

1 **Unique genetic variants underlie parallel gene expression**
2 **within a young adaptive radiation despite specialization on**
3 **highly divergent resources**

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23 **Abstract**

24 There are many cases of parallel gene expression underlying the evolution of convergent niche
25 specialization, but parallel expression could also underlie divergent specialization. We
26 investigated divergence in gene expression and genetic variation across three sympatric
27 *Cyprinodon* pupfishes endemic to San Salvador Island, Bahamas. This recent radiation consists
28 of a generalist and two derived specialists adapted to novel niches – a ‘scale-eater’ and a ‘snail-
29 eater.’ We sampled total mRNA from all three species at two early developmental stages and
30 compared gene expression with whole-genome genetic differentiation between all three species.
31 82% of genes that were differentially expressed between snail-eaters and generalists were up or
32 downregulated in the same direction between scale-eaters and generalists; however, there were
33 no shared fixed variants underlying this parallel expression. These genes showing parallel
34 expression did not exhibit increased developmental constraints, but were enriched for effects on
35 metabolic processes. Alternatively, genes showing divergent expression were enriched for effects
36 on cranial skeleton development and pigment biosynthesis, reflecting the most divergent
37 phenotypes observed between specialist species. Our findings reveal that convergent adaptation
38 to higher trophic levels between divergent niche specialists through shared genetic pathways is
39 governed by unique genetic variants.

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50 **Background**

51 Abundant research on the genetic basis of adaptive traits has revealed an overarching pattern in
52 nature – when species are faced with similar selective pressures, they often respond with the
53 same genetic solutions (1). Parallel gene expression underlying convergent traits is a well-
54 documented evolutionary phenomenon, with examples from experimental evolution studies
55 imposing uniform selection pressures on replicate populations (2,3), studies in natural systems
56 between closely related taxa (4–10), and distantly related taxa (11–13). This work has shown that
57 parallelism at the level of gene expression is common in many cases of phenotypic convergence,
58 particularly when divergence time between species is short (1,14).

59 However, few studies have investigated the extent of parallel gene expression
60 contributing to species divergence, largely because most expression studies focus on only two
61 species and are either concerned with divergent expression giving rise to divergent phenotypes
62 (15–17) or parallel expression of specific loci (11,18,19) (but see 19,20). Furthermore, while
63 there many genetic and demographic factors thought to influence the probability of parallel
64 evolution (22,23), there are no theoretical expectations for the amount of parallel genetic
65 variation contributing to parallel gene expression during ecological speciation (24,25).

66 Here we ask how both parallel and divergent expression patterns influence novel
67 phenotypes by measuring transcriptomic and genomic divergence between three sympatric
68 species of *Cyprinodon* pupfishes endemic to hypersaline lakes on San Salvador Island, Bahamas.
69 This recent radiation consists of a dietary generalist species (*C. variegatus*) and two novel
70 specialists: a ‘snail-eater’ (*C. brontotheroides*) and a ‘scale-eater’ (*C. desquamator*). Scale-eaters
71 have large jaws and elongated bodies, whereas snail-eaters have short, thick jaws and a
72 protruding nasal region that may function in crushing hard-shelled mollusks. These specialized
73 species have evolved within the past 10,000 years based on the most recent drying of San
74 Salvador lakes (26,27) and occupy unique niches within atherinomorph fishes (28). Scale-eaters
75 and snail-eaters most likely diverged from a generalist common ancestor based on phylogenetic
76 analyses of outgroup species and surveys of pupfish populations on neighboring Bahamian
77 islands (28,29).

78 We performed total mRNA sequencing to examine gene expression in lab-reared
79 individuals of all three species from different lake populations at two developmental stages. We

80 also searched for SNPs unique to each specialist and determined whether fixed variants near
81 differentially expressed genes showed signs of hard selective sweeps (30). We found divergent
82 genetic variation underlying trophic specialization, yet significant parallelism at the level of gene
83 expression in specialists. We tested whether this counterintuitive result of parallel expression
84 between divergent species may be due to 1) increased developmental constraint for genes
85 showing parallelism or that 2) specialists experience parallel selective environments and adapt to
86 higher trophic levels using similar genetic pathways.

87

88 **Materials and Methods**

89 *Study system and sample collection*

90 Individuals were caught from hypersaline lakes on San Salvador Island, Bahamas using a hand
91 net or seine net in 2011, 2013, and 2015. Whole genome resequencing was performed for wild-
92 caught individuals from a total of nine isolated lakes on San Salvador (Great Lake, Stout's Lake,
93 Oyster Lake, Little Lake, Crescent Pond, Moon Rock, Mermaid's Pond, Osprey Lake, and
94 Pigeon Creek). 14 scale-eaters were sampled from six populations; 11 snail-eaters were sampled
95 from four populations; and 13 generalists were sampled from eight populations on San Salvador.
96 Outgroup samples included one *C. laciniatus* from Lake Cunningham, New Providence Island,
97 Bahamas, one *C. bondi* from Etang Saumautre lake in the Dominican Republic, one *C. diabolis*
98 from Devil's Hole in California, and captive-bred individuals of *C. simus* and *C. maya* from
99 Laguna Chicancanab, Quintana Roo, Mexico. Sampling is further described in (31,32) Fish were
100 euthanized in an overdose of buffered MS-222 (Finquel, Inc.) following approved protocols from
101 the University of California, Davis Institutional Animal Care and Use Committee (#17455) and
102 University of California, Berkeley Animal Care and Use Committee (AUP-2015-01-7053) and
103 stored in 95-100% ethanol.

104

105 *RNA sequencing and alignment*

106 Juvenile pupfish were derived from individuals raised in a common laboratory environment (25-
107 27 C, 10-15 ppt salinity, pH 8.3) with identical diets. F₁ and F₂ fry were collected at two

108 developmental stages after hatching: 8-10 days post-fertilization and 17-20 dpf. They were
109 euthanized in an overdose of buffered MS-222, and stored in RNA later (Ambion, Inc.) at 4 C for
110 one day, followed by long-term storage at -20 C for up to one year. We extracted whole-body
111 RNA using RNeasy kits (Qiagen) from 15 larvae (8-10 dpf) (Three generalists and snail-eaters
112 from Crescent Pond and Little Lake and three scale-eaters from Crescent Pond; Table S1). We
113 also dissected 14 larvae (17-20 dpf) to isolate RNA from the anterior craniofacial region
114 containing the dentary, angular articular, maxilla, premaxilla, palatine, and associated
115 craniofacial connective tissues using fine-tipped tweezers washed with RNAase-away (Three
116 generalists and snail-eaters from Crescent Pond and Little Lake and two scale-eaters from
117 Crescent Pond; Table S1). Our decision to sample RNA at 8-10 dpf and 17-20 dpf affects how
118 we interpret patterns of gene expression. We chose to sample a range of days within each
119 developmental stage to increase noise, thereby increasing the chance of identifying
120 transcriptional variation that robustly influences larval development.

121 Libraries were prepared using the KAPA stranded mRNA-seq kit (KAPA Biosystems
122 2016) at the High Throughput Genomic Sequencing Facility at UNC Chapel Hill. Stranded
123 sequencing on one lane of Illumina 150PE Hiseq4000 resulted in 677 million raw reads. We
124 filtered raw reads using Trim Galore (v. 4.4, Babraham Bioinformatics) to remove Illumina
125 adaptors and low-quality reads (mean Phred score < 20). We mapped these reads to the
126 *Cyprinodon* reference genome using the RNA-seq aligner STAR (v. 2.5 (33)). We used the
127 featureCounts function of the Rsubread package (34) requiring paired-end and reverse stranded
128 options to generate read counts across previously annotated features. We assessed mapping and
129 count quality using MULTIQC (35).

130

131 *Differential expression analyses*

132 We quantified differences in gene expression between all three species at two developmental
133 stages. Our raw counts determined by featureCounts were normalized and used to identify
134 differentially expressed transcripts with DEseq2 (v. 3.5 (36)). We performed four pairwise tests
135 pooling species across lakes to identify differentially expressed transcripts among generalists vs.
136 snail-eaters and generalists vs. scale-eaters at 8-10 dpf and 17-20 dpf (Table. S1). Genes with
137 fewer than two read counts were discarded from all analyses (n = 1,570), along with genes

138 showing low normalized counts at a threshold determined by DEseq2 (using Cook's distance,
139 0.99 quantile (36)). Wald tests determined significant differences in expression between species
140 by comparing normalized posterior log fold change estimates and correcting for multiple testing
141 using the Benjamini–Hochberg procedure with a false discovery rate of 0.05 (37).

