control samples harboured a lower microbial diversity compared to QIA-extracted ones
350 (Supplementary Figures S4 and S5). Even though we identified distinct QIA and UCP kit
351 "fingerprints", bacteria such as *Cutibacterium acnes* and *Propionibacterium humerusii* were present
352 as background DNA across samples, independent of the DNA isolation kit. These bacteria likely
353 originated from the staff handling the samples and/or additional laboratory supplies that were used
354 during sample handling and processing [25]. Contaminant background DNA has been identified
355 previously in other DNA isolation kits [26-28], and our analysis suggests that an ultraclean production
356 of reagents and consumables reduced the amount of background DNA in the UCP DNA isolation
357 kit reagents and/or supplies. Contaminant viral DNA has been identified in previous sequencing-
358 based studies as well such as hybrid parvovirus-like virus NIH-CQV/PHV DNA from silica
359 column-based nucleic acid isolation kits [29,30], and which we detected in our samples, too
360 (https://figshare.com/s/ff0527509828d1529ad9). We did not, however, detect torque teno virus
361 DNA, as previously described for some endophthalmitis cases [31]. Overall, the UCP kit appeared to

362 be suited for the isolation of DNA from vitreous, and may potentially be useful for other human
363 body fluids and biological specimens that are assumed to have a low microbial biomass.
364
365 Our metagenomics data analysis workflow included three filtering steps (Figure 2) to reduce i)
366 human host DNA sequences, ii) false positive hits due to ambiguous and contaminant DNA
367 sequences in reference genomes, and iii) environmental background DNA sequences introduced by
368 kit reagents, potentially other laboratory supplies, as well as laboratory staff. We particularly found
369 that ambiguous/contaminant sequences in public genomes, which serve as reference in many
370 metagenomic studies, could lead to the false positive identification of microorganisms. Our initial
371 read classification, in which we used the original reference genomes, revealed *Toxoplasma gondii*
372 (false positive) across samples, even after filtering reads that mapped to the human reference
373 genome. Some microbial reference genomes appeared to harbour human DNA sequences not
374 present in the human reference genome, thus making it challenging to detect these sequences in the
375 initial human DNA sequence filtering step. This effect is especially critical when analysing clinical
376 specimens, since the patient's DNA is expected to be found in these samples. In addition, we
377 noticed that certain microbial genomes contained sequences that had a high similarity to other
378 microorganisms belonging to a different genus. These can be correct naturally occurring DNA
379 sequence regions that have a high similarity across a range of microbial taxa (including regions
380 acquired via horizontal gene transfer). In other cases they can be contaminant contigs or scaffolds in
381 primarily incomplete genome sequence assemblies. In either case, the read classification can lead to
382 a false positive identification of microorganisms. Contaminant DNA sequences in published
383 genomes have been previously found, particularly in human and animal genome assemblies [32-34].
384 Hence, we systematically examined 5,715 microbial reference and representative genomes (archaea,
385 bacteria, fungi, protozoa), and in 62% of all incomplete microbial genomes sequences were flagged
386 as ambiguous. We removed 70,478 ambiguous sequences (including human and microbial
387 contaminants), reflecting 0.35% of the bases in total from the microbial reference genomes
388 (https://figshare.com/s/045b1252bd7555b50ef0). The majority of the removed sequence fragments
389 are a correct part of the respective genome. However, a more complex and thorough analysis is
390 required in the future to decide whether a particular part should be removed or not. In our case,
391 many of the remove sequences are plasmids or sequence fragments from lower-quality assemblies.
392 Since plasmids are mobile genetic elements it is unknown at what confidence-level they contribute
393 to the taxonomy assignment because some plasmids have a broad host range. The removed
394 sequences from the lower-quality assemblies should be neglectable in our clinical study, since most
395 of the identified infectious agents are not in that category. According to the insight gained from this
396 study we recommend using curated microbial reference genomes in microbiome studies and
397 particularly for the analysis of clinical samples with an assumed low microbial biomass.
398 Additionally, subsequent to filtering we recommend to always check the sequences that were
399 removed. We provide a script that facilitates the generation of curated databases, including the one
400 used in this study (https://github.com/philDTU/endoPublication) as well as the sequences of the
401 curated genomes (ftp://ftp.cbs.dtu.dk/public//CGE/databases/CuratedGenomes).
402
403 Our analysis further demonstrated the benefit of including a variety of control samples. In fact, the
404 number of control samples in this study exceeded the number of the main samples under
405 investigation by a factor of 1.2. By analysing vitreous from endophthalmitis-negative patients,
406 aliquots of balanced salt solution (from bottle and vitrectomy infusion line), and blank DNA
407 extraction controls, we determined the background levels of organisms in the respective
408 environments. All control sample types had a similar microbiome pattern, characterized by
409 organisms found in the corresponding DNA extraction (blank) controls (QIA and UCP) and typical

skin inhabitants. We did not identify specific microorganisms for the endophthalmitis-negative patients, similar to previous cultivation-based assessments, suggesting that vitreous fluid is a sterile body part or only contains few microbial cells in individuals without eye infections [35,36]. We also did not identify any specific organisms residing in balanced salt solution that was infused into the patient's eye, in addition to the ones identified in DNA extraction controls. In all cases, we cannot however exclude that DNA sequences from other microorganisms would have been found if a deeper DNA sequencing had been performed or RNA had been isolated and analysed by deep sequencing. In our analysis of the vitreous fluid from endophthalmitis-positive patients we removed the background contaminant organisms *in silico* that were detected in the respective DNA extraction controls and were not present in higher abundance in the endophthalmitis-positive patients. To trace the origin of detected organisms, including infectious agents, additional controls in future studies could include samples from: i) the patients skin, eye lid, conjunctiva, or other body sites that are in proximity to the surgical site, ii) the surgical instruments, iii) blank tubes and/or devices used for the collection of the patient specimen, as well as iv) reference mock communities with known composition. Careful analysis of control samples may assist in the design of harmonized standards and guidelines for the sequencing-based analysis of clinical samples and other biological specimens.