142 We used permutation tests to determine whether specialist species exhibited nonrandom
143 patterns of parallel gene expression. We performed 100,000 permutations randomly sampling
144 transcript IDs from the combined number of transcripts differentially expressed between
145 generalists and each specialist followed by randomly sampling transcripts as either up or
146 downregulated relative to generalists.

147

148 *Genomic variant discovery and population genetic analyses*

149 SNP variants were called using previously outlined methods (31,32). Briefly, 42 individual DNA
150 samples extracted from muscle tissue were fragmented, barcoded with Illumina indices, and
151 quality checked using a Fragment Analyzer (Advanced Analytical Technologies, Inc.).
152 Sequencing on four lanes of Illumina 150PE Hiseq4000 resulted in 2.8 billion raw reads that
153 were mapped from 42 individuals to the *Cyprinodon* reference genome (NCBI, *C. variegatus*
154 Annotation Release 100, total sequence length = 1,035,184,475; number of scaffold = 9,259,
155 scaffold N50, = 835,301; contig N50 = 20,803; (38)). We followed Genome Analysis Toolkit (v
156 3.5) best practices and hard filter criteria to call and refine our SNP variant dataset (QD < 2.0; FS
157 < 60; MQRankSum < -12.5; ReadPosRankSum < -8 (39)). Our final SNP dataset contained 16
158 million variants and a mean sequencing coverage of 7× per individual (range: 5.2–9.3×).

159 We identified SNPs that were fixed in each specialist species. We calculated genome
160 wide F_{st} using VCFtools' 'weir-fst-pop' function for two different population comparisons
161 involving samples collected from San Salvador: generalists (n = 13) vs. snail-eaters (n = 11) and
162 generalists (n = 13) vs. scale-eaters (n = 9). Our SNP dataset included 14 scale-eaters, however,
163 we split our scale-eater population into two groups (large-jawed scale-eaters, n = 9 and small-
164 jawed scale-eaters, n = 5) based on previous evidence that these two populations are genetically
165 distinct (31,32). This allowed us to identify SNPs unique to large-jawed scale-eaters (i.e. *C.*
166 *desquamator* (40)), which were the only type of scale-eater we sampled for RNA-seq. We

167 identified which of these SNPs resided in gene regions (either exonic, intronic, or within 10kb of
168 the first or last exon) for genes showing differential expression. We determined whether these
169 regions showed signatures of hard selective sweeps using SweeD ((30); methods previously
170 described in (31)). Briefly, SweeD sections scaffolds into 1000 windows of equal size and
171 calculates a composite likelihood ratio (CLR) using a null model where the site frequency
172 spectrum of each window does not differ from that of the entire scaffold. Various demographic
173 histories can shift the neutral site frequency spectrum making it difficult to infer signatures of
174 selection (41,42). We previously estimated ancestral effective population sizes of San Salvador
175 pupfishes using MSMC (31,43) and used these estimates to correct the expected neutral site
176 frequency spectrum for the inferred recent population bottleneck in Caribbean pupfishes using
177 SweeD. Windows with fixed SNPs that showed CLR_s above the 95th percentile across their
178 respective scaffolds (>10,000bp) under the assumptions of a population decrease determined by
179 MSMC were interpreted as regions that recently experienced a hard sweep.

180

181 ***Measuring evolutionary constraint for differentially expressed genes***

182 We performed gene ontology (GO) enrichment analyses for differentially expressed genes using
183 GO Consortium resources available at geneontology.org (44). We used BlastP (v. 2.6 (45)) to
184 identify zebrafish protein orthologs with high similarity (E -value < 1) to NCBI protein
185 accessions for genes that we identified as differentially expressed between *Cyprinodon* species.
186 We used these orthologs to determine if any gene ontology categories were enriched across
187 differentially expressed genes. We grouped enriched GO categories into similar representative
188 terms using the REVIGO clustering algorithm (46).

189 We also used these orthologs to estimate pleiotropy for differentially expressed genes
190 based on the number of associated GO biological processes, protein-protein interactions (PPIs),
191 and developmental stages when they are known to be expressed (47). We again used the GO
192 Consortium (44) to determine the number of biological processes associated with each gene. We
193 examined biological process annotations only for genes from ZFIN (zfin.org) with experimental
194 evidence (GO evidence code EXP). The String protein database (v. 10; (48)) calculates a
195 combined score measuring confidence in protein interactions by considering known interactions
196 (experimentally determined and from manually curated databases) and predicted interactions.

197 We used the String database to quantify PPIs for protein products of differentially expressed
198 genes, focusing only on interactions with experimental evidence (i.e. non-zero experimental
199 evidence scores).

200 Next, we determined the number of developmental stages where a gene is known to be
201 expressed using the Bgee expression call database for zebrafish (v. 14.0 (49)). We considered
202 eight developmental stages from larval day five to juvenile day 89 from the Zebrafish Stage
203 Ontology (ZFS) that were deemed ‘gold quality,’ meaning there was no contradicting call of
204 absence of expression for the same gene, in the same developmental stage (49). We tested
205 whether levels of gene pleiotropy were different in genes showing parallel expression in
206 specialists versus divergently expressed genes by fitting a generalized linear model (negative
207 binomial family; *glm.nb* function in the R library “MASS”) on count data for pleiotropy
208 estimates. We did not measure pleiotropy for genes expressed at 17-20 dpf due to the low
209 number of zebrafish orthologs matched for genes with parallel expression in craniofacial tissues
210 (11 out of 23).

211 Finally, we used the duplicated genes database (50) to identify paralogs in our
212 differentially expressed gene dataset. We calculated whether genes showing parallel expression
213 in specialists had more paralogs than divergently expressed genes using Pearson’s Chi-square
214 test in R.

215

216 **Results**

217 *Differential expression between generalists and specialists*

218 Total mRNA sequencing across all 29 samples resulted in 677 million raw reads, which was
219 reduced to 674 million reads after quality control and filtering. 81.2% of these reads successfully
220 aligned to the reference genome and 75.5% of aligned reads mapped to annotated features with
221 an average read depth of 309× per sample. We used this dataset to identify differences in gene
222 expression associated with specialist phenotypes at two developmental stages and within two
223 groups of tissues.

224 We compared expression across 22,183 genes that showed greater than two read counts
225 out 24,383 total genes annotated in the *Cyprinodon variegatus* assembly (NCBI, *C. variegatus*
226 Annotation Release 100, (38)). These analyses revealed 1,014 genes (1,119 isoforms)
227 differentially expressed between generalists vs. snail-eaters and 5,982 genes (6,776 isoforms)
228 differentially expressed between generalists vs. scale-eaters (Fig. 1) (Benjamini and Hochberg
229 adjusted $P < 0.05$). 833 genes (923 isoforms) showed the same direction of expression (up or
230 down regulated) in both specialists relative to generalists, indicating parallel expression in
231 specialists (Fig. 2). Specifically, 497 differentially expressed isoforms showed lower expression
232 in both specialist species compared to generalists, while 424 showed higher expression in
233 specialists (Fig. S1). This is significantly more parallel expression than would be expected by
234 chance for genes ($n = 6,996$; 100,000 permutations; $P < 1.0 \times 10^{-5}$) and gene isoforms ($n =$
235 $8,998$; 100,000 permutations; $P < 7.0 \times 10^{-4}$) (Fig. S2).

236 Craniofacial morphology is the most rapidly diversifying trait in the San Salvador
237 radiation (51). In order to detect genes expressed during jaw development, we compared
238 expression within craniofacial tissue at the 17-20 dpf stage. We discovered 120 genes (123
239 isoforms) differentially expressed between generalists vs. snail-eaters and 1,903 genes (2,222
240 isoforms) differentially expressed between generalists vs. scale-eaters (Benjamini and Hochberg
241 adjusted $P < 0.05$). We observed a lower proportion of parallel expression in specialists for
242 craniofacial tissue compared to whole-body tissue. 23 genes showed the same direction of
243 expression in specialists relative to generalists, with 11 differentially expressed isoforms
244 showing lower expression in both specialist species and 14 showing higher expression. This is
245 significantly less parallel expression than would be expected by chance for genes ($n = 2,023$;
246 100,000 permutations; $P < 1.0 \times 10^{-5}$) and gene isoforms ($n = 2,345$; 100,000 permutations; $P <$
247 1.0×10^{-5}) (Fig. S2).

248 We also identified 4 genes that showed opposite patterns of expression in specialists
249 relative to generalists (Table S2). In 8-10 dpf whole-body tissue, 2 genes (*agxt2*, *si:ch211-*
250 *197h24.9*) were upregulated in snail-eaters and downregulated in scale-eaters, while one gene
251 (*plin2*) was upregulated in scale-eaters and downregulated in snail-eaters. In 17-20 dpf
252 craniofacial tissue, one gene (*mybpc2a*) was upregulated in snail-eaters and downregulated in
253 scale-eaters.

254

255 ***Parallel expression does not share the same genetic basis***

256 Similar to previous work on a smaller sample size (31), we identified 79 SNPs fixed between
257 generalists and snail-eaters and 1,543 SNPs fixed between generalists and scale-eaters. None of
258 these fixed variants were shared between specialists. Next, we determined which of these fixed
259 SNPs fell within gene regions (either exonic, intronic, or within 10kb of the first or last exon).
260 29% of SNPs fixed in snail-eaters overlapped with 21 gene regions, while 59% of SNPs fixed in
261 scale-eaters overlapped with 245 gene regions. We found 365 SNPs fixed in scale-eaters (24%)
262 within 68 gene regions that showed differential expression between generalists and scale-eaters
263 in whole-body tissue (8-10 dpf) and 123 SNPs (8%) within 26 gene regions differentially
264 expressed between generalists and scale-eaters in craniofacial tissue (17-20 dpf). We suspect that
265 some of these fixed variants are causal cis-regulatory variants responsible for species specific
266 expression patterns that ultimately give rise to phenotypic differences in scale-eaters.
267 Conversely, we only identified a single SNP fixed in snail-eaters within a gene (*tmprss2*) that
268 was differentially expressed between generalists and snail-eaters in whole-body tissue (8-10 dpf).
269 We did not find any fixed variants near genes differentially expressed between generalists and
270 snail-eaters in craniofacial tissue, possibly suggesting that fixed variants regulate expression
271 divergence at an earlier developmental.

272 Since we did not find any variants that were fixed between scale-eaters and generalists
273 that were also fixed between snail-eaters and generalists, we repeated our analysis with a lower
274 threshold of genetic divergence by examining SNPs with $F_{st} \geq 0.8$ between generalists *vs.* snail-
275 eaters and generalists *vs.* scale-eaters. Again, none of these SNPs were shared between
276 specialists. We found 11,491 divergent SNPs within 810 gene regions differentially expressed
277 between scale-eaters and generalists. We also found 297 SNPs within 27 gene regions
278 differentially expressed between snail-eaters and generalists. Strikingly, only three genes out of
279 833 that showed parallel expression in specialists contained SNPs with $F_{st} \geq 0.8$ between
280 generalists and each of the specialists. This suggests that parallel expression is not a result of the
281 same genetic variation in specialists.

282

283 ***Pleiotropic constraint and functional redundancy do not explain parallel expression***

284 We predicted that genes showing parallel expression patterns in specialists may be constrained
285 by negative pleiotropy. High gene pleiotropy is correlated with participation in more protein-
286 protein interactions (PPIs), which in turn effects multiple biological processes (52,53). Genes
287 that act across multiple developmental stages are also more pleiotropic (54). We estimated
288 pleiotropy for differentially expressed genes based on the number of protein-protein interactions
289 (PPIs), associated GO biological processes, and developmental stages when they are known to be
290 expressed. However, we did not find evidence for higher pleiotropy for genes showing parallel
291 expression patterns in specialists compared to genes showing divergent expression using any of
292 these three metrics (GLM; biological processes: $P = 0.67$; PPIs: $P = 0.09$; developmental stages:
293 $P = 0.89$) (Fig. 3).

294 Next, we predicted that genes showing divergent expression patterns in specialists might
295 be under less developmental constraint because they share redundant functions with paralogous
296 genes. We searched the duplicated genes database (50) and found that datasets for genes with
297 divergent expression patterns in specialists showed similar numbers of paralogs to genes with
298 parallel expression patterns in specialists (χ^2 , 8-10 dpf: 48 duplicated genes out of 627 genes
299 showing parallel expression and 270 duplicated genes out of 3,491 divergently expressed genes,
300 $P = 0.95$; 17-20 dpf: 2 duplicated genes out of 11 genes showing parallel expression and 41
301 duplicated genes out of 663 divergently expressed genes, $P = 0.11$).

302

303 ***Genes showing parallel expression are enriched for metabolic processes***

304 We performed GO enrichment analyses with zebrafish orthologs for genes showing parallel
305 expression patterns in specialists ($n = 620$) and genes showing divergent expression patterns in
306 scale eaters ($n = 3,349$) and snail-eaters ($n = 102$). We restricted these analyses to genes
307 expressed at 8-10 dpf because the number of genes showing parallel expression in specialists at
308 17-20 dpf ($n = 23$) was low and did not show enrichment for any biological process.

309 Genes showing parallel expression in trophic specialists were enriched for metabolic
310 processes (20% of GO terms). In contrast, genes with divergent expression patterns in specialists
311 were enriched for cranial skeletal development and pigment biosynthesis (7% and 3% of terms,

312 respectively) while only 11% of enriched categories described metabolic processes (Fig. 4, Table
313 S3). We identified genes with divergent expression between specialists enriched for effects on
314 viscerocranium, striated muscle, liver, erythrocyte, and the development of the heart, optic cup,
315 pancreas, and brain. 41 divergently expressed genes were annotated for viscerocranium
316 development, which is particularly interesting because this tissue gives rise to the most divergent
317 craniofacial structures in specialists (55,56).

318

319 ***The genetic basis of extreme craniofacial divergence***

320 We previously described 30 candidate gene regions containing variants fixed between trophic
321 specialist species associated with variation in jaw length (31). These candidates also showed
322 signatures of a recent hard selective sweep (31). Encouragingly, we found eleven of these genes
323 differentially expressed between generalists and specialists (seven at 8-10 dpf and four at 17-20
324 dpf) (Table S4).

325 We searched for variants fixed in specialists within gene regions across all 7,394 genes
326 that were differentially expressed between either generalists and snail-eaters or generalists and
327 scale-eaters. We discovered fixed SNPs in 81 of these gene regions (either intronic or within
328 10kb of the first or last exon), potentially implicating cis-acting variants regulating gene
329 expression. Further testing this hypothesis, we searched for signatures of hard selective sweeps
330 in specialists. Interestingly, 95% of these gene regions containing fixed SNPs showed signs of
331 experiencing a selective sweep (estimated by SweeD; CLR > 95th percentile across their
332 respective scaffolds) (Table S5). All of these genes regions contained SNPs fixed between
333 generalists and scale-eaters and showed differential expression in this same comparison. Five of
334 these 81 genes were indicated as strong craniofacial candidates in our previous study (31).
335 Finally, we compared this list of genes experiencing selection to those annotated for cranial
336 skeletal system development (GO:1904888) and muscle organ development (GO:0007517). This
337 revealed three genes containing fixed variation in scale-eaters that likely influence craniofacial
338 divergence through cis-acting regulatory mechanisms (*lox13b* (annotated for cranial effects);
339 *fbxo32* and *klhl40a* (annotated for muscle effects)).

340

341 **Discussion**

342 We combined RNA sequencing with genome-wide divergence scans to study the molecular
343 evolution of two trophic specialist species that rapidly diverged from a generalist common
344 ancestor within the last 10,000 years. We examined how gene expression and SNP variation
345 influence snail-eater and scale-eater niche adaptations using comparisons between each specialist
346 and their generalist sister species. We found a significant amount of parallelism at the level of
347 gene expression yet no parallelism at the level of fixed genetic variation in specialists.
348 Specifically, 82% of genes that were differentially expressed between snail-eaters and generalists
349 were up or downregulated in the same direction when comparing expression between scale-
350 eaters and generalists (Fig. 2). We show that this parallel expression is not the result of the same
351 underlying genetic variation.

352 We explored two possible explanations for this pattern: 1) developmental constraints
353 resulting from pleiotropy or functional redundancy favor parallel expression in specialist species
354 or 2) parallel selective environments act on similar genetic pathways needed for adaptation to
355 higher trophic levels.

356

357 ***1. Developmental constraints on the evolution of parallel gene expression***

358 We tested two hypotheses considering how different forms of developmental constraint could
359 promote parallel expression patterns in specialist species. One mechanism constraining the
360 probability of parallelism is pleiotropy – when the expression of one transcript influences
361 multiple phenotypic characteristics (22,47,57). Higher gene pleiotropy is correlated with
362 participation in more protein-protein interactions (PPIs), which can cause cascading downstream
363 effects on multiple biological processes (52,53). We predicted that genes showing parallel
364 expression might be under higher constraint due to negative pleiotropy. We estimated three
365 measures of gene pleiotropy (number of associated GO biological processes, protein-protein
366 interactions (PPIs), and developmental stages when they are known to be expressed) and found
367 no significant difference in any measure for genes showing parallel versus divergent expression
368 patterns in specialists (Fig. 3). This finding is consistent with some empirical evidence and

369 theoretical models of gene expression evolution that found pleiotropy constrains the variability
370 of gene expression within species, but does not hinder divergence between species (15,58).