Through our metagenomics read classification data analysis workflow we identified a single potential causing microorganism in 11 out of 12 culture-positive cases, and which in most cases agreed with the cultivation-based analyses (Figure 5). For patient C5 we did not identify a single potential causing agent and instead obtained different microbiome patterns for the two sequenced aliquots (QIA and UCP). In both samples we detected *Micrococcus luteus*, in alignment with the cultivation-based analysis. However, *Micrococcus luteus* was not the most abundant organism in the metagenomic analysis. *Escherichia coli* was the most abundant organism in the UCP-extracted sample, but which may also be a contamination introduced during sample handling. For patient I2 we revealed *Paenibacillus* spp. as a possible causing agent, whereas in the cultivation-dependent analysis the isolate was identified as a related *Bacillus* sp using MALDI-TOF. For patient I3, our metagenomic analysis suggests a potential infection by *Staphylococcus epidermidis* together with *Staphylococcus aureus*. In the cultivation-based analysis *Staphylococcus aureus* was identified as the potential causing agent. Only a few metagenomic reads were classified for patients C1 and I7, and which were regarded as contaminants. For these two patients no microorganisms could be isolated by cultivation-based methods, neither at the hospital nor the research laboratory. Therefore, these two patients are assumed to have a non-infectious (sterile) endophthalmitis.

Both, the analysis of metagenome assemblies as well as whole genome sequences of isolates can reveal the presence of antibiotic resistance genes that could potentially guide therapeutic treatment strategies, in particular when verified by results from susceptibility analysis of isolates. In addition, the specific sequence type for infectious agents, such as *Enterococcus faecalis*, *Staphylococcus epidermidis*, and other organisms, can be identified and assist in the source tracking and epidemiology of the particular agent. Detailed evolutionary relationships between isolates can be revealed if sufficient genome sequence information has been obtained. Our analysis of the identified *Enterococcus faecalis* and *Staphylococcus epidermidis* suggests that they may originate from the individuals involved in the surgery or the immediate environment, as different bacterial sequence types and resistant profiles were identified across patients. The metagenomics analysis did not reveal these bacteria to be present in the balanced salt solution samples, further pointing towards an acquisition from another source.

458 Our previous clinical microbiology research on urinary tract infections and diarrhoeal diseases [37,38]
459 had some limitations for assessing clinical metagenomics as a technology, and whose analysis could
460 now be improved by new insight gained from this study. For example, the examined urine samples
461 were pre-processed by using a centrifugation step to remove human cells [37]. In this step, also
462 microbial cells may have been removed that have a similar density than the human cells and/or
463 were attached to these. While this step can be advantageous to limit human contaminant sequences,
464 it could be of interest to examine samples with and without the sedimentation step and using the
465 DNA isolation procedure and/or data analysis pipeline described in the present study. Furthermore,
466 the presence of potential contaminant DNA sequences was not examined in the previous study. In
467 the study concerning diarrhoeal diseases one challenge was to differentiate between natural
468 intestinal inhabitants, possible infectious agents, and potential contaminants [38]. Careful
469 bioinformatics filtering steps and inclusion of control samples, as used in the present study, might
470 allow for more robust identifications in the future. To facilitate a more standardized workflow for
471 sample analysis, we have created a list of recommendations for the design and execution of
472 metagenomic sequencing projects (https://figshare.com/s/2a0709b1f0c5e18754df), in addition to
473 specific details described in this study.
474
475 In summary, we find that metagenomics analysis, supported by WGS of isolates, may be a
476 promising strategy for the identification and characterization of infectious agents from human
477 ocular body fluid. This technology may also facilitate a more robust differentiation between
478 infectious and non-infectious ("sterile") endophthalmitis. Nucleic acid extraction from patient
479 specimens, followed by high-throughput sequencing may ultimately provide more rapid insight in
480 regard to the identity of the causing agent(s) than cultivation-based techniques, in particular in light
481 of recent developments in long-read nanopore sequencing and real-time analysis [39-41]. In cases
482 where the metagenomic sequencing depth of coverage of the microorganism is sufficiently high,
483 valuable functional information such as antibiotic resistance and virulence-associated genes can be
484 revealed. Prerequisites for a robust data analysis are suitable procedures that facilitate the isolation
485 of nucleic acid from microorganisms residing in complex samples, the analysis of relevant control
486 samples, as well as high-quality genome sequence reference databases for data analysis, as
487 exemplified in this study.
488
489

490 # Methods

491 ### Vitreous Samples

492 A total of 21 vitreous samples from 21 individual patients were examined in this study. From April
493 2012 to November 2013, vitreous samples from 14 eyes with postoperative endophthalmitis
494 following cataract surgery (n=7) and intravitreal injection (n=7) were collected using vitrectomy
495 after informed consent had been obtained. At the Department of Ophthalmology, Glostrup
496 Rigshospitalet (Denmark), where vitreous was collected, all patients with suspected postoperative
497 endophthalmitis are treated with a vitrectomy independent of the presenting visual acuity. As
498 control, vitreous was collected from 7 patients without endophthalmitis during macula hole surgery
499 after informed consent had been obtained. Approximately 1-2 ml of vitreous body fluid was
500 aspirated from each eye. It was at the discretion of the vitreoretinal surgeon whether to aspirate the
501 vitreous sample before or after balanced salt solution installation. About half of each collected
502 sample was cultured in the acute clinical setting at the Department of Microbiology, Hvidovre
503 Hospital, Denmark, and the remaining material was stored at -80°C.
504