371 Gene sequence similarity can also affect the probability of parallelism (22). Genes that
372 are similar due to duplication events (paralogs) may be less constrained and therefore more likely
373 to exhibit divergent expression (7,22,59–61). Given that some paralogs are known to share
374 molecular functions even when they have had much time to diverge (62–64), we predicted that
375 the set of genes showing divergent expression might be under less constraint because it was
376 enriched for paralogs exhibiting functional redundancy. However, we did not find any difference
377 in the proportion of paralogs for genes showing parallel and divergent expression in specialists
378 (χ^2 ; 8-10 dpf: $P = 0.95$; 17-20 dpf $P = 0.11$). Overall, we find that parallel expression in
379 specialists is not the result of increased developmental constraints.

380

381 ***2. Parallel gene expression underlies convergent metabolic adaptations to a higher trophic*** 382 ***level in each specialist***

383 While the specialists are more morphologically diverged from one another than either is from
384 generalist species, particularly in their craniofacial phenotype and male reproductive coloration
385 (40,65) (Fig. 4), dietary isotope analyses show that they occupy a higher trophic level than
386 generalists (66). Fish scales and mollusks contribute to more nitrogen-rich diets in specialists
387 compared to generalist species that primarily consume algae and detritus (66). Perhaps the same
388 metabolic processes required for this type of diet are adaptive at higher trophic levels for both
389 scale-eaters and snail-eaters, which might explain patterns of parallel expression. Thus, we
390 predicted that genes showing parallel expression would affect metabolic processes that may be
391 similar between specialists, whereas genes showing divergent expression in specialists would
392 affect developmental processes.

393 GO enrichment analyses support both hypotheses. We found that 20% of GO categories
394 enriched for genes showing parallel expression described metabolic processes, and zero
395 described cranial skeletal development or pigment biosynthesis (Fig. 4A). In contrast, 10% of
396 terms showing enrichment in the divergently expressed gene set described developmental
397 processes (cranial skeletal development and pigment biosynthesis) and only 11% described

398 metabolic processes (Fig 4A, Table S4). These results may suggest that parallel expression in
399 specialists is important for adapting to a higher trophic level, in which snail-eating and scale-
400 eating present similar metabolic requirements relative to the lower trophic level of generalists.
401 This is consistent with the high macroalgae content of generalist diets relative to both specialist
402 species (51) and the shorter intestinal lengths observed in both specialists relative to the
403 generalist (CHM and JAM personal observation). In contrast, genes showing divergent
404 expression in specialists are more important for shaping divergent skeletal and pigmentation
405 phenotypes between species (Fig. 4B, C and D).

406

407 ***Parallel gene expression despite divergent genetic variation***

408 We find significant parallel gene expression across genes that are annotated for effects on
409 metabolism, yet shared expression patterns do not seem to be driven by the same fixed variants.
410 This is surprising in this young radiation given that the probability of parallel genetic variation
411 underlying phenotypic convergence increases with decreasing divergence time (1,24,67).
412 Although 95% of differentially expressed gene regions containing fixed SNPs show signs of
413 experiencing a selective sweep, and almost none of these variants in these SNPs were in exons, it
414 is still possible that fixed alleles do not regulate parallel expression of metabolic genes. Instead,
415 we reasoned that segregating variants shared between specialists might be responsible for shared
416 expression patterns. However, we still failed to find any shared variants at lower levels of
417 differentiation ($F_{st} \geq 0.8$). Another possibility is that parallel expression is not controlled by
418 shared SNPs, but rather a more complex type of genetic variation (copy number variants,
419 inversions, insertions, deletions). However, preliminary results suggest that there are no shared
420 insertion/deletions between specialists (JAM unpublished data). Many studies on parallel
421 adaptation through gene reuse describe convergence within pigmentation and skeletal
422 development pathways (6,18,23,68). Perhaps the architecture of metabolic adaptation is more
423 flexible, having more mutational targets or employing more late-acting developmental regulatory
424 networks that are less constrained than early-acting networks (67,69–73). Our findings highlight
425 the importance of understanding the probability of genetic convergence across different
426 biological processes.

427

428 *Caveats to gene expression analyses and the robustness of parallel expression*

429 We chose to sample RNA at 8-10 dpf and 17-20 dpf to identify transcriptional variation that
430 influences larval development, however, some activation of parallel gene networks is likely
431 specified at pre-hatching developmental stages (69,70). It is also possible that we did not have
432 the power to identify subtle differences in expression for genes that showed high divergence
433 between specialists and generalists. Detecting differential expression of transcripts is notoriously
434 difficult when read counts are low and variance within treatment groups is high (74,75). We
435 were able to detect differential expression for genes with a mean normalized count as low as 1.6
436 (median = 150) and \log_2 fold change as low as 0.2 (median = 1.11). Thus, we could not detect
437 any variation causing very slight fold changes in expression below 0.2. Furthermore, our scale-
438 eater sample sizes (8-10 dpf $n = 3$; 17-20 dpf $n = 2$) were lower than that of generalists and snail-
439 eaters ($n = 6$ at both stages) (Fig. S1), which likely resulted in more false positives for scale-eater
440 comparisons.

441 Finally, our results are robust in a recently published independent analysis of gene
442 expression in San Salvador pupfishes that identified many of the same genes we found
443 divergently expressed between specialists (38). We examined this dataset using the same
444 significance thresholds for differentially expressed genes as described in Lencer *et al.* for mRNA
445 extracted from all three species at 8 dpf and 15dpf ($p < 0.1$ and $|\text{Log}_2 \text{ fold change}| > 0.2$). We
446 found that 40% of genes divergently expressed between specialists in this dataset were
447 divergently expressed in our own dataset. Importantly, Lencer *et al.* only examined cranial
448 tissues at both of these developmental stages, and they did not investigate parallel expression
449 between each specialist. Next we searched for evidence of parallel expression for mRNA
450 extracted from all three species at 8 dpf. 28.8% of genes that were differentially expressed
451 between snail-eaters and generalists were up or downregulated in the same direction between
452 scale-eaters and generalists. This is a lower proportion of parallel expression than we identified
453 (Fig. 2), but this is most likely because Lencer *et al.* only sampled RNA from cranial tissues
454 (38). Thus, the majority of parallel expression between specialists likely occurs in non-cranial
455 tissues, consistent with our shared metabolic hypothesis.

456

457 *Conclusion*

458 Here we find that divergent genetic variation underlies significant parallel gene expression in
459 two novel specialist species. While there are many cases of parallel expression underlying
460 parallel ecological specialization, to our knowledge, this represents the first case of parallel
461 expression accompanying divergent specialization. Our findings will be useful in developing
462 theoretical predictions of parallelism contributing to species divergence at both the gene and
463 nucleotide level. Numerous studies have shown that the probability of parallel genetic variation
464 underlying phenotypic convergence is higher when divergence time between species is short
465 (1,24,67). Scale-eating and snail-eating species have evolved rapidly within the last 10,000
466 years, yet parallel expression patterns are not driven by the same underlying genetic variants,
467 arguing that some forms of phenotypic convergence between young species do not exhibit
468 parallelism at the nucleotide level. We show that parallel expression is not a result of
469 developmental constraint imposed by negative pleiotropy or functional redundancy, but most
470 likely reveals convergent adaptation to a higher trophic level in each trophic specialist, despite
471 their specialization on highly divergent resources.

472

473 **Competing interests**

474 We declare we have no competing interests.

475

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479

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486 associated with BioProject PRJNA391309.

487 **Author Contributions**

488 JAM wrote the manuscript. JAM extracted the RNA samples, and conducted all bioinformatic
489 and population genetic analyses. Both authors contributed to the conception and development of
490 the ideas and revision of the manuscript.