505 ### Balanced salt solution samples

506 During vitrectomy, balanced salt solution (BSS PLUS, Alcon) is infused into the eye in order to
507 keep the appropriate tension of the eye. BSS PLUS is a sterile physiological saltwater solution
508 containing bicarbonate, dextrose and glutathione. Subsequently, 2.25 mg ceftazidime and 1 mg
509 vancomycin dissolved in 0.1 ml sterile salt solution are injected into the vitreous chamber. We
510 examined 3 paired sets of samples, i.e. 6 BSS samples in total. Aliquots were taken directly from
511 separate BSS PLUS bottles before vitrectomy at different time points during the study period.
512 Subsequently, BSS was collected from the vitrectomy surgical system after the BSS bottle had been
513 connected and BSS had passed through the vitrectomy infusion line. The aliquot obtained from the
514 vitrectomy system represents the fluid that is infused into the eye of the patient. The BSS samples
515 were stored at -80°C.
516

517 ### Isolation of DNA from complex samples

518 DNA was isolated from 200 µl vitreous fluid and balanced salt solution samples using two different
519 DNA isolation procedures, i) the QIAamp DNA Mini Kit (51304, Qiagen), and ii) the QIAamp
520 UCP Pathogen Mini Kit (50214, Qiagen). For each round of DNA isolation, one extraction control
521 (blank) was included. For details, see Supplementary Methods.
522

523 ### Metagenomic sequencing

524 The DNA was prepared and sequenced according to the Nextera XT DNA Library Preparation
525 Guide, Part # 15031942 Rev. D. Sequencing was performed on an Illumina MiSeq sequencer using
526 paired-end sequencing with v3 chemistry and 2×250 cycles. A total of 90,599,659 read pairs were
527 obtained from the samples. The number of read pairs was in a range of 711,886 – 4,633,576 for the
528 samples from patients with endophthalmitis, with the exception of sample I6_QIA for which only
529 106 read-pairs were obtained (Supplementary Table S2).
530

531 ### Metagenomic sequencing data analysis

532 The metagenomics analysis was carried out in five steps. 1) Adapter and quality trimming as well as
533 low complexity filtering of raw reads was performed using BBDuk of BBMap version 35.82
534 (http://jgi.doe.gov/data-and-tools/bbtools/). 2) Removal of human-affiliated reads from samples in a
535 2-step approach: i) reads that mapped against the reference genome GRCh38.p10
536 (GCF_000001405.36), and ii) reads that aligned to human sequences in the non-redundant
537 nucleotide collection (nt) database from NCBI. 3) Detection of ambiguous sequences in public

538 reference genomes and creation of curated microbial genome database that was composed of 5751
539 different genomes: archaea (251), bacteria (5166), fungi (225), protozoa (73), viruses (35) and the
540 human reference GRCh38.p7 (Supplementary Table S4). 4) Classification of reads in samples using
541 Kraken followed by Bayesian reestimation of abundance (Bracken) [42,43], and 5) Classification of
542 reads using BLASTn of BLAST version 2.6.0 [44]. For details, see Supplementary Methods.
543
544 **Cultivation and mass spectrometry (Clinical Microbiology lab)**
545 Aliquots from the vitreous specimens were cultivated for 12 days on 5% horse blood agar,
546 chocolate agar, brain heart infusion broth under anaerobic conditions, and on anaerobic plates (SSI
547 Diagnostica, Denmark) under anaerobic conditions at 35°C according to the standard operating
548 procedure at the Department of Clinical Microbiology, Hvidovre Hospital. Species identification
549 was performed using MALDI-TOF mass spectrometry analysis (MALDI Biotyper 3.1, Bruker
550 Daltonics Microflex LT, database MBT DB-5627) from colony material. Antimicrobial
551 susceptibility was tested towards a range of compounds and the results were interpreted in
552 accordance to EUCAST breakpoints (http://www.eucast.org/clinical_breakpoints/).
553
554 **Cultivation (Research lab)**
555 To isolate bacteria and fungi from the vitreous body and balanced salt solution samples, 100 µl
556 aliquots were distributed on chocolate agar (SSI Diagnostica, Denmark) and Sabouraud agar with
557 Chloramphenicol (Fischer Scientific). The chocolate agar was incubated for 2 days at 37°C.
558 Colonies from the chocolate agar plates were harvested and stored in Protect Multipurpose TS80
559 preservation tubes (Technical Service Consultants Ltd, UK) at -80°C. One representative colony
560 morphotype per sample was selected for whole genome sequencing. No growth after incubation for
561 5 days was observed on the Sabouraud agar plates.
562
563 **Whole genome sequence analysis**
564 Isolates were sequenced (2x150 bp paired-end) on a MiSeq system (Illumina, San Diego, CA, USA)
565 as previously described [45]. Reads were adapter trimmed and filtered for phiX reads using BBduk.
566 The high-quality reads where assembled using the SPAdes assembler [46], and the genome sequence
567 assemblies analysed using the Bacterial Analysis Pipeline [47]. For details, see Supplementary
568 Methods.
569
570 **Ethics**
571 This study was performed in accordance with the Declaration of Helsinki. It was approved by the
572 Danish Data Protection Agency (journal number: 2011-41-5881) and by the local ethics committee
573 De Videnskabsetiske Komiteer - Region Hovedstaden (journal number: H-2-2011-004), and took
574 place at public clinics in the capital region of Denmark.
575
576 **Data accessibility**
577 The sequencing data generated and analyzed in this study are available from DDBJ/ENA/GenBank
578 under the umbrella project PRJEB21503, including metagenomics shotgun reads (ERS1830261-
579 ERS1830322), WGS reads (ERS1827480-ERS1827489), and WGS assemblies (ERZ468526-
580 ERZ468535). A detailed methods description and results from the data analysis are available as
581 supplemental material from the journal website and through Figshare
582 (https://figshare.com/projects/Genomics-
583 Based_Identification_of_Microorganisms_in_Human_Ocular_Body_Fluid/21038). The code for the
584 creation of the curated microbial reference genome database is accessible from Github

585 (https://github.com/philDTU/endoPublication), and the curated microbial reference genomes can be
586 downloaded from ftp://ftp.cbs.dtu.dk/public//CGE/databases/CuratedGenomes.
587

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709

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Author contributions statement

P.K., S.S.B., H.W., and S.J.P. designed the research; P.K., S.S.B., A.F.M, H.W. and S.J.P. performed the research; P.K., F.M.A., H.W., and S.J.P. contributed analytic tools; P.K., S.S.B., A.F.M., H.W., and S.J.P. analysed the data; P.K. and S.J.P. wrote the manuscript; and S.S.B, A.F.M, F.M.A., and H.W. edited the manuscript. All authors have read and approved the manuscript as submitted.

Additional information

Competing financial interests

The authors declare that they have no competing interests.

731 **Figures**

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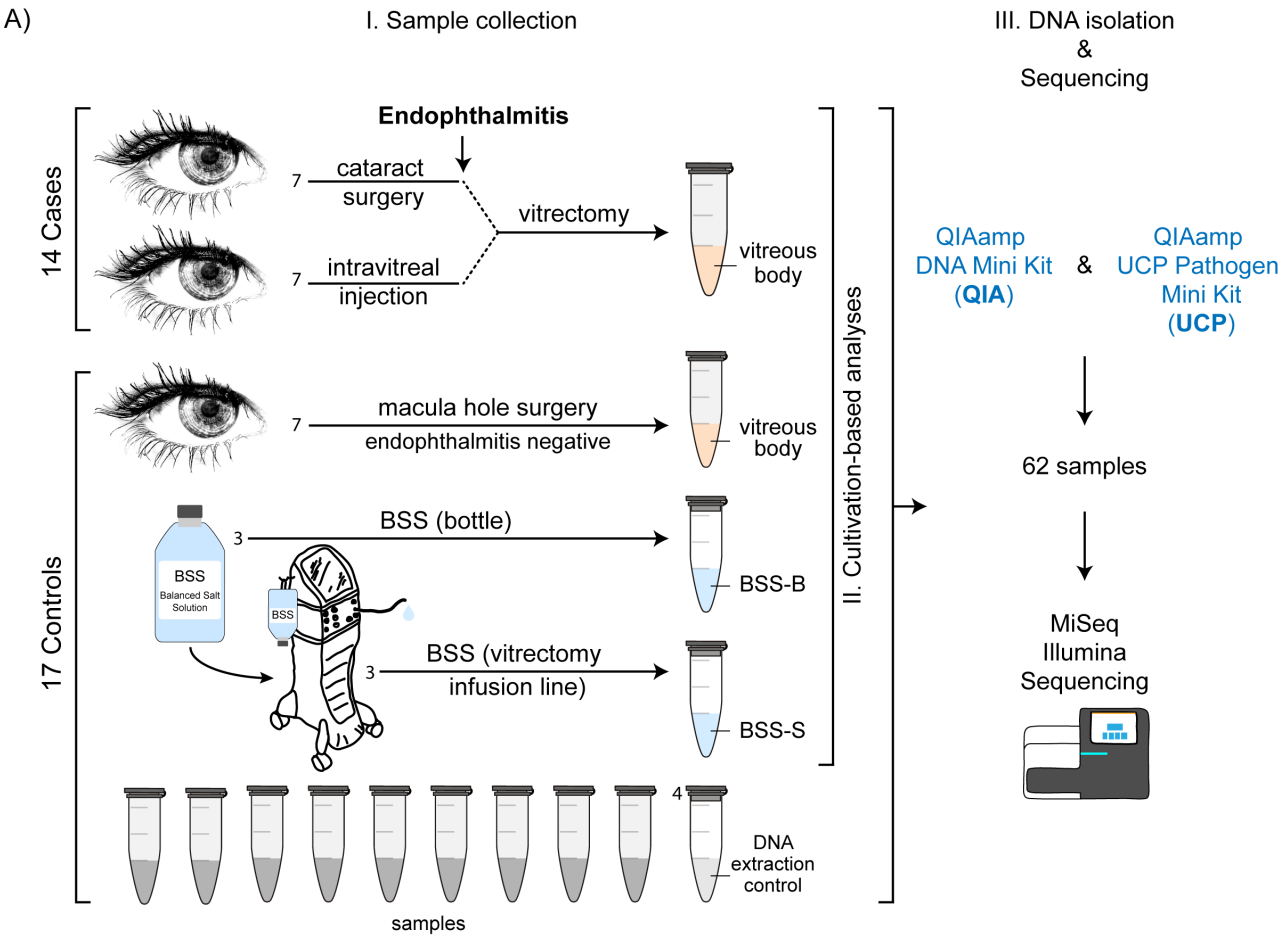