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510 **References**

- 511 1. Conte GL, Arnegard ME, Peichel CL, Schluter D. The probability of genetic parallelism
512 and convergence in natural populations. 2012;(October):5039–47.
- 513 2. Cooper TF, Rozen DE, Lenski RE. Parallel changes in gene expression after 20 , 000
514 generations of evolution in *Escherichia coli*. 2003;2002(Track II).
- 515 3. Riehle MM, Bennett AF, Lenski RE, Long AD, Michelle M, Bennett AF, et al.
516 Evolutionary changes in heat-inducible gene expression in lines of *Escherichia coli*
517 adapted to high temperature. 2003;47–58.
- 518 4. Derome N, Bernatchez L. The Transcriptomics of Ecological Convergence between 2
519 Limnetic Coregonine Fishes (*Salmonidae*). 2004;
- 520 5. Chan YF, Marks ME, Jones FC, Jr GV, Shapiro MD, Brady SD, et al. Adaptive Evolution
521 of Pelvic Reduction of a *Pitx1* Enhancer. 2010;327(January):302–6.
- 522 6. Reed RD, Papa R, Martin A, Hines HM, Kronforst MR, Chen R, et al. optix Drives the
523 Repeated. 2011;(August).
- 524 7. Nagai H, Terai Y, Sugawara T, Imai H, Nishihara H, Hori M, et al. Reverse Evolution in
525 RH1 for Adaptation of Cichlids to Water Depth in Lake Tanganyika Research article
526 Determination of Sequences Upstream. 2011;28(6):1769–76.
- 527 8. Manousaki T, Hull PM, Kusche H, Machado-schiaffino G, Franchini P, Harrod C. Parsing
528 parallel evolution □: ecological divergence and differential gene expression in the adaptive
529 radiations of thick-lipped Midas cichlid fishes from Nicaragua. 2013;22:650–69.
- 530 9. Reid NM, Proestou DA, Clark BW, Warren WC, Colbourne JK, Shaw JR, et al. The
531 genomic landscape of rapid repeated evolutionary adaptation to toxic pollution in wild
532 fish.
- 533 10. Zhao L, Wit J, Svetec N, Begun DJ. Parallel Gene Expression Differences between Low
534 and High Latitude Populations of *Drosophila melanogaster* and *D . simulans*. 2015;1–25.
- 535 11. Shapiro MD, Bell MA, Kingsley DM. Parallel genetic origins of pelvic reduction in
536 vertebrates. 2006;103(37):13753–8.
- 537 12. Miller CT, Swartz ME, Khuu PA, Walker MB, Eberhart JK, Kimmel CB. *mef2ca* is

- 538 required in cranial neural crest to effect Endothelin1 signaling in zebrafish. 2007;308:144–
539 57.
- 540 13. Shen Y, Liang L, Li G, Murphy RW, Zhang Y. Parallel Evolution of Auditory Genes for
541 Echolocation in Bats and Toothed Whales. 2012;8(6).
- 542 14. Losos JB. CONVERGENCE , ADAPTATION , AND CONSTRAINT. 2011;1827–40.
- 543 15. Bolivar P, Burri R, Dutoit L, Mugal CF, Nater A, Aken B, et al. Divergence in gene
544 expression within and between two closely related flycatcher species. 2016;2015–28.
- 545 16. Davidson JH, Balakrishnan CN. Gene Regulatory Evolution During Speciation in a
546 Songbird. 2016;6(May):1357–64.
- 547 17. Poelstra JW, Vijay N, Bossu CM, Lantz H, Ryll B, Müller I, et al. The genomic landscape
548 underlying phenotypic integrity in the face of gene flow in crows. *Science* [Internet]. 2014
549 Jun 20 [cited 2016 Mar 9];344(6190):1410–4. Available from:
550 <http://science.sciencemag.org/content/344/6190/1410.abstract>
- 551 18. Miller CT, Beleza S, Pollen AA, Schluter D, Kittles RA, Shriver MD, et al. cis -
552 Regulatory Changes in Kit Ligand Expression and Parallel Evolution of Pigmentation in
553 Sticklebacks and Humans. 2007;1179–89.
- 554 19. Quin KEO, Hofmann CM, Hofmann HA, Carleton KL. Parallel Evolution of Opsin Gene
555 Expression in African Cichlid Fishes *Research article*. 2010;27(12):2839–54.
- 556 20. Ahi EP, Kapralova KH, Pálsson A, Maier VH, Gudbrandsson J, Snorrason SS, et al.
557 Transcriptional dynamics of a conserved gene expression network associated with
558 craniofacial divergence in Arctic charr. 2014;1–19.
- 559 21. Enard W, Khaitovich P, Klose J, Heissig F, Giavalisco P, Nieselt-struwe K, et al. Intra-
560 and Interspecific Variation in Primate Gene Expression Patterns. 2002;296(April):340–4.
- 561 22. Rosenblum EB, Parent CE, Brandt EE. The Molecular Basis of Phenotypic Convergence.
- 562 23. Conte GL, Arnegard ME, Peichel CL, Schluter D. The probability of genetic parallelism
563 and convergence in natural populations. *Proc R Soc B Biol Sci* [Internet]. 2012 Oct 17
564 [cited 2015 Oct 5];279(1749):5039–47. Available from:
565 [http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=3497250&tool=pmcentrez&re
566 ndertype=abstract](http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=3497250&tool=pmcentrez&rendertype=abstract)
- 567 24. Schluter D, Clifford EA, Nemethy M, Mckinnon JS. Parallel Evolution and Inheritance of
568 Quantitative Traits. 2004;163(6).

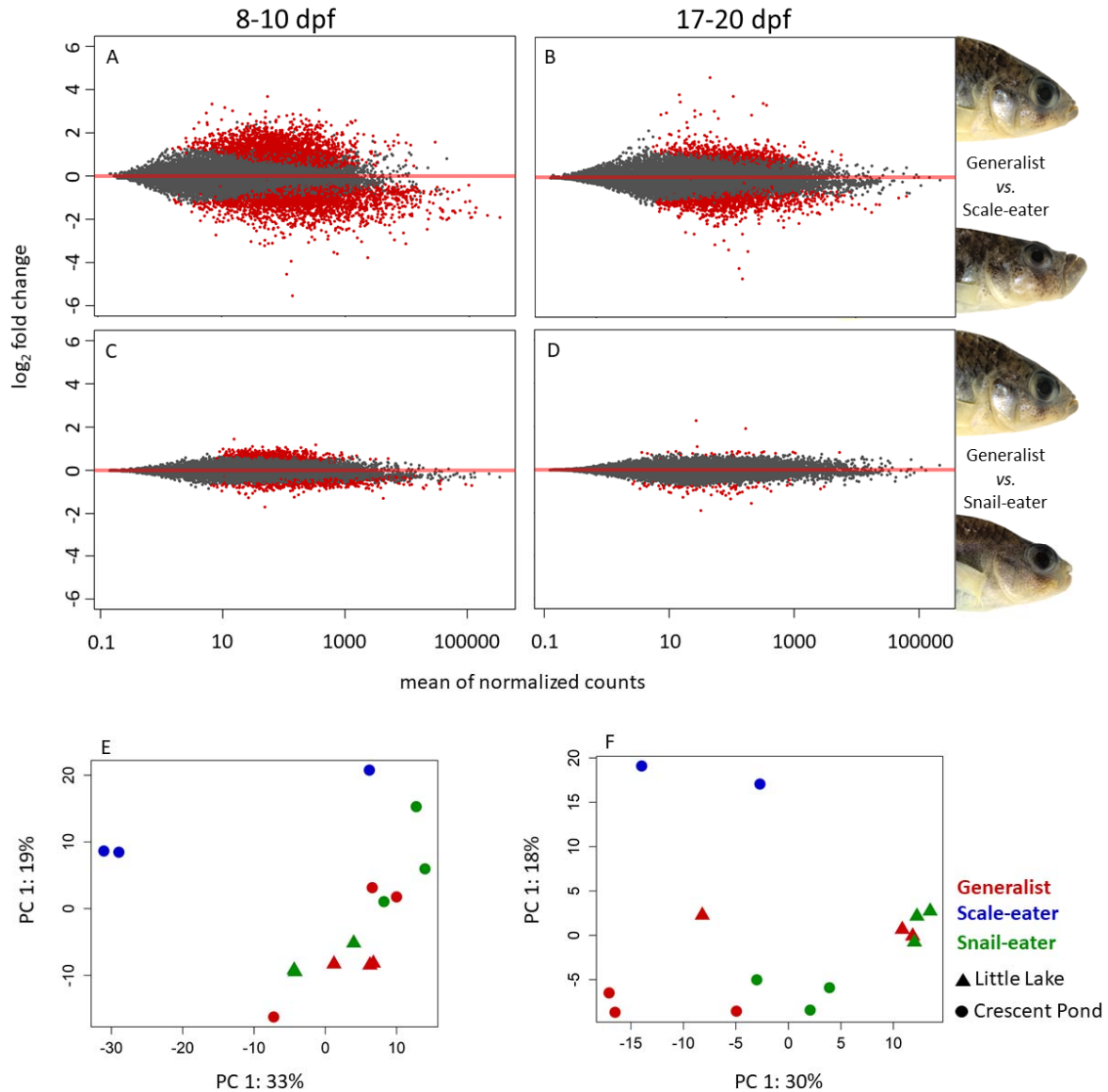
- 569 25. Pavey SA, Nosil P, Rogers SM. The role of gene expression in ecological speciation.
570 2010;1206:110–29.
- 571 26. Mylroie, J.E, Hagey FM. Terrestrial and Shallow Marine Geology of the Bahamas and
572 Bermuda. Boulder, CO: Geological Society of America; 1995. 77-90 p.
- 573 27. Turner BJ, Duvernell DD, Bunt TM, Barton MG. Reproductive isolation among endemic
574 pupfishes (Cyprinodon) on San Salvador Island, Bahamas: Microsatellite evidence. *Biol J*
575 *Linn Soc.* 2008;95(3):566–82.
- 576 28. Martin CH, Wainwright PC. Trophic novelty is linked to exceptional rates of
577 morphological diversification in two adaptive radiations of *Cyprinodon* pupfish. *Evolution*
578 [Internet]. 2011 Aug [cited 2016 Jan 14];65(8):2197–212. Available from:
579 <http://www.ncbi.nlm.nih.gov/pubmed/21790569>
- 580 29. Martin CH. Context-dependence in complex adaptive landscapes: frequency and trait-
581 dependent selection surfaces within an adaptive radiation of Caribbean pupfishes.
582 *Evolution (N Y)*. 2016;(Simpson 1944):1–57.
- 583 30. Pavlidis P, Živković D, Stamatakis A, Alachiotis N. SweeD: Likelihood-based detection
584 of selective sweeps in thousands of genomes. *Mol Biol Evol.* 2013;30(9):2224–34.
- 585 31. Mcgirr JA, Martin CH. Novel Candidate Genes Underlying Extreme Trophic
586 Specialization in Caribbean Pupfishes. 2016;34(4):873–88.
- 587 32. Richards EJ, Martin CH. Adaptive introgression from distant Caribbean islands
588 contributed to the diversification of a microendemic adaptive radiation of trophic
589 specialist pupfishes. 2017. 1-35 p.
- 590 33. Dobin A, Davis CA, Schlesinger F, Drenkow J, Zaleski C, Jha S, et al. Sequence analysis.
591 2013;29(1):15–21.
- 592 34. Liao Y, Smyth GK, Shi W. Sequence analysis featureCounts: an efficient general
593 purpose program for assigning sequence reads to genomic features. 2014;30(7):923–30.
- 594 35. Ewels P, Lundin S, Max K. Data and text mining MultiQC: summarize analysis results
595 for multiple tools and samples in a single report. 2016;32(June):3047–8.
- 596 36. Love MI, Huber W, Anders S. Moderated estimation of fold change and dispersion for
597 RNA-seq data with DESeq2. 2014;1–21.
- 598 37. Society RS. Controlling the False Discovery Rate: A Practical and Powerful Approach
599 to Multiple Testing Author (s): Yoav Benjamini and Yosef Hochberg Source: Journal