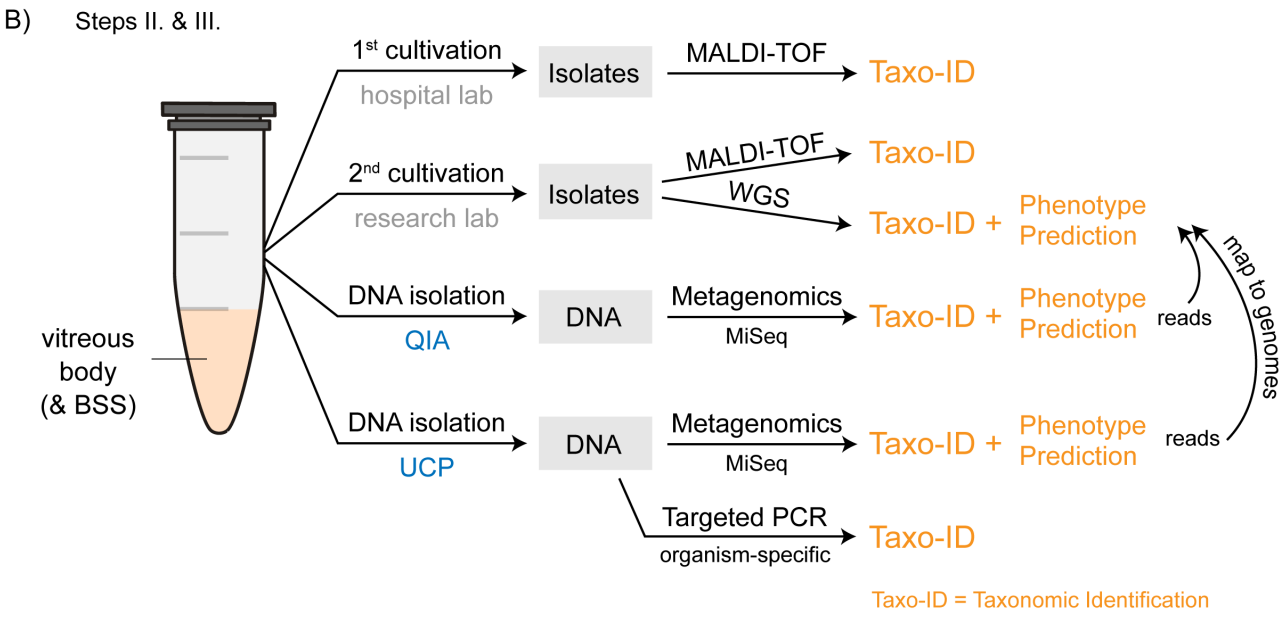

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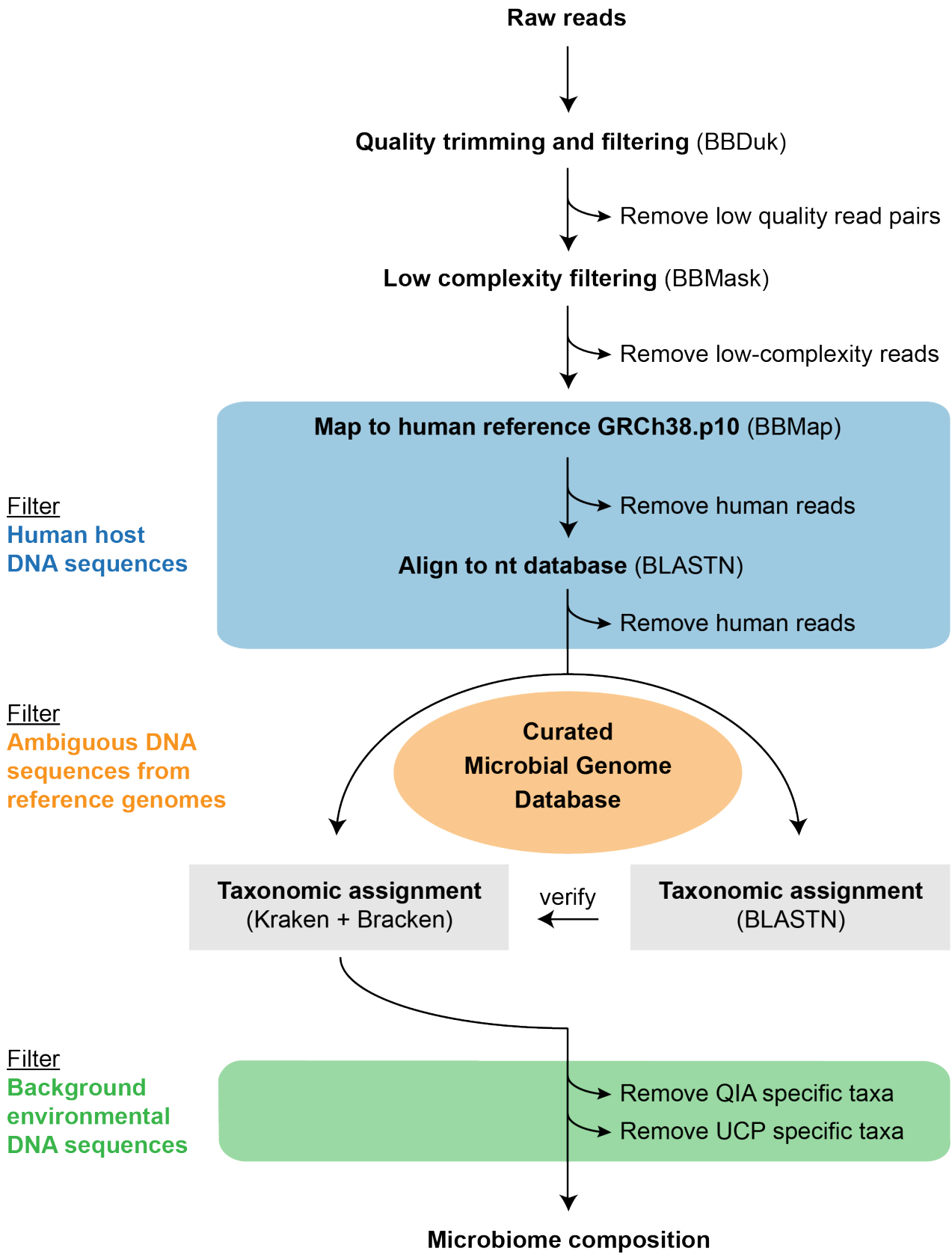
736
737 **Figure 1: Sample collection, DNA isolation, and shotgun metagenomic sequencing.**
738 A) I.) Sample collection: Vitreous body (intraocular body fluid) was collected through
739 vitrectomy from 14 patients with endophthalmitis following cataract surgery (n=7) and
740 intravitreal injection (n=7). As control, vitreous was collected from 7 patients without
741 postoperative endophthalmitis during macula hole surgery. Six aliquots (3 sample pairs)
742 were obtained from balanced salt solution (BSS) that is infused into the eye during
743 vitrectomy. Three aliquots were collected from separate BSS bottles (BSS-B), and the
744 second set of aliquots was collected from the vitrectomy surgical system (BSS-S) after it
745 had passed through the vitrectomy infusion line, respectively. The samples were
746 examined using II.) Cultivation-based analyses and III.) DNA isolation (2 methods) &
747 Metagenomic shotgun sequencing, including the examination of DNA extraction (blank)
748 controls. A total of 62 samples were sequenced using Illumina MiSeq sequencing
749 technology. B) More details to steps II.) and III.): II.) Cultivation-based analyses: Aliquots
750 of the vitreous body fluid and balanced salt solution samples were subjected to
751 cultivation-based analyses separately at the hospital and research laboratories. Obtained
752 isolates were analyzed using mass spectrometry and whole genome sequencing. III.) DNA
753 isolation & Metagenomic shotgun sequencing: Samples were extracted using two DNA
754 isolation procedures: QIAamp DNA Mini Kit (QIA), and QIAamp UCP Pathogen Mini kit
755 (UCP). A DNA extraction (blank) control was included at each round of DNA isolation, i.e.
756 one DNA extraction control for 12-14 samples in total per extraction round (more vitreous
757 samples were extracted than analyzed in this study). To verify the presence of the main
758 microorganisms detected in the metagenomics analysis, the shotgun metagenomics
759 reads were mapped to the genome assemblies of the isolates obtained from the vitreous
760 samples. Not displayed here is the mapping of metagenomic shotgun reads to microbial
761 reference genomes in the database (Provided in Figure 4). As an additional verification,
762 PCR analyses were carried out to detect the presence of the most abundant
763 microorganisms in the vitreous samples using organism-specific primer sets.

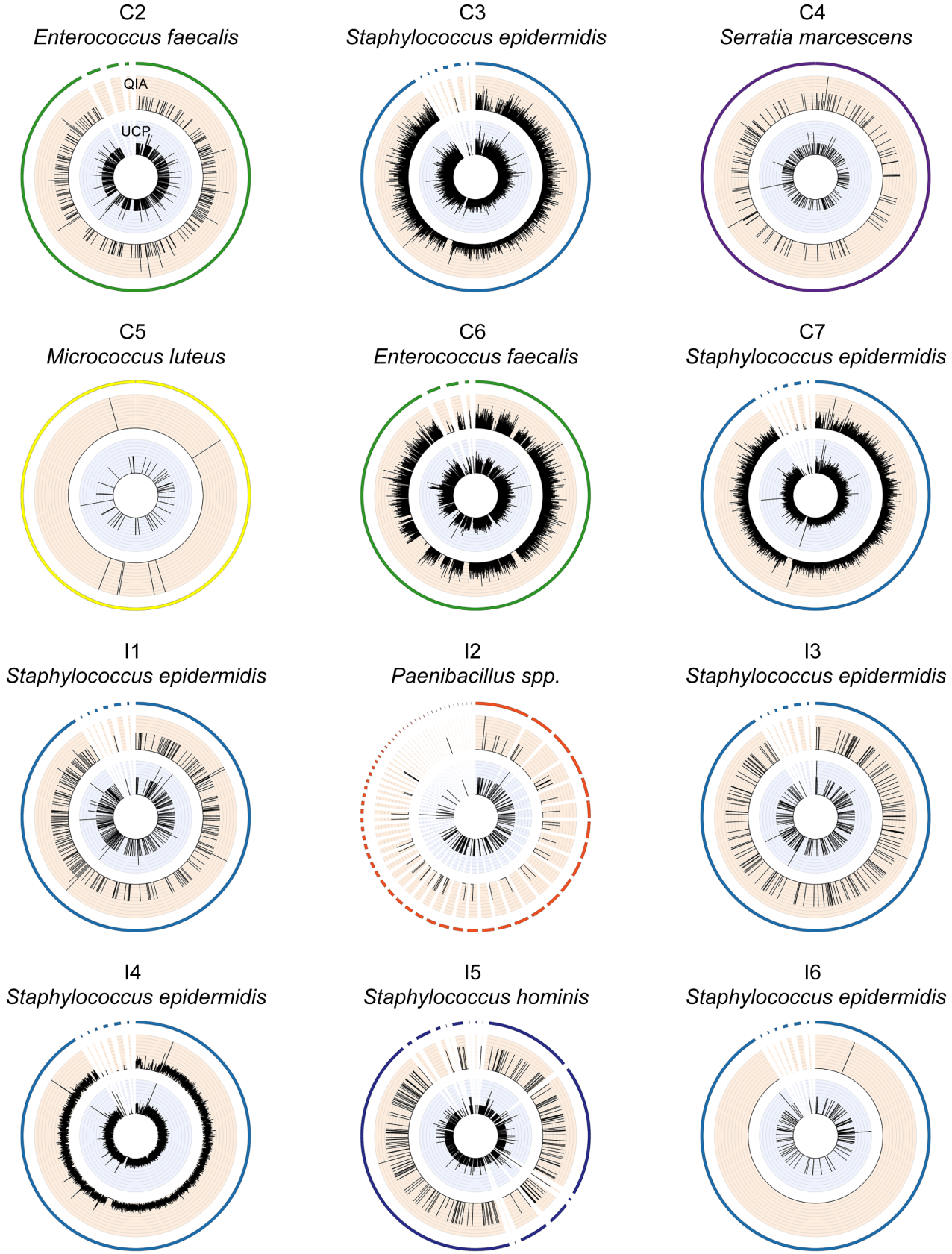

Raw reads
Quality trimming and filtering (BBDuk)
Remove low quality read pairs
Low complexity filtering (BBMask)
Remove low-complexity reads
Map to human reference GRCh38.p10 (BBMap)
Remove human reads
Align to nt database (BLASTN)
Remove human reads
Filter
Human host
DNA sequences
Filter
Ambiguous DNA
sequences from
reference genomes
Curated
Microbial Genome
Database
Taxonomic assignment
(Kraken + Bracken)
verify
Taxonomic assignment
(BLASTN)
Filter
Background
environmental
DNA sequences
Remove QIA specific taxa
Remove UCP specific taxa
Microbiome composition

764
765

766 **Figure 2: Workflow for metagenomic data analysis.** In a first step, sequencing
767 adapters, low quality bases, and reads with low complexity were removed. Subsequently,
768 reads that mapped against the human reference genome sequence, or aligned with
769 human sequences in the nt database were removed. The taxonomic classification of the
770 reads was performed with Kraken together with Bracken using a curated microbial
771 genome database containing 5750 microbial (archaea [251], bacteria [5166], fungi [225],
772 protozoa [73], viruses [35]) and 1 human reference genome sequence (for details, see
773 Supplementary Methods). Additional reads that in this step were classified as human
774 were removed. To verify the classification results, the reads were also aligned to the
775 reference genomes using BLASTn. Organisms specific for the DNA extraction (blank)
776 controls were filtered from the patient samples.

Figure 3: Selected contaminant and non-contaminant organisms based on evaluation of DNA extraction control samples. Contaminant organisms (light grey) were present in higher abundance in DNA extraction controls (green) compared to the endophthalmitis-positive samples (red). The contaminant organisms were detected in similar abundance in the endophthalmitis-negative (yellow) and/or balanced salt solution

784 samples (blue) as in the DNA extraction control samples. Organisms that were detected in
785 higher abundance in patient samples (dark grey), compared to their respective DNA
786 extraction control samples, were not regarded as sample contaminants. Read counts are
787 presented as counts per million in relation to the total non-human read counts per
788 sample, respectively. An interactive version of this figure that includes individual sample
789 information, including read counts, is available from
790 https://figshare.com/s/a4fd9d84260e8456ab72. For a detailed list of contaminant
791 organisms, see Supplementary Table S5.
792


C2
Enterococcus faecalis
QIA
UCP
C3
Staphylococcus epidermidis
C4
Serratia marcescens
C5
Micrococcus luteus
C6
Enterococcus faecalis
C7
Staphylococcus epidermidis
I1
Staphylococcus epidermidis
I2
Paenibacillus spp.
I3
Staphylococcus epidermidis
I4
Staphylococcus epidermidis
I5
Staphylococcus hominis
I6
Staphylococcus epidermidis

795 **Figure 4: Coverage of bacterial reference genomes by metagenomic reads**
796 **originating from intraocular fluid of endophthalmitis patients.** For each individual
797 patient, the metagenomic shotgun reads of the most abundant microbial organism were
798 extracted at genus-level and mapped as unpaired reads using BBmap suite to the
799 respective reference genome sequence in the database. For patients C1 and I7 a
800 particular microbial organism could not be assigned confidently in the metagenomics
801 analysis, and these patients are regarded as "sterile" endophthalmitis cases. The outer
802 most circle displays all sequences of the reference genome (including short contigs and
803 plasmids). The orange and blue inner circles display the depth of mapped reads
804 originating from the vitreous specimens that were extracted with the QIA and UCP DNA
805 extraction methods, respectively. In the two cases where metagenomics analysis and
806 culture results from the hospital were not identical regarding the most abundant organism
807 (patients C5 and I3), we examined the reads via genome mapping for all organisms
808 detected in the metagenomics analysis. The most relevant abundant organism is shown
809 here and the additional plots, as well as information about the maximum read depth for all
810 detected organisms, is available from https://figshare.com/s/c2ce2d32daf25db54904.
811