- 600 of the Royal Statistical Society . Series B (Methodological), Vol . 57 , No . 1 Published
601 by □: Wiley for the Royal Statistical Society Stable URL □:
602 <http://www.jstor.org/stable/2346101>. 2017;57(1):289–300.
- 603 38. Lencer ES, Warren WC, Harrison R, Mccune AR. The *Cyprinodon variegatus* genome
604 reveals gene expression changes underlying differences in skull morphology among
605 closely related species. 2017;1–33.
- 606 39. DePristo MA, Banks E, Poplin R, Garimella K V, Maguire JR, Hartl C, et al. A
607 framework for variation discovery and genotyping using next-generation DNA sequencing
608 data. *Nat Genet* [Internet]. 2011 May [cited 2014 Jul 9];43(5):491–8. Available from:
609 [http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=3083463&tool=pmcentrez&re](http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=3083463&tool=pmcentrez&rendertype=abstract)
610 [ndertype=abstract](http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=3083463&tool=pmcentrez&rendertype=abstract)
- 611 40. Martin CH, Wainwright PC. A remarkable species flock of *Cyprinodon* pupfishes
612 endemic to San Salvador Island, Bahamas. *Bull Peabody Museum Nat Hist* [Internet].
613 2013;54(2):231–41. Available from: [file:///Users/antoniaford/Library/Application](file:///Users/antoniaford/Library/Application%20Support/Papers2/Articles/2013/Martin/Martin_Bulletin_of_the_Peabody_Museum_of_Natural_History_2013-1.pdf)
614 [Support/Papers2/Articles/2013/Martin/Martin_Bulletin_of_the_Peabody_Museum_of_Nat](file:///Users/antoniaford/Library/Application%20Support/Papers2/Articles/2013/Martin/Martin_Bulletin_of_the_Peabody_Museum_of_Natural_History_2013-1.pdf)
615 [ural_History_2013-1.pdf](file:///Users/antoniaford/Library/Application%20Support/Papers2/Articles/2013/Martin/Martin_Bulletin_of_the_Peabody_Museum_of_Natural_History_2013-1.pdf)<http://www.bioone.org/doi/abs/10.3374/014.054.0201>
- 616 41. Galtier N, Depaulis F, Barton NH. Detecting Bottlenecks and Selective Sweeps From
617 DNA Sequence Polymorphism. 2000;
- 618 42. Nielsen R. Molecular signatures of natural selection. *Annu Rev Genet* [Internet].
619 2005;39(1):197–218. Available from: [citeulike-article-](http://dx.doi.org/10.1146/annurev.genet.39.073003.112420)
620 [id:524184%5Cnhttp://dx.doi.org/10.1146/annurev.genet.39.073003.112420](http://dx.doi.org/10.1146/annurev.genet.39.073003.112420)
- 621 43. Schiffels S, Durbin R. Inferring human population size and separation history from
622 multiple genome sequences. *Nat Publ Gr* [Internet]. 2014;46(8):919–25. Available from:
623 <http://dx.doi.org/10.1038/ng.3015>
- 624 44. Gene T, Consortium O. Gene Ontology □: tool for the. 2000;25(may):25–9.
- 625 45. Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, et al. BLAST
626 + □: architecture and applications. 2009;9:1–9.
- 627 46. Tomislav S. REVIGO Summarizes and Visualizes Long Lists of Gene Ontology Terms.
628 2011;6(7).
- 629 47. Papakostas S, Vøllestad LA, Bruneaux M, Aykanat T, Vanoverbeke J, Ning M, et al.
630 conditions. 2014;(316):1–9.

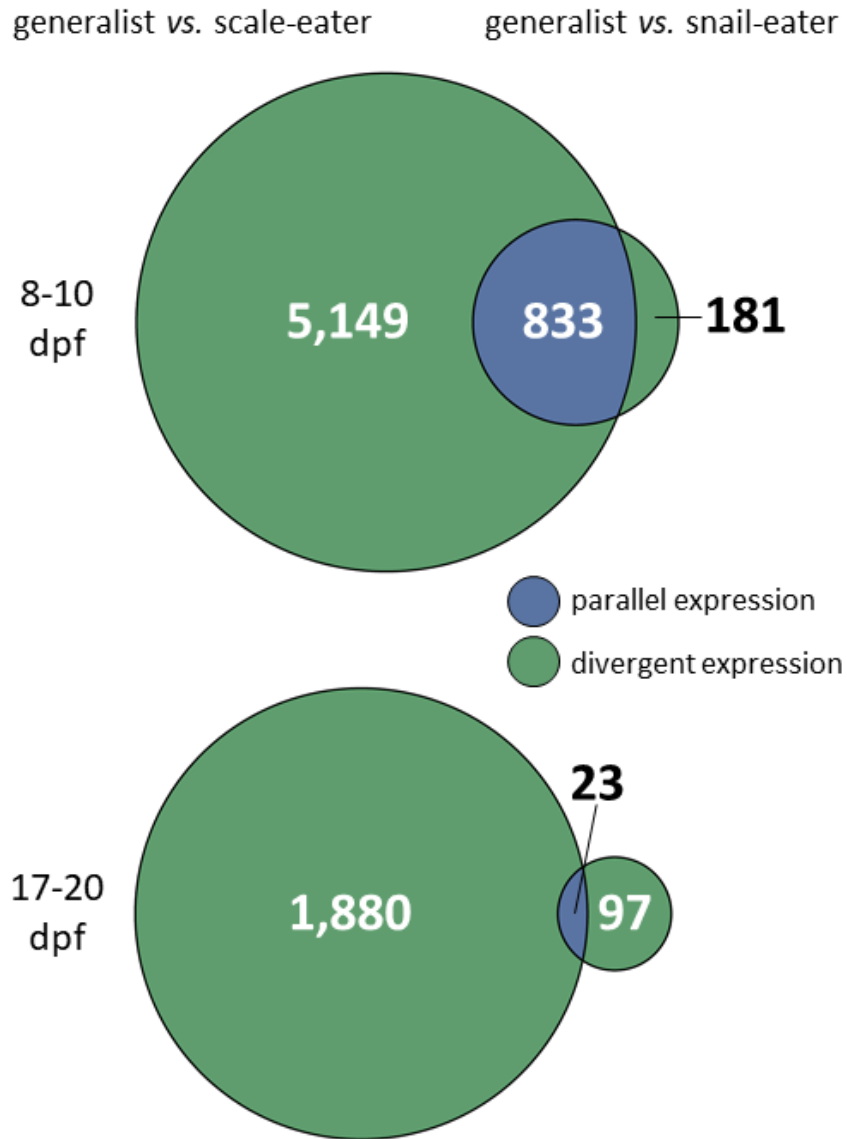
- 631 48. Szklarczyk D, Franceschini A, Wyder S, Forslund K, Heller D, Huerta-cepas J, et al.
632 STRING v10: protein – protein interaction networks , integrated over the tree of life.
633 2015;43(October 2014):447–52.
- 634 49. Bastian F, Parmentier G, Roux J, Moretti S, Lyon U De, Génomique I De, et al. Bgee:
635 Integrating and Comparing Heterogeneous Transcriptome Data Among Species. :124–31.
- 636 50. Ouedraogo M, Bettembourg C, Bretaudeau A, Sallou O, Diot C. The Duplicated Genes
637 Database: Identification and Functional Annotation of Co-Localised Duplicated Genes
638 across Genomes. 2012;7(11):1–8.
- 639 51. Martin CH, Wainwright PC. On the measurement of ecological novelty: scale-eating
640 pupfish are separated by 168 my from other scale-eating fishes. PLoS One [Internet].
641 2013;8(8):e71164. Available from:
642 [http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=3747246&tool=pmcentrez&](http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=3747246&tool=pmcentrez&rendertype=abstract)
643 [ndertype=abstract](http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=3747246&tool=pmcentrez&rendertype=abstract)
- 644 52. He X, Zhang J. Toward a Molecular Understanding of Pleiotropy.
645 2006;1891(August):1885–91.
- 646 53. Safari-alighiarloo N, Taghizadeh M, Rezaei-tavirani M, Goliaei B, Peyvandi AA. Protein-
647 protein interaction networks (PPI) and complex diseases. 2014;7(7):17–31.
- 648 54. Stern DL, Orgogozo V. THE LOCI OF EVOLUTION: HOW PREDICTABLE IS
649 GENETIC EVOLUTION? 2008;2155–77.
- 650 55. Article R. Developmental Patterning and Evolution of the Mammalian Viscerocranium:
651 1997;155(September 1996):139–55.
- 652 56. Cerny R, Lwigale P, Ericsson R, Meulemans D, Epperlein H, Bronner-fraser M.
653 Developmental origins and evolution of jaws: new interpretation of b maxillary Q and b
654 mandibular Q. 2004;276:225–36.
- 655 57. Wagner GP, Zhang J. The pleiotropic structure of the genotype – phenotype map: the
656 evolvability of complex organisms. Nat Publ Gr [Internet]. 2011;12(3):204–13. Available
657 from: <http://dx.doi.org/10.1038/nrg2949>
- 658 58. Tulchinsky AY, Johnson NA, Porter AH. Hybrid Incompatibility Despite Pleiotropic
659 Constraint in a Sequence-Based Bioenergetic. 2014;198(December):1645–54.
- 660 59. Zhang J. Parallel Functional Changes in the Digestive RNases of Ruminants and
661 Colobines by Divergent Amino Acid Substitutions. 2001;1310–7.

- 662 60. Carvalho LS, Cowing JA, Wilkie SE, Bowmaker JK, Hunt DM. The Molecular Evolution
663 of Avian Ultraviolet- and Violet-Sensitive Visual Pigments. 2006;
- 664 61. Holtzinger A, Evans T. NIH Public Access. 2008;312(2):613–22.
- 665 62. Lynch M, Force A. The Probability of Duplicate Gene Preservation by
666 Subfunctionalization. 2000;
- 667 63. Kassahn KS, Dang VT, Wilkins SJ, Perkins AC, Ragan MA. Evolution of gene function
668 and regulatory control after whole-genome duplication: Comparative analyses in
669 vertebrates. 2009;1404–18.
- 670 64. Qian W, Liao B, Chang AY, Zhang J. Maintenance of duplicate genes and their functional
671 redundancy by reduced expression. Trends Genet [Internet]. 26(10):425–30. Available
672 from: <http://dx.doi.org/10.1016/j.tig.2010.07.002>
- 673 65. MARTIN, P. ERICKSON CM. The genetic architecture of novel trophic specialists:
674 larger effect sizes are associated with exceptional oral jaw diversification in a pupfish
675 adaptive radiation. 2017;624–38.
- 676 66. Martin CH. The cryptic origins of evolutionary novelty: 1000-fold faster trophic
677 diversification rates without increased ecological opportunity or hybrid swarm. 2016;
- 678 67. Martin A, Orgogozo V. THE LOCI OF REPEATED EVOLUTION: A CATALOG OF
679 GENETIC HOTSPOTS OF PHENOTYPIC VARIATION. 2013;(McGhee 2011):1235–
680 50.
- 681 68. Kronforst MR, Barsh GS, Kopp A, Mallet J, Monteiro A, Mullen SP, et al.
682 INTERNATIONAL FEDERATION OF PIGMENT CELL SOCIETIES · SOCIETY FOR
683 MELANOMA RESEARCH PIGMENT CELL & MELANOMA of diversity and
684 convergence in animal pigmentation. 2012;
- 685 69. Garfield DA, Runcie DE, Babbitt CC, Haygood R, Nielsen WJ, Wray GA. The Impact of
686 Gene Expression Variation on the Robustness and Evolvability of a Developmental Gene
687 Regulatory Network. 2013;11(10).
- 688 70. Ferna A, Tena JJ, Gonza C, Parra-acero H, Cross JW, Rigby PWJ, et al. Comparative
689 epigenomics in distantly related teleost species identifies conserved cis -regulatory nodes
690 active during the vertebrate phylotypic period. 2014;1075–85.
- 691 71. Kalinka AT, Varga KM, Gerrard DT, Preibisch S, Corcoran DL, Jarrells J, et al.
692 developmental hourglass model. Nature [Internet]. 2010;468(7325):811–4. Available

- 693 from: <http://dx.doi.org/10.1038/nature09634>
- 694 72. Reddiex AJ, Gosden TP, Bonduriansky R, Chenoweth SF. Sex-Specific Fitness
695 Consequences of Nutrient Intake and the Evolvability of Diet Preferences. 2013;182(1).
- 696 73. Comeault AA, Serrato-capuchina A, Turissini DA, Mclaughlin PJ, David JR, Matute DR.
697 A nonrandom subset of olfactory genes is associated with host preference in the fruit fly
698 *Drosophila orena*. 2017;1–13.
- 699 74. Conesa A, Madrigal P, Tarazona S, Gomez-cabrero D, Cervera A, Mcpherson A, et al. A
700 survey of best practices for RNA-seq data analysis. 2016;1–19.
- 701 75. Lin Y, Golovnina K, Chen Z, Lee HN, Negron YLS, Sultana H, et al. Comparison of
702 normalization and differential expression analyses using RNA-Seq data from 726
703 individual *Drosophila melanogaster*. BMC Genomics [Internet]. 2016;1–20. Available
704 from: <http://dx.doi.org/10.1186/s12864-015-2353-z>
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719 **Fig. 1. Differential gene expression between generalists and trophic specialists.** Red points
720 represent genes that are differentially expressed in 8-10 dpf whole-body tissue (A, C) and 17-20
721 dpf craniofacial tissue (B, C) between generalists vs. scale-eaters (A, B) and generalist vs. snail-
722 eaters (C, D). Bottom panels show the top two principal components accounting for a combined
723 52% (8-10 dpf; E) and 48% (17-20 dpf; F) of the total variation between samples across 413
724 million reads mapped to annotated features. Triangles represent samples from Little Lake and
725 circles represent samples from Crescent Pond. Generalists are shown in red, scale-eaters in blue,
726 and snail-eaters in green.