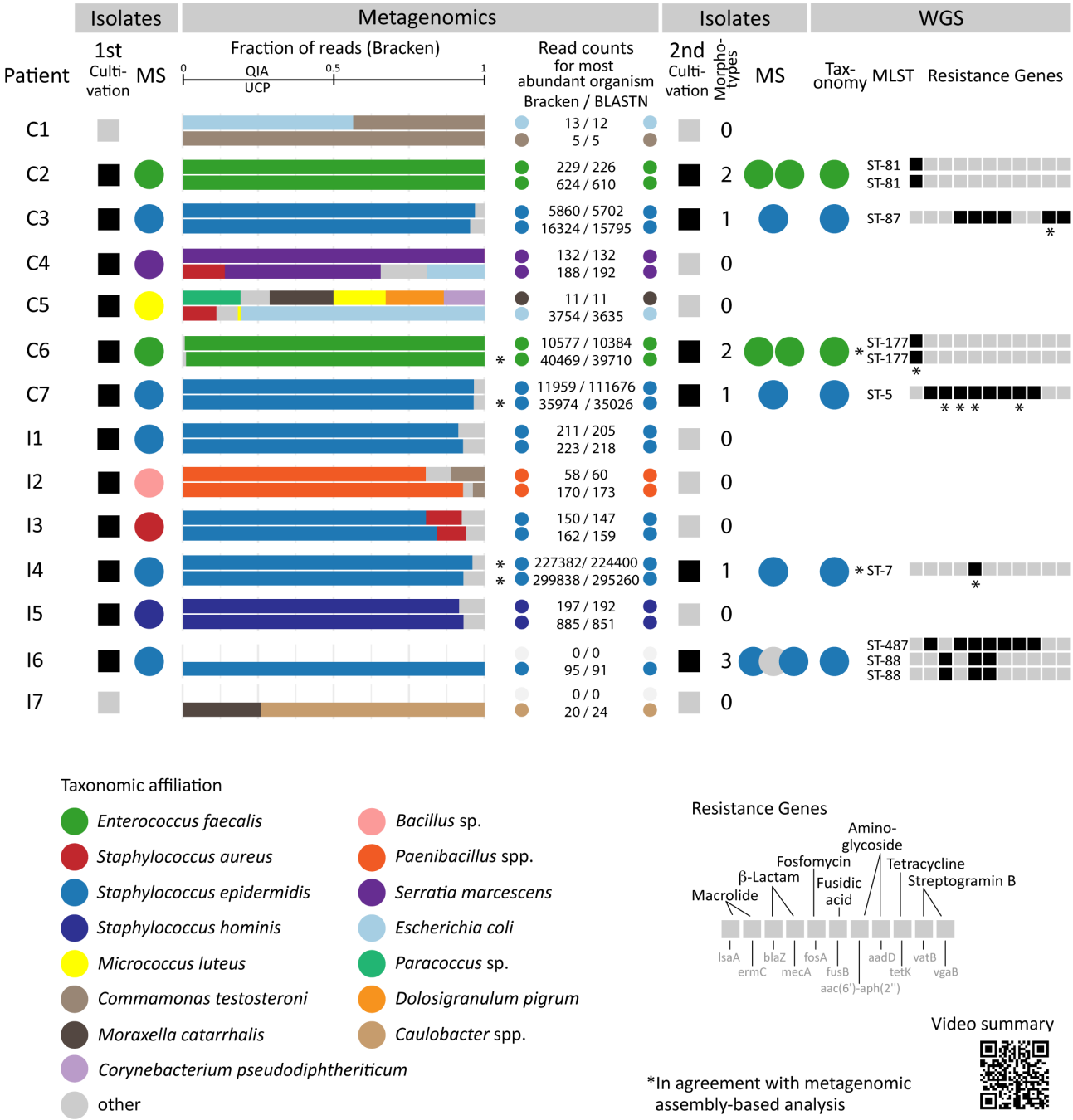


Figure 5: Summary of cultivation-based, metagenomics, and whole genome sequence analyses.
Bacterial isolates were obtained at the hospital laboratory (1st cultivation) from vitreous from endophthalmitis patients following cataract surgery (C1-7) and intravitreal injection (I1-7) and the taxonomic affiliation of the isolates were determined by MALDI-TOF mass spectrometry (MS). Vitreous was analyzed through metagenomics at the research laboratory using two DNA isolation methods (QIAamp DNA Mini Kit, QIA; QIAamp UCP Pathogen Mini kit, UCP) and the taxonomic affiliation of reads was determined. The detected amount of human DNA sequences in percent (%) is provided in the first column of the Metagenomics tab. In the horizontal bar charts, the taxonomic identity and relative

824 fraction of microbial reads for the most abundant identified organisms based on the
825 Kraken+Bracken analysis is indicated for both DNA isolation methods. The read counts
826 for the most abundant organism according to the Kraken+Bracken (all reads) and
827 BLASTN (forward read) analyses are indicated to the right. The read counts for the most
828 abundant organisms per sample as determined by Kraken, Bracken, and BLASTn
829 analyses are available through figshare at https://figshare.com/s/5feabfad1d8c495bf7a3.
830 Bacterial isolates for some samples were obtained in a second round of cultivation at the
831 research laboratory (2nd cultivation), and one representative per colony morphotype per
832 vitreous sample was subjected to MS and whole genome sequencing (WGS). The
833 taxonomic affiliation of isolates was determined through classification of assembled
834 genomes using a k-mer based approach and genomic MLST, and antibiotic resistance
835 genes were identified using ResFinder. Furthermore, metagenomic assemblies were
836 generated from the shotgun metagenomic reads and analyzed with regards to taxonomic
837 affiliation and selected functional characteristics (Supplementary Table S6). A video
838 summary is available from figshare at https://figshare.com/s/38fe043f6a8ef1710444.

839 **Table 1. Ambiguous, including contaminant, sequences in public microbial genomes**
840

	Total		Ambiguous sequences*		
	Genomes	Bases	Genomes	Bases	Bases (%)
Archaea	251	673,145,451	65	1,813,095	0.27
Bacteria	5166	20,854,687,300	2251	75,888,994	0.36
Fungi	225	6,486,874,847	126	6,642,500	0.10
Protozoa	73	2,930,167,033	53	26,447,579	0.84
Viruses	35	640,331	ND	ND	ND
Sum	5750	30,945,514,962	2922	110,858,157	0.35

841 *Genomic sequence regions ≤10 kb (incl. contigs and scaffolds) that had a match (e-value ≤ 1e-6; query
842 coverage ≥ 70%) belonging to a different genus than their stated genus definition when aligned against the
843 non-redundant nucleotide collection (nt) database from NCBI. For more details, see Supplementary Methods
844 and https://figshare.com/s/a282670f1405eae232df, https://figshare.com/s/045b1252bd7555b50ef0,
845 https://figshare.com/s/c42158cdee23f25489cd. ND=not determined