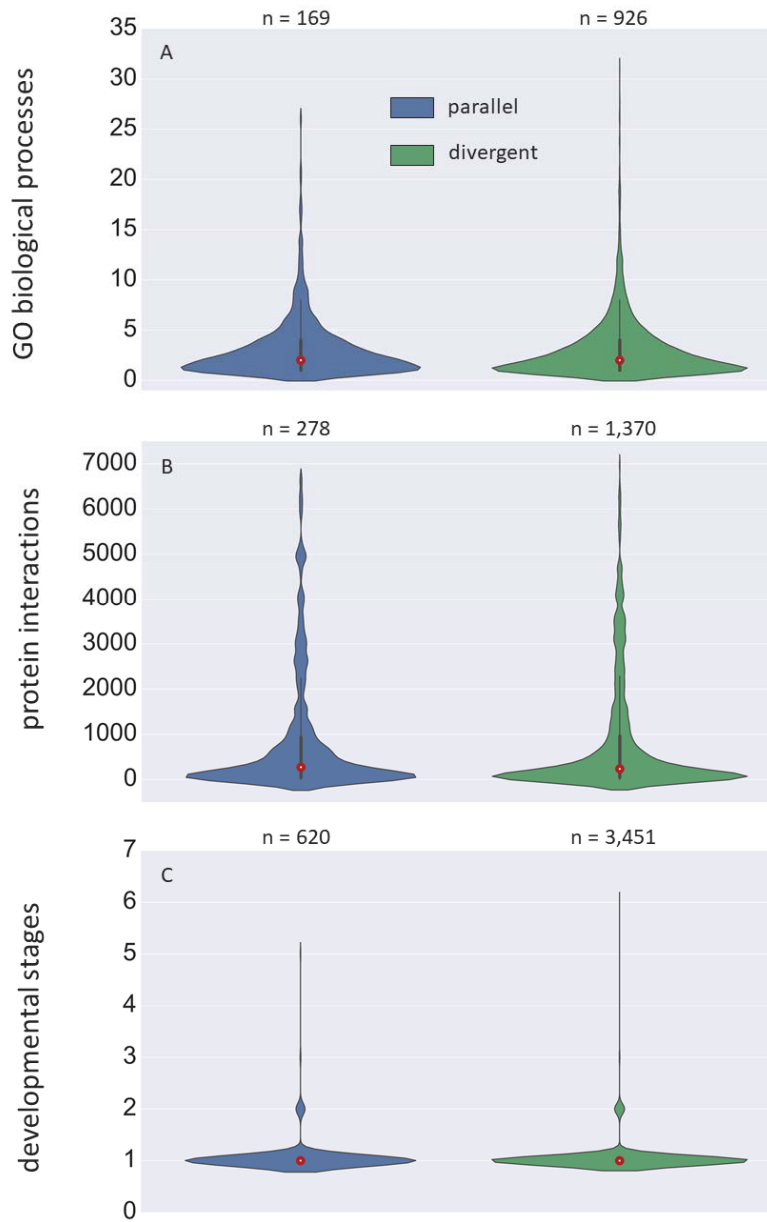


727

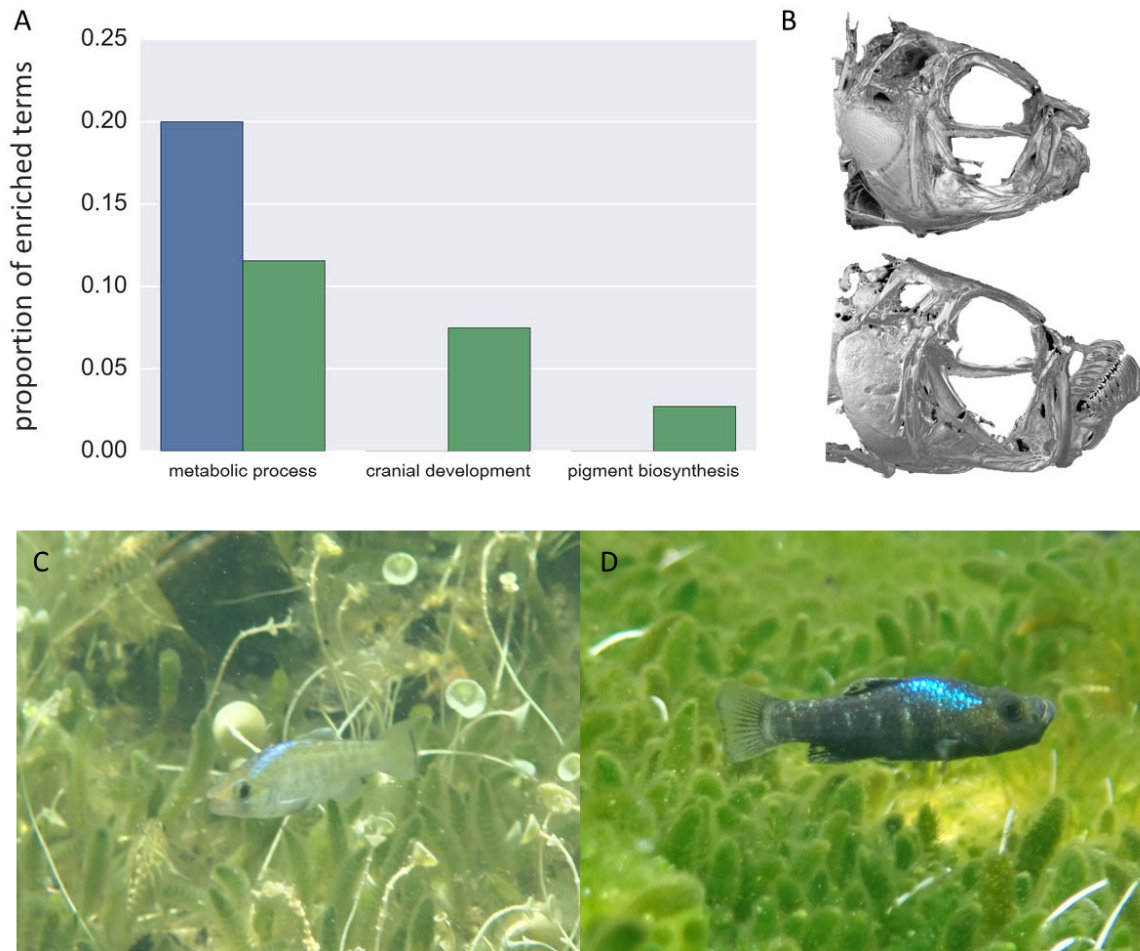
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729 **Fig. 2. Significant parallel gene expression between specialists despite divergent trophic**
730 **adaptation.** Circles illustrate genes differentially expressed in 8-10 dpf whole-body tissue (top)
731 and 17-20 dpf craniofacial tissue (bottom) for generalists vs. scale-eaters (left) and generalists vs.
732 scale-eaters (right). Genes that show the same expression patterns in specialists relative to
733 generalists are blue, and those showing divergent expression patterns unique to each specialist
734 are green. Significantly more genes show the same expression pattern (either up or
735 downregulated) in specialists relative to generalist gene expression at 8-10 dpf than expected by
736 chance (Fig. S2; 100,000 permutations; $P < 1.0 \times 10^{-5}$). Alternatively, significantly fewer genes
737 show the same expression pattern in 17-20 dpf craniofacial tissue (Fig. S2; 100,000
738 permutations; $P < 7.0 \times 10^{-4}$).

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761 **Fig. 3. Genes showing parallel expression patterns in specialists are not more pleiotropic**
762 **than genes showing divergent expression.** Violin plots show the distribution of pleiotropy
763 estimates (GO biological processes (A), protein-protein interactions (B), and developmental
764 stages expressed (C)) for genes showing parallel expression patterns (blue) and divergent
765 expression patterns (green) in specialists relative to generalists. Red dots show the median, thick
766 black bars show interquartile ranges and thin bars show 95% confidence intervals. Genes
767 showing parallel expression are not significantly more or less pleiotropic than divergently
768 expressed genes (GLM; biological processes: $P = 0.67$; PPIs: $P = 0.09$; developmental stages: P
769 $= 0.89$).



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771 **Fig. 4. Parallel gene expression underlies metabolic adaptations while divergent expression**
772 **underlies trophic morphology** A) Genes showing parallel expression in specialists (blue) and
773 genes showing divergent expression (green) are contrastingly enriched for terms describing
774 metabolic processes (parallel: %20; divergent: %11). Genes showing divergent expression are
775 enriched for cranial skeleton development (7% of terms) and pigment biosynthesis (3% of
776 terms). B) μ CT scans show drastic craniofacial divergence between snail-eaters (top) and scale-
777 eaters (bottom). Bottom panels show male breeding coloration characteristic of light snail-eaters
778 (C) and dark scale-eaters (D).